

Electronic Supplementary Information

Enantiopure natural deep eutectic solvents for metal-organic framework chiral induction

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Experimental Section

Materials

The commercially available reagents choline chloride (99% purity, Acros), *L*-proline (99% purity, Sigma-Aldrich), *D*-proline (99% purity, ABCR), *DL*-proline (98% purity, BLDpharm), *L*-menthol (\geq 99% purity, Sigma-Aldrich), *D*-menthol (\geq 99% purity, TCI Chemicals), *DL*-menthol (99% purity, Acros), thymol (\geq 99% purity, Sigma-Aldrich), $\text{TbCl}_3 \cdot 6\text{H}_2\text{O}$ (99.9% purity, Strem Chemicals or Sigma-Aldrich), 1,3,5-benzenetricarboxylic acid (prepared as described¹ or purchased from BLD pharm, 95% purity) were used as received.

Synthesis

General preparation of the deep eutectic solvents: in a round bottom flask, equipped with a stirring bar, it was added, in the respective molar ratio (see Table 1), the DES components. The flask was placed in a pre-heated oil bath at 100 °C and the mixture was stirred until it became a clear liquid solution. Due to their viscosity, the DESs were kept at 100 °C to facilitate the addition of these solvents into the reaction vessel by syringe. The DESs were freshly prepared right before each reaction.

Synthesis of MIL-103: In a 48 mL Chemglass vessel, $\text{TbCl}_3 \cdot 6\text{H}_2\text{O}$ (1 equiv., 1.71 mmol, 0.64 g), 1,3,5-benzenetricarboxylic acid (1 equiv., 1.71 mmol, 0.75 g) and NaOH 2 M solution (1.8 equiv. 3.0 mmol, 1.5 mL) were added. With the aid of a syringe, 12.5 mL of the appropriate deep eutectic solvent was added into the reaction vessel. Right after, the vessel was sonicated at 40 °C for 1 h to homogenize the reaction mixture before the reaction. Then, the vessel was placed in a pre-heated oil bath at 120 °C for 3 days. At the end of the reaction, the vessel was cooled down to room temperature. The workup of the reaction consists in washing the solid product in a Büchner funnel according to the following steps. First, distilled water (15 mL) was poured into the reaction vessel and the slurry was mixed by a spatula. After, it was filtered off and washed with more water (30 mL). Then, the brownish solid was washed with DMF until the filtrate became clear and the solid became white/off-white. The DMF was then washed off from the solid with water (30 mL), followed by ethanol (30 mL). The remaining solid product was immersed in ethanol (100 mL) for a week, and every two days the ethanol was replaced for a fresh one. Finally, the solid was again filtered off and dried in an oven at 80 °C overnight. The remaining product was analyzed by PXRD, TGA, elemental analysis, infra-red spectroscopy, and N_2 adsorption-desorption isotherms at 77 K.

Table ESI1

| MOF(Tb) | DES | Mass (g) | T _d (°C) | S _{BET} (m ² /g) | Elemental analysis (%) | CD (mdeg) |
|-----------------------------------|---------------------------------|--------------------|---------------------|--------------------------------------|--|-----------|
| <i>D</i>-pro-MIL-103 | (1:7) <i>D</i> - proline:thymol | 1.478 ^a | 588 | 630 | Found: N 0.19, C 53.25, H 2.64 Calcd: N 0.19, C 53.33, H 2.82 for Tb(BTB)(H ₂ O) _{0.75} (<i>D</i> -pro) _{0.08} | - 6.4 |
| <i>L</i>-pro-MIL-103 | (1:7) <i>L</i> -proline:thymol | 1.346 ^a | 588 | 592 | Found: N 0, C 52.05, H 2.58 Calcd: N 0, C 52.04, H 2.94 for Tb(BTB)(H ₂ O) _{1.60} | + 5.4 |
| <i>DL</i>-pro-MIL-103 | (1:7) <i>DL</i> -proline:thymol | 0.736 | 580 | 787 | Found: N 0.30, C 51.35, H 2.66 Calcd: N 0.36, C 51.47, H 3.18 for Tb(BTB)(H ₂ O) _{2.0} (<i>DL</i> -pro) _{0.17} | 0 |
| <i>D</i>-men-MIL-103 | (1:1) <i>D</i> -menthol/thymol | 0.645 | 581 | 633 | b | + 2.8 |
| <i>L</i>-men-MIL-103 | (1:1) <i>L</i> -menthol/thymol | 0.662 | 581 | 758 | b | - 2.9 |
| <i>DL</i>-men-MIL-103 | (1:1) <i>DL</i> -menthol/thymol | 0.589 | 580 | 589 | b | 0 |
| water:cyclohexanol MIL-103 | water:cyclohexanol (1:1) | 0.656 | 578 | 1087 | Found: N 0, C 53.36, H 2.45 Calcd: N 0, C 53.35, H 2.74 for Tb(BTB)(H ₂ O) _{0.75} | 0 |

^a The scale for this reaction has been doubled compared with the general protocol.

^b No stable nor repeatable CHN analysis.

X-ray diffraction

PXRD patterns were recorded on a Bruker D8 Advance diffractometer with LynxEye detector, Cu K α wavelength, in a 2θ interval of $4^\circ - 40^\circ$, step size of 0.007° and time/step of 0.35 s.

Thermogravimetric analysis

The thermal stability of the samples was accessed in a PerkinElmer Thermogravimetric Analyzer TGA 4000 under N $_2$ flow of 20 mL/min, and a heating rate of $5^\circ\text{C}/\text{min}$, up to 800°C .

Gas sorption

The textural properties of the samples were obtained by N $_2$ adsorption-desorption isotherms at 77 K, in a Micromeritics ASAP 2020 Surface Area and Porosity Analyzer. The Brunauer-Emmett-Teller (BET) method was used for obtaining the total specific surface area (S_{BET}) and the pore size (model: NLDFT for cylinder pores). The samples were activated, prior to the N $_2$ adsorption-desorption analysis, by heating the sample under vacuum at 150°C for 15 h. The MicroActive Software version 4.00 was used to analyze the data.

Elemental analysis

Elemental analyses (CHN) were performed at the *Service Commun d'Analyses* of the University of Strasbourg, in duplicate, employing a ThermoFischer Flash 2000 equipment, where the reported values for the CHN were taken as the average of two measurements.

Fourier-transform infrared spectroscopy

The infrared spectra were obtained in a FTIR-UATR PerkinElmer Spectrum Two, in the wavenumber interval of $4.000\text{ cm}^{-1} - 400\text{ cm}^{-1}$.

Scanning electron microscopy

SEM images were obtained with a Zeiss GeminiSEM 500 microscope with a FEG Schottky source and an Everhart-Thornley detector.

Circular dichroism

Diffuse-reflectance circular dichroism (DRCD) spectra (room temperature) of crystalline samples were performed on a JASCO J-1500 spectrophotometer equipped with an integrating sphere attachment (DRCD-575 accessory) in the 200-800 nm domain at a scan rate of 100 nm/min and accumulated twenty times.

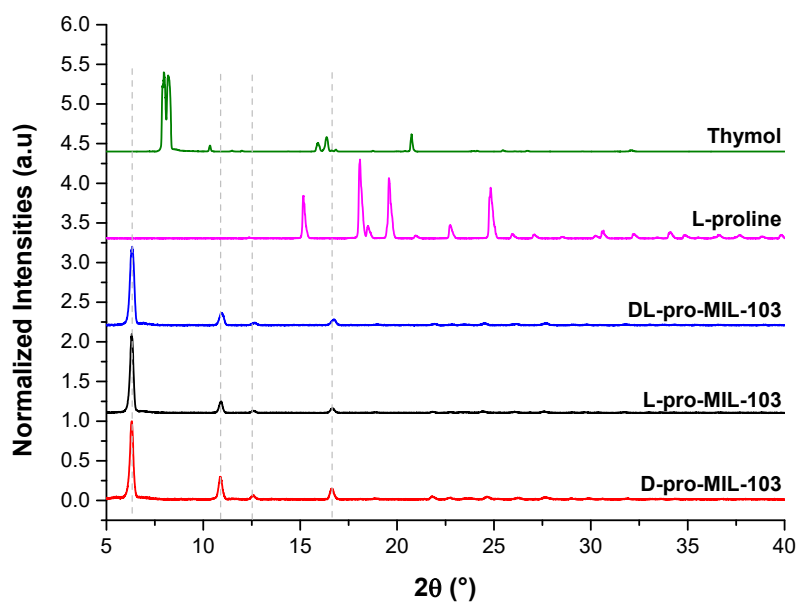


Fig. ESI1 – PXRD patterns of the DES components thymol and L-proline, and of the MIL-103(Tb) MOFs prepared in *D*-proline:thymol, *L*-proline:thymol, *DL*-proline:thymol.

