

Supplementary

## High-purity lignin from selective biomass fractionation with ternary deep eutectic solvents

Liang Ying EE <sup>a, b</sup>, Yong Kuok TAN <sup>a</sup>, Jiapei MIAO <sup>a</sup>, Hui Ting CHU <sup>a, c</sup>, Sam Fong Yau LI\* <sup>a, d</sup>

<sup>a</sup> Department of Chemistry, College of Humanities and Sciences, National University of Singapore, 3 Science Drive 3, Singapore 117543, Singapore

<sup>b</sup> Department of Chemical and Biomolecular Engineering, College of Design and Engineering, National University of Singapore, 4 Engineering Drive 4, Singapore 117585, Singapore

<sup>c</sup> Singapore Institute of Food and Biotechnology Innovation, 31 Biopolis Drive, Singapore 138669, Singapore

<sup>d</sup> NUS Environmental Research Institute, National University of Singapore, T-Lab Building, 5A Engineering Drive 1, 117411, Singapore

### Contents

1. Preparation and characterization of DES .....	2
2. DES dissolution of ground rice husks .....	7
3. Precipitation and characterization of lignin samples .....	9

## 1. Preparation and characterization of DES

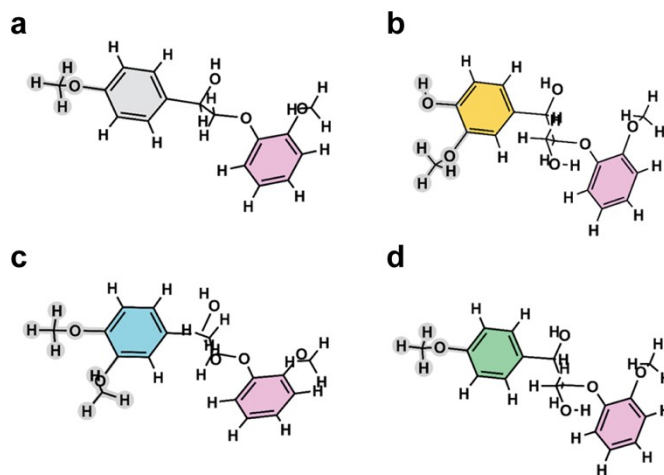


Fig. S1: a – d.  $\beta$ -O-4 Lignin model compounds for molecular simulations and theoretical calculations.

