

Supporting Information

Cobalt-nanoparticles catalysed *N*-alkylation of amides with alcohols

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1. Materials and methods

All the alcohols and amides were purchased from various chemical companies. Cobalt(II) nitrate hexahydrate (cat no.139267-100G) was provided by Sigma-Aldrich. Toluene solvent (analytic reagent grade) was obtained from Fischer scientific chemicals. Carbon powder, VULCAN® XC72R with Code XVC72R and CAS No. 1333-86-4 was obtained from Cabot Corporation Prod.

XRD powder pattern were recorded on a Panalytical X'Pert $\theta/2\theta$ -diffractometer equipped with Xcelerator detector using automatic divergence slits and Cu $k\alpha_{1/2}$ radiation (40 kV, 40 mA; $\lambda=0.15406$ nm, 0.154443 nm). Cu beta-radiation was excluded using a nickel filter foil. Measurements were performed with $0.005^{\circ}\text{s}^{-1}$. Finely crushed samples were mounted on silicon zero background holders. After data collection obtained intensities were converted from automatic to fixed divergence slits (0.25°) for further analysis. Peak positions and profile were fitted with Pseudo-Voigt function using the HighScore Plus software package (Panalytical). Phase identification was done by using the PDF-2 database of the International Center of Diffraction Data (ICDD).

XPS (X-ray Photoelectron Spectroscopy) measurements were performed on an ESCALAB 220iXL (Thermo Fisher Scientific) with monochromated Al $K\alpha$ radiation ($E = 1486.6$ eV). Samples are prepared on a stainless-steel holder with conductive double-sided adhesive carbon tape. The electron binding energies were obtained with charge compensation using a flood electron source and referenced to the C 1s core level of adventitious carbon at 284.8 eV (C-C and C-H bonds). For quantitative analysis the peaks were deconvoluted with Gaussian-Lorentzian curves using the software Unifit 2021. The peak areas were normalized by the transmission function of the spectrometer and the element specific sensitivity factor of Scofield.

The surface area and porosity were carried out by the N_2 adsorption isotherm using the Brunauer-Emmett-Teller (BET) method on an ASAP 2020 Micromeritics instrument. Before analysis, all samples were degassed at 200°C for 6 h to desorb moisture and impurities from their surfaces. The pore size distributions were calculated using the Barrett-Joyner-Halenda (BJH) model from the desorption branch.

TEM measurements were performed on a JEM-2100 operating at an accelerated voltage of 200 kV. The sample was ultrasonicated in ethanol solution, and a drop was deposited on a copper grid covered with a holey carbon membrane for observation.

GC analyses was performed on an Agilent 7890A instrument (Column: Agilent 19091J-413: $30\text{ m} \times 320\ \mu\text{m} \times 0.25\ \mu\text{m}$, carrier gas: H_2 , FID detection).

All catalytic reactions were carried out in pressure tubes by placing them into a heated aluminum block.

2. Preparation of catalysts

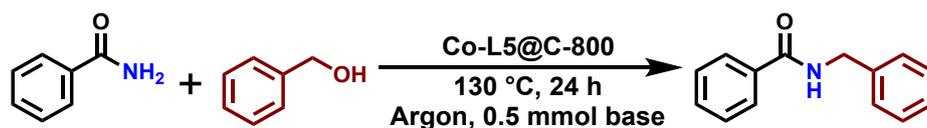
All catalysts mentioned in this work were prepared via the same procedure. A typical procedure for the preparation of the optimal catalyst (Co-L5@C-800) is as follows: In a round bottomed flask, 0.3 mmol of $\text{Co}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$, and 1 mmol of 1,2-phenylenediamine were stirred in methanol (100 mL) for 10 min at room temperature. Then, the carbon (Vulcan XC72R powder) support (2.0 g) without any treatment was added and the mixture was stirred overnight at RT (room temperature). After removing methanol by rotary evaporation at 45 °C, the resulting black solid material was grinded to give a fine powder, which was pyrolyzed under argon at 800 °C for 2 h. The cobalt mass loading in Co-L5@C-800 is found to be 0.937 wt% by ICP-OES analysis (Table 1). A series of catalysts were prepared through the same procedure by changing ligand, support, and calcination temperature. Different catalytic materials such as Co-L1@ZSM-5-800, Co-L1@TiO₂-800, Co-L1@TiO₂, Co-L1@Al₂O₃, Co-L1@MgO-Al₂O₃-800, Co @C-800, Co-L1@C-800, Co-L2@C-800, Co-L3@C-800, Co-L4@C-800, Co-L5@C-400, Co-L5@C-600 and Co-L5@C-1000, were prepared using the described protocol. The procedure, molar amounts or grams of reagents/components used in preparing these catalysts are as follows: 0.3 mmol of $\text{Co}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$, and 1 mmol of ligand were stirred in methanol (100 mL) for 10 min at room temperature. Then, the support (2.0 g) without any treatment was added and the mixture was stirred overnight at room temperature. After removing methanol by rotary evaporation at 45 °C, the resulting black solid material was grinded to give a fine powder, which was pyrolyzed under argon at a certain temperature for 2 h. Further, these obtained materials are used as catalysts.

3. Screening of bases and solvents

We screened many bases to optimize the reaction conditions (**Table S1**). It was clearly found that catalytic system without base is inactive toward the model reaction (**Table S1**, entry 1). The Na_2CO_3 , K_2CO_3 and NaHCO_3 involved systems did not show any reactivity, too (**Table S1**, entries 2-4). The bases Cs_2CO_3 , K_3PO_4 , and NaOH exhibited lower to moderate product yields (**Table S1**, entries 5-7). KOH and $t\text{-BuOK}$ showed similar yield of 96-97% (**Table S1**, entries 8-9). Due to the lower price of KOH , it was selected as the optimal base.

Next, we investigated the effect of KOH loading on the catalytic performance (**Figure S1**). The yield was gradually increased from 30% (0.05 mmol KOH) to 97% (0.3 mmol KOH). However, the yield could not be improved by further increasing the KOH loading. Thus, the optimal KOH loading is selected as 0.3 mmol.

Table S1. Screening the base for the *N*-alkylation of benzamide with benzyl alcohol.



Entry	Base type	Conversion, %	Yield, %
1	without base	0	0
2	Na ₂ CO ₃	0	0
3	K ₂ CO ₃	0	0
4	NaHCO ₃	0	0
5	CsCO ₃	88	87
6	K ₃ PO ₄	68	67
7	NaOH	89	45
8	<i>t</i> -BuOK	97	96
9	KOH	98	97

Reaction conditions: 0.5 mmol benzamide, 0.55 mmol benzyl alcohol, 60 mg catalyst, 0.5 mmol base, 1 atm argon, 3 mL toluene, 130 °C, 24 h. Yields were determined by GC based on benzamide using *n*-hexadecane as standard.

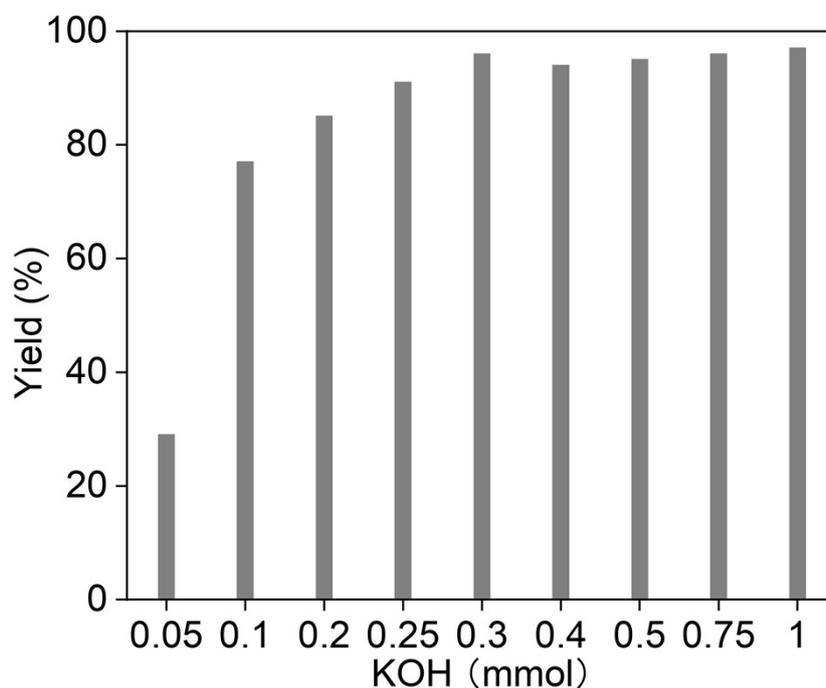
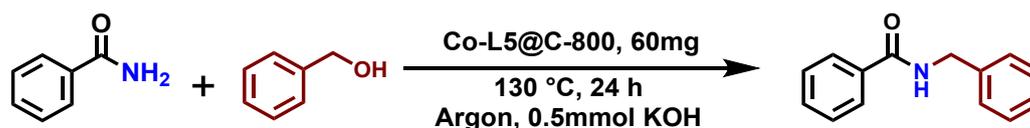


Figure S1. Screening the KOH loading for the *N*-alkylation of benzamide with benzyl alcohol. Reaction conditions: 0.5 mmol benzamide, 0.55 mmol benzyl alcohol, 60 mg catalyst, 1 atm argon, 3 mL toluene, 130 °C, 24 h. Yields were determined by GC based on benzamide using *n*-hexadecane as standard.

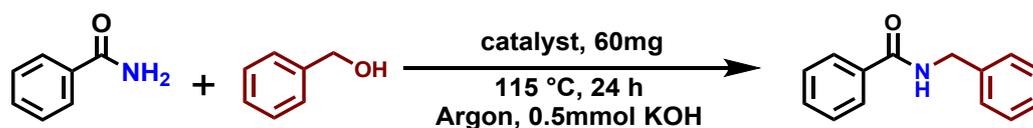
Table S2. Screening the solvents for the *N*-alkylation of benzamide with benzyl alcohol.



Entry	Solvent	Conversion, %	Yield, %
1	Tetrahydrofuran	16	15
2	Methanol	0	0
3	Ethanol	0	0
4	Toluene	98	97
5	n-hexane	35	34

Reaction conditions: 0.5 mmol benzamide, 0.55 mmol benzyl alcohol, 60 mg catalyst, 0.5 mmol KOH, 1 atm argon, 3 mL solvent, 130 °C, 24 h. Yields were determined by GC based on benzamide using n-hexadecane as standard.

Table S3. Screening the metal and ligand content.



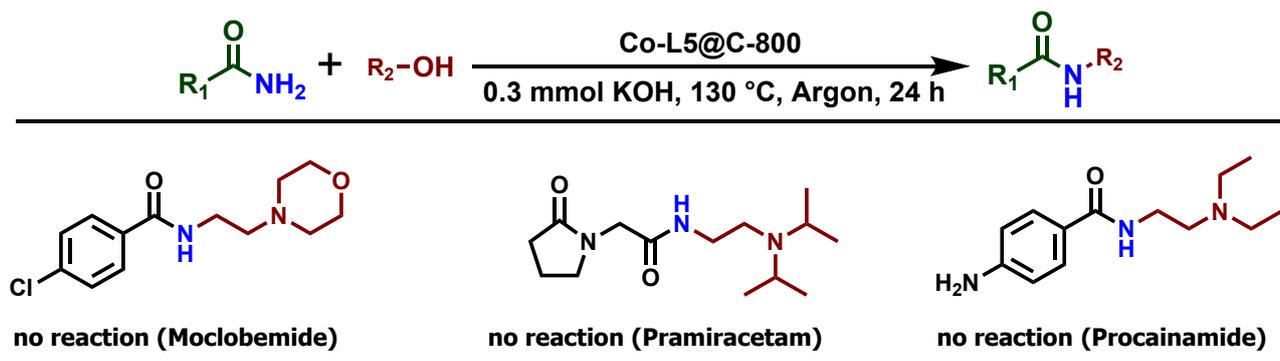
Entry	catalyst	Conversion, %	Yield, %
1	0.5Co-L5@C-800	82	82
2	Co-L5@C-800	87	87
3	1.5Co-L5@C-800	88	88
4	Co-2L5@C-800	86	86
5	Co-3.5L5@C-800	90	90

Reaction conditions: 0.5 mmol benzamide, 0.55 mmol benzyl alcohol, 60 mg catalyst, 0.5 mmol KOH, 1 atm argon, 3 mL solvent, 115 °C, 24 h. Yields were determined by GC based on benzamide using n-hexadecane as standard.

4. General procedure for catalytic reactions

A 25 mL pressure tube was charged with magnetic stirring bar, 0.5 mmol benzamide, 0.55 mmol benzyl alcohol. Then, 3 mL of toluene was added followed by the addition of catalyst (60 mg). Then, the pressure tube was flushed with argon gas three times and fitted with septum and cap. The pressure tube was placed into an aluminum block preheated at 135 °C (placed 30 minutes before counting the

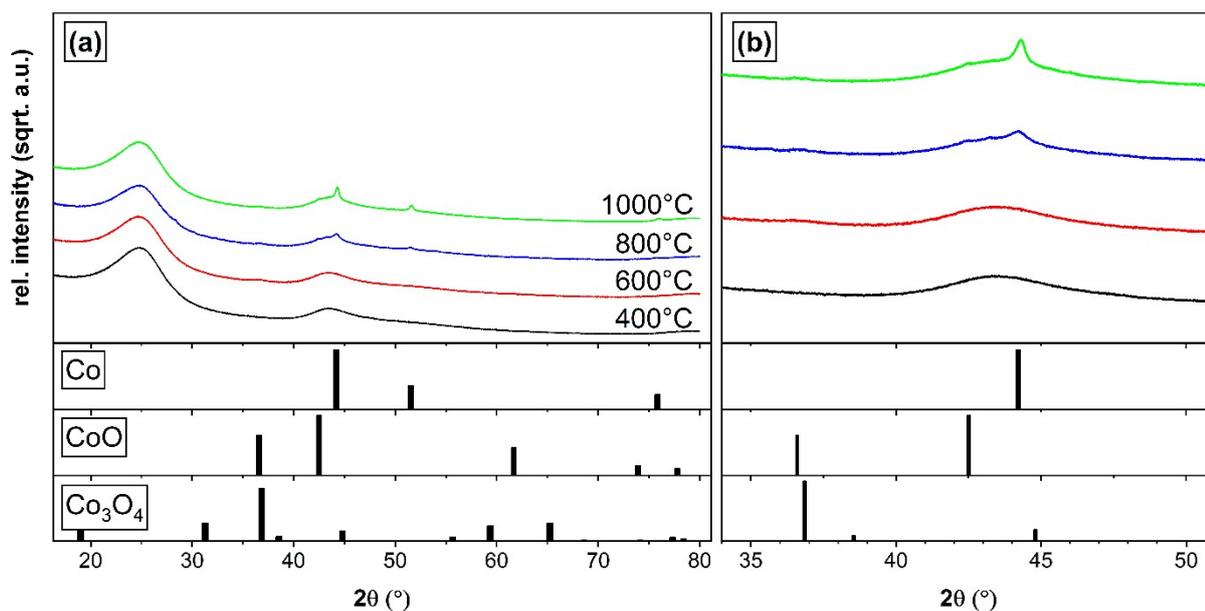
reaction time in order to attain reaction temperature) and the reactions were stirred for required time. During the reaction the inside temperature of the pressure tube was measured to be 130 °C and this temperature was used as the reaction temperature. After the completion of the reactions, the pressure tubes were taken out and products were analyzed by GC and GCMS. The corresponding products were purified by column chromatography (silica, n-hexane, ethyl acetate, methanol). The purified products were analyzed by NMR (¹H, ¹³C) and HRMS.



Scheme S1. Synthesis of selected drug molecules. Reaction conditions: 0.5mmol amide, 0.55 mmol alcohol, 60 mg Co-L5@C-800 (1.9 mol% Co), 0.3 mmol KOH, 3 mL toluene, 130 °C, 24 h, in 1 atm argon

Catalyst recycling. The procedure of catalyst recycling is as following: after the completion of each reaction, the catalyst was separated by centrifugation and washed with methanol followed by drying at 60 °C. Then, the catalyst was reused for the next run without reactivation.

5. Characterization of Co-materials



Fig

ure S2. (a) Powder diffraction pattern of Co-L5@C-400, 600, 800 and 1000 and references, respectively (Co ICDD pdf 00-015-0806, CoO ICDD pdf 01-074-2391, Co₃O₄ ICDD pdf 00-024-1467); (b) magnification of angular region $34^\circ \leq 2\theta \leq 51^\circ$.

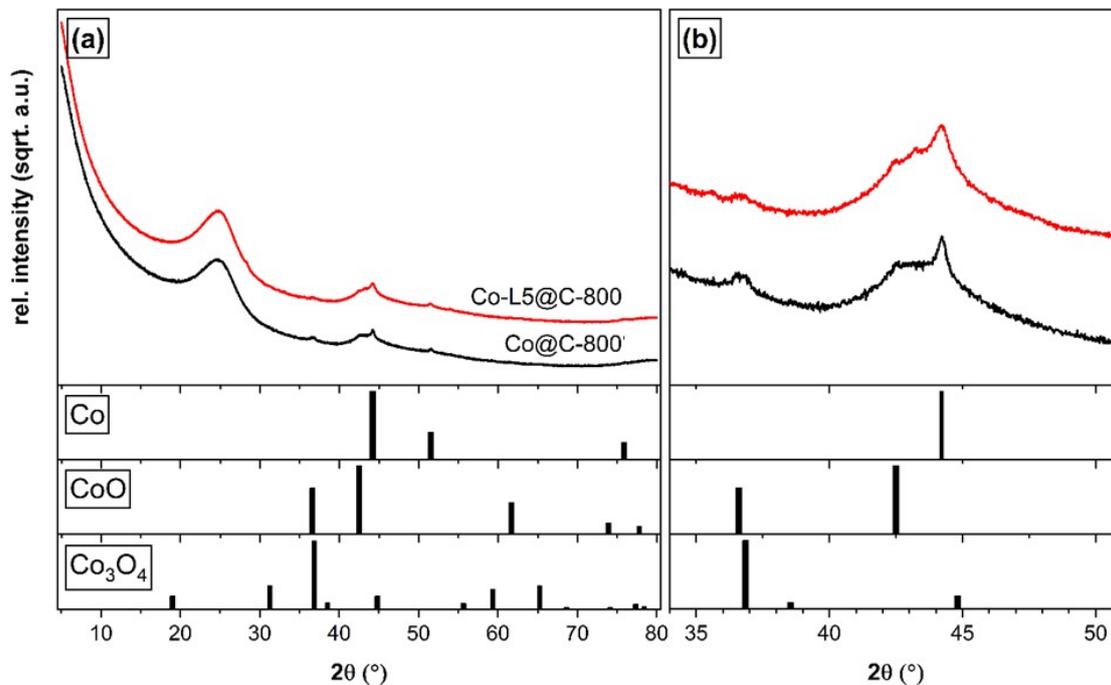


Figure S3. (a) Comparison of X-ray powder diffraction pattern of Co-L5@C-800, Co@C-800 and references, respectively (Co ICDD pdf 00-015-0806, CoO ICDD pdf 01-074-2391, Co₃O₄ ICDD pdf 00-024-1467); (b) magnification of angular region $32^\circ \leq 2\theta \leq 53^\circ$.

Table S4. Elemental distribution of the Co@C-800 derived by EDX.

Element	Family	Atomic Fraction (%)	Mass Fraction (%)
C	k	98.9	97.5
N	k	0.0	0.0
O	k	0.9	1.1
Co	k	0.3	1.4

Table S5. Elemental distribution of the Co-L5@C-800 derived by EDX.

Element	Family	Atomic Fraction (%)	Mass Fraction (%)
C	k	97.6	93.7
N	k	0.0	0.0
O	k	1.5	1.9
Co	k	0.9	4.4

Table S6. Elemental composition of the near-surface region (at %) derived by XPS.

Element	Co-L5@C-400	Co-L5@C-600	Co-L5@C-800	Co-L5@C-1000	Co@C-800
C	96.8	97.9	97.3	98.8	98.3
N	1.2	0.7	0.4	0.1	0.0
Co	0.3	0.2	0.1	0.1	0.1
O	1.2	0.8	1.6	0.6	1.0
S	0.2	0.1	0.2	0.2	0.2
Si	0.2	0.3	0.3	0.4	0.4

Table S7. The proportion of different forms of cobalt on the near-surface region (%) derived by XPS.