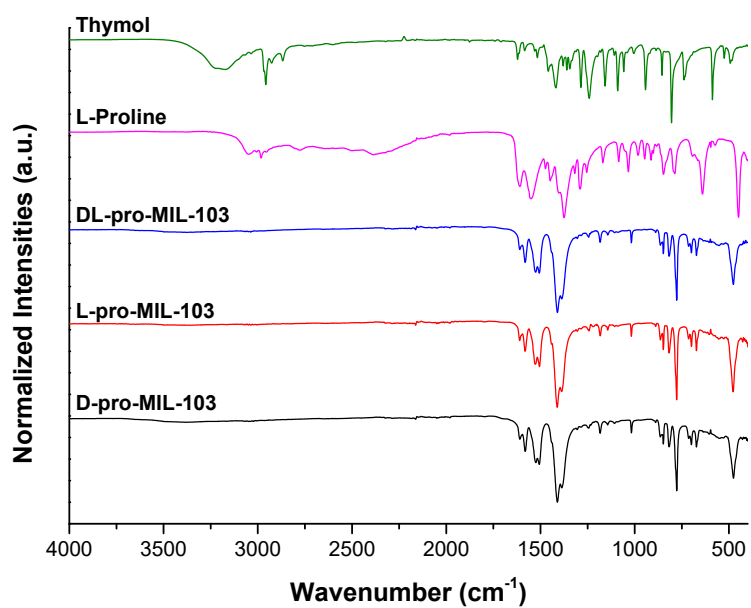


Fig. ESI2 – FT-IR spectra of the DES components thymol (green) and L-proline (pink), and of the MIL-103(Tb) prepared in *D*-proline:thymol (black), *L*-proline:thymol (red), *DL*-proline:thymol (blue).

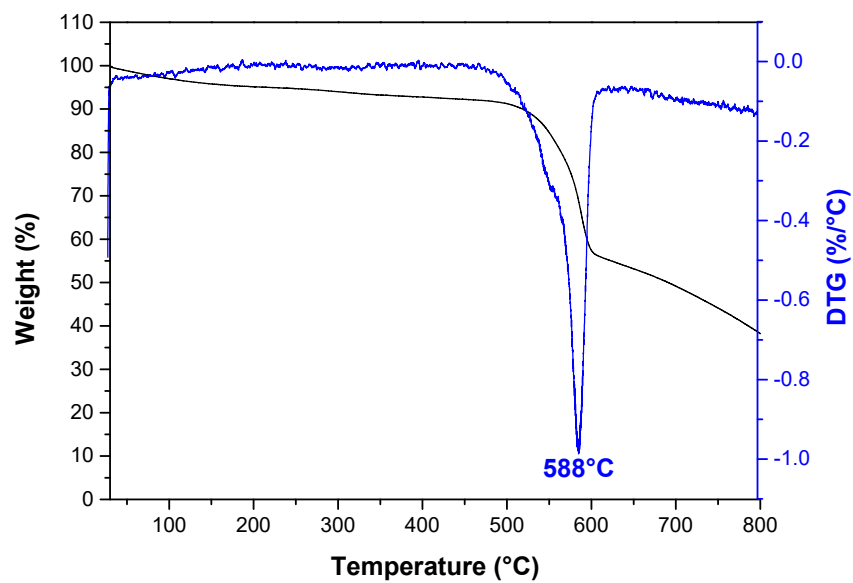


Fig. ESI3 – TGA plot of *D*-pro-MIL-103(Tb).

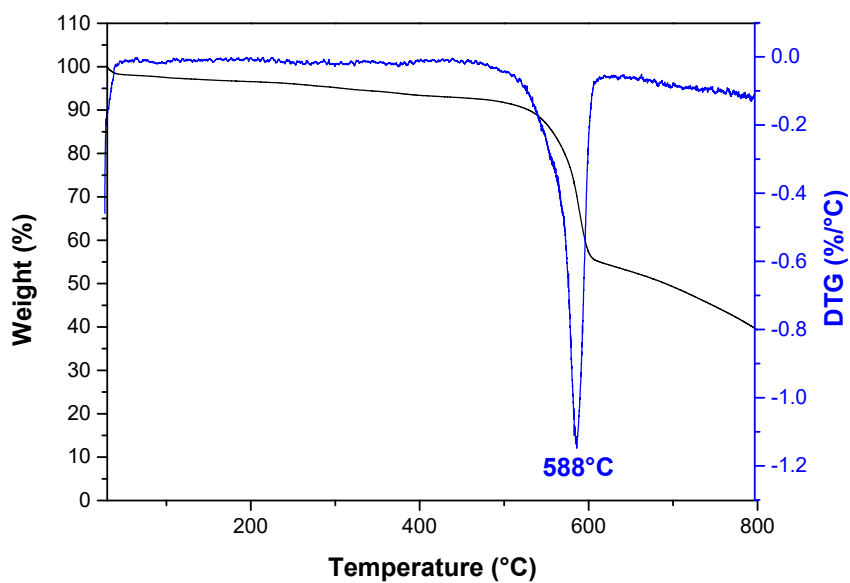


Fig. ESI4 – TGA plot of *L*-pro-MIL-103(Tb).

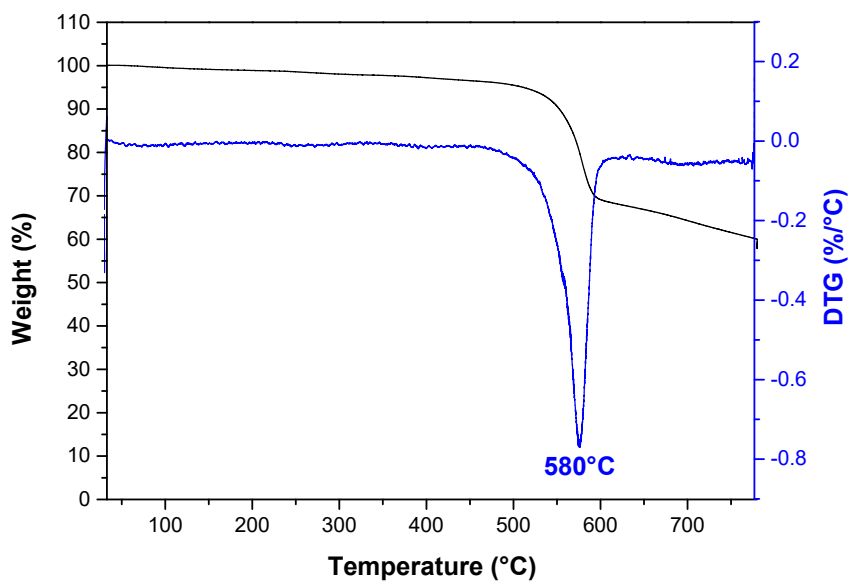


Fig. ESI5 – TGA plot of *DL*-pro-MIL-103(Tb).

