

1 **Cephalexin interaction with biosolids-derived dissolved organic matter:**
2 **binding mechanism and implications for adsorption by biochar and clay**

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24 **Table S1.** Physicochemical characteristics of biochar and montmorillonite used in adsorption
 25 study.

Material	N ₂ specific surface area (m ² g ⁻¹)	Cation exchange capacity (meq 100 g ⁻¹)	%C	%N	Particle size (μm)
Date palm petiole biochar	71.07	1.48	71.03	0.25	<180
SWy-1 ^a	31.82	76.4			1.15 ¹

26 ^a SWy-1 surface area and cation exchange capacity from The Clay Minerals Society (clays.org/sourceclays_data/)

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28 **Table S2.** Basic chemical and optical characteristics of DOM used in experiments. Properties
 29 were determined in deionized water. Standard deviations of replicated measurements (n=3) are
 30 presented in parentheses.

% C	SUVA ₂₅₄ (L mg ⁻¹ C m ⁻¹)	Fluorescence index (FI)	Humification index (HIX)	Biological fluorescence index (BIX)	pH
38.4 (0.002)	1.01 (0.06)	1.7 (0.03)	0.36 (0.01)	1.15 (0.04)	5.78 (0.03)

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36 **Table S3.** Ryan-Weber fitting parameters for cephalexin-DOM binding at experimental pH
 37 values.

pH	Log K _c	L _t (M)	Root-mean-square error
4.0	8.48	8.57 • 10 ⁻⁴	4.36 • 10 ⁻²
5.0	7.16	2.48 • 10 ⁻³	4.94 • 10 ⁻²
7.0	5.33	3.99 • 10 ⁻³	2.26 • 10 ⁻²

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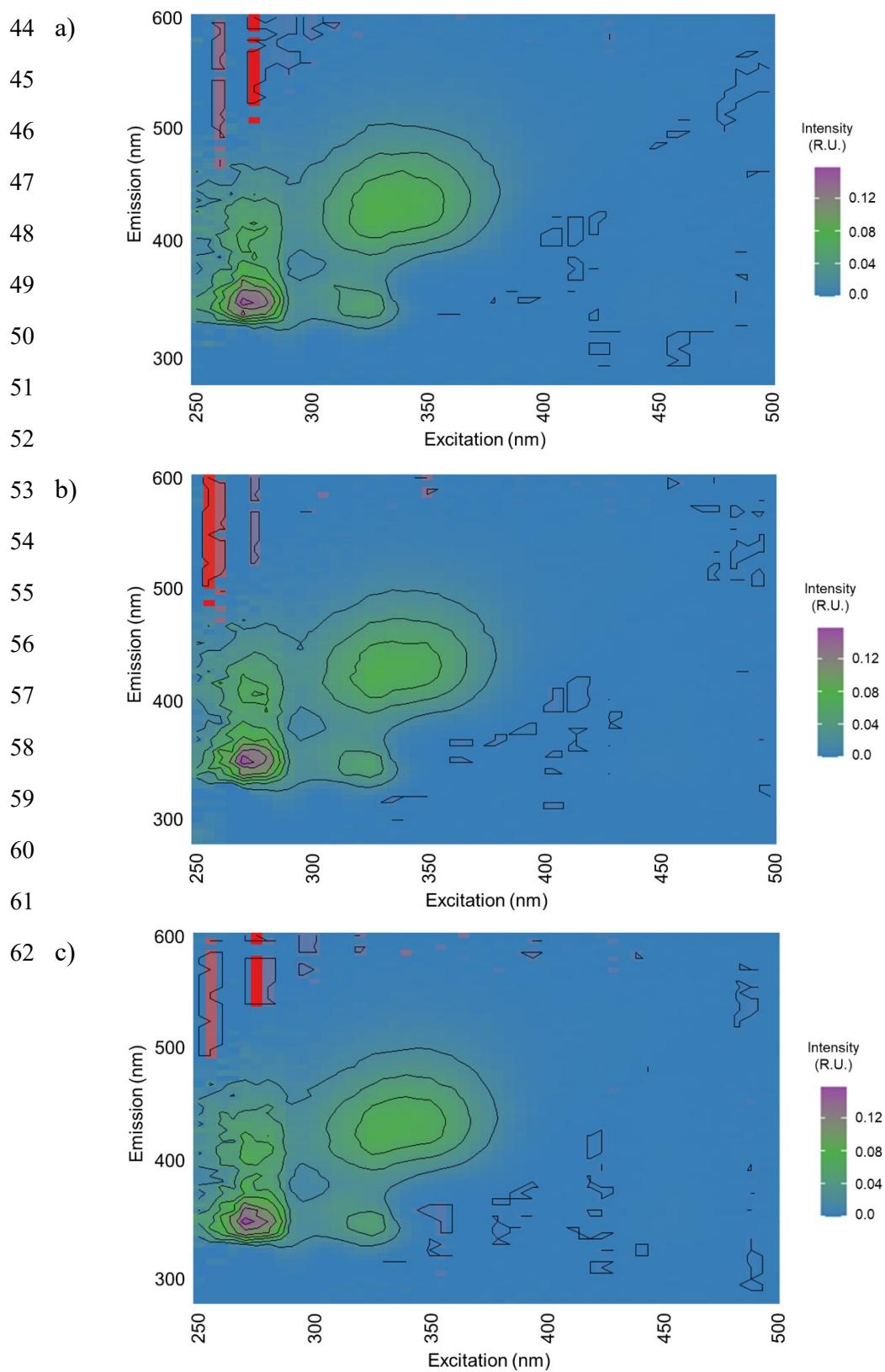
39 **Table S4.** Ryan-Weber fitting parameters for cephalexin-DOM binding at experimental
 40 background ionic strength values and cation charges.