Co form	Co-L5@C-400	Co-L5@C-600	Co-L5@C-800	Co-L5@C-1000	Co@C-800
Co ⁰	0	0	12.0	22.7	0
Co _x O _y	100	100	88.0	77.3	100

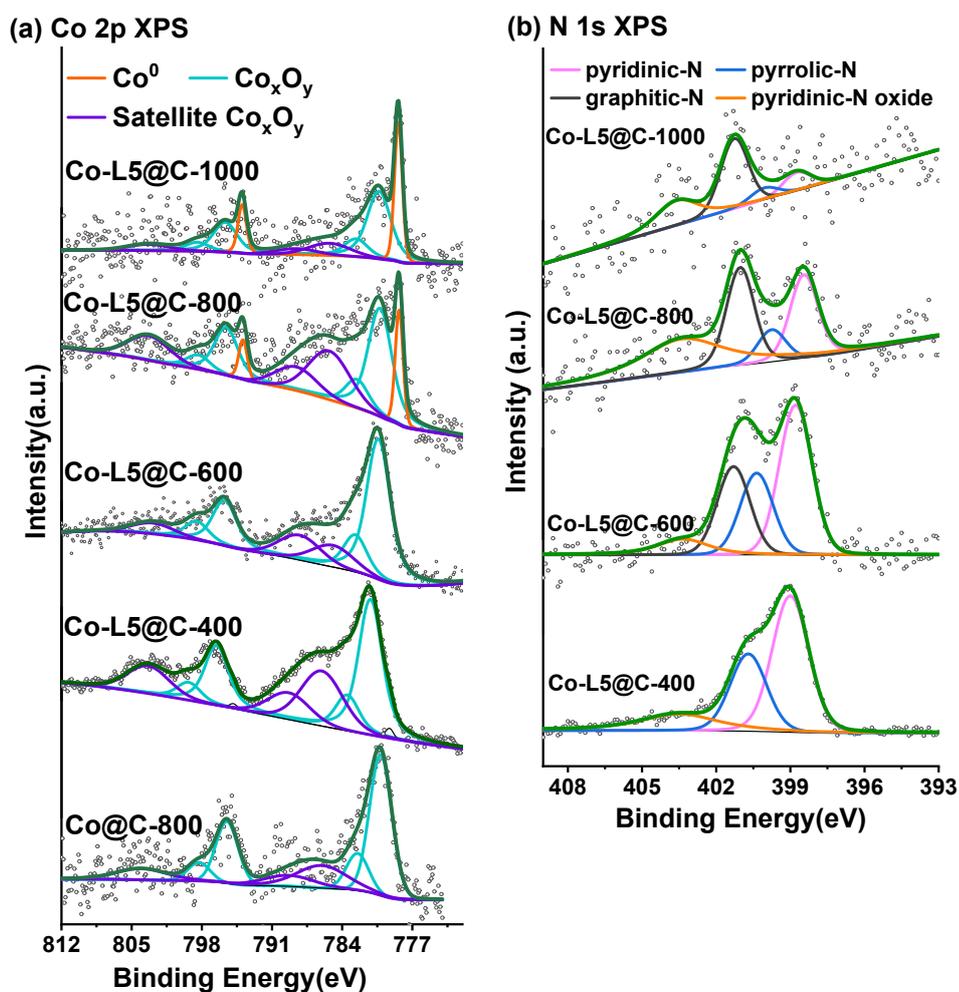


Figure S4. (a) Co 2p XPS and (b) N 1s XPS spectra of selected materials.

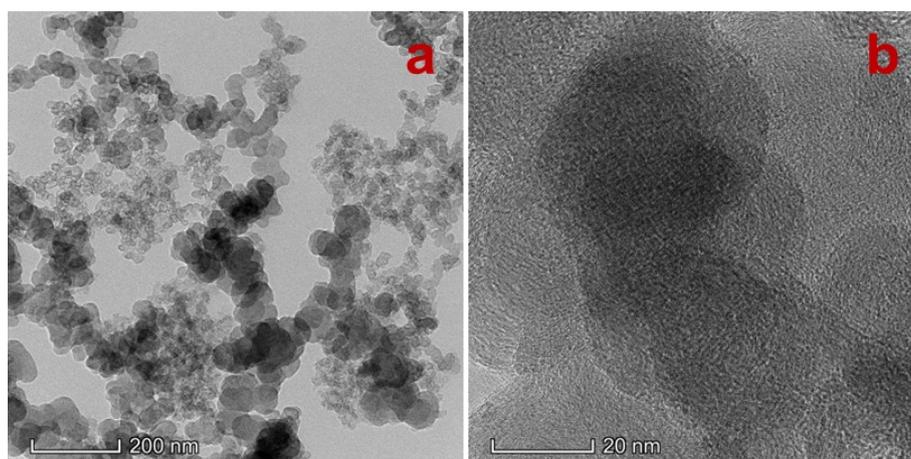


Figure S5. Structure characterization of Co-L5@C-400. (a-b) TEM images, (a) shows the metal complexes were uniformly distributed on the carbon support, (b) magnification TEM image at 20 nm.

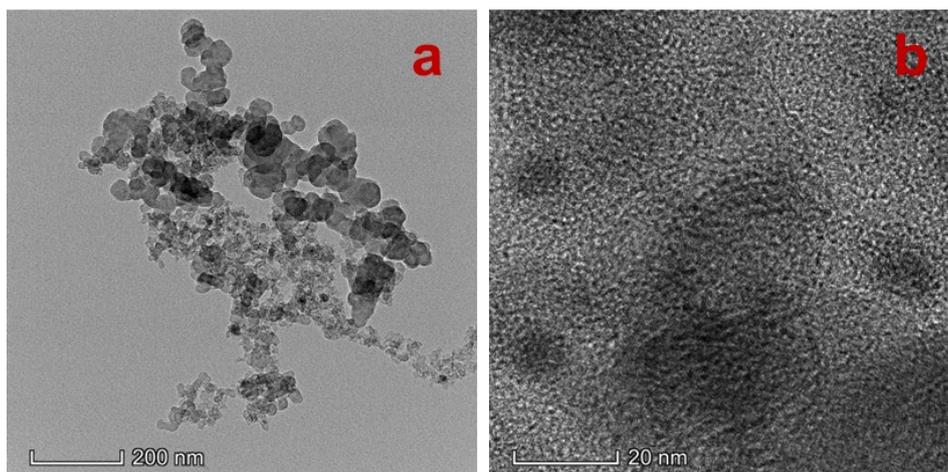


Figure S6. Structure characterization of Co-L5@C-600. (a-b) TEM images show (a) the metal complexes were uniformly distributed on the carbon support, (b) magnification TEM image at 20 nm.

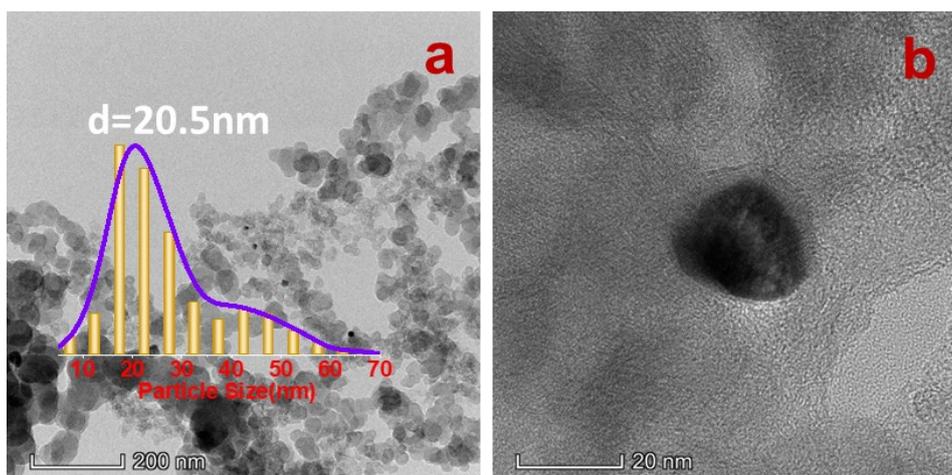


Figure S7. Structure characterization of Co-L5@C-1000. (a-b) TEM images, (a) shows the Co nanoparticles were distributed on the carbon support with an average diameter of 20.5 nm, (b) magnification TEM image at 20nm.

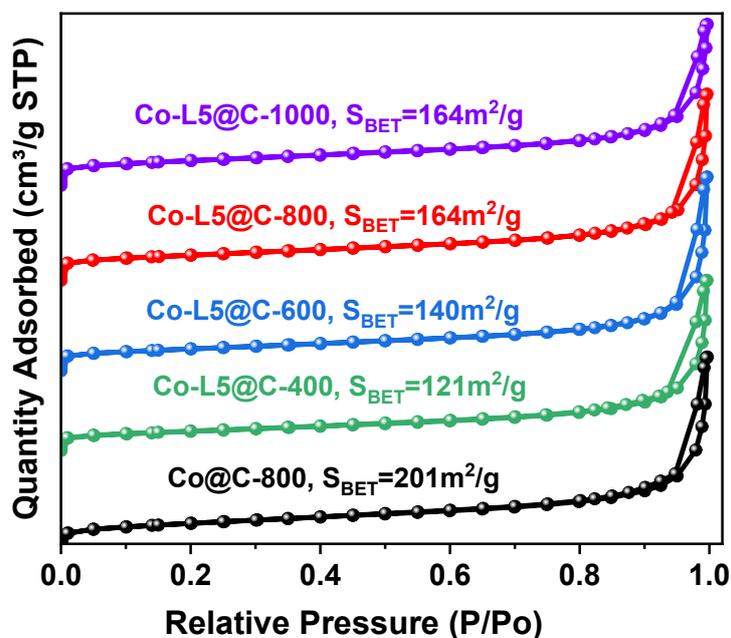


Figure S8. BET spectra of selected materials.

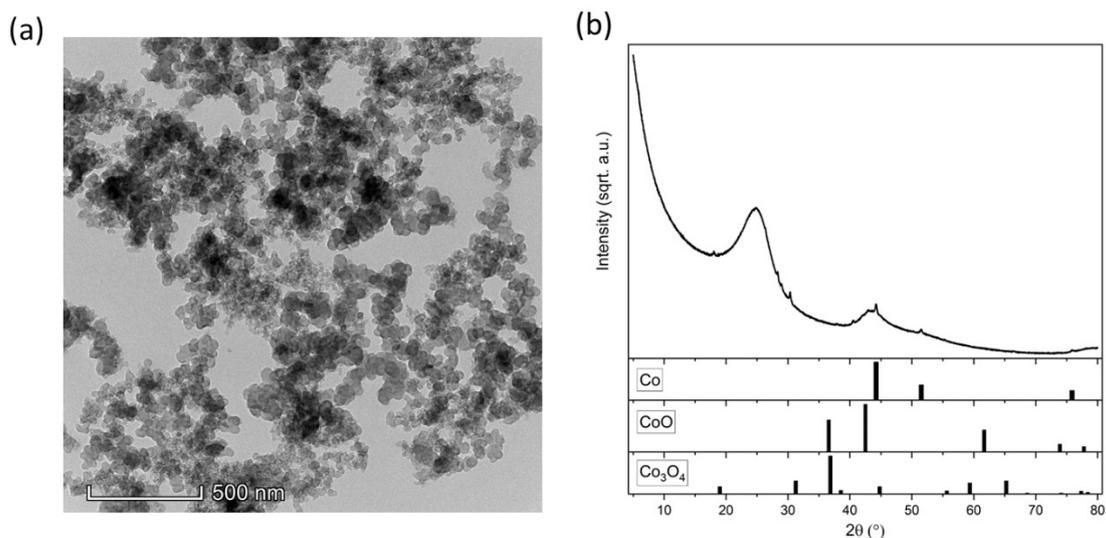


Figure S9. Characterization of catalyst Co-L5@C-800 after recycling. (a) TEM image, (b) Comparison of X-ray powder diffraction pattern of Co-L5@C-800, Co@C-800 and references, respectively (Co ICDD pdf 00-015-0806, CoO ICDD pdf 01-074-2391, Co₃O₄ ICDD pdf 00-024-1467).

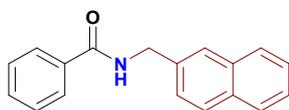
6. NMR data

NMR spectra were recorded on Bruker AV 300 and 400 spectrometers. All chemical shifts (δ) are reported in parts per million (ppm) and coupling constants (J) in Hz. Mass spectra were recorded on an AMD 402/3 or a HP 5973 mass selective detector.

HRMS data were recorded on (1) ESI-HRMS: HPLC System 1200 /ESI-TOF-MS 6210 (Agilent) and (2) EI-HRMS: Mass Spectrometer MAT 95XP (Thermo Electron), 70eV. GC-MS was performed on an ISQ Trace 1300 in the electron ionization (EI) mode.

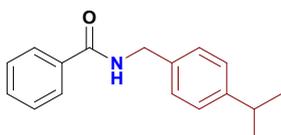
Isolated yields are reported. In the following, chemical shifts of the products are given.

1a: N-(naphthalen-2-ylmethyl)benzamide^[1]



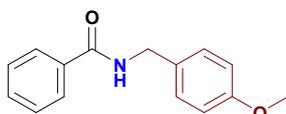
¹H NMR (300 MHz, CDCl₃) δ 7.87 – 7.77 (m, 6H), 7.54 – 7.39 (m, 6H), 6.75 (s, 1H), 4.79 (dd, J = 5.8, 0.8 Hz, 2H). **¹³C NMR (75 MHz, CDCl₃)** δ 167.50, 135.70, 134.37, 133.40, 132.82, 131.60, 128.62, 127.78, 127.73, 127.06, 126.52, 126.33, 126.00, 77.52, 77.10, 76.67, 44.25.

1b: N-(4-isopropylbenzyl)benzamide^[2]



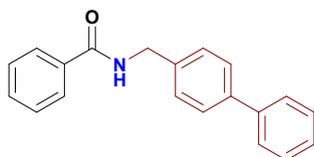
¹H NMR (300 MHz, DMSO) δ 9.03 (t, J = 5.9 Hz, 1H), 7.99 – 7.88 (m, 2H), 7.58 – 7.42 (m, 3H), 7.29 – 7.23 (m, 2H), 7.22 – 7.17 (m, 2H), 4.48 (dd, J = 5.8, 1.7 Hz, 2H), 2.86 (pd, J = 7.0, 2.1 Hz, 1H), 1.20 (d, J = 2.6 Hz, 3H), 1.18 (d, J = 2.6 Hz, 3H). **¹³C NMR (75 MHz, DMSO)** δ 166.61, 147.36, 147.24, 137.57, 134.89, 131.64, 128.74, 127.80, 127.73, 127.07, 126.62, 126.38, 42.90, 40.83, 40.55, 40.27, 39.99, 39.71, 39.44, 39.16, 33.62, 24.45, 24.40.

1c: N-(4-methoxybenzyl)benzamide^[2]



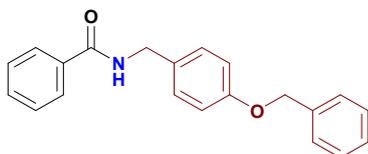
¹H NMR (300 MHz, CDCl₃) δ 7.74 – 7.64 (m, 2H), 7.42 – 7.34 (m, 1H), 7.18 – 7.13 (m, 2H), 6.79 – 6.74 (m, 2H), 6.59 (s, 1H), 4.44 (d, *J* = 5.6 Hz, 2H), 3.69 (s, 3H). **¹³C NMR (75 MHz, CDCl₃)** δ 167.37, 159.06, 134.46, 131.48, 130.38, 129.27, 128.62, 128.54, 127.02, 114.12, 113.90, 77.54, 77.12, 76.69, 55.32, 43.57.

1d: N-(4-phenylbenzyl)benzamide^[3]



¹H NMR (300 MHz, DMSO) δ 9.12 (t, *J* = 6.0 Hz, 1H), 8.01 – 7.89 (m, 2H), 7.68 – 7.61 (m, 4H), 7.59 – 7.29 (m, 8H), 4.55 (d, *J* = 6.2 Hz, 2H). **¹³C NMR (75 MHz, DMSO)** δ 166.71, 140.48, 139.44, 139.19, 139.03, 134.80, 131.74, 129.38, 128.81, 128.33, 127.78, 127.74, 127.51, 127.10, 127.05, 127.02, 126.86, 42.85, 40.82, 40.54, 40.26, 39.98, 39.70, 39.43, 39.15.

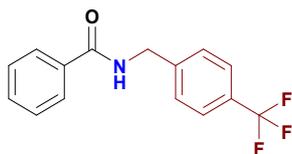
1e: N-(4-(benzyloxy)benzyl)benzamide



¹H NMR (300 MHz, CDCl₃) δ 7.88 – 7.75 (m, 1H), 7.58 – 7.21 (m, 9H), 7.02 – 6.94 (m, 2H), 6.57 (s, 1H), 5.09 (d, *J* = 2.7 Hz, 2H), 4.66 – 4.54 (m, 2H). **¹³C NMR (75 MHz, CDCl₃)** δ 167.40, 158.37, 158.31, 137.00, 136.93, 134.42, 131.54, 130.59, 129.34, 128.67, 128.64, 128.59, 128.03, 128.00, 127.48, 127.01, 115.38, 115.14, 114.93, 77.53, 77.11, 76.68, 70.07, 70.05, 43.63.

HRMS (ESI-TOF): *m/z* calcd. for C₂₁H₁₉NO₂+H⁺ ([M+H⁺]) 318.1494, found 318.1502.

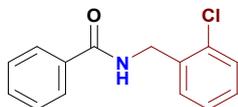
1f: N-(4-(trifluoromethyl)benzyl)benzamide^[3]



¹H NMR (300 MHz, DMSO) δ 8.94 (t, *J* = 5.8 Hz, 1H), 8.01 – 7.87 (m, 2H), 7.58 – 7.43 (m, 3H), 7.30 – 7.23 (m, 1H), 7.21 – 7.12 (m, 3H), 4.49 (d, *J* = 5.9 Hz, 2H). **¹³C NMR (75 MHz, DMSO)** δ

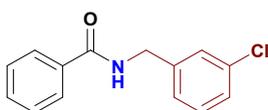
166.69, 137.65, 135.95, 134.87, 131.67, 130.35, 128.77, 127.78, 127.75, 127.20, 126.19, 41.17, 40.83, 40.55, 40.28, 40.00, 39.72, 39.44, 39.16.

1g: N-(2-chlorobenzyl)benzamide^[4]



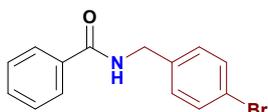
¹H NMR (400 MHz, CDCl₃) δ 7.85 – 7.77 (m, 2H), 7.53 – 7.29 (m, 6H), 7.25 – 7.22 (m, 1H), 6.88 (d, J = 59.5 Hz, 1H), 4.67 (dd, J = 34.3, 5.8 Hz, 2H). **¹³C NMR (101 MHz, CDCl₃)** δ 167.51, 167.49, 138.30, 135.66, 134.38, 134.25, 133.62, 131.61, 131.53, 130.16, 129.55, 128.95, 128.75, 128.59, 128.56, 127.87, 127.55, 127.14, 127.07, 127.05, 77.44, 77.32, 77.12, 76.80, 44.07, 41.99.

1h: N-(3-chlorobenzyl)benzamide^[5]



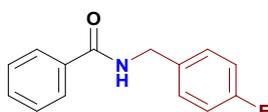
¹H NMR (300 MHz, DMSO) δ 9.05 (t, J = 6.2 Hz, 1H), 7.94 – 7.87 (m, 2H), 7.63 – 7.44 (m, 4H), 7.34 – 7.31 (m, 3H), 4.58 – 4.45 (m, 2H). **¹³C NMR (75 MHz, DMSO)** δ 140.17, 131.71, 128.79, 128.75, 127.70, 127.65, 127.19, 43.05, 40.81, 40.53, 40.25, 39.97, 39.70, 39.42, 39.14.

1j: N-(4-bromobenzyl)benzamide^[1]



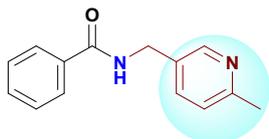
¹H NMR (300 MHz, CDCl₃) δ 7.58 – 7.47 (m, 4H), 7.37 (dddd, J = 8.3, 3.8, 2.8, 0.9 Hz, 4H), 7.31 – 7.12 (m, 1H), 4.68 (d, J = 7.1 Hz, 2H). **¹³C NMR (75 MHz, CDCl₃)** δ 140.68, 139.89, 128.81, 127.49, 127.35, 127.12, 77.46, 77.04, 76.62, 65.43, 65.14, 29.73.

1k: N-(4-fluorobenzyl)benzamide^[6]



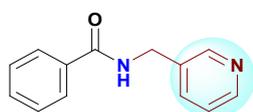
¹H NMR (400 MHz, CDCl₃) δ 7.83 – 7.77 (m, 2H), 7.54 – 7.48 (m, 1H), 7.45 – 7.40 (m, 2H), 7.28 (s, 2H), 7.11 – 6.94 (m, 2H), 4.62 – 4.54 (m, 2H). **¹³C NMR (101 MHz, CDCl₃)** δ 167.48, 167.40, 163.74, 163.42, 161.29, 160.98, 134.41, 134.26, 134.17, 134.13, 131.63, 131.54, 129.57, 129.49, 129.37, 129.29, 128.60, 128.58, 127.03, 126.99, 115.66, 115.63, 115.45, 115.42, 115.09, 77.41, 77.09, 76.77, 43.32.