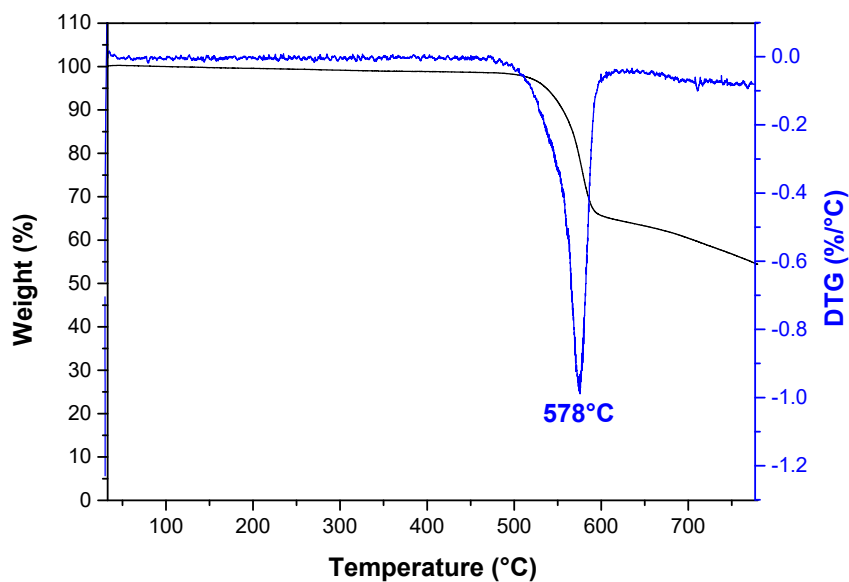


Fig. ESI6 – TGA plot of water:cyclohexanol MIL-103(Tb).

Table ESI2. Data presented in Fig. 4

| MOF | CD (mdeg) |
|---|------------------|
| <i>L</i> -pro-MIL-103(Tb) before activation | +5.0 (at 317 nm) |
| <i>D</i> -pro-MIL-103(Tb) before activation | -7.7 (at 316 nm) |
| <i>L</i> -pro-MIL-103(Tb) after activation | +5.5 (at 318 nm) |
| <i>D</i> -pro-MIL-103(Tb) after activation | -6.4 (at 318 nm) |

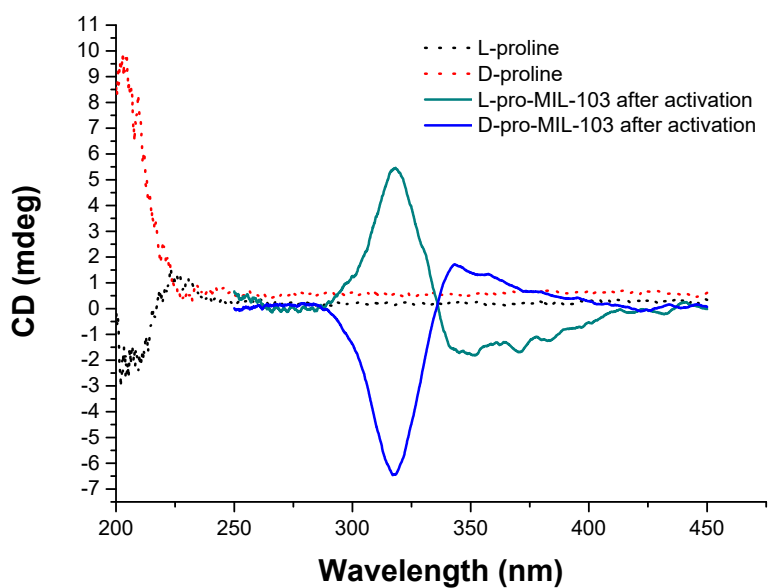


Fig. ESI7 – Circular dichroism spectra of the chiral MIL-103(Tb) MOFs prepared in different solvents: *D*-proline:thymol, and *L*-proline:thymol (after activation) and comparison with the DES components *D*- and *L*-proline.

Table ESI3. Extension of proline-based MOFs to other lanthanides (no sample was activated – the MOF was just filtered off and washed with DMF, water, and ethanol). Data presented in Fig. 5.

| | CD (mdeg) |
|----------------------------|--|
| <i>L</i> -pro-MIL-103 (Tb) | +6.8 (at 323 nm) |
| <i>D</i> -pro-MIL-103 (Tb) | -5.9 (at 323 nm) |
| <i>L</i> -pro-MIL-103 (Eu) | +13.7 (at 317 nm) |
| <i>D</i> -pro-MIL-103 (Eu) | -17.3 (at 318 nm) |
| <i>L</i> -pro-MIL-103 (Nd) | +40.2 (at 317 nm), +6.0 (at 585 nm), +4.2 (741 nm) |
| <i>D</i> -pro-MIL-103 (Nd) | -44.5 (at 318 nm), -6.0 (at 580 nm), -4.1 (740 nm) |

* For MIL-103(Tb), the batches are different from the MIL-103(Tb) ones listed in table ESI2.

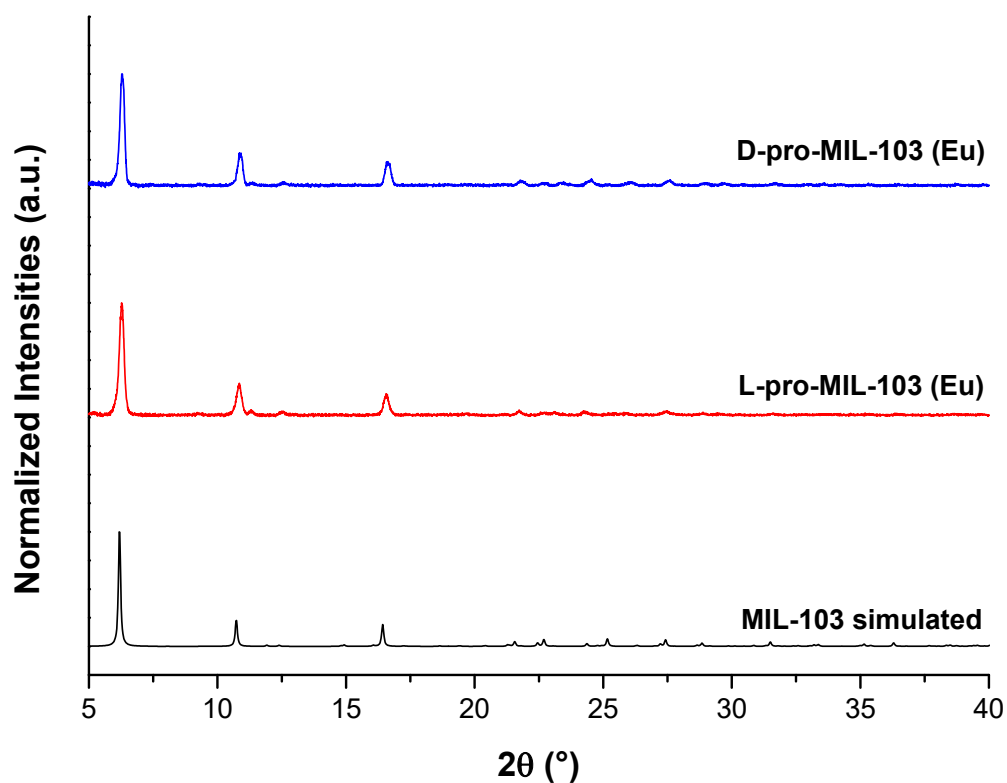


Fig. ESI8 – PXRD patterns of the MIL-103(Eu) MOFs prepared in *D*-proline:thymol and *L*-proline:thymol.

