

## Supporting Information

# Engineered green alga *Chlamydomonas reinhardtii* as a whole-cell photosynthetic biocatalyst for stepwise photoproduction of H<sub>2</sub> and ε-caprolactone

Vilja Siitonen<sup>a</sup>, Anna Probst<sup>b</sup>, Gábor Tóth<sup>a</sup>, Robert Kourist<sup>c</sup>, Michael Schroda<sup>b</sup>, Sergey Kosourov<sup>a</sup> and Yagut Allahverdiyeva<sup>a\*</sup>

### Affiliations

a. Molecular Plant Biology, Department of Life Technologies, University of Turku, 20014 Turku, Finland. \*email: [allahve@utu.fi](mailto:allahve@utu.fi)

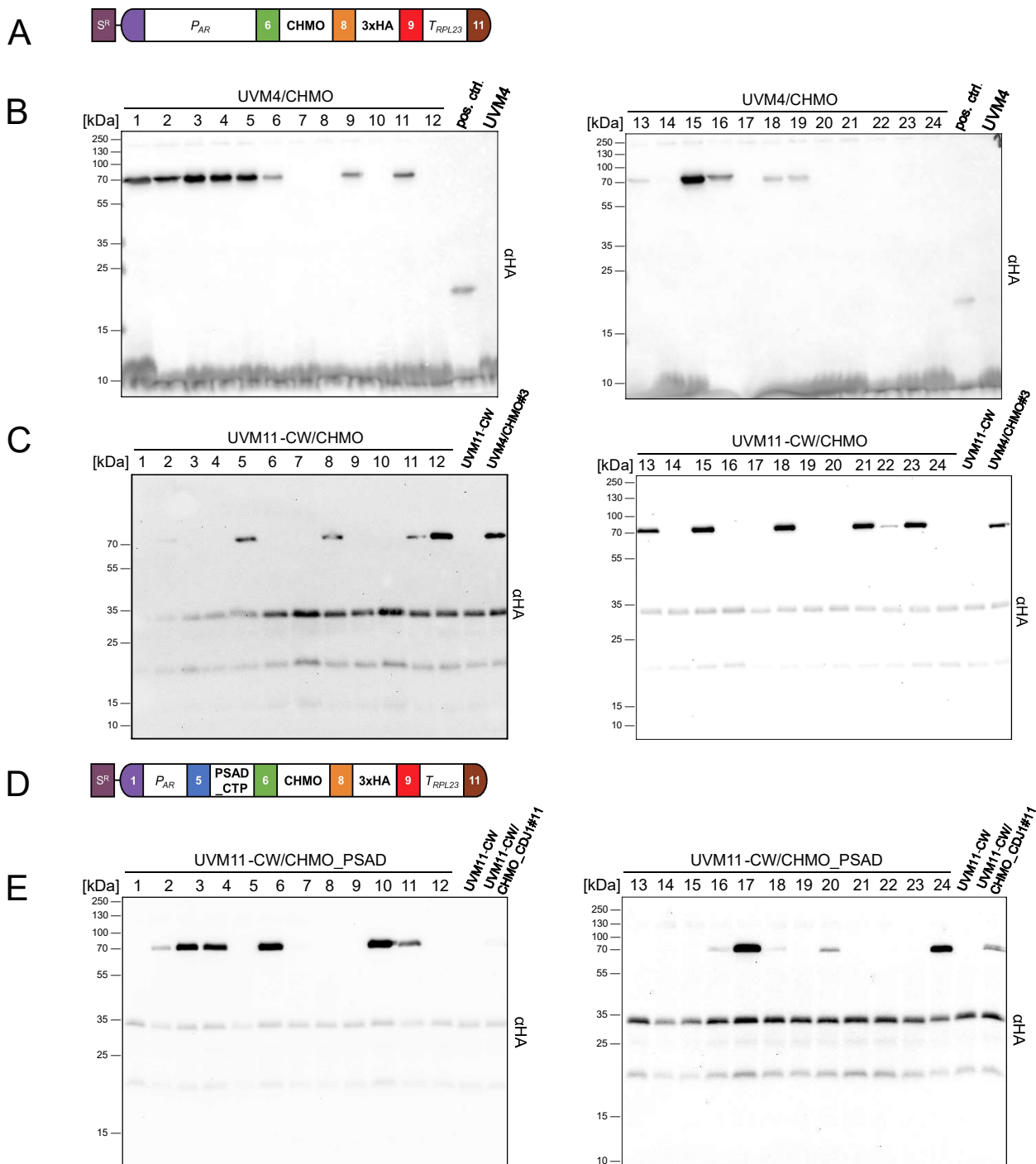
b. Molecular Biotechnology & Systems Biology, Rheinland-Pfälzische Technische Universität Kaiserslautern-Landau, 67663 Kaiserslautern, Germany