1m: N-((6-methylpyridin-3-yl)methyl)benzamide^[8]



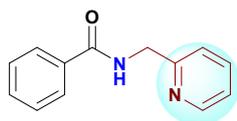
¹H NMR (300 MHz, DMSO) δ 9.08 (t, *J* = 5.9 Hz, 1H), 8.43 (dd, *J* = 2.3, 0.8 Hz, 1H), 7.93 – 7.86 (m, 2H), 7.64 – 7.60 (m, 1H), 7.56 – 7.43 (m, 3H), 7.20 (d, *J* = 7.9 Hz, 1H), 4.47 (d, *J* = 5.9 Hz, 2H), 2.44 (d, *J* = 1.9 Hz, 3H). **¹³C NMR (75 MHz, DMSO)** δ 166.77, 156.78, 156.69, 148.61, 136.00, 134.66, 132.50, 131.76, 128.79, 127.70, 123.16, 122.99, 40.81, 40.66, 40.53, 40.25, 39.97, 39.70, 39.42, 39.14, 24.15, 24.11.

1n: N-(pyridin-3-ylmethyl)benzamide^[9]



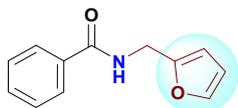
¹H NMR (300 MHz, DMSO) δ 9.26 – 9.07 (m, 1H), 8.60 – 8.44 (m, 2H), 8.06 – 7.86 (m, 2H), 7.81 – 7.67 (m, 1H), 7.57 – 7.31 (m, 4H), 4.58 – 4.48 (m, 2H). **¹³C NMR (75 MHz, DMSO)** δ 166.85, 151.17, 151.12, 149.31, 148.58, 148.51, 148.48, 137.07, 135.62, 134.58, 131.80, 129.75, 128.80, 128.50, 127.75, 123.94, 40.90, 40.78, 40.50, 40.22, 39.94, 39.66, 39.39, 39.11.

1o: N-(pyridin-2-ylmethyl)benzamide^[8]



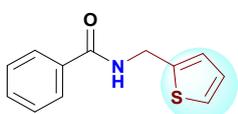
¹H NMR (300 MHz, CDCl₃) δ 8.58 – 8.49 (m, 1H), 7.92 – 7.38 (m, 6H), 7.21 (s, 2H), 4.82 – 4.70 (m, 2H). **¹³C NMR (75 MHz, CDCl₃)** δ 167.45, 156.37, 148.92, 148.48, 136.92, 136.73, 134.33, 131.49, 128.56, 128.52, 128.50, 127.13, 122.45, 122.28, 122.24, 120.58, 77.54, 77.11, 76.69, 44.76.

1p: N-(furan-2-ylmethyl)benzamide^[6]



¹H NMR (300 MHz, DMSO) δ 8.97 (t, $J = 5.7$ Hz, 1H), 8.05 – 7.79 (m, 2H), 7.61 – 7.40 (m, 4H), 6.40 (ddd, $J = 3.8, 3.2, 1.8$ Hz, 1H), 6.28 (dp, $J = 2.8, 0.9$ Hz, 1H), 4.48 (dd, $J = 5.8, 0.9$ Hz, 2H). **¹³C NMR (75 MHz, DMSO)** δ 166.53, 152.92, 142.56, 142.46, 134.58, 131.77, 128.77, 127.75, 110.95, 110.75, 107.31, 107.29, 40.82, 40.54, 40.26, 39.99, 39.71, 39.43, 39.15, 36.51.

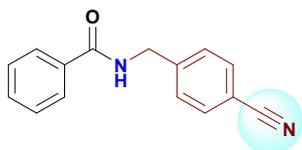
1q: N-(thiophen-2-ylmethyl)benzamide^[6]



¹H NMR (300 MHz, CDCl₃) δ 7.83 – 7.78 (m, 2H), 7.55 – 7.49 (m, 1H), 7.47 – 7.40 (m, 2H), 7.26 (s, 1H), 7.05 (dq, $J = 3.4, 0.9$ Hz, 1H), 6.99 (dd, $J = 5.1, 3.5$ Hz, 1H), 6.67 (d, $J = 17.0$ Hz, 1H), 4.82 (dd, $J = 5.6, 0.8$ Hz, 2H).

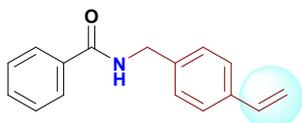
¹³C NMR (75 MHz, CDCl₃) δ 167.20, 140.83, 134.16, 131.66, 128.61, 128.59, 127.04, 126.98, 126.90, 126.25, 125.36, 124.97, 77.50, 77.07, 76.65, 39.11, 38.84.

1r: N-(4-cyanobenzyl)benzamide^[2]



¹H NMR (300 MHz, DMSO) δ 9.09 (t, $J = 6.0$ Hz, 1H), 7.94 – 7.88 (m, 2H), 7.86 – 7.80 (m, 2H), 7.58 – 7.45 (m, 2H), 7.42 – 7.25 (m, 3H), 4.54 (t, $J = 5.8$ Hz, 2H). **¹³C NMR (75 MHz, DMSO)** δ 168.29, 146.37, 133.09, 129.04, 128.83, 128.01, 127.81, 127.35, 126.40, 62.94, 40.80, 40.53, 40.25, 39.97, 39.69, 39.41, 39.13.

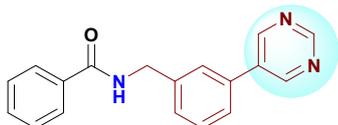
1s: N-(4-vinylbenzyl)benzamide



¹H NMR (300 MHz, DMSO) δ 9.04 (t, $J = 6.0$ Hz, 1H), 7.94 – 7.86 (m, 2H), 7.59 – 7.40 (m, 5H), 7.34 – 7.26 (m, 2H), 6.72 (dd, $J = 17.7, 10.9$ Hz, 1H), 5.80 (dd, $J = 17.7, 1.1$ Hz, 1H), 5.27 – 5.19 (m, 1H), 4.48 (d, $J = 6.0$ Hz, 2H). **¹³C NMR (75 MHz, DMSO)** δ 166.66, 136.87, 131.72, 128.79, 127.95, 127.70, 126.55, 114.29, 42.87, 40.83, 40.55, 40.28, 40.00, 39.72, 39.44, 39.16.

HRMS (ESI-TOF): m/z calcd. for C₁₆H₁₅NO+H⁺ ([M+H⁺]) 238.1232, found 238.1229.

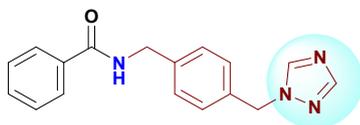
1t: N-(3-(pyrimidin-5-yl)benzyl)benzamide



¹H NMR (300 MHz, DMSO) δ 9.17 (d, $J = 0.9$ Hz, 1H), 9.09 (d, $J = 1.7$ Hz, 3H), 7.97 – 7.82 (m, 2H), 7.76 – 7.70 (m, 1H), 7.66 (ddd, $J = 7.5, 1.9, 1.4$ Hz, 1H), 7.53 – 7.39 (m, 5H), 4.57 (dd, $J = 5.9, 3.8$ Hz, 2H). **¹³C NMR (75 MHz, DMSO)** δ 166.83, 157.78, 157.71, 155.16, 155.14, 141.35, 134.76, 134.23, 133.72, 131.75, 129.85, 129.60, 128.80, 128.21, 127.76, 127.46, 126.46, 125.90, 125.75, 125.38, 43.08, 40.80, 40.52, 40.25, 39.97, 39.69, 39.41, 39.13.

HRMS (ESI-TOF): m/z calcd. for C₁₈H₁₅N₃O+Na⁺ ([M+Na⁺]) 312.1107, found 312.1110.

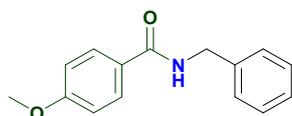
1u: N-(4-((1H-1,2,4-triazol-1-yl) methyl)benzyl)benzamide



¹H NMR (300 MHz, DMSO) δ 9.08 (t, $J = 6.0$ Hz, 1H), 8.68 (s, 1H), 8.00 (s, 1H), 7.96 – 7.90 (m, 2H), 7.55 – 7.43 (m, 3H), 7.37 – 7.23 (m, 4H), 5.41 (s, 2H), 4.50 (d, $J = 6.0$ Hz, 2H). **¹³C NMR (75 MHz, DMSO)** δ 166.76, 152.18, 144.60, 139.99, 135.23, 134.80, 131.72, 128.78, 128.41, 128.01, 127.74, 52.37, 42.87, 40.80, 40.53, 40.25, 39.97, 39.69, 39.41, 39.13.

HRMS (ESI-TOF): m/z calcd. for C₁₇H₁₆N₄O+H⁺ ([M+H⁺]) 293.1402, found 293.1403.

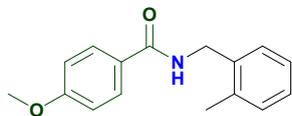
2a: N-benzyl-4-methoxybenzamide^[11]



¹H NMR (400 MHz, DMSO) δ 8.92 (t, $J = 6.0$ Hz, 1H), 7.95 – 7.87 (m, 2H), 7.33 (d, $J = 4.6$ Hz, 4H), 7.28 – 7.21 (m, 1H), 7.05 – 6.98 (m, 2H), 4.49 (d, $J = 6.0$ Hz, 2H), 3.81 (s, 3H). **¹³C NMR (101**

MHz, DMSO) δ 166.19, 162.07, 140.39, 129.56, 128.72, 127.65, 127.14, 127.03, 113.98, 55.80, 43.01, 40.60, 40.39, 40.18, 39.97, 39.76, 39.55, 39.34.

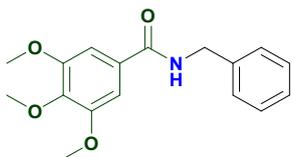
2b: 4-methoxy-N-(2-methylbenzyl)benzamide



¹H NMR (300 MHz, CDCl₃) δ 7.82 – 7.72 (m, 2H), 7.41 – 7.09 (m, 4H), 6.98 – 6.87 (m, 2H), 6.36 (s, 1H), 4.62 (d, J = 5.3 Hz, 2H), 3.85 (s, 3H), 2.37 (s, 3H). **¹³C NMR (75 MHz, CDCl₃)** δ 166.82, 162.21, 136.60, 136.01, 130.59, 128.81, 128.63, 127.81, 126.61, 126.26, 113.76, 77.52, 77.09, 76.67, 55.42, 42.26, 19.08.

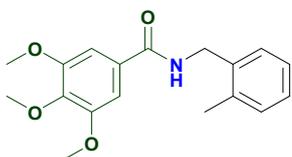
HRMS (ESI-TOF): m/z calcd. for C₁₆H₁₇NO₂+H⁺ ([M+H⁺]) 256.1337, found 256.1340.

2d: N-benzyl-3,4,5-trimethoxybenzamide^[13]



¹H NMR (400 MHz, DMSO) δ 9.01 (t, J = 6.0 Hz, 1H), 7.35 – 7.32 (m, 4H), 7.28 – 7.23 (m, 3H), 4.50 (d, J = 5.9 Hz, 2H), 3.84 (s, 6H), 3.71 (s, 3H). **¹³C NMR (101 MHz, DMSO)** δ 166.03, 153.07, 140.43, 140.17, 129.88, 128.77, 128.51, 127.72, 127.24, 127.09, 126.88, 105.28, 60.54, 56.45, 43.17, 40.59, 40.38, 40.17, 39.96, 39.76, 39.55, 39.34.

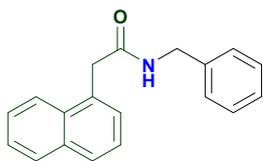
2e: 3,4,5-trimethoxy-N-(2-methylbenzyl)benzamide



¹H NMR (300 MHz, CDCl₃) δ 7.36 – 7.31 (m, 1H), 7.28 – 7.20 (m, 3H), 7.03 (s, 2H), 6.17 (s, 1H), 4.67 (d, J = 5.3 Hz, 2H), 3.91 (d, J = 5.4 Hz, 9H), 2.41 (s, 3H). **¹³C NMR (75 MHz, CDCl₃)** δ 153.26, 130.72, 128.87, 128.01, 126.32, 104.40, 77.45, 77.03, 76.61, 60.93, 56.41, 42.51, 19.13.

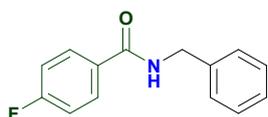
HRMS (ESI-TOF): m/z calcd. for C₁₈H₂₁NO₄+H⁺ ([M+H⁺]) 316.1549 found 316.1553.

2f: N-benzyl-2-(naphthalen-1-yl)acetamide^[14]



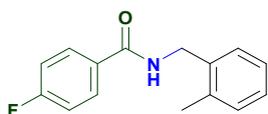
¹H NMR (300 MHz, DMSO) δ 8.65 (t, $J = 5.9$ Hz, 1H), 8.16 – 8.08 (m, 1H), 7.96 – 7.90 (m, 1H), 7.86 – 7.79 (m, 1H), 7.55 – 7.50 (m, 2H), 7.48 – 7.43 (m, 2H), 7.32 – 7.21 (m, 5H), 4.30 (d, $J = 5.9$ Hz, 2H), 3.98 (s, 2H). **¹³C NMR (75 MHz, DMSO)** δ 170.55, 139.97, 133.83, 133.25, 132.44, 128.84, 128.71, 128.33, 127.70, 127.54, 127.23, 126.37, 126.11, 125.99, 124.78, 42.74, 40.83, 40.55, 40.27, 39.99, 39.72, 39.44, 39.16.

2g: N-benzyl-4-fluorobenzamide^[6]



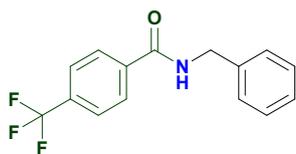
¹H NMR (400 MHz, DMSO) δ 9.07 (dt, $J = 10.9, 5.9$ Hz, 1H), 8.04 – 7.85 (m, 2H), 7.52 – 7.45 (m, 1H), 7.40 – 7.28 (m, 5H), 7.25 (dddd, $J = 7.1, 4.3, 3.2, 1.7$ Hz, 1H), 4.49 (dd, $J = 6.0, 3.1$ Hz, 2H). **¹³C NMR (101 MHz, DMSO)** δ 166.69, 165.64, 165.60, 140.16, 140.04, 134.80, 131.72, 131.27, 131.24, 130.41, 130.32, 128.80, 128.76, 128.51, 127.71, 127.68, 127.66, 127.24, 127.20, 126.88, 115.83, 115.61, 43.13, 43.07, 40.59, 40.38, 40.17, 39.96, 39.75, 39.54, 39.33.

2h: 4-fluoro-N-(2-methylbenzyl)benzamide^[15]



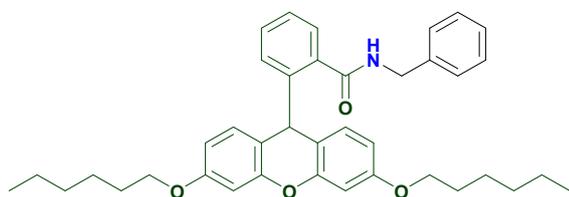
¹H NMR (300 MHz, CDCl₃) δ 7.86 – 7.74 (m, 2H), 7.52 – 7.40 (m, 1H), 7.34 – 7.28 (m, 1H), 7.28 – 7.18 (m, 3H), 7.13 – 7.07 (m, 1H), 6.38 (s, 1H), 4.64 (dd, $J = 5.4, 4.1$ Hz, 2H), 2.38 (d, $J = 1.9$ Hz, 3H). **¹³C NMR (75 MHz, CDCl₃)** δ 167.30, 166.42, 166.23, 163.08, 136.63, 136.60, 135.76, 135.67, 134.35, 131.57, 130.67, 130.54, 129.38, 129.27, 128.70, 128.62, 127.97, 127.94, 126.96, 126.32, 115.76, 115.47, 77.49, 77.07, 76.65, 42.40, 42.36, 19.08.

2i: N-benzyl-4-(trifluoromethyl)benzamide^[7]



¹H NMR (300 MHz, DMSO) δ 9.29 (t, J = 5.9 Hz, 1H), 8.14 – 8.06 (m, 2H), 7.90 – 7.82 (m, 2H), 7.36 – 7.21 (m, 5H), 4.52 (d, J = 6.0 Hz, 2H). **¹³C NMR (75 MHz, DMSO)** δ 165.55, 139.76, 138.56, 131.86, 131.44, 128.79, 128.65, 127.73, 127.31, 126.22, 125.91, 125.86, 125.81, 125.75, 122.61, 43.24, 40.81, 40.53, 40.25, 39.98, 39.70, 39.42, 39.14.

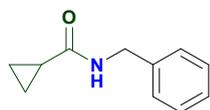
2j: N-benzyl-2-(3,6-bis(hexyloxy)-9H-xanthen-9-yl)benzamide



¹H NMR (300 MHz, CDCl₃) δ 7.42 – 7.30 (m, 6H), 7.25 (ddd, J = 8.1, 6.9, 1.5 Hz, 1H), 7.10 (ddd, J = 22.9, 8.0, 0.9 Hz, 4H), 6.70 (d, J = 2.5 Hz, 2H), 6.61 (t, J = 5.8 Hz, 1H), 6.52 (dd, J = 8.5, 2.6 Hz, 2H), 5.83 (s, 1H), 4.62 (d, J = 5.8 Hz, 2H), 3.95 (t, J = 6.5 Hz, 4H), 1.88 – 1.76 (m, 4H), 1.56 – 1.34 (m, 12H), 1.03 – 0.92 (m, 6H). **¹³C NMR (75 MHz, CDCl₃)** δ 169.97, 158.83, 151.53, 151.49, 146.16, 138.23, 135.14, 131.25, 130.80, 130.74, 128.85, 128.29, 127.94, 127.67, 126.40, 126.17, 116.83, 110.73, 101.71, 77.63, 77.20, 76.78, 68.17, 44.12, 38.54, 31.66, 29.26, 25.81, 22.74, 22.69, 14.15.

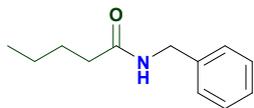
HRMS (ESI-TOF): m/z calcd. for C₃₉H₄₅NO₄+H⁺ ([M+H⁺]) 592.3427, found 592.3438.

2k: N-benzylcyclopropanecarboxamide^[16]



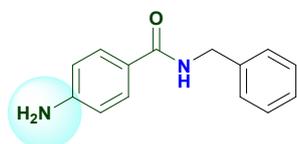
¹H NMR (300 MHz, DMSO) δ 8.55 (s, 1H), 7.36 – 7.21 (m, 5H), 4.29 (d, J = 5.9 Hz, 2H), 1.66 – 1.55 (m, 1H), 0.94 – 0.55 (m, 4H). **¹³C NMR (75 MHz, DMSO)** δ 173.03, 140.16, 128.76, 128.71, 127.75, 127.62, 127.21, 127.14, 42.71, 42.43, 40.82, 40.55, 40.27, 39.99, 39.72, 39.44, 39.16, 14.13, 14.03, 6.71.

2l: N-benzylpentanamide^[17]



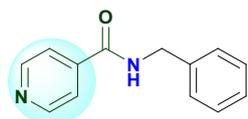
¹H NMR (300 MHz, DMSO) δ 8.29 (s, 1H), 7.52 – 6.90 (m, 5H), 4.26 (d, J = 5.9 Hz, 2H), 2.19 – 2.10 (m, 2H), 1.58 – 1.45 (m, 2H), 1.35 – 1.20 (m, 2H), 0.92 – 0.82 (m, 3H). **¹³C NMR (75 MHz, DMSO)** δ 172.58, 140.24, 128.71, 127.61, 127.13, 42.43, 40.83, 40.55, 40.27, 39.99, 39.72, 39.44, 39.16, 35.54, 27.95, 22.30, 14.19.

2m: 4-amino-N-benzylbenzamide^[18]



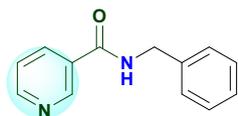
¹H NMR (300 MHz, CDCl₃) δ 7.69 – 7.59 (m, 2H), 7.42 – 7.29 (m, 5H), 6.73 – 6.58 (m, 2H), 6.34 (s, 1H), 4.63 (d, J = 5.7 Hz, 2H), 4.32 – 3.55 (m, 2H). **¹³C NMR (75 MHz, CDCl₃)** δ 167.11, 149.68, 138.66, 128.76, 128.73, 127.91, 127.48, 123.88, 114.16, 77.48, 77.06, 76.63, 43.98.