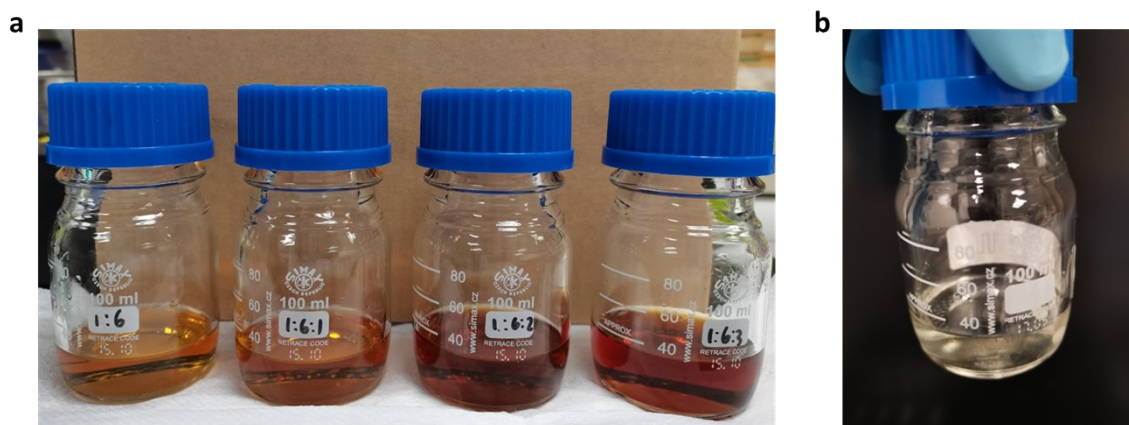
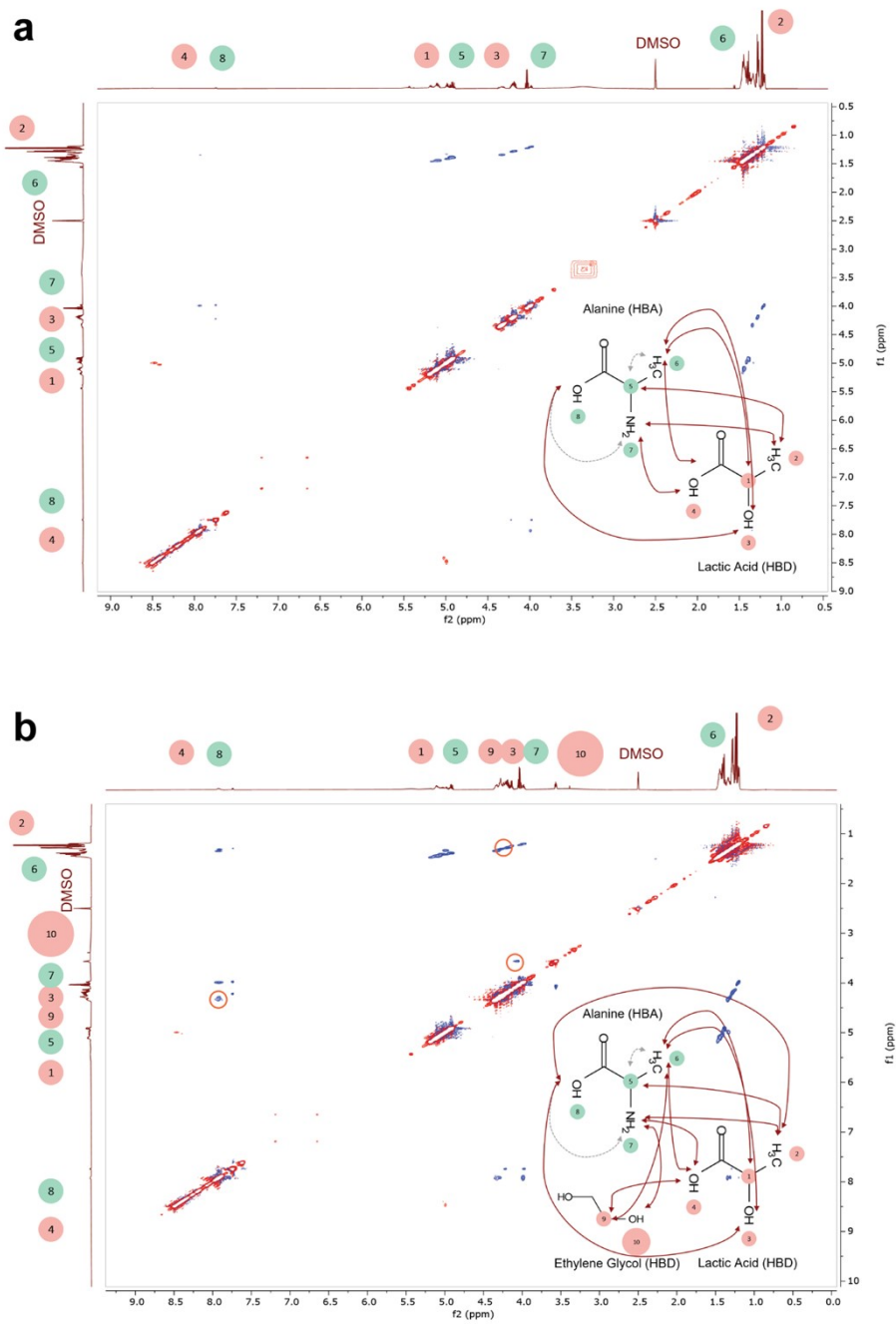
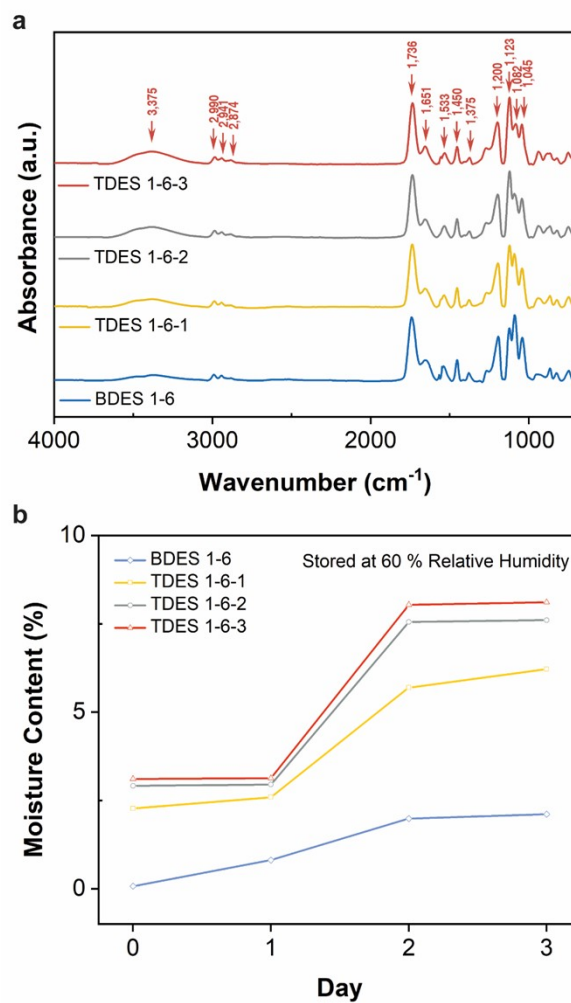


