

## Electrochemical Hydrogen Isotope Exchange of Amines Controlled by Alternating Current Frequency

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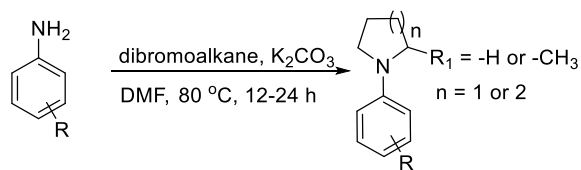
## 1. General information

All reactions were carried out in oven-dried glassware under a positive pressure of argon using Schlenk line techniques. TLC was performed using pre-coated glass plates with 230-400 mesh silica gel impregnated with a fluorescent indicator (250 nm). Visualization was accomplished using a UV light. Flash chromatography was performed on silica gel flash chromatography columns, Teledyne Isco Combi Flash R<sub>f</sub> system utilizing normal phase pre-column cartridges and gold high-performance columns. Purifications were performed using Ethyl acetate: Hexane (v:v) eluting with indicated gradient mixture. All proton (<sup>1</sup>H) nuclear magnetic resonance spectra were recorded on a 400 or 600 MHz spectrometer. All carbon (<sup>13</sup>C) nuclear magnetic resonance spectra were recorded on a 101 or 151 MHz NMR spectrometer. All fluorine (<sup>19</sup>F) nuclear magnetic resonance spectra were recorded on a 564 MHz NMR spectrometer. Chemical shifts were expressed in parts per million (δ scale) and were referenced to residual CDCl<sub>3</sub> (<sup>1</sup>H: δ 7.27 ppm, <sup>13</sup>C: δ 77.00 ppm). The data are presented as follows: chemical shift (δ), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, quint = quintet, sept= septate, m = multiplet, dd = doublet of doublet, dt = doublet of triplet, td = triplet of doublet), coupling constant in hertz (Hz), number of protons. High-resolution TOF mass spectrometry utilizing electrospray ionization in positive mode or electron ionization was performed to confirm the identity of the new compounds. All AC experiments were carried out using a function generator (RIGOL DG1062) combined with modulated power supply (ACCEL instruments TS200-0A), and glassy carbon (vitreous) plates (100 mm × 100 mm) were purchased from SPI Supplies. DC experiments were carried out using an IKA ElectraSyn 2.0. The discharge voltage, frequency, and waveform were monitored using an oscilloscope (SIGLENT Technologies SDS1202X-E). Voltammetry experiments were carried out using a CHI 650E potentiostat. All the mass spectra were recorded on a Thermo LTQ-Orbitrap spectrometry utilizing electrospray ionization in positive mode.

## 2. General experimental procedures for substrates synthesis

### 2A. N-aryl pyrrolidines/piperidines

Compounds **2** and **14-24** were synthesized using the following procedure.

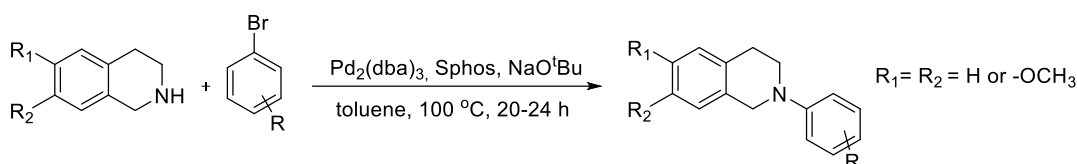


According to a literature procedure,<sup>1</sup> potassium carbonate (K<sub>2</sub>CO<sub>3</sub>) (30 mmol, 1.2 equiv.), appropriate aniline (25 mmol, 1 equiv.), and anhydrous *N,N*-dimethylformamide (DMF) (30 mL) was added in an oven-dried two-neck round-bottom flask under argon atmosphere. The resulting suspension was stirred for 5 min, and appropriate dibromoalkane (30

mmol, 1.2 equiv.) was added. The reaction mixture was heated to 80 °C for 12-24 h. After completion, the reaction mixture was cooled to room temperature and diluted with ice-cold water (100 mL). The aqueous layer was extracted with ethyl acetate (100 mL x 2), and the separated organic layer was washed with brine (50 mL) and dried over sodium sulfate. The organic layer was concentrated in *vacuo*, and the resulting crude product was purified using flash silica gel column chromatography to afford *N*-aryl pyrrolidines. All synthesized substrates are known compounds, and their analytical data were consistent with that reported in the literature.<sup>2-6</sup>

## 2B. *N*-aryl tetrahydroisoquinolines

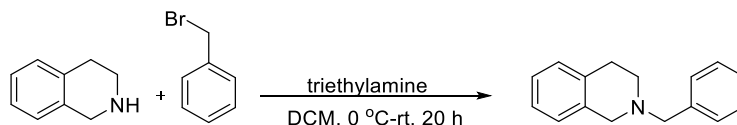
Compounds **3-13** were synthesized according to the following procedure.



According to a literature procedure,<sup>7</sup> an oven-dried pressure tube equipped with a stir bar was charged with Pd<sub>2</sub>(dba)<sub>3</sub> (3 mol %), Sphos (8 mol%), and anhydrous toluene (5 mL) under argon atmosphere. The resulting mixture was stirred for a minute, and bromoarene (2.5 mmol, 1 equiv.), tetrahydroisoquinoline (3 mmol, 1.2 equiv.), and NaO<sup>t</sup>Bu (3.5 mmol, 1.4 equiv.) were added. The pressure tube was sealed using a Teflon cap under the stream of argon and heated to 100 °C for 20-24 h. After completion, the reaction mixture was cooled to room temperature and quenched by adding water (20 mL). The aqueous layer was extracted using ethyl acetate (20 mL x 2). The combined organic layer was washed with brine (50 mL), dried over sodium sulfate, and concentrated under reduced pressure to obtain the crude product. The obtained crude product was purified using flash silica-gel column chromatography to afford the *N*-aryl tetrahydroisoquinolines. All synthesized substrates are known compounds, and their analytical data were consistent with that reported in the literature<sup>8-9</sup>.

## 2C. *N*-benzyl tetrahydroisoquinoline

Compound **1** was synthesized according to the following procedure.



According to a literature procedure,<sup>10</sup> an oven-dried two-neck round-bottom flask was charged with tetrahydroisoquinoline (10 mmol, 1 equiv.), triethylamine (30 mmol, 3 equiv.), and anhydrous dichloromethane (20 mL) using syringe under argon atmosphere. The resulting suspension was cooled to 0 °C (ice bath), and benzyl bromide

(15 mmol, 1.5 equiv.) was added dropwise. The reaction mixture was stirred for 20 h. After completion, the reaction mixture was filtered, and the organic layer was washed with brine (20 mL). The separated organic layer was dried over sodium sulfate, and the solvent was evaporated under a vacuum resulting mixture was purified using flash silica-gel column chromatography to obtain *N*-benzyl tetrahydroisoquinoline.

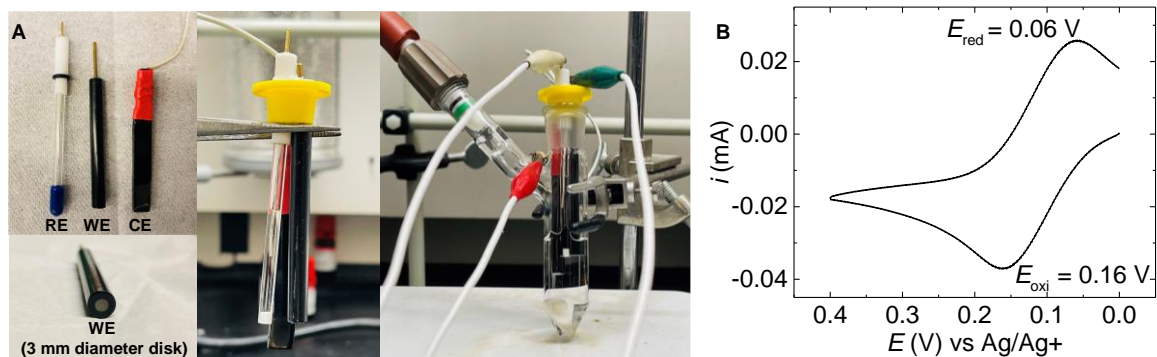
### 3. Experimental setups for electrochemical HIE reactions and voltammetric measurements.



**Fig. S1.** Photographs of the experimental setup for DC electrolysis using a commercial IKA ElectraSyn 2.0.



**Fig. S2.** Photographs of the experimental setup for AC electrolysis consisting of a TS200 amplifier from Accel Instruments and RIGOL DG1062 waveform generator, a Schlenk tube, and two glassy carbon electrodes.



**Fig. S3.** (A) Photographs of the three-electrode setup for electrochemical measurements. A 3-mm-diameter glassy carbon disk electrode as the working electrode (WE), a glassy carbon plate as the counter electrode (CE), and an Ag/Ag<sup>+</sup> as the reference electrode (RE). The Ag/Ag<sup>+</sup> electrode was prepared by filling the glass tube with 10 mM AgNO<sub>3</sub> and LiClO<sub>4</sub> (0.1 M, supporting electrolyte) in an anhydrous DMA solution. (B) Cyclic voltammogram of ferrocene in anhydrous DMA containing 0.1 M LiClO<sub>4</sub>. Scan rate = 0.1 V/s. The Ag/Ag<sup>+</sup> reference electrode potential was calibrated using ferrocene/ferrocenium ( $E_{Fc/Fc^+} = 0.11$  vs. Ag/Ag<sup>+</sup>).

#### 4. *iR* correction

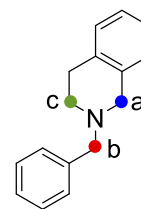
Due to the *iR* drop associated with the charging and discharging of the electrical double layer during AC electrolysis, the actual voltage available for electrochemical reactions ( $V_{ec}$ ) is lower than the amplitude of the applied AC waveform ( $V_{peak}$ ) with a frequency of  $1/2t$ , as estimated by the following equation:<sup>11</sup>

$$V_{ec} = V_{peak} (1 - \exp(-2t/R_{electrolyte}C_{EDL}))$$
, where  $R_{electrolyte}$  and  $C_{EDL}$  were the solution resistance and electrical double-layer capacitance of the AC electrolysis setup, respectively.  $R_{electrolyte}$  and  $C_{EDL}$  were measured to be 14.7  $\Omega$  and 0.15 mF using previously reported electrochemical techniques.<sup>12</sup>  $V_{ec}$  is the potential difference between the peak potentials for amine oxidation and methyl thioglycolate reduction in their cyclic voltammograms. For **1** and **2**,  $V_{ec}$  was 3.5 V and 3.1 V, respectively, according to **Fig. 1B-C**.

#### 5. Determining the relative $k_{HAT}$

The HAT kinetics is correlated with the BDE of  $\alpha$ -amino C-H bond following the Evans-Polanyi principle. More specifically,  $\Delta G^\ddagger = 0.47 \times \Delta BDE$ ,<sup>13</sup> where  $\Delta BDE$  is the BDE difference between two  $\alpha$ -amino C-H bonds and  $\Delta G^\ddagger$  is the activation energy barrier difference for HAT at the two sites. Taking **1** as the example, the BDEs of C-H bonds at sites **a** and **b** are 77.0 kcal/mol and 82.3 kcal/mol, respectively.

Thus,  $\Delta G^\ddagger = 0.47 \times (82.3 - 77.0)$  kcal/mol = 2.65 kcal/mol. The relative HAT kinetics for **a** and **b** ( $k_a/k_b$ ) are calculated from  $\Delta G^\ddagger$  using the Arrhenius equation,

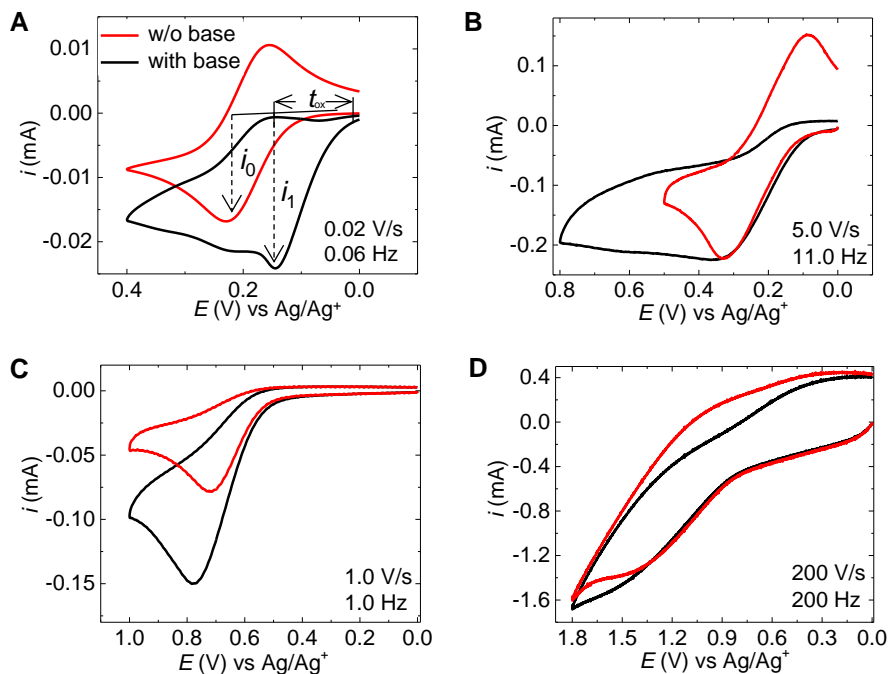


$$\frac{k_a}{k_b} = e^{-\frac{\Delta G^\ddagger}{RT}} = 0.011$$

Where  $R$  is the universal gas constant (8.314 J/(mol·K)),  $T$  is 298 K.

#### 6. Determining the relative $k_{PT}$

Relative deprotonation kinetics for **1** and **2** were estimated using a voltammetric method previously developed by our group.<sup>12</sup> Briefly, we varied the scan rate and compared the cyclic voltammograms of an amine with and without base (NaOAc was used as the base in **Fig. S4**). From the scan rates at which  $i_0 \approx i_1$  for **1** (200 V/s) and **2** (5 V/s), we can estimate the relative deprotonation kinetics to be 20 : 1. Note that we assume that the relative deprotonation kinetics does not significantly vary as the base changes from NaOAc used in the voltammetric study to thiolate used in our electrochemical HIE reactions.



**Fig. S4.** (A), (B) Cyclic voltammograms of **2** (2mM) in the presence (black curve) and absence (red curve) of NaOAc in anhydrous DMA containing LiClO<sub>4</sub> (0.1 M) at the scan rate of 0.02 V/s and 5.0 V/s respectively. (C), (D) Cyclic voltammograms of **1** (2mM) in the presence (black curve) and absence (red curve) of NaOAc in anhydrous DMA containing LiClO<sub>4</sub> (0.1 M) at the scan rate of 1.0 V/s and 200 V/s respectively.

## 7. Determining the equivalent AC frequency for the DC electrolysis conditions:

We first calculated the diffusion coefficients ( $D$ ) of **1** ( $4.34 \times 10^{-6} \text{ cm}^2/\text{s}$ ) and **2** ( $8.49 \times 10^{-6} \text{ cm}^2/\text{s}$ ) from their cyclic voltammograms at different scan rates ( $v$ ) using the Randles - Sevcik equation,<sup>14</sup>

$$i_p = 0.4463 n^{3/2} F^{3/2} A \frac{D^{1/2} c v^{1/2}}{(RT)^{1/2}}$$

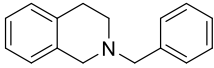
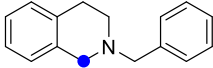
where  $i_p$  is the peak current in the voltammogram,  $n$  is the number of electrons transferred in the redox event ( $= 1$ ),  $F$  is Faraday constant (96485 C/mol),  $A$  is working electrode surface area,  $c$  is concentration,  $R$  is the universal gas constant (8.314 J/mol K), and  $T$  is 298 K. Then, the diffusion time ( $t$ ) between the two electrodes with a separation distance of  $d$  can be estimated by the following diffusion equation:

$$d = (2Dt)^{1/2}$$

For the IKA setup with  $d = 5 \text{ mm}$ ,  $t = 2.9 \times 10^4 \text{ s}$  or an equivalent  $f = 3.5 \times 10^{-5} \text{ Hz}$  for **1** and  $6.8 \times 10^{-5} \text{ Hz}$  for **2**. Similarly, for our home-built setup with  $d = 1 \text{ mm}$ ,  $f = 8.7 \times 10^{-4} \text{ Hz}$  for **1** and  $1.7 \times 10^{-3} \text{ Hz}$  for **2**.

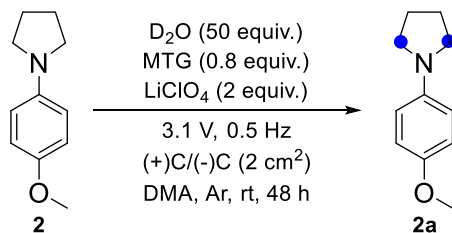
## 8. Reaction optimization for **1** and **2**

**Table S1.** Optimization table for **1**

|  | $\xrightarrow[\text{DMA, Ar, rt, 24 h}]{\begin{array}{l} \text{D}_2\text{O (50 equiv.)} \\ \text{MTG (0.3 equiv.)} \\ \text{LiClO}_4 \text{ (2 equiv.)} \\ \text{3.5 V, DC} \\ \text{(+)/C/(-)C (2 cm}^2\text{)} \end{array}}$ |  |
|---|--|--|
| Entry   | Variation from the standard conditions   | D-incorporation %  |
| 1   | None   | 60   |
| 2   | $(i\text{-Pr})_3\text{SiSH}^*$   | 54   |
| 3   | No thiol   | 00   |
| 4   | No voltage   | 00   |

**Reaction scale:** 0.25 mmol **1**, 12.5 mmol  $\text{D}_2\text{O}$ , 0.5 mmol  $\text{LiClO}_4$ , 4 mL DMA, Ar, constant voltage, 24 h, D-incorporation from  $^1\text{H NMR}$ . \* voltage amplitude = 3

**Table S2.** Optimization table for **2**



| Entry | Variation from the standard conditions            | D-incorporation (%) |
|-------|---|---------------------|
| 01    | None  | 89                  |
| 02    | No voltage  | 00                  |
| 03    | No thiol  | 00                  |
| 04    | 0.3 eq MTG  | 60                  |
| 05    | 0.5 eq MTG  | 50                  |
| 06    | 1.0 eq MTG  | 88                  |
| 07    | 1.2 eq MTG  | 81                  |
| 08    | 0.3 eq ( <i>i</i> -Pr) <sub>3</sub> SiSH as thiol | 07                  |
| 09    | NMP as the solvent (24 h)                         | 12                  |
| 10    | 2 eq NaOAc as the base                            | 50                  |
| 11    | 36 h  | 71                  |
| 12    | 30 eq $D_2O$ (0.3 eq MTG)                         | 52                  |

**Reaction scale:** 0.25 mmol **2**, 12.5 mmol  $D_2O$ , 0.2 mmol MTG, 0.5 mmol  $LiClO_4$ , 4 mL DMA, Ar, constant voltage = 3.1 V, 48 h, D-incorporation from  $^1H$  NMR.

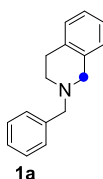
**Reproducibility of HIE reaction:**

The HIE reactions of both **1** and **2** were repeated four times. The D-incorporation and yield for **1a** are 58-60% and 42-45%, respectively. In the case of **2a**, the D-incorporation and yield are 86-89% and 40-44%, respectively. Hence, these results confirmed the reproducibility of these HIE reactions.



## 9. Experimental details and characterization data

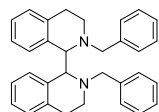
### 2-benzyl-1,2,3,4-tetrahydroisoquinoline (1, 1a)



**1:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.46 – 7.42 (m, 2H), 7.40 – 7.35 (m, 2H), 7.32 – 7.29 (m, 1H), 7.16 – 7.14 (m, 3H), 7.02 (d,  $J = 5.4$  Hz, 1H), 3.73 (s, 2H), 3.68 (s, 2H), 2.94 (d,  $J = 4.1$  Hz, 2H), 2.79 (dd,  $J = 9.4, 5.6$  Hz, 2H).

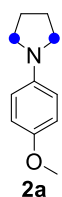
**1a: Reaction condition:** DC: 3.5 V; 24 h, 25 mg, 45% yield, 60 D%. The crude product was purified using flash silica-gel column chromatography to afford **1a** as a colorless oil.  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.44 (d,  $J = 7.3$  Hz, 2H), 7.36 (t,  $J = 7.3$  Hz, 2H), 7.32-7.29 (m, 1H), 7.17 – 7.09 (m, 3H), 7.02 – 6.98 (m, 1H), 3.76 (s, 2H), 3.71 – 3.66 (m, 0.77H), 2.95 (t,  $J = 5.9$  Hz, 2H), 2.83 (t,  $J = 5.6$  Hz, 2H). Deuterium incorporation of **1a** determined by liquid chromatography-mass spectrometry (LCMS)/IsoPat2 analysis<sup>15</sup>: 79% D (raw data and its fitting results are provided in Appendix B).

### 2,2'-dibenzyl-1,1',2,2',3,3',4,4'-octahydro-1,1'-biisoquinoline (Dimer)



$^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.42 (d,  $J = 7.4$  Hz, 2H), 7.37 – 7.31 (m, 8H), 7.29 – 7.26 (m, 2H), 7.02 – 6.91 (m, 6H), 4.20 (s, 2H), 4.07 (d,  $J = 13.2$  Hz, 2H), 3.54 (d,  $J = 13.2$  Hz, 2H), 3.32 – 3.25 (m, 2H), 2.75 – 2.67 (m, 2H), 2.64 – 2.56 (m, 2H), 2.54 – 2.48 (m, 2H).

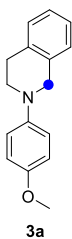
### 1-(4-methoxyphenyl)pyrrolidine (2, 2a)



**2:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  6.91 – 6.83 (m, 2H), 6.59 – 6.54 (m, 2H), 3.79 (s, 3H), 3.25 (d,  $J = 6.3$  Hz, 4H), 2.04 – 1.99 (m, 4H).

**2a: Reaction condition:** AC: 0.5 Hz, 3.1 V (peak amplitude); 48 h, 18 mg, 40% yield, 89% D. The crude product was purified using flash silica-gel column chromatography (Ethyl acetate: Hexane = 10:90 v/v) to afford **2a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  6.89 – 6.83 (m, 2H), 6.56 (d,  $J = 8.5$  Hz, 2H), 3.77 (s, 3H), 3.23 (s, 0.44H), 1.99 (d,  $J = 4.2$  Hz, 4H). Deuterium incorporation of **2a**: 85% D (mass).

### 2-(4-methoxyphenyl)-1,2,3,4-tetrahydroisoquinoline (3, 3a)

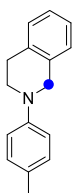


**3:**  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.23 – 7.11 (m, 4H), 7.00 (d,  $J = 9.0$  Hz, 2H), 6.90 – 6.87 (m, 2H), 4.32 (s, 2H), 3.80 (s, 3H), 3.47 (t,  $J = 5.9$  Hz, 2H), 3.01 (t,  $J = 5.8$  Hz, 2H).

**3a: Reaction condition:** DC: 3.2 V; 24 h, 36 mg, 60% yield, 93 D%. The crude product was purified using flash silica-gel column chromatography to afford **3a** as a white solid.  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.20 –

7.14 (m, 4H), 7.02 – 6.98 (m, 2H), 6.91 – 6.87 (m, 2H), 4.32 – 4.29 (m, 0.14H), 3.80 (s, 3H), 3.47 (t,  $J = 5.9$  Hz, 2H), 3.00 (t,  $J = 5.9$  Hz, 2H). Deuterium incorporation of **3a**: 97% D (mass).

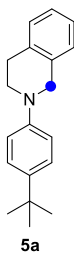
#### 2-(*p*-tolyl)-1,2,3,4-tetrahydroisoquinoline (**4, 4a**)



**4:**  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.20 – 7.14 (m, 4H), 7.11 (d,  $J = 8.4$  Hz, 2H), 6.93 (d,  $J = 8.5$  Hz, 2H), 4.37 (s, 2H), 3.53 (t,  $J = 5.9$  Hz, 2H), 3.00 (t,  $J = 5.8$  Hz, 2H), 2.29 (s, 3H).

**4a: Reaction condition:** DC: 3.3 V; 18 h, 17 mg, 30% yield, 70 D%. The crude product was purified using flash silica-gel column chromatography to afford **4a** as a white solid.  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.20 – 7.14 (m, 4H), 7.11 (d,  $J = 8.4$  Hz, 2H), 6.93 (d,  $J = 8.3$  Hz, 2H), 4.36 (d,  $J = 11.0$  Hz, 0.63H), 3.57 – 3.48 (m, 2H), 3.00 (t,  $J = 5.8$  Hz, 2H), 2.30 (s, 3H). Deuterium incorporation of **4a**: 38% D (mass).

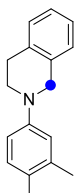
#### 2-(4-(*tert*-butyl)phenyl)-1,2,3,4-tetrahydroisoquinoline (**5, 5a**)



**5:**  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ ) 7.34 (d,  $J = 8.9$  Hz, 2H), 7.22 – 7.15 (m, 4H), 6.97 (d,  $J = 8.8$  Hz, 2H), 4.41 (s, 2H), 3.56 (t,  $J = 5.8$  Hz, 2H), 3.01 (t,  $J = 5.8$  Hz, 2H), 1.33 (s, 9H).

**5a: Reaction condition:** DC: 3.3 V; 24 h, 7 mg, 10% yield, 86 D%. The crude product was purified using flash silica-gel column chromatography to afford **5a** as a white solid.  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.36 – 7.29 (m, 2H), 7.22 – 7.14 (m, 4H), 6.96 (d,  $J = 8.7$  Hz, 2H), 4.40 – 4.37 (m, 0.26H), 3.55 (t,  $J = 5.8$  Hz, 2H), 2.99 (t,  $J = 5.8$  Hz, 2H), 1.31 (s, 9H). Deuterium incorporation of **5a**: 87% D (mass).

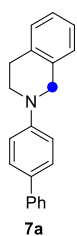
#### 2-(3,4-dimethylphenyl)-1,2,3,4-tetrahydroisoquinoline (**6, 6a**)



**6:**  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.20 – 7.14 (m, 4H), 7.06 (d,  $J = 8.2$  Hz, 1H), 6.85 (s, 1H), 6.78 (d,  $J = 8.2$  Hz, 1H), 4.37 (s, 2H), 3.52 (t,  $J = 5.8$  Hz, 2H), 3.00 (t,  $J = 5.7$  Hz, 2H), 2.27 (s, 3H), 2.21 (s, 3H).

**6a:** DC: 3.2 V; 24 h, 24 mg, 40% yield, 90 D%. The crude product was purified using flash silica-gel column chromatography to afford **6a** as a white solid.  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.20-7.15 (m, 4H), 7.06 (d,  $J = 8.2$  Hz, 1H), 6.84 (s, 1H), 6.78 (dd,  $J = 8.2, 2.3$  Hz, 1H), 4.34 (s, 0.23H), 3.52 (t,  $J = 5.9$  Hz, 2H), 3.00 (t,  $J = 5.8$  Hz, 2H), 2.27 (s, 3H), 2.21 (s, 3H). Deuterium incorporation of **6a**: 90 %D (mass).

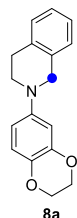
### 2-([1,1'-biphenyl]-4-yl)-1,2,3,4-tetrahydroisoquinoline (7, 7a)



**7:**  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.75 (t,  $J = 8.6$  Hz, 4H), 7.42 (t,  $J = 7.7$  Hz, 2H), 7.30 (d,  $J = 7.3$  Hz, 1H), 7.24 – 7.16 (m, 4H), 7.07 (d,  $J = 8.6$  Hz, 2H), 4.49 (s, 2H), 3.64 (t,  $J = 5.9$  Hz, 2H), 3.03 (t,  $J = 5.8$  Hz, 2H).

**7a: Reaction condition:** DC: 3.3 V; 36 h, 6 mg, 8% yield, 88 D%. The crude product was purified using flash silica-gel column chromatography to afford **7a** as a white solid.  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.75 (t,  $J = 8.6$  Hz, 4H), 7.42 (t,  $J = 7.7$  Hz, 2H), 7.30 (d,  $J = 7.4$  Hz, 1H), 7.23 – 7.18 (m, 4H), 7.06 (d,  $J = 8.6$  Hz, 2H), 4.49 – 4.46 (m, 0.24H), 3.64 (t,  $J = 5.9$  Hz, 2H), 3.03 (t,  $J = 5.9$  Hz, 2H). Deuterium incorporation of **7a**: 85% D (mass).

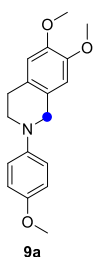
### 2-(2,3-dihydrobenzo[b][1,4]dioxin-6-yl)-1,2,3,4-tetrahydroisoquinoline (8, 8a)



**8:**  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.19 – 7.12 (m, 4H), 6.82 – 6.80 (m, 1H), 6.56 (s, 2H), 4.30 (s, 2H), 4.28 – 4.25 (m, 2H), 4.24 – 4.21 (m, 2H), 3.45 (t,  $J = 5.8$  Hz, 2H), 2.98 (t,  $J = 5.7$  Hz, 2H).

**8a: Reaction condition:** DC: 3.2 V; 24 h, 17 mg, 25% yield, 84 D%. The crude product was purified using flash silica-gel column chromatography to afford **8a** as a white solid.  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.21 – 7.10 (m, 4H), 6.85 – 6.78 (m, 1H), 6.57 – 6.54 (m, 2H), 4.34 (d,  $J = 25.2$  Hz, 0.17H), 4.28 – 4.25 (m, 2H), 4.24 – 4.21 (m, 2H), 3.46 (t,  $J = 5.9$  Hz, 2H), 2.98 (t,  $J = 5.9$  Hz, 2H). Deuterium incorporation of **8a**: 79% D (mass).

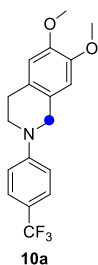
### 6,7-dimethoxy-2-(4-methoxyphenyl)-1,2,3,4-tetrahydroisoquinoline (9, 9a)



**9:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.00 (s, 2H), 6.88 (d,  $J = 9.0$  Hz, 2H), 6.64 (d,  $J = 7.1$  Hz, 2H), 4.24 (s, 2H), 3.88 (d,  $J = 3.0$  Hz, 6H), 3.79 (s, 3H), 3.45 (t,  $J = 5.2$  Hz, 2H), 2.91 (s, 2H).

**9a: Reaction condition:** DC: 3.2 V; 24 h, 45 mg, 60% yield, 93 D%. The crude product was purified using flash silica-gel column chromatography to afford **9a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.00 – 6.98 (m, 2H), 6.89 – 6.86 (m, 2H), 6.64 (d,  $J = 4.9$  Hz, 2H), 4.21 (s, 0.14H), 3.87 (d,  $J = 3.9$  Hz, 6H), 3.79 (s, 3H), 3.44 (t,  $J = 5.9$  Hz, 2H), 2.90 (t,  $J = 5.8$  Hz, 2H). Deuterium incorporation of **9a**: 90% D (mass).

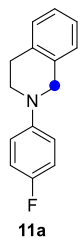
### 6,7-dimethoxy-2-(4-(trifluoromethyl)phenyl)-1,2,3,4-tetrahydroisoquinoline (10, 10a)



**10:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.50 (d,  $J = 8.7$  Hz, 2H), 6.95 (d,  $J = 8.7$  Hz, 2H), 6.68 (d,  $J = 5.1$  Hz, 2H), 4.42 (s, 2H), 3.89 (d,  $J = 3.2$  Hz, 6H), 3.63 (t,  $J = 5.8$  Hz, 2H), 2.91 (t,  $J = 5.7$  Hz, 2H).

**10a: Reaction condition:** DC: 3.5 V; 15 h, 23 mg, 27% yield, 65 D%. The crude product was purified using flash silica-gel column chromatography to afford **10a** as a white solid.  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.51

(d,  $J = 8.8$  Hz, 2H), 6.95 (d,  $J = 8.7$  Hz, 2H), 6.68 (d,  $J = 3.0$  Hz, 2H), 4.41 (d,  $J = 9.8$  Hz, 0.68H), 3.89 (d,  $J = 2.2$  Hz, 6H), 3.63 (t,  $J = 5.8$  Hz, 2H), 2.91 (t,  $J = 5.8$  Hz, 2H).  $^{19}\text{F NMR}$  (471 MHz,  $\text{CDCl}_3$ )  $\delta$  -61.11. Deuterium incorporation of **10a**: 60% D (mass).



### 2-(4-fluorophenyl)-1,2,3,4-tetrahydroisoquinoline (11, 11a)

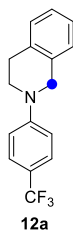
**11:**  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.21 – 7.14 (m, 4H), 7.03 – 6.95 (m, 4H), 4.35 (s, 2H), 3.51 (t,  $J = 5.9$  Hz, 2H), 3.01 (t,  $J = 5.8$  Hz, 2H).

**11a: DC:** 3.3 V; 24 h, 23 mg, 40% yield, 82 D%. The crude product was purified using flash silica-gel column

chromatography to afford **11a** as white solid  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.21 – 7.14 (m, 4H), 7.02 – 6.95 (m, 4H), 4.34 (d,  $J = 10.1$  Hz, 0.36H), 3.51 (t,  $J = 5.9$  Hz, 2H), 3.00 (t,  $J = 5.8$  Hz, 2H).  $^{19}\text{F NMR}$  (471 MHz,  $\text{CDCl}_3$ )  $\delta$  -125.70.

Deuterium incorporation of **11a**: 80% D (mass).

### 2-(4-(trifluoromethyl)phenyl)-1,2,3,4-tetrahydroisoquinoline (12, 12a)

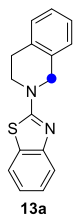


**12:**  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.51 (d,  $J = 8.8$  Hz, 2H), 7.24 – 7.18 (m, 4H), 6.96 (d,  $J = 8.7$  Hz, 2H), 4.50 (s, 2H), 3.65 (t,  $J = 5.9$  Hz, 2H), 3.01 (t,  $J = 5.8$  Hz, 2H).

**12a: Reaction condition:** DC: 3.5 V; 15 h, 7 mg, 10% yield, 70 D%. The crude product was purified using flash silica-gel column chromatography to afford **12a** as a white solid.  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.51 (d,  $J = 8.8$

Hz, 2H), 7.24 – 7.18 (m, 4H), 6.95 (d,  $J = 8.6$  Hz, 2H), 4.48 (d,  $J = 9.8$  Hz, 0.63H), 3.64 (t,  $J = 6.1$  Hz, 2H), 3.01 (t,  $J = 5.9$  Hz, 2H).  $^{19}\text{F NMR}$  (376 MHz,  $\text{CDCl}_3$ )  $\delta$  -61.12 (s). Deuterium incorporation of **12a**: 80% D (mass).

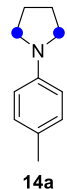
### 2-(3,4-dihydroisoquinolin-2(1H)-yl)benzo[d]thiazole (13,13a)



**13:**  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.63 (t,  $J = 8$  Hz, 2H), 7.32 (t,  $J = 7.7$  Hz, 1H), 7.25 – 7.20 (m, 4H), 7.09 (t,  $J = 7.6$  Hz, 1H), 4.84 (s, 2H), 3.90 (t,  $J = 5.9$  Hz, 2H), 3.05 (t,  $J = 5.8$  Hz, 2H).

**13a:** DC: 3.9 V; 12 h, 13 mg, 20% yield, 35 D%. The crude product was purified using flash silica-gel column chromatography to afford **6a** as a white solid.  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.63 (dd,  $J = 7.8, 5.0$  Hz, 2H), 7.32 (t,  $J = 8.3$  Hz, 1H), 7.26 – 7.18 (m, 4H), 7.10 (t,  $J = 8.1$  Hz, 1H), 4.83 (d,  $J = 10.5$  Hz, 1.37H), 3.90 (t,  $J = 5.9$  Hz, 2H), 3.05 (t,  $J = 5.9$  Hz, 2H). Deuterium incorporation of **13a**: 29% (mass).

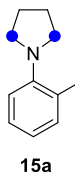
### 1-(p-tolyl)pyrrolidine (14, 14a)



**14:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.09 (d,  $J = 8.1$  Hz, 2H), 6.56 (d,  $J = 8.5$  Hz, 2H), 3.33 – 3.28 (m, 4H), 2.31 (s, 3H), 2.06 – 2.02 (m, 4H).

**14a:** Reaction condition: AC: 0.5 Hz, 3.3 V (peak amplitude); 48 h, 14 mg, 34% yield, 77% D. The crude product was purified using flash silica-gel column chromatography (Ethyl acetate: Hexane = 10:90 v/v) to afford **14a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.08 – 7.03 (m, 2H), 6.52 (d,  $J = 8.0$  Hz, 2H), 3.25 (q,  $J = 6.6$  Hz, 0.92H), 2.27 (s, 3H), 1.99 (d,  $J = 4.2$  Hz, 4H). Deuterium incorporation of **14a**: 50% (mass).

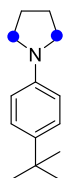
### 1-(o-tolyl)pyrrolidine (15, 15a)



**15:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.13 (t,  $J = 7.0$  Hz, 2H), 6.91 (d,  $J = 8.0$  Hz, 1H), 6.85 (t,  $J = 7.4$  Hz, 1H), 3.21 (d,  $J = 6.3$  Hz, 4H), 2.35 (s, 3H), 1.95 (qd,  $J = 7.1, 4.6, 3.1$  Hz, 4H).

**15a:** Reaction condition: AC: 0.5 Hz, 3.1 V (peak amplitude); 48 h, 16 mg, 38% yield, 36% D. The crude product was purified using flash silica-gel column chromatography (Ethyl acetate: Hexane = 10:90 v/v) to afford **15a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.14 (t,  $J = 7.0$  Hz, 2H), 6.93 – 6.90 (m, 1H), 6.86 (t,  $J = 7.3$  Hz, 1H), 3.24 – 3.17 (m, 2.51H), 2.36 (s, 3H), 1.97 – 1.94 (m, 4H). Deuterium incorporation of **15a**: 36% (mass).

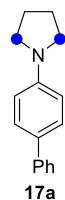
### 1-(4-(tert-butyl)phenyl)pyrrolidine (16, 16a)



**16:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.31 (d,  $J = 8.8$  Hz, 2H), 6.58 (d,  $J = 8.7$  Hz, 2H), 3.33 – 3.29 (m, 4H), 2.04 – 2.00 (m, 4H), 1.34 (s, 9H).

**16a: Reaction condition:** AC: 0.5 Hz, 3.3 V (peak amplitude); 48 h, 20mg, 38% yield, 75% D. The crude product **16a** was purified using flash silica-gel column chromatography (Ethyl acetate: Hexane = 10:90 v/v) to afford **16a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.27 – 7.24 (m, 2H), 6.53 (d,  $J = 8.2$  Hz, 2H), 3.25 (q,  $J = 6.4$  Hz, 1.02H), 1.97 (d,  $J = 8.2$  Hz, 4H), 1.29 (s, 9H). Deuterium incorporation of **16a**: 70% (mass).

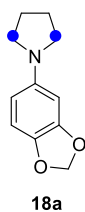
### 1-([1,1'-biphenyl]-4-yl)pyrrolidine (17, 17a)



**17:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.59 (d,  $J = 7.8$  Hz, 2H), 7.54 (d,  $J = 8.9$  Hz, 2H), 7.42 (t,  $J = 7.8$  Hz, 2H), 7.27 (d,  $J = 14.7$  Hz, 1H), 6.67 (d,  $J = 8.8$  Hz, 2H), 3.40 – 3.32 (m, 4H), 2.09 – 2.01 (m, 4H).

**17a: Reaction condition:** AC: 0.5 Hz, 3.3 V (peak amplitude); 48 h, 13 mg, 23% yield, 52% D. The crude product was purified using flash silica-gel column chromatography (Ethyl acetate: Hexane = 10:90 v/v) to afford **17a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.60 – 7.55 (m, 2H), 7.55 – 7.50 (m, 2H), 7.41 (t,  $J = 7.8$  Hz, 2H), 7.28 – 7.24 (m, 1H), 6.66 (d,  $J = 8.1$  Hz, 2H), 3.39 – 3.31 (m, 1.91H), 2.08 – 1.98 (m, 4H). Deuterium incorporation of **17a**: 46% (mass).

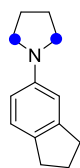
### 1-(benzo[d][1,3]dioxol-5-yl)pyrrolidine (18, 18a)



**18:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  6.74 (d,  $J = 8.4$  Hz, 1H), 6.24 (d,  $J = 2.4$  Hz, 1H), 5.98 (dd,  $J = 8.4, 2.4$  Hz, 1H), 5.86 (s, 2H), 3.27 – 3.17 (m, 4H), 2.02 – 1.96 (m, 4H).

**18a: Reaction condition:** AC: 0.5 Hz, 3.1 V (peak amplitude); 48 h, 14 mg, 28% yield, 75% D. The crude product was purified using flash silica-gel column chromatography (Ethyl acetate: Hexane = 10:90 v/v) to afford **18a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  6.73 (d,  $J = 8.4$  Hz, 1H), 6.24 (s, 1H), 5.98 (d,  $J = 8.4$  Hz, 1H), 5.86 (s, 2H), 3.21 (q,  $J = 6.4$  Hz, 0.98H), 1.99 (d,  $J = 4.1$  Hz, 4H). Deuterium incorporation of **18a**: 48% (mass).

### 1-(2,3-dihydro-1H-inden-5-yl)pyrrolidine (19, 19a)

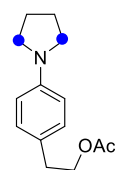


19a

**19:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.12 (d,  $J = 8.1$  Hz, 1H), 6.53 (s, 1H), 6.43 (d,  $J = 8.2$  Hz, 1H), 3.33 – 3.27 (m, 4H), 2.88 (dt,  $J = 19.3, 7.3$  Hz, 4H), 2.09 (q,  $J = 7.3$  Hz, 2H), 2.04 – 1.99 (m, 4H).

**19a: Reaction condition:** AC: 0.5 Hz, 3.3 V (peak amplitude); 48 h, 20 mg, 42% yield, 24% D. The crude product was purified using flash silica-gel column chromatography (Ethyl acetate: Hexane = 10:90 v/v) to afford **19a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.10 (d,  $J = 8.1$  Hz, 1H), 6.52 (s, 1H), 6.46 – 6.38 (m, 1H), 3.28 (q,  $J = 6.0$  Hz, 3.05H), 2.86 (dt,  $J = 28.9, 7.3$  Hz, 4H), 2.06 (p,  $J = 7.3$  Hz, 2H), 2.00 (q,  $J = 5.5, 4.0$  Hz, 4H). Deuterium incorporation of **19a**: 23% (mass).

#### 4-(pyrrolidin-1-yl)phenethyl acetate (**20**, **20a**)

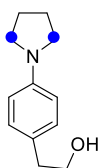


20a

**20:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.09 (d,  $J = 8.5$  Hz, 2H), 6.54 (d,  $J = 8.5$  Hz, 2H), 4.24 (t,  $J = 7.2$  Hz, 2H), 3.31 – 3.26 (m, 4H), 2.85 (t,  $J = 7.2$  Hz, 2H), 2.06 (s, 3H), 2.03 – 1.98 (m, 4H).

**20a: Reaction condition:** AC: 0.5 Hz, 3.3 V (peak amplitude); 48 h, 21 mg, 36% yield, 89% D. The crude product was purified using flash silica-gel column chromatography (Ethyl acetate: Hexane = 10:90 v/v) to afford **20a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.09 (d,  $J = 8.1$  Hz, 2H), 6.52 (d,  $J = 7.9$  Hz, 2H), 4.23 (t,  $J = 7.3$  Hz, 2H), 3.25 (s, 0.44H), 2.84 (t,  $J = 7.2$  Hz, 2H), 2.05 (s, 3H), 1.99 (s, 4H). Deuterium incorporation of **20a**: 75% (mass).

#### 2-(4-(pyrrolidin-1-yl)phenyl)ethan-1-ol (**21**, **21a**)

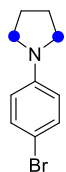


21a

**21:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.10 (d,  $J = 8.0$  Hz, 2H), 6.55 (s, 2H), 3.81 (q,  $J = 6.3$  Hz, 2H), 3.27 (d,  $J = 6.3$  Hz, 4H), 2.78 (t,  $J = 6.5$  Hz, 2H), 2.00 (d,  $J = 6.1$  Hz, 4H).

**21a: Reaction condition:** AC: 0.5 Hz, 3.3 V (peak amplitude); 48 h, 20 mg, 40% yield, 54% D. The crude product was purified using flash silica-gel column chromatography (Ethyl acetate: Hexane = 10:90 v/v) to afford **21a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.13 – 7.06 (m, 2H), 6.55 (d,  $J = 8.1$  Hz, 2H), 3.81 (t,  $J = 6.6$  Hz, 2H), 3.28 (dd,  $J = 12.4, 6.2$  Hz, 1.82H), 2.78 (t,  $J = 6.6$  Hz, 2H), 2.03 – 1.97 (m, 4H). Deuterium incorporation of **21a**: 41% (mass).

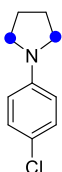
### 1-(4-bromophenyl)pyrrolidine (**22**, **22a**)



**22:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.29 (d,  $J = 8.9$  Hz, 2H), 6.43 (d,  $J = 8.9$  Hz, 2H), 3.28 – 3.22 (m, 4H), 2.05 – 1.97 (m, 4H).

**22a:** **Reaction condition:** AC: 0.5 Hz, 3.5 V (peak amplitude); 48 h, 23 mg, 40% yield, 23% D. The crude product was purified using flash silica-gel column chromatography (Ethyl acetate: Hexane = 10:90 v/v) to afford **22a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.30 – 7.27 (m, 2H), 6.44 (d,  $J = 8.5$  Hz, 2H), 3.25 (td,  $J = 8.0$ , 7.3, 4.2 Hz, 3.1H), 2.04 – 1.99 (m, 4H). Deuterium incorporation of **22a**: 41% (mass).

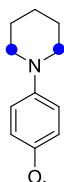
### 1-(4-chlorophenyl)pyrrolidine (**23**, **23a**)



**23:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.17 (d,  $J = 8.9$  Hz, 2H), 6.48 (d,  $J = 8.9$  Hz, 2H), 3.28 – 3.24 (m, 4H), 2.05 – 2.00 (m, 4H).

**23a:** **Reaction condition:** AC: 0.5 Hz, 3.4 V (peak amplitude); 48 h, 19 mg, 42% yield, 36% D. The crude product was purified using flash silica-gel column chromatography (Ethyl acetate: Hexane = 10:90 v/v) to afford **23a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.19 – 7.13 (m, 2H), 6.48 (d,  $J = 8.5$  Hz, 2H), 3.28 – 3.21 (m, 2.54H), 2.03 – 1.99 (m, 4H). Deuterium incorporation of **23a**: 32% (mass).