2n: N-benzylisonicotinamide^[8]



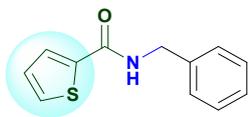
¹H NMR (300 MHz, DMSO) δ 9.35 (t, J = 6.0 Hz, 1H), 8.77 – 8.71 (m, 2H), 7.84 – 7.79 (m, 2H), 7.37 – 7.23 (m, 5H), 4.52 (d, J = 6.0 Hz, 2H). **¹³C NMR (75 MHz, DMSO)** δ 165.17, 150.75, 141.71, 139.57, 128.82, 127.75, 127.37, 121.74, 43.19, 40.80, 40.52, 40.24, 39.97, 39.69, 39.41, 39.13.

2o: N-benzylnicotinamide^[19]



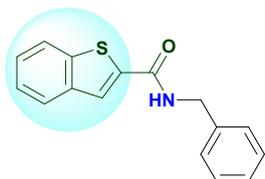
¹H NMR (300 MHz, CDCl₃) δ 8.83 (d, J = 89.3 Hz, 2H), 8.15 (d, J = 7.9 Hz, 1H), 6.97 (s, 1H), 4.65 (d, J = 5.6 Hz, 2H). **¹³C NMR (75 MHz, CDCl₃)** δ 165.52, 152.07, 147.83, 137.79, 135.37, 128.85, 127.97, 127.77, 77.49, 77.06, 76.64, 44.19.

2p: N-benzylthiophene-3-carboxamide^[19]



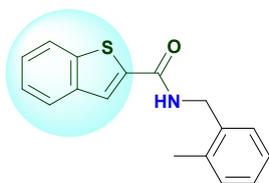
¹H NMR (400 MHz, DMSO) δ 8.88 (t, J = 6.1 Hz, 1H), 8.18 (dd, J = 3.0, 1.3 Hz, 1H), 7.61 – 7.54 (m, 2H), 7.36 – 7.31 (m, 4H), 7.25 (ddd, J = 8.7, 3.7, 2.5 Hz, 1H), 4.46 (d, J = 6.0 Hz, 2H). **¹³C NMR (101 MHz, DMSO)** δ 162.52, 140.14, 138.15, 129.26, 128.76, 128.51, 127.69, 127.30, 127.23, 42.74, 40.60, 40.39, 40.18, 40.08, 39.97, 39.76, 39.55, 39.34.

2q: N-benzylbenzo[b]thiophene-2-carboxamide^[20]



¹H NMR (400 MHz, DMSO) δ 9.35 (t, J = 6.0 Hz, 1H), 8.16 (d, J = 0.8 Hz, 1H), 8.06 – 8.00 (m, 1H), 7.97 – 7.92 (m, 1H), 7.49 – 7.42 (m, 2H), 7.38 – 7.32 (m, 4H), 7.29 – 7.23 (m, 1H), 4.52 (d, J = 6.0 Hz, 2H). **¹³C NMR (101 MHz, DMSO)** δ 162.03, 140.70, 140.33, 139.71, 139.66, 128.84, 128.59, 128.51, 127.83, 127.39, 126.89, 126.69, 125.67, 125.39, 125.34, 123.27, 43.19, 40.58, 40.37, 40.17, 40.09, 39.96, 39.75, 39.54, 39.33.

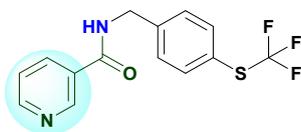
2r: N-(2-methylbenzyl) benzo[b]thiophene-2-carboxamide



¹H NMR (300 MHz, DMSO) δ 9.19 (t, J = 5.7 Hz, 1H), 8.19 (d, J = 0.8 Hz, 1H), 8.09 – 7.99 (m, 1H), 7.97 – 7.91 (m, 1H), 7.51 – 7.41 (m, 2H), 7.33 – 7.26 (m, 1H), 7.21 – 7.15 (m, 3H), 4.49 (d, J = 5.7 Hz, 2H), 2.34 (s, 3H). **¹³C NMR (75 MHz, DMSO)** δ 161.94, 140.68, 140.34, 139.69, 137.20, 136.15, 130.45, 128.14, 127.45, 126.66, 126.26, 125.65, 125.37, 123.27, 41.33, 40.82, 40.54, 40.26, 39.98, 39.71, 39.43, 39.15, 19.21.

HRMS (ESI-TOF): m/z calcd. for C₁₇H₁₅NOS+H⁺ ([M+H⁺]) 282.0952, found 282.0952.

2s: N-(4-((trifluoromethyl)thio)benzyl)nicotinamide

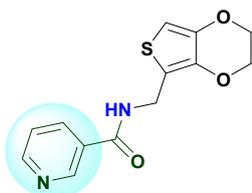


¹H NMR (300 MHz, CDCl₃) δ 9.07 (d, J = 2.3 Hz, 1H), 8.73 (dd, J = 4.9, 1.7 Hz, 1H), 8.21 (ddd, J = 7.9, 2.3, 1.7 Hz, 1H), 7.68 – 7.63 (m, 2H), 7.46 – 7.41 (m, 3H), 6.97 (s, 1H), 4.77 – 4.68 (m, 2H).

¹³C NMR (75 MHz, CDCl₃) δ 165.47, 152.08, 147.60, 141.02, 136.78, 136.53, 135.74, 129.98, 128.86, 127.65, 123.78, 77.46, 77.03, 76.61, 43.56.

HRMS (ESI-TOF): m/z calcd. for C₁₄H₁₁N₂OSF₃+H⁺ ([M+H⁺]) 313.0622, found 313.0622.

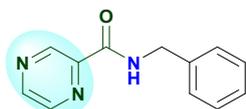
2t: N-((2,3-dihydrothieno[3,4-b][1,4]dioxin-5-yl)methyl)nicotinamide



¹H NMR (300 MHz, CDCl₃) δ 9.02 (ddd, J = 16.1, 2.3, 0.9 Hz, 1H), 8.76 (ddd, J = 12.2, 4.9, 1.7 Hz, 1H), 8.20 – 8.12 (m, 1H), 7.45 – 7.38 (m, 1H), 6.59 (d, J = 4.9 Hz, 1H), 6.29 (s, 1H), 4.68 (d, J = 5.4 Hz, 2H), 4.28 – 4.20 (m, 4H). **¹³C NMR (75 MHz, CDCl₃)** δ 152.29, 147.83, 135.32, 123.57, 98.30, 77.45, 77.03, 76.61, 64.82, 64.60, 35.44.

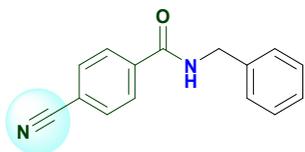
HRMS (ESI-TOF): m/z calcd. for C₁₃H₁₂N₂O₃S+H⁺ ([M+H⁺]) 277.0647, found 277.0654.

2u: N-benzylpyrazine-2-carboxamide^[21]



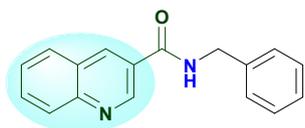
¹H NMR (300 MHz, DMSO) δ 9.49 (s, 1H), 9.21 (dd, J = 1.5, 0.3 Hz, 1H), 8.88 (dd, J = 2.5, 0.3 Hz, 1H), 8.75 (dd, J = 2.5, 1.5 Hz, 1H), 7.35 – 7.21 (m, 5H), 4.52 (d, J = 6.4 Hz, 2H). **¹³C NMR (75 MHz, DMSO)** δ 148.02, 144.07, 139.71, 128.75, 127.84, 127.30, 42.85, 40.83, 40.55, 40.27, 40.00, 39.72, 39.44, 39.16.

2v: N-benzyl-4-cyanobenzamide^[22]



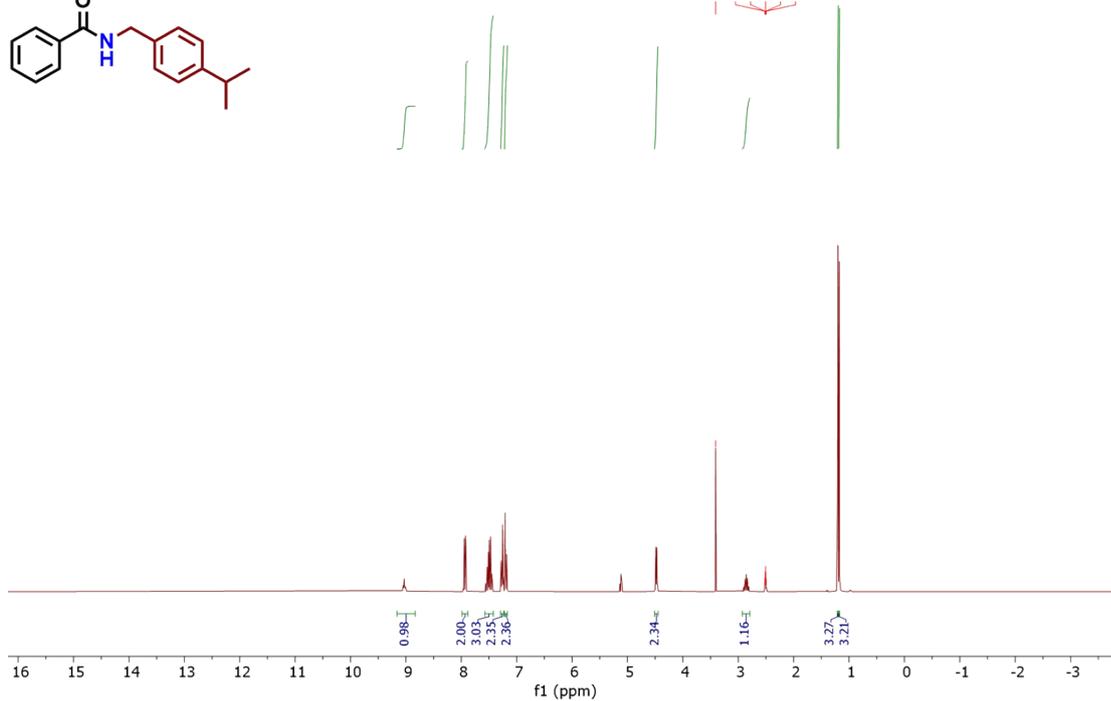
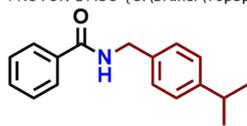
¹H NMR (300 MHz, DMSO) δ 9.30 (t, J = 6.0 Hz, 1H), 8.08 – 8.02 (m, 2H), 8.01 – 7.94 (m, 2H), 7.35 – 7.25 (m, 5H), 4.50 (dd, J = 5.8, 2.1 Hz, 2H). **¹³C NMR (75 MHz, DMSO)** δ 165.32, 139.66, 138.77, 132.95, 128.81, 128.59, 127.74, 127.34, 118.82, 114.12, 43.27, 40.83, 40.55, 40.27, 39.99, 39.72, 39.44, 39.16.

2w: N-benzylquinoline-3-carboxamide^[23]

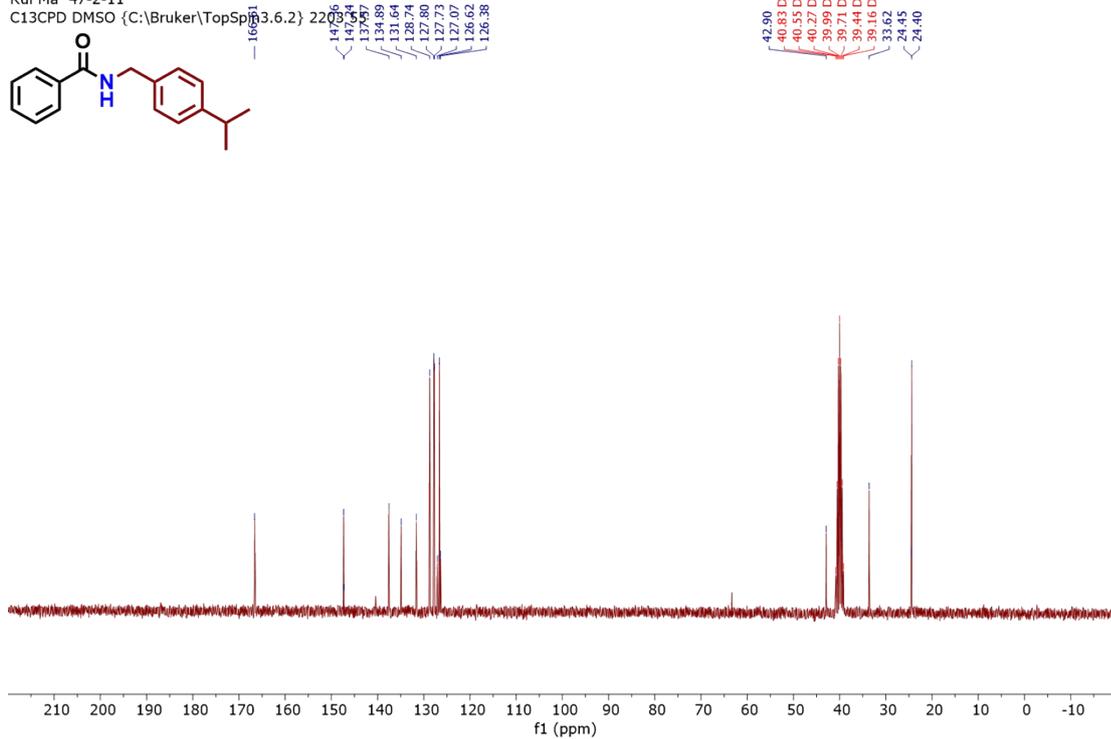
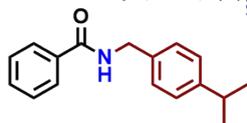


¹H NMR (300 MHz, CDCl₃) δ 8.76 (dd, J = 4.3, 1.7 Hz, 1H), 8.19 – 8.13 (m, 1H), 7.98 – 7.85 (m, 3H), 7.41 (d, J = 5.5 Hz, 1H), 7.28 – 7.09 (m, 6H), 4.52 (d, J = 5.7 Hz, 2H). **¹³C NMR (75 MHz, CDCl₃)** δ 167.01, 151.84, 149.18, 138.19, 136.98, 132.35, 129.71, 128.71, 127.81, 127.54, 127.48, 127.35, 121.82, 77.58, 44.19.

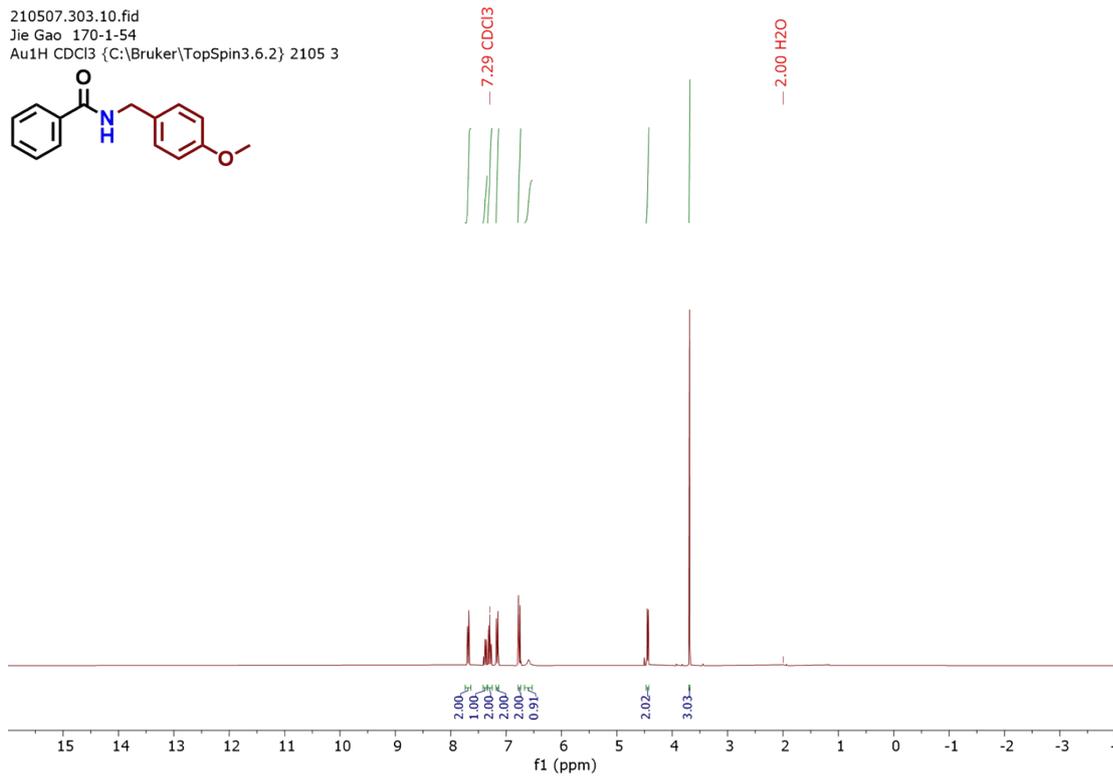
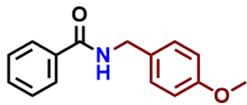
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Rui Ma 47-2-11
PROTON DMSO (C:\Bruker\TopSpin3.6.2) 2203 55



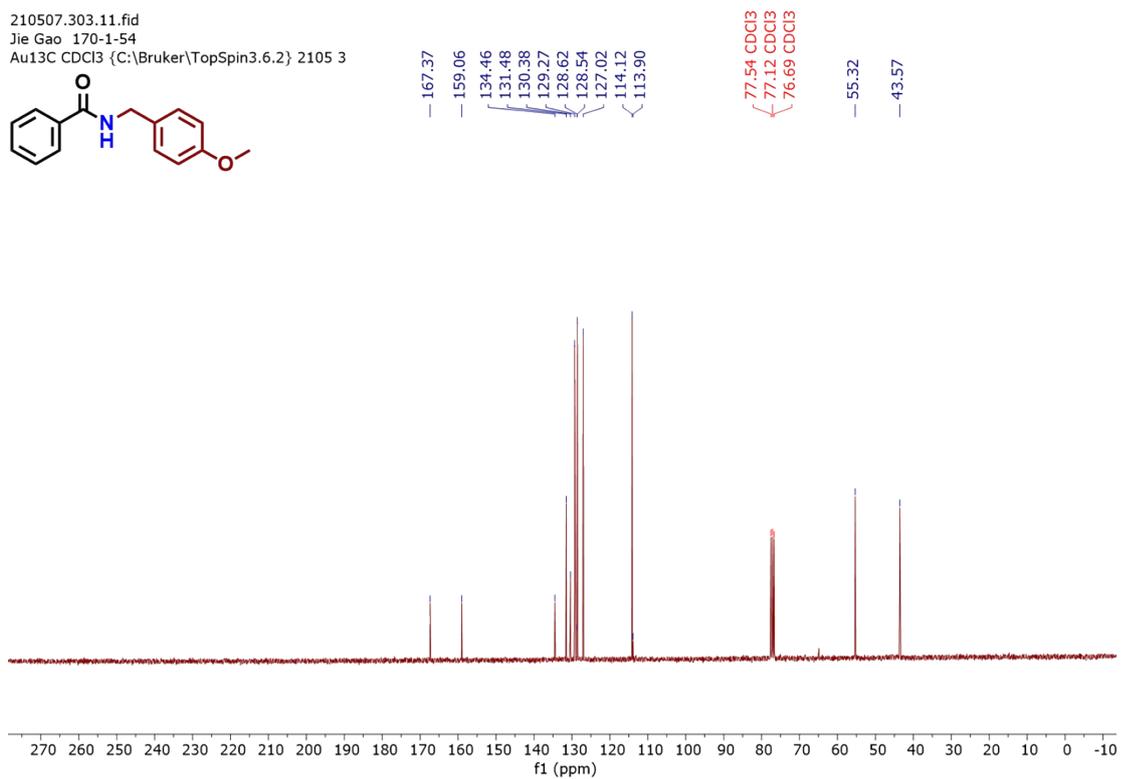
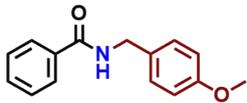
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C13CPD DMSO (C:\Bruker\TopSpin3.6.2) 2203 55