Fig. S2: Image of a. prepared BDES and TDES samples based on alanine, lactic acid, and ethylene glycol at different molar ratios, and b. lactic acid, tartaric acid, and choline chloride at molar ratio 4:1:1.



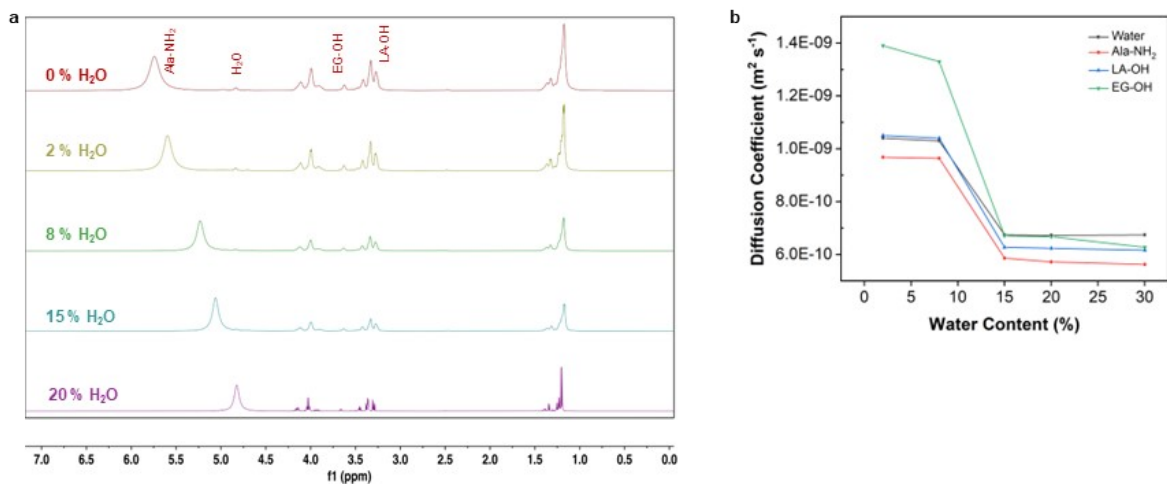
**Fig. S3:**  $^1\text{H}$ - $^1\text{H}$  2D NOESY NMR spectra of **a.** BDES and **b.** TDES with illustration of intramolecular and intermolecular hydrogen bond interactions.



