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Supporting Information

Unlocking the photo-dehydrogenation ability of naphthalene monoimide towards the synthesis of quinazolinones

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1) General Information:

<u>Reagent Information</u>: All starting compounds employed in this study were procured from commercial suppliers. Potassium *tert*-butoxide, potassium hydroxide, and sodium hydroxide were purchased from Avra Synthesis Pvt. Ltd., India. 2-aminobenzonitrile; 1,8-naphthalic anhydride; and 2,6-diisopropylaniline were purchased from Sigma-Aldrich. These chemicals were used without further purification. Glassware were dried overnight at 160 °C. Solvents such as xylene, toluene, acetonitrile, THF were used as received from the suppliers (Finar Chemicals). For thin layer chromatography (TLC), aluminum foil (from Merck) coated with silica and fluorescent indicator 254 nm was used. Column chromatography was performed using SD Fine silica gel 60-120 mesh using a gradient of hexane and ethyl acetate/ diethyl ether as mobile phase.

Analytical Information: All isolated compounds were characterized by ¹H NMR, ¹³C NMR spectroscopies and high-resolution mass spectrometry. ¹H NMR and ¹³C NMR spectra were recorded on a 400 MHz Bruker Biospin Advance III FT-NMR spectrometer. NMR shifts are reported as delta (δ) units in parts per million (ppm), and coupling constants (J) are reported in Hertz (Hz). Chemical shifts (δ) are quoted to the nearest 0.01 ppm relative to the residual protons in CDCl₃ (δ 7.26 ppm). Carbon chemical shifts are internally referenced to the deuterated solvent signals in $CDCl_3$ (δ 77.1 ppm). High-resolution mass spectra (HRMS) were recorded on a Waters QTOF mass spectrometer. Fluorescence quenching experiments of the NMI catalyst were carried out in a Cary Eclipse Fluorescence Spectrofluorometer. The X-band EPR spectrum was collected in a JEOL Model: JES-FA200 machine operating at a microwave frequency of 9.4 GHz, modulation frequency of 100 kHz, 5 mW power, along with 30 seconds of sweep time. UV-Vis was recorded using Varian Cary 60 (Agilent Technologies) spectrophotometer. The total volume in the cuvette was fixed at 1 ml and a cuvette of path length 1 cm was used. The photochemical reactions were conducted with eplite 50 W blue LED (455 nm). The reaction tube was kept 7-8 cm away from the light source.

2. General synthetic procedure for 2-(2,6-diisopropylphenyl)-1*H*- benzo[de]isoquinoline -1,3(2*H*)-dione (NMI) Catalyst¹:

The NMI molecule was synthesized according to the previously reported literature procedure¹. To a solution of 1,8-Naphthalic anhydride (5.05 mmol, 1.0 g) in glacial acetic acid (10 mL) taken in a sealed tube, 2,6-diisopropylaniline (6.06 mmol, 1.2 mL) was added and heated at 110 °C for 6 h with continuous stirring. After that, the reaction mixture was cooled to room temperature followed by filtration and washing with diethyl ether, and the resulting solution was dried. Further purification was done by column chromatography using ethyl acetate/hexane solvent mixture (3:7) as the eluent to afford **NMI** as a greenish-white shiny solid (1.156 g, 64%) The desired products were fully characterized by ¹H, ¹³C NMR spectroscopies.

¹H NMR (400 MHz, CDCl₃) δ 8.68 (dd, J = 7.3, 1.2 Hz, 2H), 8.31 (dd, J = 8.3, 1.1 Hz, 2H), 7.82 (dd, J = 8.2, 7.2 Hz, 2H), 7.48 (dd, J = 8.2, 7.3 Hz, 1H), 7.34 (d, J = 7.8 Hz, 2H), 2.76 (s, 2H), 1.17 (d, J = 6.9 Hz, 12H).¹³C{¹H} NMR (100 MHz, CDCl₃) δ 164.3, 145.7, 134.3, 131.9, 131.8, 130.9, 129.5, 127.1, 124.1, 122.8, 29.1, 24.0.



Figure S1. ¹H NMR spectrum (400 MHz) of NMI in CDCl₃.



Figure S2. ¹³C NMR spectrum (100 MHz) of NMI in CDCl₃

3. Table S1: Optimization of the reaction condition



Entry	Catalyst (mol%)	Base (equiv.)	Time (h)	Solvent	Yield* (%)
1	5	KO ^t Bu (1)	16	Toluene	18
2	10	KO ^t Bu (1)	16	Toluene	28
3	10	KO ^t Bu (1)	16	Xylene	88
4	10	KO ^t Bu (0.5)	16	Xylene	45
5	10	KO ^t Bu (1)	12	Xylene	58
6	15	KO ^t Bu (1)	20	Xylene	86
7	5	KOH (1)	16	Xylene	10
8	10	NaOH (1)	16	Xylene	8
9	10	KO ^t Bu (1)	16	MeCN	trace
10	15	KO ^t Bu (1)	16	THF	0
11	10	KO ^t Bu (1)	16	MeOH	trace
12	-	KO ^t Bu (1)	16	Xylene	12
13	10	-	16	Xylene	n.r

* *Reaction conditions*: NMI (x mol %, with respect to 2-amino benzonitrile), 2-amino benzonitrile (0.5 mmol), alcohol (0.75 mmol), KO^tBu (y mmol), solvent (2 mL) in Blue LED, 16 h (isolated yield).

4. General procedure

a) General procedure for the synthesis of Quinazolinones using benzyl and aliphatic alcohols with 2-aminobenzonitrile.



A 15 mL vial tube was charged with 10 mol% of **NMI**, 0.5 mmol of *o*-aminobenzonitrile, KO^tBu (0.5 mmol), and alcohol (0.75 mmol) in 2 mL xylene. The reaction mixture was stirred under blue light irradiation for 16 h in an open atmosphere. Upon completion, the reaction mixture was worked up in ethyl acetate and brine mixture. Then the organic layer was collected, dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by column chromatography using hexane/ethyl acetate (4:1 to 2:1 gradually) as eluent to afford pure products. The desired products were fully characterized by ¹H, ¹³C NMR spectroscopies.

b) General synthetic procedure for the synthesis of 2*H*-benzo[1,2,4]thiadiazine-1,1dioxide derivatives or quinazolinone derivatives from aliphatic alcohols:



A 15 mL vial was charged with 10 mol% of **NMI**, 0.5 mmol of 2-aminobenzenesulfonamide or 2-aminobenzonitrile, KO^tBu (0.5 mmol), and aromatic/aliphatic alcohol (0.75 mmol) in 2 mL xylene. The reaction mixture was stirred under blue light irradiation for 24 h in an oil bath at 60 °C. Upon completion, the reaction mixture was cooled and worked up in ethyl acetate and brine mixture. Then the organic layer was collected, dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by column chromatography using hexane/ethyl acetate (1:1

to 1:4) as eluent to afford pure products. The desired products were fully characterized by ¹H, ¹³C NMR spectroscopies.

5. Mechanistic investigation:

a) Procedure for radical quenching experiment:



In a typical reaction, a 15 mL vial was charged with **NMI** (10 mol%), benzyl alcohol (0.75 mmol), 2-aminobenzonitrile (0.5 mmol) KO^tBu (0.5 mmol), and the varying equivalent of TEMPO dissolved in 2 mL xylene. The reaction mixture was stirred under blue light irradiation for 16 h in an open atmosphere, to maintain the identical conditions to the optimized protocol. The yield of the reaction decreased drastically with the addition of TEMPO, as probed by GC-MS analysis. On addition of 1.2 equivalent of TEMPO complete quenching of the reaction occurs.