### 1-(4-methoxyphenyl)piperidine (**24**, **24a**)

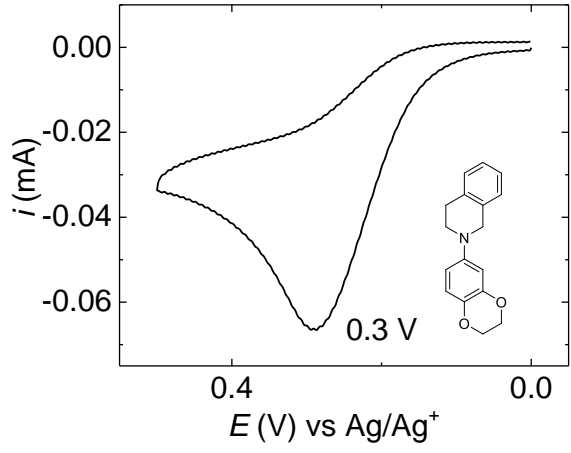
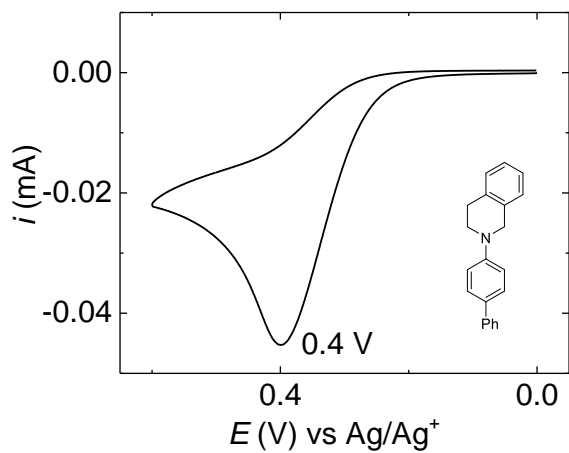
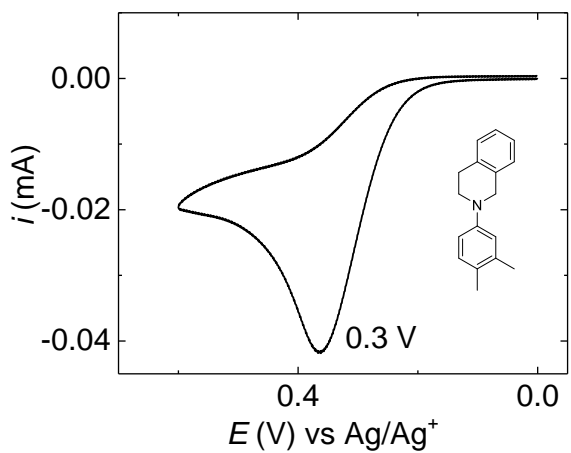
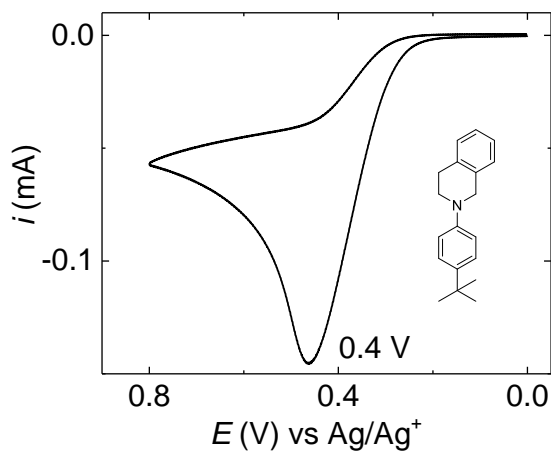
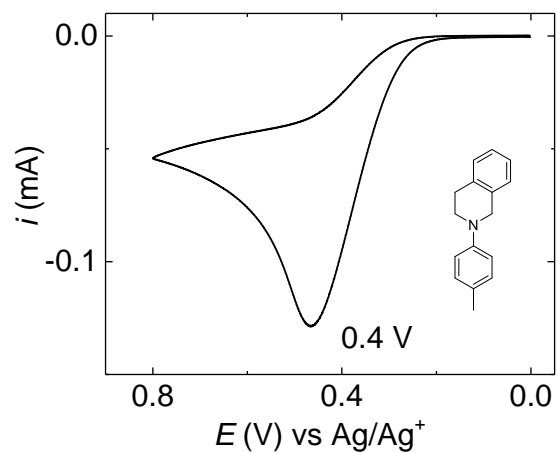
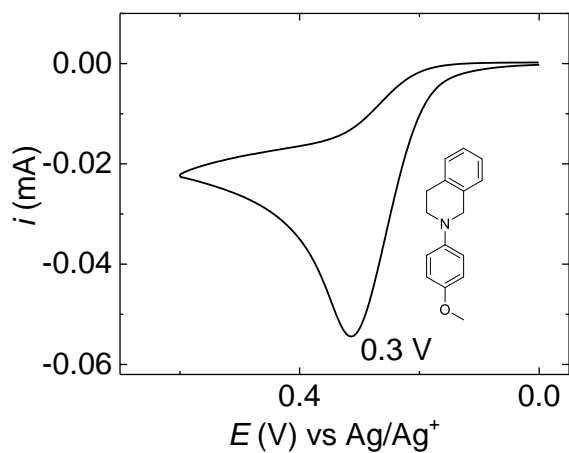


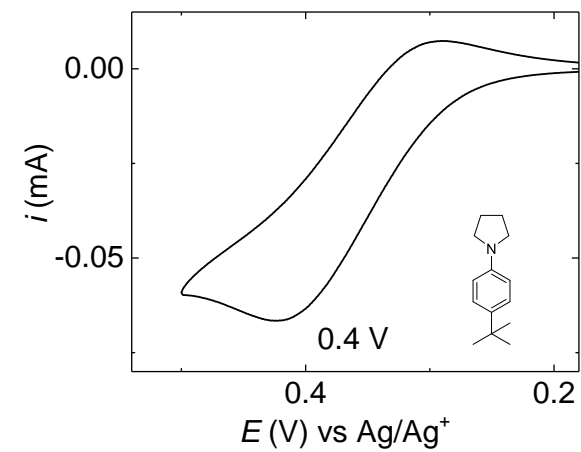
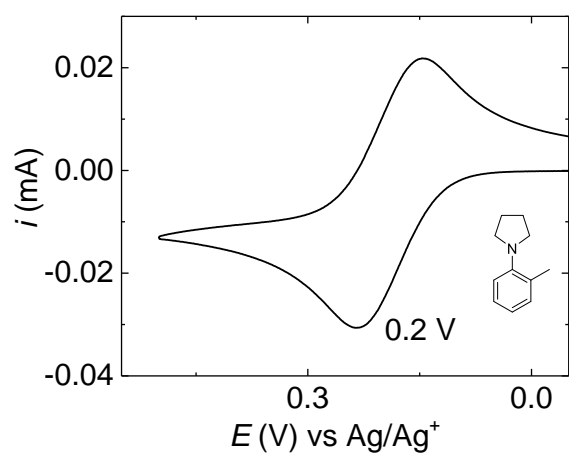
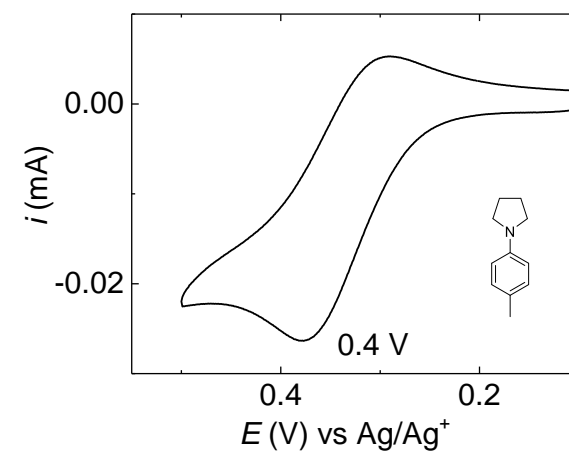
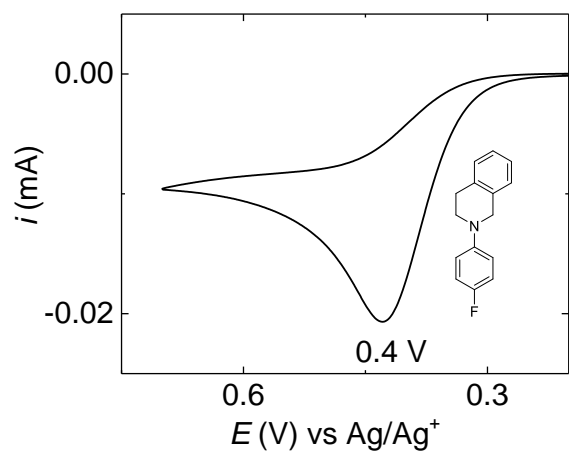
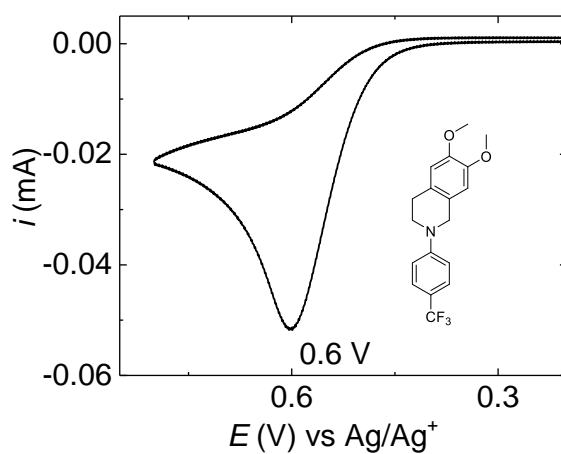
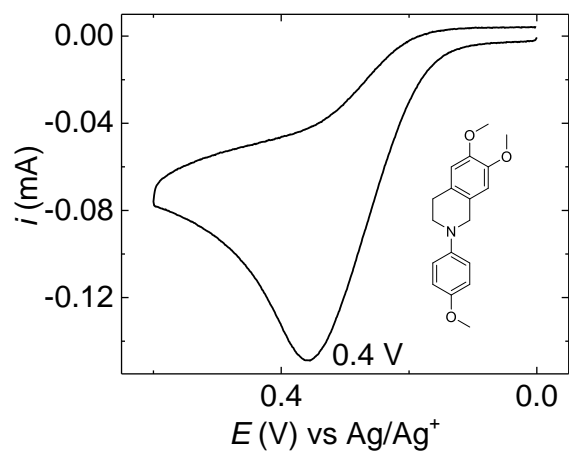
**24:**  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  6.93 (d,  $J = 9.0$  Hz, 2H), 6.86 – 6.81 (m, 2H), 3.78 (s, 3H), 3.06 – 3.01 (m, 4H), 1.74 (p,  $J = 5.8$  Hz, 4H), 1.56 (q,  $J = 6.1$  Hz, 2H).

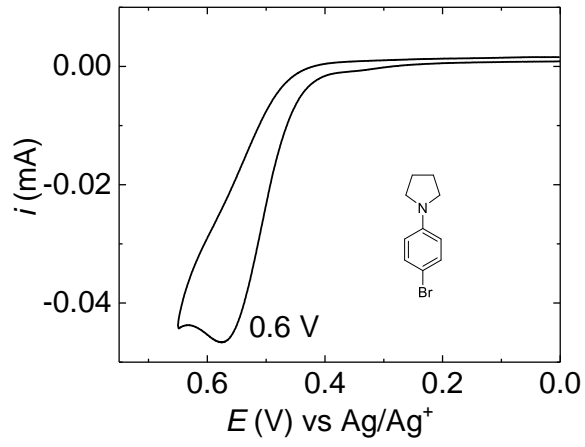
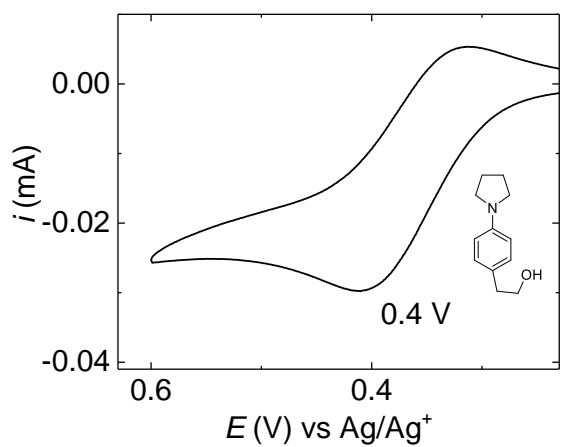
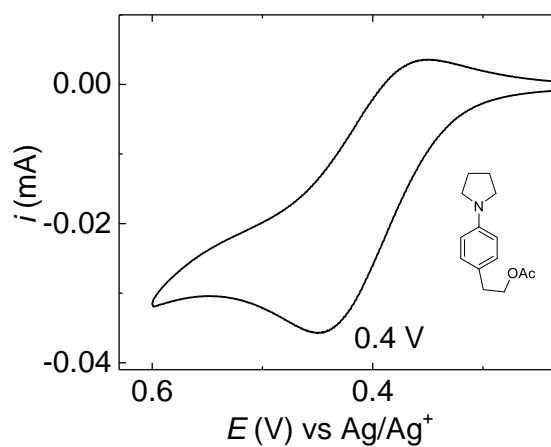
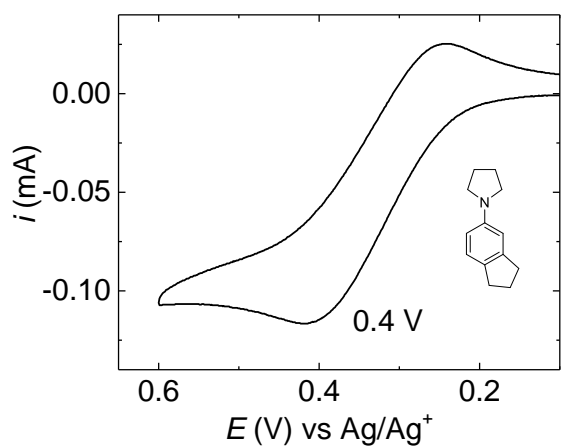
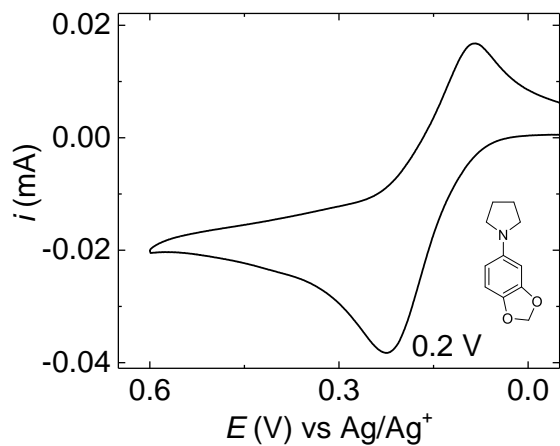
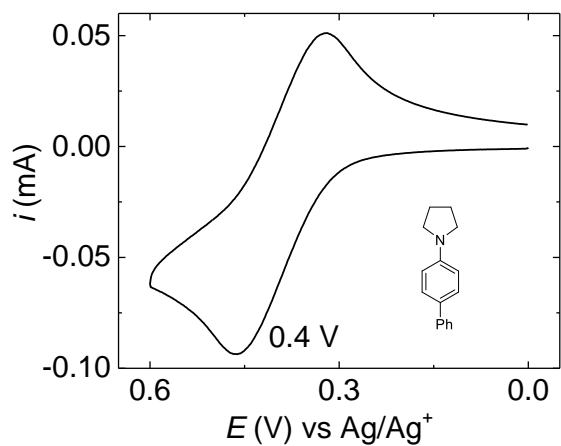
**24a:** **Reaction condition:** AC: 50 Hz, 3.4 V (peak amplitude); 48 h, 25 mg, 51% yield, 50% D. The crude product was purified using flash silica-gel column chromatography (Ethyl acetate: Hexane = 10:90 v/v) to afford **24a** as a white solid.  $^1\text{H NMR}$  (600 MHz,  $\text{CDCl}_3$ )  $\delta$  6.93 (d,  $J = 8.7$  Hz, 2H), 6.87 – 6.81 (m, 2H), 3.78 (s, 3H), 3.02 (dt,  $J = 16.9$ , 5.4 Hz, 1.99H), 1.73 (q,  $J = 5.8$  Hz, 4H), 1.56 (tdd,  $J = 9.5$ , 5.9, 3.9 Hz, 2H). Deuterium incorporation of **24a**: 45% (mass).

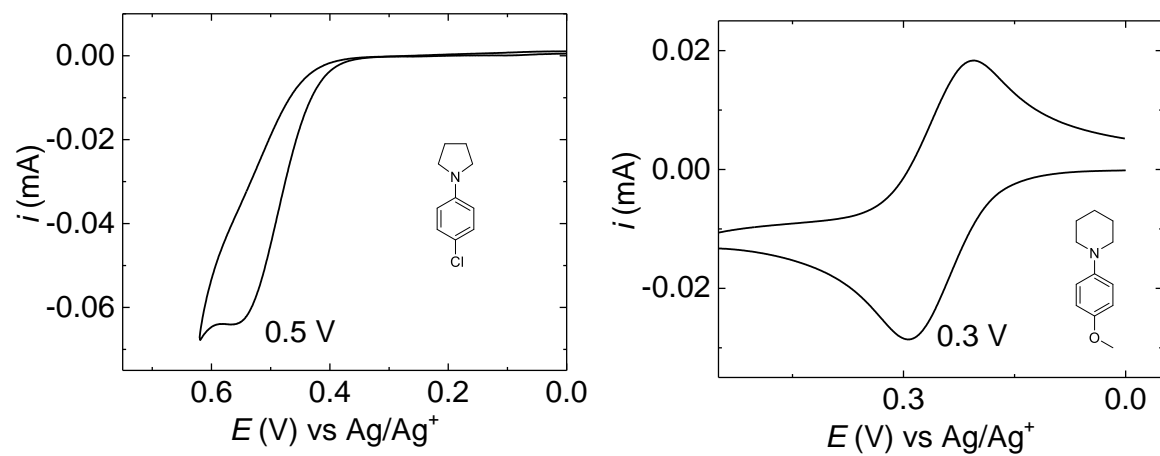


10. Oxidation potentials of tetrahydroisoquinolines (3-13), pyrrolidine (14-23), and piperidines (24)







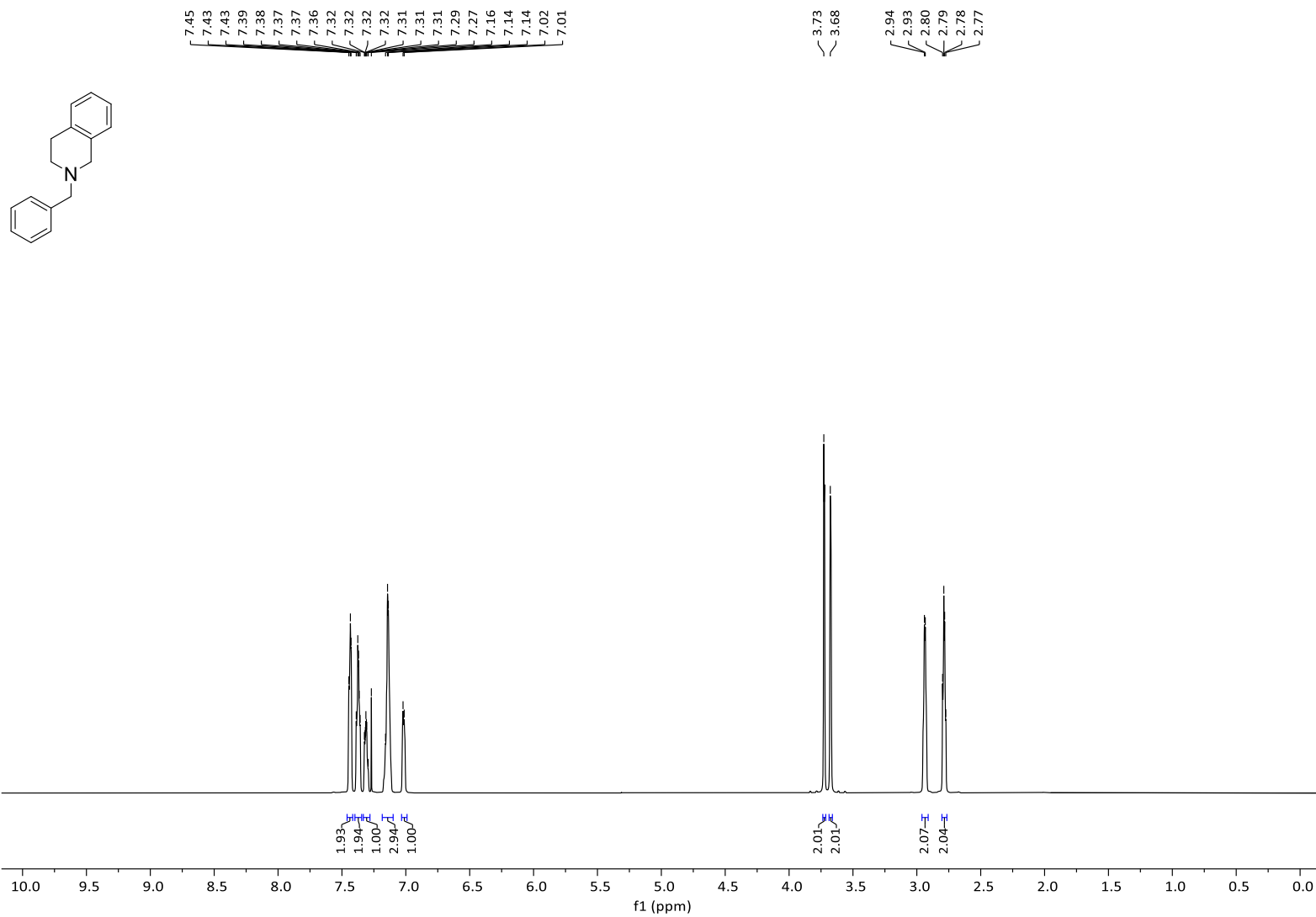


**Fig. S5.** Cyclic voltammograms of **3-24** in anhydrous DMA containing 0.1 M  $\text{LiClO}_4$ . Scan rate 0.1 V/s.

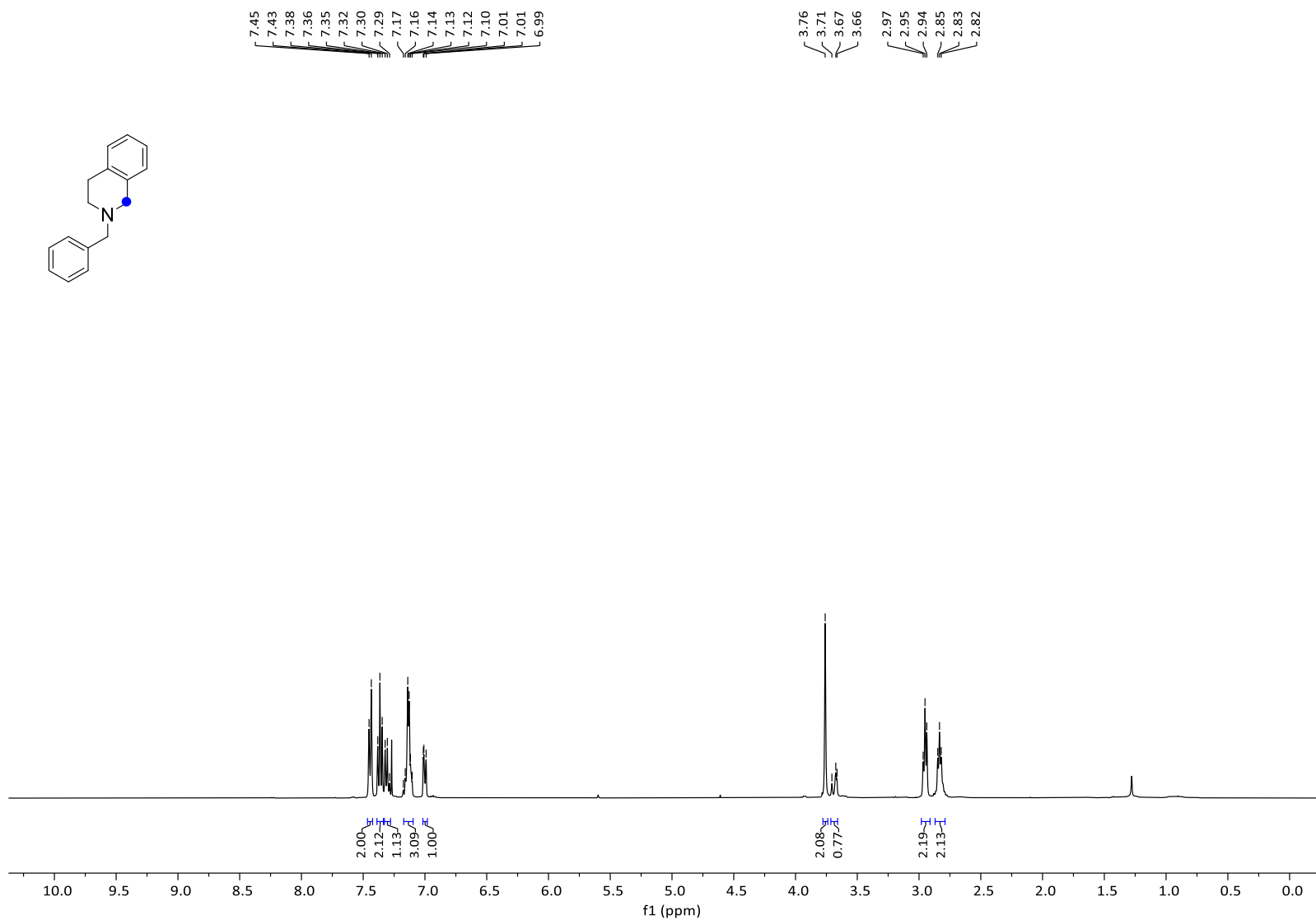
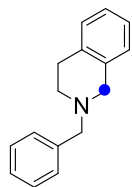
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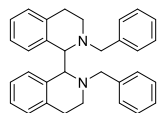
Appendix A. Copies of  $^1\text{H}$  and  $^{19}\text{F}$  NMR spectra



$^1\text{H}$  NMR spectrum of **01** (600 MHz,  $\text{CDCl}_3$ )

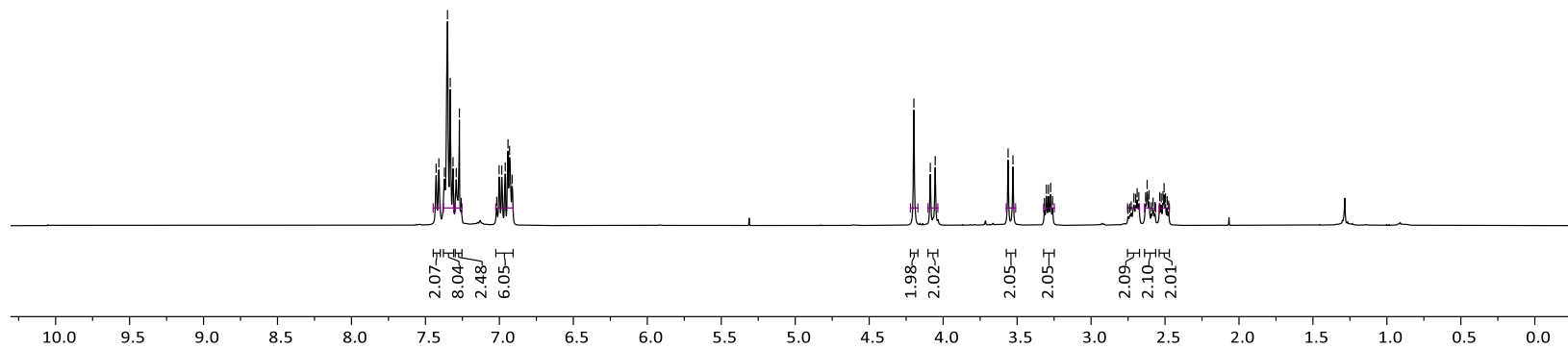


$^1\text{H}$  NMR spectrum of **1a** (600 MHz,  $\text{CDCl}_3$ )



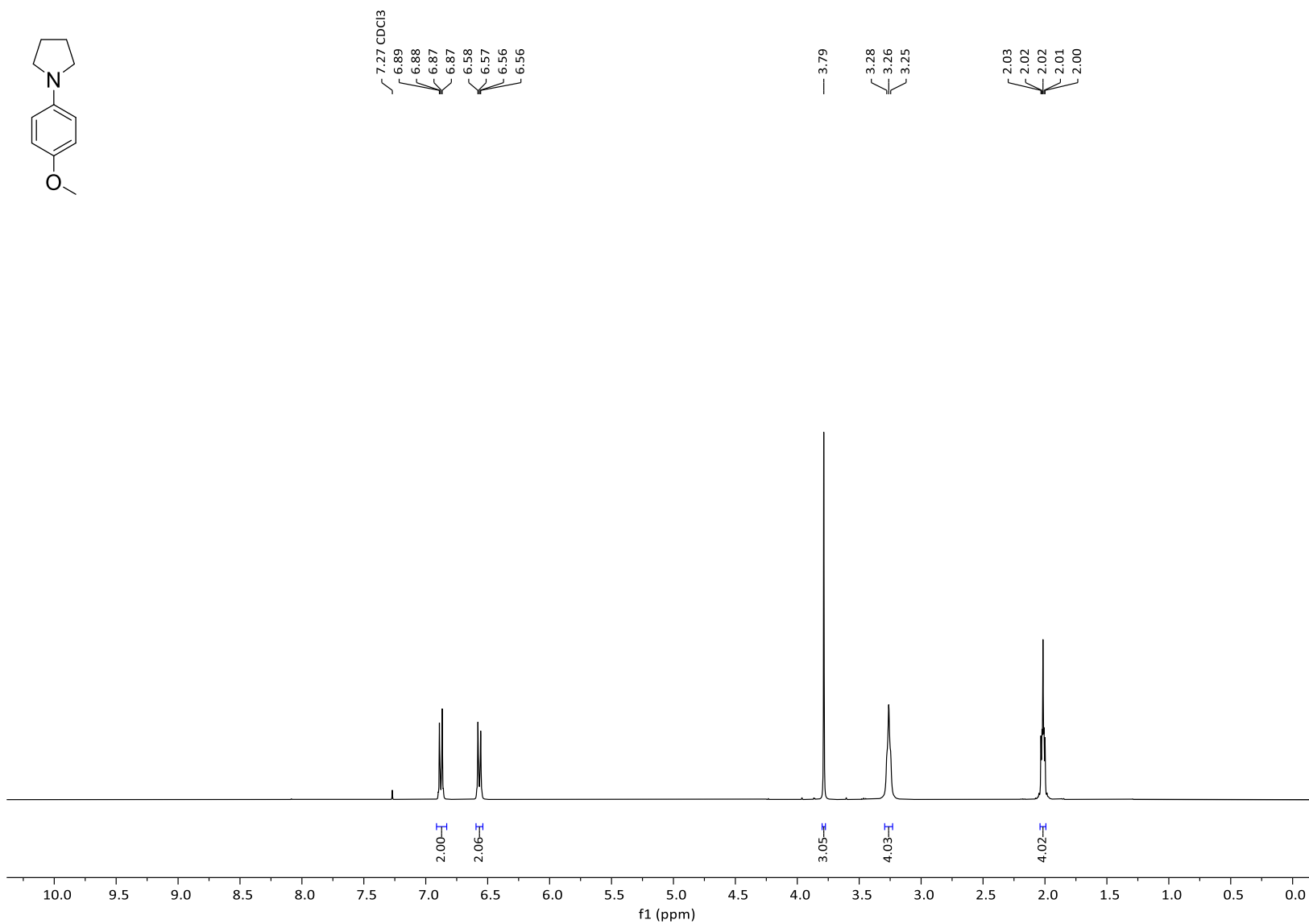
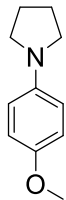
7.43  
7.41  
7.37  
7.35  
7.33  
7.31  
7.29  
7.27  
7.26  
7.02  
7.00  
6.98  
6.96  
6.94  
6.93  
6.91

4.20  
4.09  
4.05  
3.56  
3.53  
3.31  
3.30  
3.29  
3.27  
3.26  
2.75  
2.74  
2.73  
2.71  
2.70  
2.69  
2.68  
2.63  
2.62  
2.61  
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2.53  
2.51  
2.51  
2.50  
2.49  
2.48

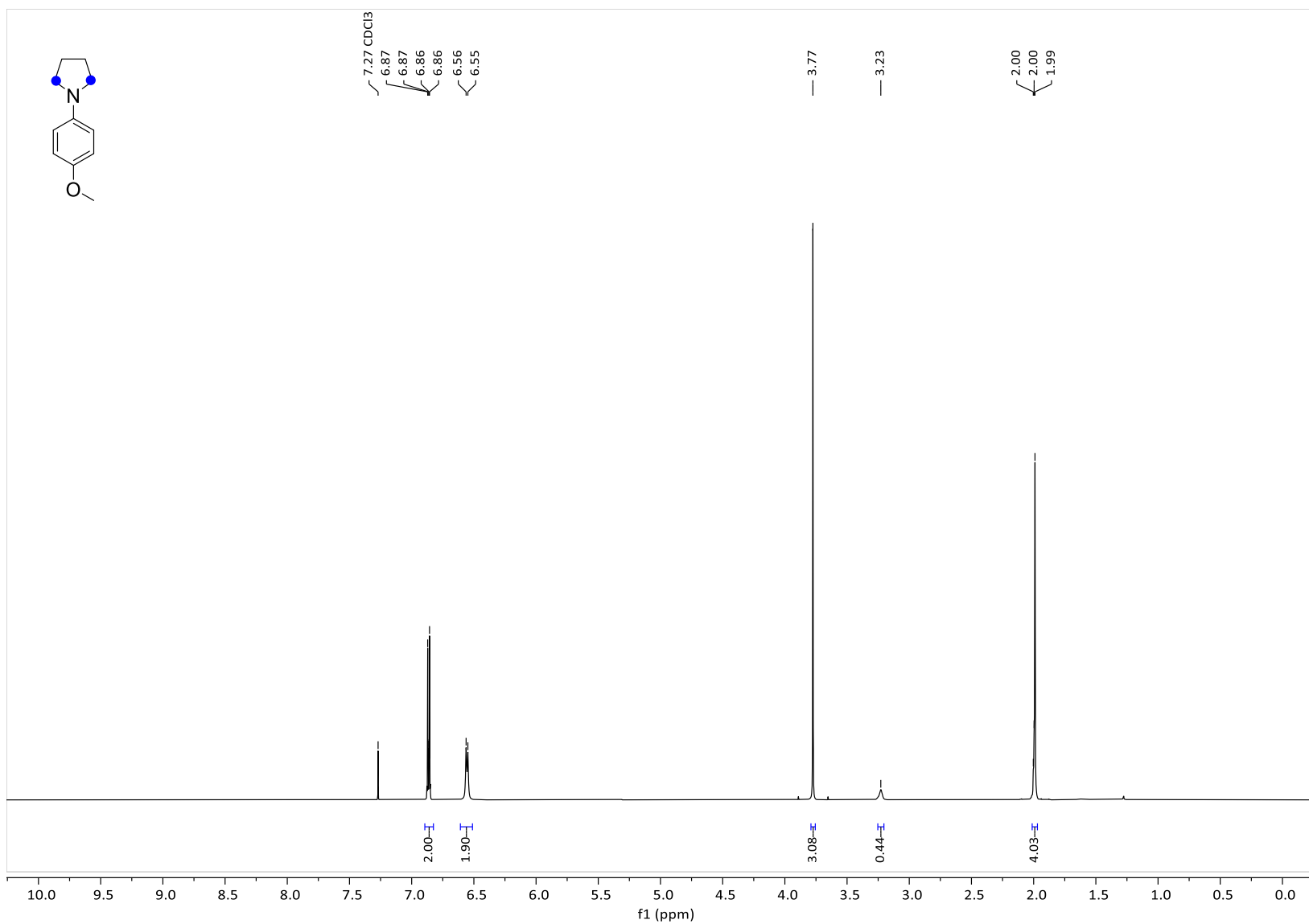


<sup>1</sup>H NMR spectrum of dimer (600 MHz, CDCl<sub>3</sub>)

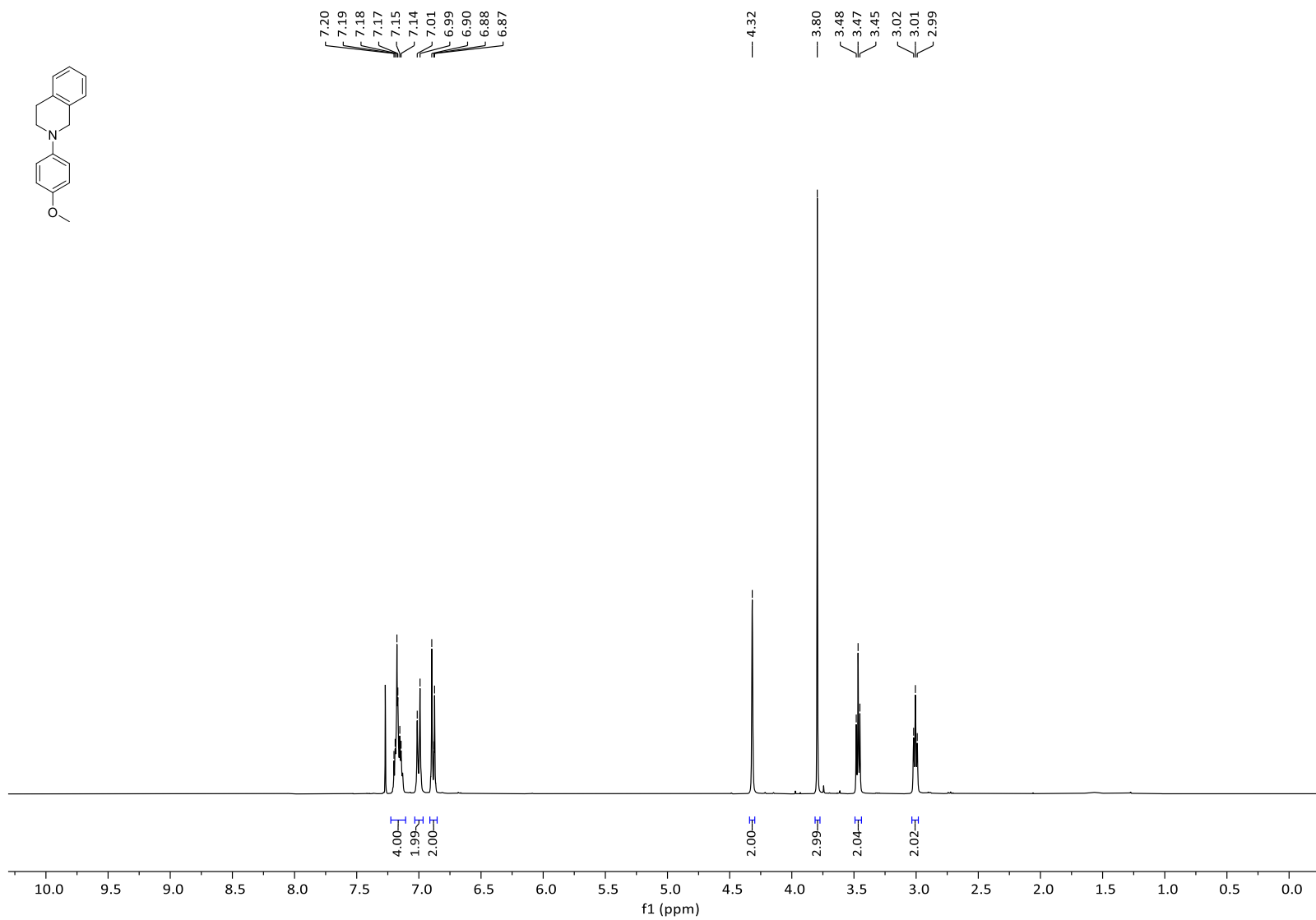
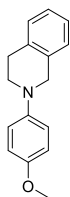




<sup>1</sup>H NMR spectrum of **02** (600 MHz, CDCl<sub>3</sub>)

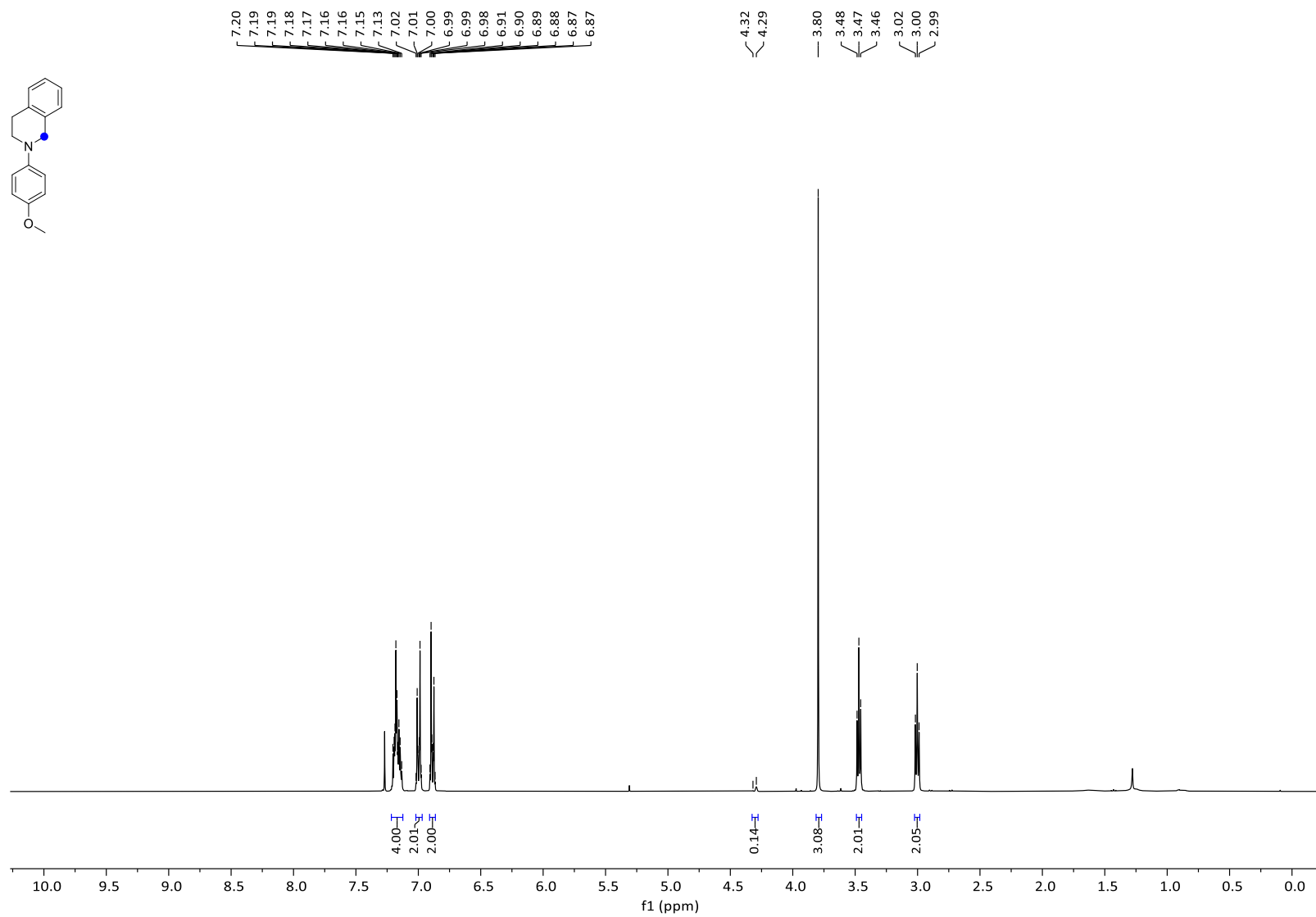
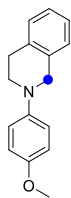


<sup>1</sup>H NMR spectrum of **2a** (600 MHz, CDCl<sub>3</sub>)

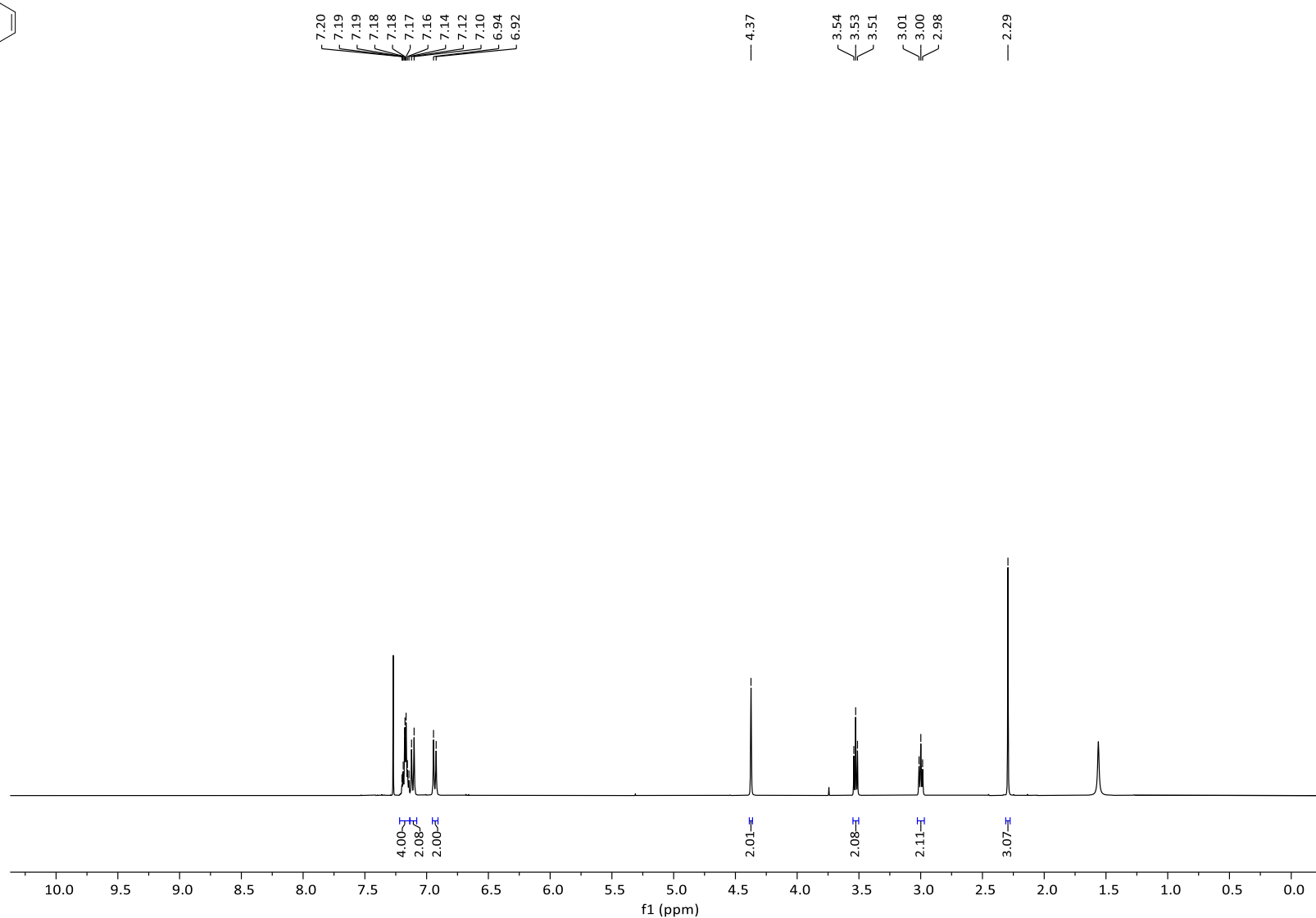
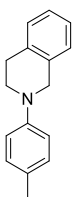


NMR spectrum of **03** (600 MHz, CDCl<sub>3</sub>)

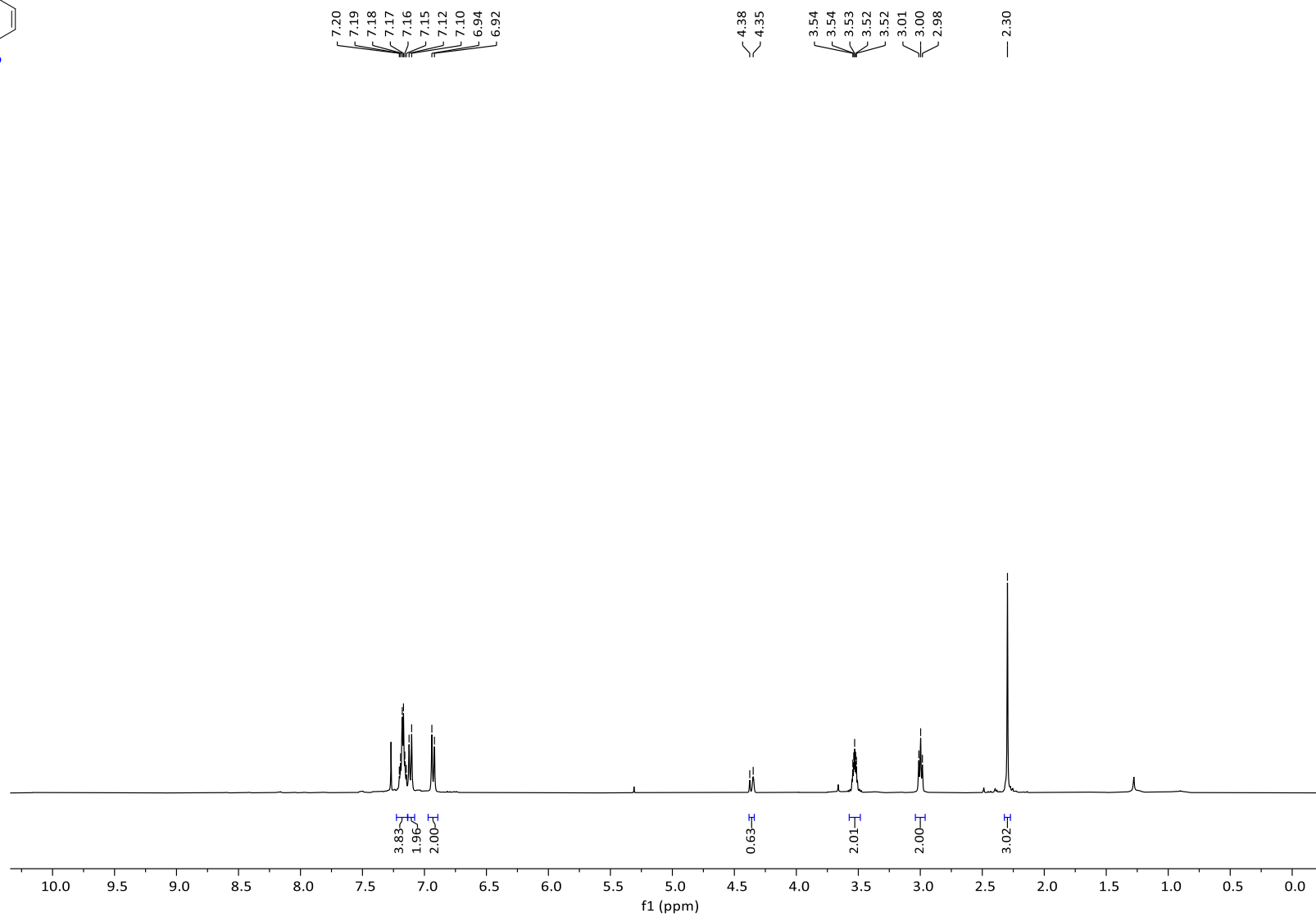
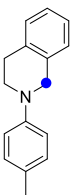
<sup>1</sup>H