c. Institute of Molecular Biotechnology, Graz University of Technology, Petersgasse 14, 8010, Graz, Austria.

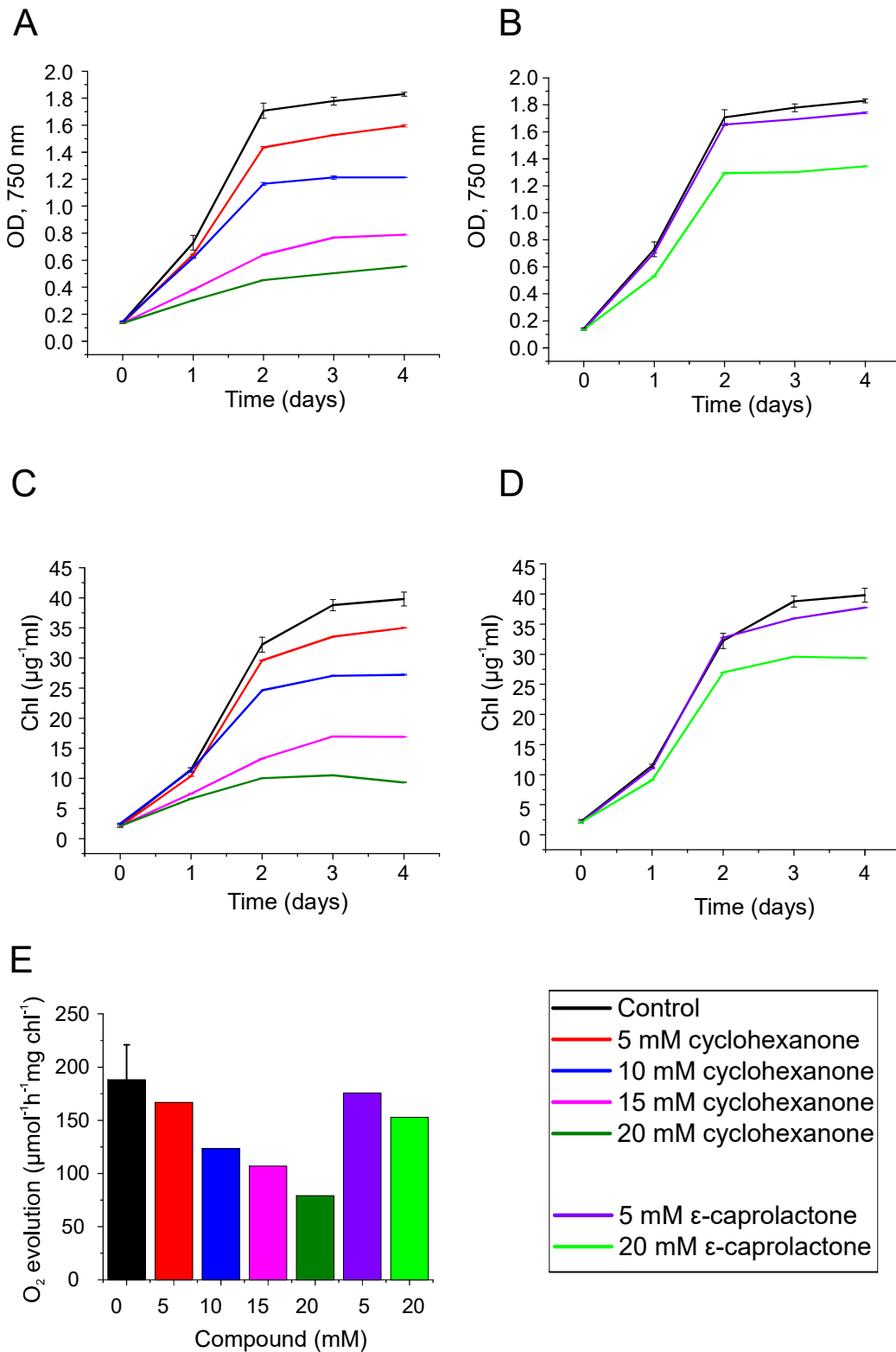
CDS CHMO  
1st Intron RBCS2  
2nd Intron RBCS2  
3rd Intron RBCS2  
Plasmid backbone pAGM1287