Product formation on varying equivalent of TEMPO addition

Tempo (x Equiv.)	Product formation
0.6	14%
1.2	0%

b) Observation of color changes during the reaction:

The molecule **NMI** when dissolved in xylene gives a colorless solution. Then KO^tBu was added to it and kept the solution in dark. There was no visual color change under the dark condition. On the other hand, as soon as visible light is irradiated on the reaction mixture, the solution first turns into orange, which slowly darkens and changed to wine-red. The resulting wine-red solution was analysed by both UV-vis and EPR spectroscopies.



Figure S3. The progression of color during exposing the NMI and KO^tBu in xylene to visible light. The molecule NMI alone in xylene is a colorless solution.

c) EPR analysis of NMI radical anion:

In a solution of **NMI** (0.028 mmol, 10 mg) in xylene, KO^tBu (0.178 mmol, 20 mg) was added. Under the dark condition, this solution is completely colorless. Then the reaction mixture was irradiated with white light for 5 minutes which prompted a sharp color change to wine red. The X-band (9.4 GHz) EPR measurement of this solution was carried out at room temperature. A sharp signal at g = 2.005 was observed, which indicated the formation of a **NMI** radical anion.



Figure S4. X-band (9.4 GHz) EPR signal of **NMI** radical anion obtained by adding KO^tBu and after shining visible light. During the data acquisition the modulation frequency was 100 kHz, and the power was 5 mW.

6. UV-Visible absorption spectra:



Figure S5. UV-visible spectra of NMI in xylene (6.7×10^{-5} M solution was taken)



Figure S6. UV-visible spectra of NMI in xylene after the addition of KO^tBu and light excitation.

7. Fluorescence quenching experiment and Stern-Volmer plot:

The Stern-Volmer quenching experiments were carried out to explore the intramolecular reductive quenching of the photocatalyst **NMI** by varying concentration of KO^tBu. The emission spectra were collected from a Cary Eclipse Fluorescence Spectrofluorometer. The **NMI** was dissolved in deaerated xylene and the solution was excited at 345 nm. The monitoring wavelength was the corresponding maxima of the emission band; $\lambda_{max} = 382$ nm (for **NMI**). Relative fluorescence intensities were measured for solutions containing **NMI** (6.7× 10⁻⁵ M) with varying KO^tBu concentration (15 mM to 65 mM). There was no change in the shape of the fluorescence spectrum but the intensity was decreasing gradually with increasing quencher (KO^tBu) concentration.

Preparation of stock sample: In a Schlenk flask, 2×10^{-3} M solution of **NMI** was prepared in 5 mL dry xylene. The stock solution was diluted to 6.7×10^{-5} M in 3 mL of dry xylene. For each quencher concentration, samples were prepared separately. The Stern-Volmer relationship (equation 1) was established by taking the ratio of the integrated fluorescence intensities (I_0/I_n) in the absence and presence of electron donors and the concentration of donors used as quenchers [Q]. In this present case, the [Q] is KO^tBu.



Figure S7. Fluorescence emission spectra of **NMI** (λ_{max} - 381 nm) and its intensity quenching by addition of successive aliquots of KO^tBu. Experiments were carried out in oxygen-free Xylene solution at 298 K with **NMI** concentration of 10⁻⁵ M, excited at 345 nm.

$$I_0/I = 1 + K_{SV}[Q]....(1)$$

Where I_0 is the fluorescence intensity without the quencher, I is the intensity in the presence of the quencher, [Q] is the concentration of added quencher and K_{SV} is the Stern-Volmer quenching constant.



Figure S8. Stern-Volmer quenching of **NMI** showing a linear correlation with increasing quencher (KO^tBu) concentration.

8. Detection of H₂O₂ during alcohol oxidation:

For oxidation of alcohols, presence of H_2O_2 in the reaction mixture was analyzed by UV–Vis spectroscopy² using the iodometric assay based on peak of I_3^- at $\lambda_{max} = 345$ nm; $\varepsilon = 26\ 000$ M⁻¹ cm⁻¹ upon reaction with KI. In a typical reaction, a 5 mL vial was charged with benzyl alcohol (1 mmol), KO^tBu (1 mmol), **NMI** (10 mol%) in 5 mL dry xylene and was closed with a rubber septum. The resulting solution was spurged with O₂. The reaction mixture was stirred for 6 h under blue light. To the reaction mixture, 10 mL of water + 10 mL of DCM was added. The aqueous part was then separated. To the separated aqueous layer, dilute H_2SO_4 (pH = 2) was added to stop further oxidation. Then, 1 mL of a 10% solution of KI and a few drops of a 3% solution of ammoniummolybdate was added. The produced H_2O_2 oxidizes I⁻ to I₂, which reacts with an excess of I⁻ to form I₃⁻. The chemical reactions are as follows²

- (i) $H_2O_2 + 2I^- + 2H^+ \rightarrow 2H_2O + I_2$
- (ii) $I_2(aq) + I^- \rightarrow I_3^-$



Figure S9: UV-Visible spectrum of I_3^- ion formation in the presence of H_2O_2 .

9. Mechanistic insight into the reaction:

a) Photo catalysed alcohol dehydrogenation:



A 15 mL vial tube was charged with 10 mol% of **NMI**, KO^tBu (0.5 mmol), and 4-methoxy benzyl alcohol (0.5 mmol) in 2 mL xylene. The reaction mixture was stirred under blue light irradiation for 6 h in an open atmosphere. Upon completion, the reaction mixture was worked up in ethyl acetate and brine mixture. Then the organic layer was collected, dried and analyzed by ¹H NMR spectroscopy. It is found that 86% of 4-methoxy benzaldehyde formed after 6 h under photochemical condition.

b) KO^tBu mediated hydration of *O*-aminobenzonitrile (1):



A 15 mL vial was charged with 0.5 mmol of 2-aminobenzonitrile and KO^tBu (0.5 mmol) in 2 mL xylene. The reaction mixture was stirred in an open atmosphere for 12 h. Upon completion, the reaction mixture was extracted with ethyl acetate and evaporated *in vacuo*. Then the residue was purified by column chromatography on silica gel using hexane and ethyl acetate as eluent (3:2), giving 63% isolated yield of 2-aminobenzamide.

c) Synthesis of the intermediate 2-phenyl-2,3-dihydroquinazolin-4(1H)-one:



A 15 mL vial tube was charged with 0.5 mmol of 2-aminobenzamide and KO^tBu (0.5 mmol) in 2 mL xylene. The reaction mixture was stirred in an open atmosphere for 12 h. Upon completion, the reaction mixture was extracted with ethyl acetate and evaporated *in vacuo*. Then the resultant residue was purified by column chromatography using silica gel employing ethyl acetate in hexane (3:2) as an eluent. The intermediate product **11** was isolated in 60% yield. This control reaction proves that this condensation and cyclization event only requires a base. The molecule **11** was fully characterized by ¹H and ¹³C NMR spectroscopies.



d) NMI catalysed dehydrogenation of intermediate 2-phenyl-2,3-dihydroquinazolin-4(1*H*)-one, 11:



A 15 mL vial tube was charged with 0.5 mmol of 2-phenyl-2,3-dihydroquinazolin-4(1*H*)-one, 11, 10 mol% of catalyst **NMI** and KO^tBu (0.5 mmol) in 2 mL xylene. The reaction mixture was stirred under blue light irradiation in an open atmosphere for 12 h. Upon completion, the reaction mixture was extracted with ethyl acetate and evaporated *in vacuo*. Then the resultant residue was purified by column chromatography using silica employing ethyl acetate in hexane (1:3) as an eluent. The desired product **3a** was separated in 92% isolated yield, suggesting effective dehydrogenation of the cyclic aminal intermediate by **NMI** under photoredox condition.