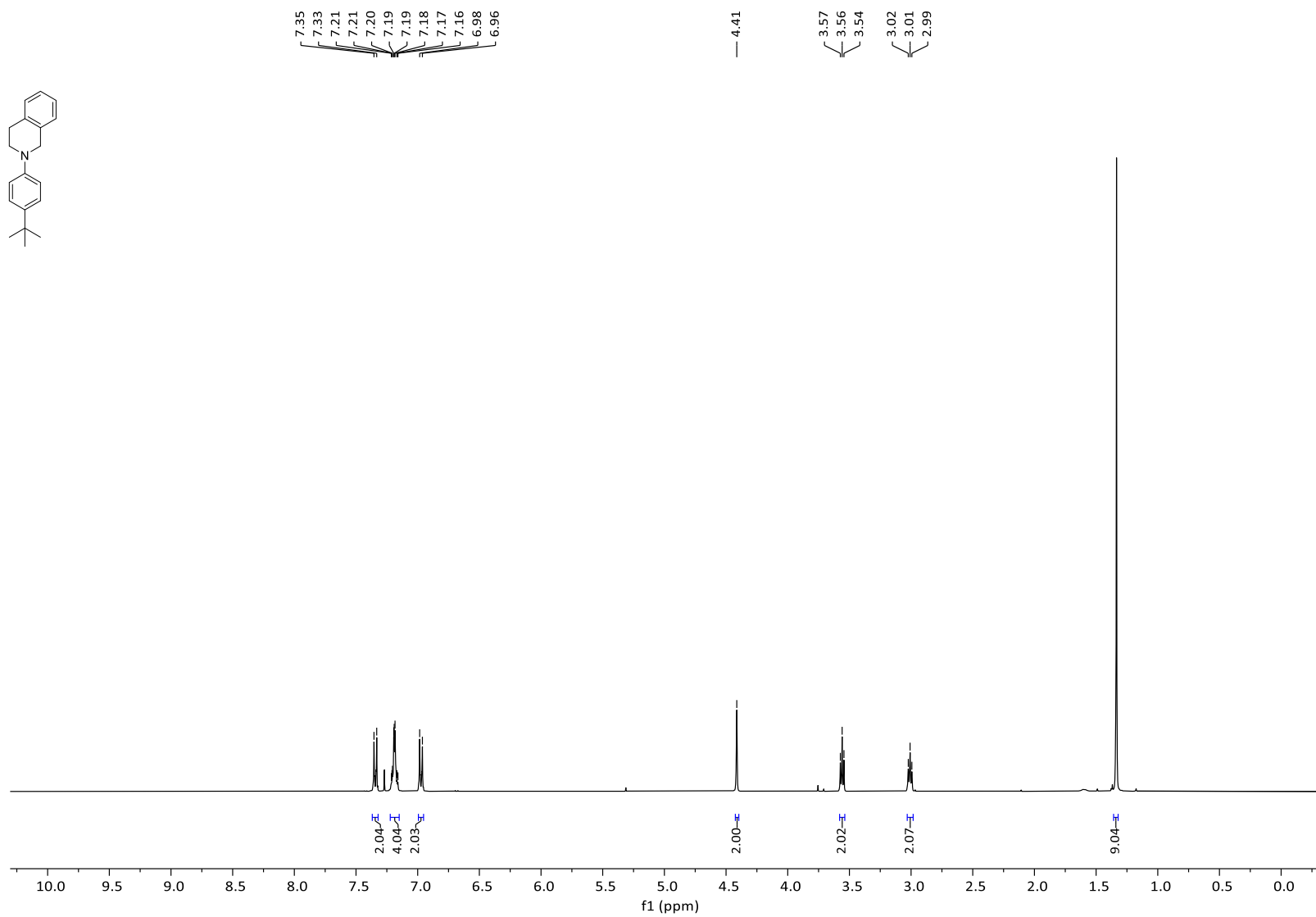
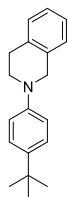
<sup>1</sup>H NMR spectrum of **3a** (600 MHz, CDCl<sub>3</sub>)



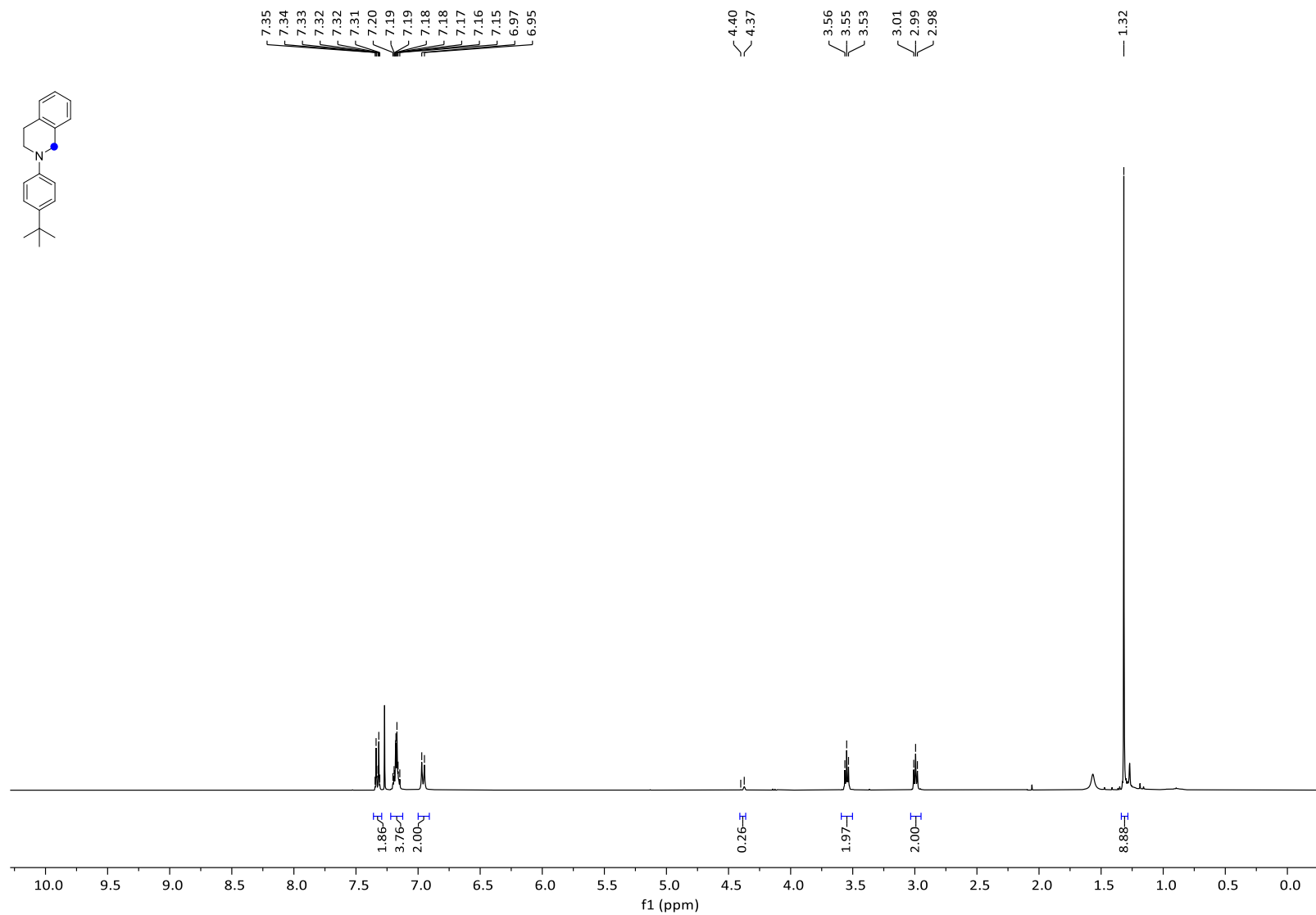
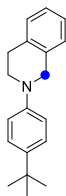
<sup>1</sup>H NMR spectrum of **04** (600 MHz, CDCl<sub>3</sub>)



<sup>1</sup>H NMR spectrum of **4a** (600 MHz, CDCl<sub>3</sub>)

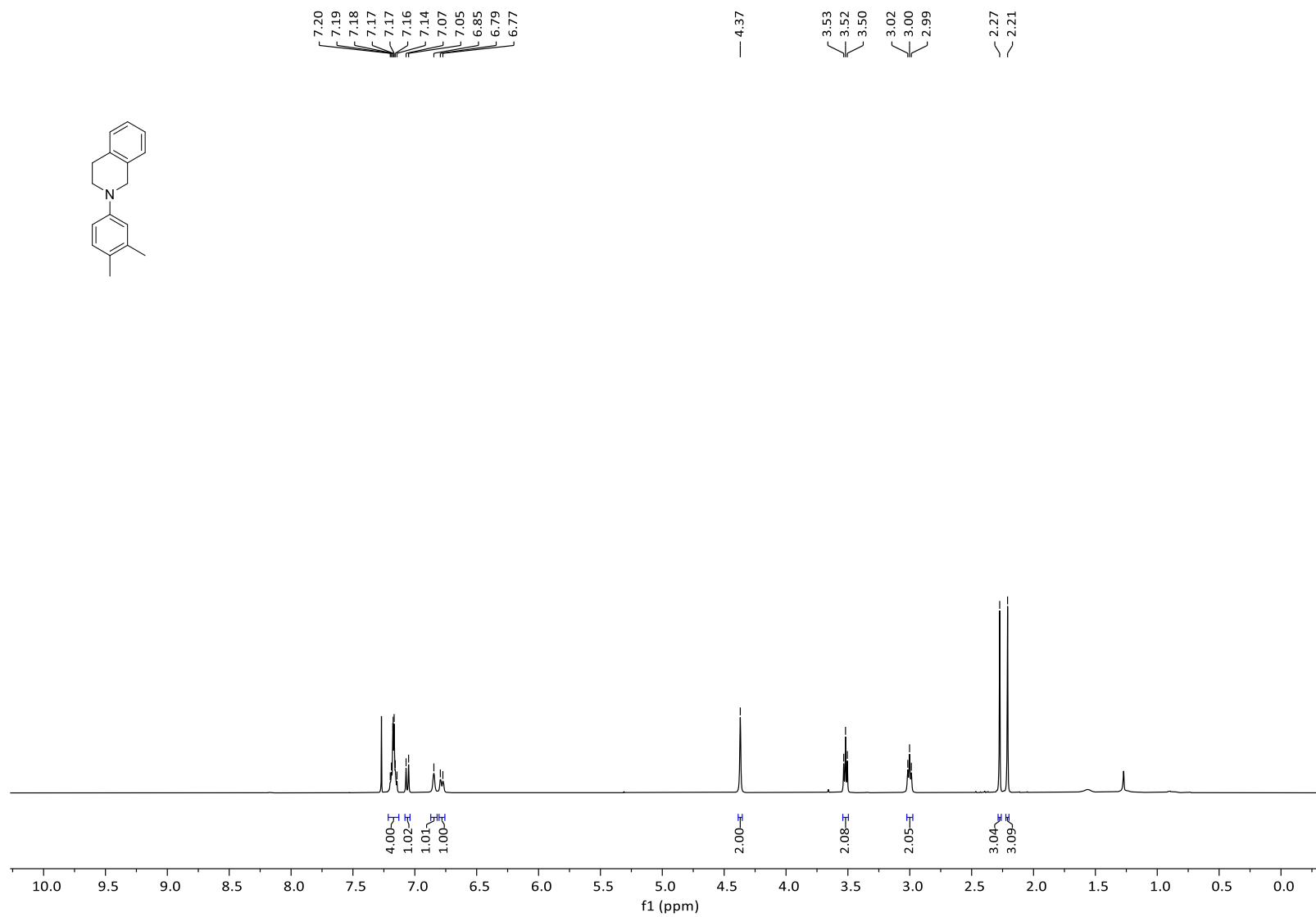
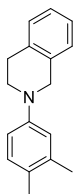


<sup>1</sup>H NMR spectrum of **05** (600 MHz, CDCl<sub>3</sub>)

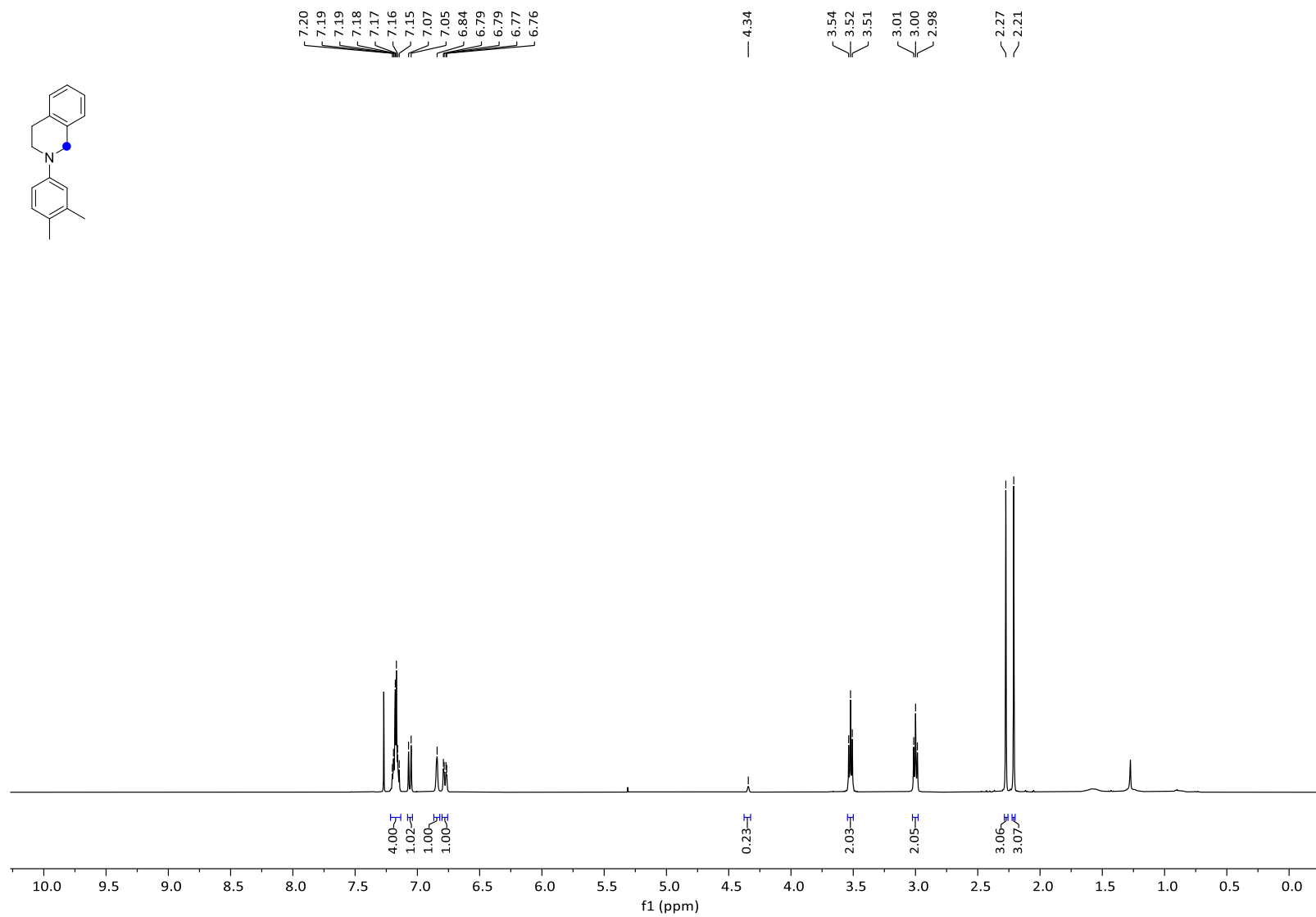
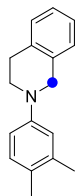


$^1\text{H}$  NMR spectrum of 5a (600 MHz,  $\text{CDCl}_3$ )

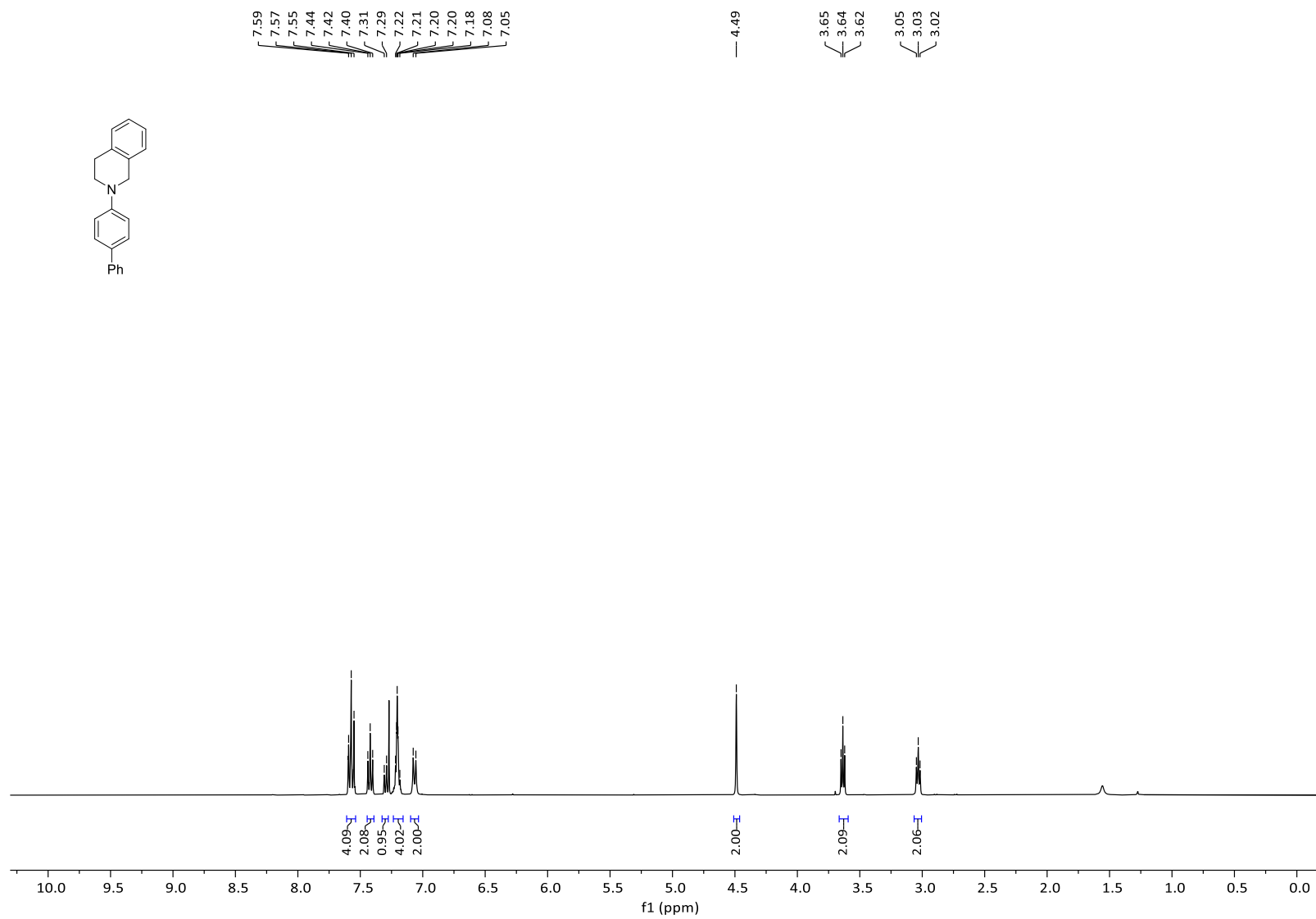
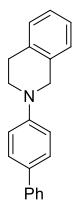




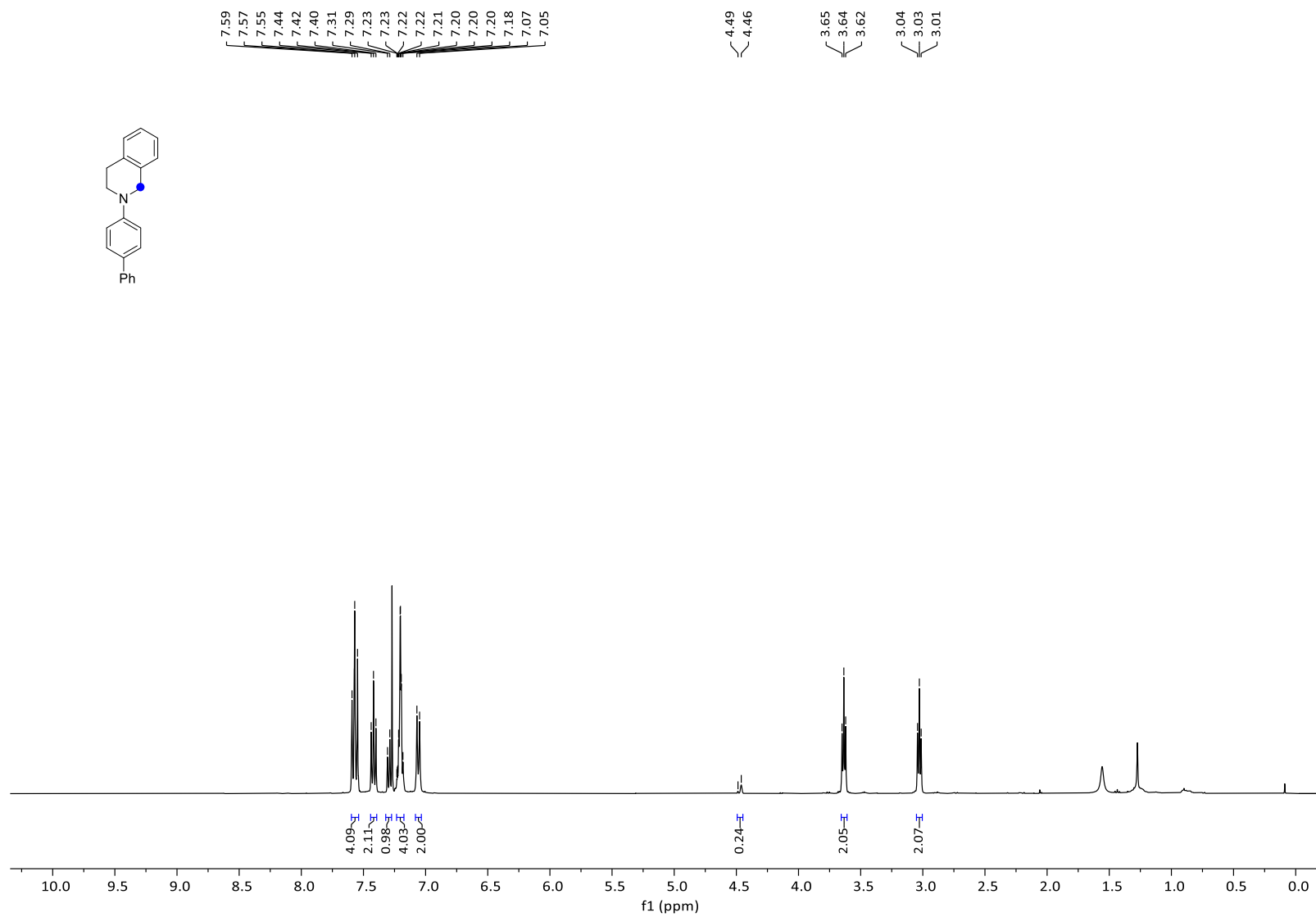
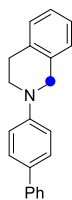
$^1\text{H}$  NMR spectrum of **06** (600 MHz,  $\text{CDCl}_3$ )



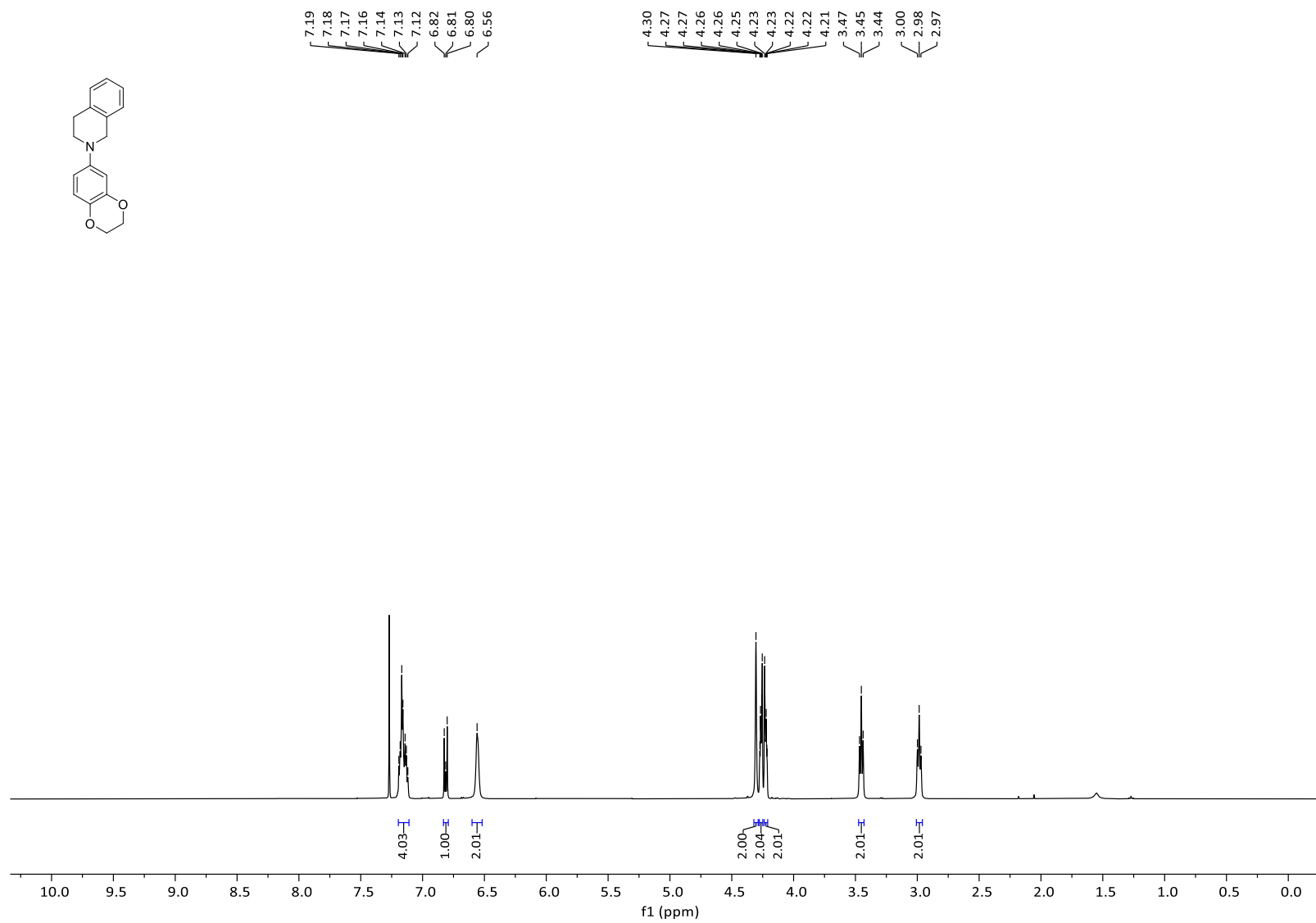
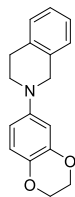
$^1\text{H}$  NMR spectrum of **6a** (600 MHz,  $\text{CDCl}_3$ )



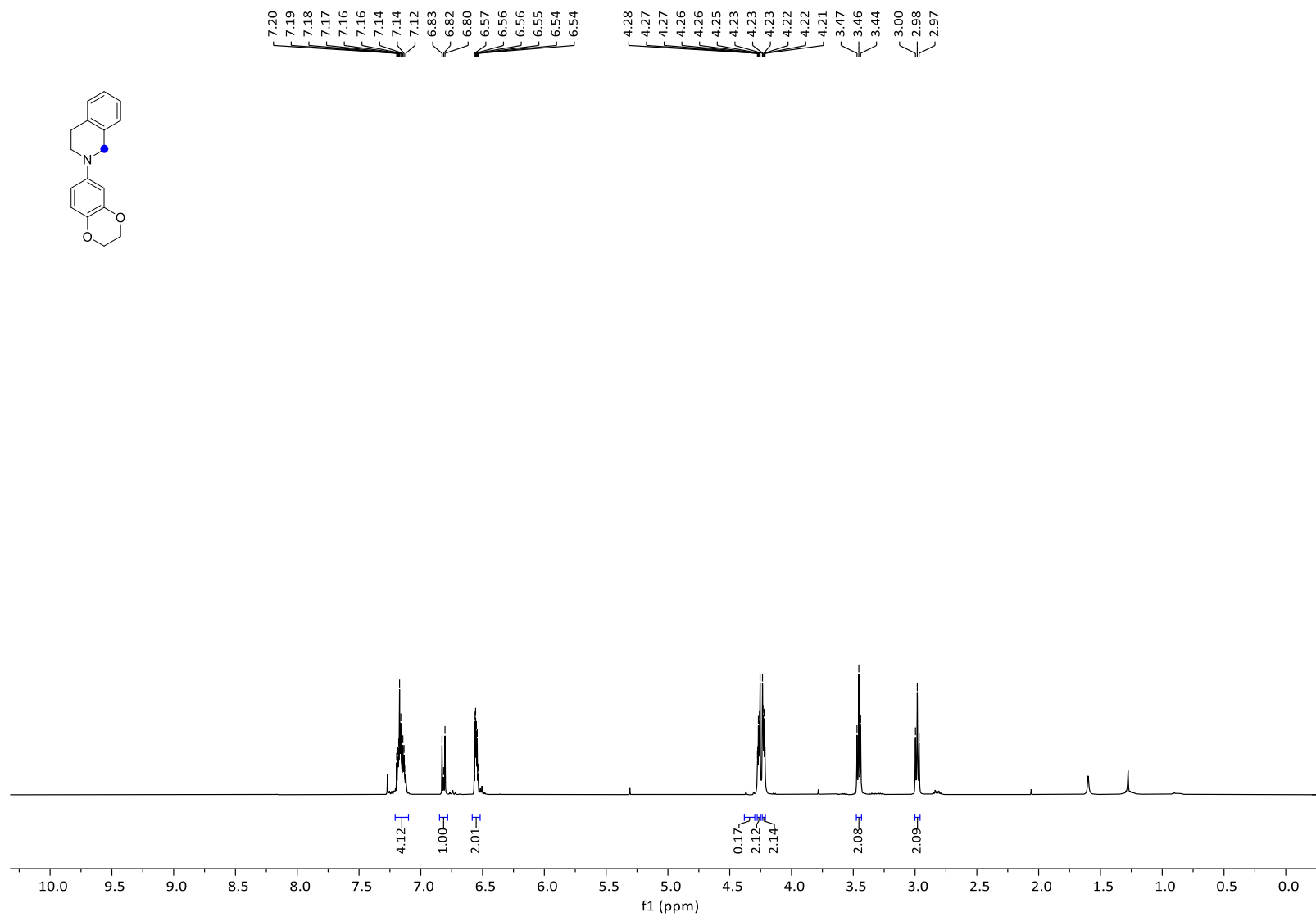
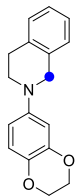
<sup>1</sup>H NMR spectrum of **07** (600 MHz, CDCl<sub>3</sub>)



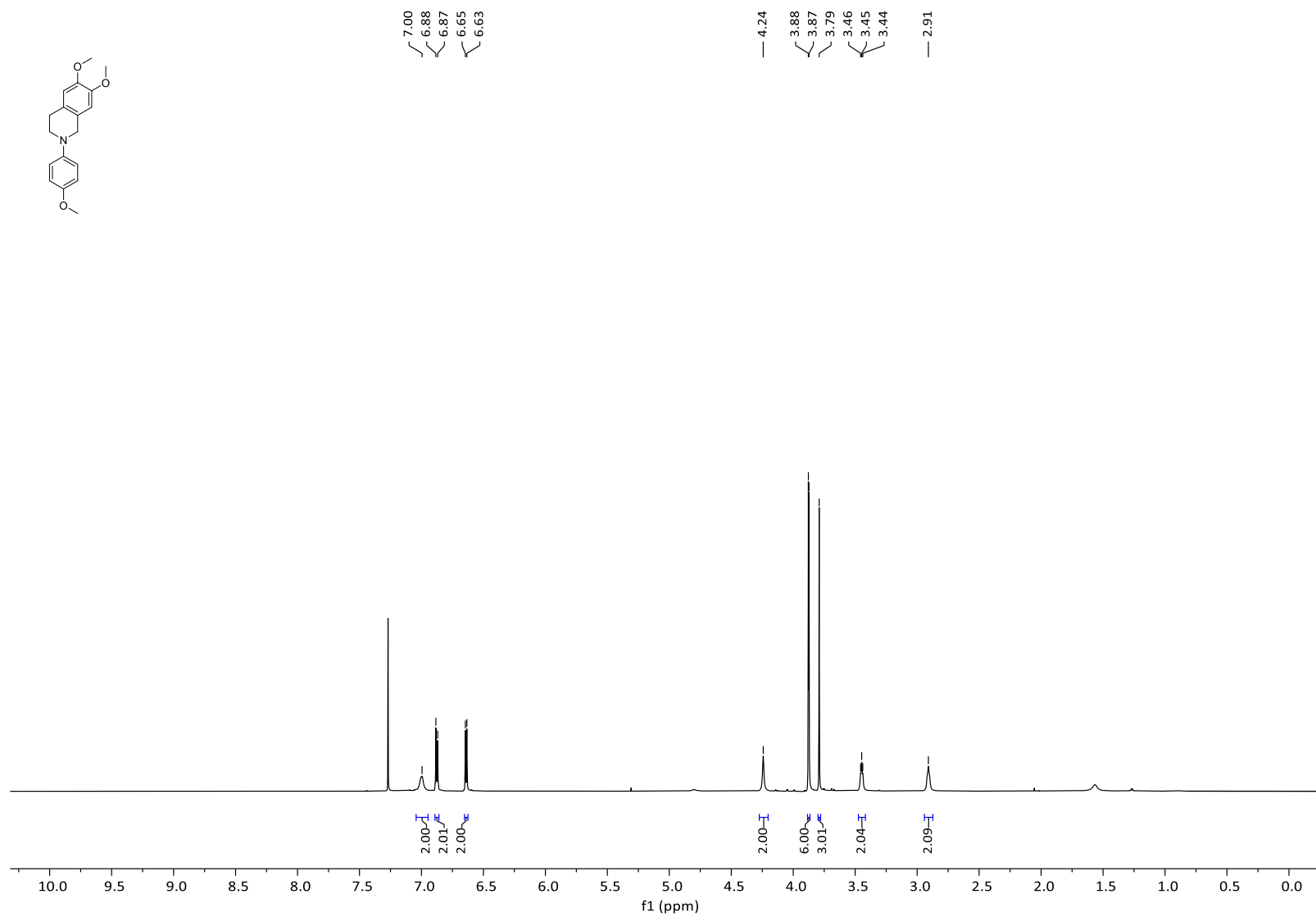
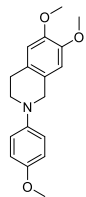
$^1\text{H}$  NMR spectrum of **7a** (600 MHz,  $\text{CDCl}_3$ )



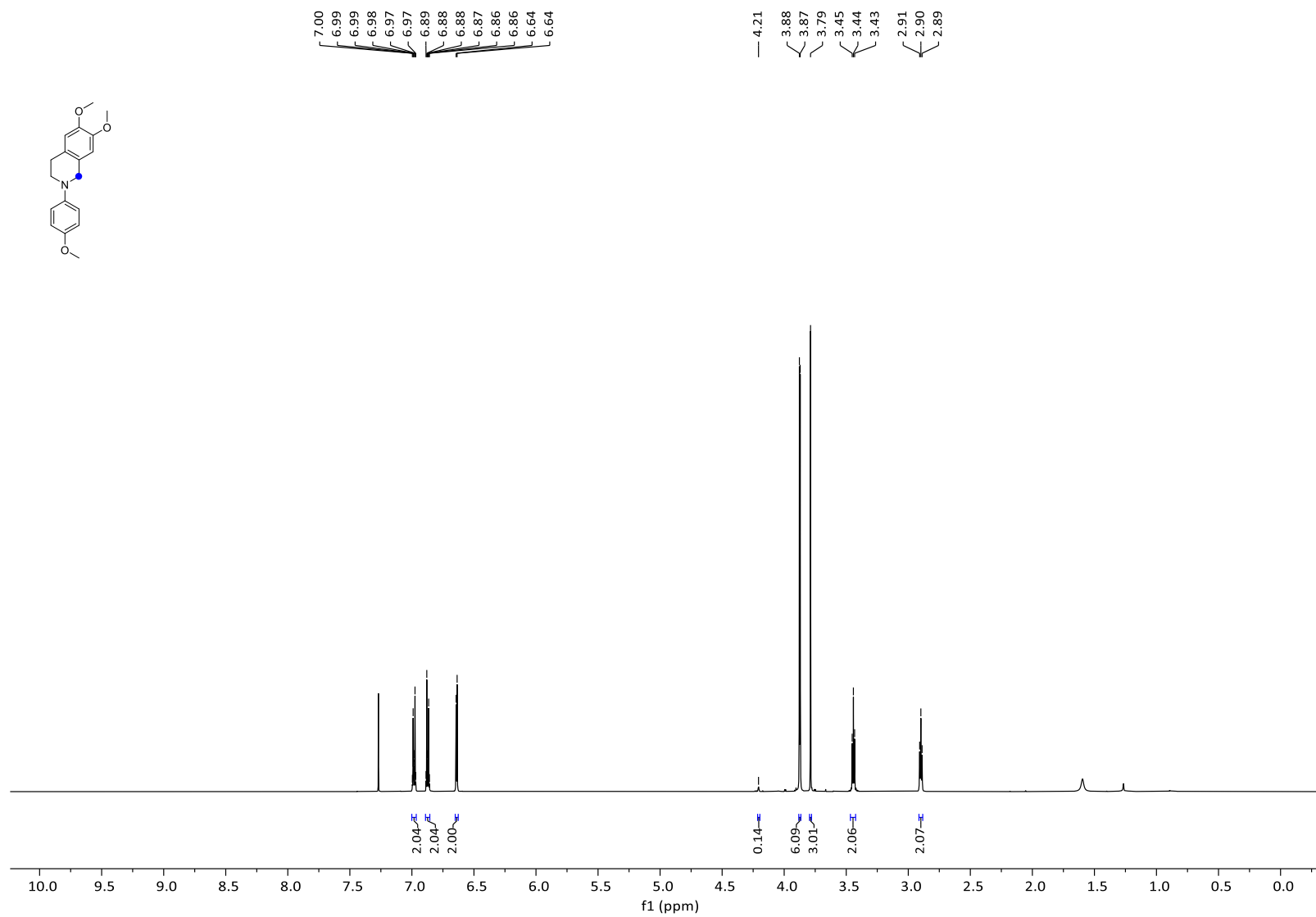
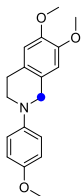
<sup>1</sup>H NMR spectrum of **08** (600 MHz, CDCl<sub>3</sub>)



$^1\text{H}$  NMR spectrum of **8a** (600 MHz,  $\text{CDCl}_3$ )

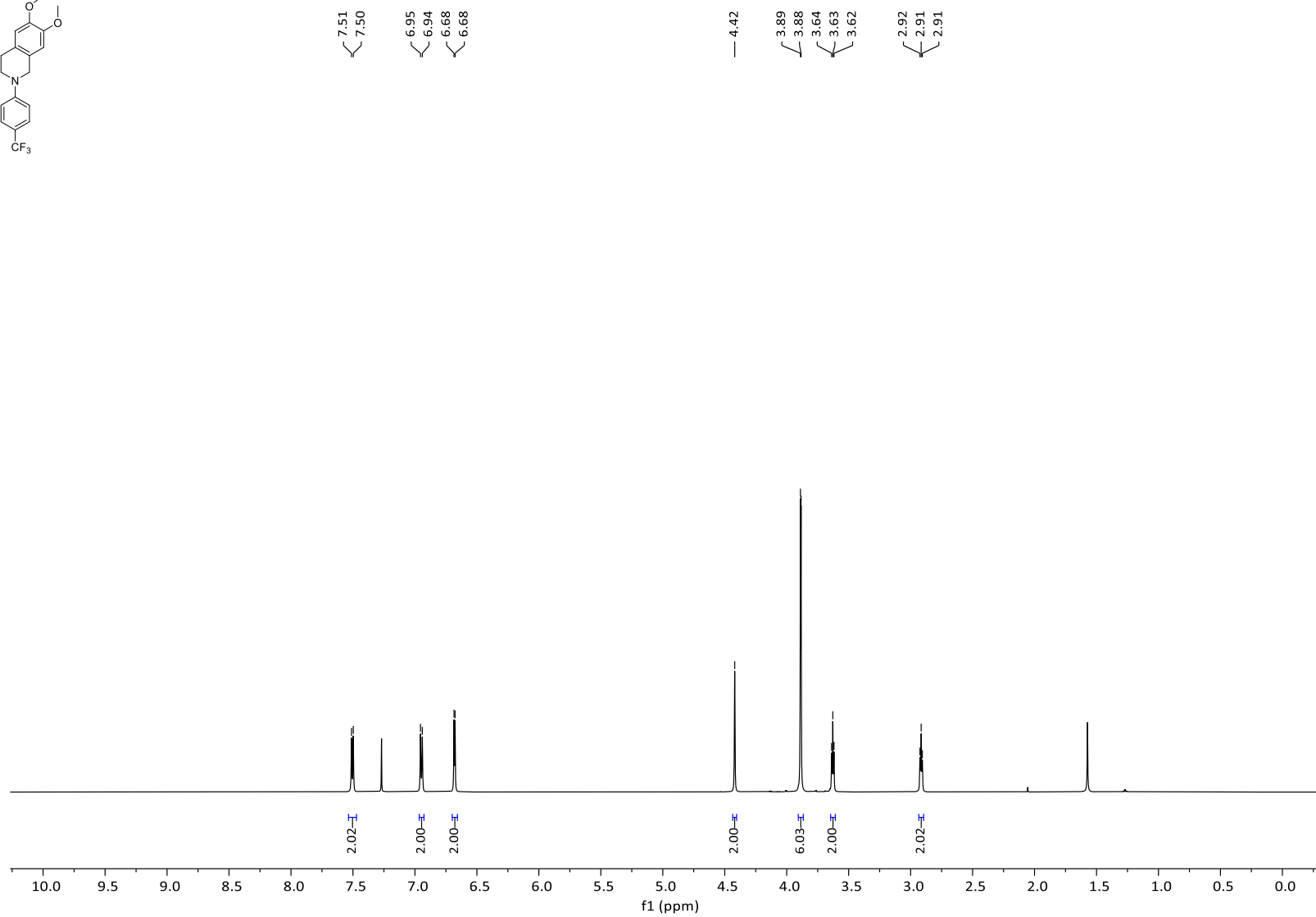
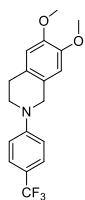


<sup>1</sup>H NMR spectrum of **09** (600 MHz, CDCl<sub>3</sub>)

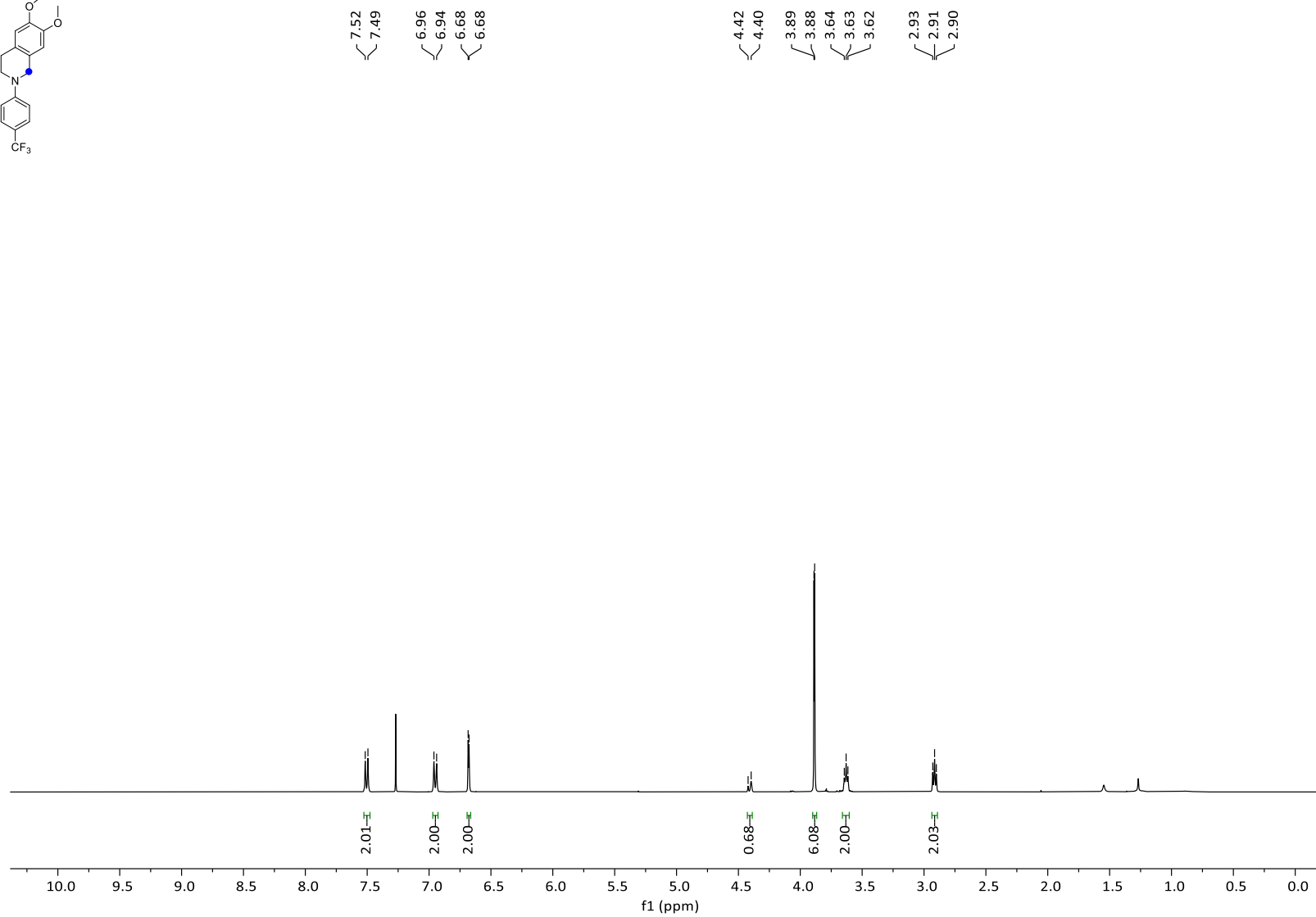
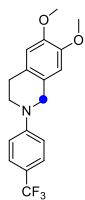


<sup>1</sup>H NMR spectrum of **9a** (600 MHz, CDCl<sub>3</sub>)

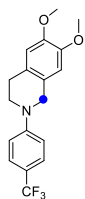




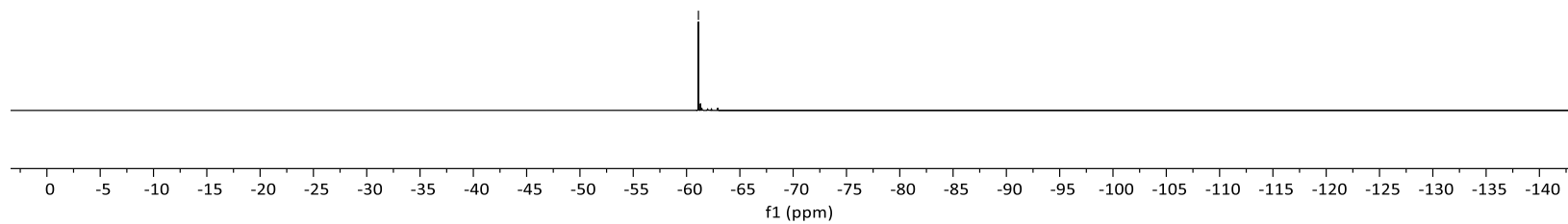
$^1\text{H}$  NMR spectrum of **10** (600 MHz,  $\text{CDCl}_3$ )



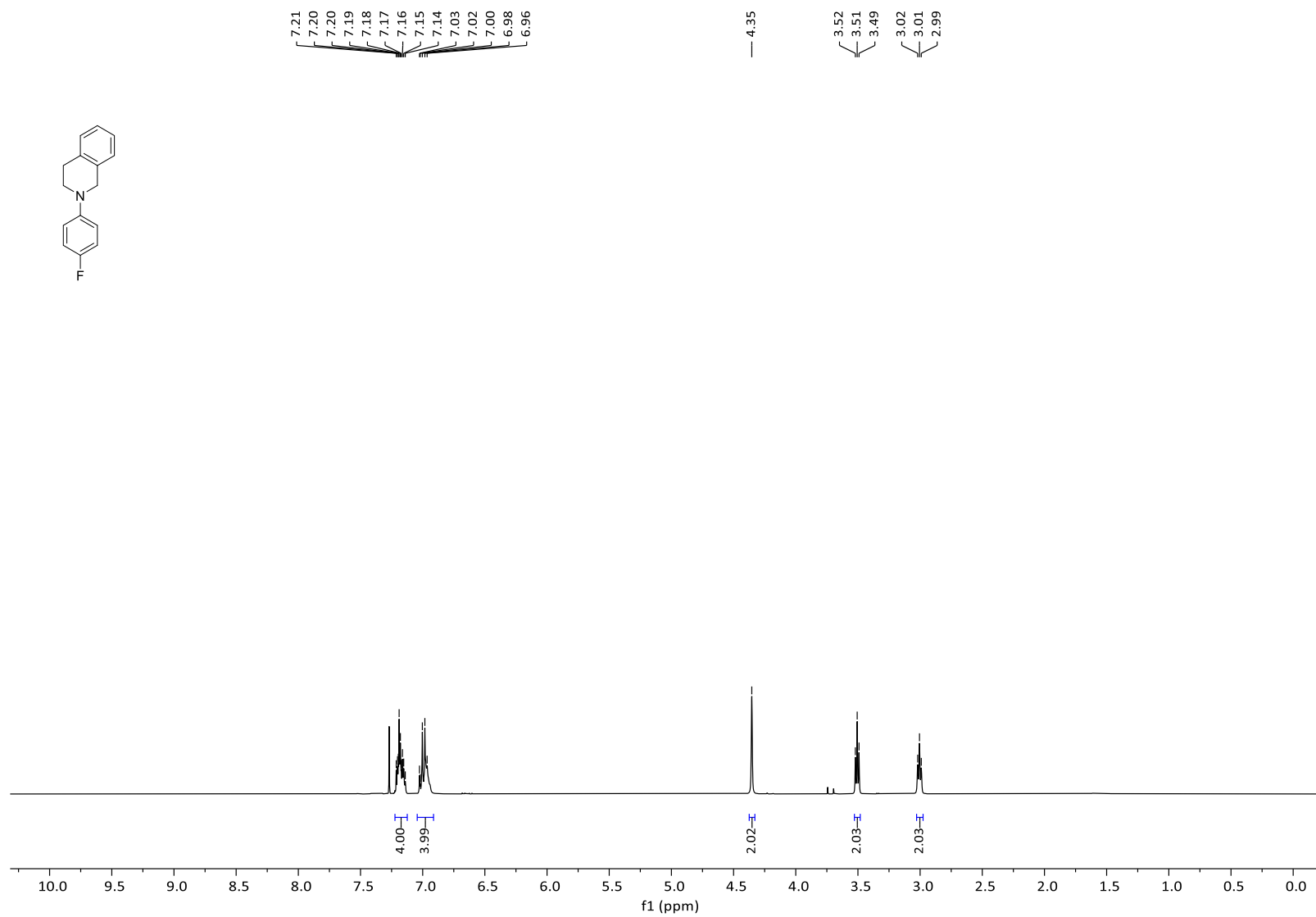
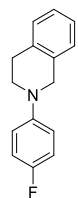
$^1\text{H}$  NMR spectrum of **10a** (600 MHz,  $\text{CDCl}_3$ )



