1 Supplementary information

Encapsulation efficiency (EE). One milliliter of astaxanthin nanoparticles were mixed with 1.0 mL of anhydrous ethanol, centrifuged at 10000 ×g for 10 min, and the supernatant was collected. The operation was repeated until the supernatant was colorless. The astaxanthin of the supernatant was regarded as the content of free astaxanthin, which was measured using a UV-visible spectrophotometer (SP-754, Shanghai Spectrum, Shanghai) at 476 nm. The EE was calculated according to equation (1).

9 EE (%)9 = (1 - content of free astaxanthin/total amount of astaxanthin) × 10010 (1)

The size and zeta potential of the particles with the highest EE were determined at 25 °C using a nanoparticle size and zeta potential analyzer (Malvern Zetasizer Nano 3 ZSE, Malvern Panalytical, UK).

X-ray diffraction (XRD). The crystalline structures of astaxanthin, MFGM-GA,
and astaxanthin nanoparticles were determined using an X-ray diffractometer (XRD,
D-MAX 2500, Rigaku, Japan). We used Cu Ka radiation at 40 kV and 30 mA, and
scans were made between 5 and 55 degrees at a scanning rate of 10 degrees/min.

Thermal analysis. The thermal weight loss behaviors of astaxanthin, carrier, and astaxanthin nanoparticles were determined using a Pyris 1 TGA thermogravimetric analyzer (TGA, PerkinElmer, Inc., UC). Appropriate amounts (about 3 mg) of samples were taken flatly into an aluminum tray and heated from 30 °C to 500 °C at a heating rate of 10 °C/min under a nitrogen flow of 100 mL/min.

Gastrointestinal stability. To study the digestive behavior of astaxanthin 23 nanoparticles, a model of gastrointestinal digestion of astaxanthin was built. The 24 astaxanthin nanoparticles and the aqueous solution containing a comparable amount 25 of free astaxanthin (0.5 mL) were mixed with an equal amount of ultrapure water. 26 Then, 1 mL of simulated gastric fluid (pH 7.0) was added. It was shaken for 1 h at 27 100 rpm in a water bath at 37 °C in the dark. Sampling and analysis were performed 28 at 30 and 60 min. Subsequently, 2 mL of simulated intestinal fluid (pH 7.0) was added 29 and shaken in a water bath (100 rpm) for 2 h at 37 °C in the dark. The digested 30 samples were centrifuged at $10000 \times g$ for 15 min, and the supernatant was collected 31 to determine the bioaccessibility of astaxanthin. The digestibility of astaxanthin was 32 calculated according to the equation (2). 33

34

Digestibility of astaxanthin (%) =
$$A_s/A_0 \times 100$$
 (2)

35 Where: A_0 denoted absorbance of the supernatant at 476 nm without digestion; A_s 36 denoted absorbance of supernatant at 476 nm after digestion.

	← 7 days	•	10 days
	Acclimation	1	Normal saline+Normal saline
Control			
	Acclimation		3% DSS+Normal saline
Model			
	Acclimation	3%	% DSS+Intervention materials
Intervention		L-AN:	Supplementation with 20, 40, and
		M-AN:	60 mg/kg of equivalent AST
		H-AN:	respectively;
		Pos:	50 mg/kg of SASP;
		H-A:	60 mg/kg of free AST;
		H-N:	2513 mg/kg of MFGMP-GA;

38

39 Figure S1. Schematic diagram of the animal experiment.



42 Figure S2. Effect of astaxanthin nanoparticles on Caco-2 cells. (A) Relationship 43 between TEER of Caco-2 monolayers and incubation time; (B) Effect of H_2O_2 on cell 44 viability. Different lowercase letters (a, b, c, and d) represent statistically significant 45 differences at the level of p < 0.05.

46

Score	Weight loss (%)	Stool character	Fecal occult blood
0	0	Normal	Normal
1	1-5	Soft but still formed	Weak positive blood
2	6-10	Soft stool	Positive blood
3	11-15	Loose stools	Visible bleeding
4	>15	Watery stool	gross bleeding

47 Table S1 DAI scoring criteria of mice

The DAI was recorded as the equation:

DAI

= (body weight score + stool consistency score + rectal

50

49

Score	Intestinal epithelium	Inflammation severity	
0	Normal	None	
1	Goblet cells damage	Infiltration limited to the crypt	
2	Extensive goblet cell damage	Infiltrate present in muscular mucosa	
3	Ioss of goblet cells	Covering large areas of muscular mucosa, mucosal edema	
4	Iarge areas without crypts	Infiltration in submucosa	

51 Table S2 The pathology scoring criteria of colon

Time (d)	Activity (AP)	Activity (BL)	AP/BL
 5	4.93±0.12	5.12±0.07	0.96 ^d
10	6.22±0.23	5.22±0.13	1.19°
15	14.46±0.31	8.96±0.44	1.61 ^b
20	18.87±0.12	10.01±0.02	1.89ª

54 Table S3 The ratio of alkaline phosphatase activity on AP to BL

55 ^{a-d} Different letters in each column indicate significant differences (p < 0.05).

	Cardiac	Liver index	Kidney	Spleen index	Thymus
	Index (%)	(%)	index (%)	(%)	index (%)
Con	0.56±0.01ª	4.14±0.12 ^a	1.11±0.11 ^b	0.26±0.08 ^b	0.16±0.01ª
Mod	$0.51{\pm}0.03^{a}$	3.66 ± 0.14^{bc}	1.32±0.09ª	0.36±0.11ª	0.18±0.01ª
L-AN	0.52±0.12 ^a	3.77 ± 0.15^{b}	1.02±0.01 ^b	$0.28{\pm}0.01^{ab}$	0.16±0.02ª
M-AN	$0.50{\pm}0.06^{a}$	$3.96{\pm}0.08^{ab}$	1.05±0.03 ^b	$0.29{\pm}0.03^{ab}$	0.16±0.01ª
H-AN	0.51±0.11ª	4.04±0.34ª	1.09±0.09 ^b	0.25±0.01 ^b	0.16±0.03ª
Pos	$0.44{\pm}0.05^{b}$	3.56±0.21°	1.15±0.11 ^b	0.18±0.02°	0.17 ± 0.05^{a}
H-A	0.52±0.13ª	3.76 ± 0.18^{b}	1.20±0.08 ^{ab}	0.22±0.07°	0.16±0.04ª
H-N	0.53±0.11ª	3.81±0.15 ^b	$1.24{\pm}0.06^{ab}$	0.20±0.06°	0.16±0.03ª

57 Table S4 The visceral index of mice in each group

⁵⁸ ^{a-c} Different letters in each column indicate significant differences (p < 0.05).