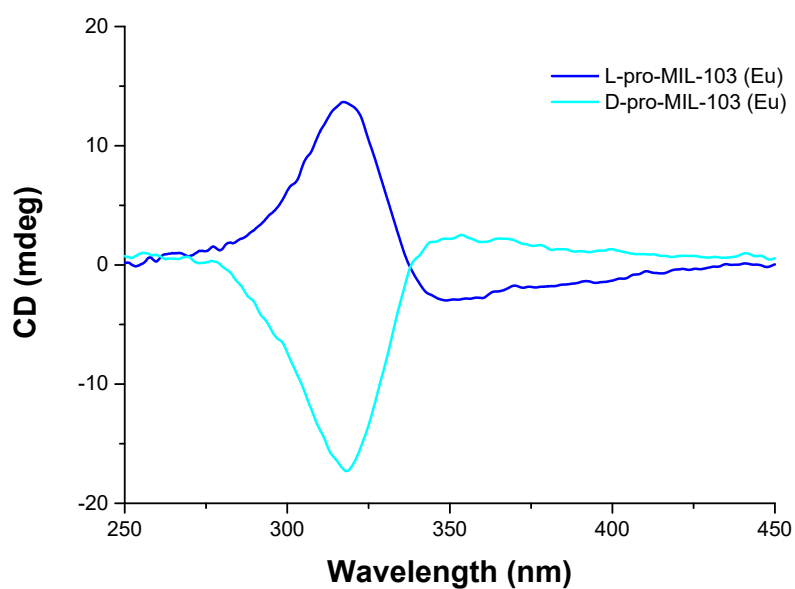


Fig. ESI9 – Circular dichroism spectra of the chiral MIL-103(Eu) MOFs prepared in different solvents: *D*-proline:thymol, and *L*-proline:thymol.

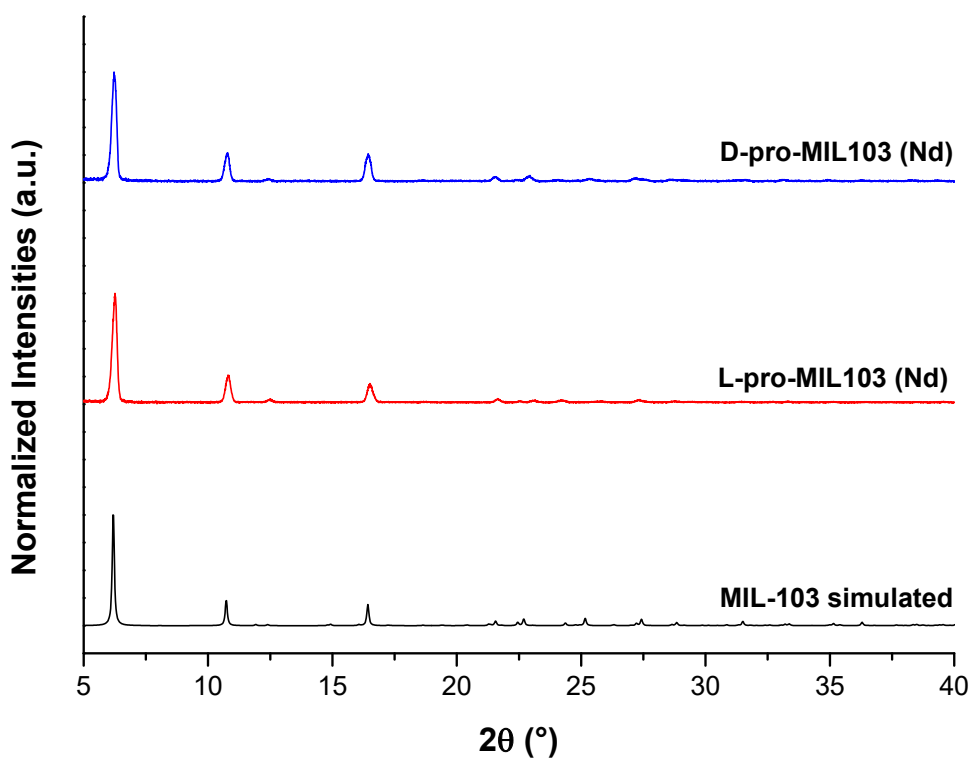


Fig. ESI10 – PXRD patterns of the MIL-103(Nd) MOFs prepared in *D*-proline:thymol and *L*-proline:thymol.

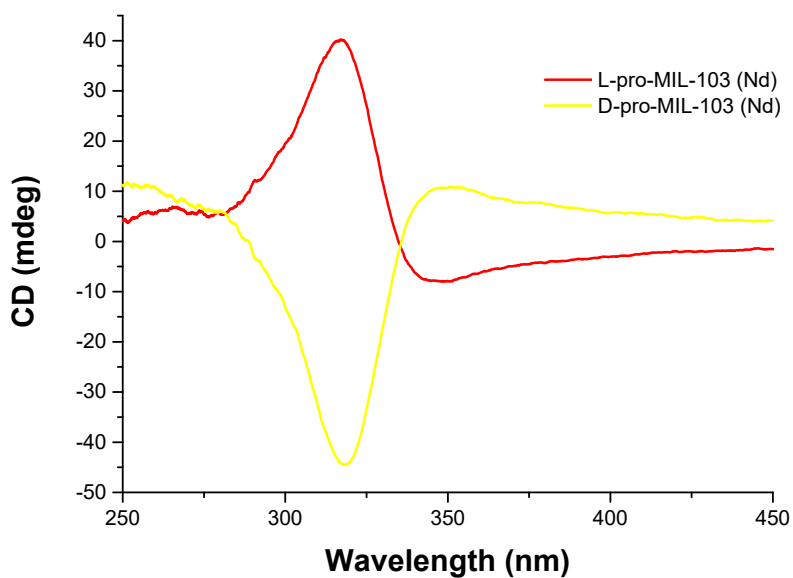


Fig. ESI11 – Circular dichroism spectra of the chiral MIL-103(Nd) MOFs prepared in different solvents: *D*-proline:thymol, and *L*-proline:thymol.

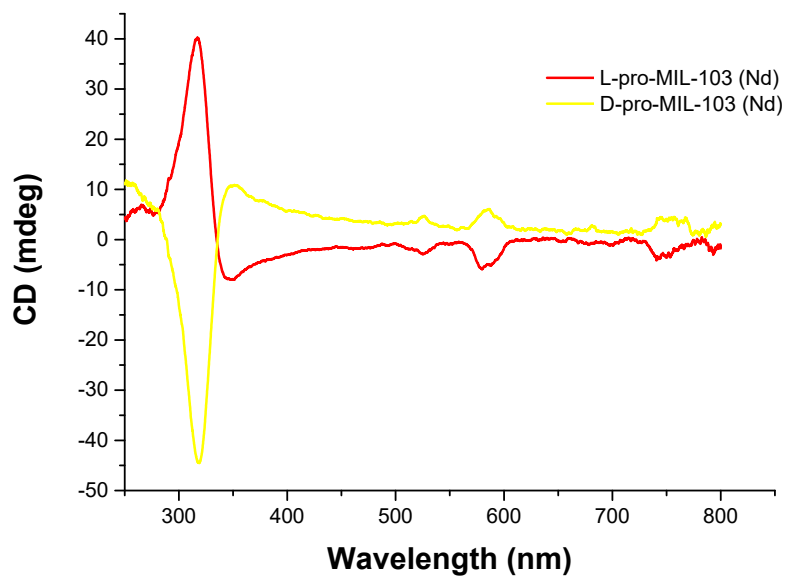


Fig. ESI12 – Circular dichroism spectra of the chiral MIL-103 MOFs (Nd) prepared in different solvents: *D*-proline:thymol, and *L*-proline:thymol.