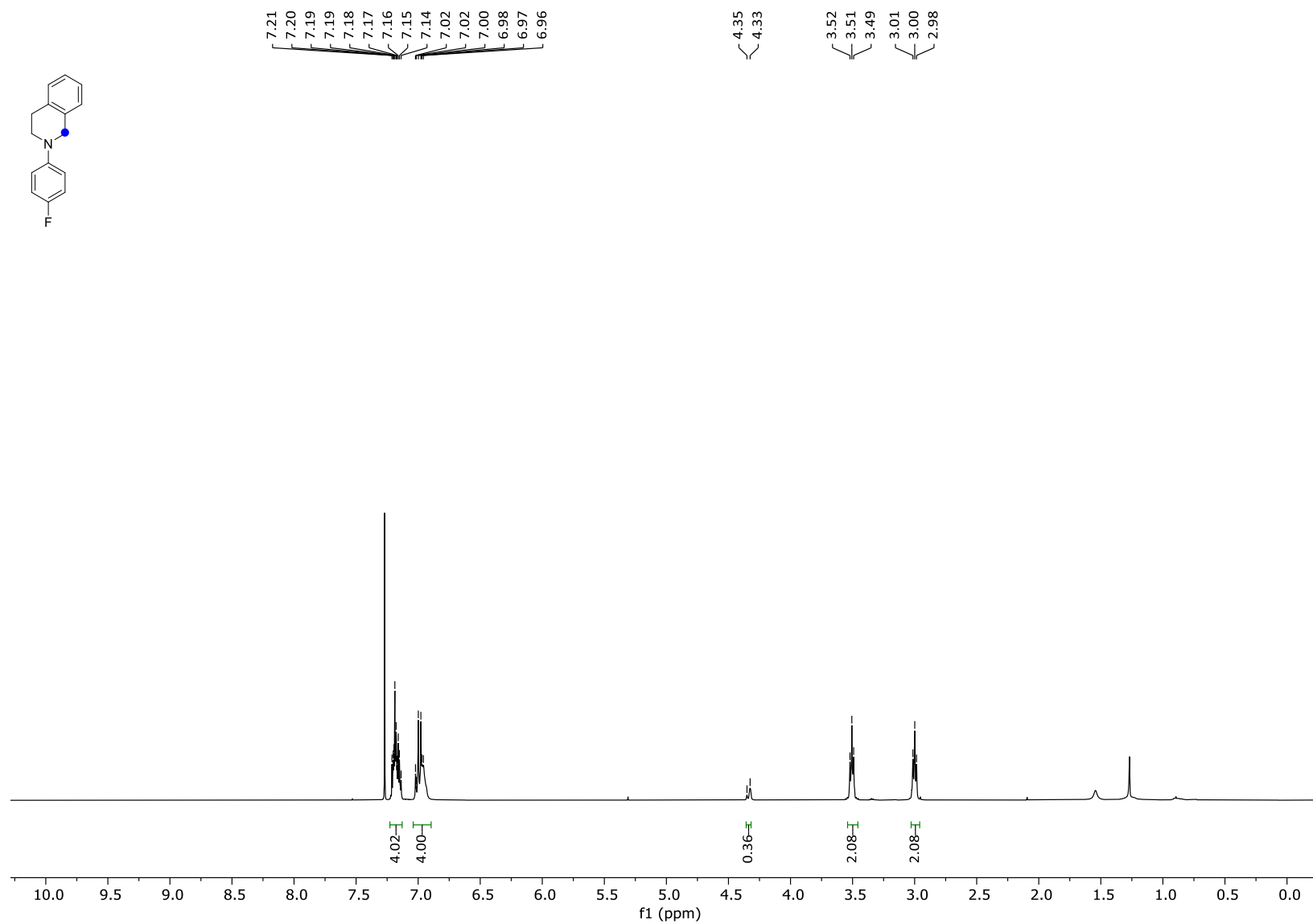
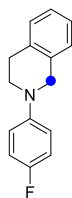
— -61.11



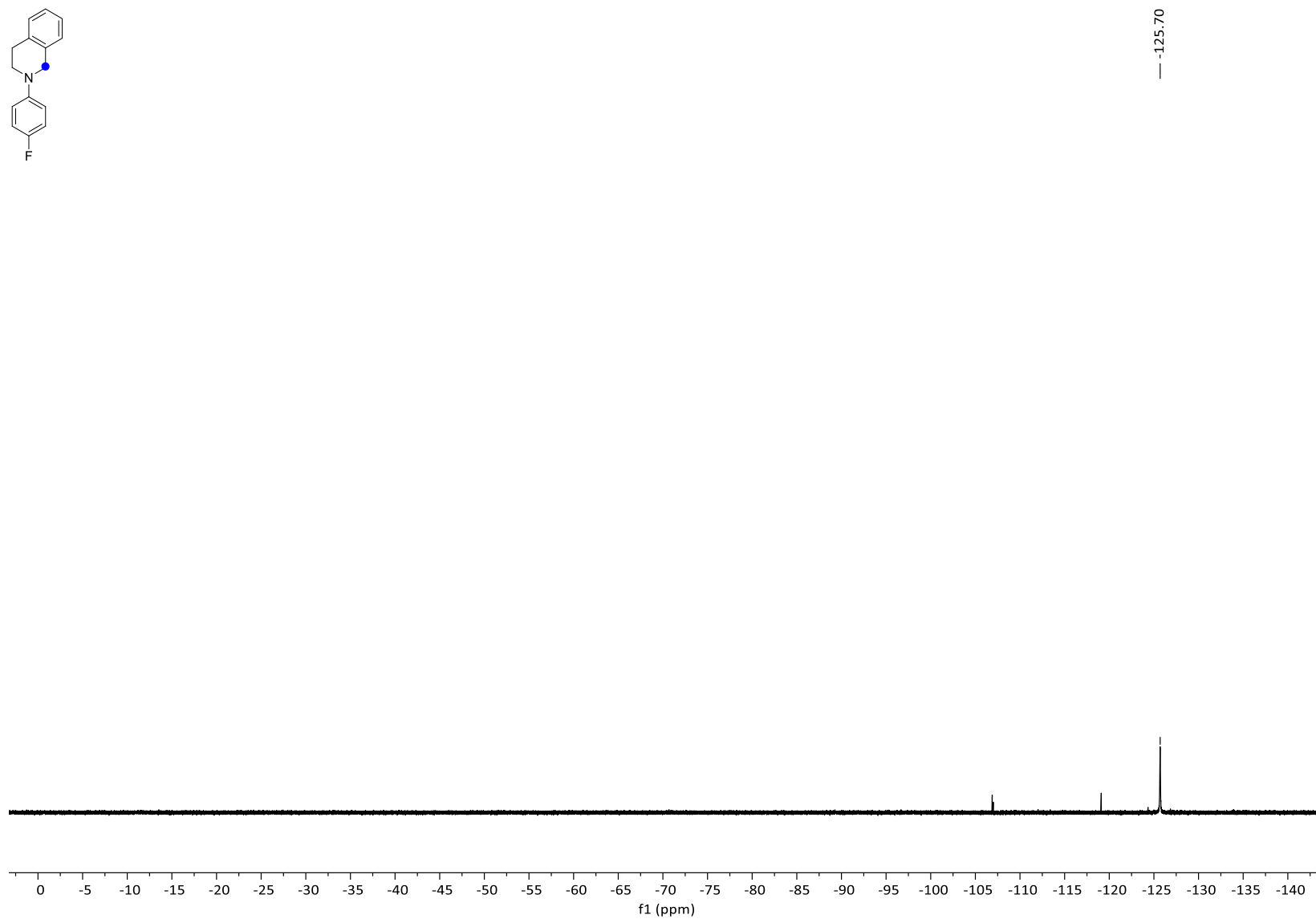
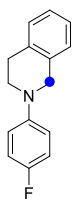
$^{19}\text{F}$  NMR spectrum of **10a** (471 MHz,  $\text{CDCl}_3$ )



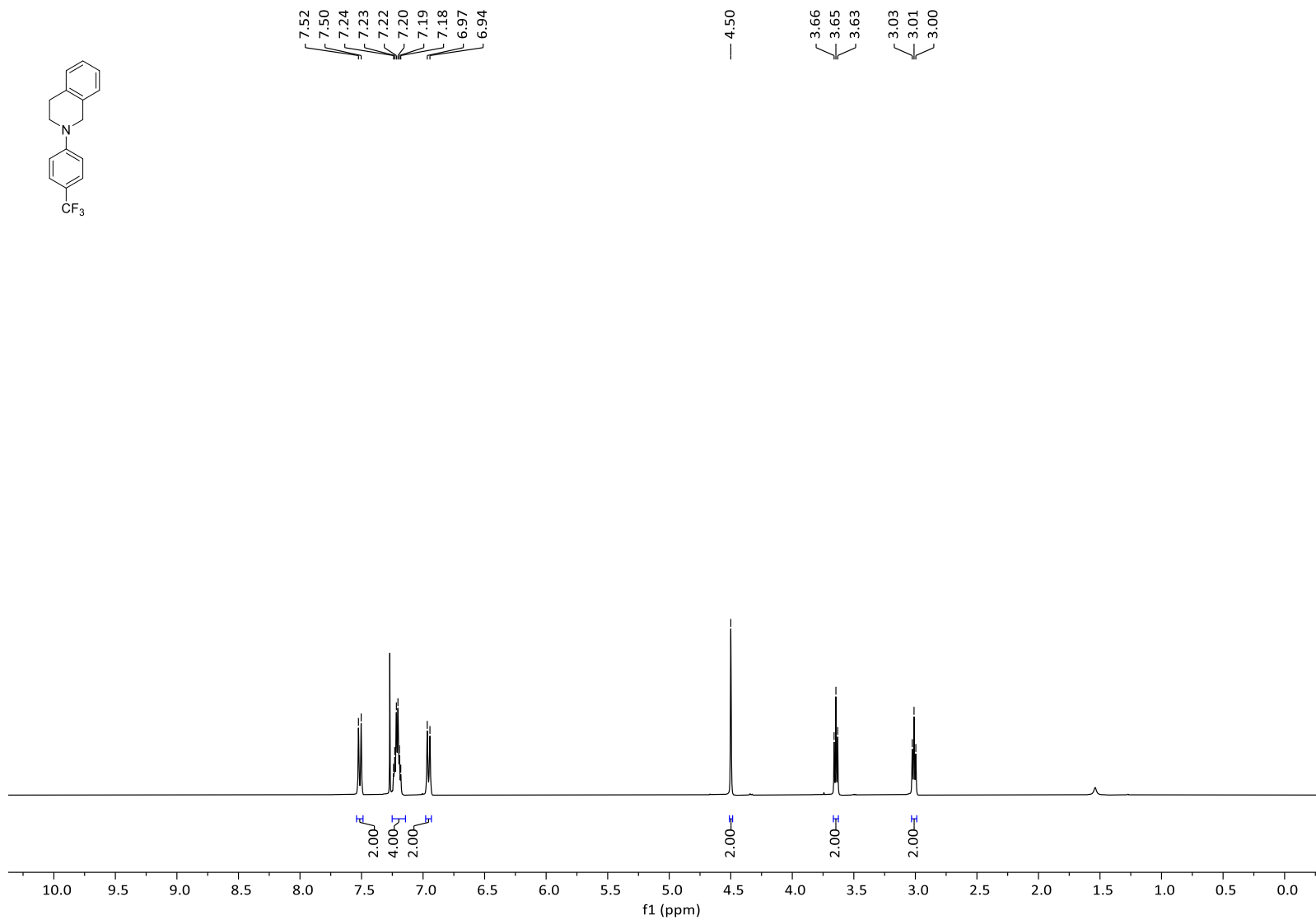
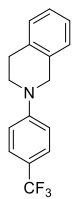
<sup>1</sup>H NMR spectrum of **11** (600 MHz, CDCl<sub>3</sub>)



$^1\text{H}$  NMR spectrum of **11a** (600 MHz,  $\text{CDCl}_3$ )

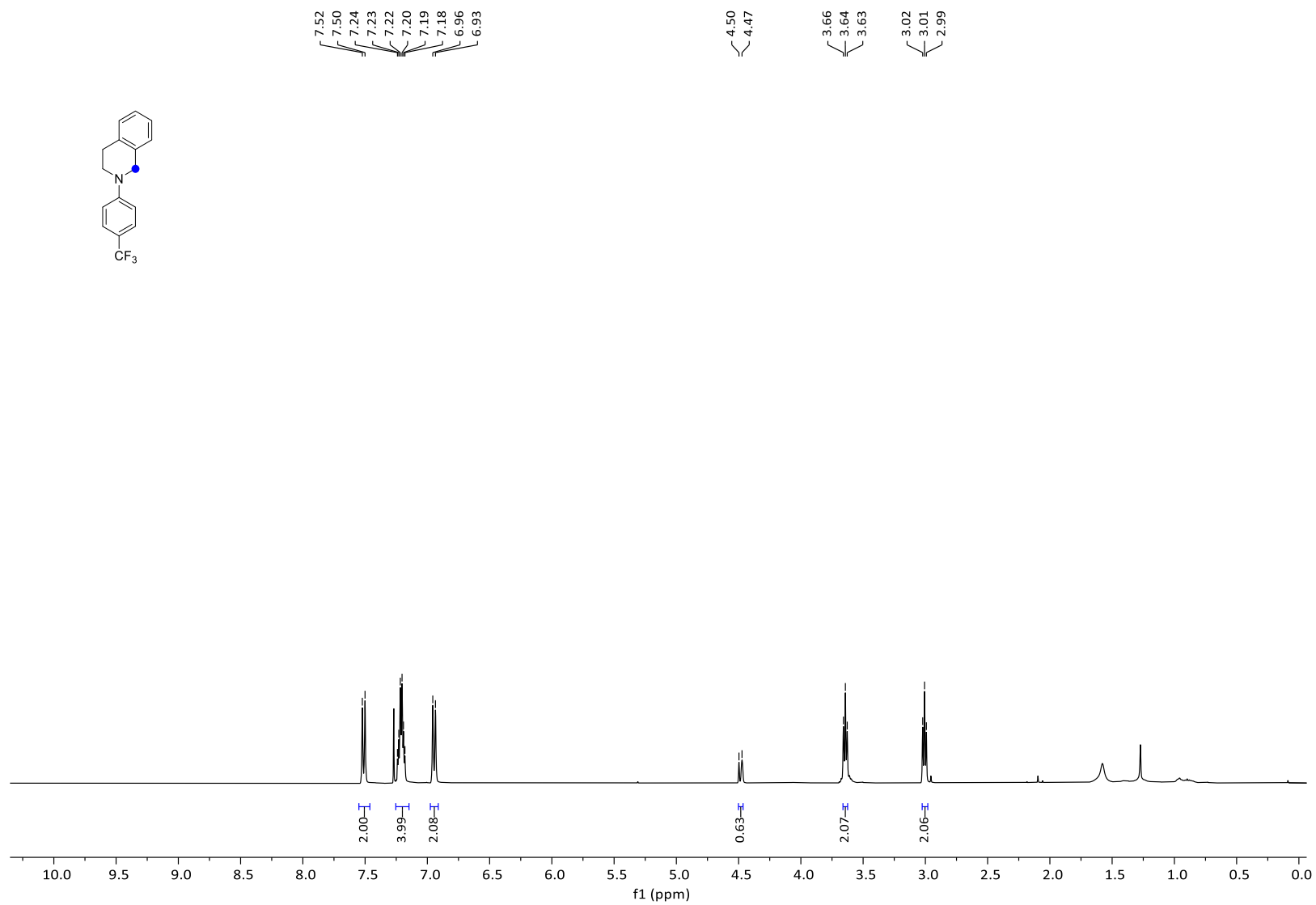
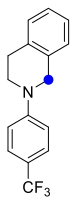


$^{19}\text{F}$  NMR spectrum of **11a** (471 MHz,  $\text{CDCl}_3$ )



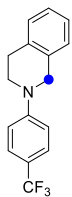
NMR spectrum of **12** (600 MHz, CDCl<sub>3</sub>)

<sup>1</sup>H

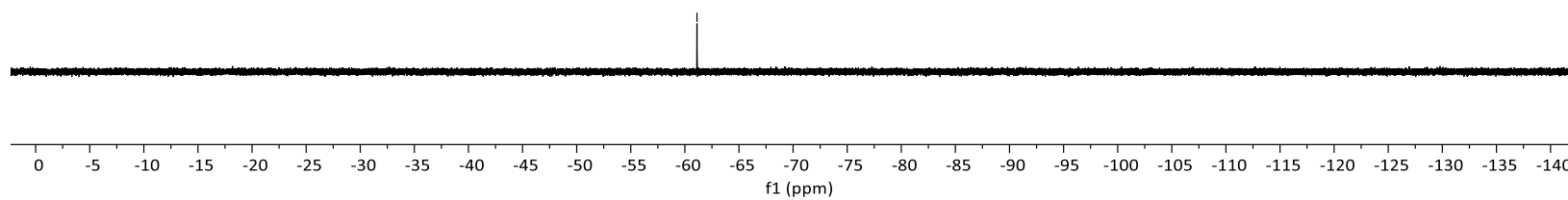


$^1\text{H}$  NMR spectrum of **12a** (600 MHz,  $\text{CDCl}_3$ )

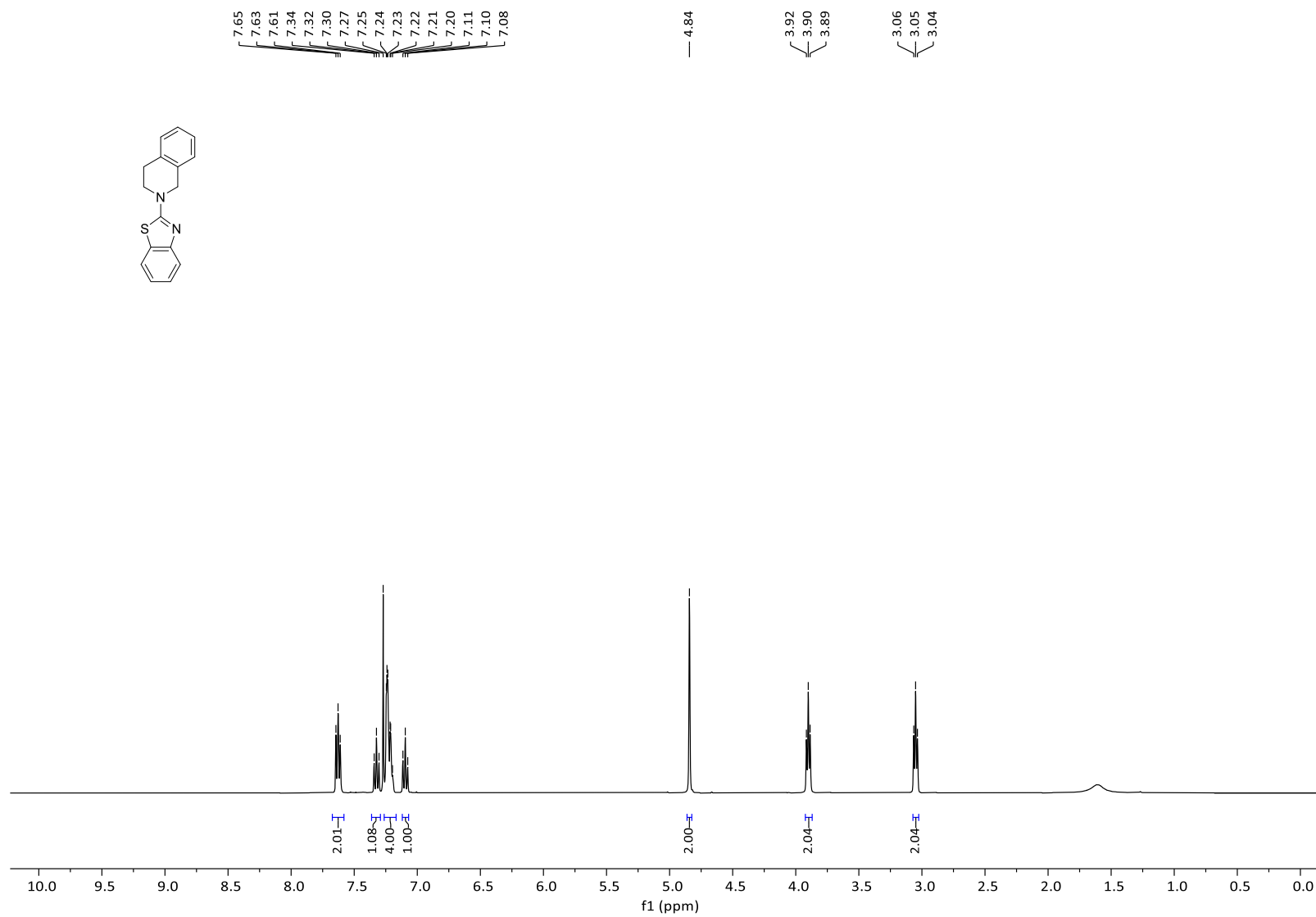
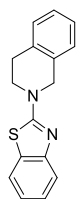




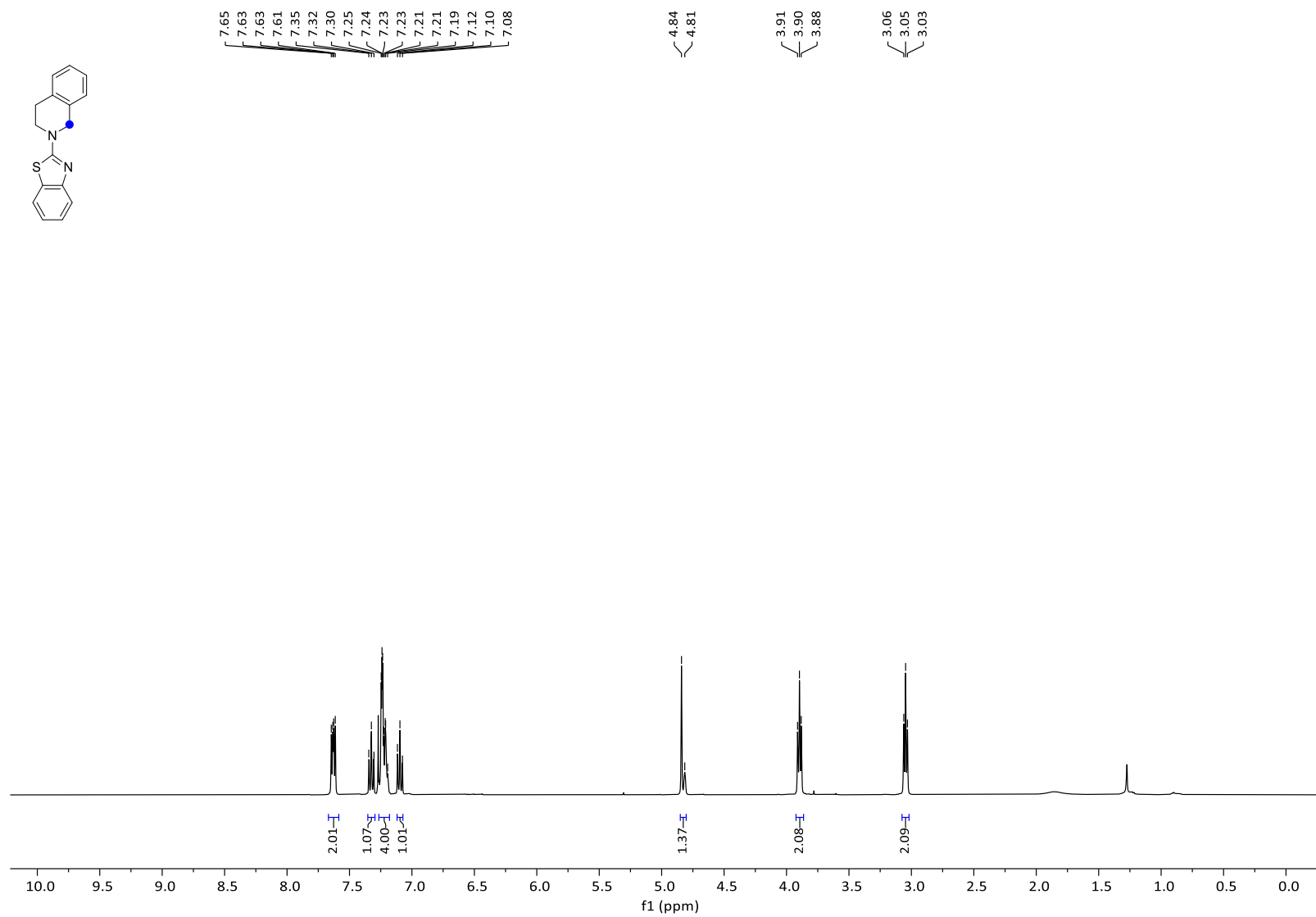
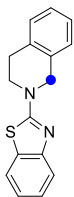
— -61.12



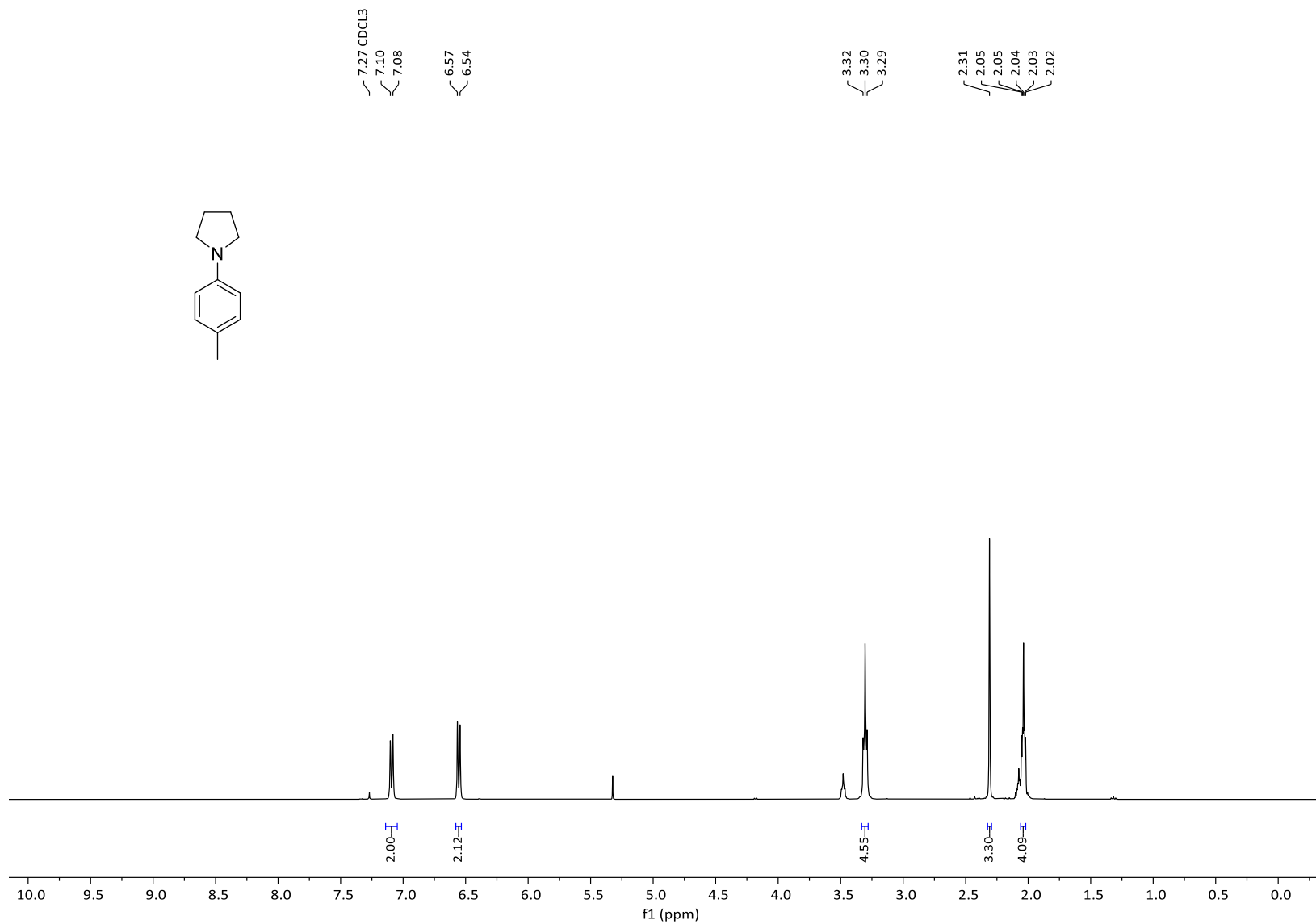
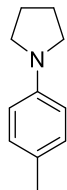
$^{19}\text{F}$  NMR spectrum of **12a** (376 MHz,  $\text{CDCl}_3$ )



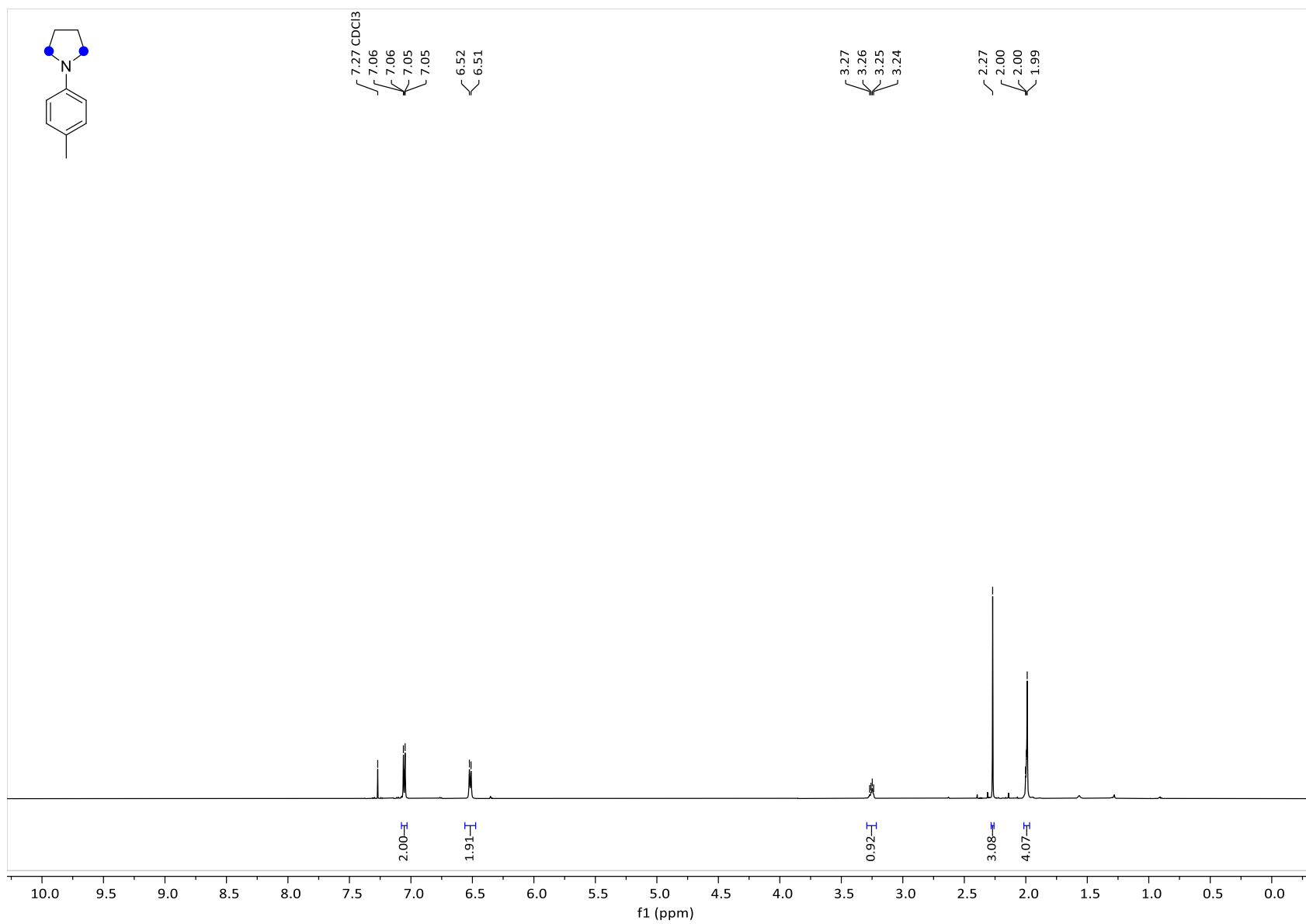
<sup>1</sup>H NMR spectrum of **13** (600 MHz, CDCl<sub>3</sub>)



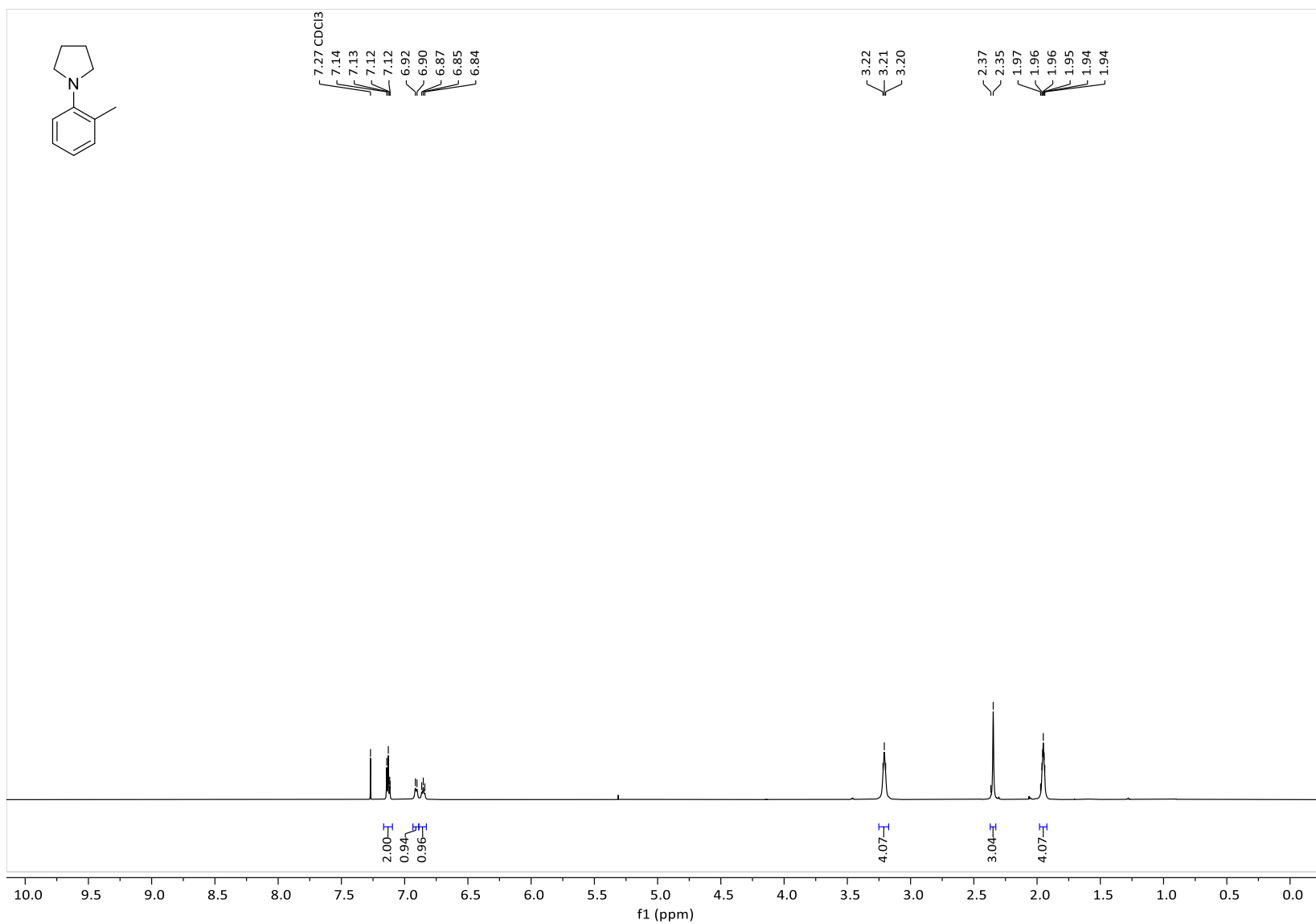
$^1\text{H}$  NMR spectrum of **13a** (600 MHz,  $\text{CDCl}_3$ )



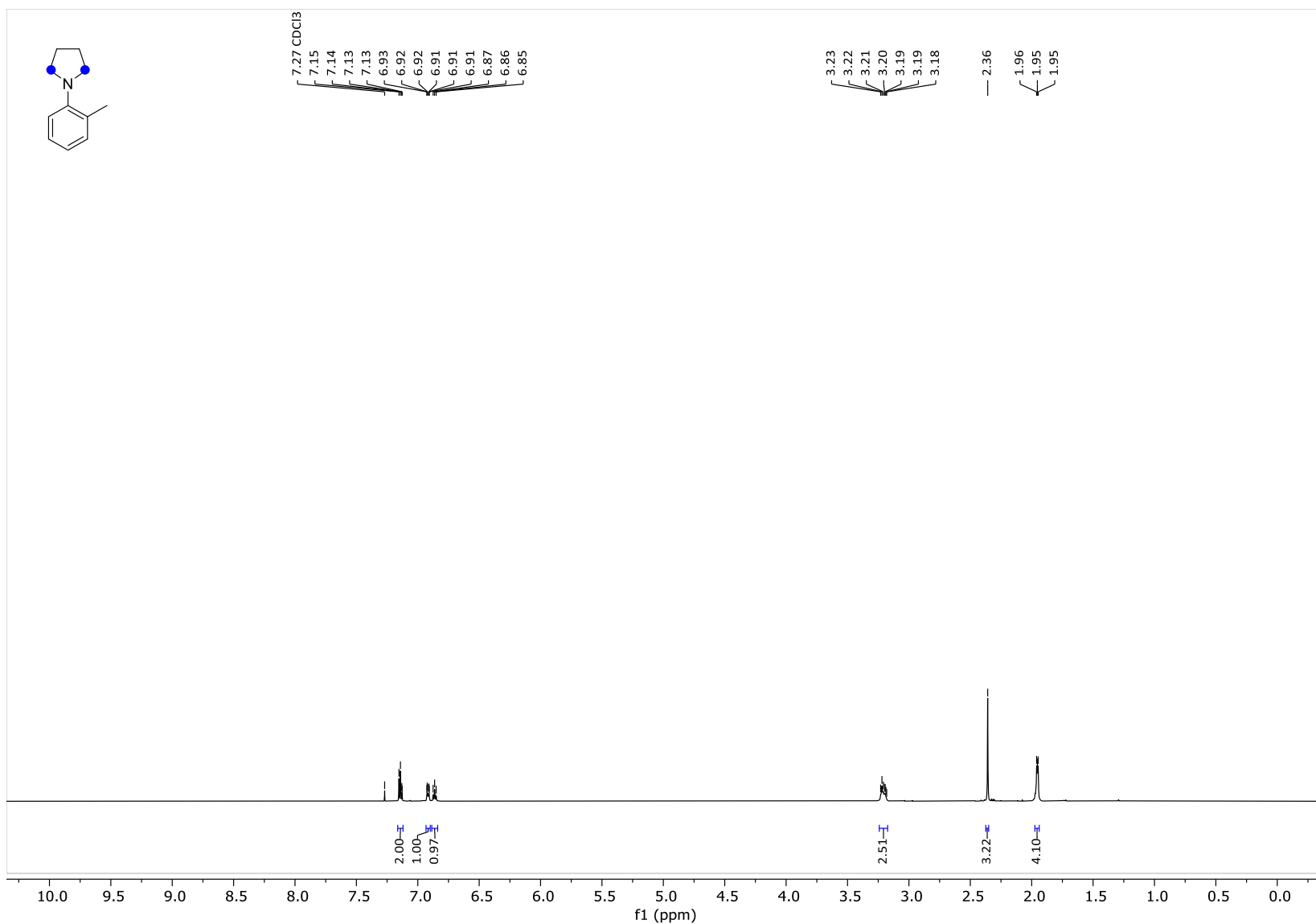
$^1\text{H}$  NMR spectrum of **14** (600 MHz,  $\text{CDCl}_3$ )



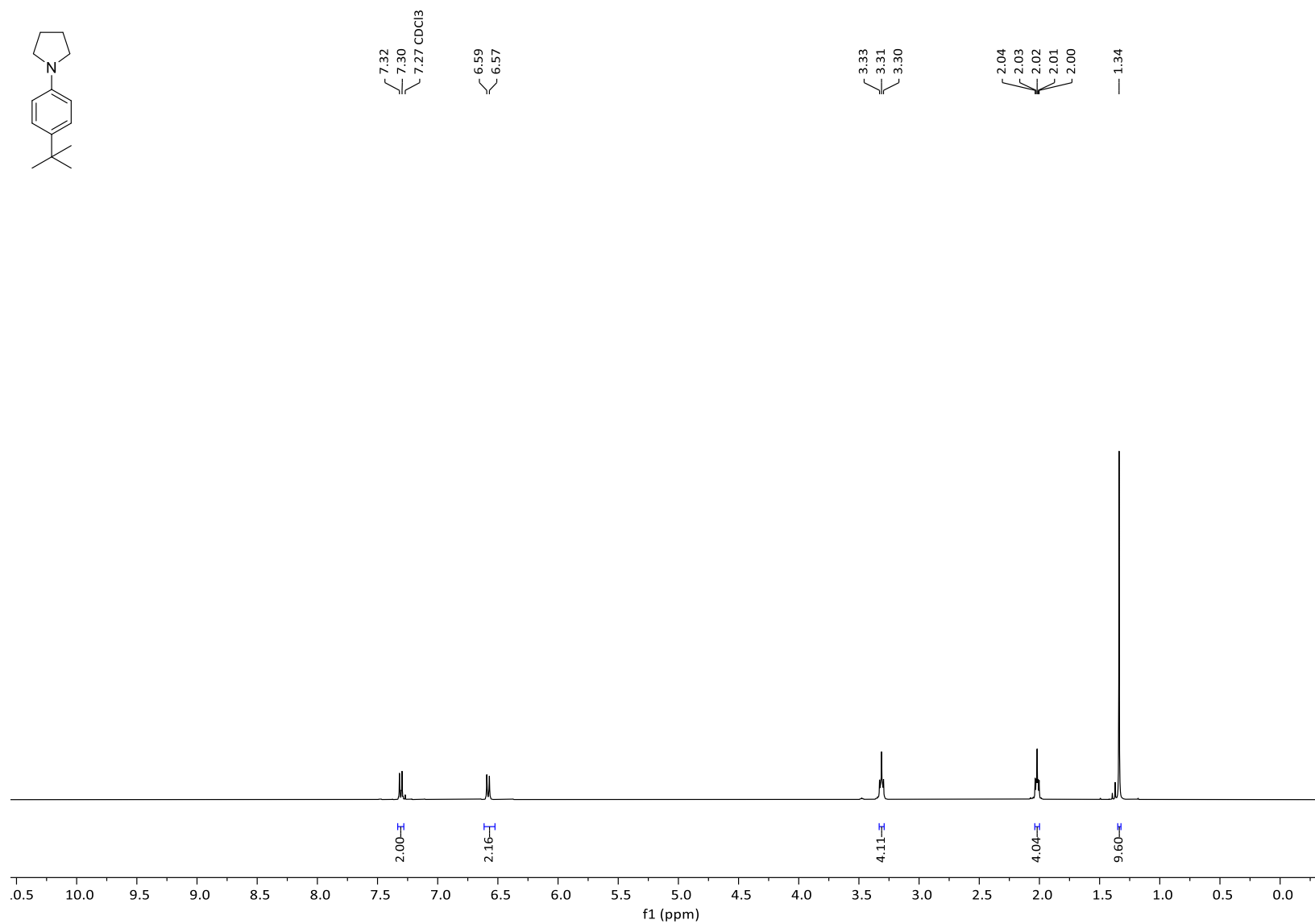
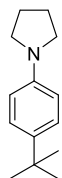
<sup>1</sup>H NMR spectrum of **14a** (600MHz, CDCl<sub>3</sub>)



<sup>1</sup>H NMR spectrum of **15** (600 MHz, CDCl<sub>3</sub>)

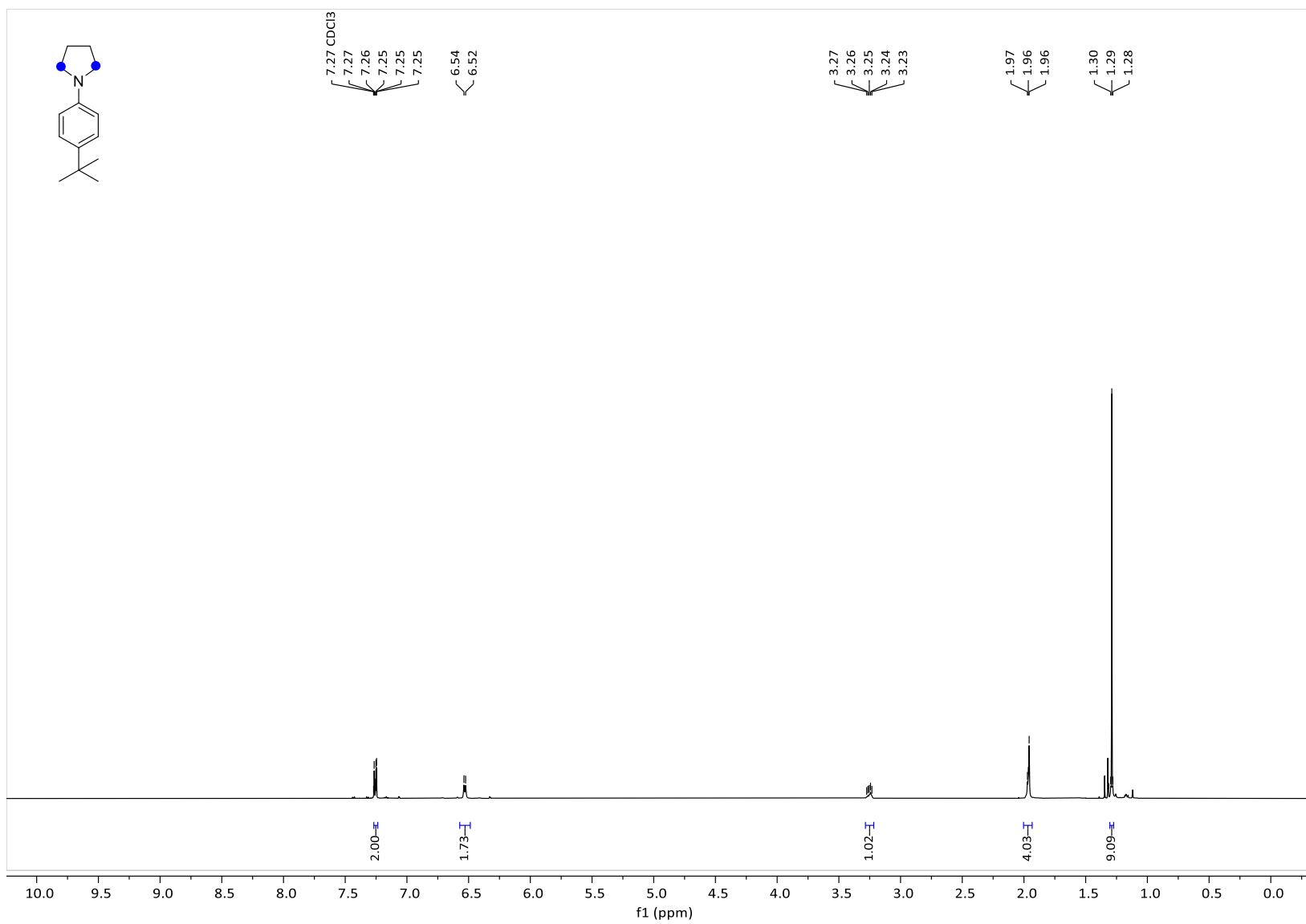


<sup>1</sup>H NMR spectrum of **15a** (600 MHz, CDCl<sub>3</sub>)