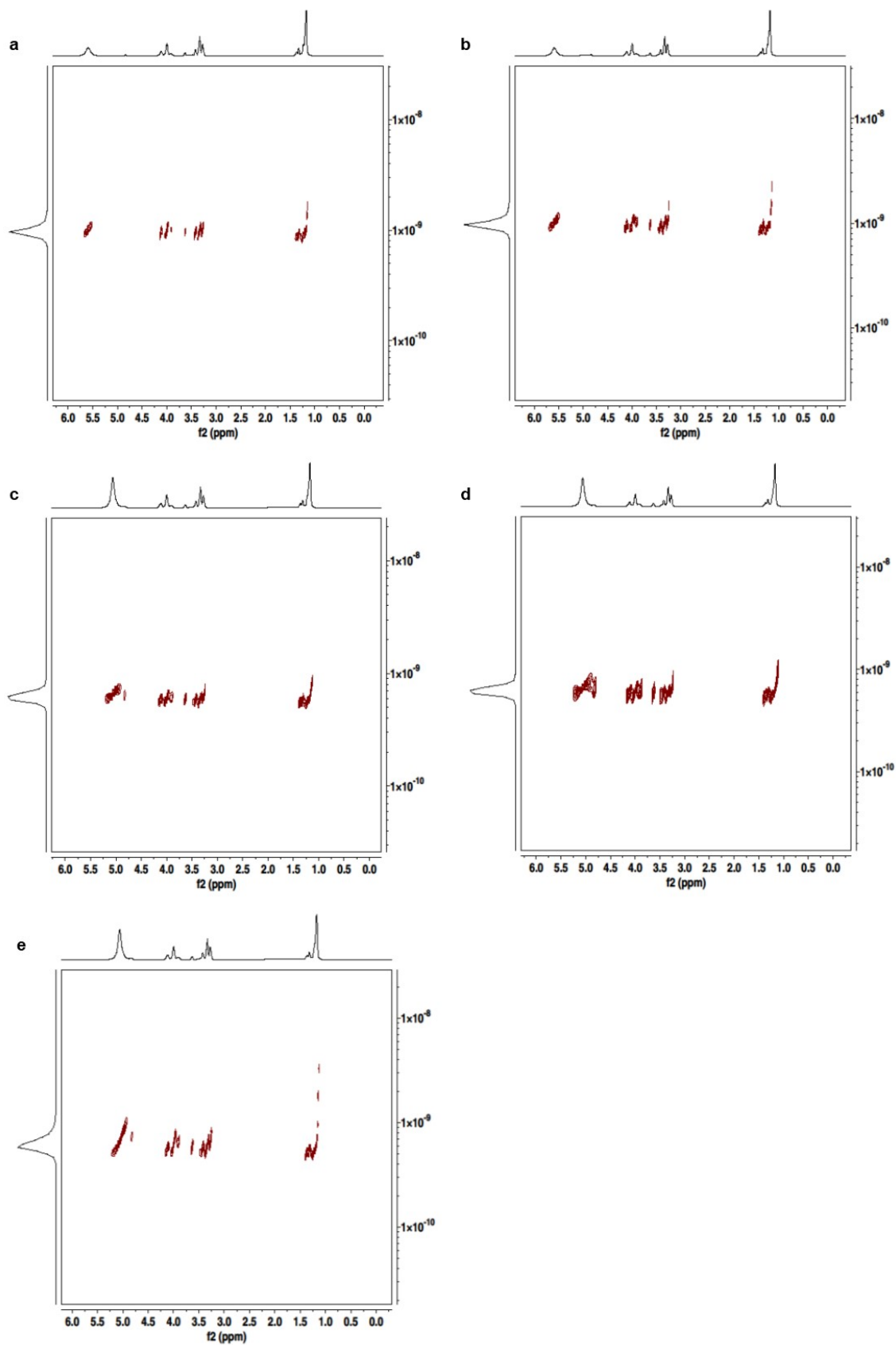
**Fig. S4:** a. ATR-FTIR spectra and b. moisture stability of deep eutectic solvent samples (stored in relative humidity 60 % and 24 °C).

**Table S1:** DSC & rheological analysis results of DES samples.

Sample	DSC Analysis		Viscosity (cP at 80 °C)
	Glass Transition Temperature (°C)	Vitrification Temperature (°C)	
BDES 1-6	-10.1	-	21.2
TDES 1-6-1	-61.7	-	80.7
TDES 1-6-2	-59.2	-	39.1
TDES 1-6-3	-56.8	-8.1	25.9



**Fig. S5:** **a.** <sup>1</sup>H {<sup>13</sup>C-decoupled} NMR spectra and **b.** diffusion coefficient determined from <sup>1</sup>H DOSY NMR analysis of TDES 1-6-3 with different water content.