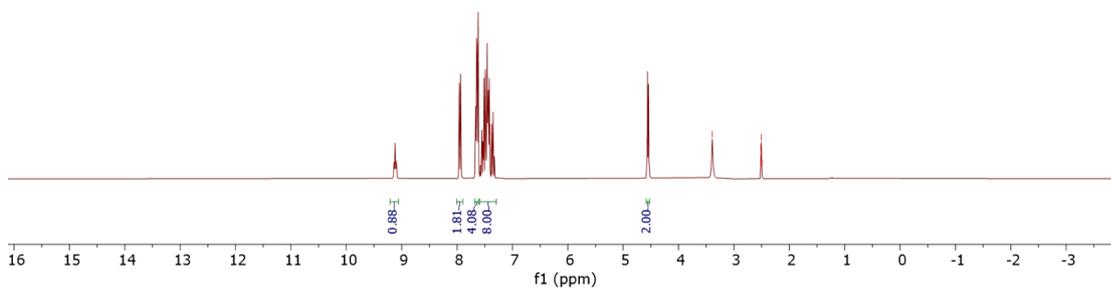
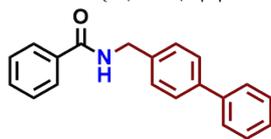
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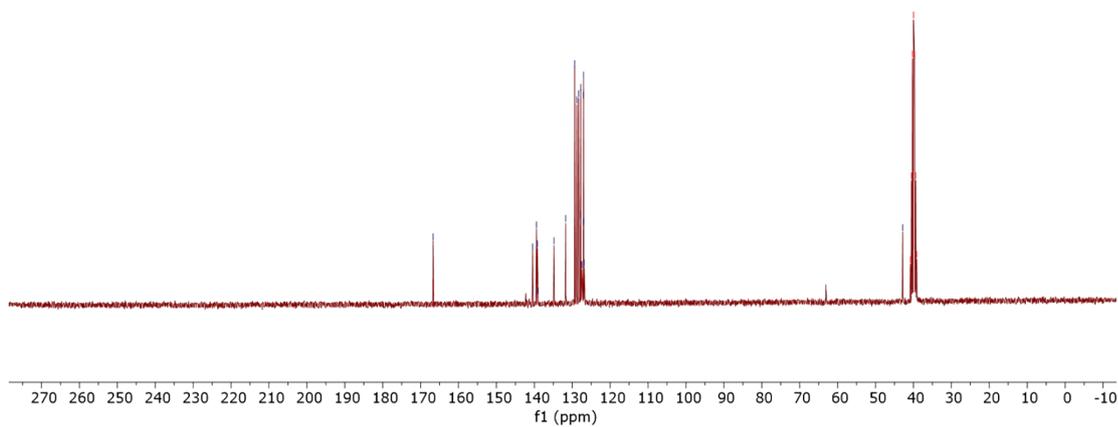
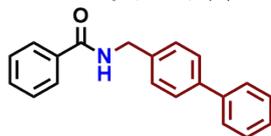
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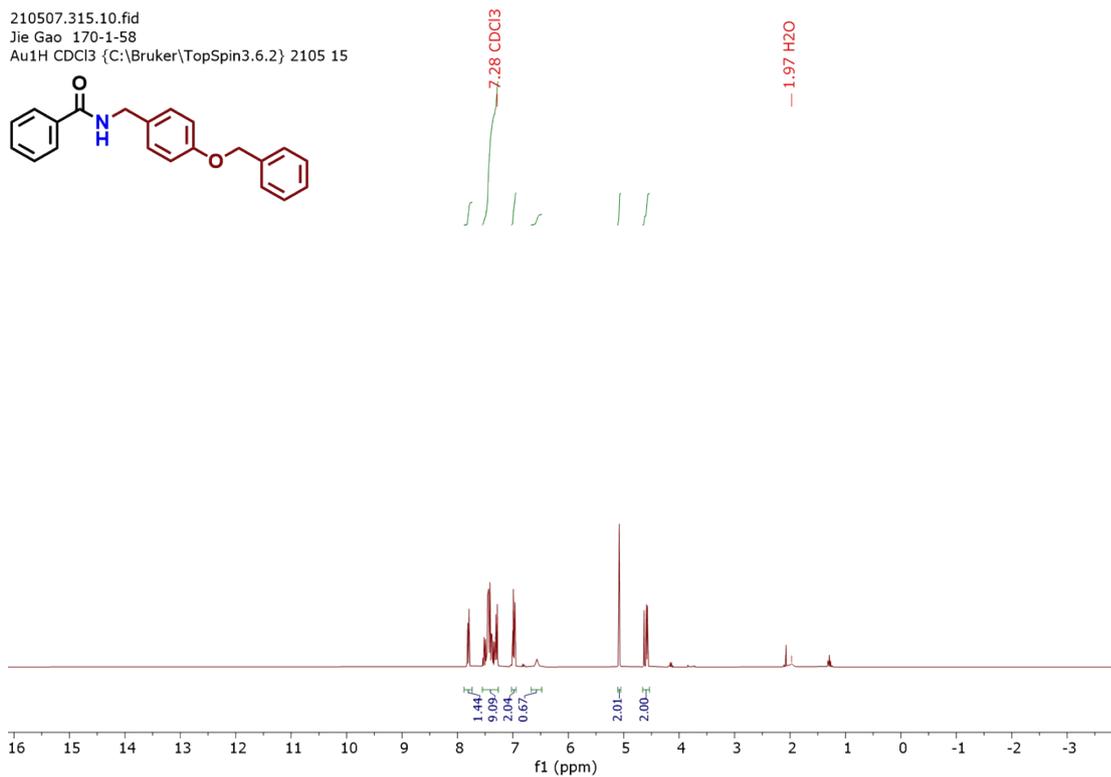
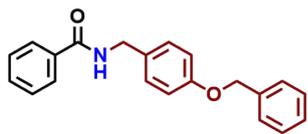
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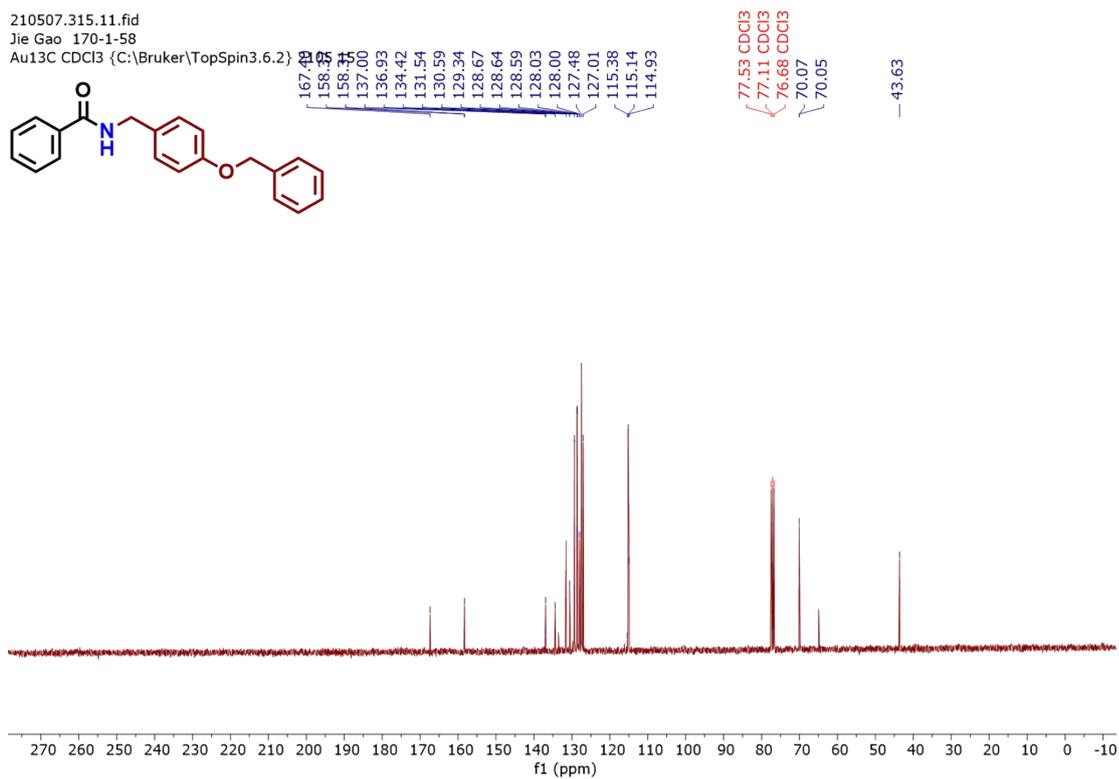
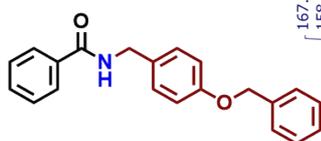
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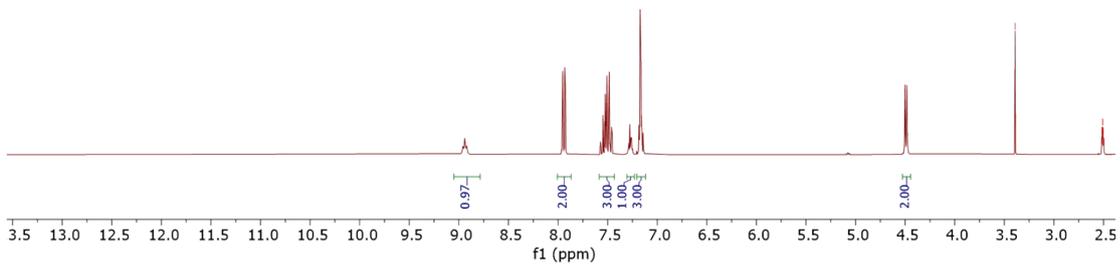
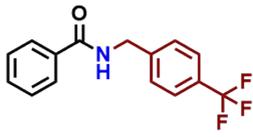
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Au1H CDCl3 (C:\Bruker\TopSpin3.6.2) 2105 15