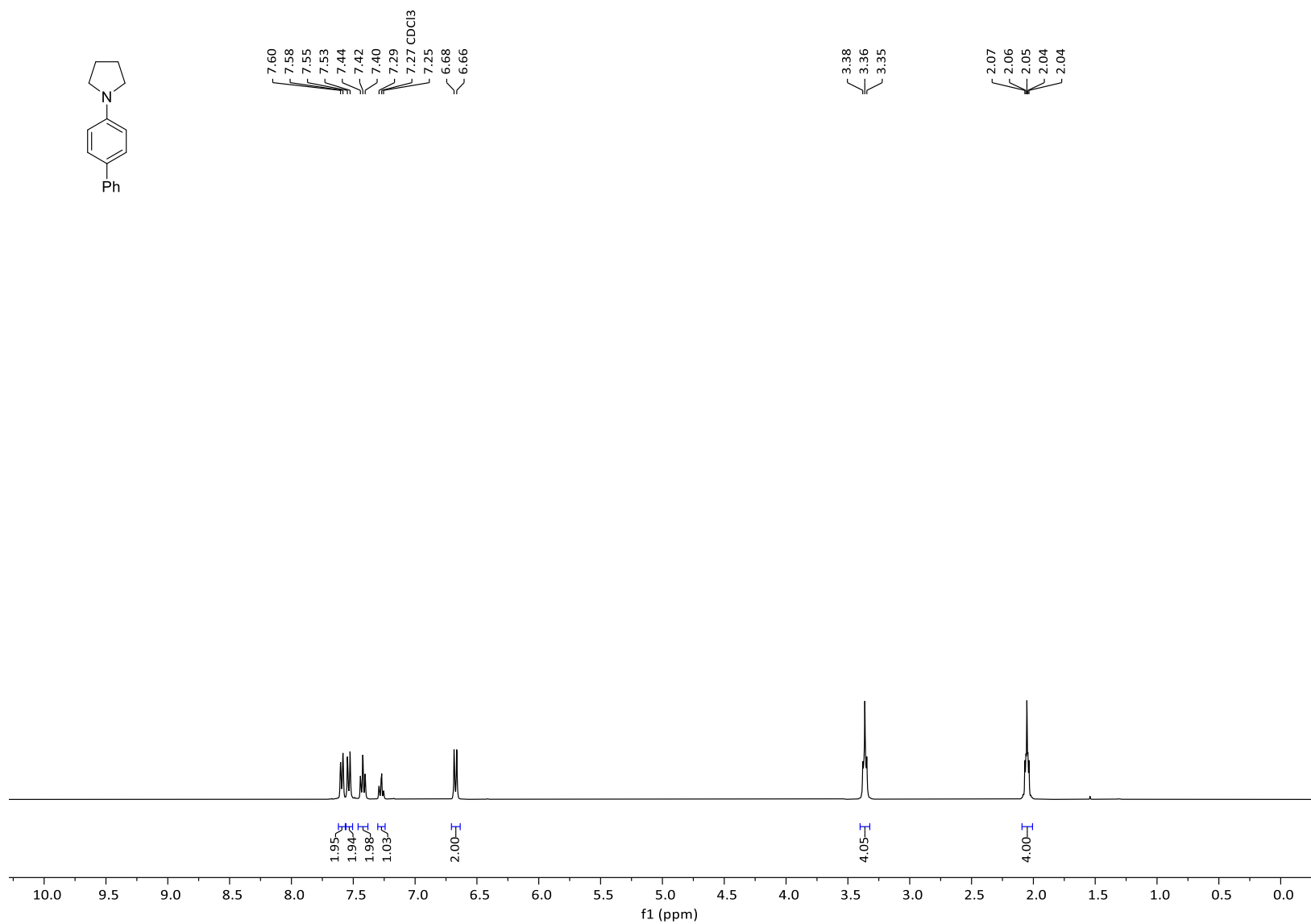
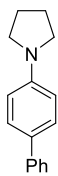


$^1\text{H}$  NMR spectrum of **16** (600 MHz,  $\text{CDCl}_3$ )

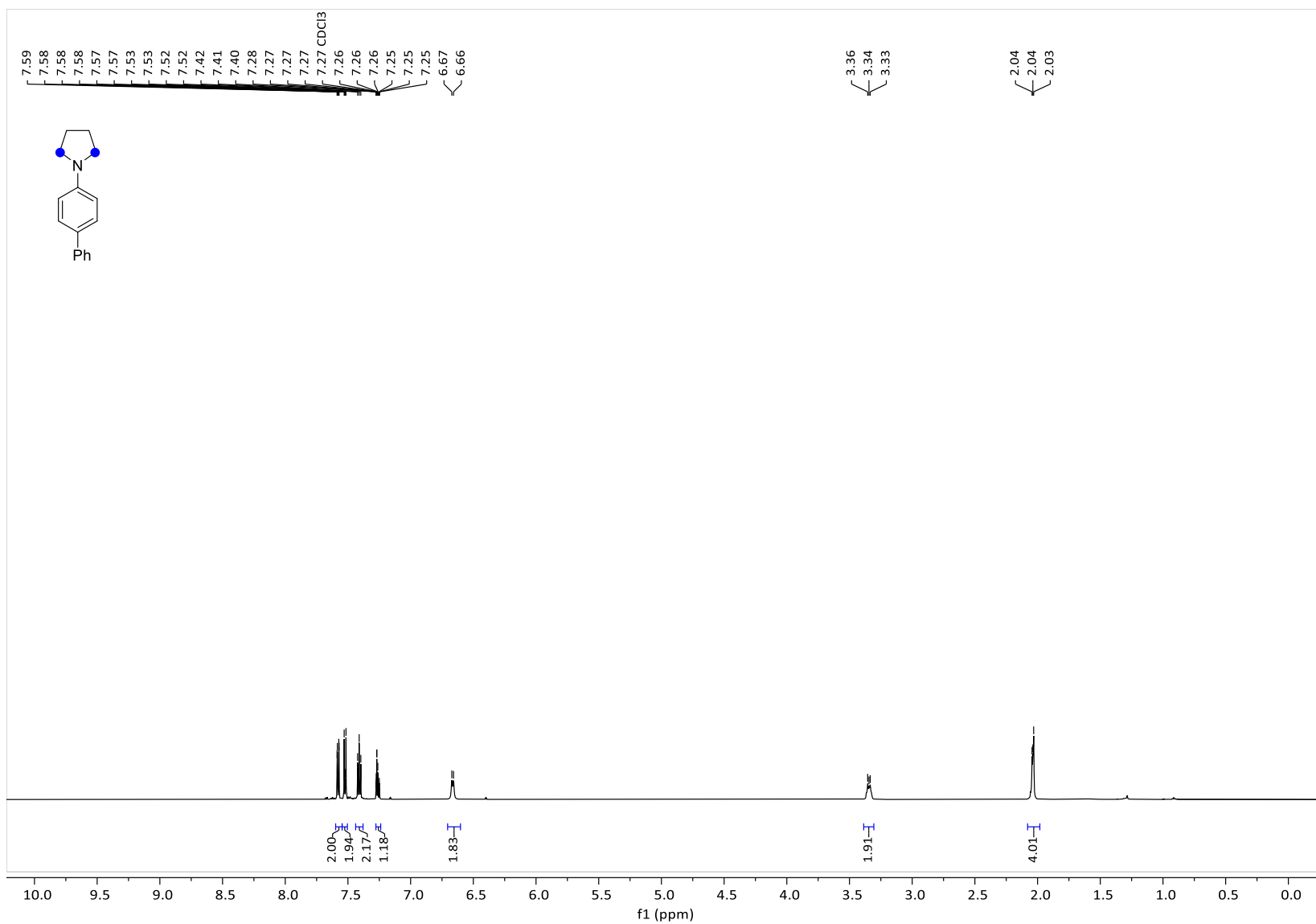




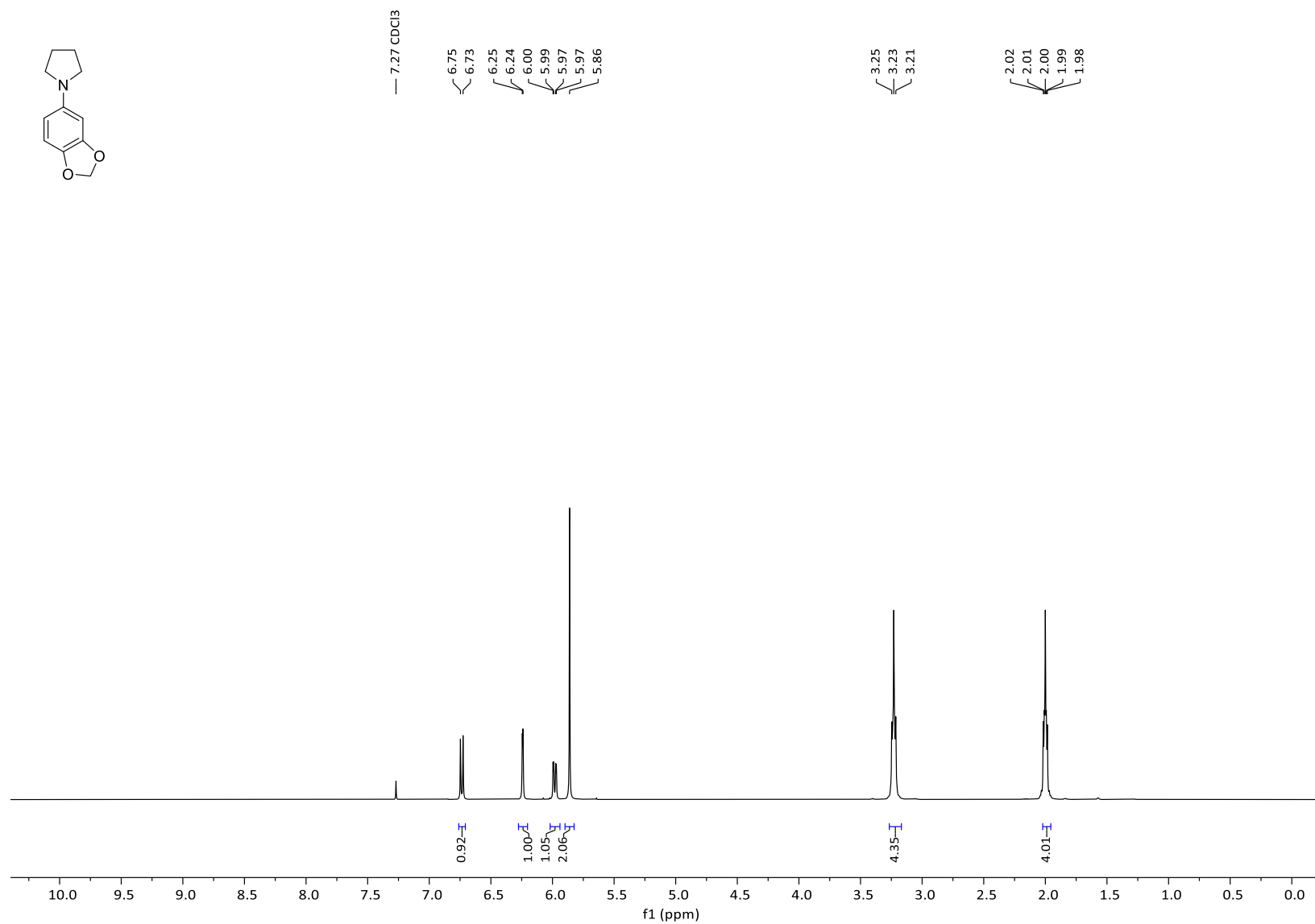
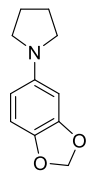
<sup>1</sup>H NMR spectrum of **16a** (600 MHz, CDCl<sub>3</sub>)



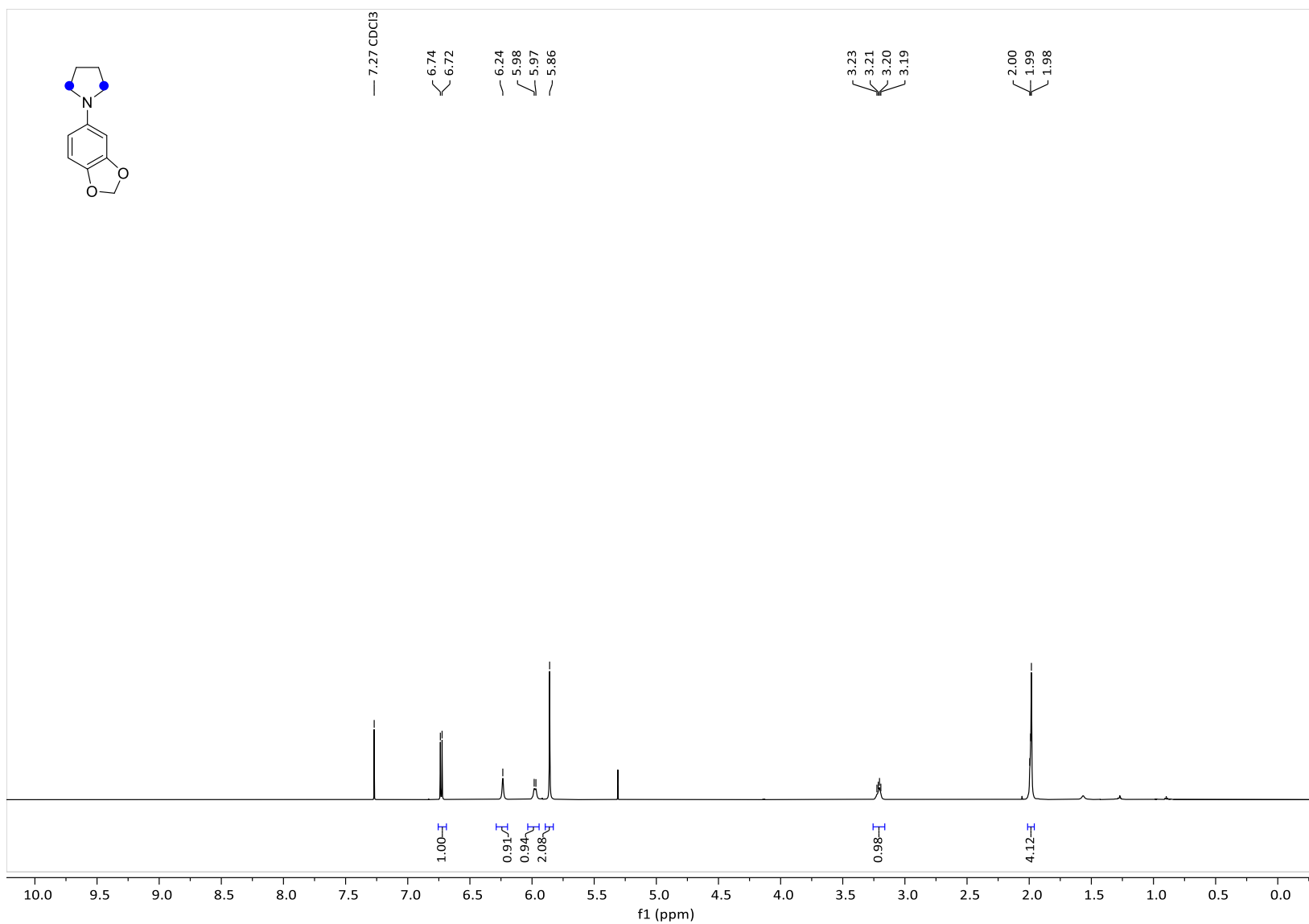
<sup>1</sup>H NMR spectrum of **17** (600 MHz, CDCl<sub>3</sub>)



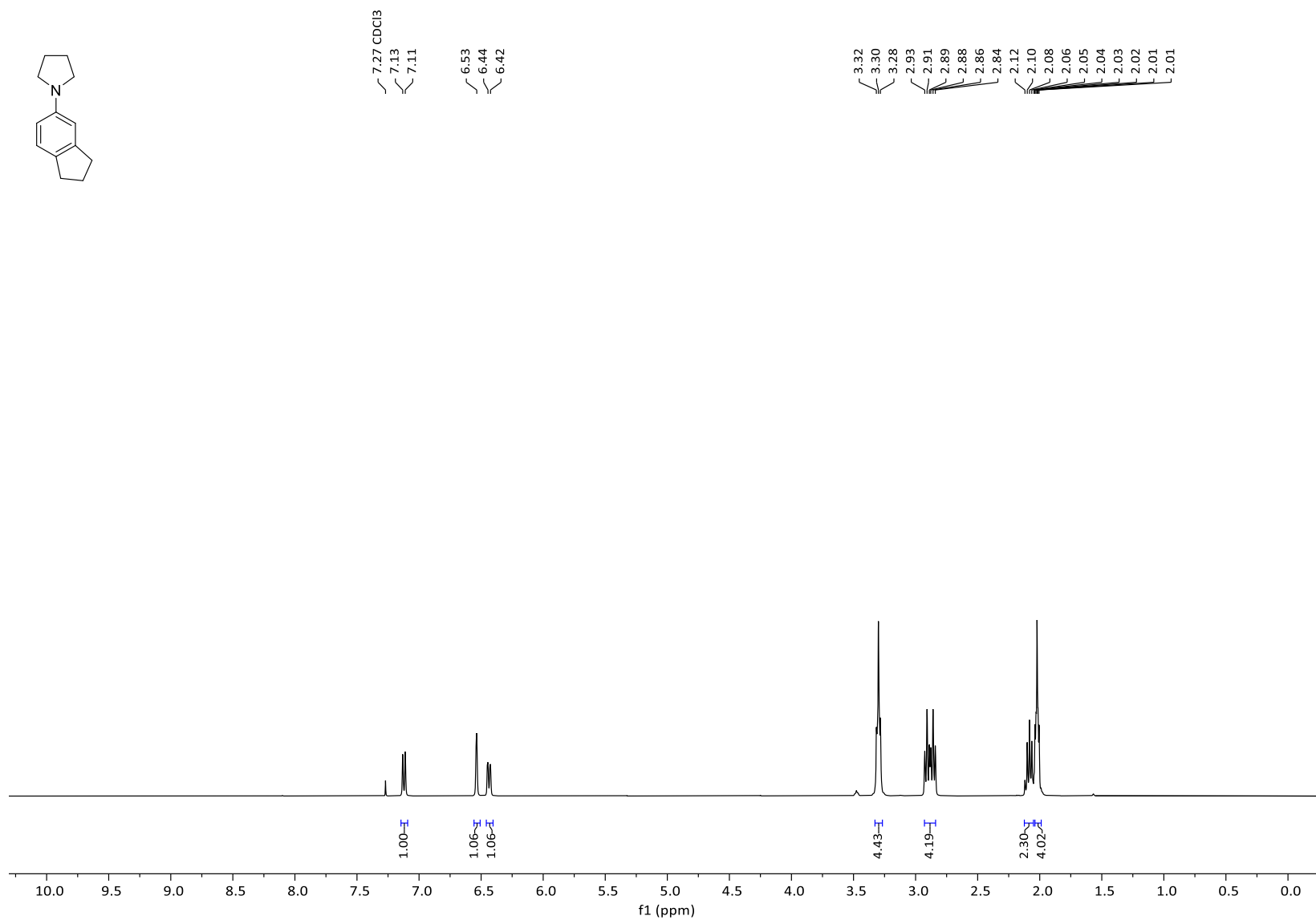
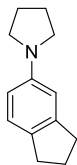
<sup>1</sup>H NMR spectrum of **17a** (600 MHz, CDCl<sub>3</sub>)



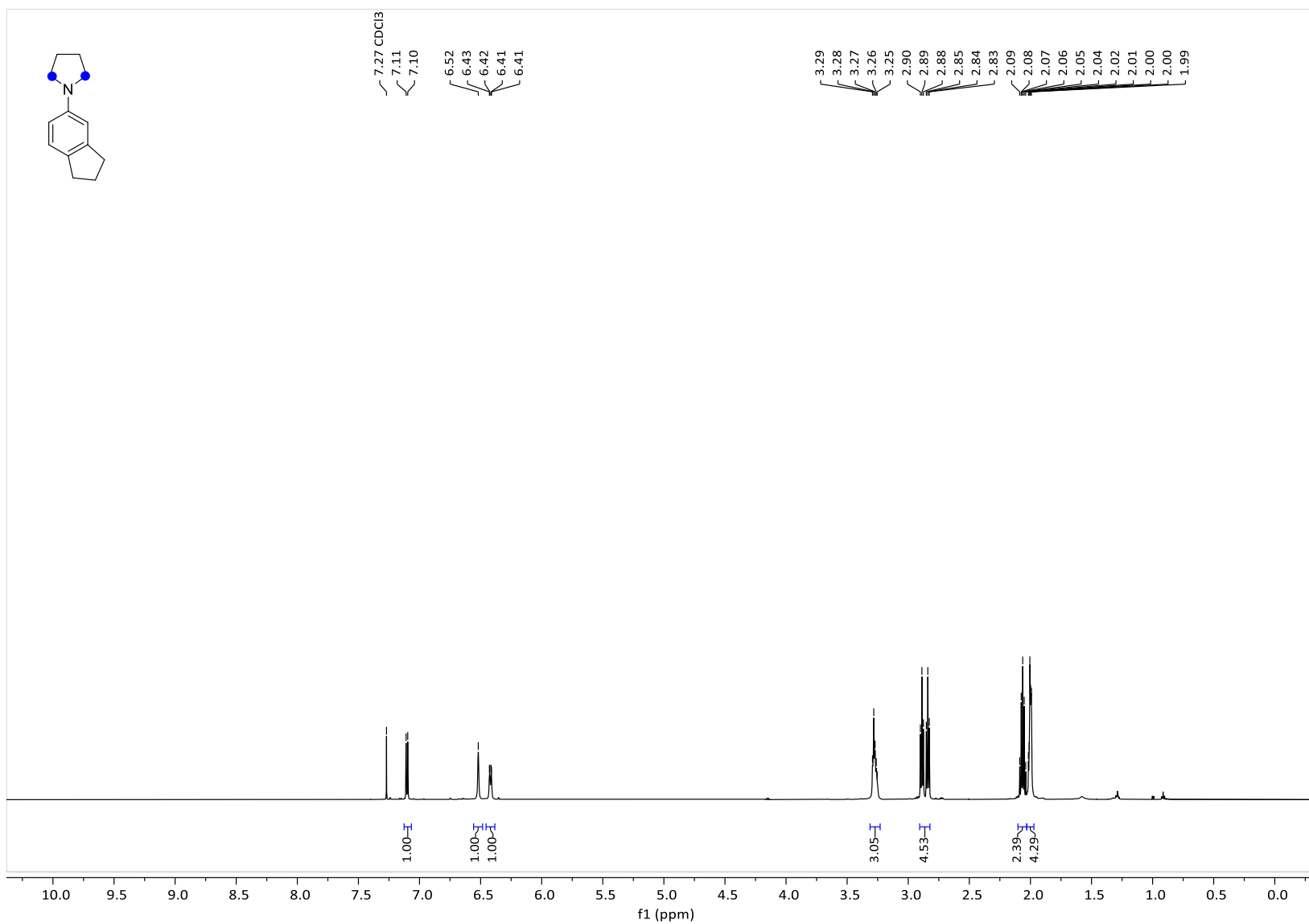
<sup>1</sup>H NMR spectrum of **18** (600 MHz, CDCl<sub>3</sub>)



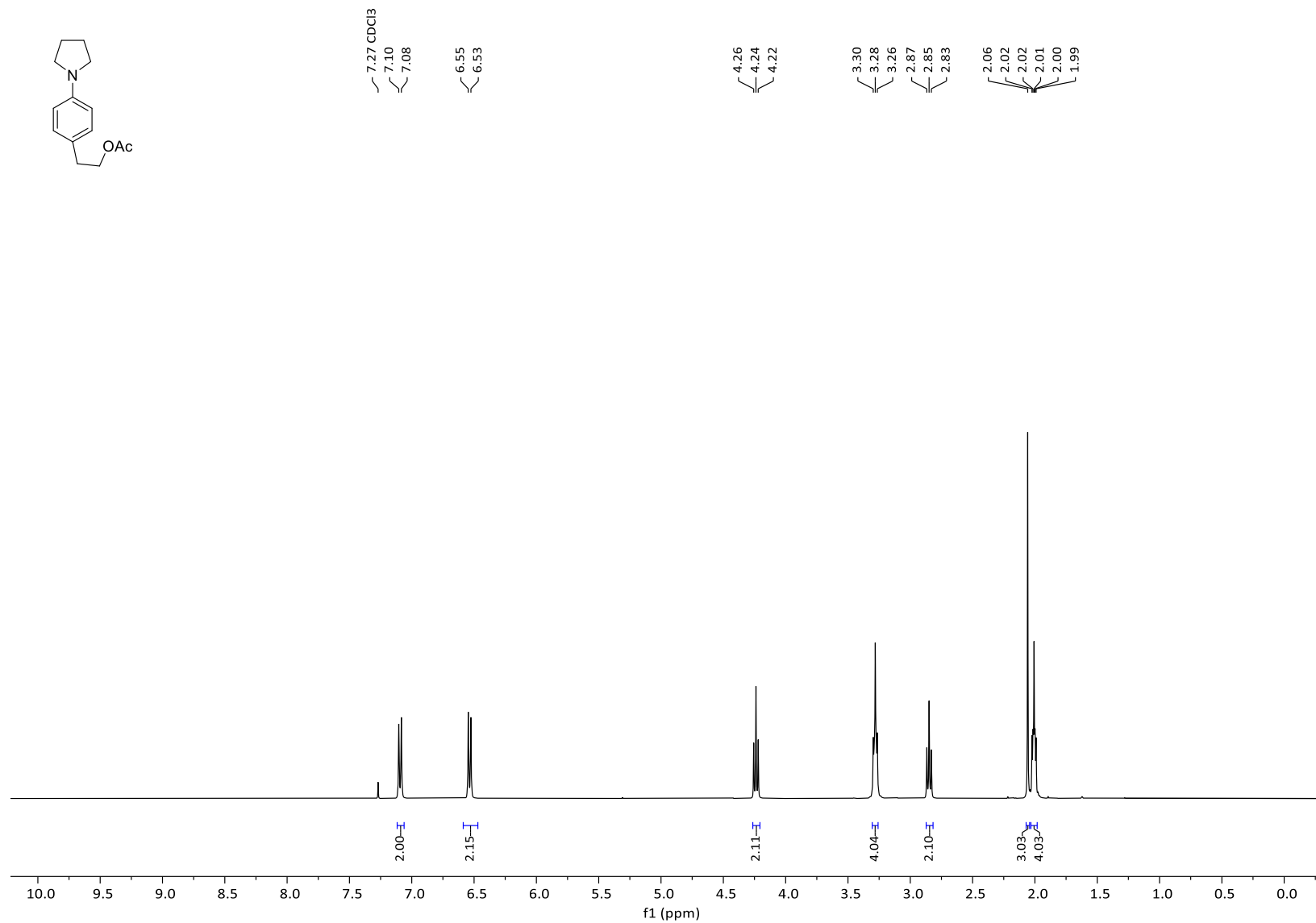
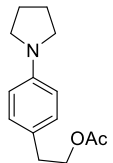
<sup>1</sup>H NMR spectrum of **18a** (600 MHz, CDCl<sub>3</sub>)



<sup>1</sup>H NMR spectrum of **19** (600 MHz, CDCl<sub>3</sub>)

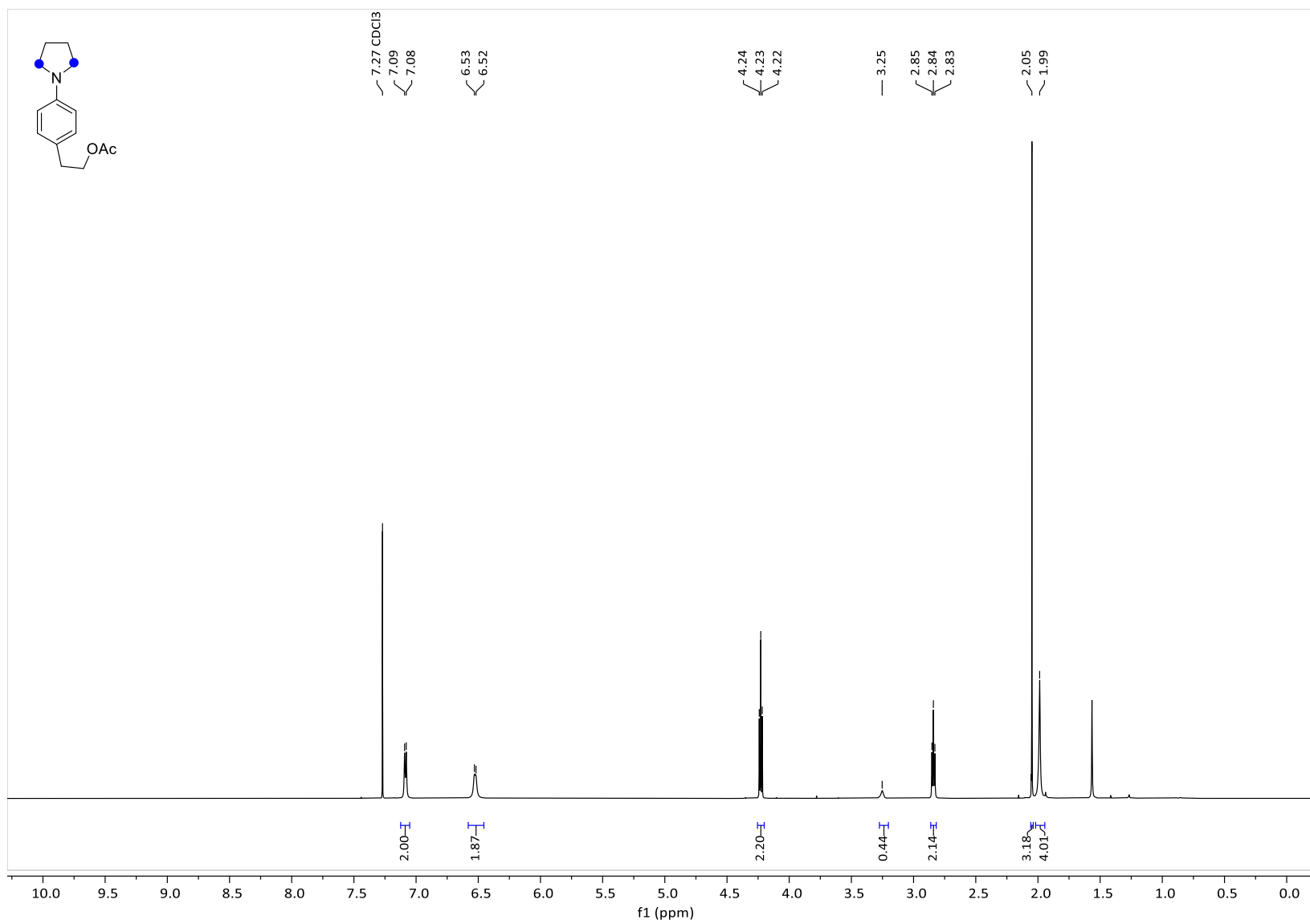


<sup>1</sup>H NMR spectrum of **19a** (600 MHz, CDCl<sub>3</sub>)

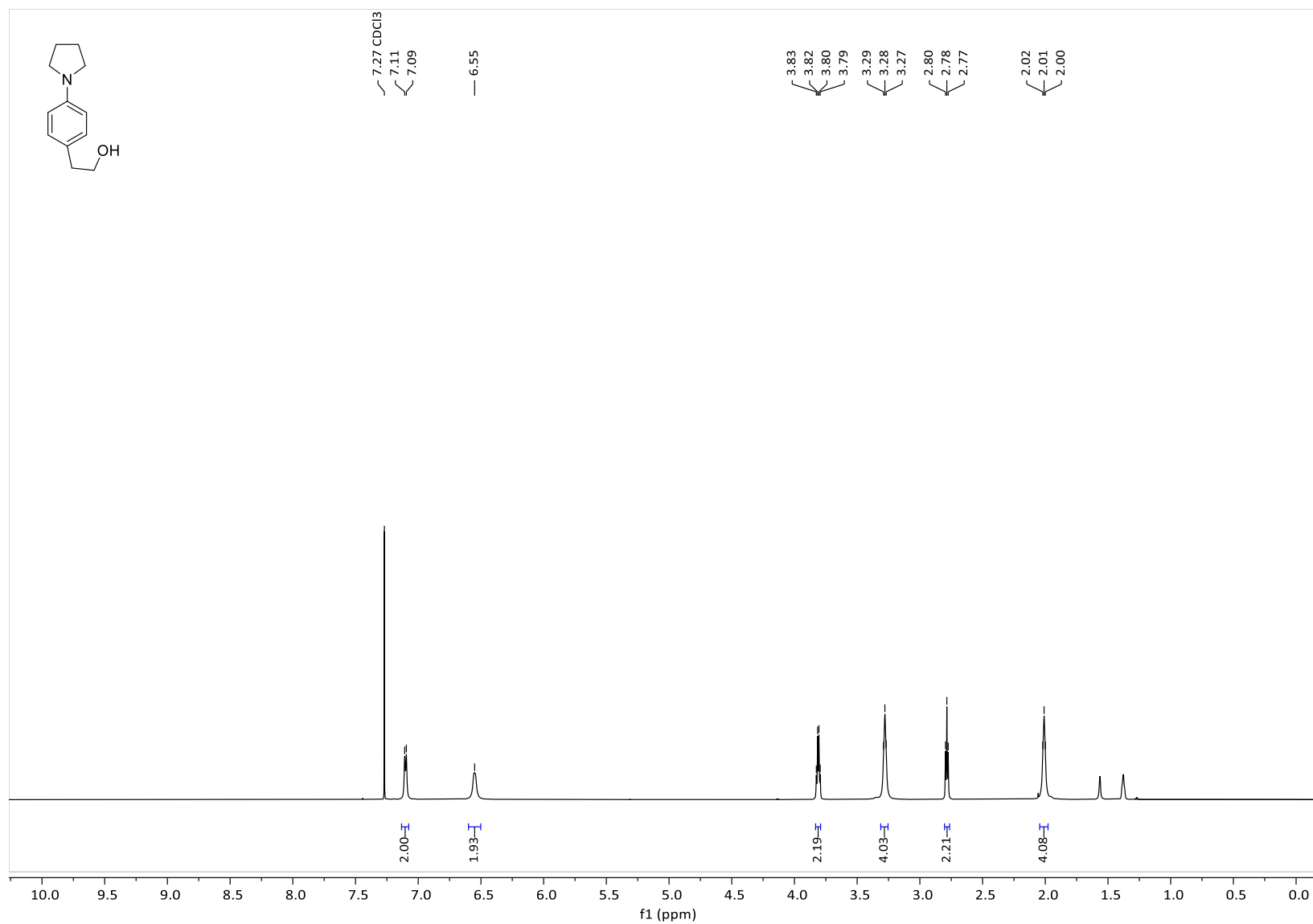


$^1\text{H}$  NMR spectrum of **20** (600 MHz,  $\text{CDCl}_3$ )

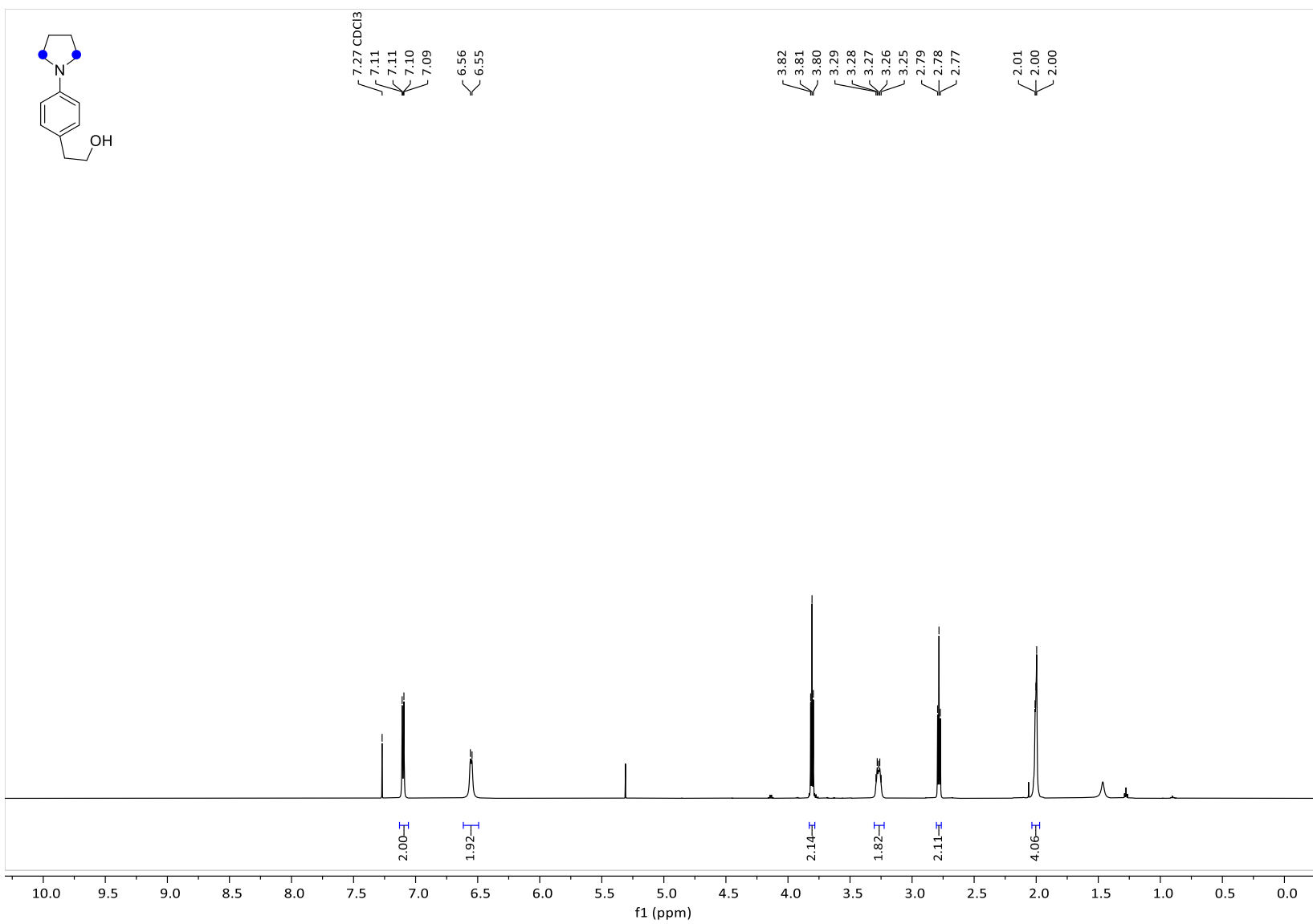




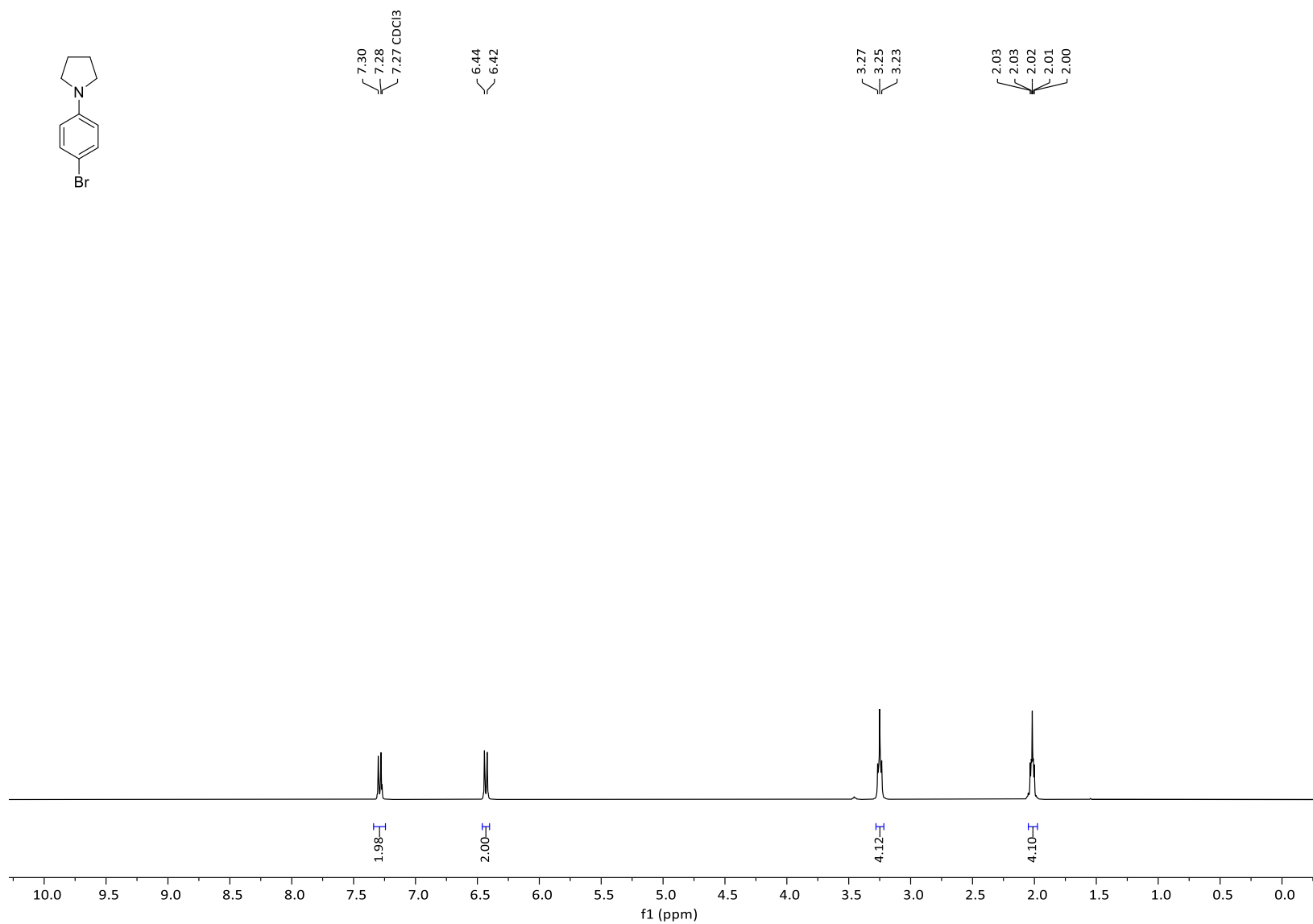
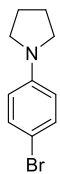
<sup>1</sup>H NMR spectrum of **20a** (600 MHz, CDCl<sub>3</sub>)



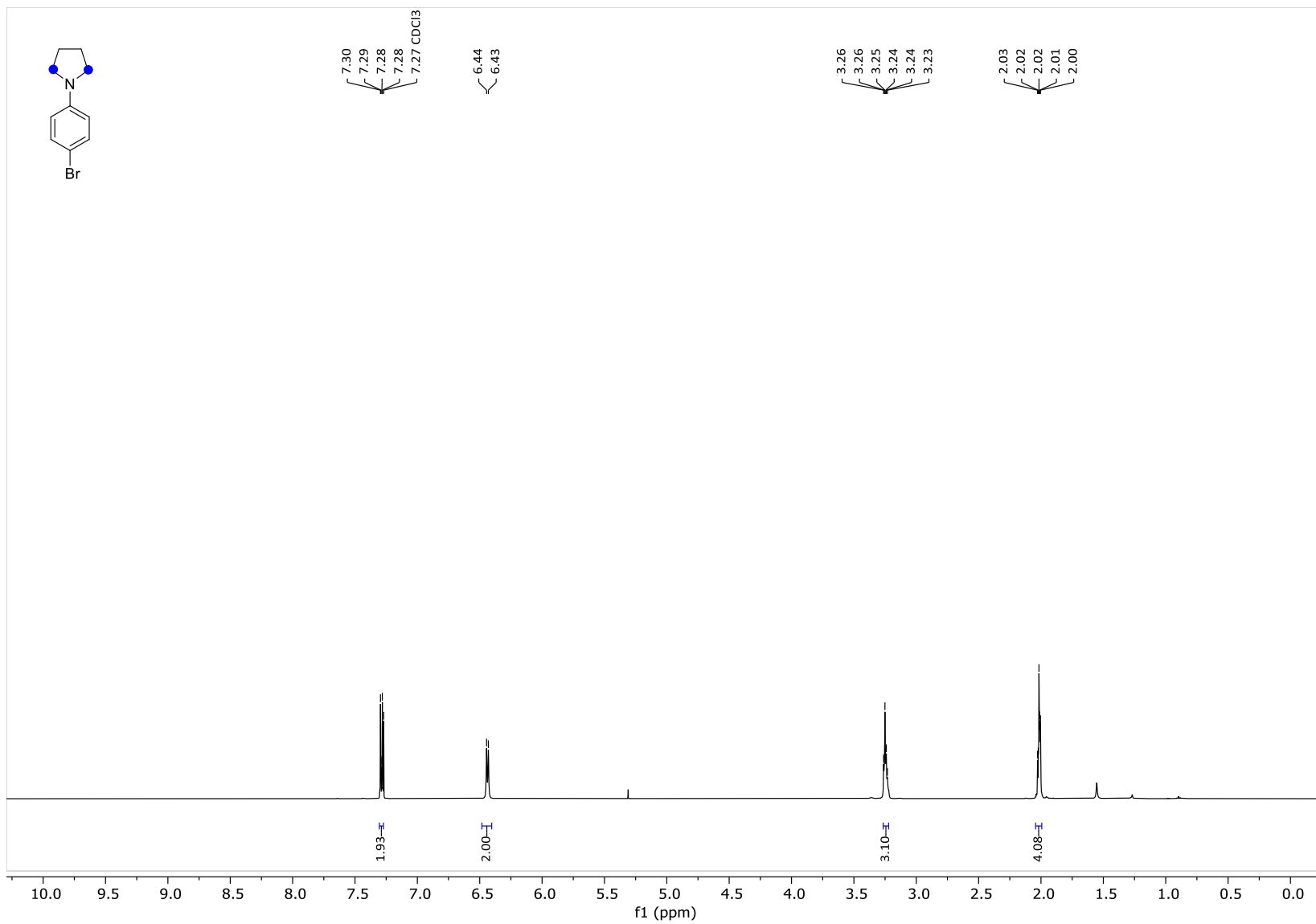
<sup>1</sup>H NMR spectrum of **21** (600 MHz, CDCl<sub>3</sub>)



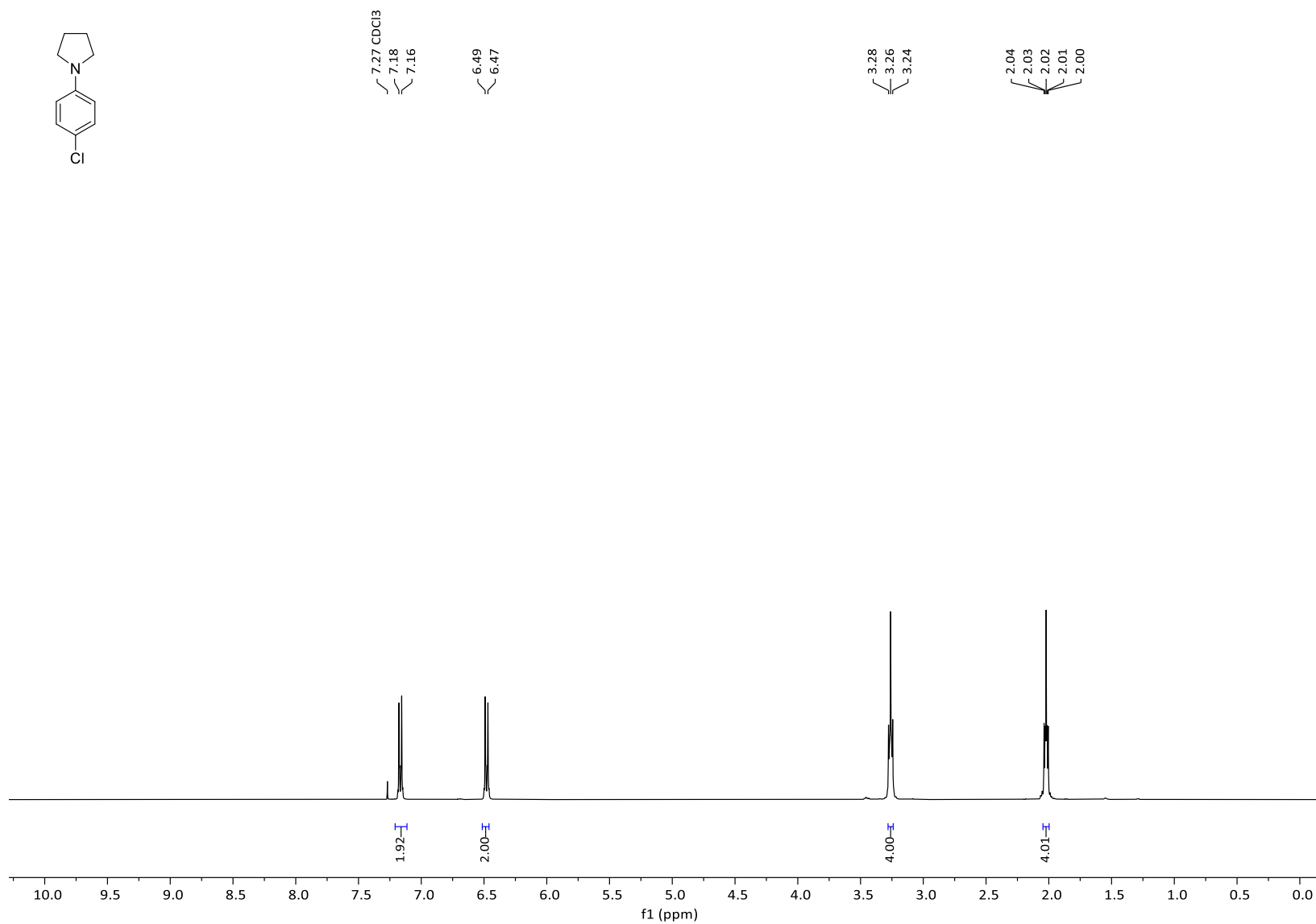
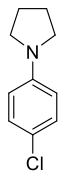
<sup>1</sup>H NMR spectrum of **21a** (600 MHz, CDCl<sub>3</sub>)