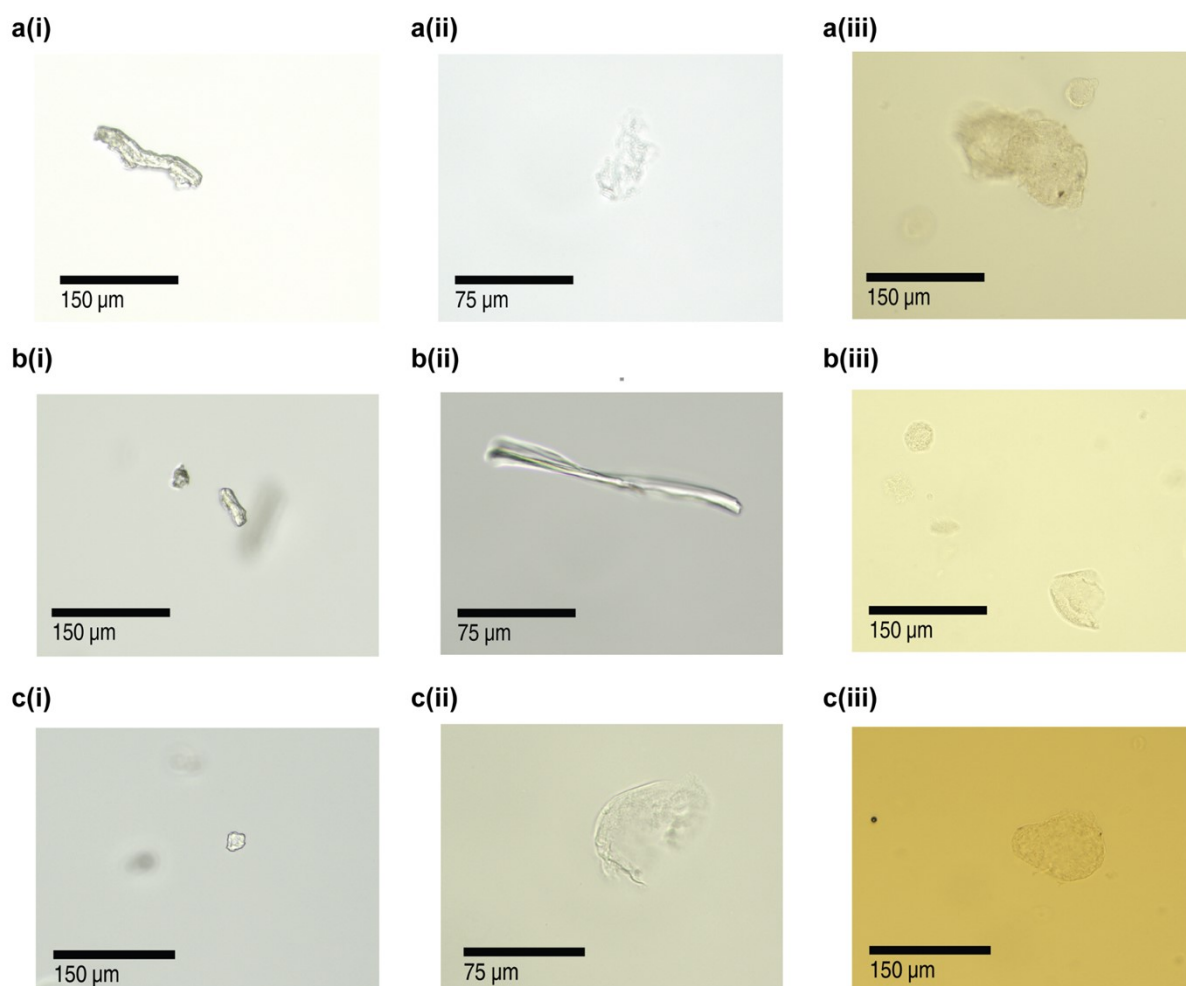
**Fig. S6:**  $^1\text{H}$  DOSY NMR spectra (Bayesian transform) of TDES 1-6-3 with **a.** 2 %, **b.** 8 %, **c.** 15 %, **d.** 20 %, and **e.** 30 % w/w water content.