ACTCTGTGGTCTCAaatgagccagaagatggacttcgacgccatcgtTatcggcggcggttcggcggcctgtacgccgtgaagaagctgcgcgac  
gagctggagctgaaggtgcaggccttcgacaaggccaccgcagctggccggcactggactggaaccgctaccccggcgccctgaccgacaccgag  
acGcacctgtactgctacagctgggacaaggagctgctgcagagcctggagatcaagaagaagtaagctgcagGTGAGTCGACGAGCAAGCCCGCG  
GATCAGGCAGCGTGTTCAGATTTGACTTGCAACGCCCGCATTTGTGTGCAGCAAGGCTTTTGGCTCCTCTGTCTGCTCAAGCAGCATCTAAC  
CCTGCGTCGCGGTTTTCCATTTGCAGggccccgacgtgcgcaagtacctgcagcaggtggccgagaagcagcagctgaagaagagctaccagttcaa  
caccgccgtgcagagcgcccactacaacgagggccgacgcctgtgggaggtgaccaccgagtaaggcgacaagtaaccgcccgttctctgatcac  
cgccctgggctgtgagcgcccccaacctgccaacatcaagggcatcaaccagttcaagggTgagctgaccacacTagccgTtggcccgcgca  
cgtgagcttcgagggTaagcgcgtgggctgtatcgccaccggcagcaccggcgtgcagGTGAGCTTGCAGGGTTCGAGCAACTCCAGCAACGA  
ACAGTGCCCAAGTCAGGAATTCGAGTCagccTGGGCTTTTCGGCGGCTTTTTCTTGGGCAACAGCTTGACTCATGCCAGCGCGGCTTGTCCAGC  
CTCACTTGTAGCTTTCCAGCTGCTACCAGCCGGGTATACGACAGCGACAGAGCCATAGCGTGGAACTACTTATTTGGGTTGCCGAAGTAGCGGTCCG  
GAGCGTGAGTTCTTGGTCAAGCCGCCCTTATCCGGTTCCTGTCCGTGTCTTTGTCCCTCGTTTACCCTTCGCGGCACCCTTCATCCCTTGTCTG  
CAGgtgatcacccgctggccccctggccaagcacctgacctgttccagcgcagcggcagtaacagctgcccacggaacgccccctgagc  
gaggagagctgaagaagatcaaggacaactacgacaagatctgggacggcgtgtggaacagcgcctggccttcggcctgaacgagagcaccgtg  
cccgcctgagcgtgagcgcgagggagcgaagggcgtgtcgagaaggcctggcagaccggcggcggttccgcttcatgttcgagacGttcggc  
gacatcgccaccaacatggagggcaacatcgagggccagaacttcatcaagggaagatcgccgagatcgtgaaggacccccgcatcgcccagaag  
ctgatgccccagGTAAGTCTGGCGAGAGCCCGACGGTCCACTGTGGCACTGGGTAGCTTTTGGCACACGGGTCCAtTGTGGCACTGGTTAGCTT  
GGCACCGGGACAGCGCCTATCTCACCGCGGGAACTGACGCATACCCCTGCTCGTGTTCAGCACggaaAAGCAAGGGGCCAAATTCATCTTTGG  
TGGTTCTGTGCGCTGGTGAACCTCTTCTCCCTCCCATTTCCCGTGGCGCCGACGacctgtacgccaagcggccccctgtgcgacagcggcta  
ctacaacaccttcaaccgcaaacgtgcgccctggaggagctgaaggccaaccccatcgtggagatcaccgagaacggcgtgaagctggagaacgg  
cgacttcgtggagctggacatgctgatctgcccaccggcttcgacgcccgtggcaggaactacgtgcatggacatccagggcaagaacggcct  
ggccatgaaggactactggaaggagggccccagcagctacatgggctgacctgaacaactacccaacatgttcatggtgctgggccccaacgg  
ccccttaccacacctgccccagcagcagagcaggtggagtgatcagcgacaccatccagtaaccgtggagaacaacgtggagagcagcga  
ggccaccaaggagggcggagagcagtgaccagacctgcccacacatcgccgagatgacctgttcccaaggcccagagctggatcttcggcgc  
caacatccccggcaagaagaacaccgtgtacttctacctggcgccctgaaggagtaccgcagcgcctggccaactgcaagaaccacgcctacga  
gggcttcgacatccagctgcagcgcagcagatcaagcagcccgcacgcttcgTGAGACCAGAAAGTGGCTCTTCAGTGGACGAAAGGGCCTCG  
TGATACGCCTATTTTTATAGTTAATGTATGATAATAATGGTTTCTTAGACGTCAGGTGGCACTTTTCGGGAAATGTGCGCGAAACCCCTATTT  
GTTTTATTTTCTAAATACATTCAAATATGTATCCGCTCATGAGACAATAACCTGATAAATGCTTCAATAATATGAAAAAGGAAGAGTATGCGCT  
CACGCAACTGGTCCAGAACCTTGACCGAACGCAGCGGTGGTAAACGGCGCAGTGGCGGTTTTTCATGGCTTGTATGACTGTTTTTTTTGGGGTACAGT  
CTATGCCCTCGGGCATCCAAGCAGCAAGCGGTTACGCCGTGGGTGATGTTTGTATGTTATGGAGCAGCAACGATGTTACGCAGCAGGGCAGTCCGC  
CTAAAACAAAGTTAAACATCATGAGGGAAGCGGTGATCGCCGAAGTATCGACTCAACTATCAGAGGTAGTTGGCGTCATCGAGCGCCATCTCGAAC  
CGACGTTGCTGGCCGTACATTTGTACGGCTCCGCAGTGGATGGCGGCCTGAAGCCACACAGCGATATTGATTTGCTGGTTACGGTGACCGTAAGGC  
TTGATGAAAACACGCGCGAGCTTTGATCAACGACCTTTTGGAAACTTCGGCTTCCCTGGAGAGAGCGAGATTCTCCGCGCTGTAGAAGTCACCA  
TTGTTGTGCACGACGACATCATTCCGTGGCGTTATCCAGCTAAGCGCGAAGTGAATTTGGAGAATGGCAGCGCAATGACATTTCTGCAGGTATCT  
TCGAGCCAGCCAGATCGACATTTGATCTGGCTATCTTGCTGACAAAAGCAAGAGAACATAGCGTTGCCTTGGTAGGTCCAGCGCGGAGGAACCTCT  
TTGATCCGGTTCCTGAACAGGATCTATTTGAGGCGCTAAATGAAACCTTAACGCTATGGAACCTCGCCGCCGACTGGGCTGGCGATGAGCGAAATG  
TAGTGCTTACGTTGTCCCGCATTTGGTACAGCGCAGTAACCGGCAAAATCGCCCGAAGGATGTCGCTGCCGACTGGGCAATGGAGCGCCTGCCGG  
CCCAGTATCAGCCCGTCATACTTGAAGCTAGACAGGCTTATCTTGGACAAGAAGAAGATCGCTTGGCCTCGCGCGCAGATCAGTTGGAAGAATTTG  
TCCATTACGTA AAAAGGCGAGATCACCAAGGTAGTCGGCAAATAACTGTACAGCAAGTTTACTCATATATACTTTAGATTGATTTAAAACCTTCATT  
TTTAATTTAAAAGGATCTAGGTGAAGATCCTTTTTGATAATCTCATGACCAAATCCCTTAACGTGAGTTTTCGTTCCACTGAGCGTCAGACCCCG  
TAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTTTTCTGCGCGTAATCTGCTGCTTGCAAACAAAAAACACCAGCTACCAGCGGTGGTTTGT  
TGCCGGATCAAGAGCTACCAACTCTTTTTCCGAAGGTAACCTGGCTTCAGCAGAGCGCAGATACCAAATACTGTCTTCTAGTGTAGCCGTAGTTAG  
GCCACCACTTCAAGAACTCTGTAGCACCAGCTACATACCTCGCTCTGCTAATCTGTTACCAAGTGGCTGCTGCCAGTGGCGATAAGTCGTGTCTTA  
CCGGGTTGGACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTCCGGCTGAACGGGGGGTTCGTGCACACAGCCAGCTTGGAGCGAACGACCT  
ACACCGAACTGAGATACCTACAGCGTGTAGCTATGAGAAAAGCGCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTATCCGGTAAGCGGCAGGGTCCG  
GAACAGGAGAGCGCACGAGGGAGCTTCCAGGGGAAACGCCCTGGTATCTTTATAGTCTGTGCGGTTTTCCGCACCTCTGACTTGTAGCGTCGATTTT  
TGTGATGCTCGTCAGGGGGGGGAGCCTATGAAAAACGCCAGCAACCGCGCTTTTTACGGTTCCTGGCCTTTTGTGCTGCTTTTGTGCTCATATGT  
TCTTTCTGCGTTATCCCTGATTCTGTGGATAACCGTATTACCGCTTTGAGTGTAGCTGATACCGCTCGCCGAGCCGAACGACCGAGCGCAGCG  
AGTCAGTGTAGCGAGGAAGCGGAAGAGCGCCCAATACGCAAACCGCCTCTCCCGCGCGTTGGCCGATTCAATTAATC