Background ionic strength condition	Background cation	Log K _c	L _t (M)	Root-mean-square error
DDIW	N/A	6.00	2.41 • 10 ⁻³	4.29 • 10 ⁻²
I = 0.10 M	Na ⁺	6.41	2.16 • 10 ⁻³	1.99 • 10 ⁻²
I = 0.10 M	Ca ⁺²	5.93	1.78 • 10 ⁻³	4.31 • 10 ⁻²

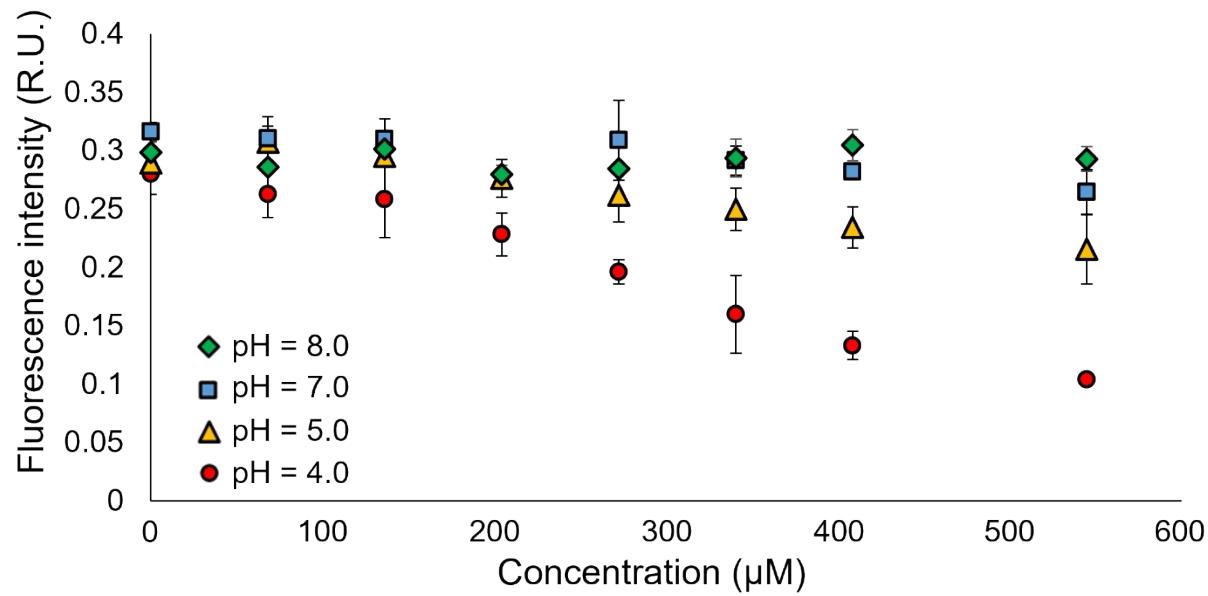
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43 **Figure S1.** EEMs of 50 ppm cephalaxin collected at a) pH = 4.0, b) pH = 5.0 and c) pH = 7.0.