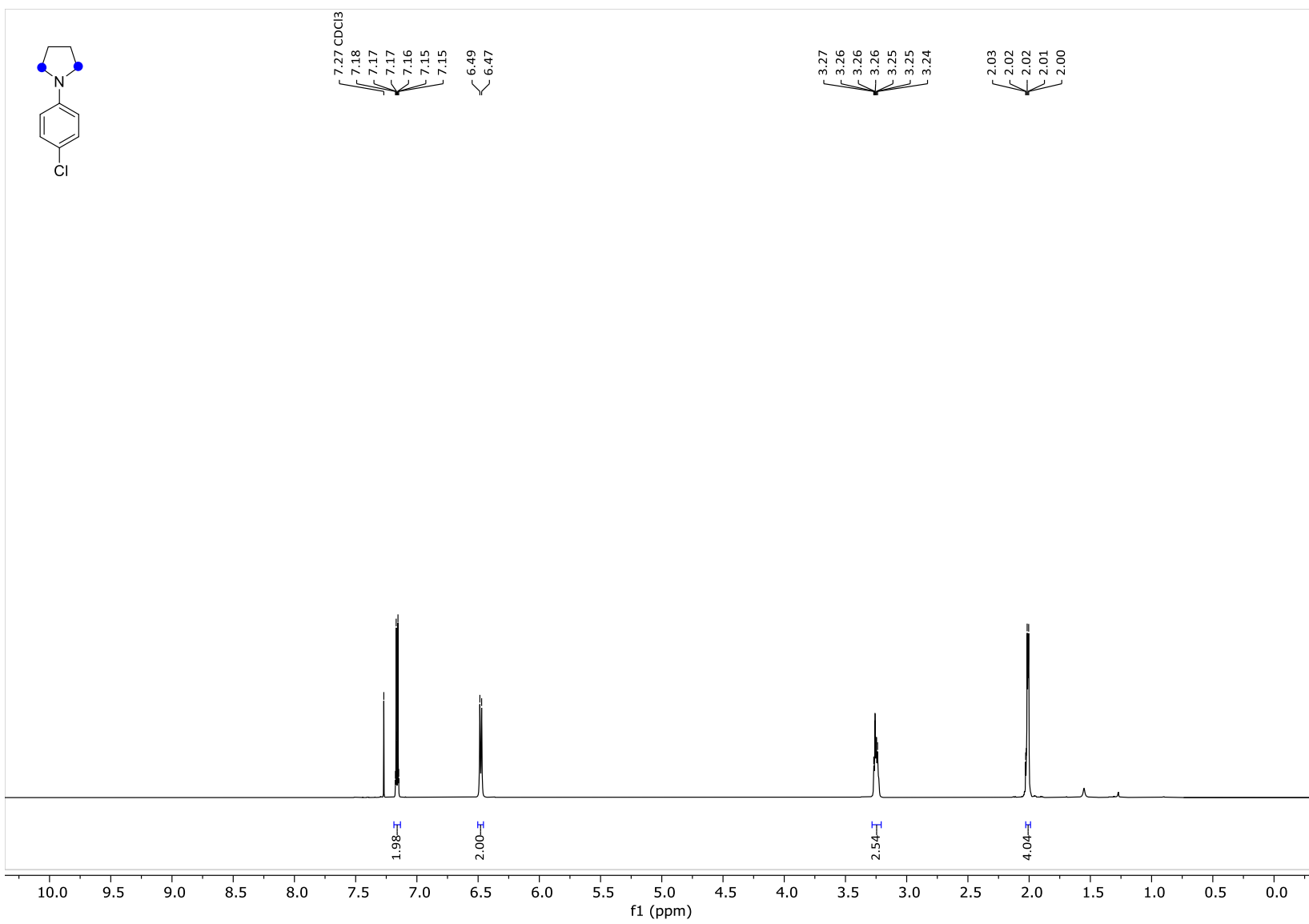
<sup>1</sup>H NMR spectrum of **22** (600 MHz, CDCl<sub>3</sub>)



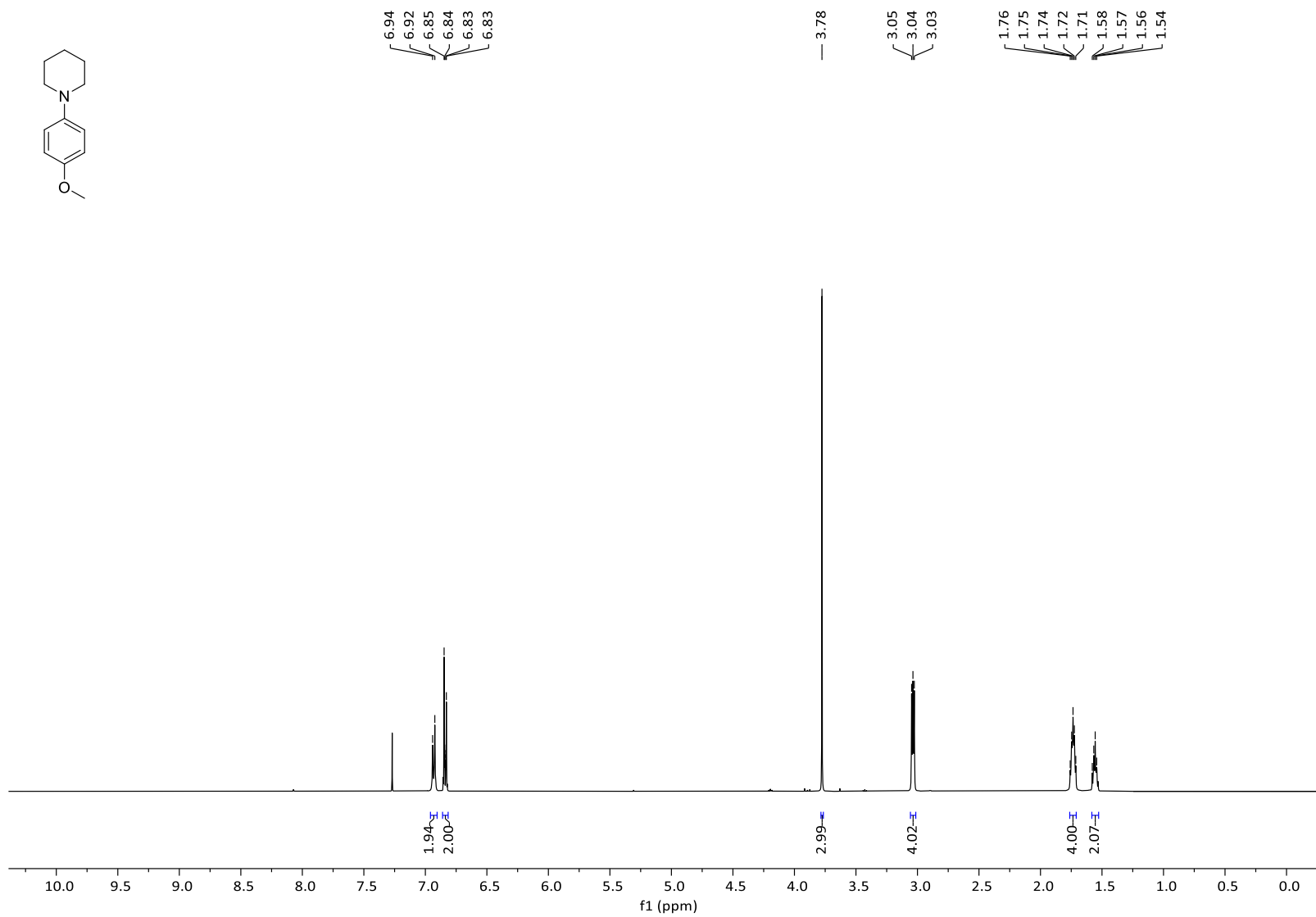
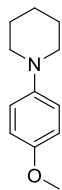
$^1\text{H}$  NMR spectrum of **22a** (600 MHz,  $\text{CDCl}_3$ )



<sup>1</sup>H NMR spectrum of **23** (600 MHz, CDCl<sub>3</sub>)

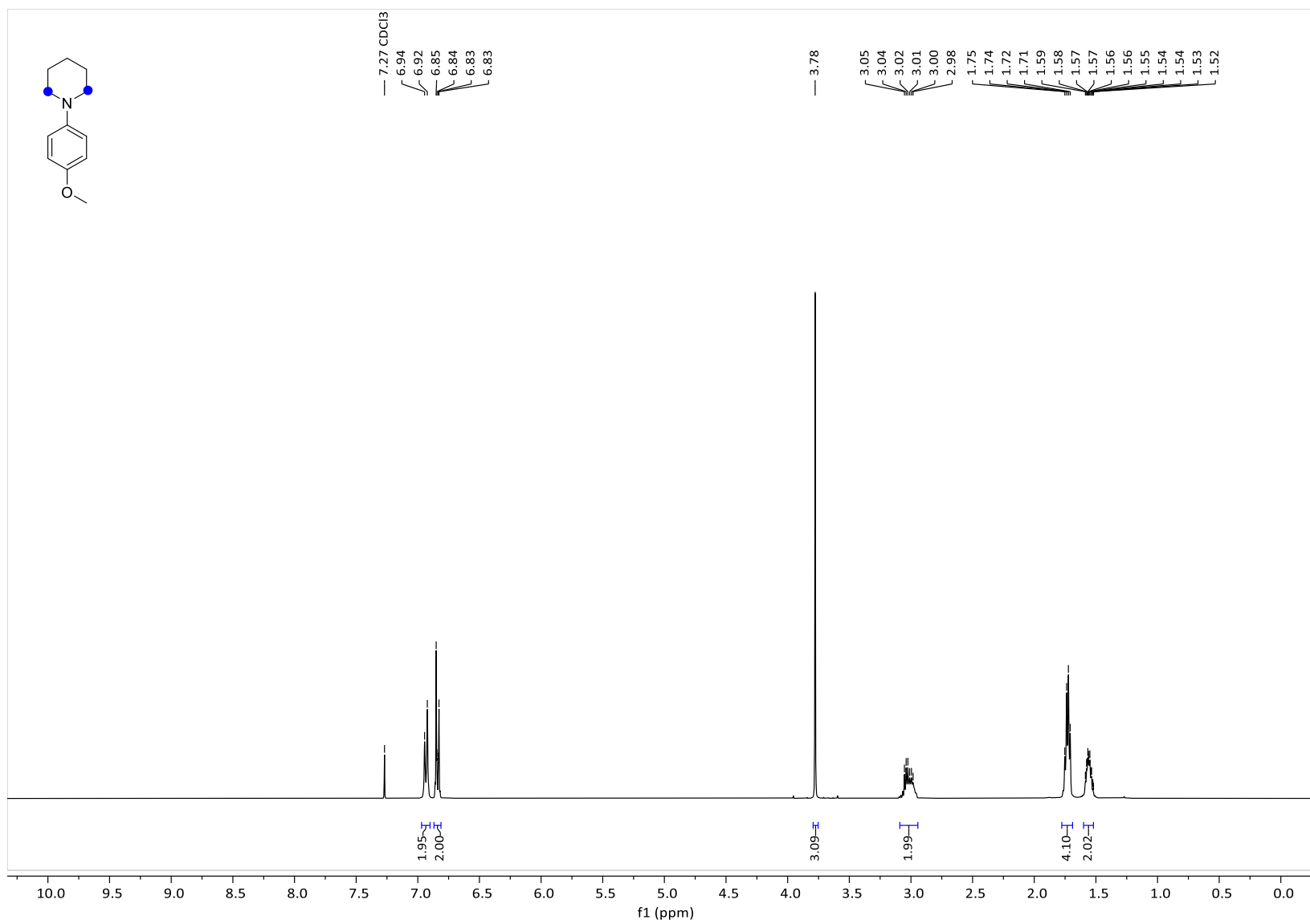


<sup>1</sup>H NMR spectrum of **23a** (600 MHz, CDCl<sub>3</sub>)



$^1\text{H}$  NMR spectrum of **24** (600 MHz,  $\text{CDCl}_3$ )

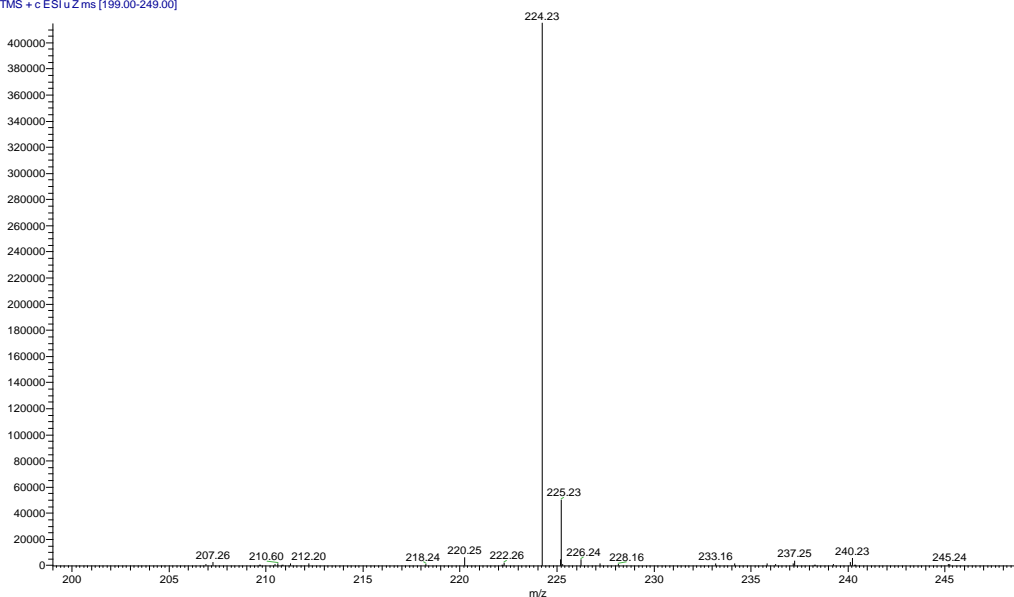




<sup>1</sup>H NMR spectrum of **24a** (600 MHz, CDCl<sub>3</sub>)

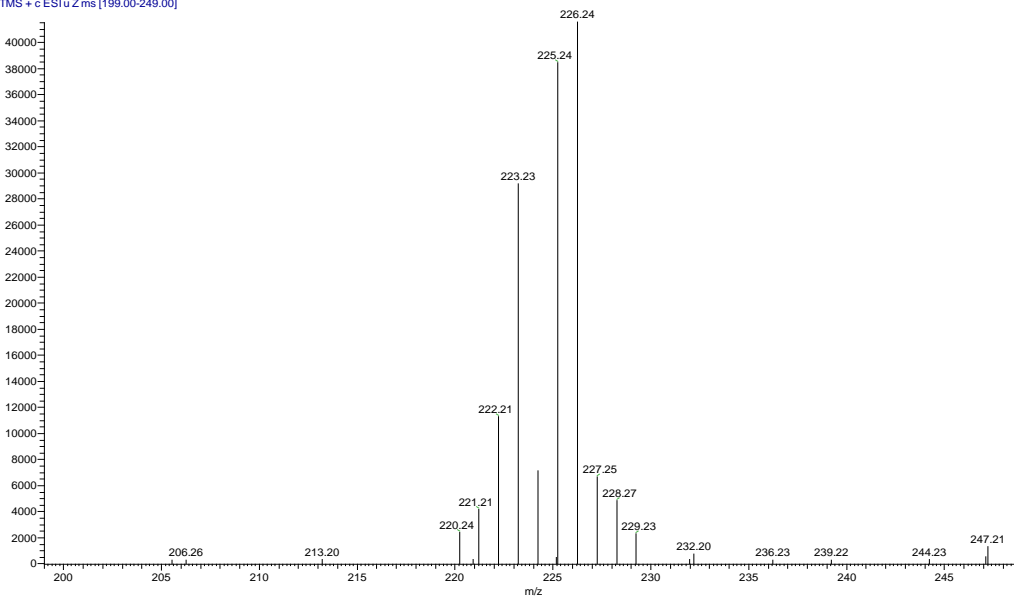
# Appendix B. Copies of LC-MS raw data and fitting results

NB-01-25-SM\_esipos #9 RT: 0.53 AV: 1 NL: 4.15E5  
T: ITMS + c ESI u Z ms [199.00-249.00]

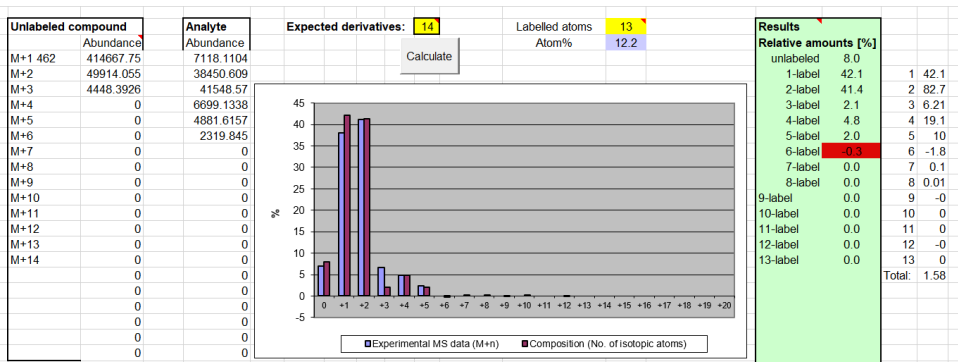


Mass spectrum of 1.

NB-01-25R-D-2\_esipos #9 RT: 0.46 AV: 1 NL: 4.15E4  
T: ITMS + c ESI u Z ms [199.00-249.00]

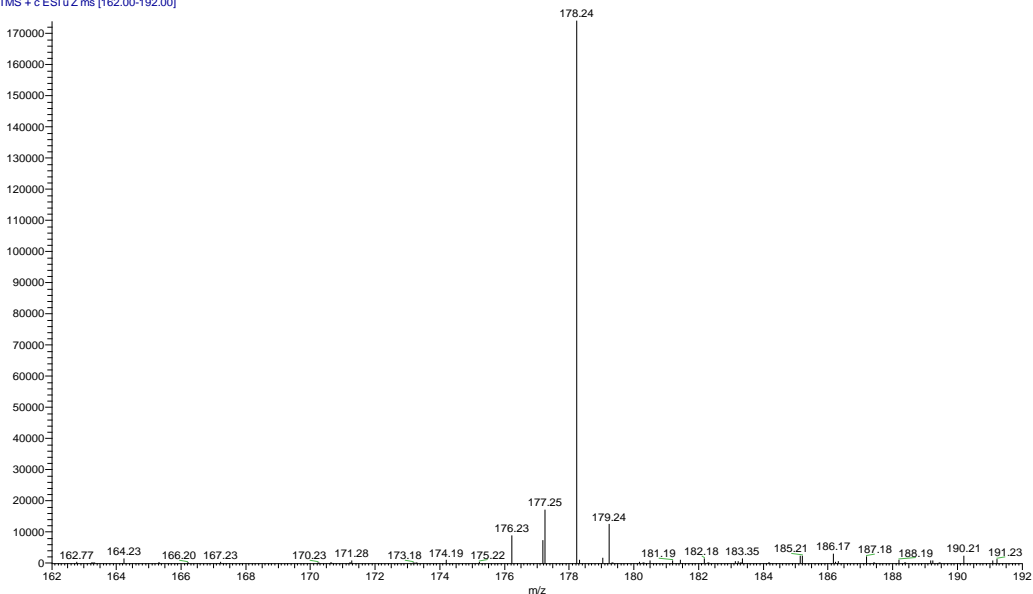


Mass spectrum of 1a.



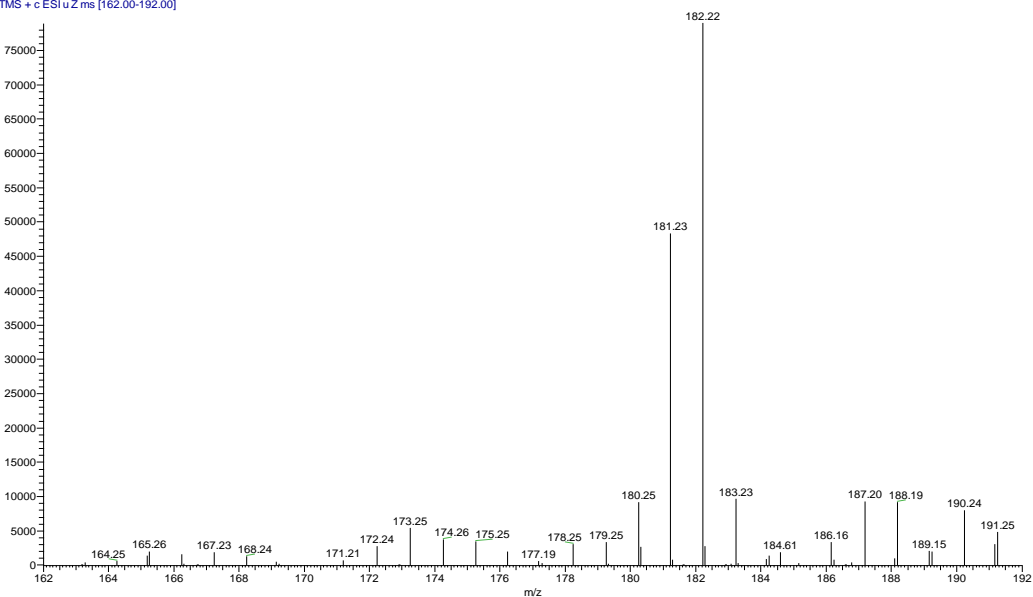
Mass spectrum and isopat analysis of 1a.

DG-03-145-SM\_230213170655 #12 RT: 0.47 AV: 1 NL: 1.74E!  
T: FTMS + c ESI u Z ms [162.00-192.00]

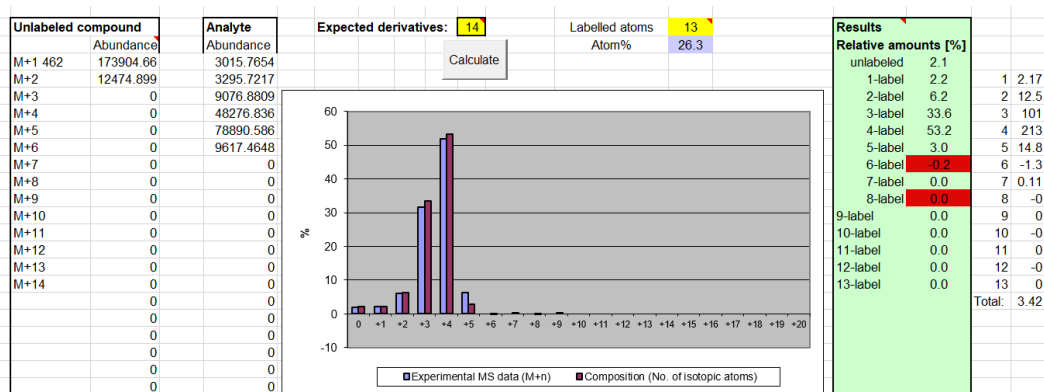


Mass spectrum of 2.

DG-03-145-D\_230213170655 #11 RT: 0.43 AV: 1 NL: 7.89E4  
T: FTMS + c ESI u Z ms [162.00-192.00]

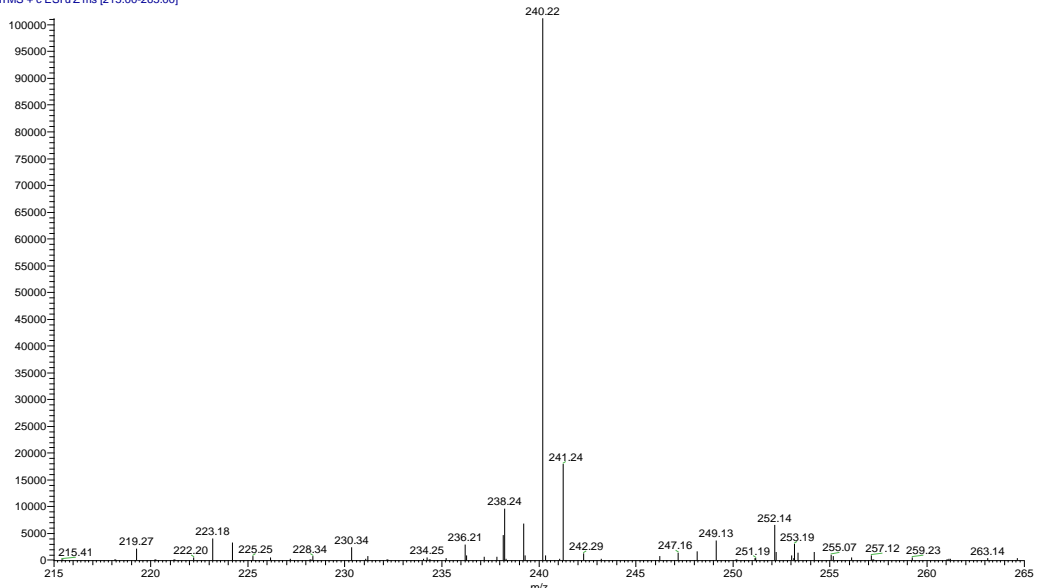


Mass spectrum of 2a.



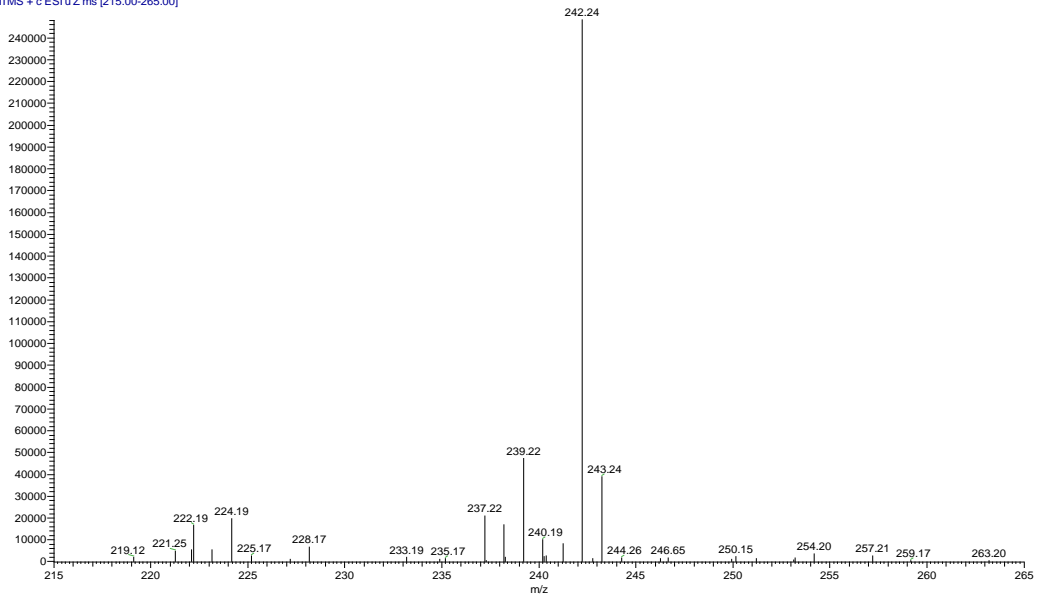
Mass spectrum and isotop analysis of 2a.

NB-01-20-SM esipos #5 RT: 0.26 AV: 1 NL: 1.01E5  
T: ITMS + c ESI u Z ms [215.00-265.00]

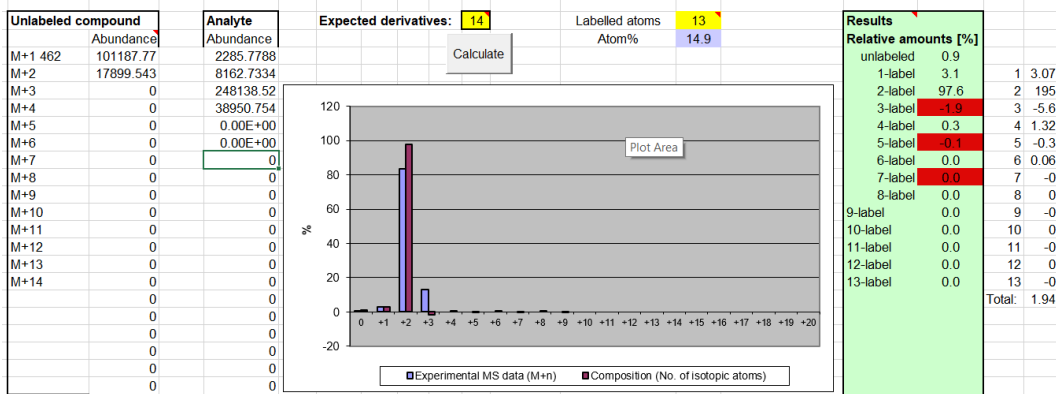


Mass spectrum of 3.

NB-01-20-D esipos #6 RT: 0.33 AV: 1 NL: 2.48E5  
T: ITMS + c ESI u Z ms [215.00-265.00]

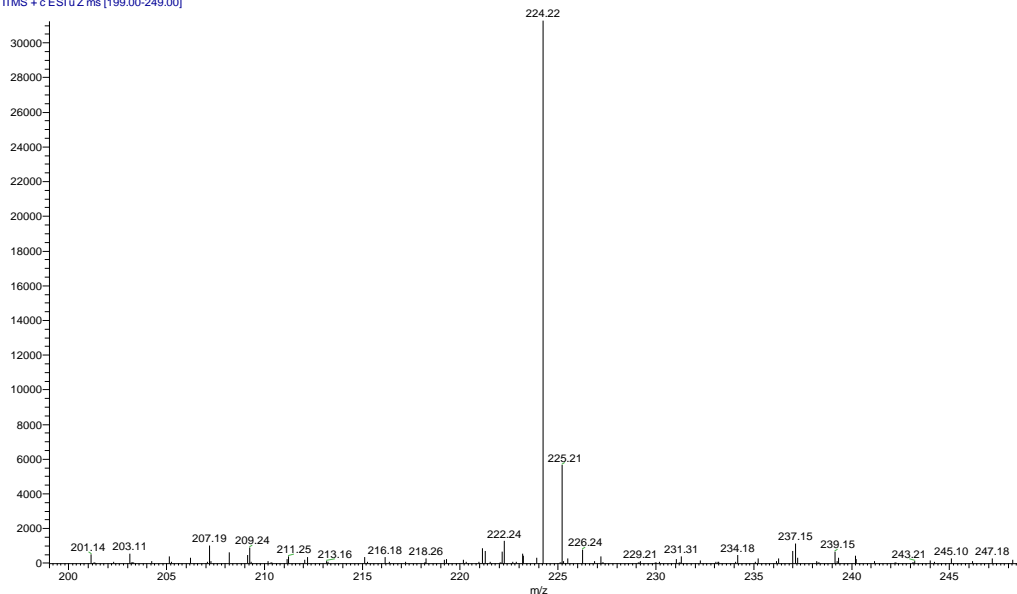


Mass spectrum of 3a.



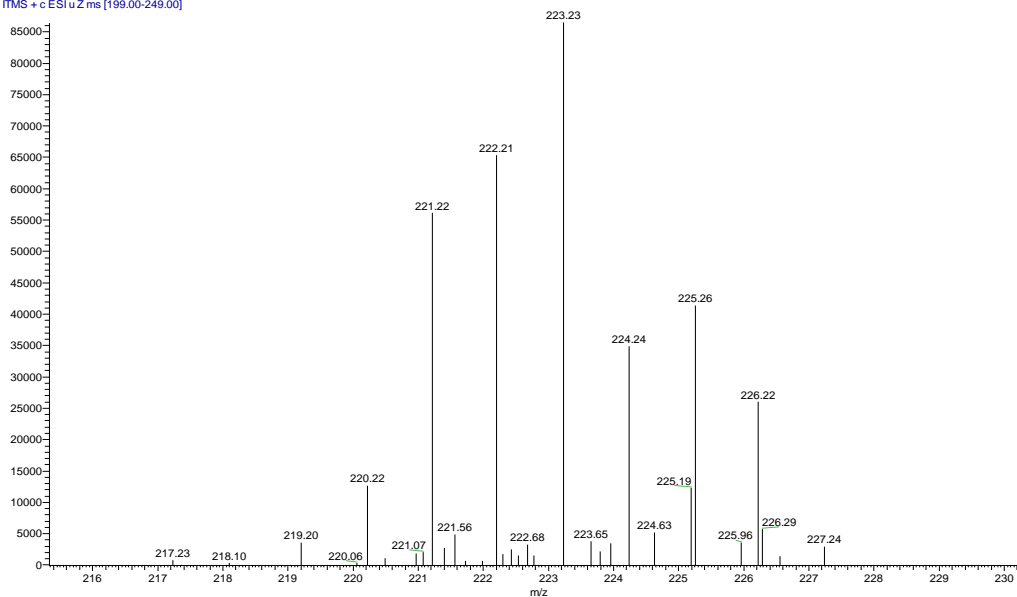
Mass spectrum and isotopic analysis of 3a.

NB-01-72-SM esipos #7 RT: 0.40 AV: 1 NL: 3.12E4  
T: ITMS + c ESI u Z ms [199.00-249.00]

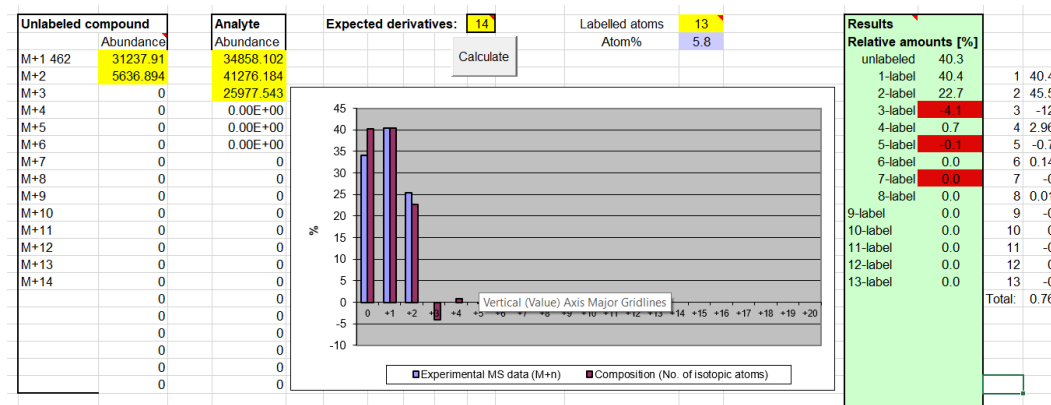


Mass spectrum of 4.

NB-01-72R-D esipos #5 RT: 0.26 AV: 1 NL: 8.64E4  
T: ITMS + c ESI u Z ms [199.00-249.00]

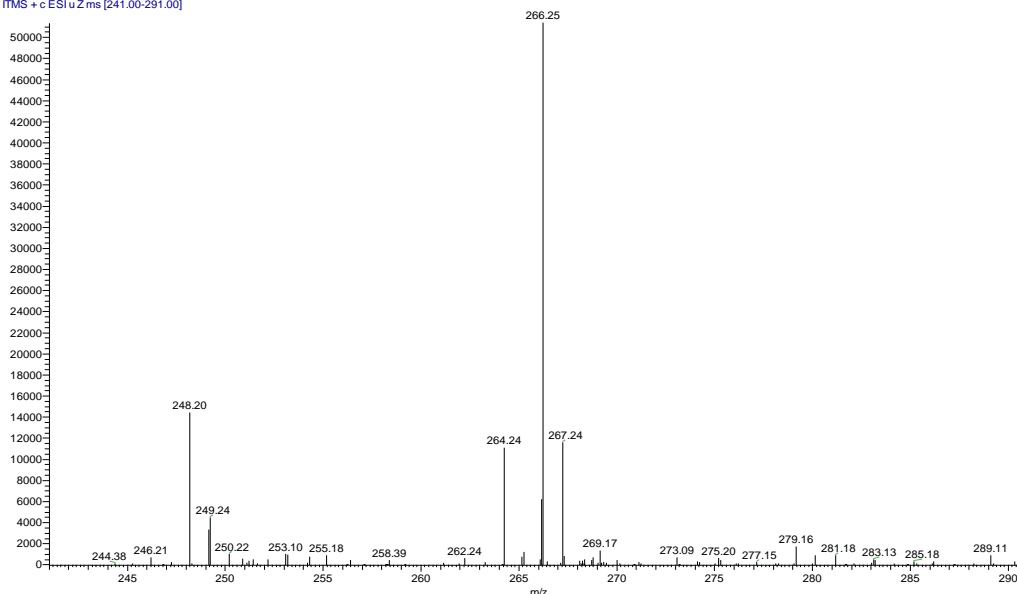


Mass spectrum of 4a.



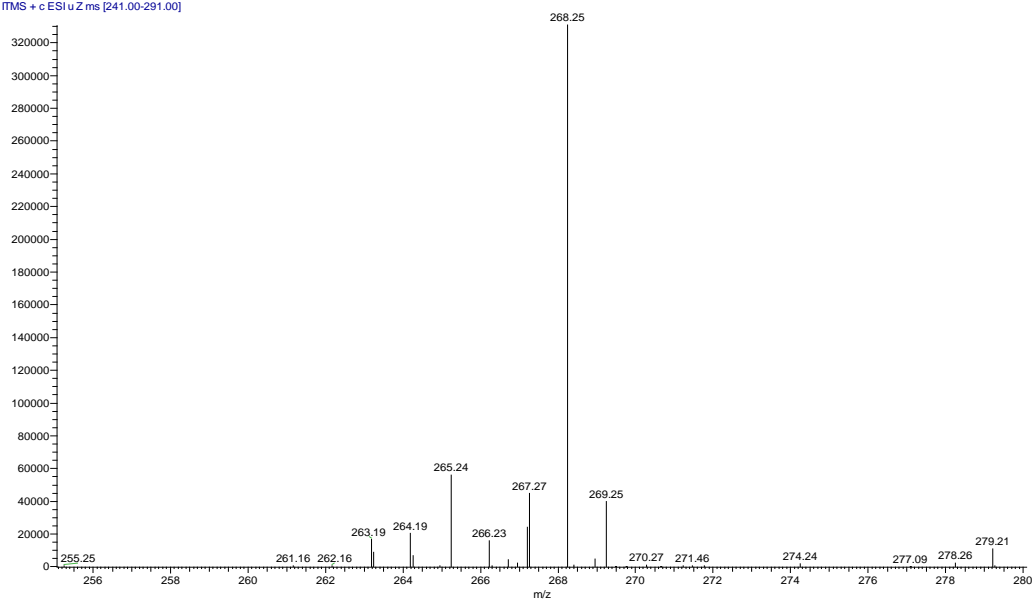
Mass spectrum and isotop analysis of 4a.

NB-01-66-SM esipos #7 RT: 0.40 AV: 1 NL: 5.13E4  
T: ITMS + c ESI u Z ms [241.00-291.00]

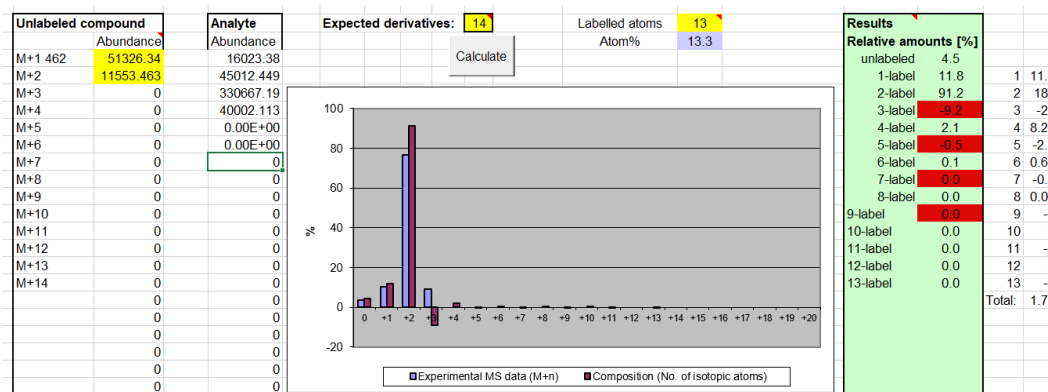


Mass spectrum of 5.

NB-01-66-D esipos #5 RT: 0.26 AV: 1 NL: 3.31E5  
T: ITMS + c ESI u Z ms [241.00-291.00]

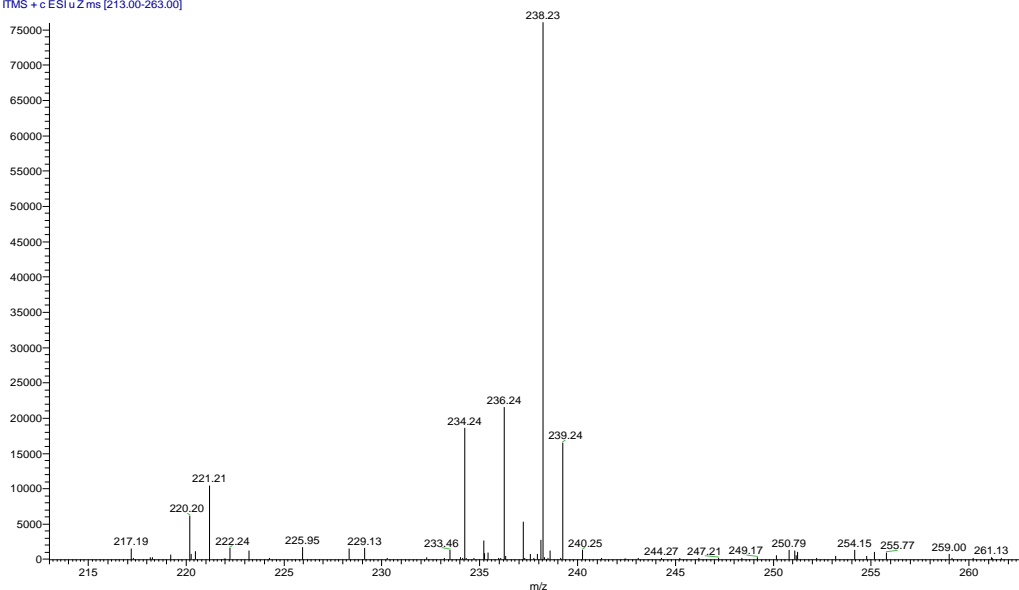


Mass spectrum of 5a.



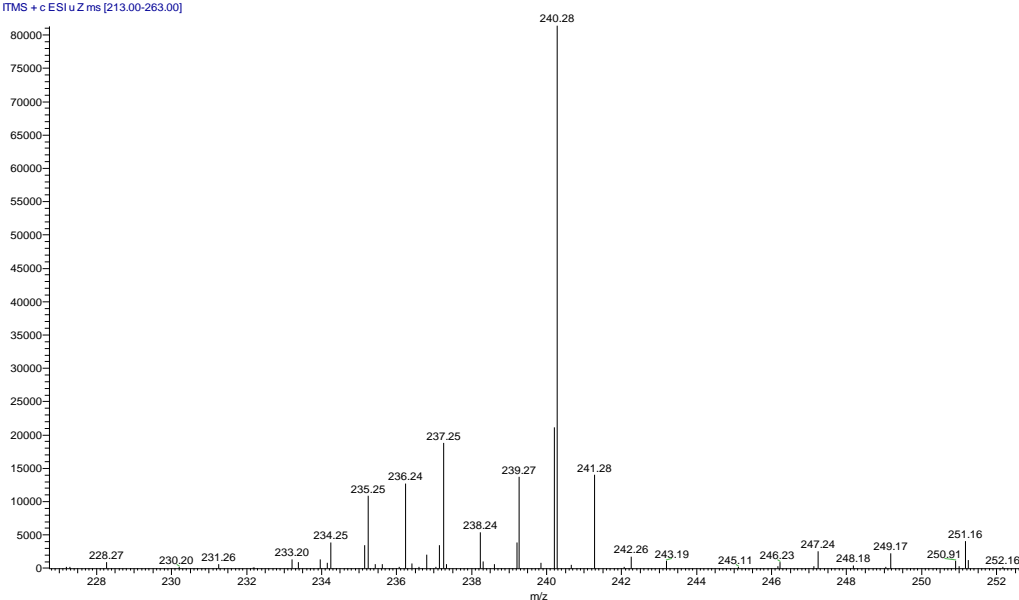
Mass spectrum and isotop analysis of 5a.

NB-01-21-SM esipos #7 RT: 0.40 AV: 1 NL: 7.60E4  
T: ITMS + c ESI u Z ms [213.00-263.00]

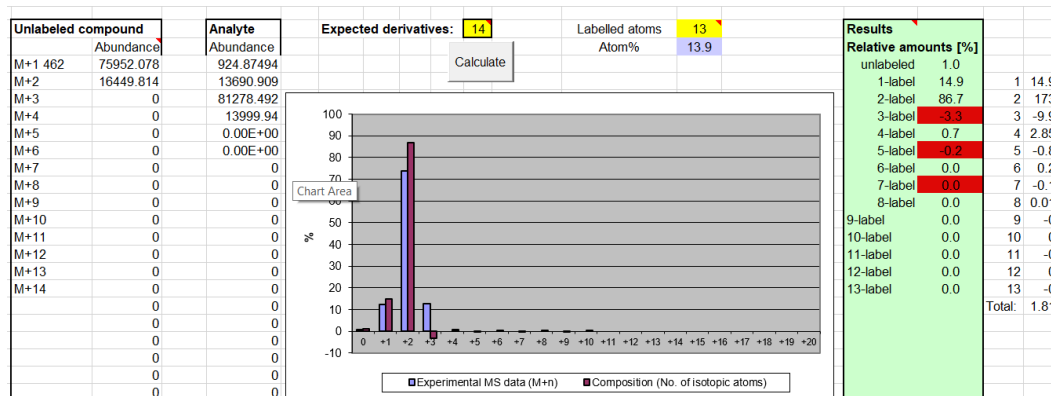


Mass spectrum of 6.

NB-01-21-D esipos #6 RT: 0.33 AV: 1 NL: 8.13E4  
T: ITMS + c ESI u Z ms [213.00-263.00]

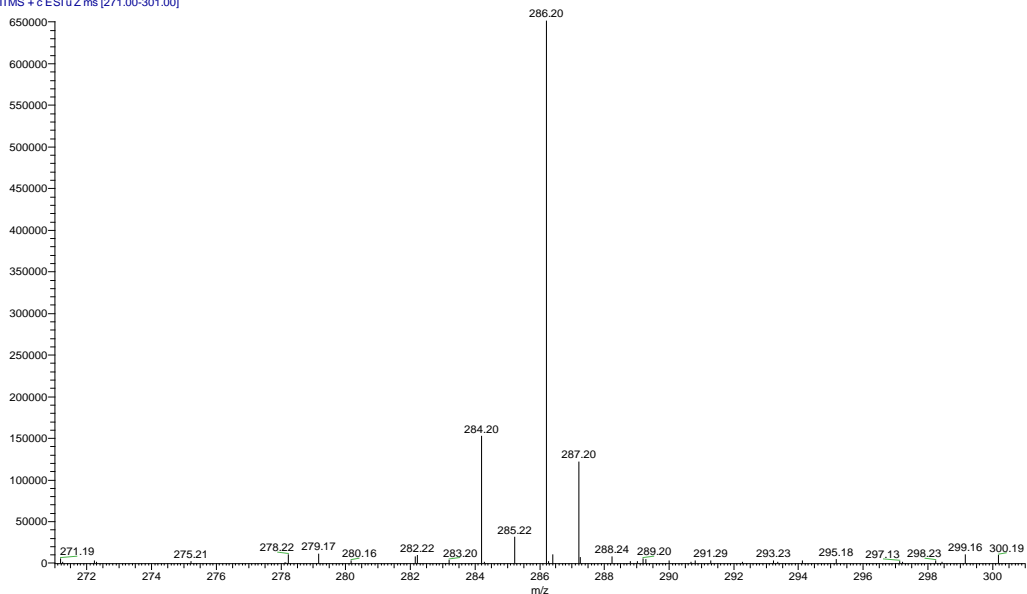


Mass spectrum of 6a.



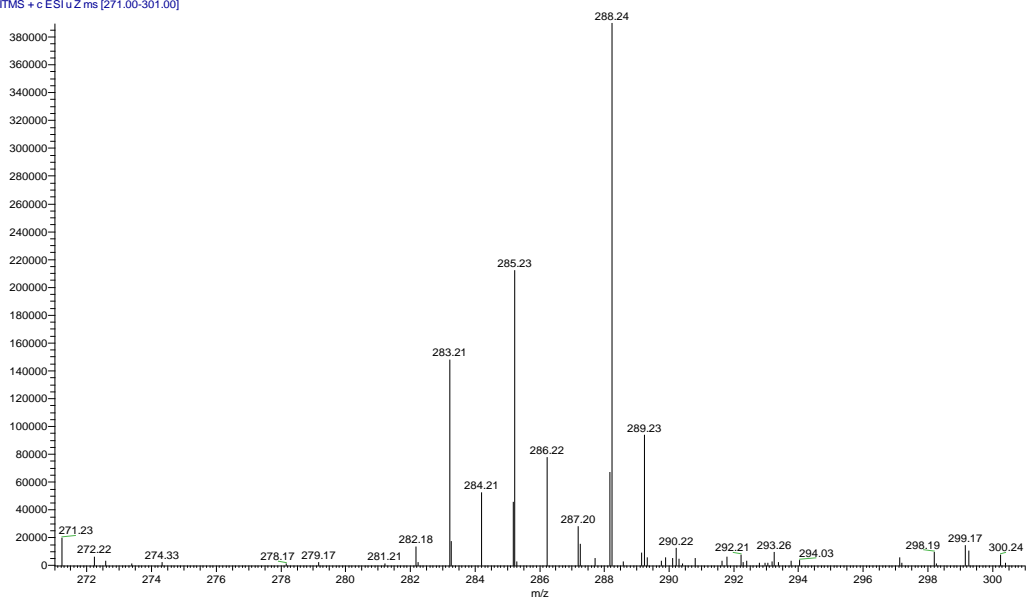
Mass spectrum and isotop analysis of 6a.