Analytical data:

(E)-1-benzylidene-1*H*-indene (3a) ³:



White solid, 88% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.55 (s, 1H), 8.17 (ddd, J = 9.4, 7.9, 1.5 Hz, 3H), 7.88 – 7.80 (m, 1H), 7.77 – 7.72 (m, 1H), 7.60 – 7.49 (m, 4H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.5, 152.5, 149.0, 134.8, 133.0, 131.6, 128.8, 128.0, 128.0, 127.7, 126.8, 126.1, 121.2.

2-(p-tolyl) quinazolin-4(3H)-one (3b) ³:



White solid, 81% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.59 (s, 1H), 8.26 (dd, J = 7.9, 1.5 Hz, 1H), 8.23 – 8.18 (m, 2H), 7.94 (ddd, J = 8.6, 7.1, 1.6 Hz, 1H), 7.84 (dd, J = 8.3, 1.2 Hz, 1H), 7.62 (ddd, J = 8.2, 7.1, 1.2 Hz, 1H), 7.46 (d, J = 8.0 Hz, 2H), 2.62 (p, J = 1.9 Hz, 3H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.6, 152.6, 149.0, 141.8, 134.9, 130.2, 129.5, 128.0, 127.6, 126.7, 126.1, 121.1, 21.2

2-(4-isopropylphenyl) quinazolin-4(3H)-one (3c):



White solid, 76% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.48 (s, 1H), 8.22 – 8.06 (m, 3H), 7.82 (ddd, J = 8.5, 7.0, 1.6 Hz, 1H), 7.73 (dd, J = 8.3, 1.2 Hz, 1H), 7.50 (ddd, J = 8.1, 7.1, 1.2 Hz, 1H), 7.40 (d, J = 8.2 Hz, 2H), 2.96 (hept, J = 6.9 Hz, 1H), 1.23 (d, J = 6.9 Hz, 6H).¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.7, 152.7, 152.6, 149.3, 135.0, 130.8, 128.3, 127.9, 127.0, 126.9, 126.3, 121.4, 33.8, 24.0.

2-(4-ethylphenyl) quinazolin-4(3H)-one (3d):



White solid, 83% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.48 (s, 1H), 8.15 (dd, J = 7.9, 1.6 Hz, 1H), 8.13 – 8.08 (m, 2H), 7.82 (ddd, J = 8.5, 7.0, 1.6 Hz, 1H), 7.72 (dd, J = 8.3, 1.2 Hz, 1H), 7.50 (ddd, J = 8.1, 7.0, 1.2 Hz, 1H), 7.40 – 7.34 (m, 2H), 2.68 (q, J = 7.6 Hz, 2H), 1.21 (t, J = 7.6 Hz, 3H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.7, 152.7, 149.3, 148.4, 135.4, 130.6, 128.5, 128.2, 127.9, 126.3, 121.4, 28.5, 15.8

2-(4-(tert-butyl)phenyl)quinazolin-4(3H)-one (3e)⁴:



White solid, 87% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.50 (s, 1H), 8.14 (td, J = 7.0, 1.7 Hz, 3H), 7.83 (s, 1H), 7.73 (dd, J = 8.3, 1.2 Hz, 1H), 7.60 – 7.46 (m, 3H), 1.32 (s, 9H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.3, 154.3, 152.,2 148.9, 134.6, 129.9, 127.6, 127.5, 126.4, 125.9, 125.5, 120.9, 34.7, 30.9.

2-phenylquinazolin-4(3H)-one (3f)³:



White solid, 78% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.45 (s, 1H), 8.17 (dd, J = 7.9, 1.6 Hz, 1H), 7.86 – 7.81 (m, 1H), 7.69 (d, J = 8.1 Hz, 1H), 7.56 – 7.50 (m, 2H), 7.42 (dd, J = 7.4, 1.5 Hz, 1H), 7.37 – 7.32 (m, 2H), 2.38 (s, 3H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.0, 154.6, 148.9, 136.3, 134.7, 134.5, 130.8, 130.1, 129.4, 127.6, 126.9, 126.0, 125.9, 121.2, 19.8.

2-(4-methoxyphenyl) quinazolin-4(3H)-one (3g)³:



White solid, 73% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.42 (s, 1H), 8.21 – 8.17 (m, 2H), 8.13 (d, J = 6.5 Hz, 1H), 7.81 (s, 1H), 7.73 – 7.68 (m, 1H), 7.48 (s, 1H), 7.09 (d, J = 8.9 Hz, 2H), 3.85 (s, 3H) ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.9, 161.9, 152.6, 149.2, 135.3, 129.5, 127.3, 126.2, 124.0, 120.7, 114.8, 56.3.

2-(2-methoxyphenyl) quinazolin-4(3H)-one (3h)³:



White solid, 70% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.11 (s, 1H), 8.15 (dd, J = 8.0, 1.5 Hz, 1H), 7.83 (s, 1H), 7.70 (dt, J = 7.9, 1.5 Hz, 2H), 7.53 (dt, J = 2.9, 0.9 Hz, 2H), 7.19 (dd, J = 8.5, 1.0 Hz, 1H), 7.09 (d, J = 1.0 Hz, 1H), 3.86 (s, 3H). ¹³C {¹H} NMR (100 MHz, DMSO- d_6) δ 160.8, 156.7, 152.0, 148.6, 134.0, 131.8, 130.1, 127.0, 126.2, 125.4, 122.3, 120.6, 120.0, 111.5, 55.3.

2-(3-methoxyphenyl) quinazolin-4(3H)-one (3i)³:



White solid, 66% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.54 (s, 1H), 8.17 – 8.14 (m, 1H), 7.86 – 7.74 (m, 4H), 7.53 (t, J = 6.9 Hz, 1H), 7.46 (t, J = 8.0 Hz, 1H), 7.15 (dd, J = 7.8, 3.1 Hz, 1H), 3.86 (s, 3H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.1, 159.2, 152.9, 134.5, 133.9, 129.6, 127.3, 126.5, 125.7, 120.8, 120.0, 117.5, 112.3, 55.2.

2-(3,5-dimethoxyphenyl) quinazolin-4(3H)-one (3j)⁴:



White solid, 72% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.52 (s, 1H), 8.15 (dd, J = 7.9, 1.6 Hz, 1H), 7.88 – 7.81 (m, 1H), 7.78 – 7.73 (m, 1H), 7.53 (ddd, J = 8.1, 7.0, 1.2 Hz, 1H), 7.38 (d, J = 2.3 Hz, 2H), 6.70 (t, J = 2.3 Hz, 1H), 3.84 (s, 6H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 163.1, 152.5, 152.1, 149.1, 135.3, 127.8, 126.9, 126.4, 125.2, 121.7, 121.1, 111.9, 111.1, 56.2.