## 2. DES dissolution of ground rice husks

**Table S2:** DES dissolution selectivity in rice husk pre-treatment.

	Untreated Rice Husk	BDES 1-6 Treated			TDES 1-6-1 Treated			TDES 1-6-2 Treated			TDES 1-6-3 Treated		
		1	2	3	1	2	3	1	2	3	1	2	3
Hemicellulose (%)	23.4	59.1	35.0	37.5	51.5	34.6	30.2	42.1	44.1	38.2	45.4	41.5	20.5
Cellulose (%)	36.5	29.1	49.2	45.4	32.9	50.8	48.0	36.0	41.4	42.9	34.8	42.6	57.7
Lignin (%)	27.1	11.2	10.1	11.2	5.6	9.1	14.4	7.8	10.0	12.7	6.7	10.9	13.2
Ash (%)	13.0	0.6	5.7	5.9	10.0	5.5	7.4	14.1	4.5	6.2	13.1	5.0	8.6

<sup>a</sup> DES was recycled for two more cycles of pretreatment of new rice husks samples



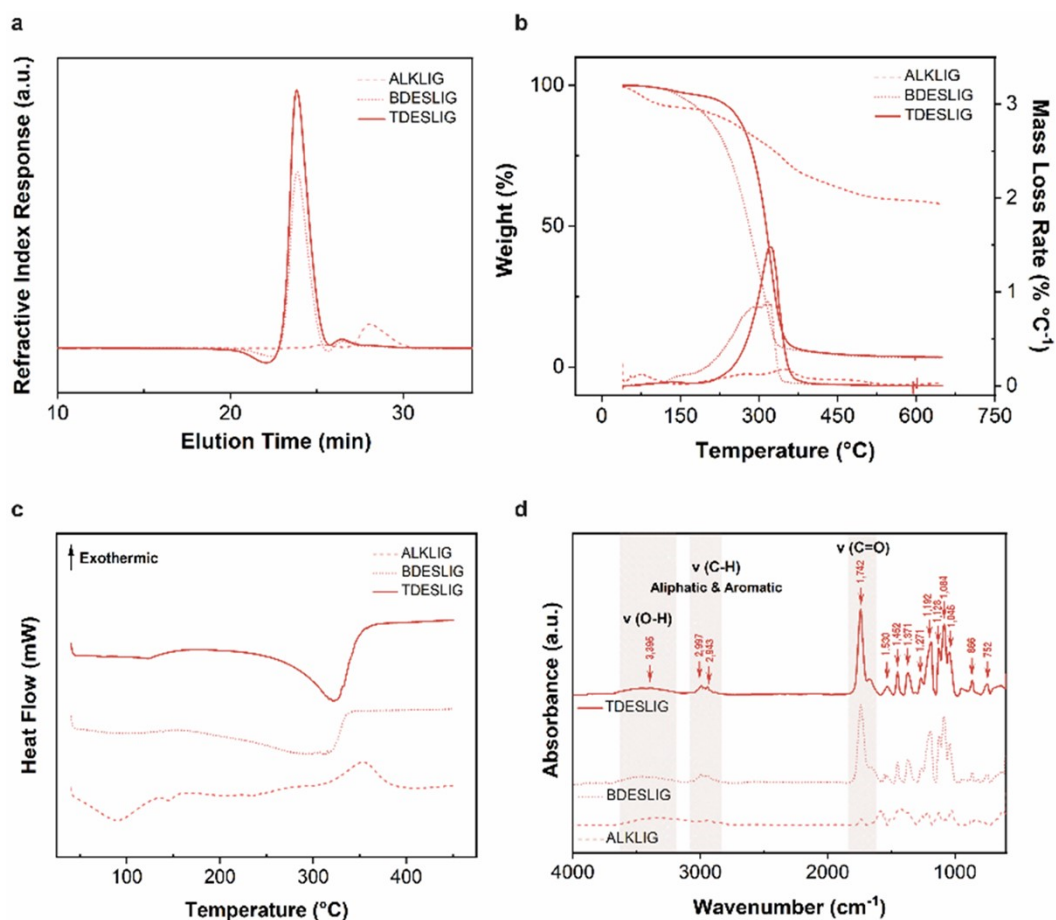
**Fig. S7:** Optical microscopic images of cellulose, hemicellulose, and lignin from **a(i) – a(iii)**. alkali-pretreatment, **b(i) – b(iii)**. BDES-pretreatment, and **c(i) – c(iii)**. TDES-pretreatment of rice husks.