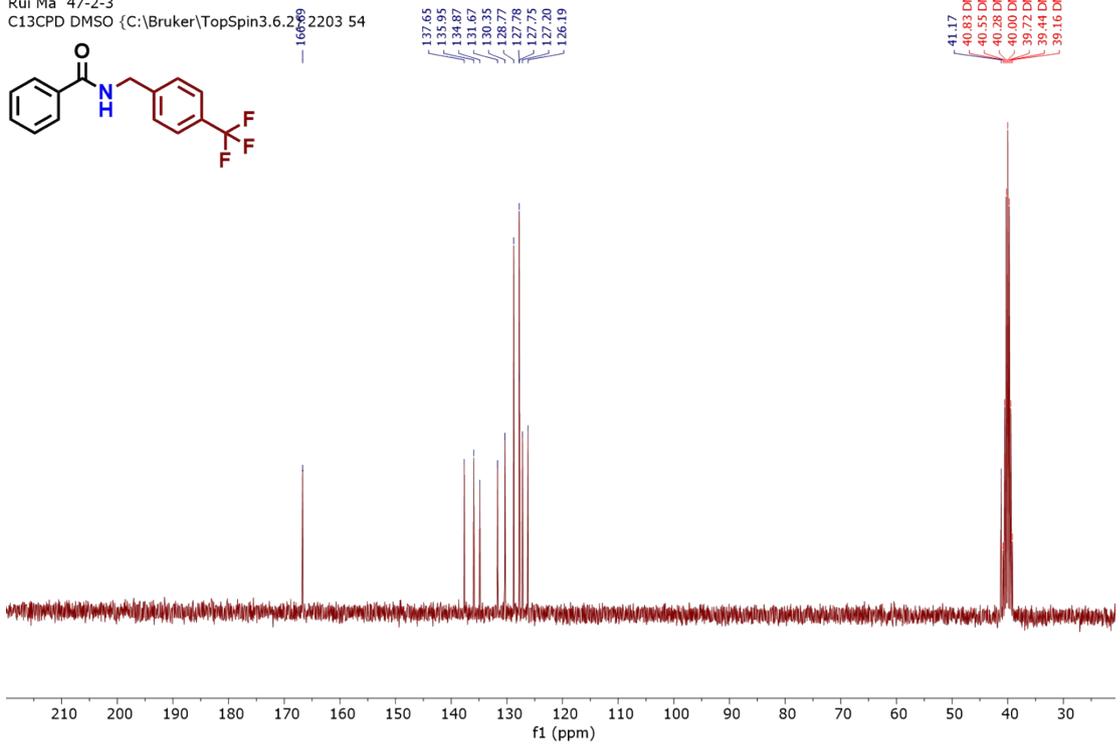
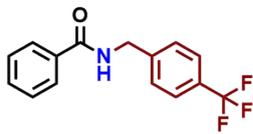
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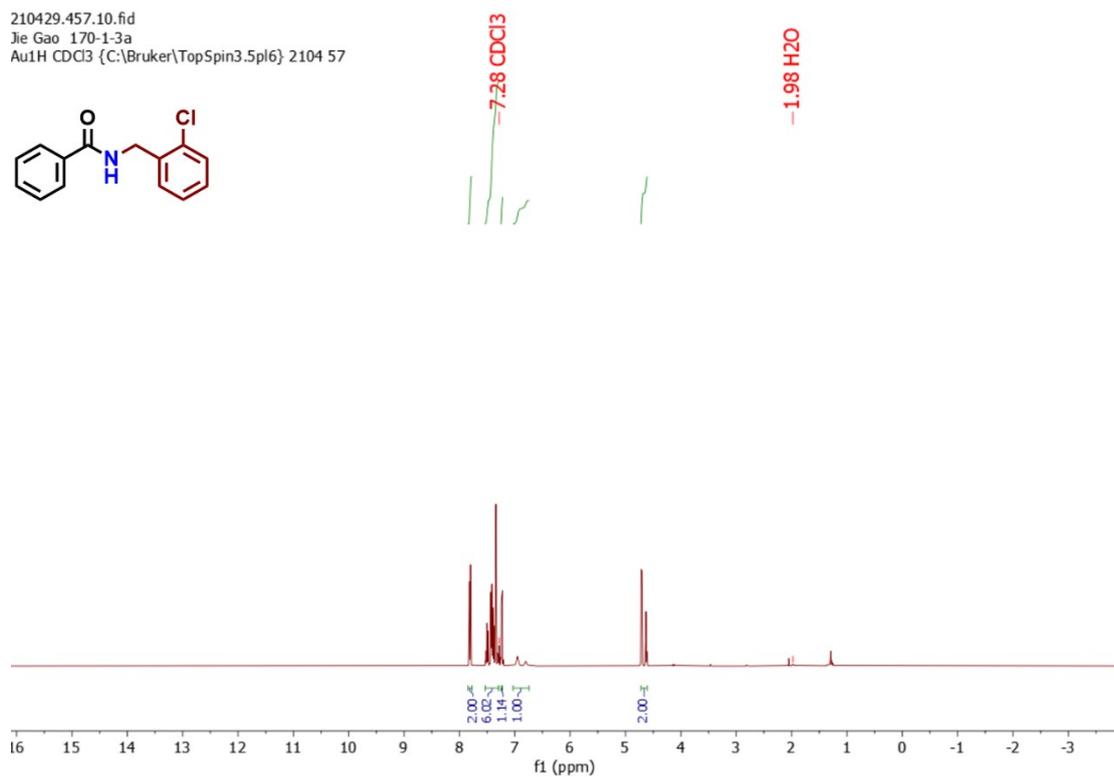
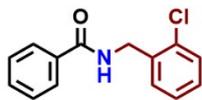
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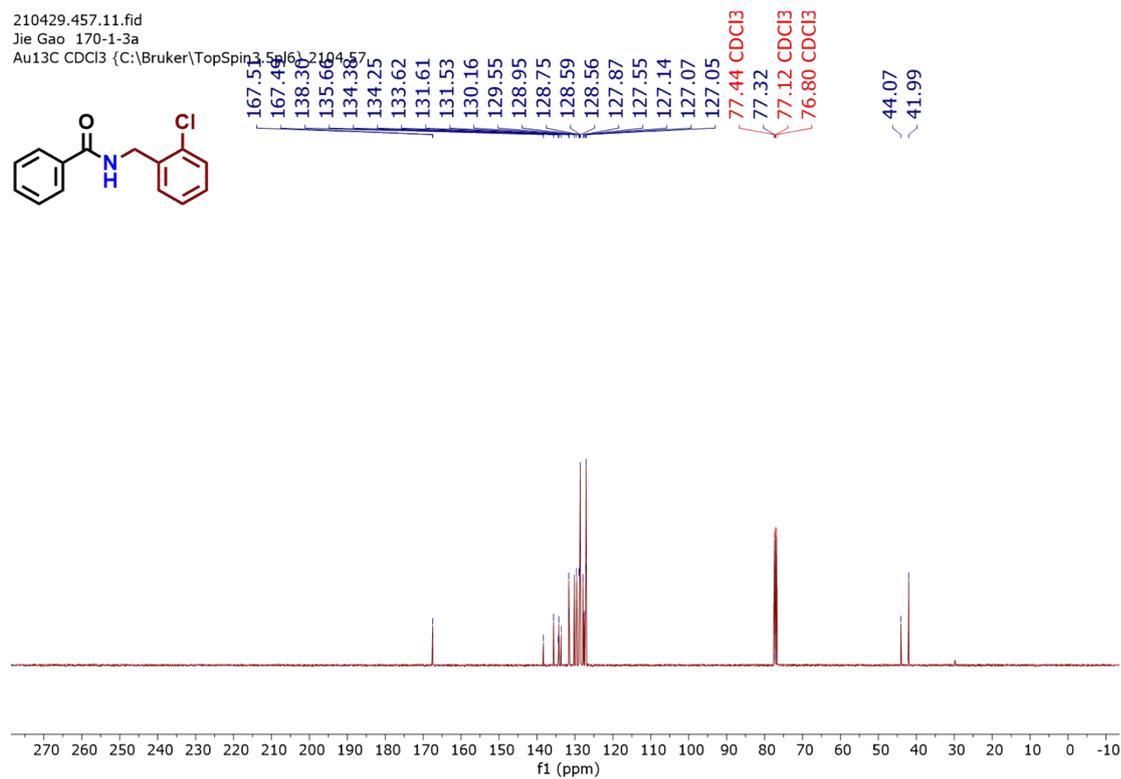
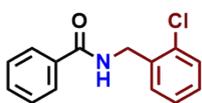
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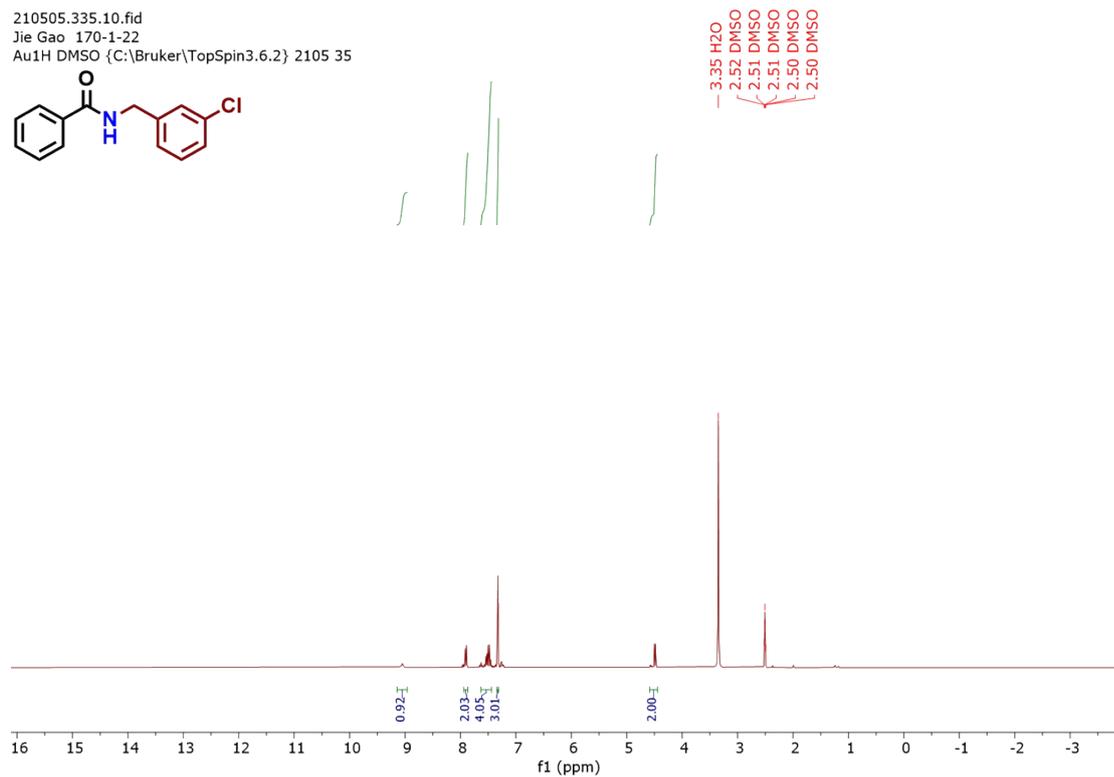
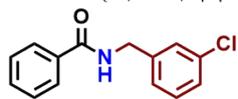
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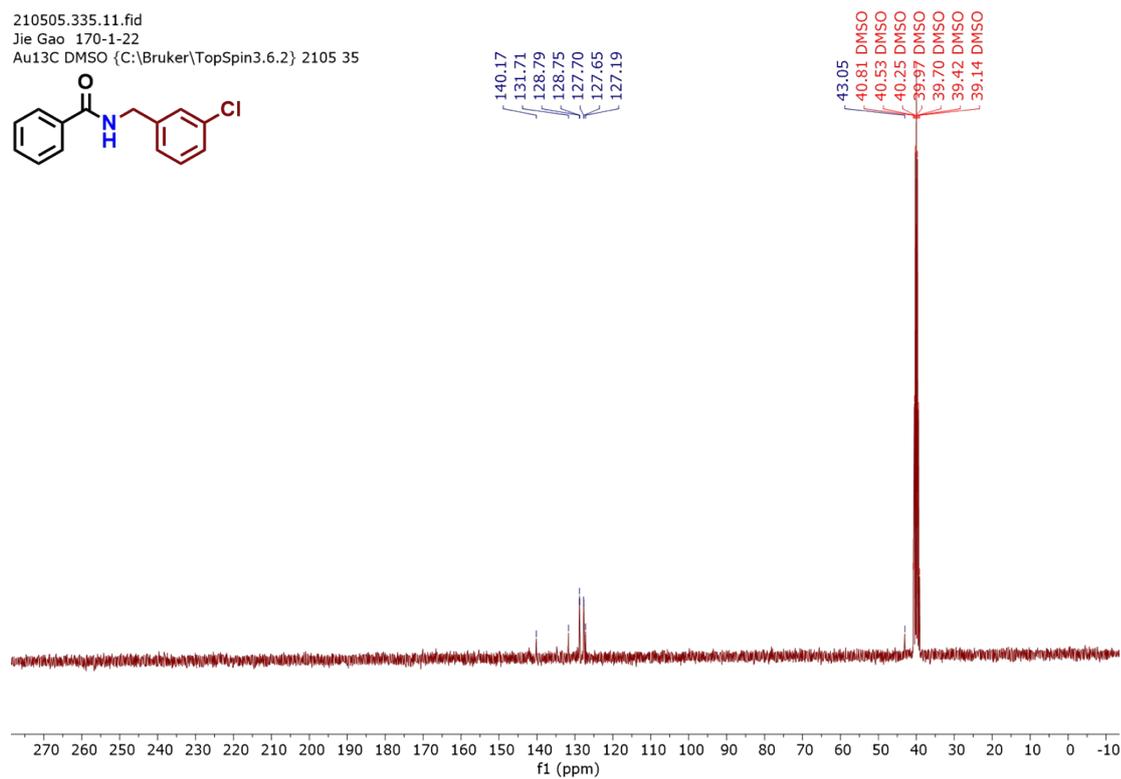
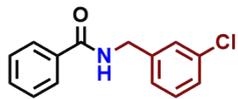
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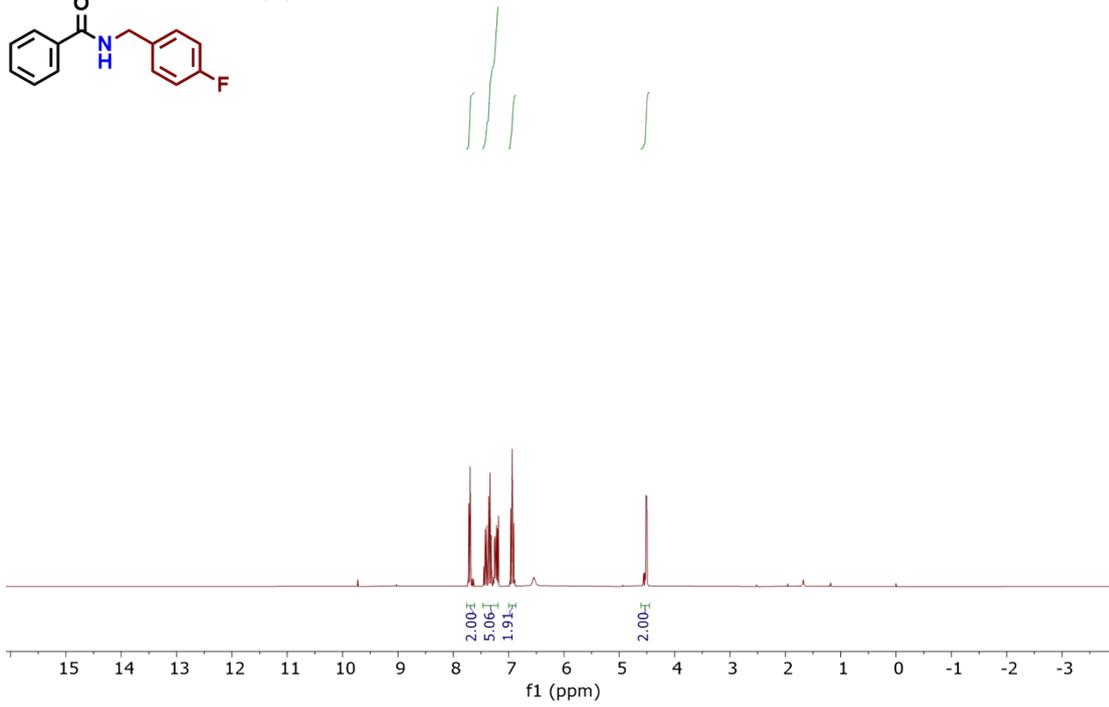
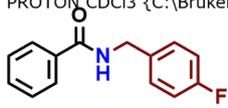
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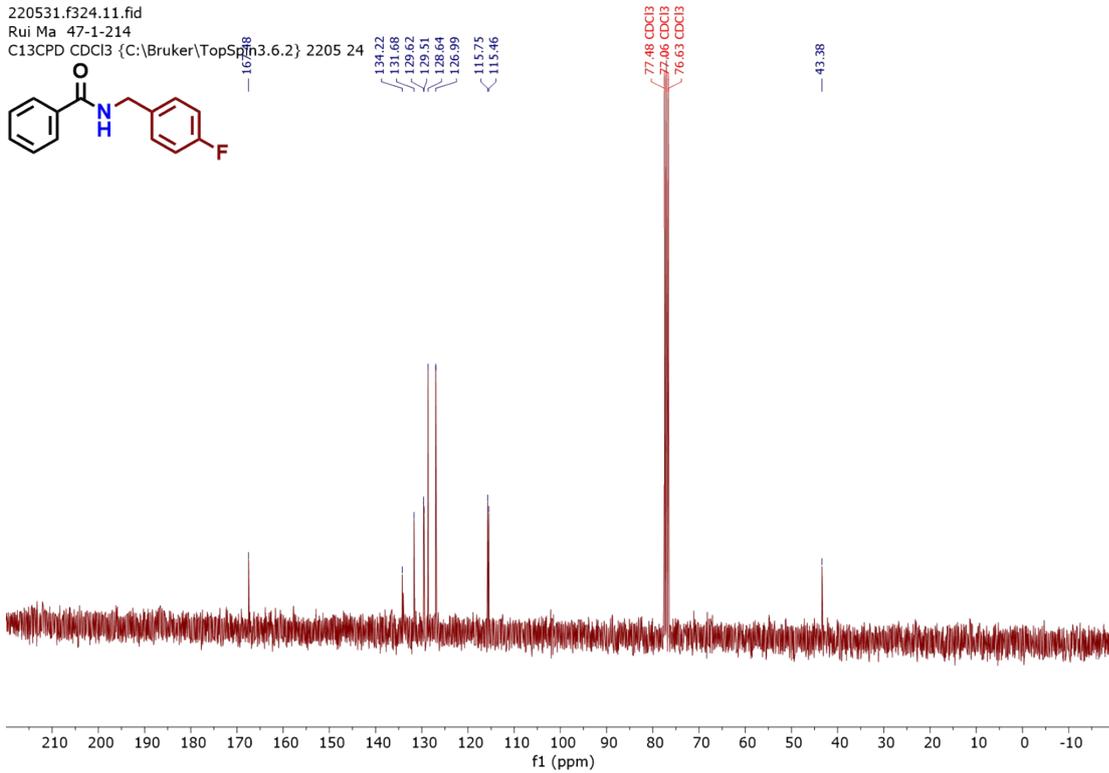
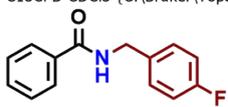
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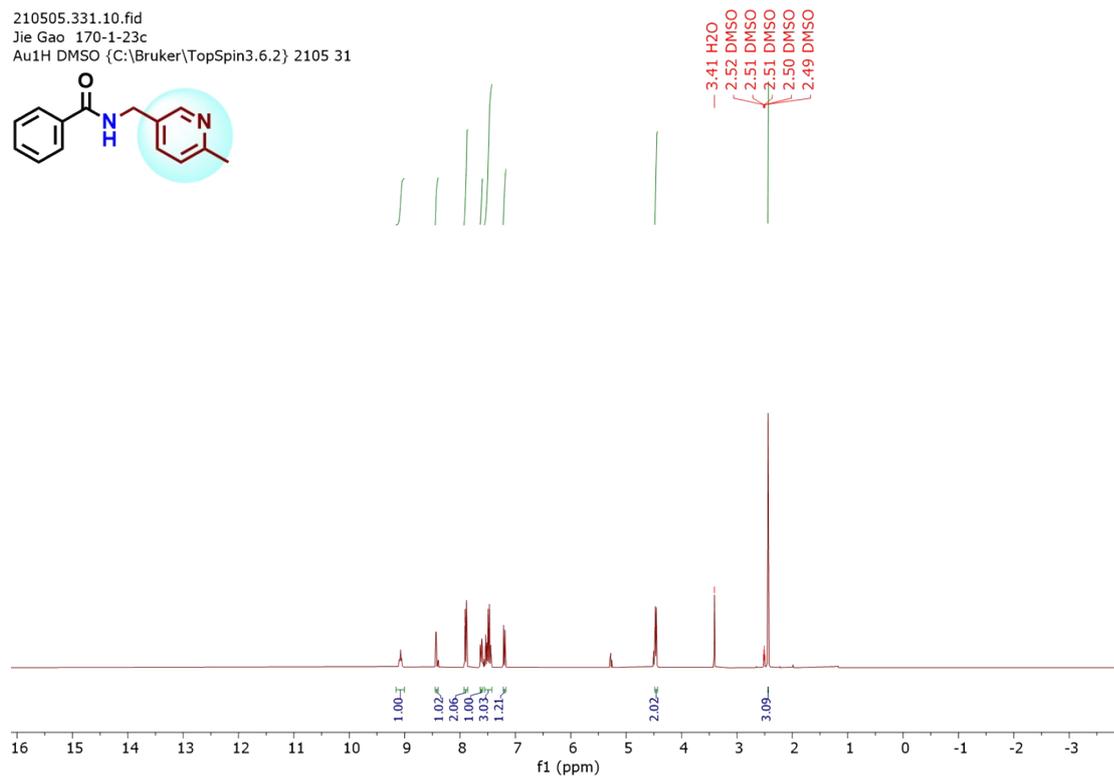
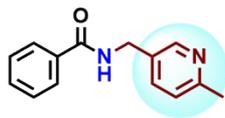
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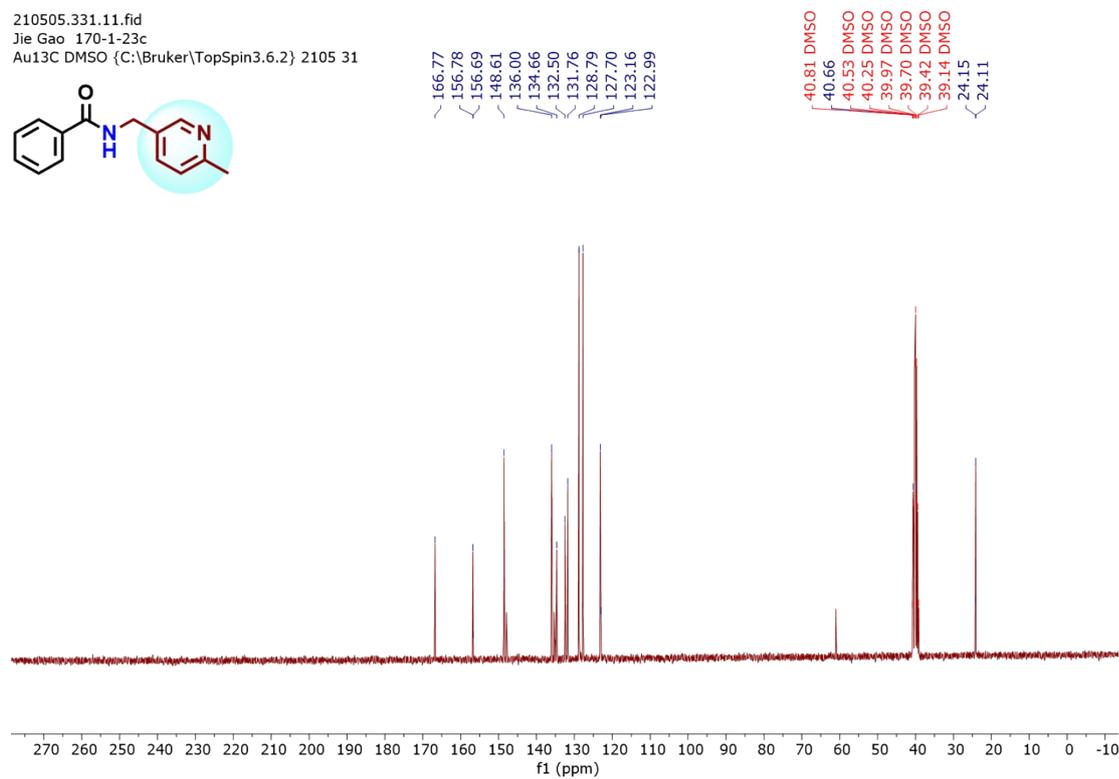
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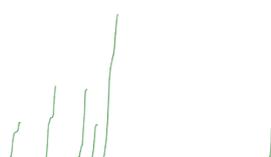
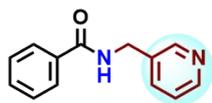
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Au1H DMSO {C:\Bruker\TopSpin3.6.2} 2105 31



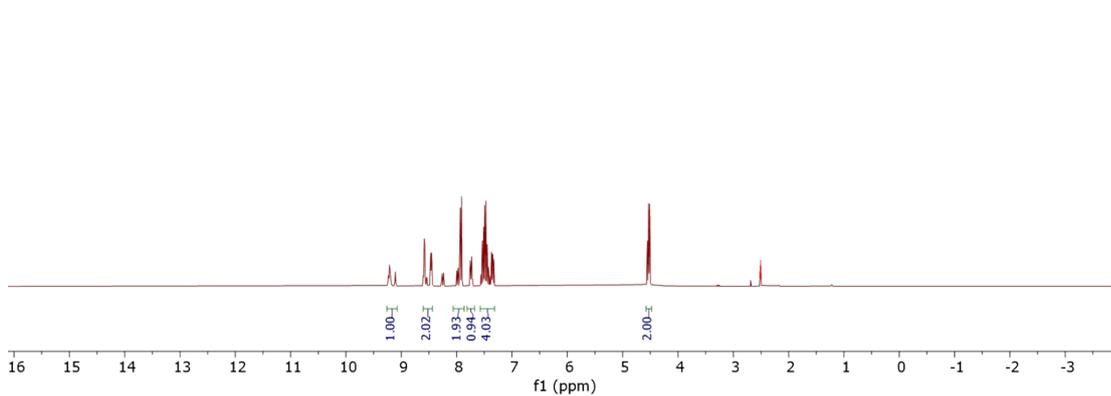
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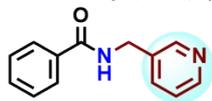
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2.51 DMSO
2.51 DMSO
2.50 DMSO
2.49 DMSO

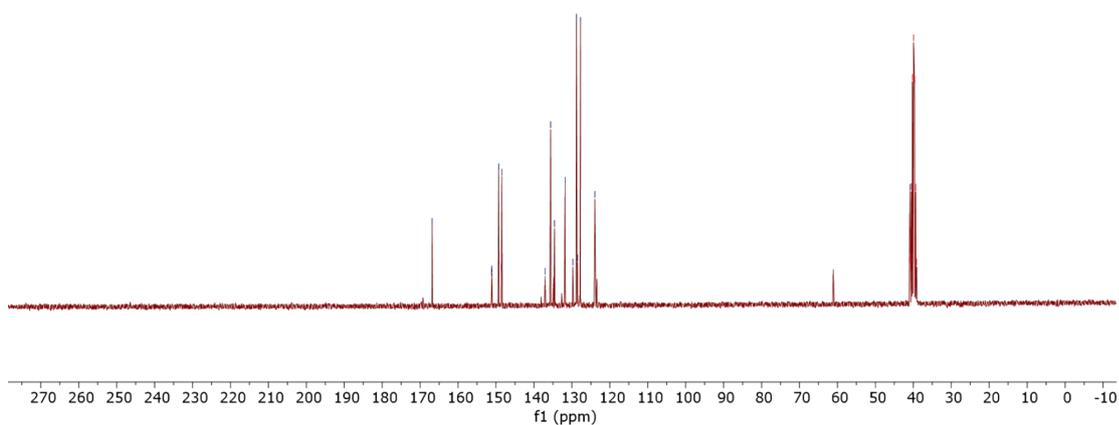


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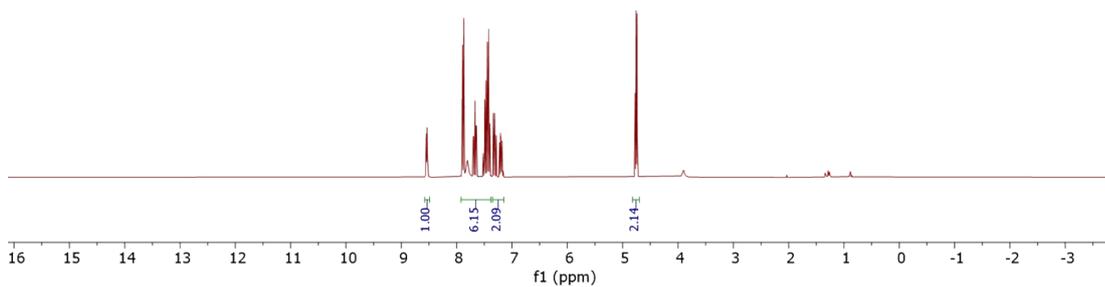
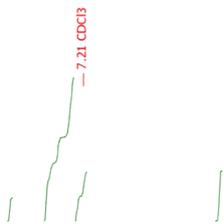
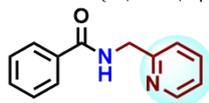