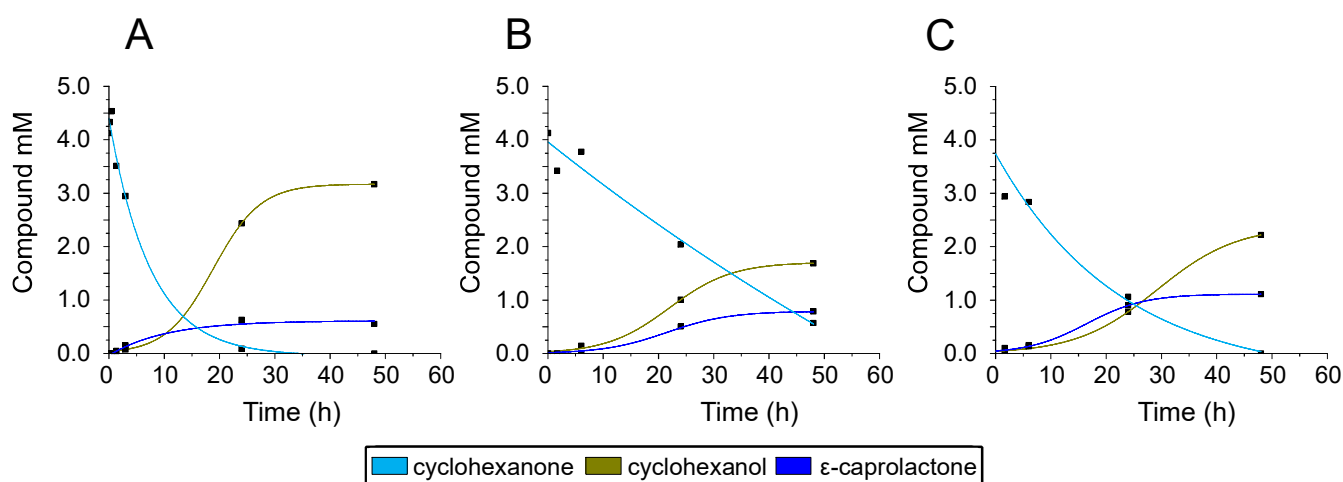
**Figure S1. Sequence of pMBS596 containing the CHMO coding sequence in B3–B4 for level 0.** The coding sequence of *Acinetobacter* NCIB 9871 CHMO (purple) was codon adapted and *Chlamydomonas* RBCS introns one (blue), two (green) and three (orange) were incorporated to ensure high-level transgene expression. Sequences of recipient vector pAGM1287 are in grey.