**Table S3:**  $^{13}\text{C}$  qNMR assignment of precipitated lignin samples.

$\delta$ (ppm)	Assignment	Binary DES Lignin (mmol g $^{-1}$ )	Ternary DES Lignin (mmol g $^{-1}$ )	Alkali Lignin (mmol g $^{-1}$ )
50.0 – 48.1	C- $\beta$ in G type $\beta$ -5, $\beta$ - $\beta$ units	-	-	0.02
58.4 – 54.5	-OCH $_3$	9.3	4.1	0.3
61.0 – 58.5	C- $\gamma$ in G type $\beta$ -O-4 units	1.4	0.6	-
62.6 – 61.6	C- $\gamma$ in G type $\beta$ -5, $\beta$ -1 units	3.2	4.9	-
67.0 – 64.8	C- $\gamma$ in G type $\beta$ -O-4 units with $\alpha$ -C=O	20.2	38.5	-
68.0 – 66.6	C- $\alpha$ in G type $\beta$ -O-4 units (threo), C- $\gamma$ in G type $\beta$ - $\beta$	3.6	7.4	-
72.6 – 68.0	C- $\alpha$ in G type $\beta$ -O-4 units (erythro)	183.3	172.2	-
117.9 – 111.5	C-5 in G type	-	-	0.2
150.5 – 146.0	C-3/C-5 of S type, C-3 in G type $\beta$ -O-4 units	-	-	0.2
171.0 – 167.5	C=O in $\phi$ -COOH, Ester C=O in $\phi$ -C(=O)OR	177.6	192.7	-
173.4 – 167.5	Ester C=O in R- C(=O)OCH $_3$	38.5	29.2	-
175.7 – 173.2	C=O in $\phi$ -CHO	24.3	47.4	-
178.2 – 176.0	C=O in $\phi$ -CH=CH-CHO, C=O in $\phi$ -C(=O)CH(-O $\phi$ )-C-	4.9	1.4	-



### 3. Precipitation and characterization of lignin samples



**Fig. S8:** a. Gel-permeation chromatograms, b. TGA and derivative profile, c. DSC thermogram, and d. ATR FT-IR spectra of precipitated lignin samples from alkali-, BDES-, and TDES-pretreatments.

**Table S4:** GPC analysis results of precipitated lignin samples.

	Retention Time (min)	$M_n$	$M_w$	$M_p$	$M_z$	$M_{z+1}$
	20.4	6,324	6,584	6,453	6,845	7,101
ALK LIG	22.9	1,817	1,890	1,688	1,967	2,049
	25.2	341	355	336	371	388
BDES LIG	22.4	2,785	3,066	2,313	3,413	3,807
	23.9	721	795	862	865	930
TDES LIG	22.1	3,127	3,538	2,750	4,088	4,765
	23.8	718	794	881	865	929

**Table S5:** DSC & TGA analysis results of precipitated lignin samples.

Sample	DSC Analysis			TGA Analysis	
	Degradation Peak (°C)	$\Delta H_{\text{enthalpy}}$ (J g <sup>-1</sup> )	Onset Temperature (°C)	Peak Degradation Temperature (°C)	Total mass change (%)
ALK LIG	353.5	111.4	230.8	349.5	41.7
BDES LIG	312.1	-418.8	236.0	315.9	96.7
TDES LIG	324.2	-322.2	284.1	323.0	96.5

**Table S6:** <sup>1</sup>H-<sup>13</sup>C 2D HSQC NMR assignment of precipitated lignin samples.

$\delta_{\text{H}}/\delta_{\text{C}}$ (ppm)	Assignment
5.2/ 67.3	Hibbert's Ketone
5.0/ 70.0	C $\alpha$ -H $\alpha$ in $\beta$ -O-4' linkages to free hydroxyl
4.9/ 62.2	C $\gamma$ -H $\gamma$ in cinnamyl alcohol end-groups
4.4/ 59.6	C $\gamma$ -H $\gamma$ in $\beta$ -O-4' linkages to acetylated hydroxyl
4.1/ 72.9	C $\gamma$ -H $\gamma$ of resinol substructure
3.8/ 55.6	Methoxy-
3.7/ 62.5	C $\gamma$ -H $\gamma$ of phenylcoumaran substructure
3.6/ 53.8	C $\beta$ -H $\beta$ of phenylcoumaran substructure
3.4/ 55.7	C $\beta$ -H $\beta$ of resinol substructure
7.6/ 141.7	C2-H2/C6-H6 of pBA substructure
7.5/ 130.0	C2-H2/C6-H6 of p-coumarate substructure
6.8/ 118.7	C6-H6 in guaiacyl units
6.7/ 115.1	C5-H5 in guaiacyl units
6.4/ 110.3	C2-H2 in guaiacyl units
6.2/ 106.4	C2-H2/C6-H6 in syringyl units