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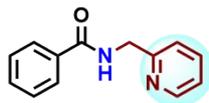
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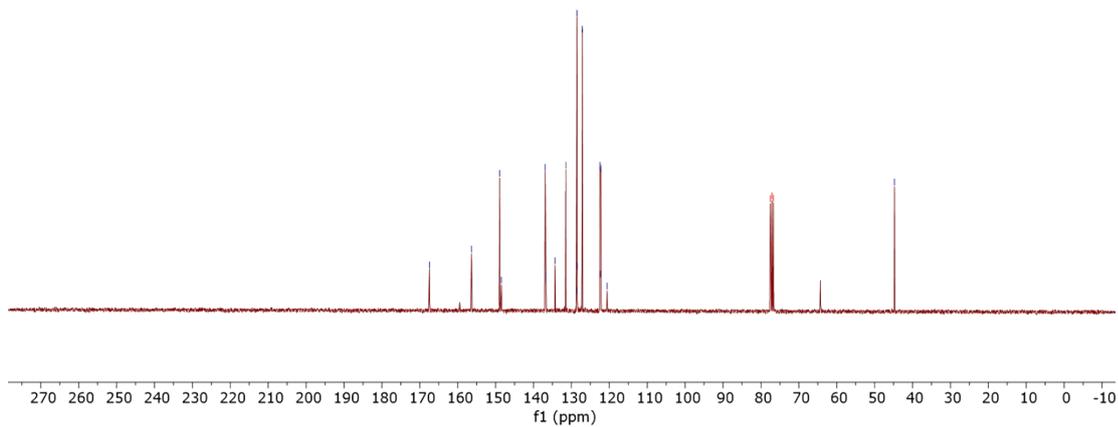
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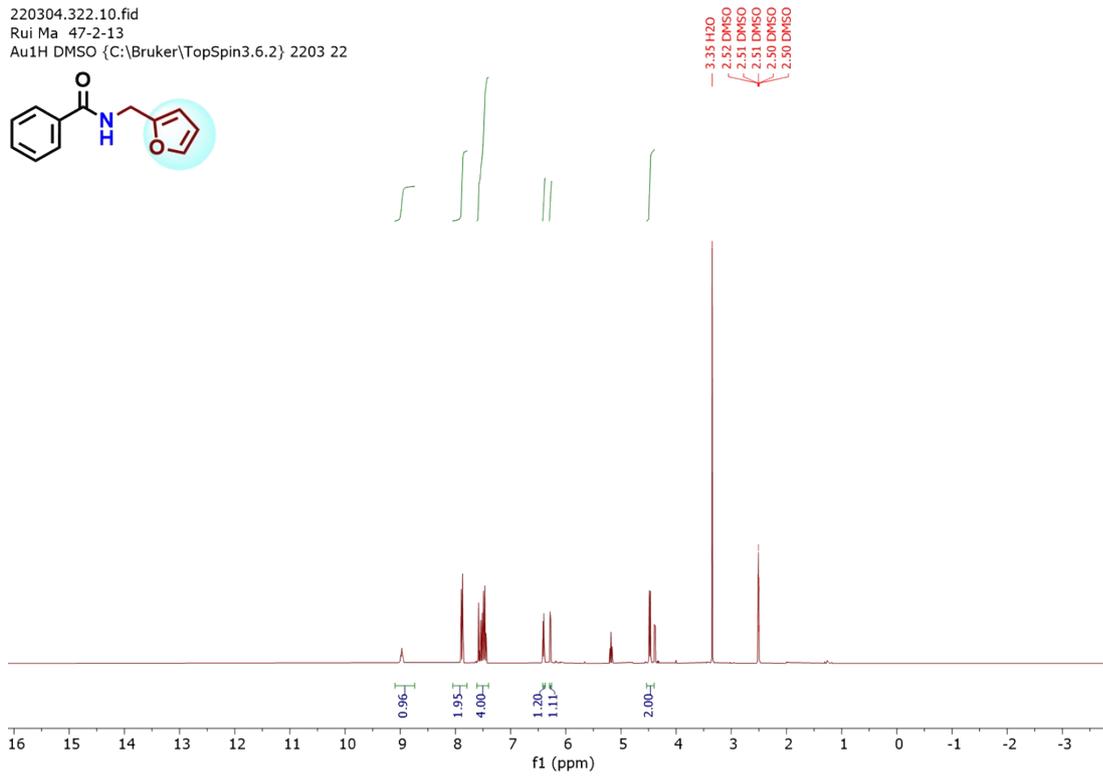
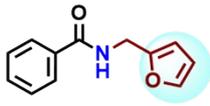
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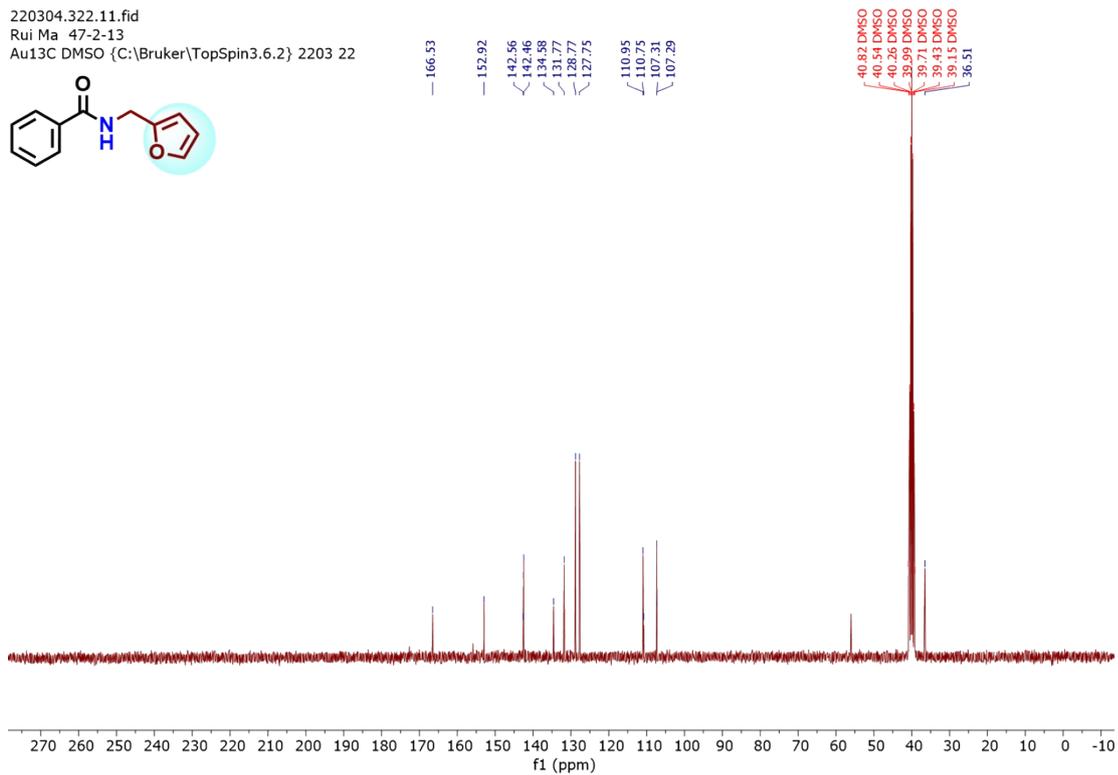
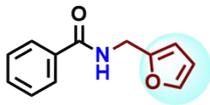
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77.11 CDCl3
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44.76



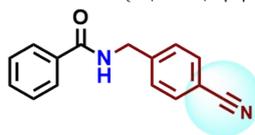
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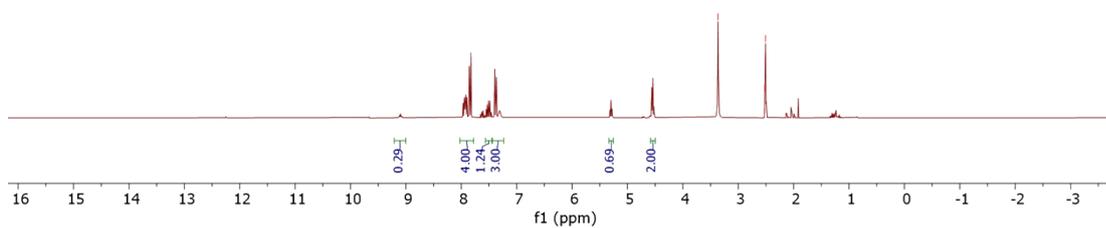
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Au13C DMSO {C:\Bruker\TopSpin3.6.2} 2203 22



220531.f321.10.fid
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PROTON DMSO {C:\Bruker\TopSpin3.6.2} 2205 21



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2.51 DMSO

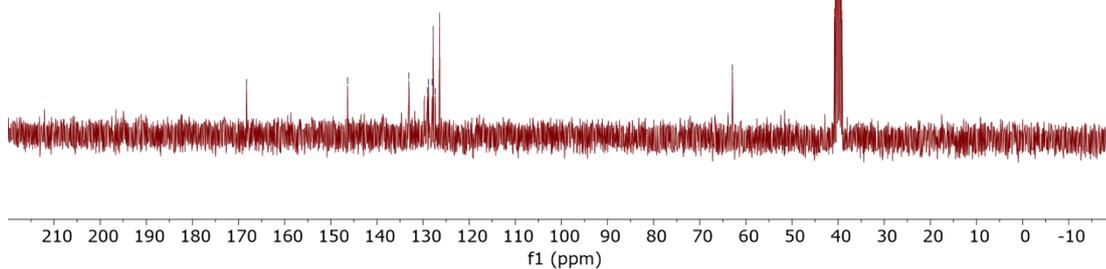


220531.f321.11.fid
Rui Ma 47-1-216b

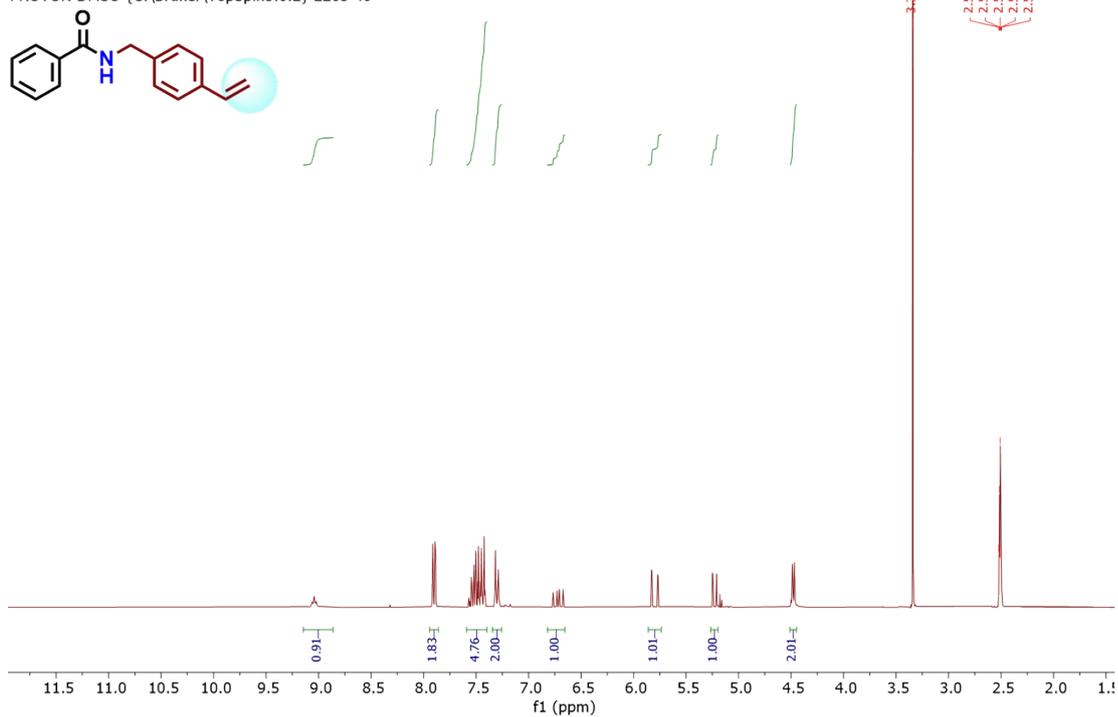


166.39
146.27
133.99
129.94
128.83
128.81
127.81
127.35
126.40

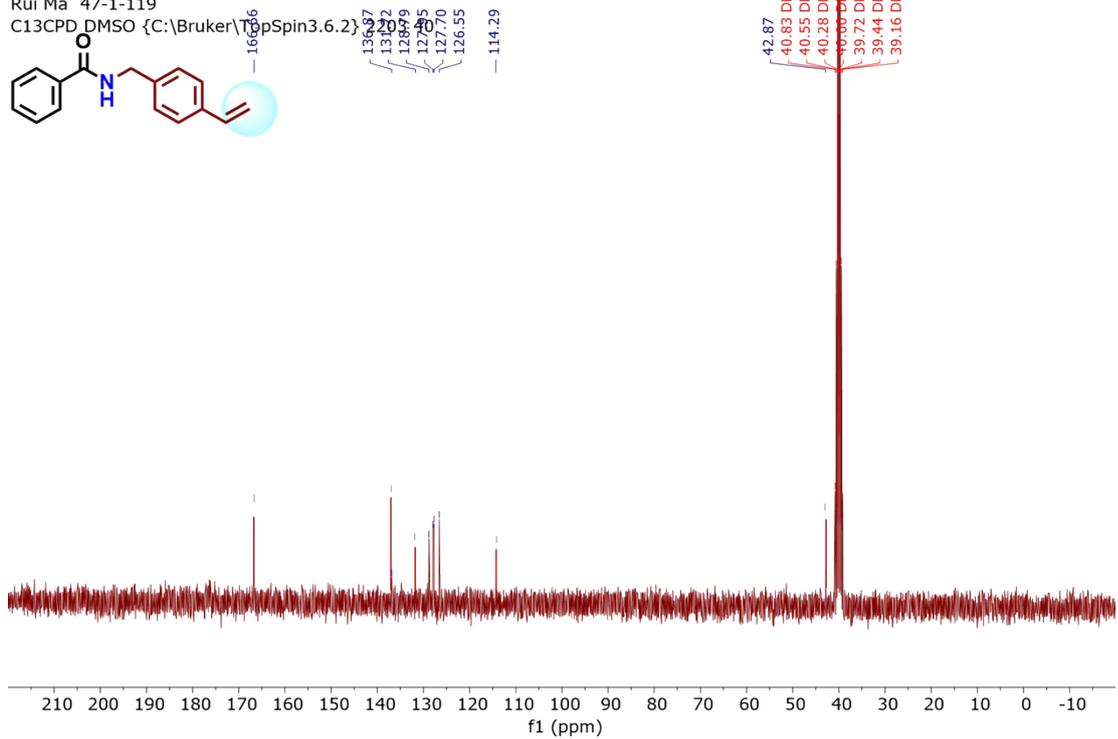
62.94
40.80 DMSO
40.53 DMSO
40.25 DMSO
39.69 DMSO
39.41 DMSO
39.13 DMSO



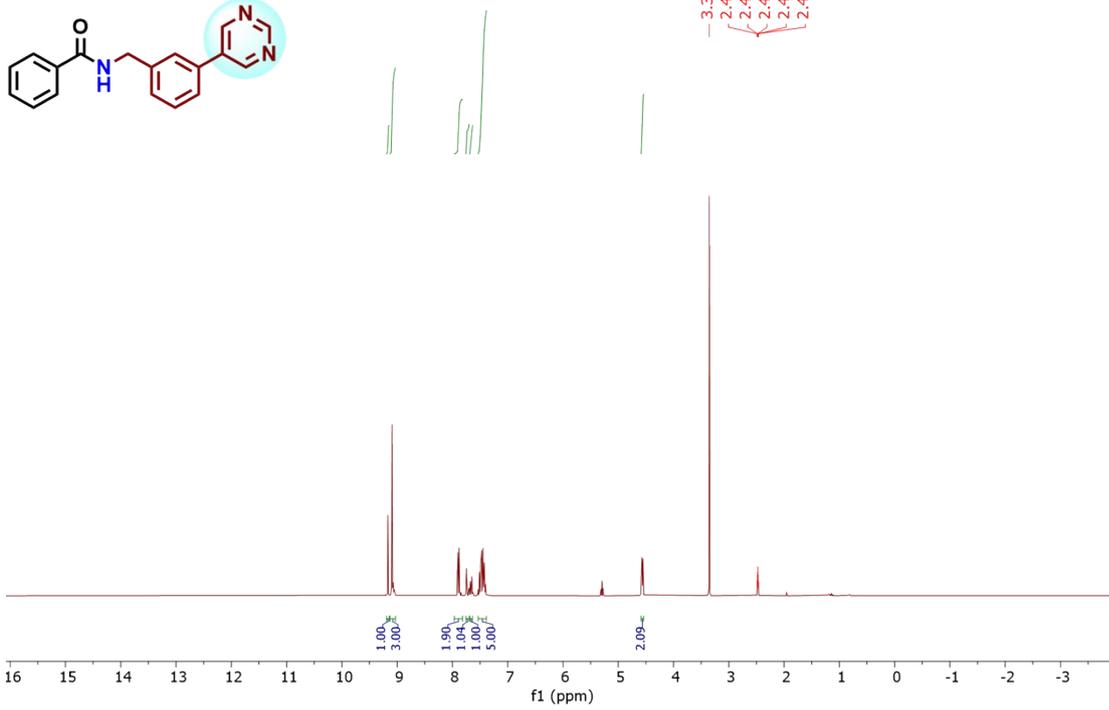
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Rui Ma 47-1-119
PROTON DMSO {C:\Bruker\TopSpin3.6.2} 2203 40



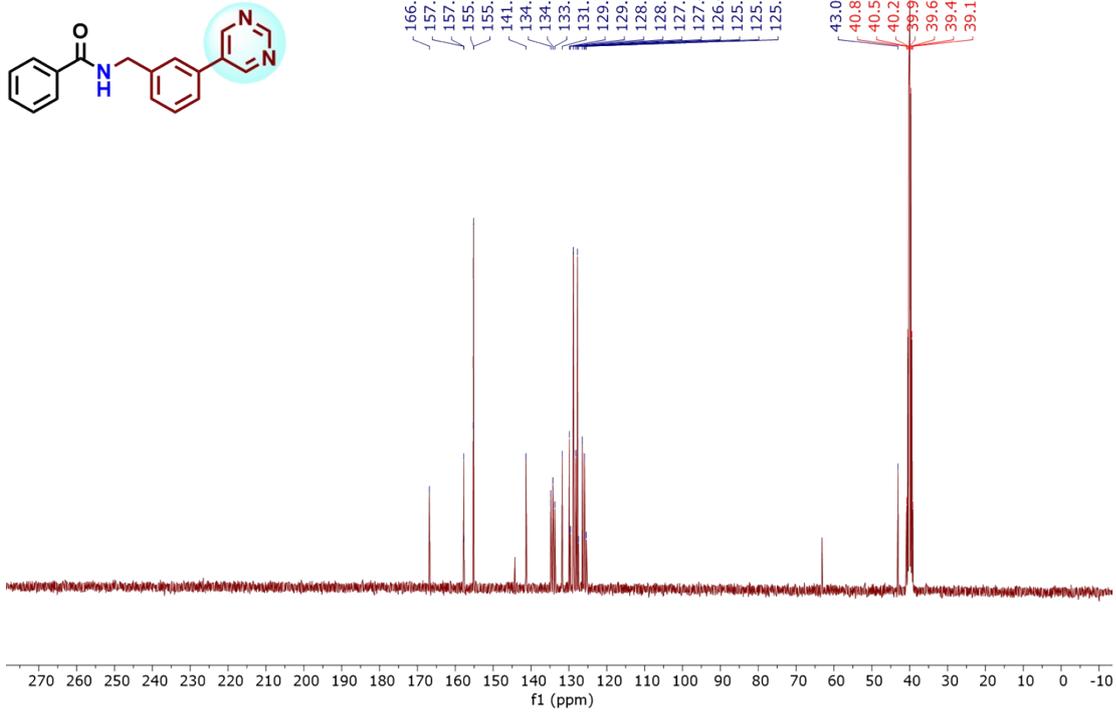
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C13CPD DMSO {C:\Bruker\TopSpin3.6.2}



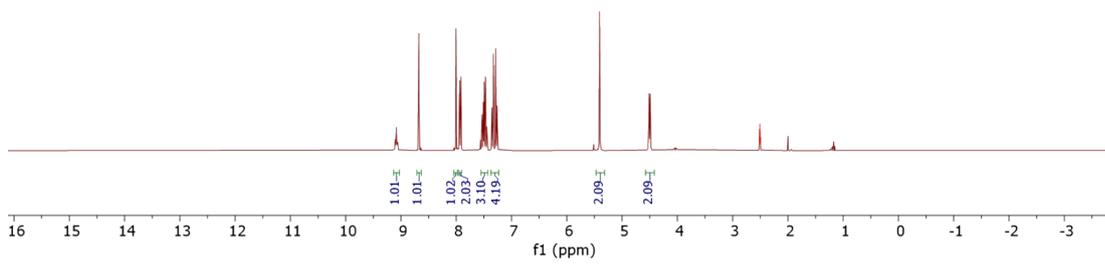
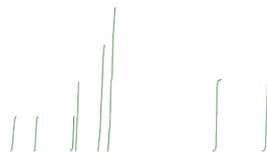
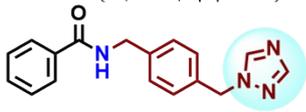
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Jie Gao 170-1-68b
Au1H DMSO {C:\Bruker\TopSpin3.6.2} 2105 10



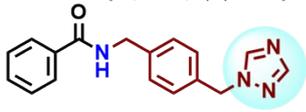
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Jie Gao 170-1-68b
Au13C DMSO {C:\Bruker\TopSpin3.6.2} 2105 10