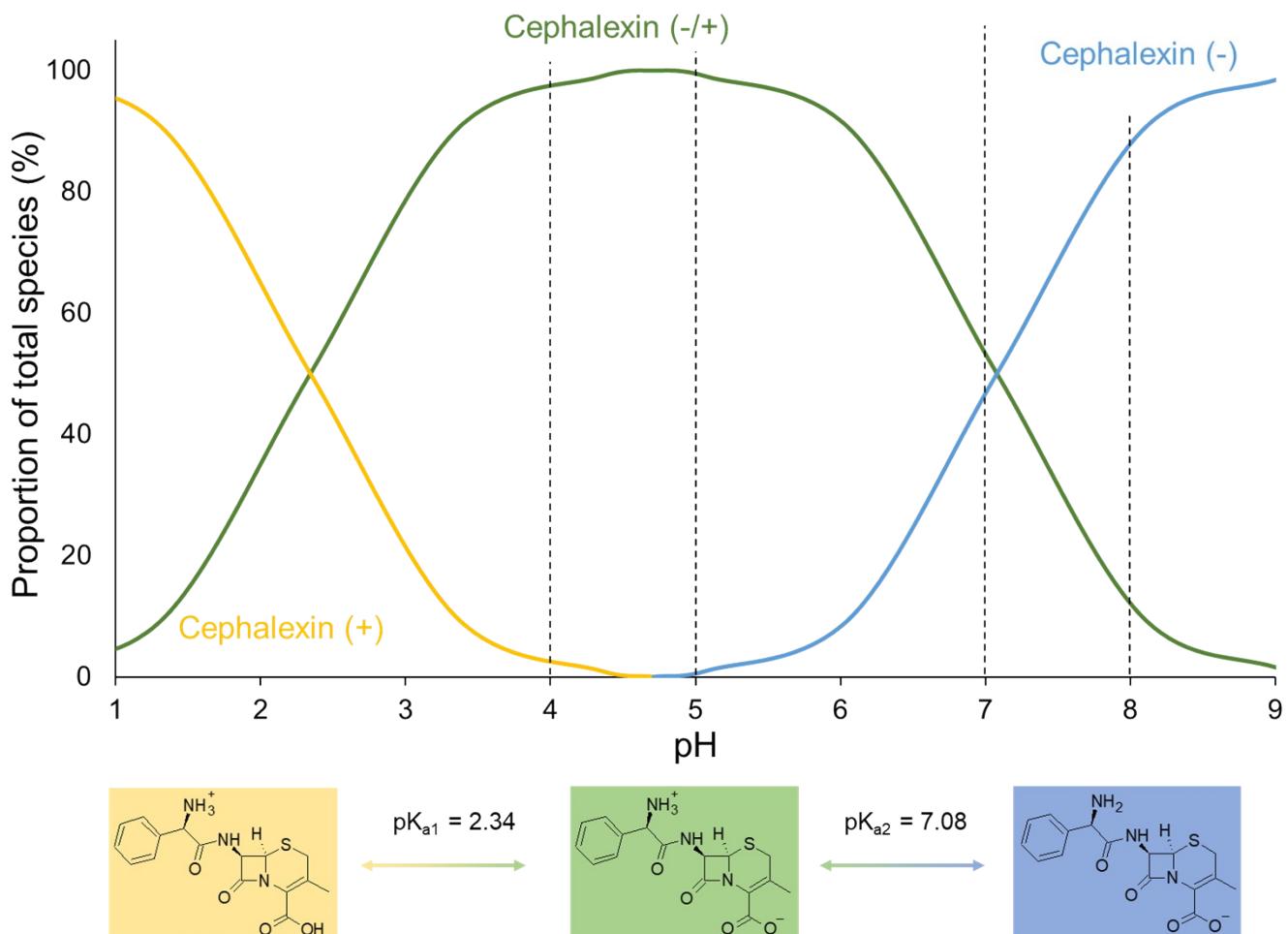
63 **Figure S2.** Fluorescence intensities versus cephalexin concentration for $[DOM] = 2.5 \text{ mg C L}^{-1}$
64 in synthetic wastewater at pH = 4.0 (red), 5.0 (yellow), 7.0 (blue) and 8.0 (green). Error bars