2-(2-fluorophenyl) quinazolin-4(3H)-one (3k)³:



White solid, 60% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.59 (s, 1H), 8.18 (d, J = 7.7 Hz, 1H), 7.91 – 7.68 (m, 3H), 7.58 (dt, J = 21.2, 7.1 Hz, 2H), 7.38 (q, J = 8.9 Hz, 2H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.36, 160.67, 158.18 (d, ¹J_{C-F} = 249 Hz), 149.78, 148.52, 134.43, 132.73, 132.64, 130.9, 130.9, 127.3, 126.9, 125.7, 124.5, 124.4, 122.2, 122.1, 121.0, 116.1, 115.9.

2-(2-chlorophenyl) quinazolin-4(3H)-one (3l)³:



White solid, 71% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.64 (s, 1H), 8.18 (dd, J = 7.9, 1.6 Hz, 1H), 7.89 – 7.82 (m, 1H), 7.69 (ddd, J = 15.8, 7.8, 1.4 Hz, 2H), 7.64 – 7.55 (m, 3H), 7.51 (dd, J = 7.4, 1.5 Hz, 1H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.4, 152.3, 148.6, 134.6, 133.8, 131.6, 131.5, 130.9, 129.6, 127.5, 127.3, 127.1, 125.9, 121.3.

2-(2-bromophenyl) quinazolin-4(3H)-one (3m)³:



White solid, 66% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.65 (s, 1H), 8.18 (dd, J = 8.0, 1.6 Hz, 1H), 7.86 (ddd, J = 8.5, 7.2, 1.6 Hz, 1H), 7.70 (ddd, J = 15.3, 7.9, 1.5 Hz, 2H), 7.65 – 7.53 (m, 3H), 7.50 (td, J = 7.4, 1.4 Hz, 1H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.4, 152.3, 148.6, 134.6, 133.8, 131.7, 130.9, 129.6, 127.5, 127.1, 125.9, 121.3.

2-(2-(trifluoromethyl) phenyl) quinazolin-4(3H)-one (3n)⁵:



White solid, 73% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.67 (s, 1H), 8.15 (dd, J = 8.1, 1.5 Hz, 1H), 7.89 – 7.86 (m, 1H), 7.84 – 7.81 (m, 1H), 7.80 – 7.77 (m, 1H), 7.76 – 7.72 (m, 2H), 7.65 (dd, J = 8.3, 1.2 Hz, 1H), 7.57 – 7.52 (m, 1H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.6, 152.6, 148.6, 134.9, 132.7, 130.9, 130.7, 127.7, 127.4 (q, ²J_{C-F} = 31 Hz), 126.7, 126.6 (q, ³J_{C-F} = 4 Hz), 126.1,125.4 122.7 (q, ¹J_{C-F} = 272.1 Hz), 121.4. ¹⁹F NMR (376 MHz, DMSO- d_6) δ -57.08.

2-(4-fluorophenyl) quinazolin-4(3H)-one (3o)³:



White solid, 71% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.57 (s, 1H), 8.31 – 8.19 (m, 2H), 8.15 (dd, J = 7.9, 1.5 Hz, 1H), 7.89 – 7.79 (m, 1H), 7.73 (d, J = 8.1 Hz, 1H), 7.52 (t, J = 7.5 Hz, 1H), 7.39 (t, J = 8.8 Hz, 2H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 164.7, 162.2 (d, ¹J_{C-} F = 248 Hz), 161.7, 150.8, 148.0, 134.0, 129.8, 129.7, 128.6, 128.6, 126.9, 126.0, 125.3, 120.3, 115.1, 114.9.

2-(4-chlorophenyl) quinazolin-4(3H)-one(3p)³:



White solid, 72% yield. ¹H NMR (400 MHz, DMSO-d6) δ 12.63 (s, 1H), 8.22 – 8.19 (m, 2H), 8.16 (dd, J = 7.9, 1.6 Hz, 1H), 7.85 (d, J = 1.6 Hz, 1H), 7.75 (d, J = 8.1 Hz, 1H), 7.65 – 7.61

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(m, 2H), 7.54 (s, 1H). ${}^{13}C{}^{1}H$ NMR (100 MHz, DMSO-*d*₆) δ 162.2, 151.4, 148.6, 136.3, 134.7, 131.6, 129.7, 128.7, 127.6, 126.8, 125.9, 121.0.

2-(4-bromophenyl) quinazolin-4(3H)-one (3q)³:



White solid, 65% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.62 (s, 1H), 8.24 – 8.13 (m, 3H), 7.84 (dd, J = 6.9, 1.5 Hz, 1H), 7.77 – 7.70 (m, 1H), 7.67 – 7.59 (m, 2H), 7.57 – 7.51 (m, 1H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.2, 151.3, 148.5, 136.3, 134.7, 131.5, 129.6, 128.7, 127.5, 126.8, 125.8, 120.9.

2-cyclobutylquinazolin-4(3H)-one (3r)⁷:



White solid, 81% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.06 (s, 1H), 8.07 (dd, J = 7.9, 1.6 Hz, 1H), 7.84 – 7.70 (m, 1H), 7.70 – 7.57 (m, 1H), 7.51 – 7.37 (m, 1H), 3.49 (t, J = 8.6 Hz, 1H), 2.40 (td, J = 9.2, 2.6 Hz, 2H), 2.28 – 2.19 (m, 2H), 2.04 – 1.91 (m, 1H), 1.87 – 1.77 (m, 1H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.7, 159.1, 148.9, 135.1, 127.0, 126.0, 125.3, 123.1, 37.5, 26.2, 19.3.

2-cyclopentylquinazolin-4(3H)-one (3s) 7:



White solid, 88% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.13 (s, 1H), 8.07 (d, J = 7.9 Hz, 1H), 7.74 (d, J = 7.6 Hz, 1H), 7.58 (d, J = 8.1 Hz, 1H), 7.44 (d, J = 7.5 Hz, 1H), 3.08 – 2.92

(m, 1H), 2.02 - 1.83 (m, 4H), 1.80 - 1.54 (m, 4H). ${}^{13}C{}^{1}H$ NMR (100 MHz, DMSO- d_6) δ 161.7, 160.2, 148.7, 134.0, 126.8, 125.7, 125.5, 120.7, 43.6, 30.6, 25.2.

2-cyclohexylquinazolin-4(3H)-one (3t)⁶:



White solid, 78% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.10 (s, 1H), 8.07 (dd, J = 7.9, 1.5 Hz, 1H), 7.76 (ddd, J = 8.5, 7.1, 1.6 Hz, 1H), 7.59 (dd, J = 8.3, 1.2 Hz, 1H), 7.48 – 7.39 (m, 1H), 2.57 (s, 1H), 1.96 – 1.74 (m, 4H), 1.58 (dd, J = 12.3, 3.3 Hz, 3H), 1.38 – 1.15 (m, 3H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.0, 160.8, 149.0, 134.3, 127.0, 126.0, 125.7, 121.0, 42.9, 30.2, 25.5, 25.4.

2-(naphthalen-1-yl) quinazolin-4(3H)-one (3u)³:



White solid, 68% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.68 (s, 1H), 8.22 (dd, J = 8.0, 1.5 Hz, 1H), 8.19 – 8.09 (m, 2H), 8.08 – 8.00 (m, 1H), 7.87 (ddd, J = 8.5, 7.1, 1.6 Hz, 1H), 7.80 (dd, J = 7.2, 1.3 Hz, 1H), 7.74 (dd, J = 8.2, 1.2 Hz, 1H), 7.67 – 7.56 (m, 4H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.0, 153.7, 148.7, 134.6, 133.2, 131.8, 130.4, 130.3, 128.4, 127.7, 127.5, 127.1, 126.9, 126.4, 125.9, 125.3, 125.1, 121.3.