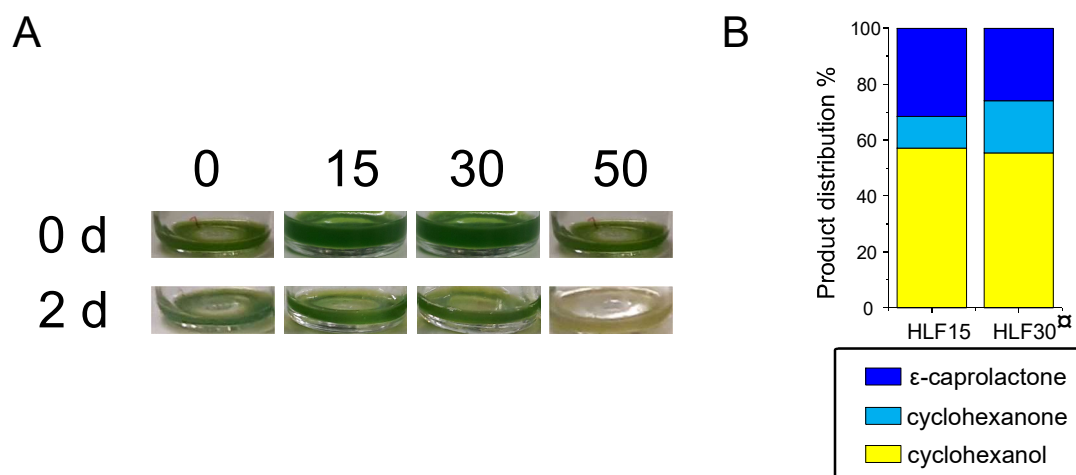
**Figure S2: Generation of *Chlamydomonas* lines expression CHMO-3xHA in the cytosol and the chloroplast stroma.** A) Level 2 device consisting of the *aadA* cassette ( $S^R$ ) and the transcription unit for the expression of CHMO-3xHA in the cytosol (CHMO). The coding sequence of CHMO (B3–B4) was fused to the PAR (HSP70A-RBCS2 promoter, A1–B2), a 3xHA-tag (B5) and TRPL23 (RPL23 terminator, B6–C1). B) Screening for transformants expressing CHMO-3xHA in the cytosol. The level 2 construct shown in A) was transformed into *Chlamydomonas* UVM4. Whole-cell protein corresponding to 2  $\mu$ g chlorophyll of 24 spectinomycin-resistant transformants was separated by SDS-PAGE and immunodetected with an antibody against HA. C) Screening for transformants overexpressing CHMO-3xHA in the cytosol. The level 2 construct shown in A) was transformed into *Chlamydomonas* UVM11-CW. Whole-cell protein corresponding to 2  $\mu$ g chlorophyll of 24 spectinomycin-transformants was separated by SDS-PAGE and immunodetected with an antibody against HA. Additionally detected bands at ~35 kDa and ~20 kDa are cross reactions of the HA antibody with proteins in this strain background. D) Level 2 device consisting of the *aadA* cassette ( $S^R$ ) and the transcription unit for the expression of CHMO-3xHA in the stroma (CHMO\_PSAD). The coding sequence of CHMO (B3–B4) was fused to PAR (HSP70A-RBCS2 promoter, A1–B1), the chloroplast transit peptide of PSAD (B2), a 3xHA-tag (B5) and TRPL23 (RPL23 terminator, B6–C1). E) Screening for transformants overexpressing CHMO-3xHA in the chloroplast stroma. The level 2 construct shown in A) was transformed into *Chlamydomonas* UVM11-CW. Whole-cell protein corresponding to 2  $\mu$ g chlorophyll of 24 spectinomycin-transformants was separated by SDS-PAGE and immunodetected with an antibody against HA. The additionally detected bands at ~35 kDa and ~20 kDa are known cross reactions of the antibody in this strain background.