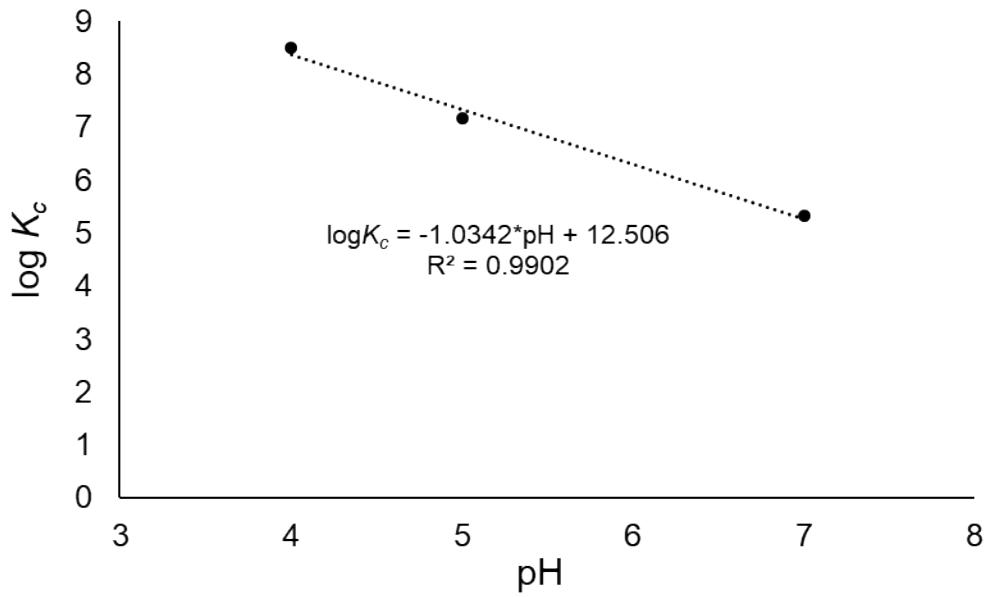
65 represent standard deviations of experimental replicates (n=3).

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67 **Figure S3.** Cephalexin charge and chemical speciation variation with pH. Experimental pH
68 values are denoted by the dashed vertical lines. Dissociation constants were determined
69 experimentally by Mrestani et al.²

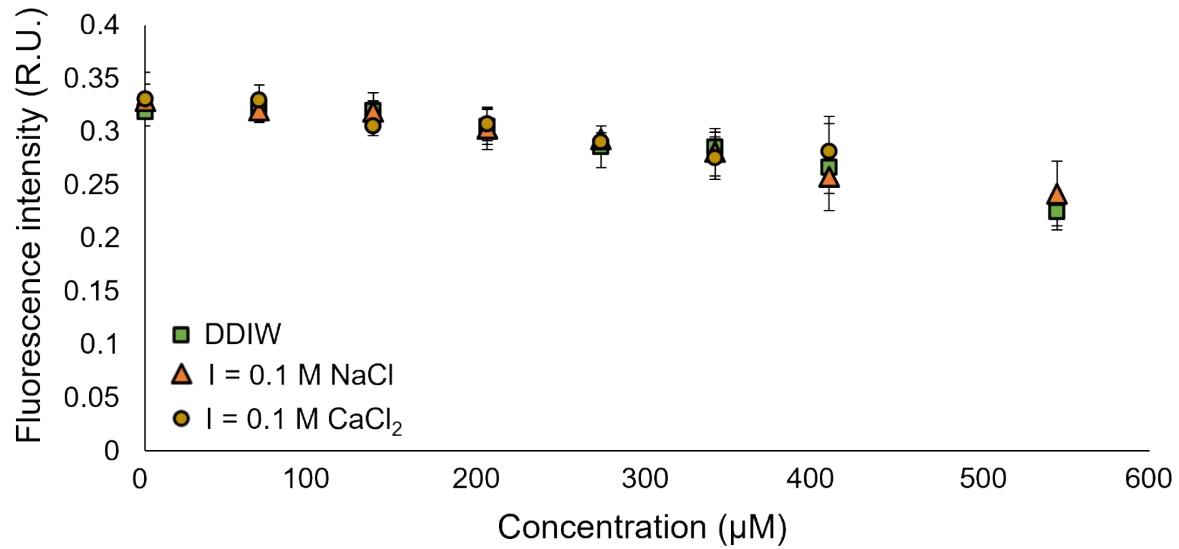


73 **Figure S4.** Relationship between $\log K_c$ and pH for cephalexin-DOM binding in a synthetic
74 wastewater matrix



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78 **Figure S5.** Fluorescence intensities versus cephalexin concentration for $[DOM] = 2.5 \text{ mg C L}^{-1}$
79 in ultrapure water (green), $I = 0.1 \text{ M NaCl}$ (orange) and $I = 0.1 \text{ M CaCl}_2$ (brown). Error bars
80 represent standard deviations of experimental replicates ($n=3$).



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83 **References**

- 84 1. M.A. Anderson and P.M. Bertsch, Electrophoretic Mobility and Particle Size of Clays
85 Using Laser Doppler Velocimetry-Photon Correlation Spectroscopy, Soil Sci. Soc. Am.
86 J., 1993, 57, 1641-1643.
- 87 2. Y. Mrestani, R. Neubert, A. Munk and M. Wiese, Determination of dissociation constants
88 of cephalosporins by capillary zone electrophoresis, J. Chromatogr. A, 1998, 803, 273-
89 278.