2-(anthracen-9-yl) quinazolin-4(3*H*)-one (3v)³:



White solid, 74% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.86 (s, 1H), 8.84 (s, 1H), 8.32 (dd, J = 8.0, 1.5 Hz, 1H), 8.25 – 8.19 (m, 2H), 7.94 – 7.88 (m, 1H), 7.79 (ddd, J = 17.8, 8.5, 1.2 Hz, 3H), 7.66 (ddd, J = 8.1, 7.2, 1.2 Hz, 1H), 7.61 – 7.53 (m, 4H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.8, 153.1, 148.9, 134.6, 130.6, 129.1, 128.8, 128.6, 127.6, 127.1, 126.1, 125.7, 125.1, 121.7.

2-([1,1'-biphenyl]-4-yl)quinazolin-4(3H)-one (3w)³:



White solid, 68% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.60 (s, 1H), 8.32 – 8.28 (m, 2H), 8.17 (dd, J = 7.9, 1.5 Hz, 1H), 7.88 – 7.84 (m, 3H), 7.80 – 7.76 (m, 3H), 7.55 – 7.50 (m, 3H), 7.44 (d, J = 7.3 Hz, 1H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.4, 142.9, 139.0, 134.8, 131.6, 129.2, 128.5, 128.3, 126.9, 126.9, 126.7, 126.0.

2-(furan-2-yl)quinazolin-4(3*H*)-one (3x)³:



White solid, 71% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.49 (s, 1H), 8.11 (dd, J = 7.9, 1.5 Hz, 1H), 7.98 (d, J = 1.7 Hz, 1H), 7.83 – 7.74 (m, 1H), 7.67 (d, J = 8.1 Hz, 1H), 7.62 (d, J = 3.6 Hz, 1H), 7.47 (t, J = 7.5 Hz, 1H), 6.73 (dd, J = 3.6, 1.7 Hz, 1H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.6, 148.7, 146.6, 146.1, 144.0, 134.6, 127.3, 126.5, 126.0, 121.2, 114.5, 112.5.

2-(thiophen-2-yl)quinazolin-4(3*H*)-one (3y)³:



White solid, 73% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.48 (s, 1H), 8.60 (dd, J = 2.9, 1.3 Hz, 1H), 8.13 (dd, J = 8.0, 1.5 Hz, 1H), 7.87 (s, 1H), 7.82 (ddd, J = 8.5, 7.1, 1.6 Hz, 1H), 7.75 – 7.68 (m, 2H), 7.50 (ddd, J = 8.1, 7.1, 1.2 Hz, 1H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.7, 148.5, 147.9, 135.0, 134.2, 128.3, 127.0, 126.9, 126.6, 126.0, 125.5, 120.6.

2-(1-methyl-1H-pyrrol-2-yl) quinazolin-4(3H)-one (3z)6:



White solid, 60% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.07 (s, 1H), 8.09 (dd, J = 7.9, 1.6 Hz, 1H), 7.77 (ddd, J = 8.4, 7.1, 1.6 Hz, 1H), 7.62 (dd, J = 8.2, 1.1 Hz, 1H), 7.42 (ddd, J = 8.0, 7.1, 1.2 Hz, 1H), 7.22 (dd, J = 4.0, 1.8 Hz, 1H), 7.10 (t, J = 2.1 Hz, 1H), 6.15 (dd, J = 4.0, 2.5 Hz, 1H), 4.07 (s, 3H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.4, 148.2, 146.2, 133.8, 129.5, 125.2, 125.1, 123.3, 119.8, 114.2, 107.1, 37.0.

2-(benzofuran-5-yl) quinazolin-4(3H)-one (3za):



White solid, 64% yield.¹H NMR (400 MHz, DMSO- d_6) δ 12.70 (s, 1H), 10.11 (s, 1H), 8.39 – 8.35 (m, 2H), 8.17 (dd, J = 7.9, 1.5 Hz, 1H), 8.08 – 8.05 (m, 2H), 7.87 (ddd, J = 8.4, 7.0, 1.6 Hz, 1H), 7.78 (dd, J = 8.2, 1.2 Hz, 1H), 7.56 (ddd, J = 8.1, 7.0, 1.2 Hz, 1H).¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 192.9, 162.2, 151.5, 148.5, 137.9, 137.8, 134.8, 129.6, 128.6, 128.4, 127.8, 127.2, 126.0, 121.2. **HRMS** (ESI, m/z) calcd. for C₁₆H₁₀N₂O₂ [M+H]+ : 263.0815; found: 263.1261

2-(1H-indol-5-yl) quinazolin-4(3H)-one (3zb):



White solid, 67% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 11.64 (s, 1H), 8.41 (s, 1H), 8.19 (dd, J = 8.1, 1.5 Hz, 1H), 7.95 – 7.91 (m, 1H), 7.88 – 7.80 (m, 2H), 7.70 – 7.58 (m, 3H), 7.57 (q, J = 3.2 Hz, 1H), 6.68 (d, J = 2.5 Hz, 1H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.5, 160.2, 157.1, 138.9, 136.0, 128.7, 128.4, 127.8, 127.2, 126.9, 123.3, 122.8, 122.0, 120.1, 112.4, 103.2. **HRMS** (ESI, m/z) calcd. for C₁₆H₁₁N₃O [M + H]⁺ : 262.0975; found: 262.0985

2-(pyridin-2-yl) quinazolin-4(3H)-one (3zc)³:



White solid, 59% yield.¹H NMR (400 MHz, DMSO- d_6) δ 11.87 (s, 1H), 8.79 – 8.75 (m, 1H), 8.48 – 8.45 (m, 1H), 8.20 (dd, J = 8.0, 1.5 Hz, 1H), 8.09 (d, J = 1.7 Hz, 1H), 7.91 – 7.80 (m, 2H), 7.70 – 7.65 (m, 1H), 7.58 (s, 1H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.1, 150.2, 149.3, 149.0, 148.7, 138.3, 135.0, 128.0, 127.6, 126.8, 126.4, 122.4, 122.3.

(E)-2-(4-styrylphenyl) quinazolin-4(3H)-one (3zd)⁴:



White solid, 63% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.35 (s, 1H), 8.11 (dd, J = 8.0, 1.6 Hz, 1H), 7.95 (d, J = 16.2 Hz, 1H), 7.80 (d, J = 7.0 Hz, 1H), 7.67 (dd, J = 8.0, 6.0 Hz, 3H), 7.50 – 7.42 (m, 4H), 7.01 (d, J = 16.2 Hz, 1H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.8, 151.4, 149.0, 138.3, 135.0, 134.6, 129.8, 129.1, 127.7, 127.2, 126.3, 125.9, 121.1, 121.1.

2-benzylquinazolin-4(3*H*)-one (3ze):



White solid, 75% yield, ¹H NMR (400 MHz, DMSO- d_6) δ 12.44 (s, 1H), 8.08 (dd, J = 8.0, 1.5 Hz, 1H), 7.77 (s, 1H), 7.61 (dd, J = 8.3, 1.1 Hz, 1H), 7.47 (d, J = 1.0 Hz, 1H), 7.41 – 7.36 (m, 2H), 7.32 (t, J = 7.6 Hz, 2H), 7.25 (d, J = 7.2 Hz, 1H), 3.94 (s, 2H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.91, 156.01, 148.94, 136.60, 134.44, 128.91, 128.54, 126.96, 126.84, 126.26, 125.74, 120.77, 40.80.