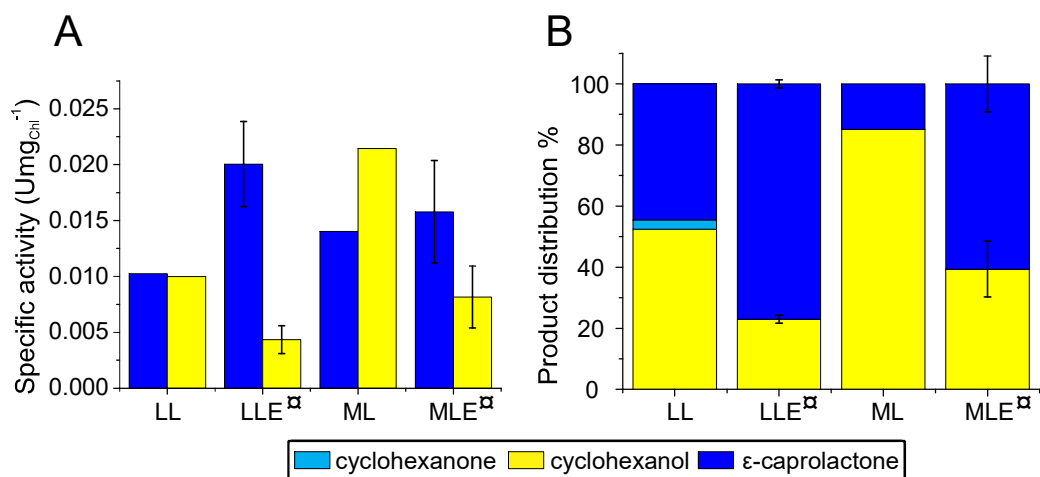
**Figure S3. Evaluating toxicity of substrate and product towards the *Chlamydomonas* UVM4 strain.** A) Following growth by OD<sub>750 nm</sub> with no additives (black), 5 mM cyclohexanone (red), 10 mM cyclohexanone (blue), 15 mM cyclohexanone (pink), and 20 mM cyclohexanone (green). B) Following growth by OD<sub>750 nm</sub> without additives (black), with 5 mM ε-caprolactone (purple) and with 20 mM ε-caprolactone (light green). C) Following growth by total chlorophyll concentration, same samples as in A. D) Following growth by total chlorophyll concentration, same samples as in B. E) Oxygen evolution on the 2nd day of cultivation (0mM, 5mM, 15mM, and 20mM cyclohexanone; 5 mM and 20 mM ε-caprolactone) and on the 3rd day of cultivation (0 mM and 10 mM cyclohexanone). The presented standard deviations in the curves are from two biological replicates for the control and from three technical replicates for all others.