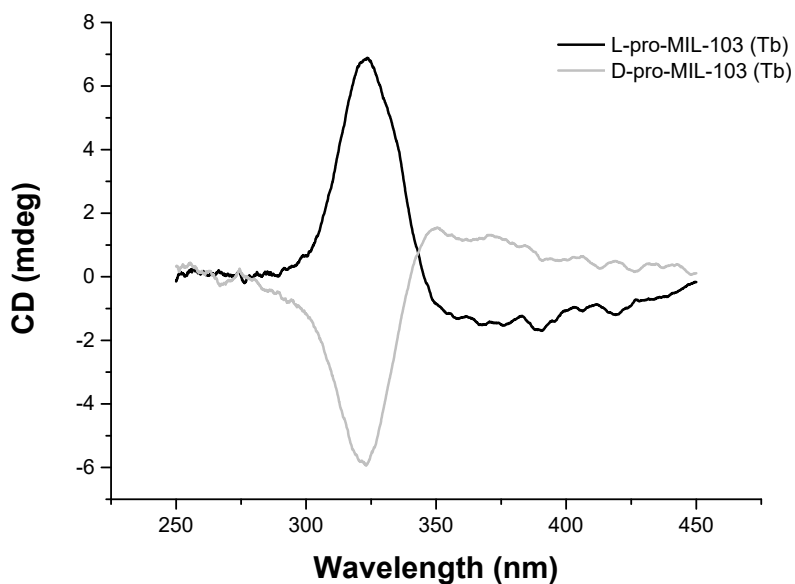


Fig. ESI13 – Circular dichroism spectra of the chiral MIL-103(Tb) MOFs prepared in different solvents: *D*-proline:thymol, and *L*-proline:thymol.

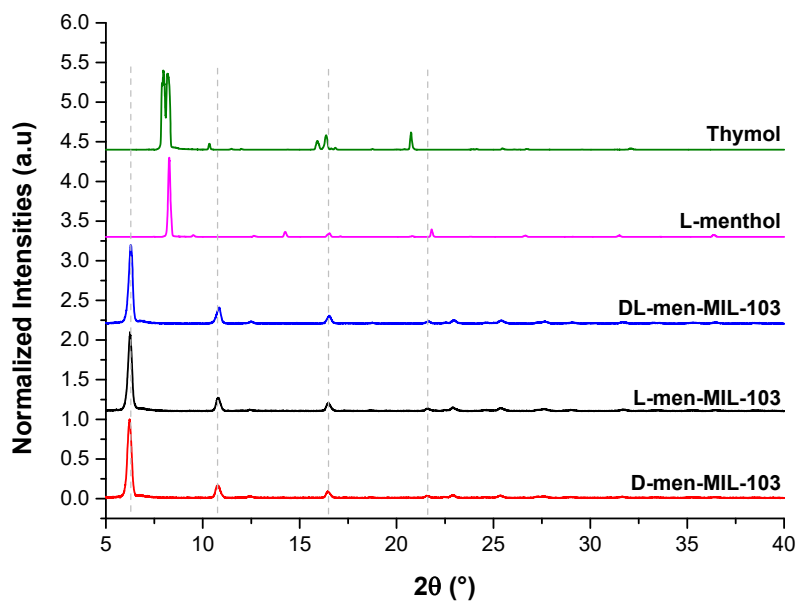


Fig. ESI14 - PXRD patterns of the DES components thymol and *L*-menthol, and of the MIL-103(Tb) MOFs prepared in *D*-menthol:thymol, *L*-menthol:thymol, *DL*-menthol:thymol.

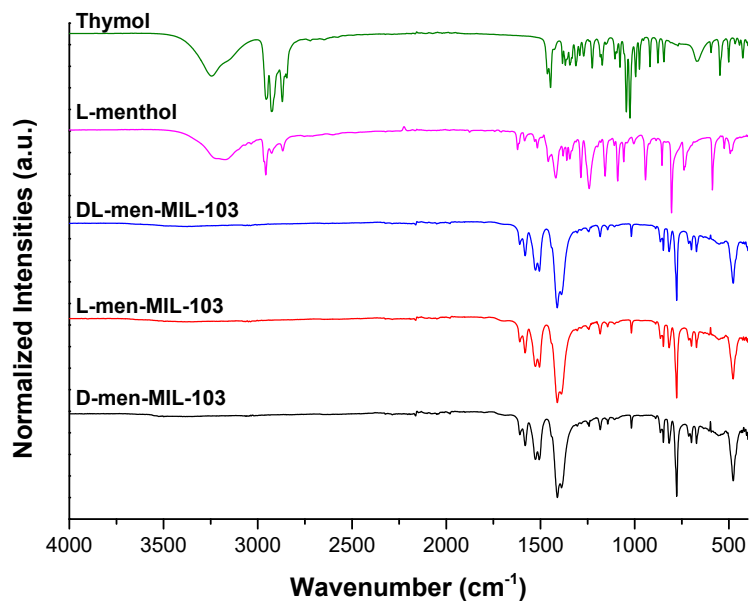


Fig. ESI15 – FT-IR spectra of the DES components thymol (green) and *L*-menthol (pink), and of the MIL-103(Tb) MOFs prepared in *D*-menthol:thymol (black), *L*-menthol:thymol (red), *DL*-menthol:thymol (blue).

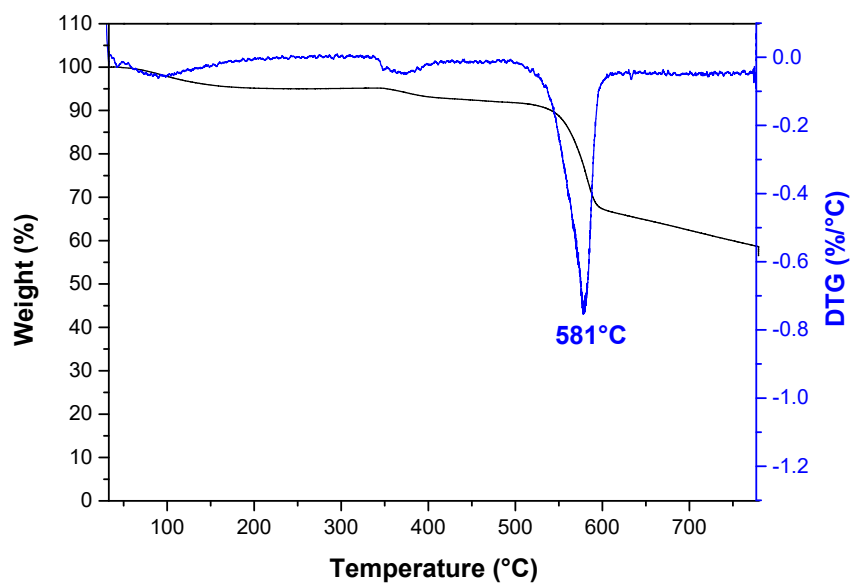


Fig. ESI16 – TGA plot of *D*-men-MIL-103(Tb).

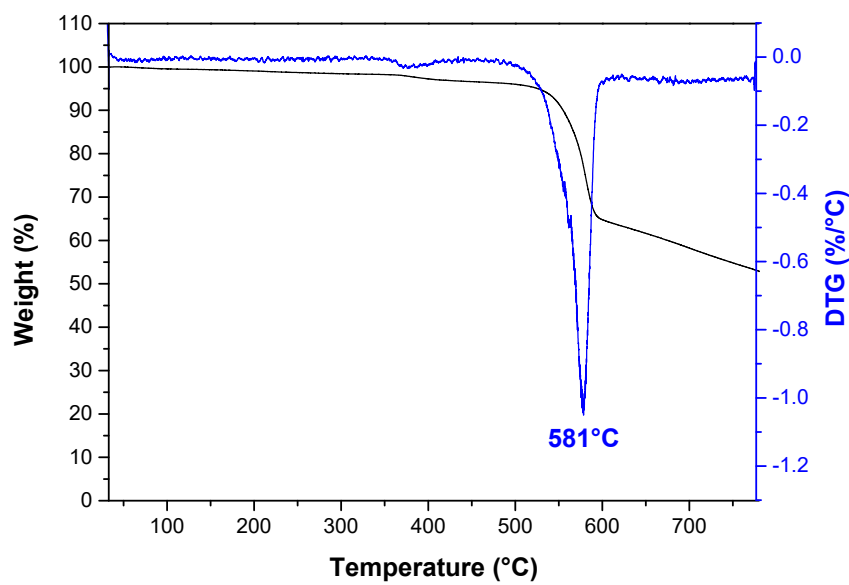


Fig. ESI17 – TGA plot of *L*-men-MIL-103(Tb).