2-(quinolin-2-yl)quinazolin-4(3H)-one (3zf):



Yellow solid, 78% yield. ¹H NMR (400 MHz, Chloroform-d) δ 11.22 (s, 1H), 8.62 (d, J = 8.6 Hz, 1H), 8.44 – 8.26 (m, 2H), 8.13 (d, J = 8.5 Hz, 1H), 7.92 – 7.84 (m, 2H), 7.84 – 7.72 (m, 2H), 7.62 (dd, J = 8.3, 6.8 Hz, 1H), 7.58 – 7.47 (m, 1H). ¹³C NMR {¹H} (100 MHz, Chloroform-d) δ 161.6, 149.2, 149.1, 148.1, 146.9, 137.7, 134.7, 130.6, 129.8, 129.4, 128.4, 128.3, 127.9, 127.7, 126.9, 122.7, 118.5.

6-chloro-2-phenylquinazolin-4(3H)-one (7a)⁴:



White solid, 87% yield.¹H NMR (400 MHz, DMSO- d_6) δ 12.73 (s, 1H), 8.20 – 8.15 (m, 2H), 8.08 (d, J = 2.5 Hz, 1H), 7.85 (d, J = 2.6 Hz, 1H), 7.76 (d, J = 8.7 Hz, 1H), 7.61 – 7.53 (m, 3H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.8, 151.4, 149.0, 138.3, 135.0, 134.5, 129.82, 129.1, 127.7, 127., 126.3, 125.9, 121.1, 121.1.

6-chloro-2-(o-tolyl) quinazolin -4(3H)-one (7b):



White solid, 71% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.62 (s, 1H), 8.09 (d, J = 2.5 Hz, 1H), 7.85 (dd, J = 8.7, 2.5 Hz, 1H), 7.70 (d, J = 8.7 Hz, 1H), 7.49 (dd, J = 7.6, 1.4 Hz, 1H), 7.42 (dd, J = 7.4, 1.5 Hz, 1H), 7.36 – 7.29 (m, 2H), 2.37 (s, 3H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 160.5, 152.8, 147.3, 134.8, 133.6, 131.8, 131.4, 131.4, 130.9, 129.8, 129.6, 127.3, 124.9, 23.3.

6-chloro-2-(p-tolyl) quinazolin-4(3H)-one (7c)⁷:



White solid, 77% yield.¹H NMR (400 MHz, DMSO- d_6) δ 12.63 (s, 1H), 8.10 (d, J = 2.5 Hz, 1H), 7.87 (dd, J = 8.7, 2.5 Hz, 1H), 7.72 (d, J = 8.7 Hz, 1H), 7.51 (dd, J = 7.6, 1.4 Hz, 1H), 7.43 (dd, J = 7.4, 1.5 Hz, 1H), 7.37 – 7.30 (m, 2H), 2.38 (s, 3H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.0, 155.1, 136.4, 134.8, 134.2, 131.1, 130.8, 130.3, 129.4, 125.9, 125.0, 19.8.

6-chloro-2-(4- methoxyphenyl) quinazolin-4(3H)-one (7d)8:



White solid, 64% yield. ¹H NMR (400 MHz, DMSO- d_6) δ 12.59 (s, 1H), 8.20 – 8.17 (m, 2H), 8.06 (d, J = 2.5 Hz, 1H), 7.84 (dd, J = 8.7, 2.6 Hz, 1H), 7.72 (d, J = 8.7 Hz, 1H), 7.11 – 7.08 (m, 2H), 3.85 (s, 3H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.1, 161.4, 152.4, 147.8, 134.7, 130.3, 129.5, 124.8, 124.5, 121.9, 114.1, 55.5.

6-chloro-2-(4-chlorophenyl) quinazolin-4(3H)-one (7e)8:



White solid,68% yield.¹H NMR (400 MHz, DMSO- d_6) δ 12.84 (s, 1H), 8.11 (d, J = 2.5 Hz, 1H), 7.89 (dd, J = 8.7, 2.6 Hz, 1H), 7.74 (d, J = 8.8 Hz, 1H), 7.67 (dd, J = 7.5, 1.7 Hz, 1H), 7.62 (dd, J = 8.0, 1.4 Hz, 1H), 7.57 (td, J = 7.6, 1.7 Hz, 1H), 7.50 (td, J = 7.4, 1.4 Hz, 1H). ¹³C NMR {¹H} (100 MHz, DMSO- d_6) δ 160.5, 152.8, 147.3, 134.8, 133.6, 131.8, 131.4, 130.9, 129.8, 129.6, 127.3, 124.9.

2-(4-chlorophenyl)-7-methylquinazolin-4(3H)-one (8a)⁹:



White solid,78% yield.¹H NMR (400 MHz, DMSO-*d*₆) δ 12.46 (s, 1H), 8.17 (dt, *J* = 6.6, 1.7 Hz, 2H), 8.03 (d, *J* = 8.1 Hz, 1H), 7.65 – 7.49 (m, 4H), 7.33 (dd, *J* = 8.1, 1.7 Hz, 1H), 2.46 (s, 3H). ¹³C{¹H} NMR (100 MHz, DMSO-*d*₆) δ 162.6, 152.8, 149.3, 145.5, 133.2, 131.8, 129.1, 128.5, 128.2, 127.6, 126.2, 119.0, 21.5.

2-(4-methoxyphenyl)-7-methylquinazolin-4(3*H*)-one (8b)¹⁰:



White solid, 84% yield.¹H NMR (400 MHz, DMSO- d_6) δ 12.33 (s, 1H), 8.20 – 8.16 (m, 2H), 8.01 (d, J = 8.0 Hz, 1H), 7.51 (d, J = 1.5 Hz, 1H), 7.30 (dd, J = 8.1, 1.7 Hz, 1H), 7.10 – 7.06 (m, 2H), 3.85 (s, 3H), 2.46 (s, 3H).¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.9, 151.9, 149.1, 145.0, 129.4, 127.6, 126.9, 125.7, 124.9, 118.3, 114.0, 55.4, 21.4.

7-methyl-2-(p-tolyl) quinazolin-4(3H)-one (8c) ⁷:



White solid,82% yield.¹H NMR (400 MHz, DMSO- d_6) δ 12.38 (s, 1H), 8.08 (d, J = 7.9 Hz, 2H), 8.02 (d, J = 8.0 Hz, 1H), 7.53 (s, 1H), 7.33 (t, J = 8.3 Hz, 3H), 2.46 (s, 3H), 2.38 (s, 3H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 162.2, 152.3, 149.0, 145.1, 141.5, 130.0, 129.3, 127.3, 127.7, 127.1, 125.8, 118.5, 21.4, 21.1.

3-phenyl-2*H*-benzo[e][1,2,4]thiadiazine 1,1-dioxide (9a)³:



White solid,62% yield.¹H NMR (400 MHz, DMSO- d_6) δ 12.22 (s, 1H), 8.08 – 8.03 (m, 2H), 7.89 – 7.85 (m, 1H), 7.76 – 7.69 (m, 2H), 7.64 (dt, J = 7.3, 2.2 Hz, 3H), 7.50 (s, 1H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 155.1, 135.8, 133.37, 133.1, 132.2, 129.1, 128.5, 126.5, 123.6, 121.7, 118.8.