NB-01-32-SM esipos #8 RT: 0.30 AV: 1 NL: 6.51E5  
T: ITMS + c ESI u z ms [271.00-301.00]

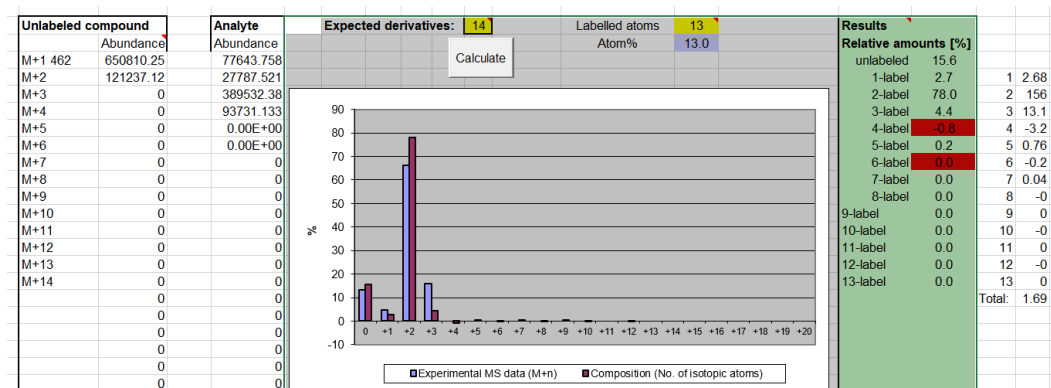


Mass spectrum of 7.

NB-01-32-D esipos #9 RT: 0.34 AV: 1 NL: 3.90E5  
T: ITMS + c ESI u z ms [271.00-301.00]



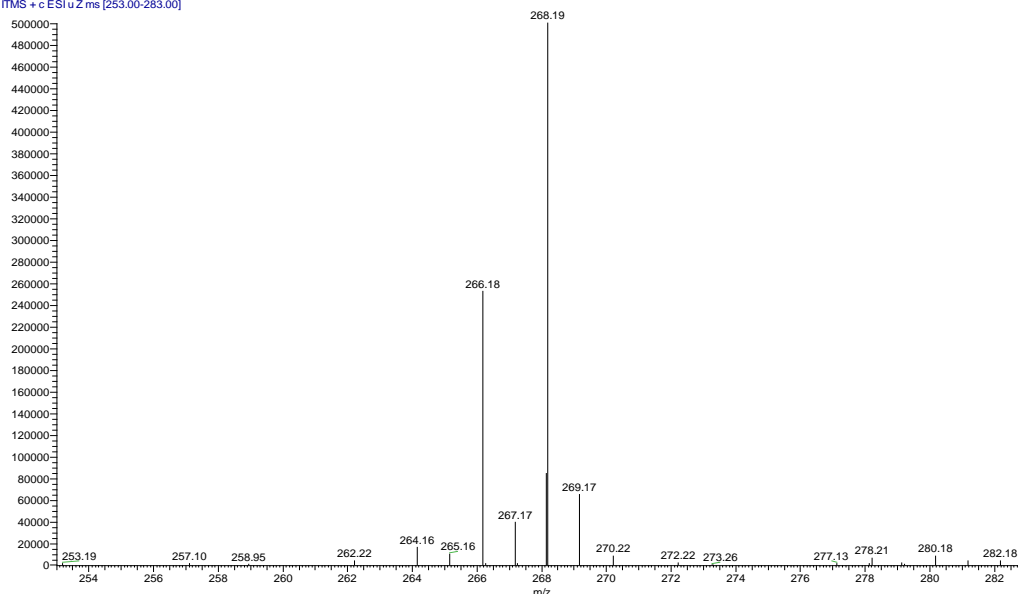
Mass spectrum of 7a.



Mass spectrum and isotop analysis of 7a.

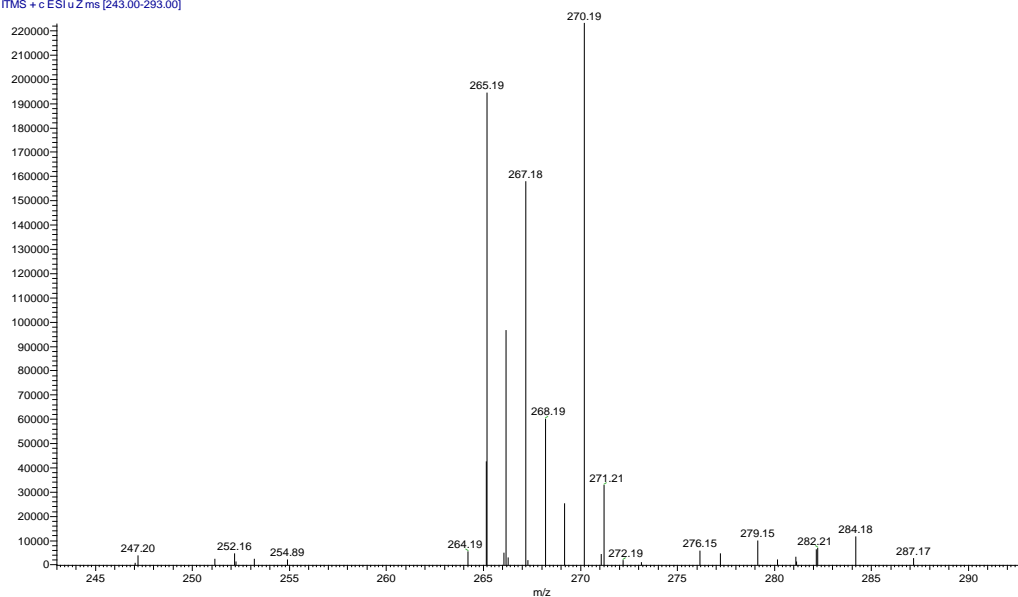


NB-01-31-SM\_230213163712 #8 RT: 0.30 AV: 1 NL: 5.00E5  
T: ITMS + c ESI u Z ms [253.00-283.00]

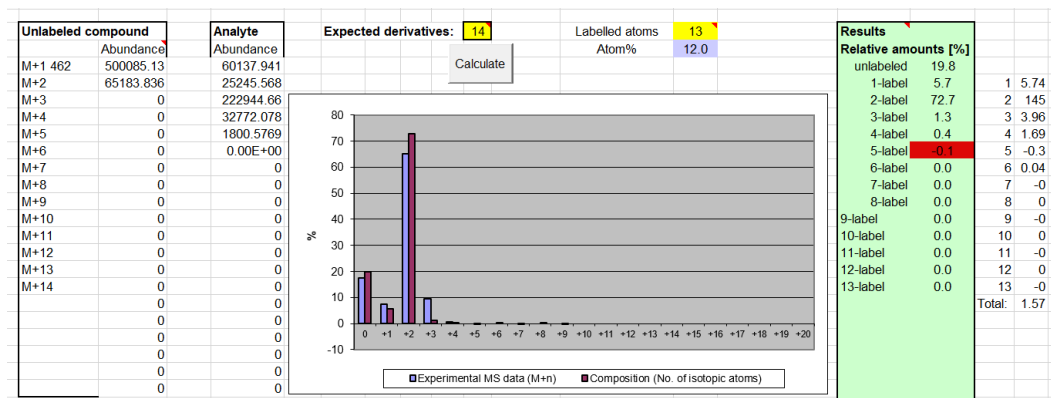


Mass spectrum of 8.

NB-01-31\_esipos #4 RT: 0.20 AV: 1 NL: 2.23E5  
T: ITMS + c ESI u Z ms [243.00-293.00]

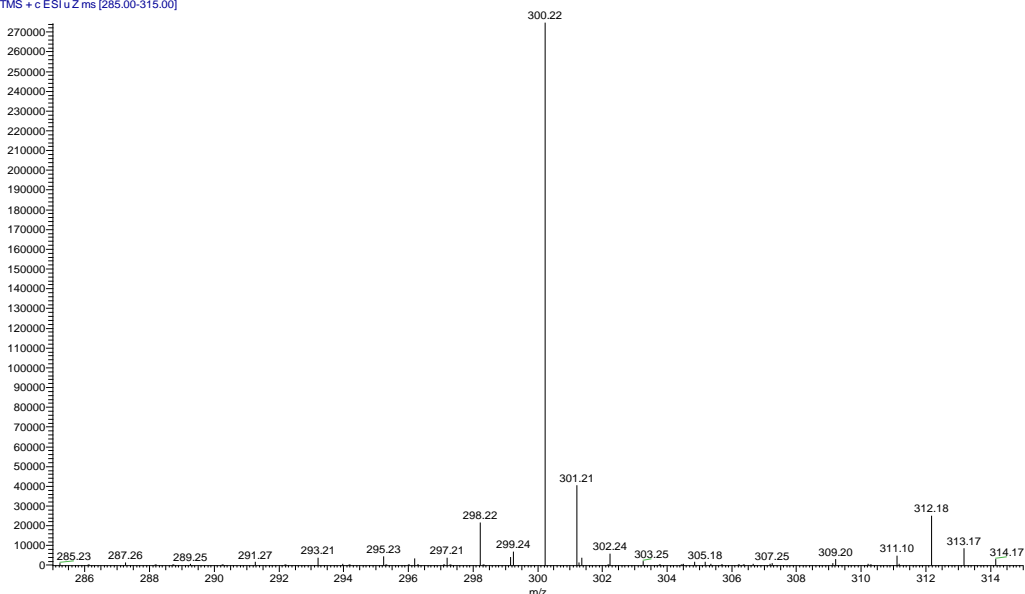


Mass spectrum of 8a.



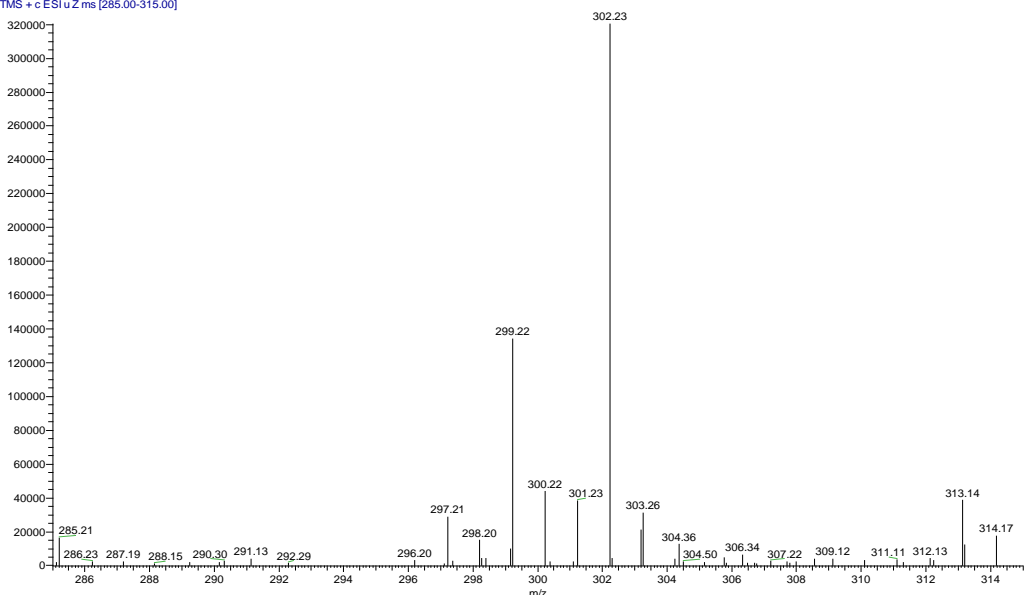
Mass spectrum and isotop analysis of 8a.

NB-01-29-SM esipos #9 RT: 0.34 AV: 1 NL: 2.74E5  
T: ITMS + c ESI u z ms [285.00-315.00]

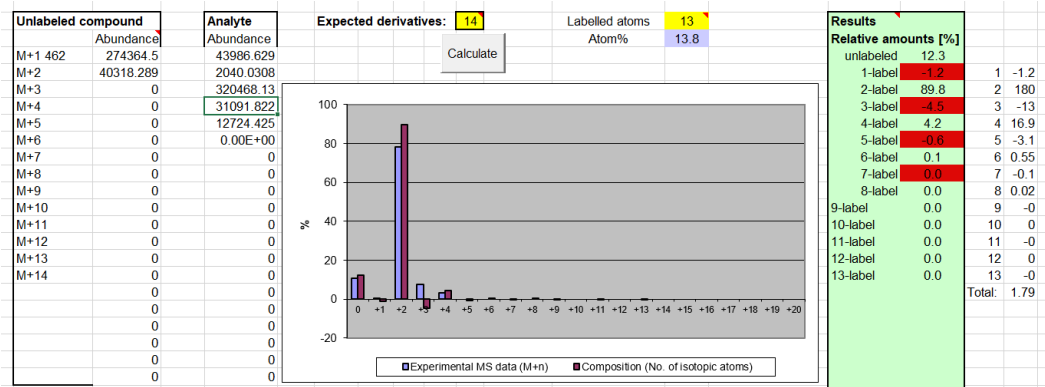


Mass spectrum of 9.

NB-01-29-D esipos #8 RT: 0.29 AV: 1 NL: 3.20E5  
T: ITMS + c ESI u z ms [285.00-315.00]

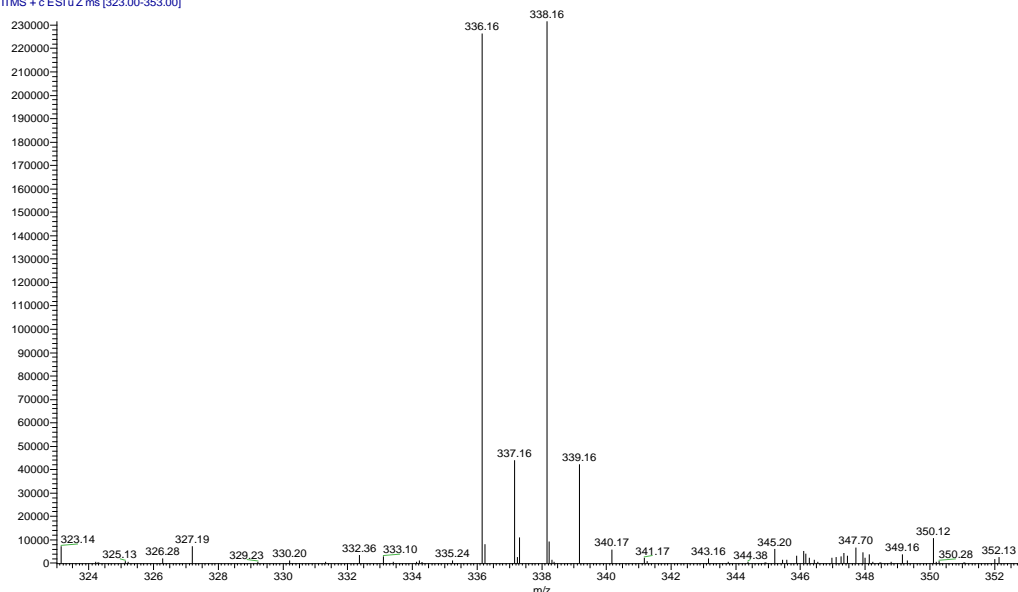


Mass spectrum of 9a.



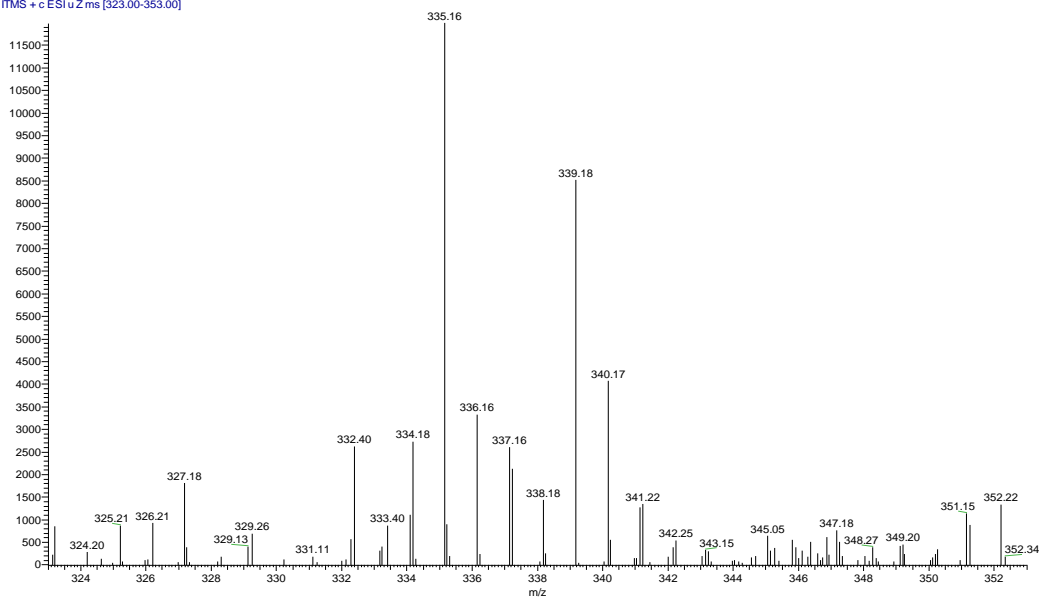
Mass spectrum and isotopic analysis of 9a.

NB-01-48-SM\_230213163712 #8 RT: 0.29 AV: 1 NL: 2.32E5  
T: ITMS + c ESI u Z ms [323.00-353.00]

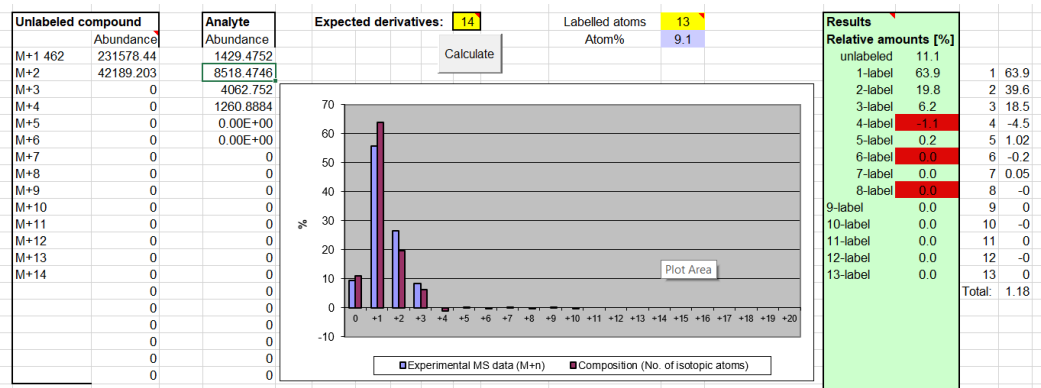


Mass spectrum of 10.

NB-01-48R-17h-D2\_230213170655 #13 RT: 0.50 AV: 1 NL: 1.1  
T: ITMS + c ESI u Z ms [323.00-353.00]

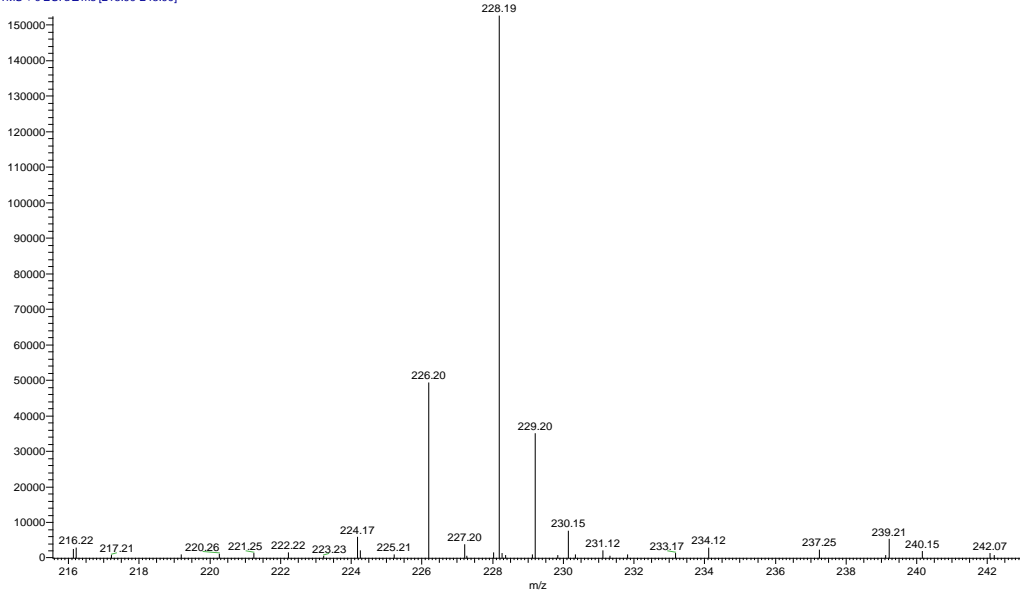


Mass spectrum of 10a.



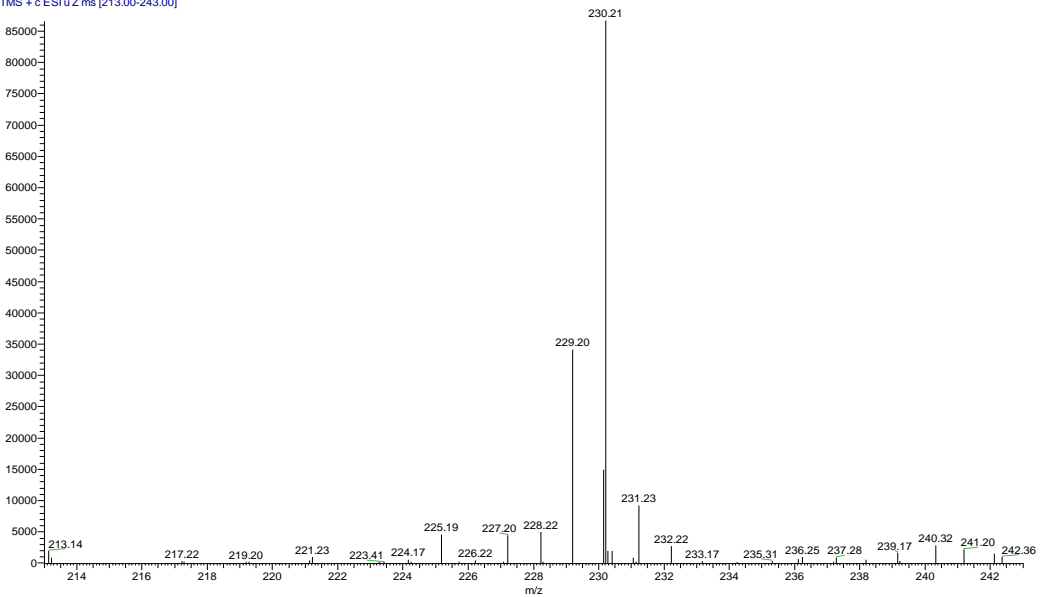
Mass spectrum and isopat analysis of 10a.

NB-01-47-SM 230213163712 #9 RT: 0.34 AV: 1 NL: 1.52E5  
T: ITMS + c ESI u Z ms [213.00-243.00]

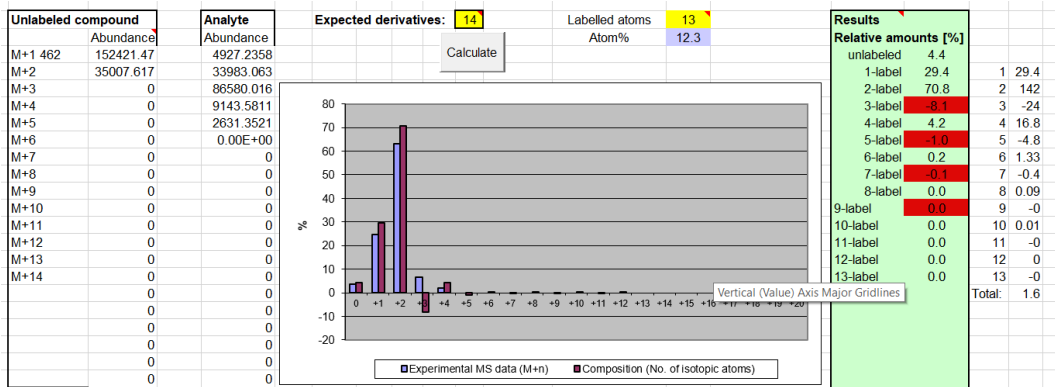


Mass spectrum of 11.

NB-01-47-R2-D 230213163712 #10 RT: 0.38 AV: 1 NL: 8.66E  
T: ITMS + c ESI u Z ms [213.00-243.00]

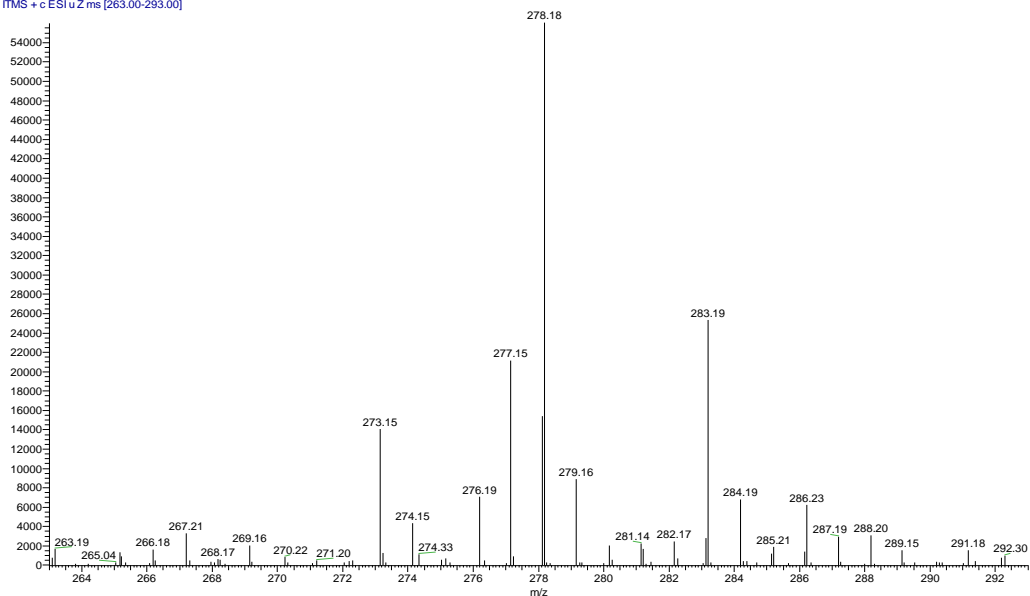


Mass spectrum of 11a.



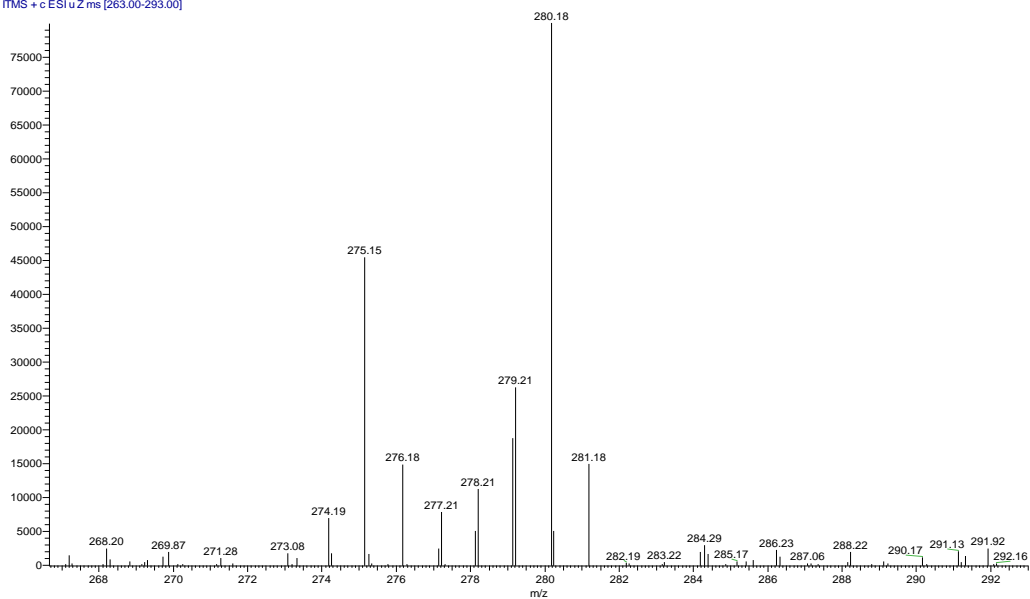
Mass spectrum and isotopic analysis of 11a.

NB-01-58-SM\_230213163712 #9 RT: 0.34 AV: 1 NL: 5.60E4  
T: ITMS + c ESI u Z ms [263.00-293.00]

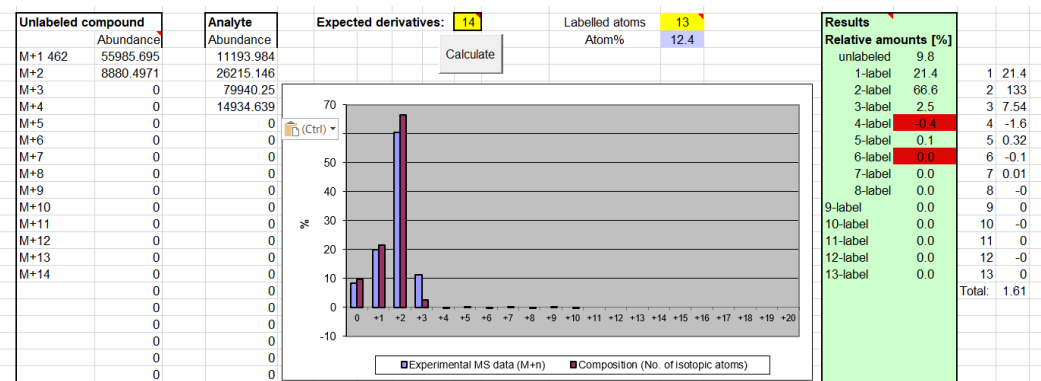


Mass spectrum of 12.

NB-01-58-D\_230213165658 #7 RT: 0.25 AV: 1 NL: 7.99E4  
T: ITMS + c ESI u Z ms [263.00-293.00]

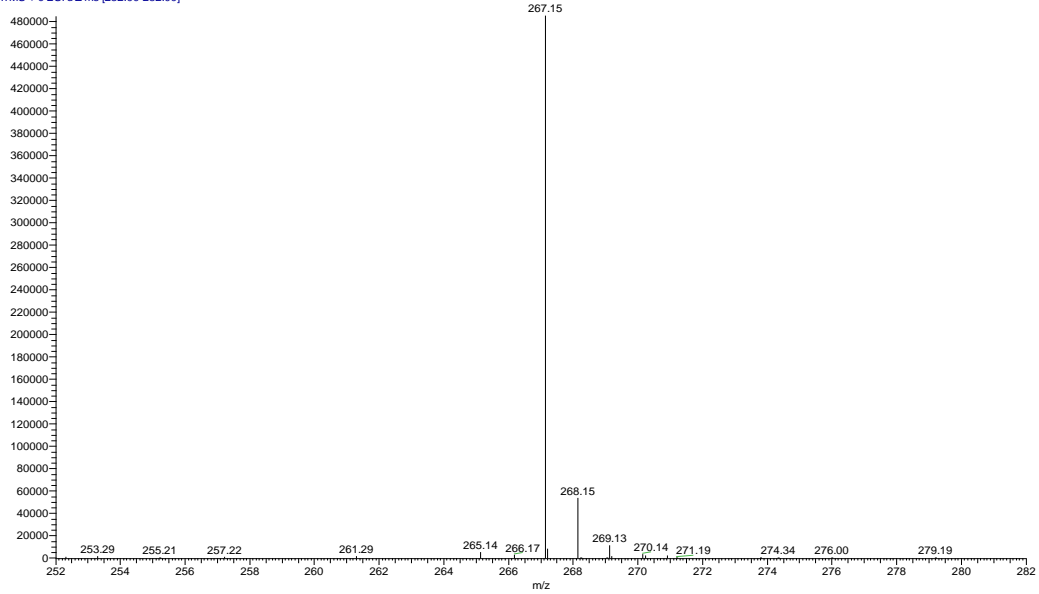


Mass spectrum of 12a.



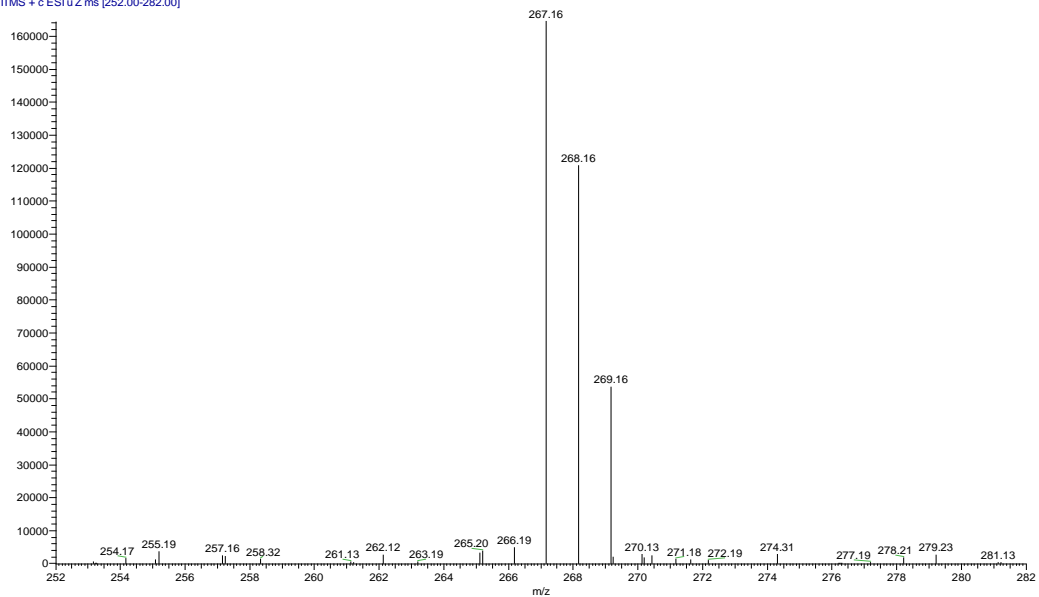
Mass spectrum and isopat analysis of 12a.

NB-01-62-SM\_230213165658 #8 RT: 0.30 AV: 1 NL: 4.85E5  
T: ITMS + c ESI u Z ms [252.00-282.00]

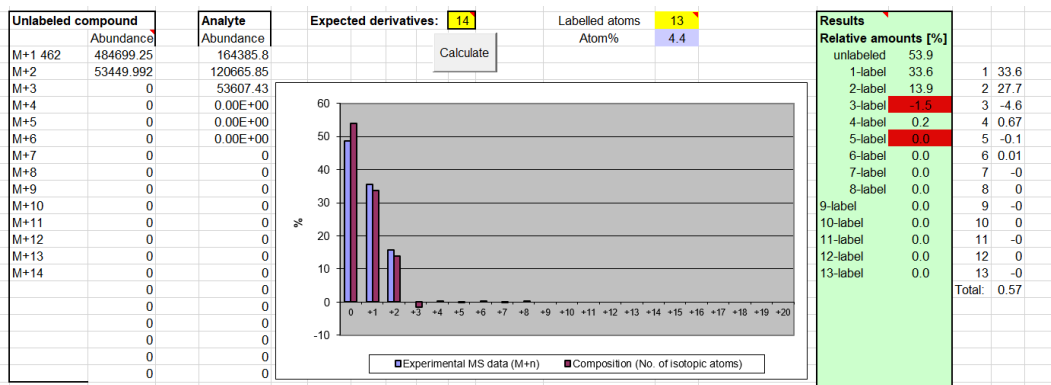


Mass spectrum of 13.

NB-01-62-D\_230213170655 #7 RT: 0.25 AV: 1 NL: 1.64E5  
T: ITMS + c ESI u Z ms [252.00-282.00]

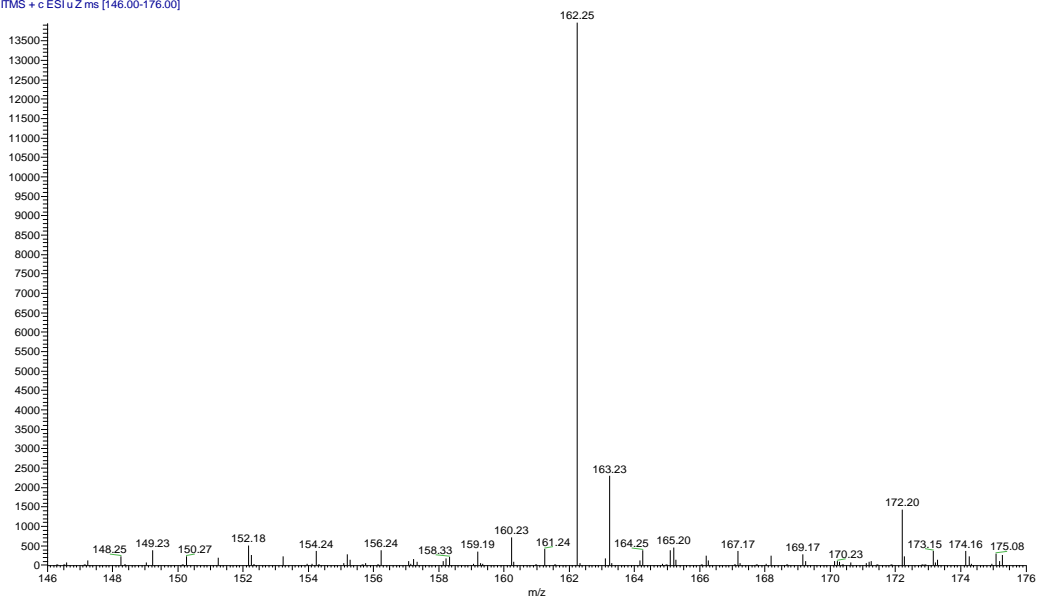


Mass spectrum of 13a.



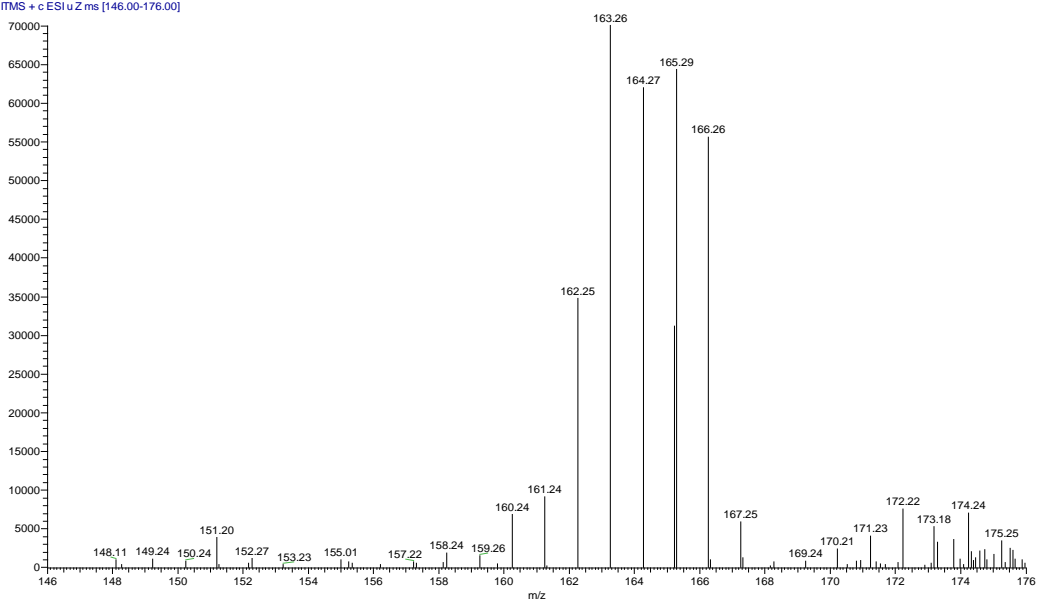
Mass spectrum and isopat analysis of 13a.

WD-6\_230213170655 #5 RT: 0.18 AV: 1 NL: 1.39E4  
T: FTMS + c ESI u Z ms [146.00-176.00]

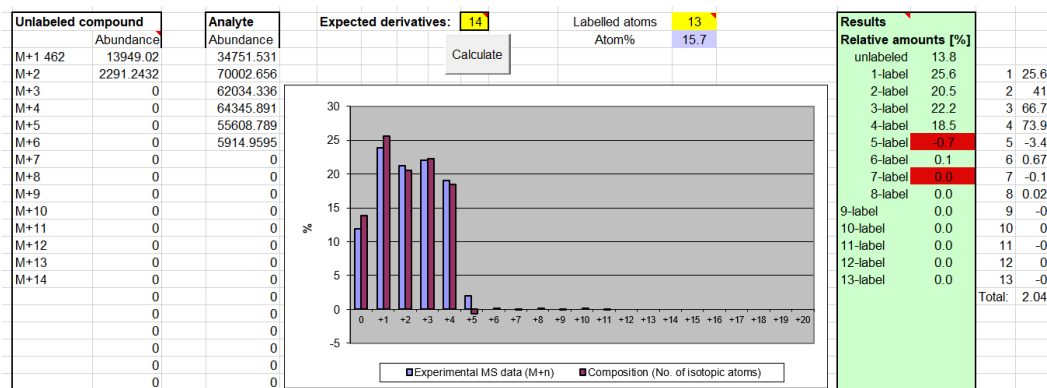


Mass spectrum of 14.

DG-03-161-D\_230213170655 #7 RT: 0.26 AV: 1 NL: 7.00E4  
T: FTMS + c ESI u Z ms [146.00-176.00]

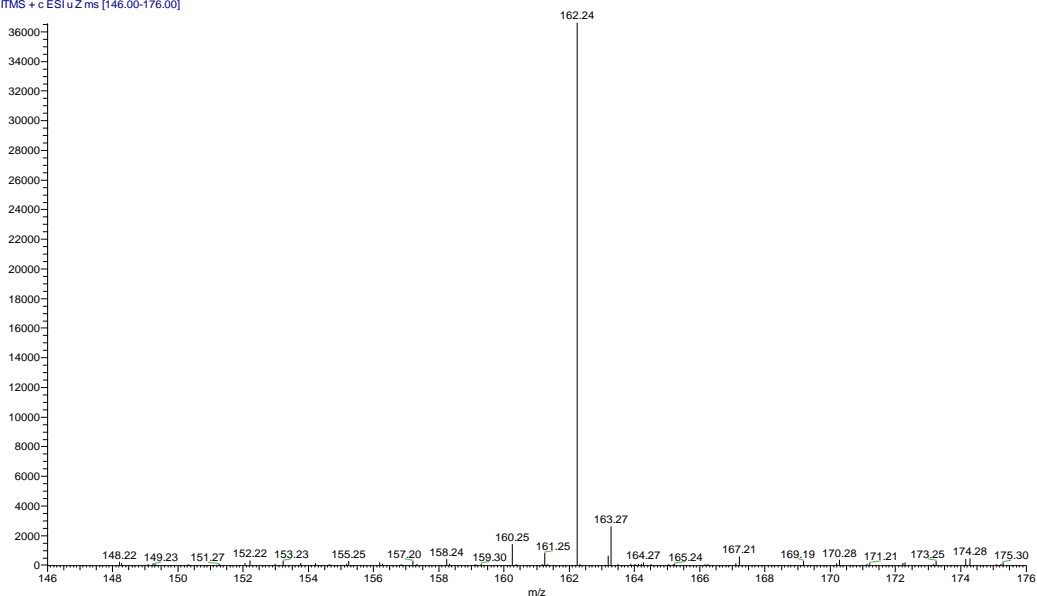


Mass spectrum of 14a.



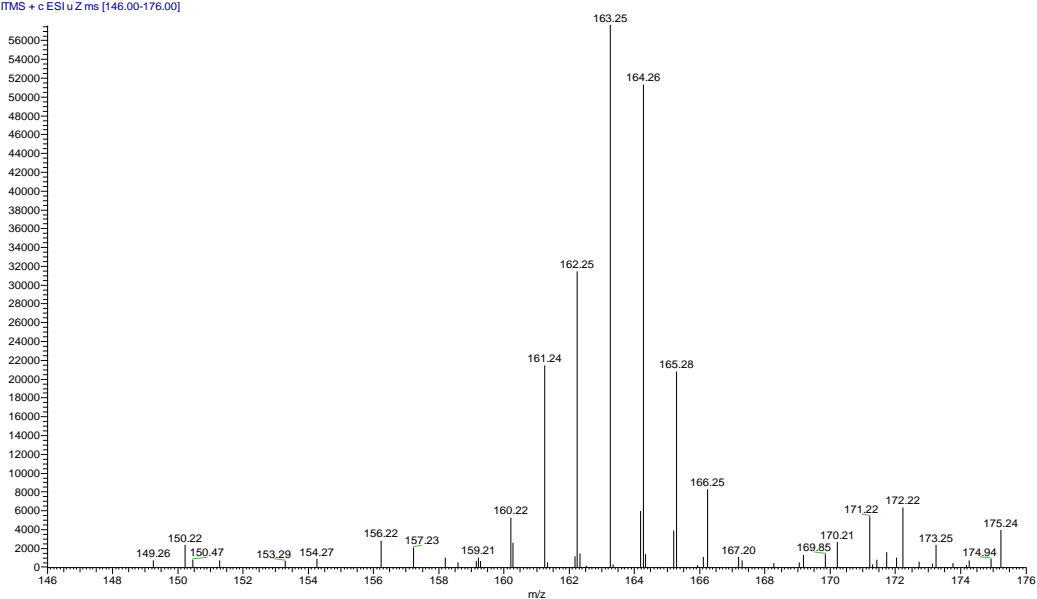
Mass spectrum and isopat analysis of 14a.

DG-03-203-SM\_230213170655 #8 RT: 0.31 AV: 1 NL: 3.66E4  
T: FTMS + c ESI u z ms [146.00-176.00]

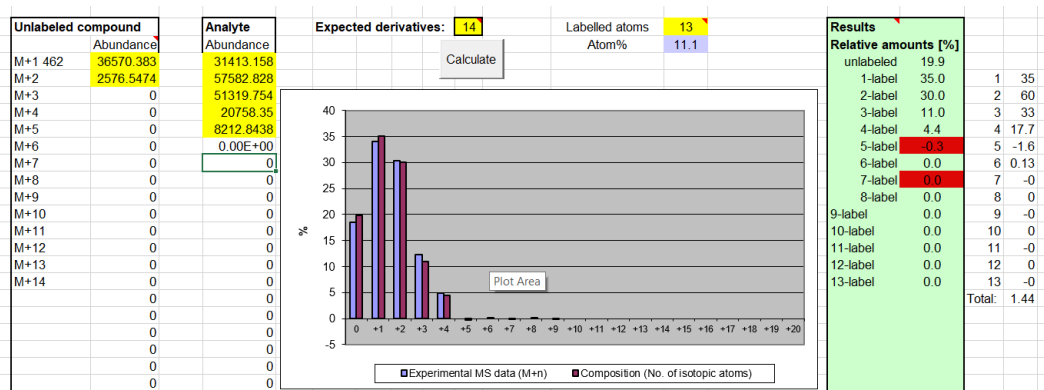


Mass spectrum of 15.

DG-03-203-D\_230213170655 #8 RT: 0.31 AV: 1 NL: 5.76E4  
T: FTMS + c ESI u z ms [146.00-176.00]



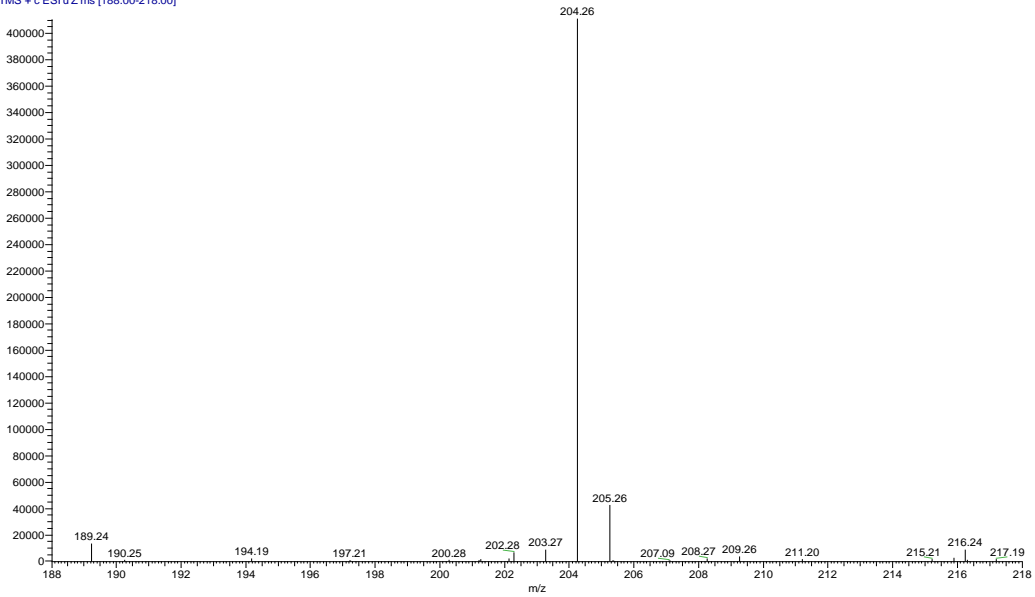
Mass spectrum of 15a.



Mass spectrum and isopat analysis of 15a.