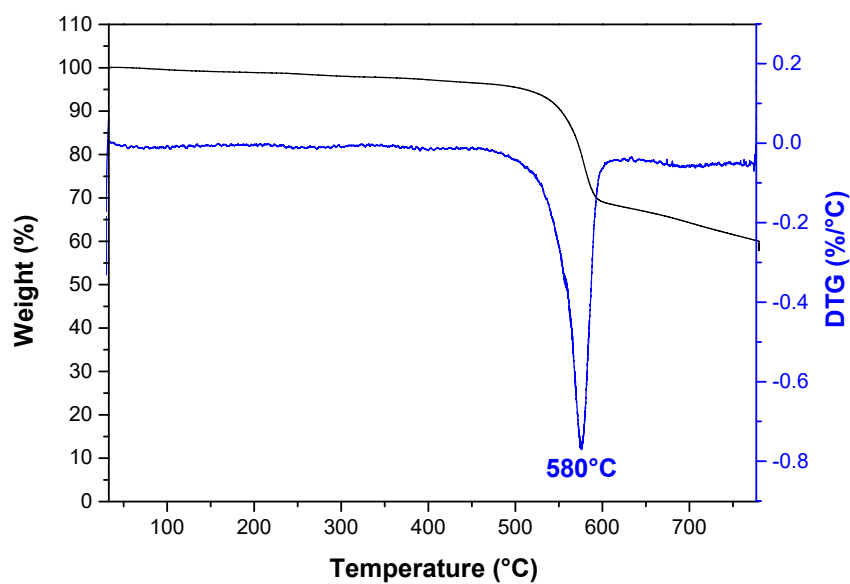


Fig. ESI18 – TGA plot of *DL*-men-MIL-103(Tb).

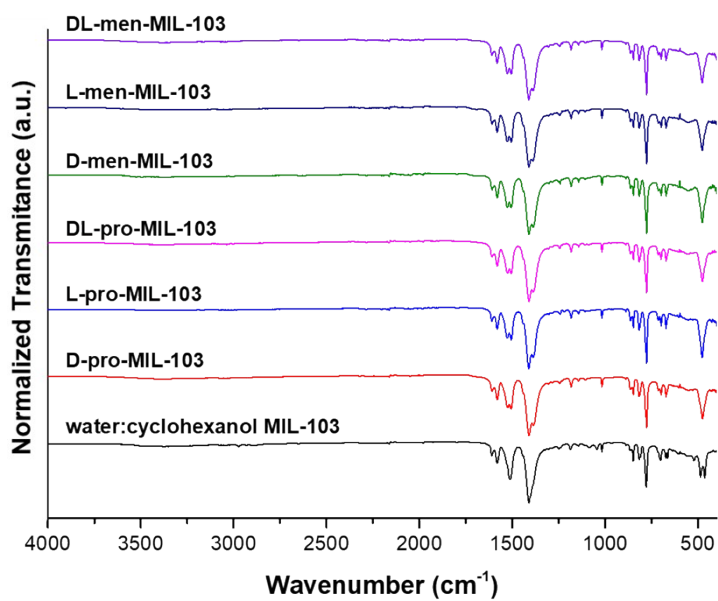


Fig. ESI19. FT-IR spectra of the MIL-103(Tb) MOFs prepared in different solvents: water:cyclohexanol, *D*-proline:thymol, *L*-proline:thymol, *DL*-proline:thymol, *D*-menthol:thymol, *L*-menthol:thymol, *DL*-menthol:thymol.

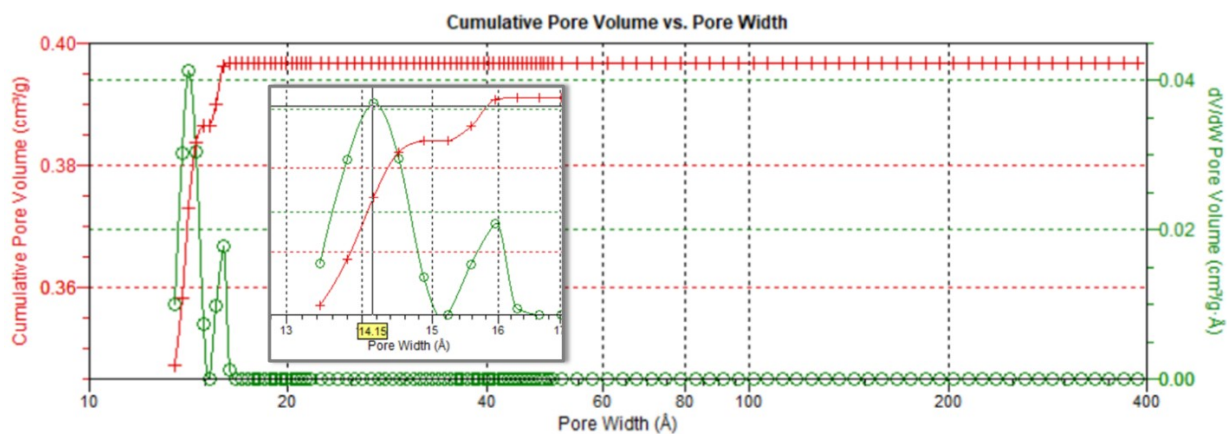


Fig. ESI20. Pore size distribution for the water:cyclohexanol MIL-103 MOF.

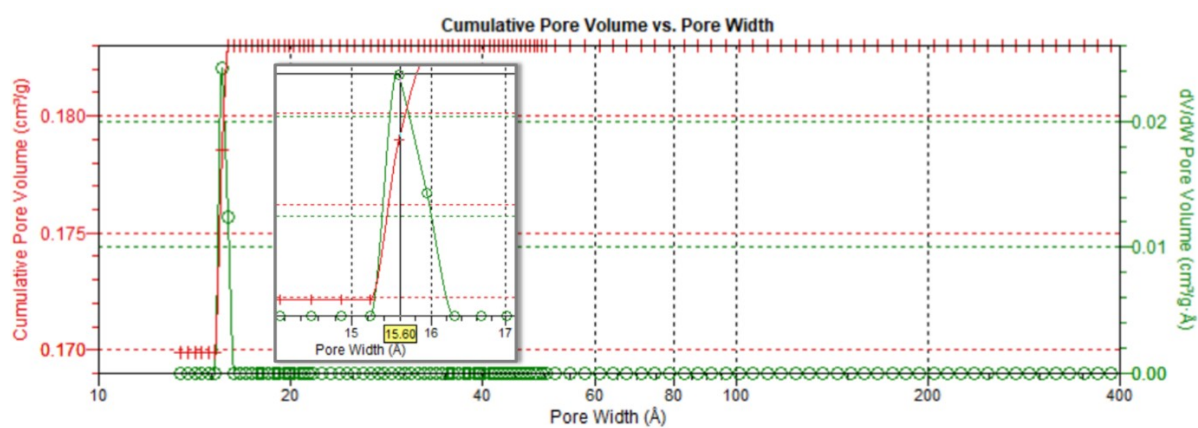


Fig. ESI21. Pore size distribution for the L-pro-MIL-103 MOF.

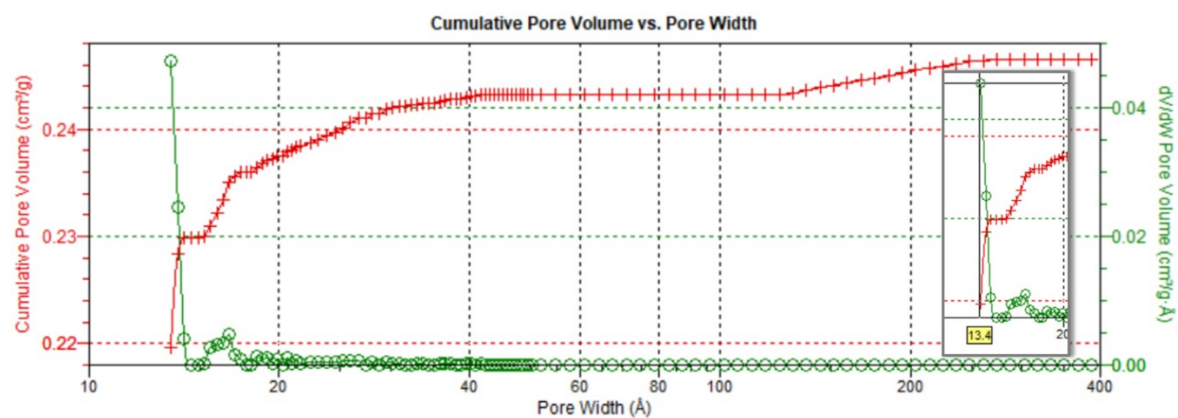


Fig. ESI22. Pore size distribution for the D-pro-MIL-103 MOF.