3-(p-tolyl)-2*H*-benzo[e][1,2,4]thiadiazine 1,1-dioxide (9b)³:



White solid,59% yield.¹H NMR (400 MHz, DMSO- d_6) δ 12.10 (s, 1H), 7.99 – 7.95 (m, 2H), 7.85 (dd, J = 7.9, 1.4 Hz, 1H), 7.77 – 7.71 (m, 1H), 7.64 (dd, J = 8.4, 1.1 Hz, 1H), 7.52 – 7.47 (m, 1H), 7.44 (d, J = 8.0 Hz, 2H), 2.42 (s, 3H).¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 154.7, 143.3, 135.5, 133.1, 129.5, 128.9, 128.3, 126.7, 123.34, 121.5, 118.4, 21.1.

3-(4-ethylphenyl)-2*H*-benzo[e][1,2,4]thiadiazine 1,1-dioxide (9c):

White solid,56% yield.¹H NMR (400 MHz, DMSO- d_6) δ 12.11 (s, 1H), 8.06 – 7.89 (m, 2H), 7.86 (dd, J = 8.0, 1.4 Hz, 1H), 7.73 (ddd, J = 8.6, 7.2, 1.5 Hz, 1H), 7.63 (dd, J = 8.5, 1.2 Hz, 1H), 7.54 – 7.37 (m, 3H), 2.72 (q, J = 7.6 Hz, 2H), 1.22 (d, J = 1.7 Hz, 3H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 154.7, 149.4, 135.6, 133.1, 129.2, 128.4, 128.3, 126.7, 123.4, 121.5, 118.5, 28.2, 15.3.

2-pentylquinazolin-4(3H)-one (10a):



White solid, 66% yield ¹H NMR (400 MHz, DMSO- $d_6 \delta$ 12.18 (s, 1H), 8.07 (dd, J = 7.9, 1.6 Hz, 1H), 7.76 (ddd, J = 8.5, 7.1, 1.6 Hz, 1H), 7.59 (dd, J = 8.2, 1.1 Hz, 1H), 7.45 (ddd, J = 8.1, 7.1, 1.2 Hz, 1H), 2.62 – 2.56 (m, 2H), 1.76 – 1.67 (m, 2H), 1.31 (dt, J = 7.5, 3.0 Hz, 4H), 0.90 – 0.84 (m, 3H).¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.85, 157.55, 148.99, 134.30, 126.82, 125.93, 125.70, 120.79, 34.49, 30.79, 26.53, 21.87, 13.89.

2-heptylquinazolin-4(3H)-one (10b):

White solid, 71% yield, ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.17 (s, 1H), 8.07 (dd, *J* = 7.9, 1.5 Hz, 1H), 7.76 (ddd, *J* = 8.5, 7.1, 1.6 Hz, 1H), 7.58 (dd, *J* = 8.3, 1.1 Hz, 1H), 7.47 – 7.43 (m, 1H), 2.61 – 2.55 (m, 2H), 1.71 (p, *J* = 7.3 Hz, 2H), 1.35 – 1.22 (m, 8H), 0.88 – 0.82 (m, 3H).). ¹³C{¹H} NMR (100 MHz, DMSO-*d*₆) δ 164.91, 157.54, 150.51, 133.75, 126.81, 125.93, 125.69, 122.53, 34.19, 30.75, 28.51, 26.81, 22.08, 16.84.

2-pentadecylquinazolin-4(3H)-one (10c):



White solid, 80% yield. ¹H NMR (400 MHz, CDCl₃) δ 11.66 (s, 1H), 8.28 (dd, J = 7.9, 1.5 Hz, 1H), 7.77 (ddd, J = 8.5, 7.0, 1.6 Hz, 1H), 7.70 (dd, J = 8.2, 1.2 Hz, 1H), 7.47 (ddd, J =

8.1, 6.9, 1.3 Hz, 1H), 2.86 – 2.72 (m, 2H), 1.94 – 1.82 (m, 2H), 1.49 – 1.41 (m, 2H), 1.38 – 1.20 (m, 22H), 0.87 (t, *J* = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 164.26, 157.06, 149.59, 134.93, 127.35, 126.48, 126.38, 120.65, 36.17, 32.06, 29.83, 29.79, 29.77, 29.61, 29.51, 29.42, 29.38, 27.73, 22.84, 14.28.

2-hexadecylquinazolin-4(3H)-one (10d):



White solid, 85% yield, ¹H NMR (400 MHz, CDCl₃) δ 12.06 (s, 1H), 8.29 (dd, J = 8.0, 1.5 Hz, 1H), 7.81 – 7.73 (m, 1H), 7.70 (dd, J = 8.3, 1.3 Hz, 1H), 7.46 (ddd, J = 8.1, 6.9, 1.3 Hz, 1H), 2.84 – 2.73 (m, 2H), 1.91 – 1.86 (m, 2H), 1.41 (dt, J = 37.2, 7.8 Hz, 4H), 1.32 – 1.21 (m, 22H), 0.87 (t, J = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 164.43, 157.10, 149.53, 134.79, 127.22, 126.31, 126.23, 120.49, 36.02, 31.94, 29.71, 29.68, 29.65, 29.50, 29.39, 29.30, 29.28, 27.64, 22.71, 14.15.

2-(2,4,4-trimethylpentyl)quinazolin-4(3*H*)-one (10e):



White solid, 72% yield, ¹H NMR (400 MHz, DMSO- d_6) δ 12.22 (s, 1H), 8.07 (dd, J = 8.0, 1.5 Hz, 1H), 7.76 (ddd, J = 8.5, 7.1, 1.6 Hz, 1H), 7.60 (dd, J = 8.3, 1.2 Hz, 1H), 7.44 (ddd, J = 8.2, 7.1, 1.2 Hz, 1H), 2.57 – 2.51 (m, 1H), 2.42 (dd, J = 13.5, 7.6 Hz, 1H), 2.15 (ddq, J = 10.8, 7.0, 3.5 Hz, 1H), 1.32 (dd, J = 14.0, 3.2 Hz, 1H), 1.09 (dd, J = 13.9, 6.8 Hz, 1H), 0.93 (d, J = 6.6 Hz, 3H), 0.82 (s, 9H). ¹³C{¹H} NMR (100 MHz, DMSO- d_6) δ 161.90, 156.70, 148.92, 134.31, 126.85, 125.95, 125.68, 120.74, 49.78, 43.89, 30.80, 29.78, 28.31, 22.53.



Figure S12. ¹H NMR spectrum (400 MHz) of 3a in DMSO-d₆



Figure S13. ¹³C NMR spectrum (100 MHz) of 3a in DMSO-d₆



Figure S14. ¹H NMR spectrum (400 MHz) of **3b** in DMSO-d₆.