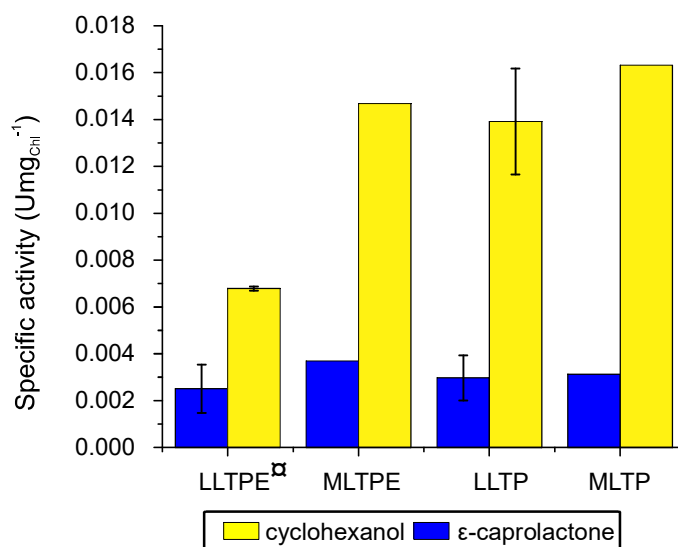
**Figure S4. Effects of additives to the reaction with UVM4/CHMO clone 3.** Illumination was set to  $165 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ . A) No additives. B) 30 mM fomepizole. C) 1.7 % (v/v) Ethanol.