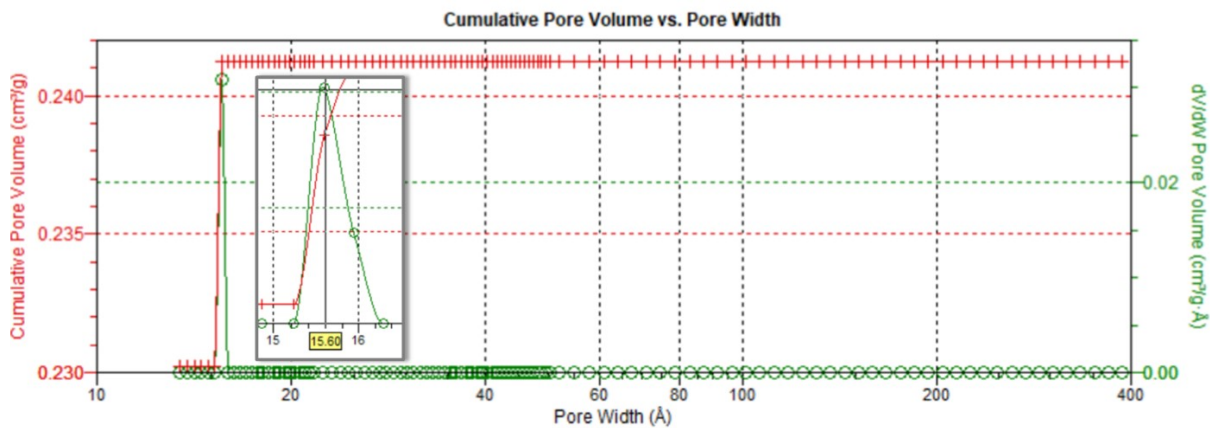


Fig. ESI23.. Pore size distribution for the *DL*-pro-MIL-103 MOF.

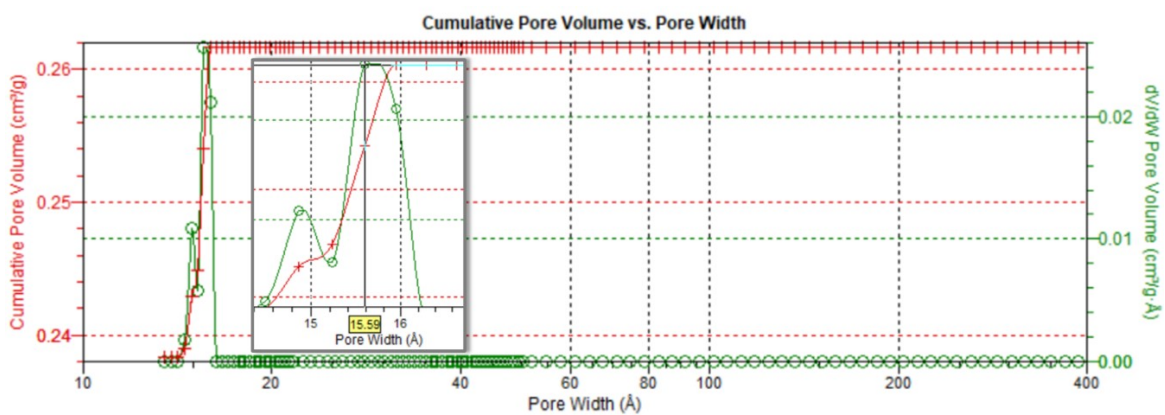


Fig. ESI24. Pore size distribution for the *L*-men-MIL-103 MOF.

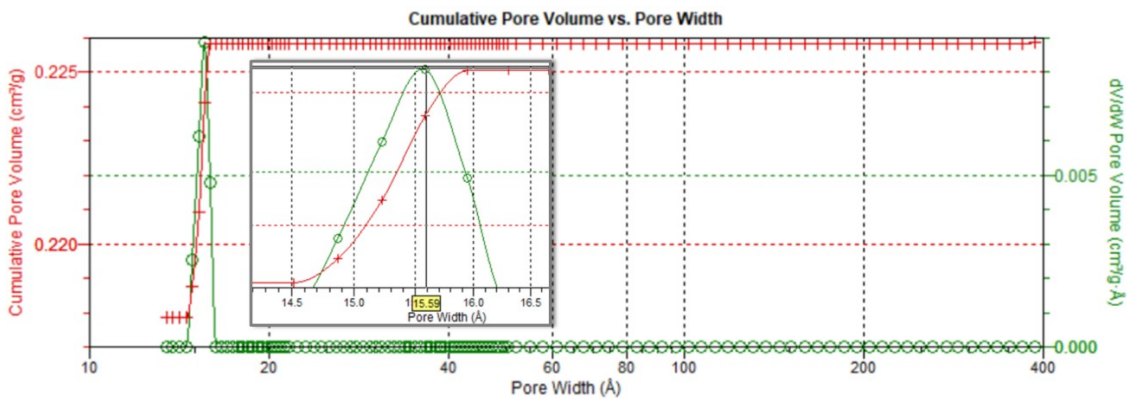


Fig. ESI25. Pore size distribution for the *D*-men-MIL-103 MOF.

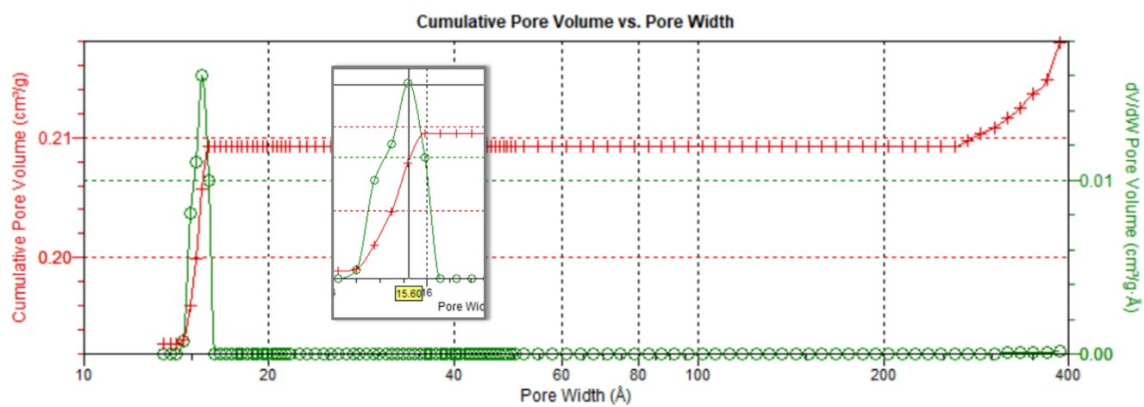


Fig. ESI26. Pore size distribution for the DL-men-MIL-103 MOF.