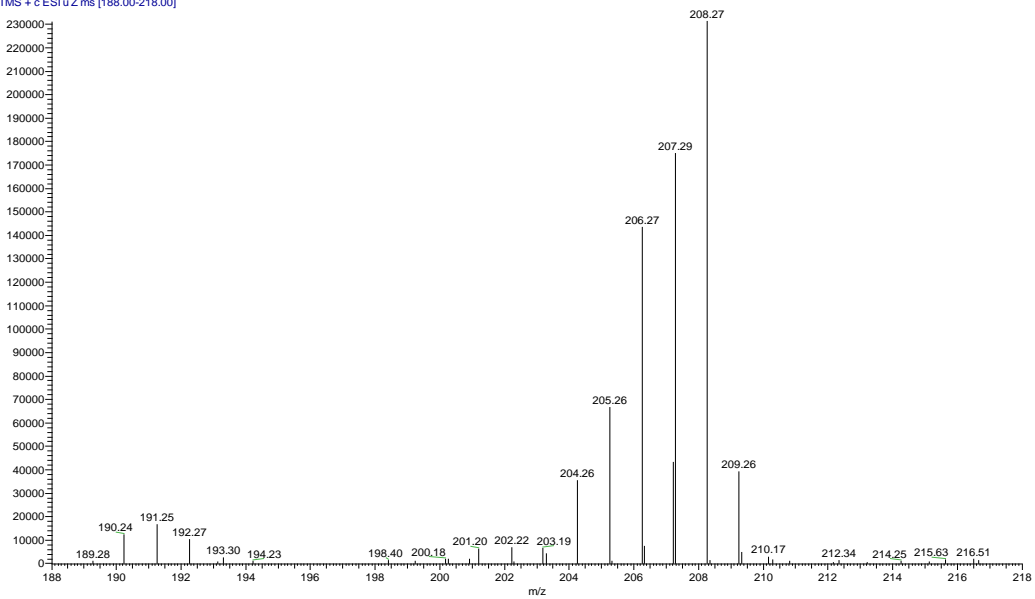


DG-03-142-SM\_230213170655 #5 RT: 0.17 AV: 1 NL: 4.11E5  
T: FTMS + c ESI u Z ms [188.00-218.00]

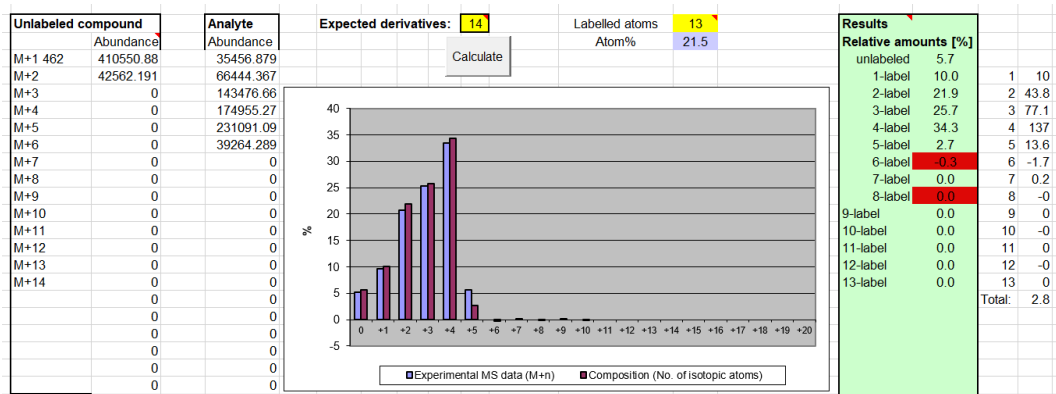


Mass spectrum of 16.

DG-03-142-D-2\_230213170655 #7 RT: 0.26 AV: 1 NL: 2.31E5  
T: FTMS + c ESI u Z ms [188.00-218.00]

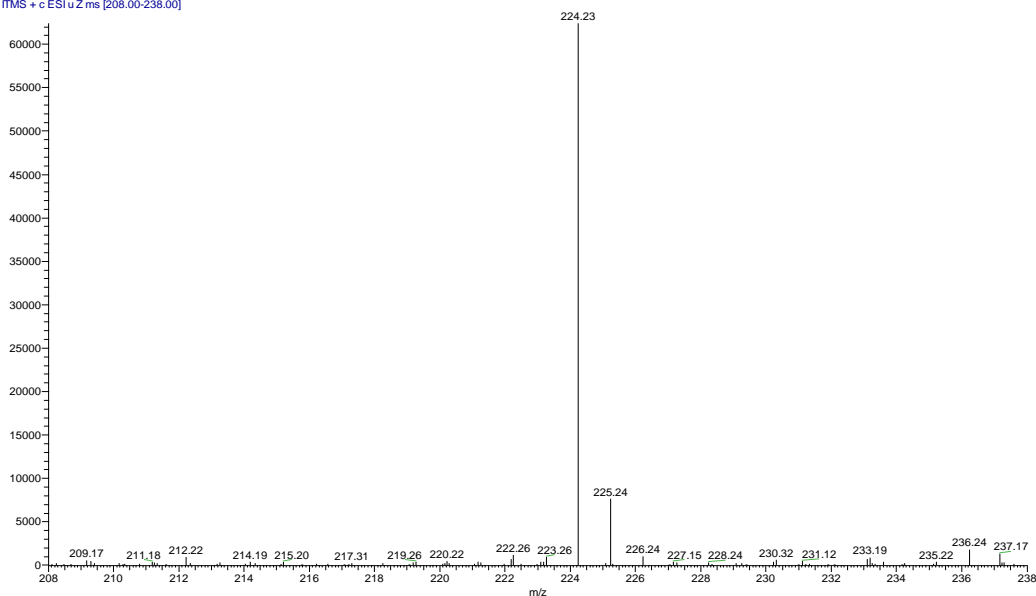


Mass spectrum of 16a.



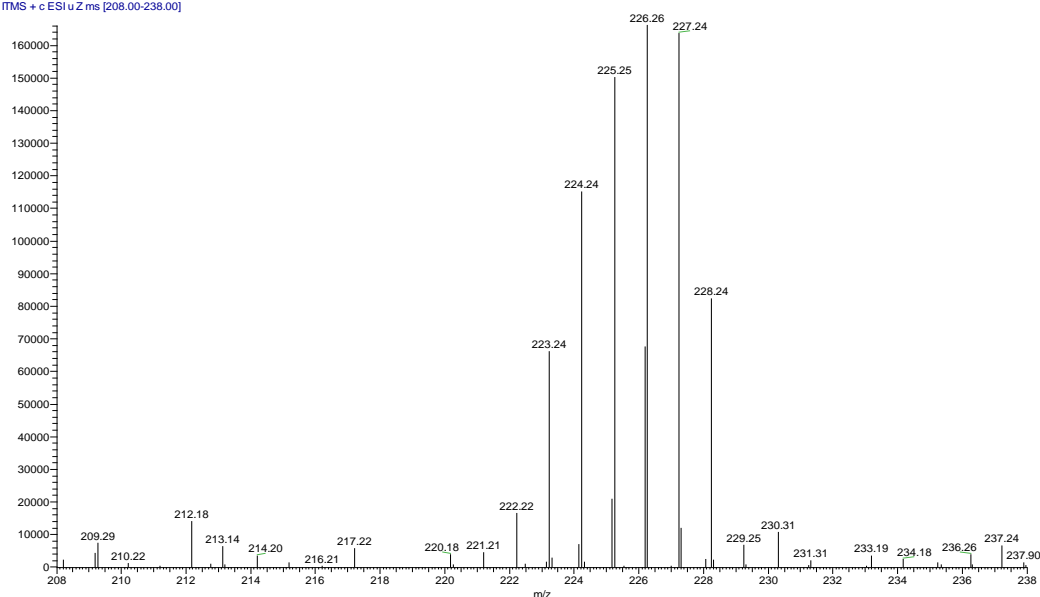
Mass spectrum and isotop analysis of 16a.

DG-03-215-SM\_230213170655 #8 RT: 0.30 AV: 1 NL: 6.24E4  
T: FTMS + c ESI u Z ms [208.00-238.00]

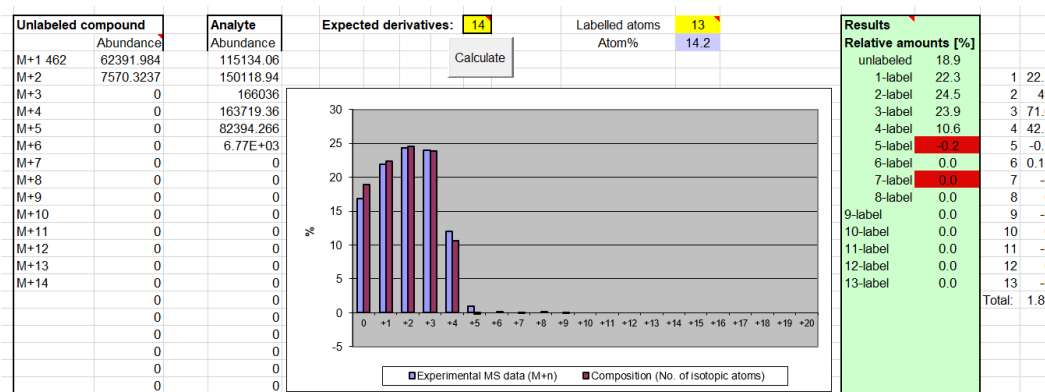


Mass spectrum of 17.

DG-03-215-D\_230213170655 #6 RT: 0.21 AV: 1 NL: 1.66E5  
T: FTMS + c ESI u Z ms [208.00-238.00]

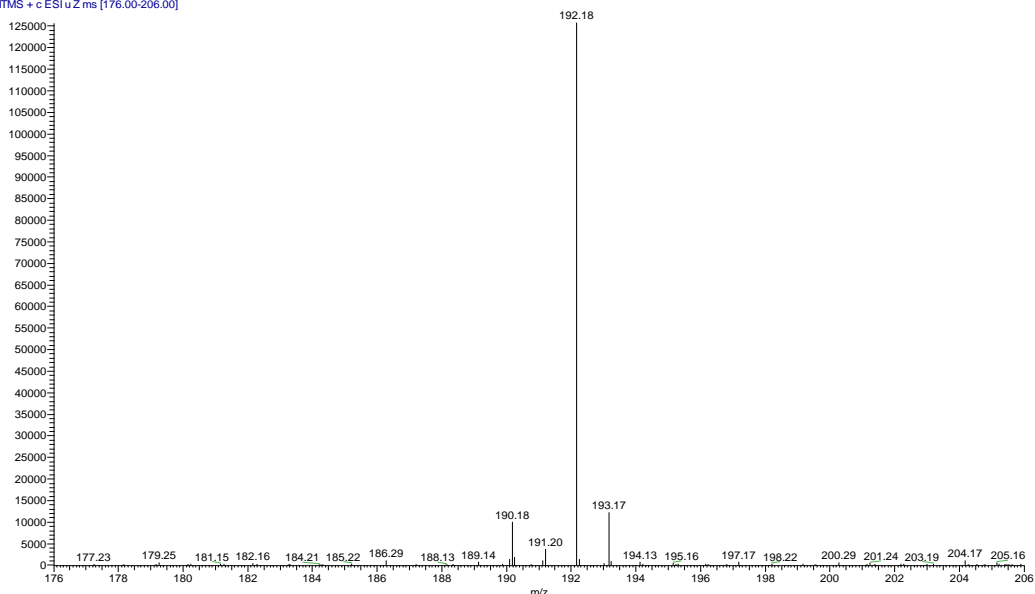


Mass spectrum of 17a.



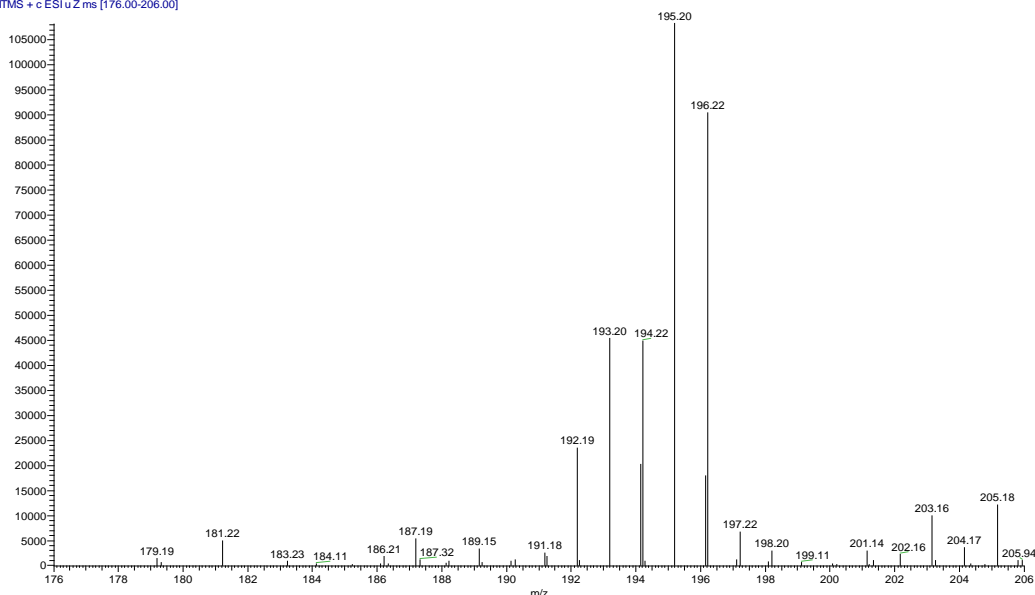
Mass spectrum and isopat analysis of 17a.

DG-03-141-SM\_230213170655 #7 RT: 0.26 AV: 1 NL: 1.26E5  
T: FTMS + c ESI u z ms [176.00-206.00]

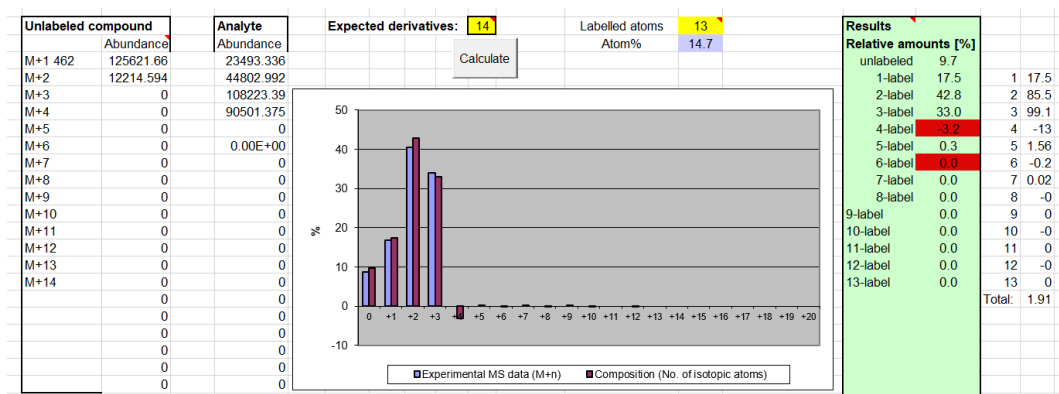


Mass spectrum of 18.

DG-03-141-D\_230213170655 #8 RT: 0.30 AV: 1 NL: 1.08E5  
T: FTMS + c ESI u z ms [176.00-206.00]

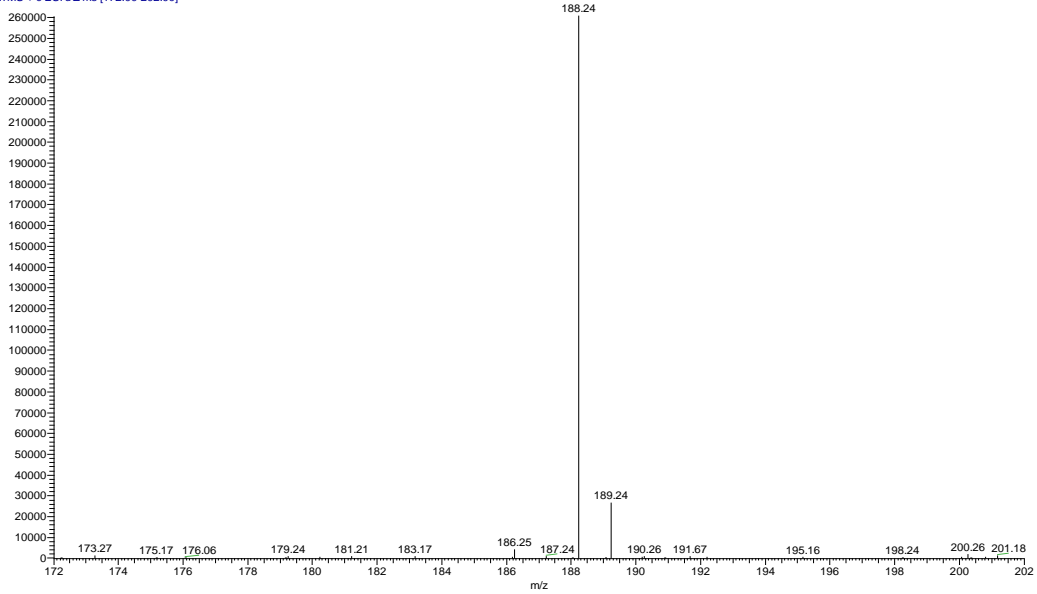


Mass spectrum of 18a.



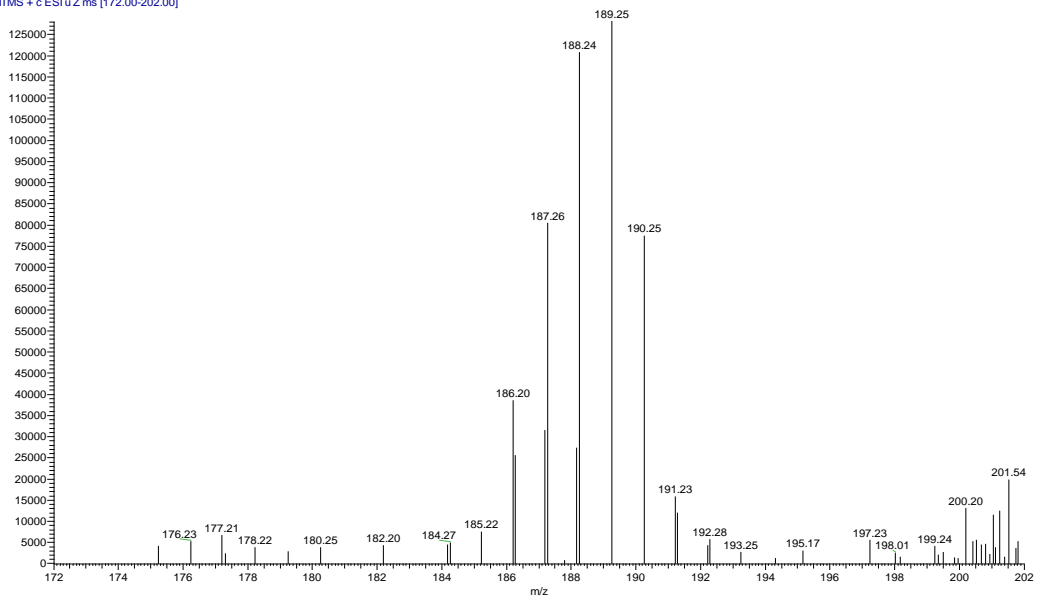
Mass spectrum and isotopic analysis of 18a.

DG-03-160-SM\_esi\_pos #6 RT: 0.21 AV: 1 NL: 2.61E5  
T: FTMS + c ESI u Z ms [172.00-202.00]

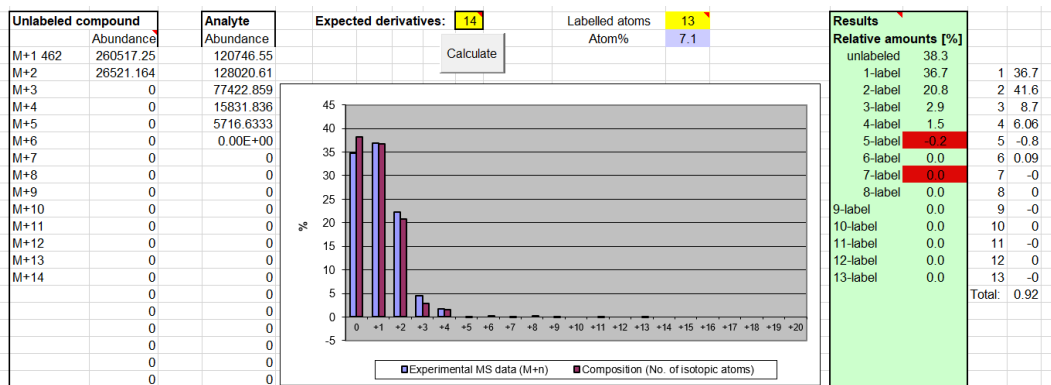


Mass spectrum of 19.

DG-03-160-D\_esi\_pos #4 RT: 0.13 AV: 1 NL: 1.28E5  
T: FTMS + c ESI u Z ms [172.00-202.00]

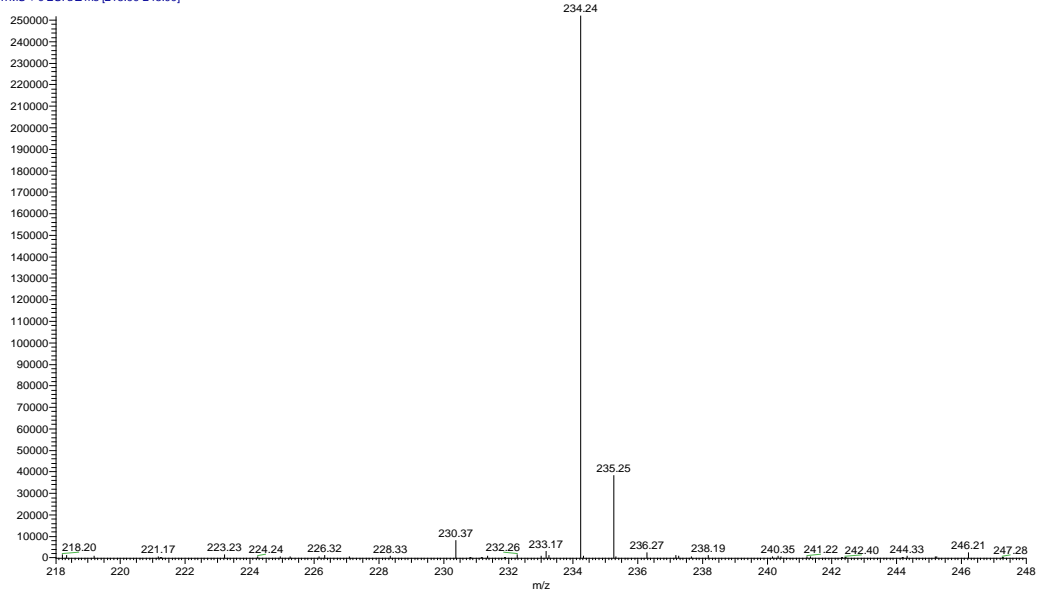


Mass spectrum of 19a.



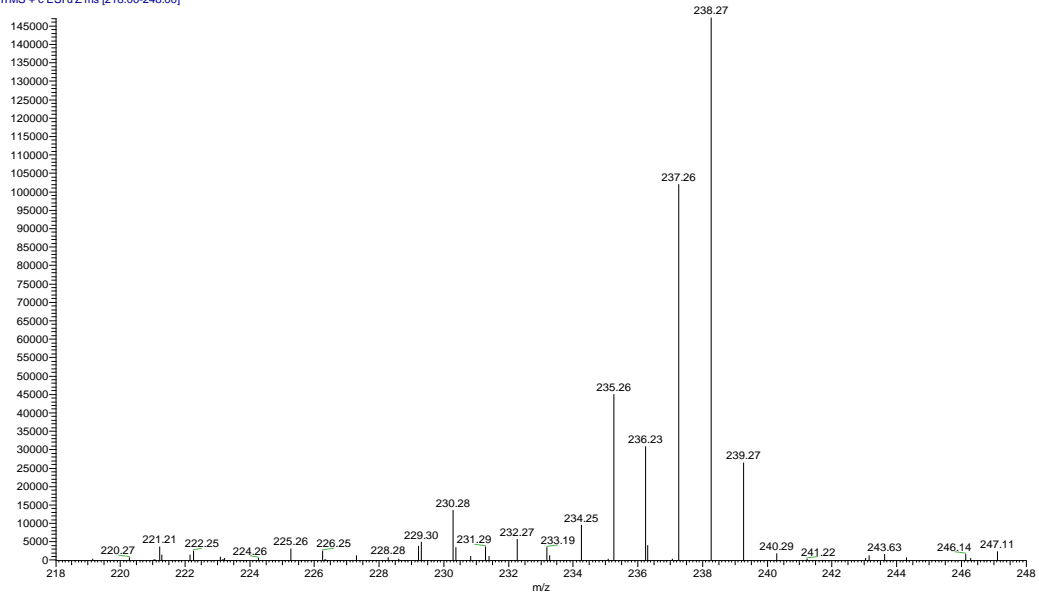
Mass spectrum and isotopic analysis of 19a.

DG-03-173-SM esi\_pos #6 RT: 0.21 AV: 1 NL: 2.52E5  
T: FTMS + c ESI u Z ms [218.00-248.00]

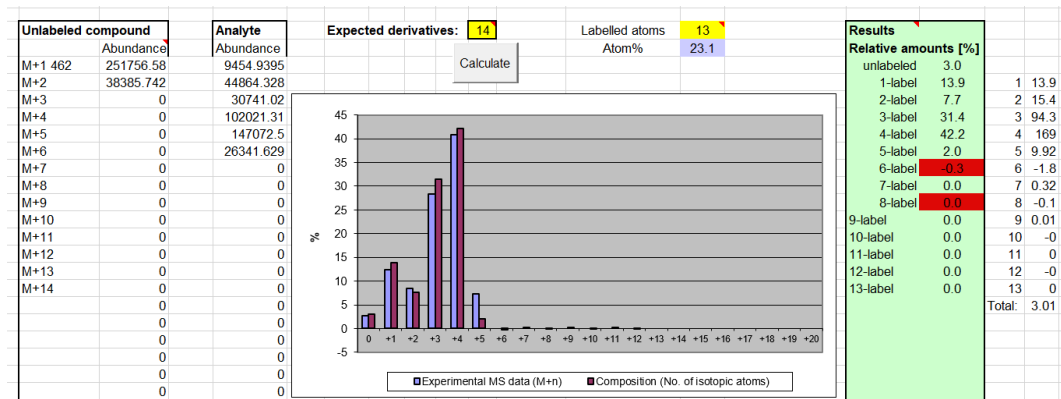


Mass spectrum of 20.

DG-03-173-D esi\_pos #6 RT: 0.21 AV: 1 NL: 1.47E5  
T: FTMS + c ESI u Z ms [218.00-248.00]

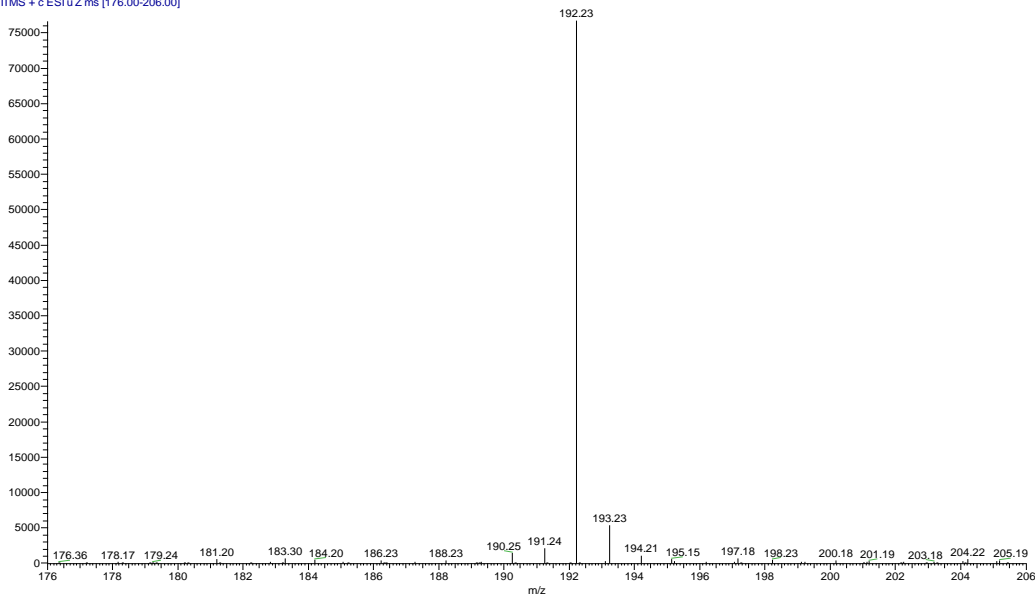


Mass spectrum of 20a.



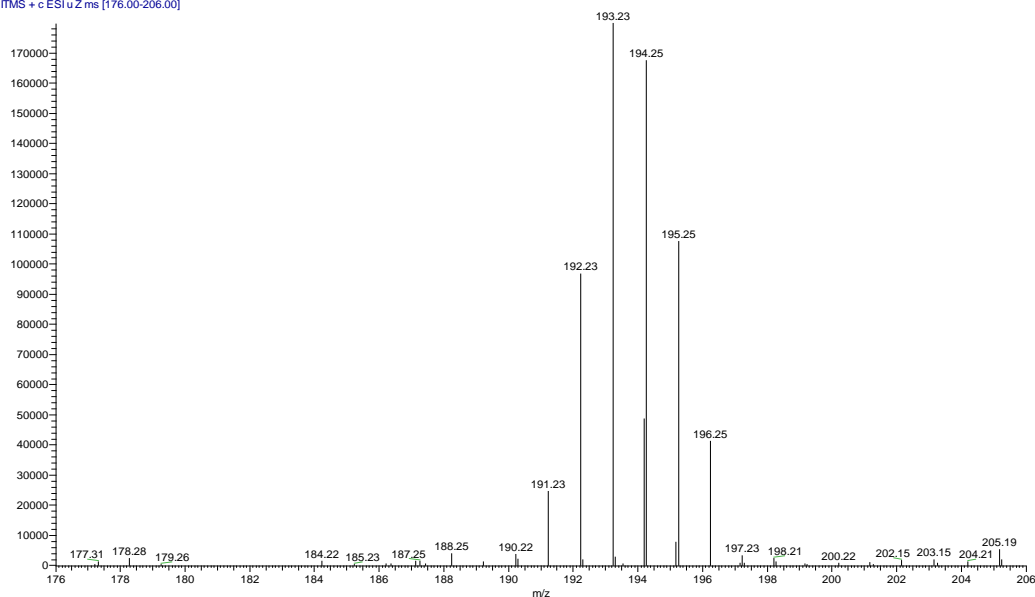
Mass spectrum and isotopic analysis of 20a.

DG-03-197-SM\_esi\_pos #6 RT: 0.21 AV: 1 NL: 7.66E4  
T: FTMS + c ESI u Z ms [176.00-206.00]

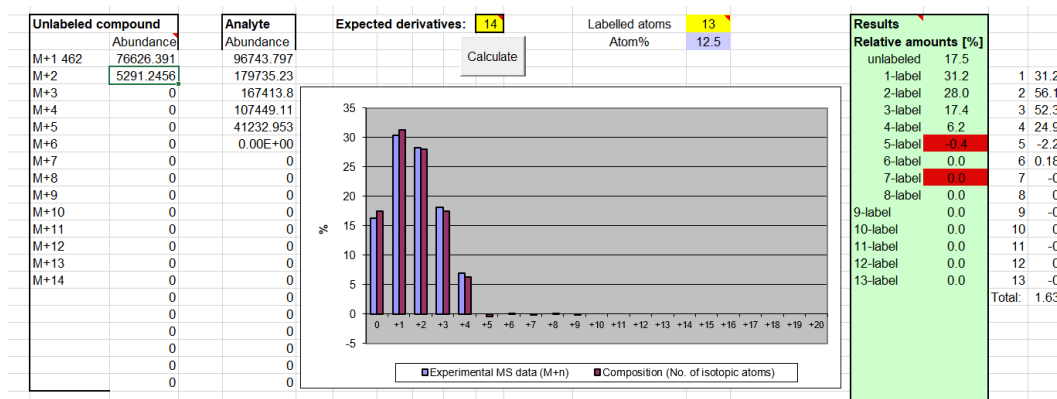


Mass spectrum of 21.

DG-03-197-D\_esi\_pos #10 RT: 0.38 AV: 1 NL: 1.80E5  
T: FTMS + c ESI u Z ms [176.00-206.00]

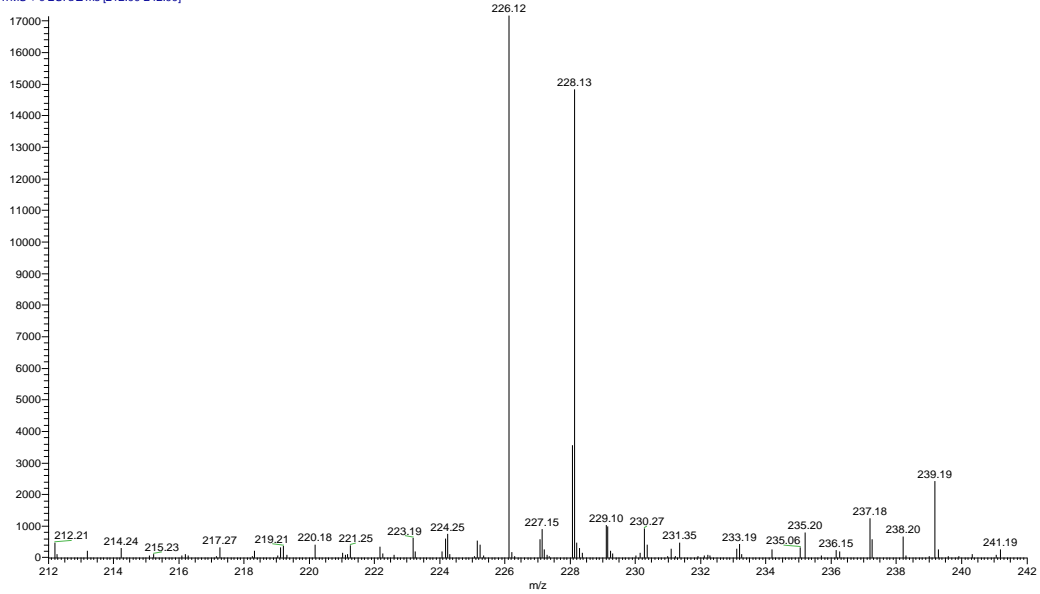


Mass spectrum of 21a.



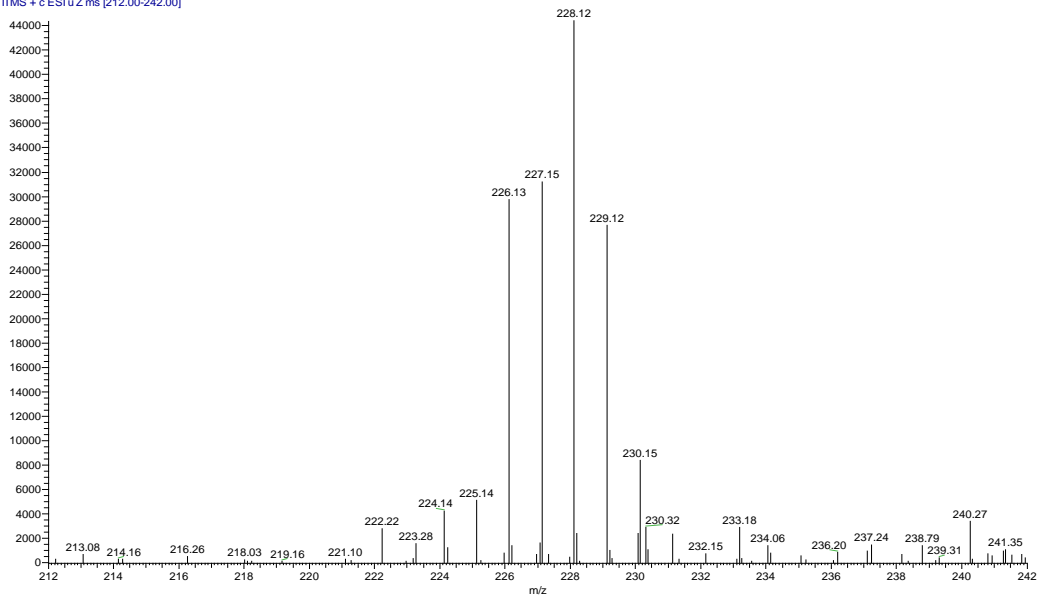
Mass spectrum and isotop analysis of 21a.

DG-03-153-SM\_esi\_pos #6 RT: 0.21 AV: 1 NL: 1.71E4  
T: FTMS + c ESI u Z ms [212.00-242.00]

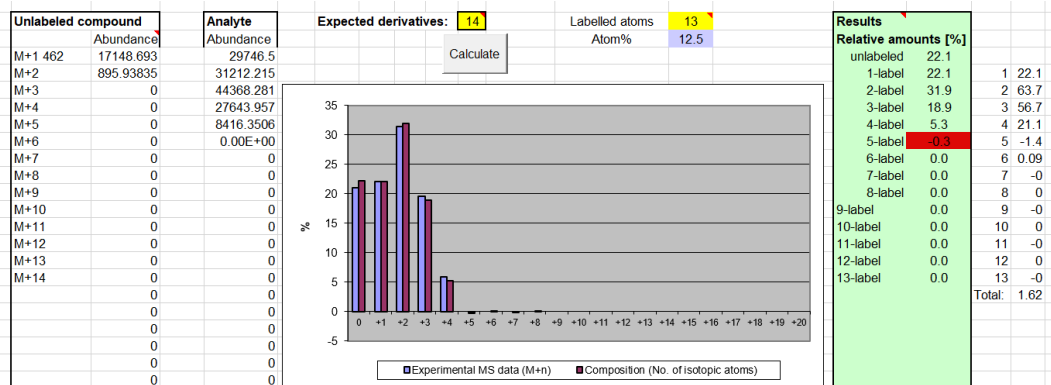


Mass spectrum of 22.

DG-03-153-D\_esi\_pos #8 RT: 0.30 AV: 1 NL: 4.44E4  
T: FTMS + c ESI u Z ms [212.00-242.00]

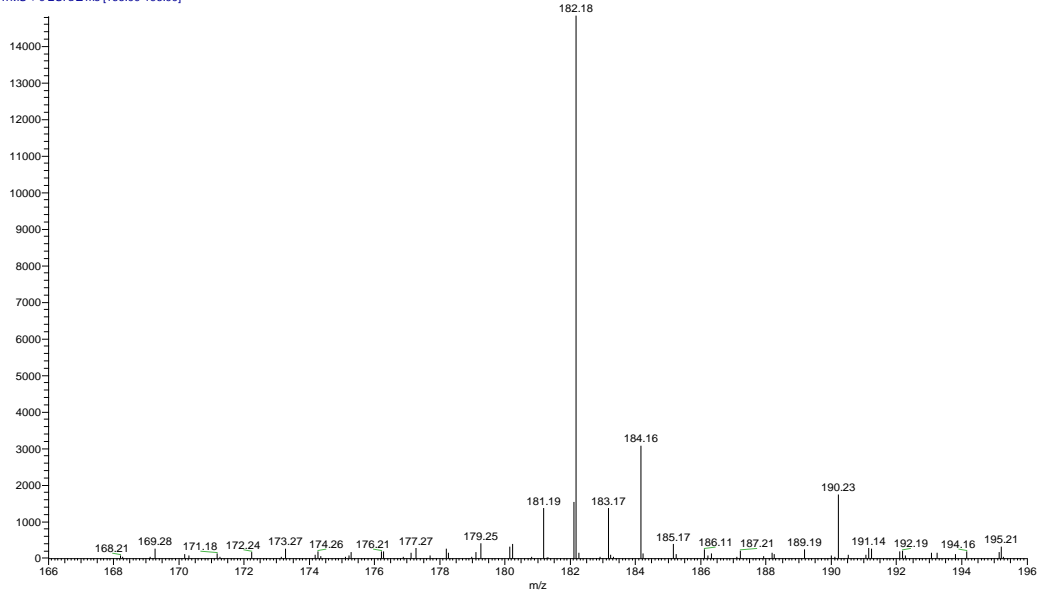


Mass spectrum of 22a.



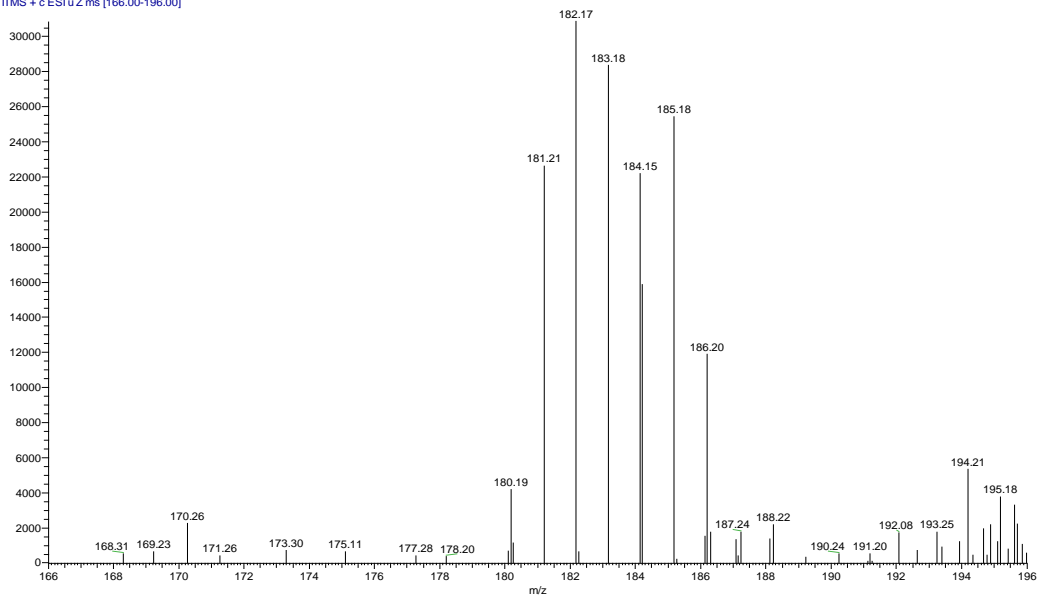
Mass spectrum and isopat analysis of 22a.

DG-03-151-SM\_esi\_pos #4 RT: 0.13 AV: 1 NL: 1.48E4  
T: FTMS + c ESI u Z ms [166.00-196.00]

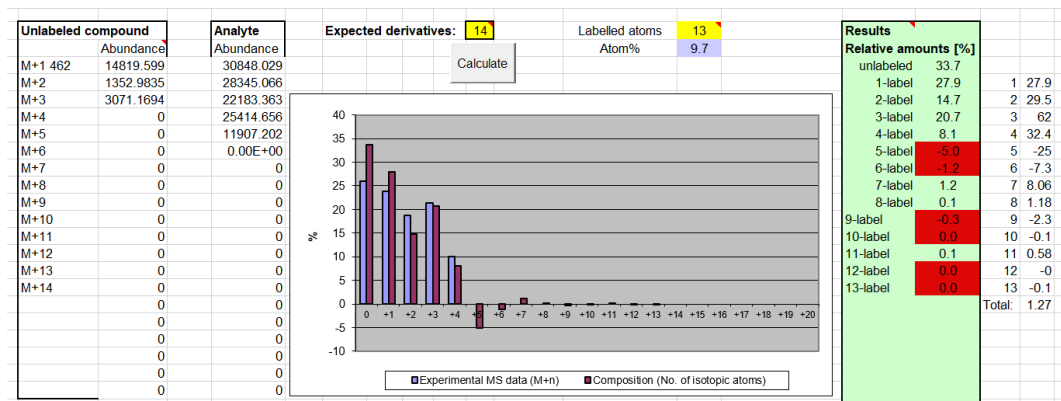


Mass spectrum of 23.

DG-03-151-D\_esi\_pos #6 RT: 0.21 AV: 1 NL: 3.08E4  
T: FTMS + c ESI u Z ms [166.00-196.00]



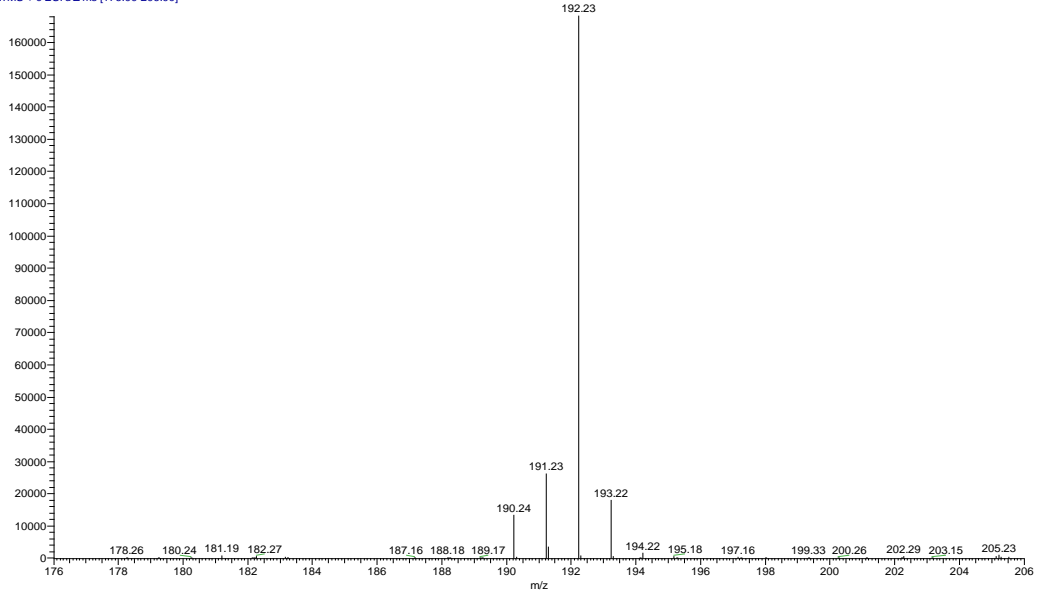
Mass spectrum of 23a.



Mass spectrum and isotop analysis of 23a.

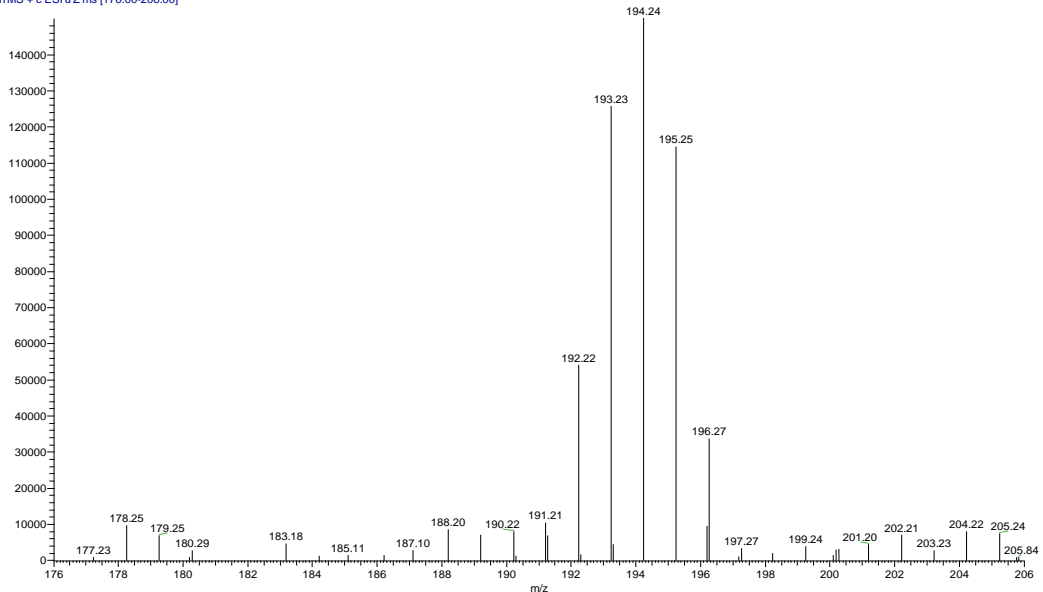


DG-03-225-SM\_esi\_pos #6 RT: 0.13 AV: 1 NL: 1.68E5  
T: ITMS + c ESI u z ms [176.00-206.00]

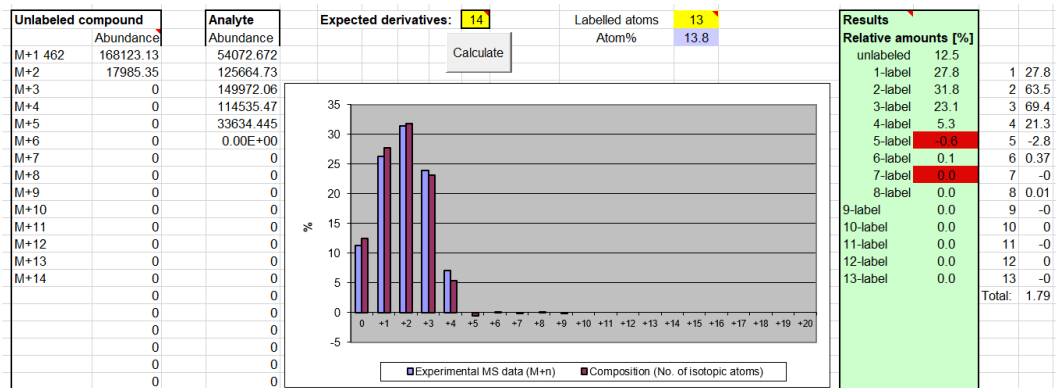


Mass spectrum of 24.

DG-03-225-D\_esi\_pos #6 RT: 0.21 AV: 1 NL: 1.50E5  
T: ITMS + c ESI u z ms [176.00-206.00]



Mass spectrum of 24a.



Mass spectrum and isopat analysis of 24a.