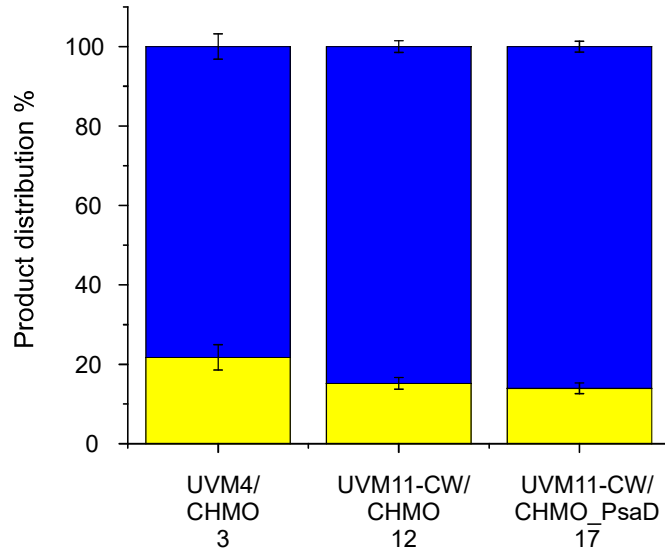
**Figure S5. Effect of fomepizole on UVM4/CHMO clone 3.** A) Effect of fomepizole on *Chlamydomonas* cells after 2 days of biotransformation in ML with 0, 15, 30, and 50 mM of fomepizole. B) Comparison of the effect of 15 mM (HLF15) and 30 mM (HLF30) fomepizole on product distribution after 48 h in HL.  $\alpha$  Same data as in Fig. 2B HLF to facilitate comparison.



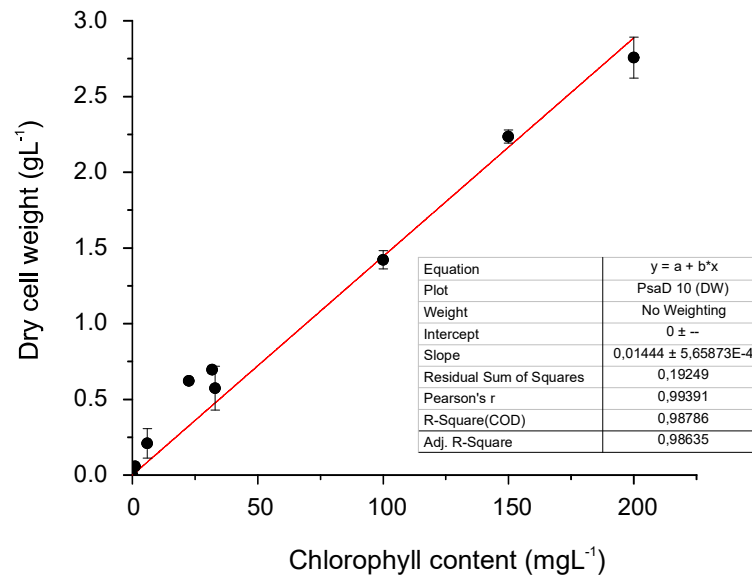
**Figure S6. Comparing the effect of ethanol addition under different light conditions using UVM4/CHMO clone 3.** A) Specific activity (Umg<sub>Chl</sub><sup>-1</sup>) under the following conditions: LL) LL without additives. LLE) LL with ethanol [1.7%, (v/v)]. ML) ML without additives. MLE) ML with ethanol [1.7%, (v/v)]. B) Product distribution after 48 h of cultivation under the conditions described in A). Presented standard deviations are from three biological replicates. <sup>α</sup> Data of the samples with ethanol are the same as in Fig. 2 to facilitate comparison.



**Figure S7. Comparing different conditions for phototrophic biotransformation using UVM4/CHMO clone 3.** Specific activity (Umg<sub>Chl</sub><sup>-1</sup>) under the following conditions: LLTPE) LL with ethanol [1.7%, (v/v)]. MLTPE) ML with ethanol [1.7%, (v/v)]. LLTP) LL without ethanol. MLTP) ML without ethanol. <sup>α</sup> Data for LLTPE are the same as in Fig. 2 to facilitate comparison. Presented standard deviation are from 2–3 technical replicates.



**Figure S8. The ratios of the compounds present in different strains after 48h of biotransformation. Standard deviations are from three biological replicates.**



**Figure S9. Chlorophyll and cell dry weight (DCW) correlation curve for UVM11-CW/CHMO\_PSA D clone 10. Standard deviations are from three biological replicates.**