Figure S15. ¹³C NMR spectrum (100 MHz) of **3b** in DMSO-d₆



Figure S16. ¹H NMR spectrum (400 MHz) of 3c in DMSO-d₆



Figure S17. ¹³C NMR spectrum (100 MHz) of 3c in DMSO-d₆



Figure S18. ¹H NMR spectrum (400 MHz) of 3d in DMSO-d₆



Figure S19. ¹³C NMR spectrum (100 MHz) of 3d in DMSO-d₆



Figure S20. ¹H NMR spectrum (400 MHz) of 3e in DMSO-d₆



Figure S21. ¹³C NMR spectrum (100 MHz) of 3e in DMSO-d₆


Figure S22. ¹H NMR spectrum (400 MHz) of 3f in DMSO-d₆



Figure S23. ¹³C NMR spectrum (100 MHz) of 3f in DMSO-d₆



Figure S24. ¹H NMR spectrum (400 MHz) of 3g in DMSO-d₆





Figure S26. ¹H NMR spectrum (400 MHz) of **3h** in DMSO-d₆



Figure S27. ¹³C NMR spectrum (100 MHz) of **3h** in DMSO-d₆



Figure S28. ¹H NMR spectrum (400 MHz) of 3i in DMSO-d₆



Figure S29. ¹³C NMR spectrum (100 MHz) of 3i in DMSO-d₆



Figure S31. ¹³C NMR spectrum (100 MHz) of 3j in DMSO-d₆



Figure S33. ¹³C NMR spectrum (100 MHz) of 3k in DMSO-d₆



Figure S34. ¹H NMR spectrum (400 MHz) of 3l in DMSO-d₆



Figure S35. ¹³C NMR spectrum (100 MHz) of 3l in DMSO-d₆



Figure S36. ¹H NMR spectrum (400 MHz) of 3m in DMSO-d₆



Figure S37. ¹³C NMR spectrum (100 MHz) of 3m in DMSO-d₆



Figure S38. ¹H NMR spectrum (400 MHz) of 3n in DMSO-d₆



Figure S39. ¹³C NMR spectrum (100 MHz) of 3n in DMSO-d₆



Figure S40. ¹⁹F NMR spectrum (376 MHz) of **3n** in DMSO-d₆



Figure S41. ¹H NMR spectrum (400 MHz) of **30** in DMSO-d₆



Figure S42. ¹³C NMR spectrum (100 MHz) of **30** in DMSO-d₆



Figure S43. ¹H NMR spectrum (400 MHz) of 3p in DMSO-d₆



Figure S44. ¹³C NMR spectrum (100 MHz) of **3p** in DMSO-d₆



Figure S46. ¹³C NMR spectrum (100 MHz) of 3q in DMSO-d₆





Figure S50. ¹³C NMR spectrum (100 MHz) of 3s in DMSO-d₆



Figure S51. ¹H NMR spectrum (400 MHz) of 3t in DMSO-d₆



Figure S52. ¹³C NMR spectrum (100 MHz) of 3t in DMSO-d₆



Figure S54. ¹³C NMR spectrum (100 MHz) of **3u** in DMSO-d₆



Figure S56. ¹³C NMR spectrum (100 MHz) of 3v in DMSO-d₆



Figure S57. ¹H NMR spectrum (400 MHz) of 3w in DMSO-d₆



Figure S58. ¹³C NMR spectrum (100 MHz) of 3w in DMSO-d₆



Figure S59. ¹H NMR spectrum (400 MHz) of 3x in DMSO-d₆



Figure S60. ¹³C NMR spectrum (100 MHz) of 3x in DMSO-d₆



Figure S61. ¹H NMR spectrum (400 MHz) of 3y in DMSO-d₆



Figure S62. ¹³C NMR spectrum (100 MHz) of 3y in DMSO-d₆





Figure S64. $^{\rm 13}\rm C$ NMR spectrum (100 MHz) of 3z in DMSO-d_6



Figure S65. ¹H NMR spectrum (400 MHz) of 3za in DMSO-d₆



Figure S66. ¹³C NMR spectrum (100 MHz) of 3za in DMSO-d₆





Figure S67. ¹H NMR spectrum (400 MHz) of 3zb in DMSO-d₆



Figure S68. ¹³C NMR spectrum (100 MHz) of **3zb** in DMSO-d₆



Figure S69. ¹H NMR spectrum (400 MHz) of 3zc in DMSO-d₆



Figure S70. ¹³C NMR spectrum (100 MHz) of 3zc in DMSO-d₆



Figure S71. ¹H NMR spectrum (400 MHz) of 3zd in DMSO-d₆



Figure S72. ¹³C NMR spectrum (100 MHz) of 3zd in DMSO-d₆



Figure S73. ¹³C NMR spectrum (100 MHz) of 3ze in DMSO-d₆



Figure S74. ¹³C NMR spectrum (100 MHz) of 3ze in DMSO-d₆







Figure S76. ^{13}C NMR spectrum (100 MHz) of 3zf in DMSO-d6



Figure S77. ¹H NMR spectrum (400 MHz) of 7a in DMSO-d₆



Figure S78. ¹³C NMR spectrum (100 MHz) of 7a in DMSO- d_6



Figure S79. ¹H NMR spectrum (400 MHz) of 7b in DMSO-d₆



Figure S80. ¹³C NMR spectrum (100 MHz) of 7b in DMSO-d₆



Figure S81. ¹H NMR spectrum (400 MHz) of 7c in DMSO-d₆



Figure S82. ¹³C NMR spectrum (100 MHz) of 7c in DMSO-d₆



Figure S83. ¹H NMR spectrum (400 MHz) of 7d in DMSO-d₆



Figure S84. ¹³C NMR spectrum (100 MHz) of 7d in DMSO- d_6



Figure S85. ¹H NMR spectrum (400 MHz) of 7e in DMSO-d₆



Figure S86. ¹³C NMR spectrum (100 MHz) of 7e in DMSO-d₆



Figure S88. ¹³C NMR spectrum (100 MHz) of 8a in DMSO-d₆



Figure S90. ¹³C NMR spectrum (100 MHz) of 8b in DMSO-d₆



Figure S92. ¹³C NMR spectrum (100 MHz) of 8c in DMSO-d₆


Figure S93. ¹H NMR spectrum (400 MHz) of 9a in DMSO-d₆



Figure S94. ¹³C NMR spectrum (100 MHz) of 9a in DMSO-d₆



Figure S95. ¹H NMR spectrum (400 MHz) of 9b in DMSO-d₆



Figure S96. ¹³C NMR spectrum (100 MHz) of 9b in DMSO-d₆



Figure S97. ¹H NMR spectrum (400 MHz) of 9c in DMSO-d₆



Figure S98. ¹³C NMR spectrum (100 MHz) of 9c in DMSO-d6



Figure S100. ¹³C NMR spectrum (100 MHz) of 10a in DMSO-d₆









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Figure S108. ¹³C NMR spectrum (100 MHz) of 10e in DMSO-d₆

References:

- R. Regar, K.S. Mehra, Bhowal, and R. J. Sankar, *Eur. J. Org. Chem.* 2019, 47, 6278-6284.
- 2. A. K. Bains, Y. Ankit, and D. Adhikari. ChemSusChem. 2021, 14, 324.
- 3. P. R. Thorve and B. Maji, Catal. Sci. Technol. 2021,11, 1116-1124
- 4. Y. Hu, L. Chen and B. Li RSC Adv. 2016, 6, 65196-65204
- R.Cheng, L.Tang, T. Guo, D.Negrerie, Y. Du and K. Zhao RSC Adv. 2014, 4, 26434-26438
- W. Liu, G. Wu, W. Gao, J. Ding, X. Huang, M. Liu and H. Wu, Org. Chem. Front. 2018, 5, 2734-2738
- J. Sun, T. Tao, D. Xu, H. Cao, Q. Kong, X. Wang, Y. Liu, J. Zhao, Y. Wang, and Y. Pan, *Tetrahedron Lett.* 2018, 21, 2099-2102
- S.M. Patel, H. Chada, S. Biswal, S. Sharma, and D. S. Sharada, *Synthesis*. 2019, 51, 3160-3170
- M. Kumar, Richa, S. Sharma, V. Bhatt and N. Kumar, *Adv. Synth. Catal.*, 2015, 357, 2862 -2868
- C. B. Reddy, S. Ram, A. Kumar, R. Bharti and P. Das, *Chem.-Eur. J.*, 2019, 25, 4067 -4071