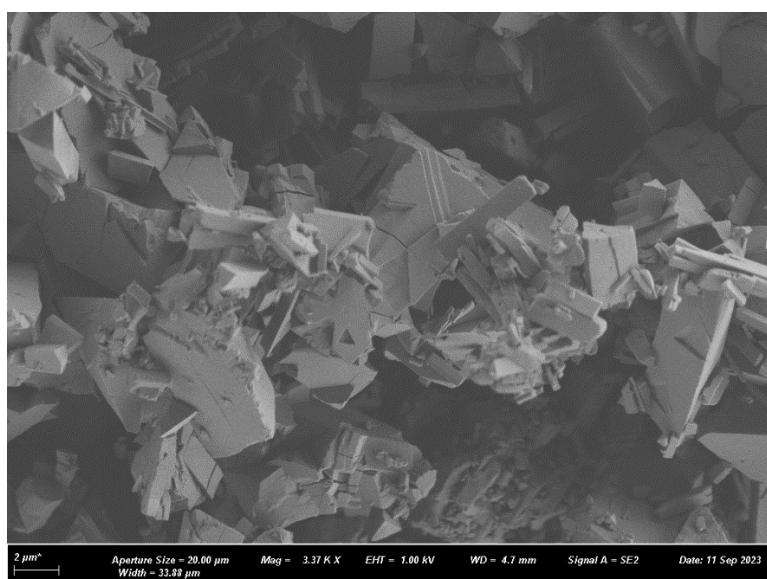


Fig. ESI27. SEM micrograph of the water:cyclohexanol MIL-103(Tb) MOF.

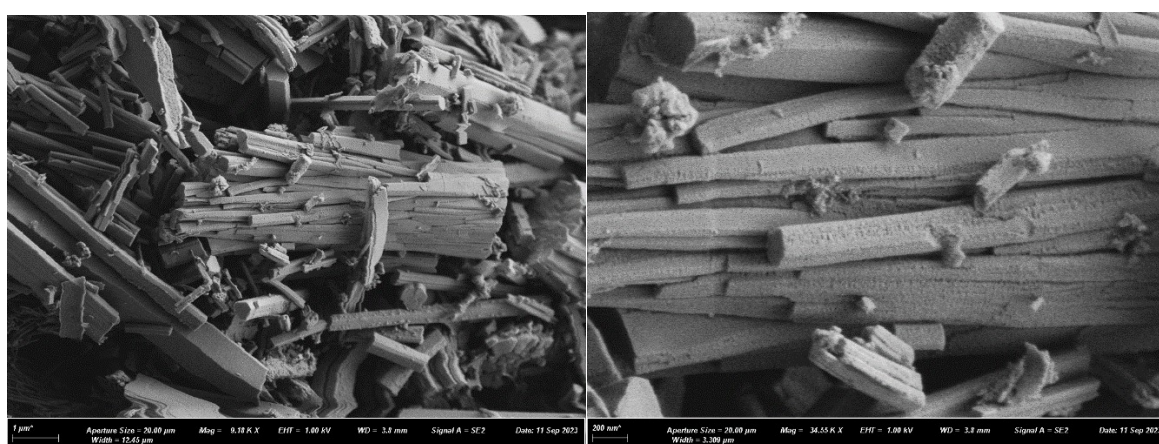


Fig. ESI28. SEM micrographs of the *L*-pro MIL-103(Tb) MOF.



Fig. ESI29. SEM micrograph of the *D*-pro MIL-103(Tb) MOF.



Fig. ESI30. SEM micrograph of the *DL*-pro MIL-103(Tb) MOF.

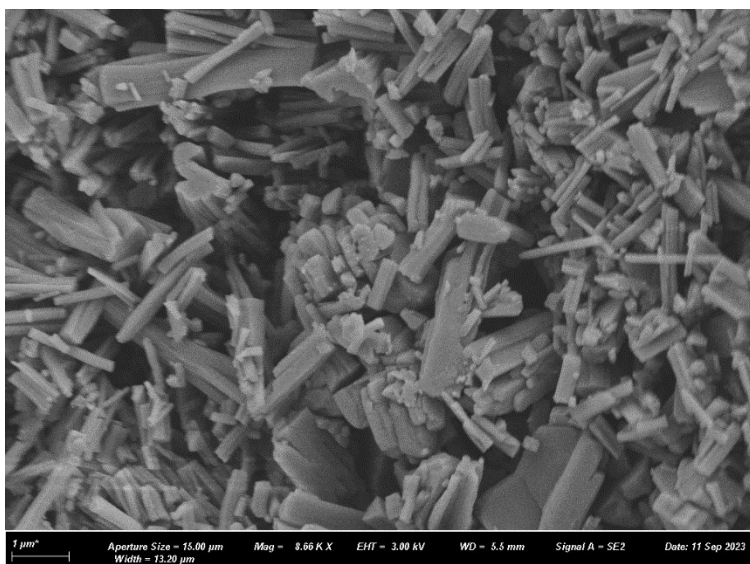


Fig. ESI31. SEM micrograph of the *L*-men MIL-103(Tb) MOF.

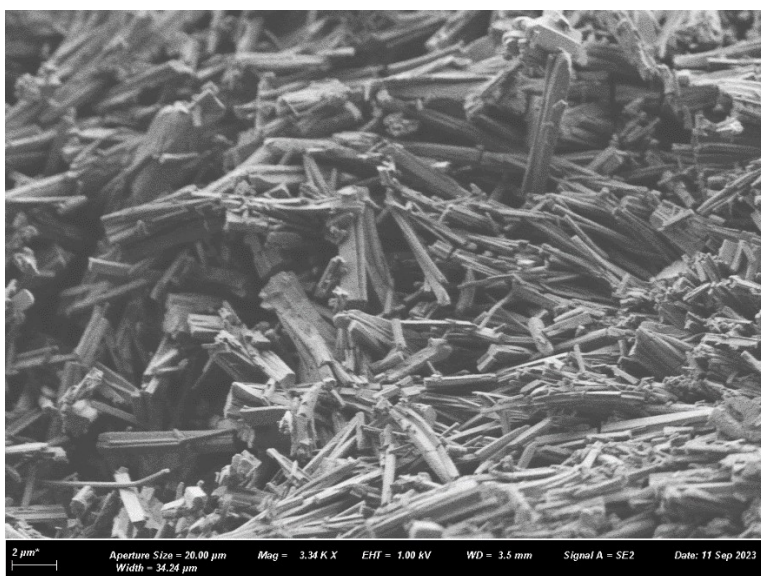


Fig. ESI32. SEM micrograph of the *D*-men MIL-103(Tb) MOF.

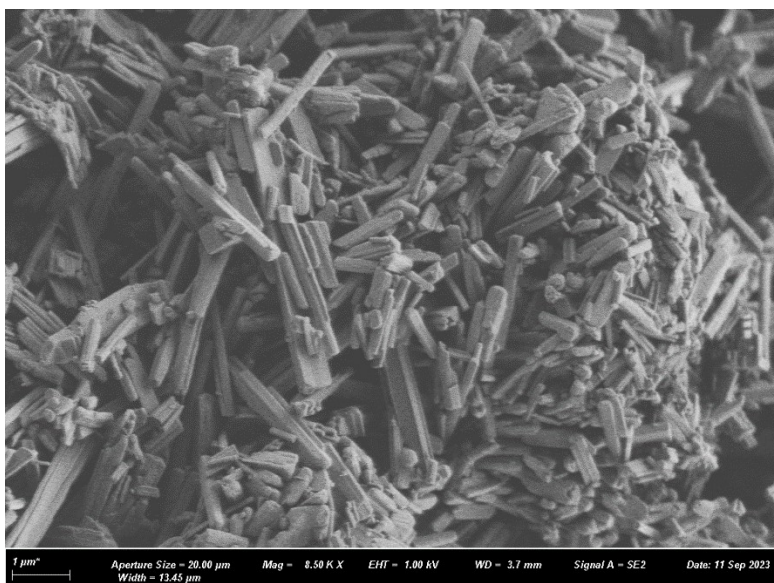


Fig. ESI33. SEM micrograph of the *DL*-men MIL-103(Tb) MOF.

Reference

1. D. J. Tranchemontagne, L. Dudek and O. M. Yaghi, *Inorg. Synth.*, 2010, **35**, 102