220518.316.10.fid
Rui Ma 47-1-203b
Au1H DMSO {C:\Bruker\TopSpin3.6.2} 2205 16

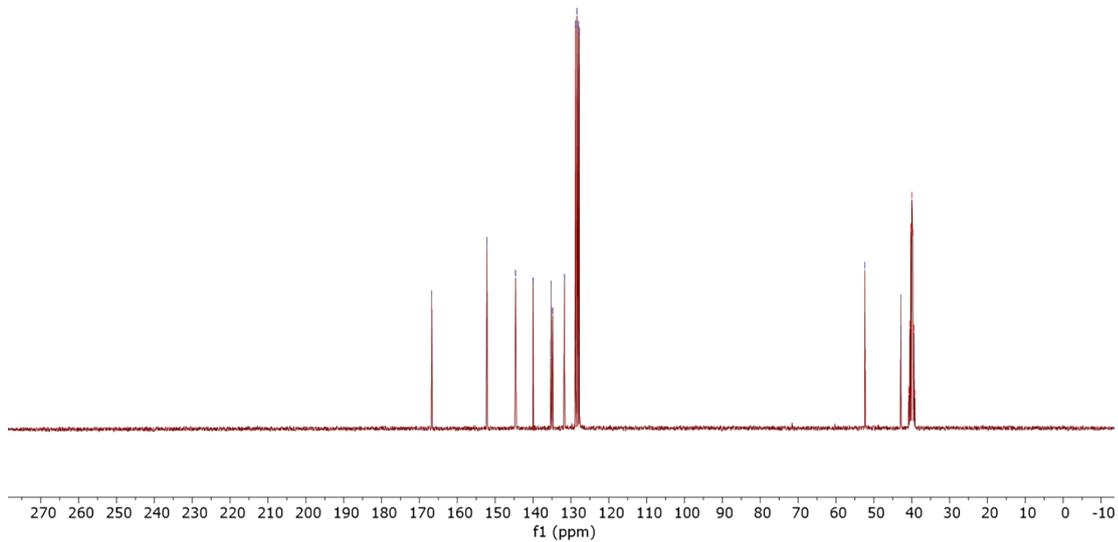


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Au13C DMSO {C:\Bruker\TopSpin3.6.2} 2205 16

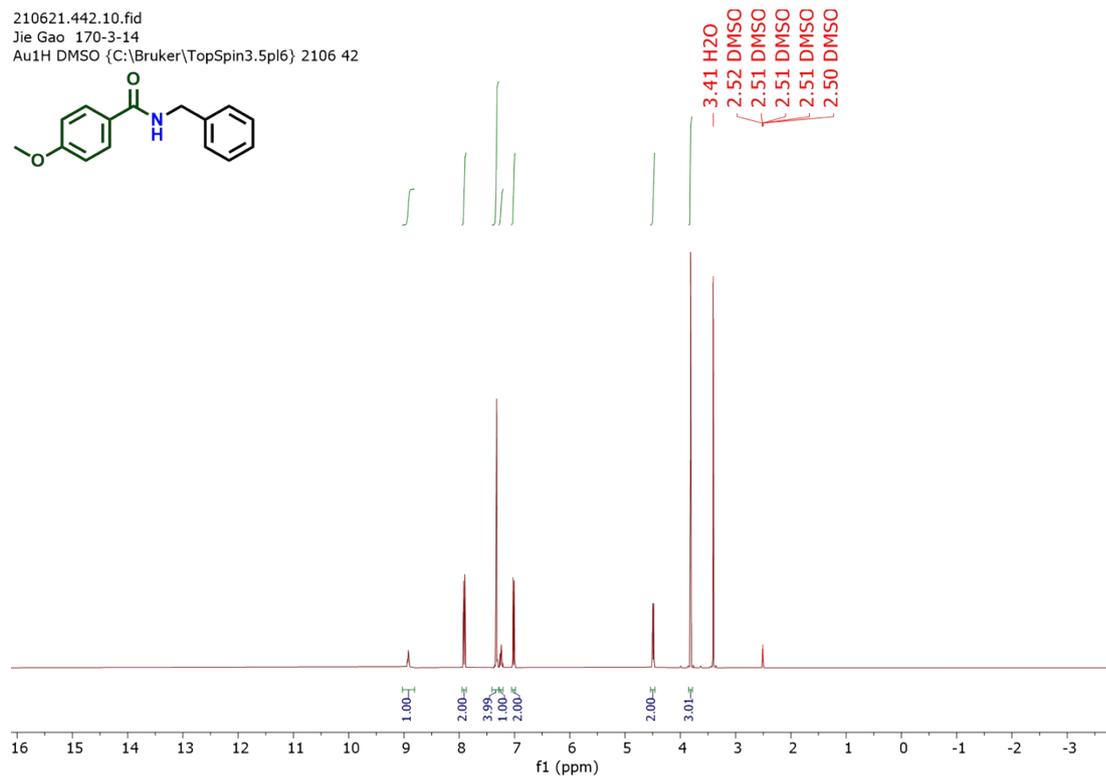
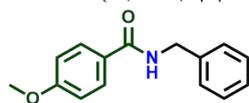


166.76
152.18
144.60
139.99
135.23
134.80
131.72
129.76
128.44
128.01
127.74

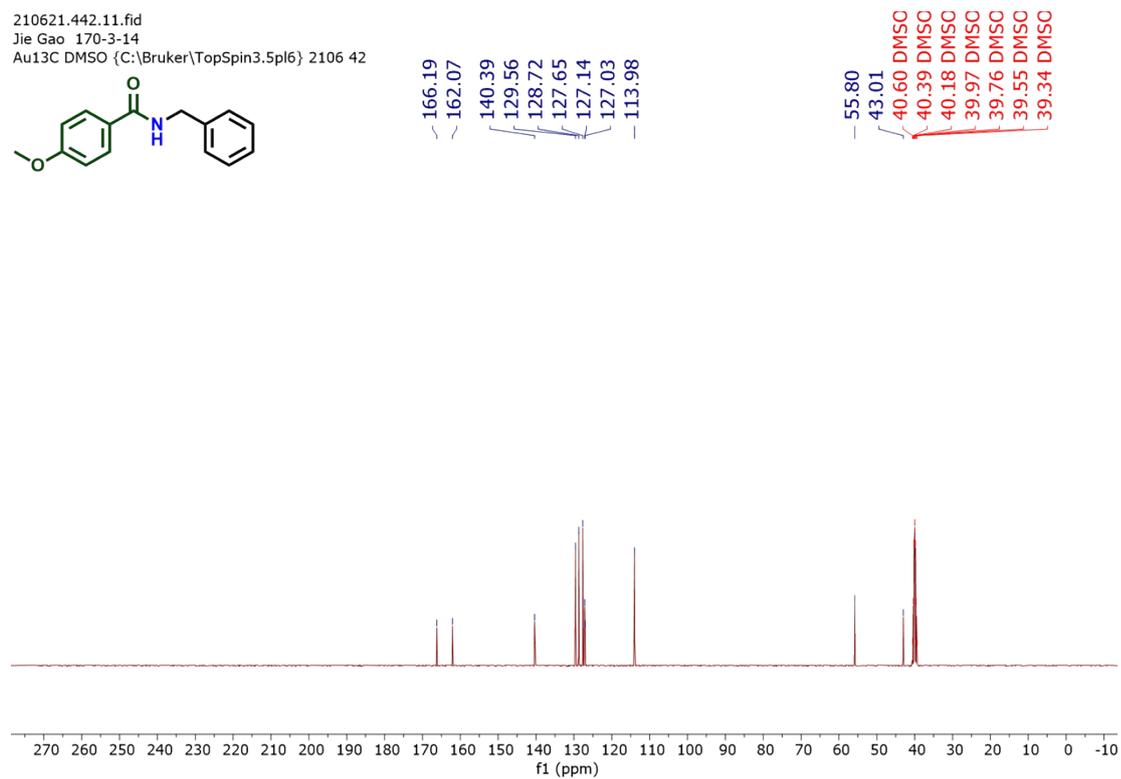
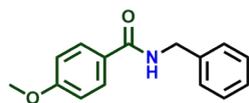
52.37
42.87
40.80 DMSO
40.53 DMSO
40.25 DMSO
39.97 DMSO
39.41 DMSO
39.13 DMSO



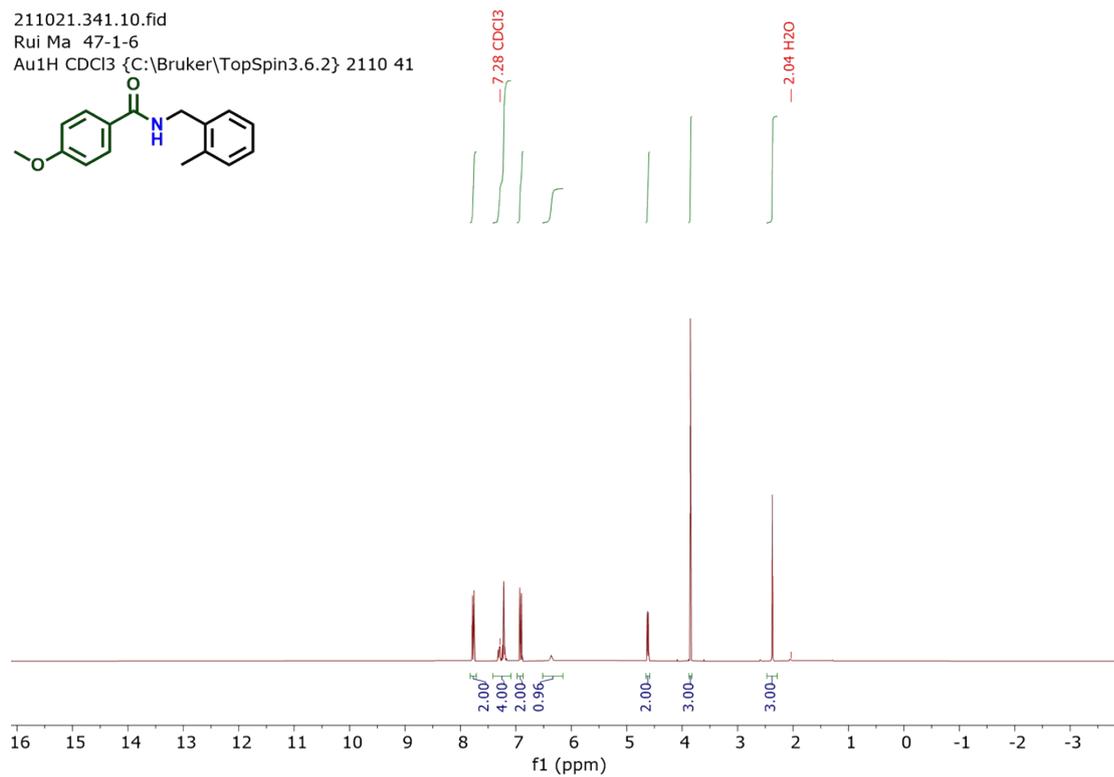
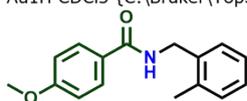
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Jie Gao 170-3-14
Au1H DMSO {C:\Bruker\TopSpin3.5pl6} 2106 42



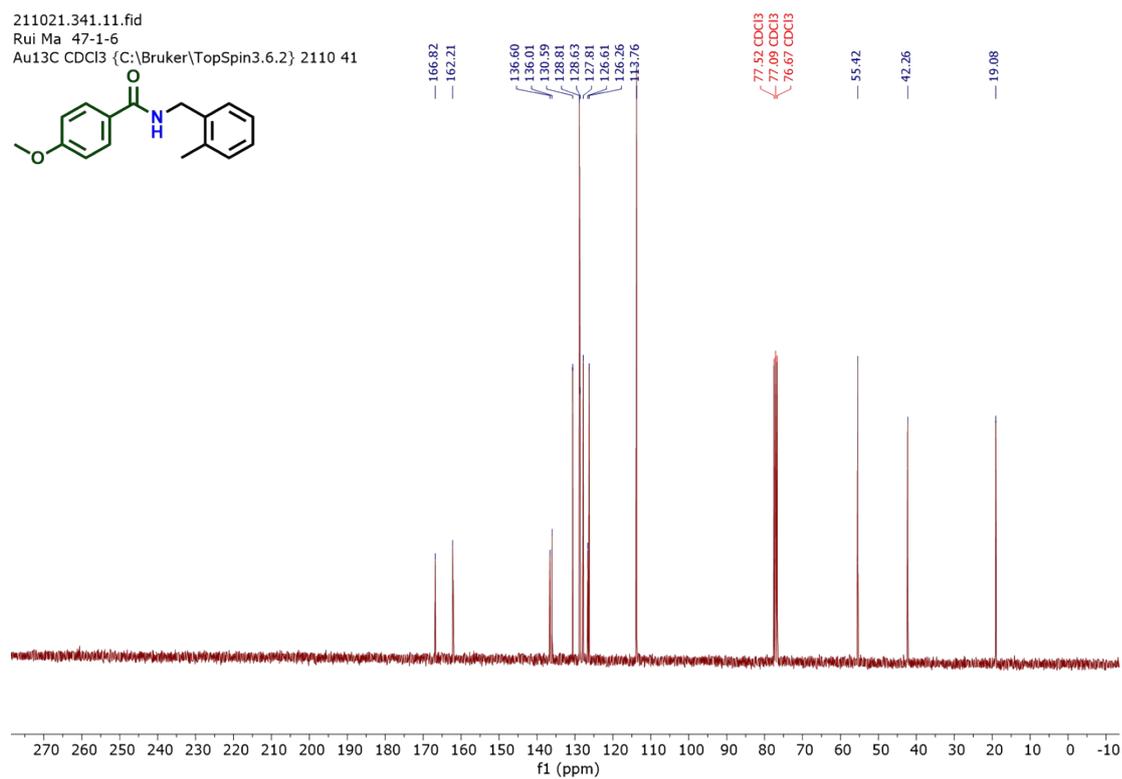
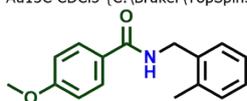
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Jie Gao 170-3-14
Au13C DMSO {C:\Bruker\TopSpin3.5pl6} 2106 42



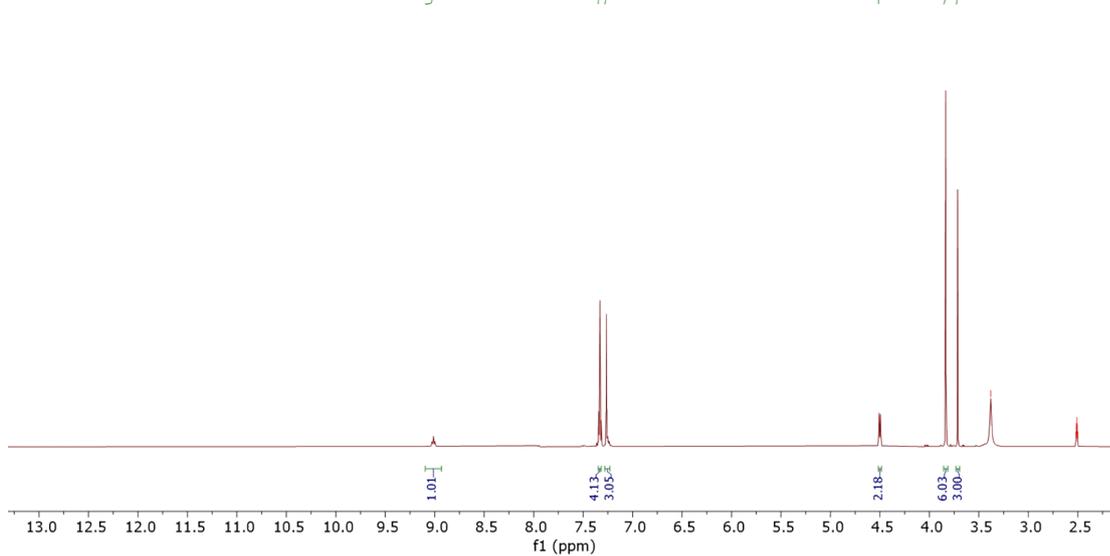
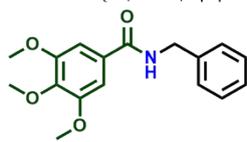
211021.341.10.fid
Rui Ma 47-1-6
Au1H CDCl3 {C:\Bruker\TopSpin3.6.2} 2110 41



211021.341.11.fid
Rui Ma 47-1-6
Au13C CDCl3 {C:\Bruker\TopSpin3.6.2} 2110 41

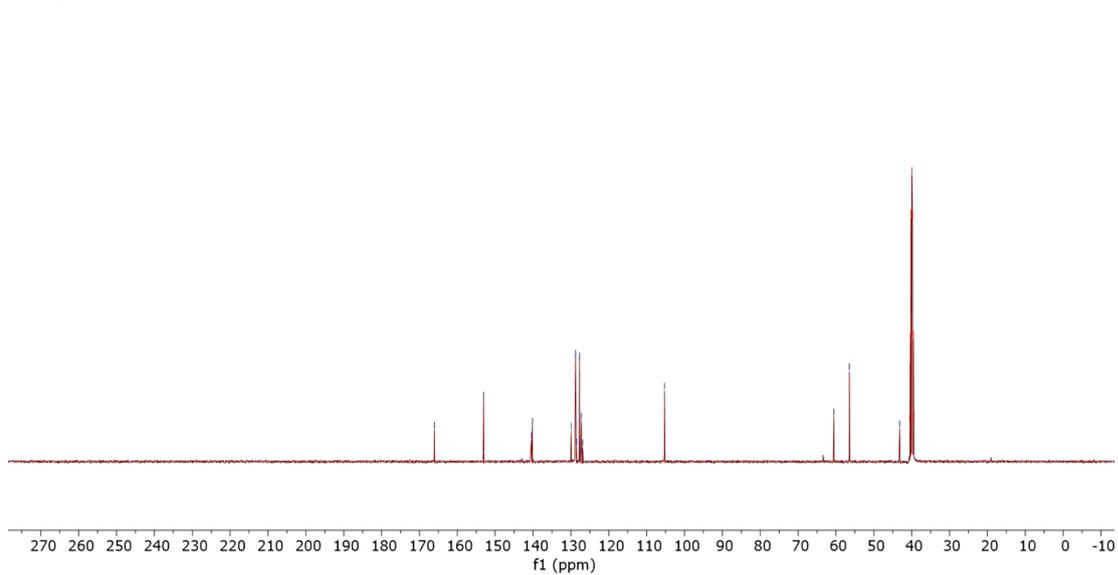
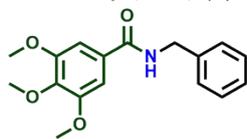


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Jie Gao 170-3-15
Au1H DMSO (C:\Bruker\TopSpin3.5pl6) 2106 58

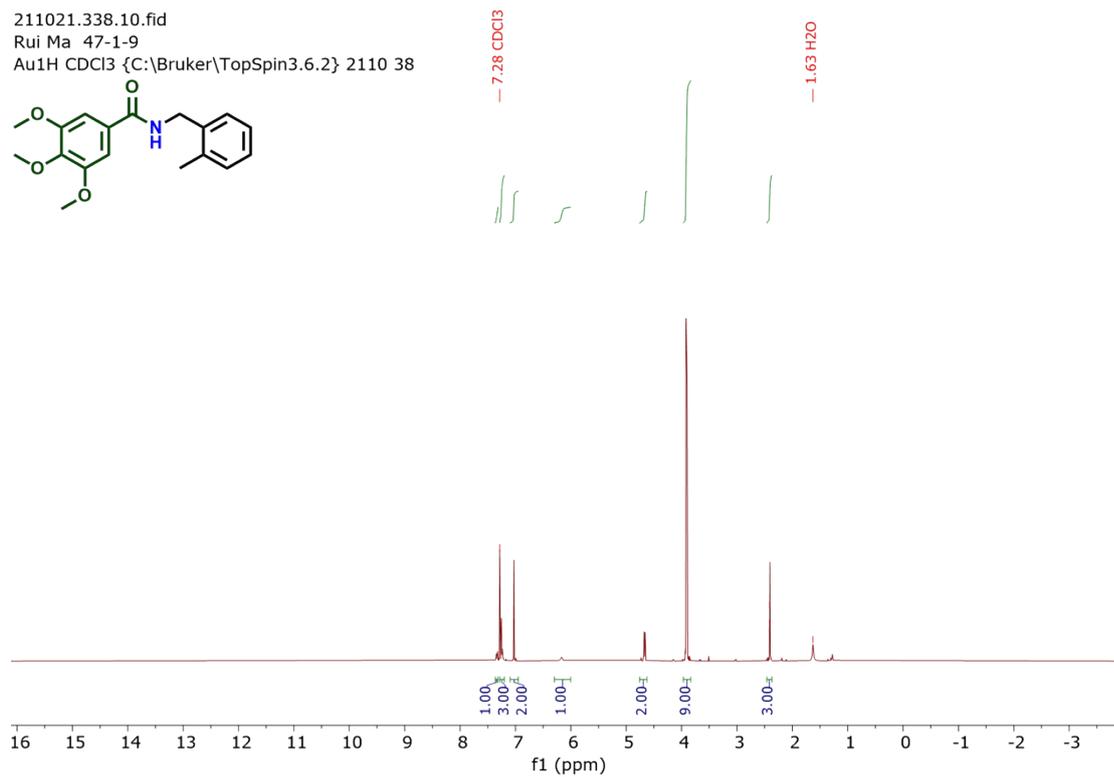
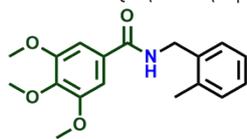


3.38 H2O
2.52 DMSO
2.51 DMSO
2.51 DMSO
2.50 DMSO

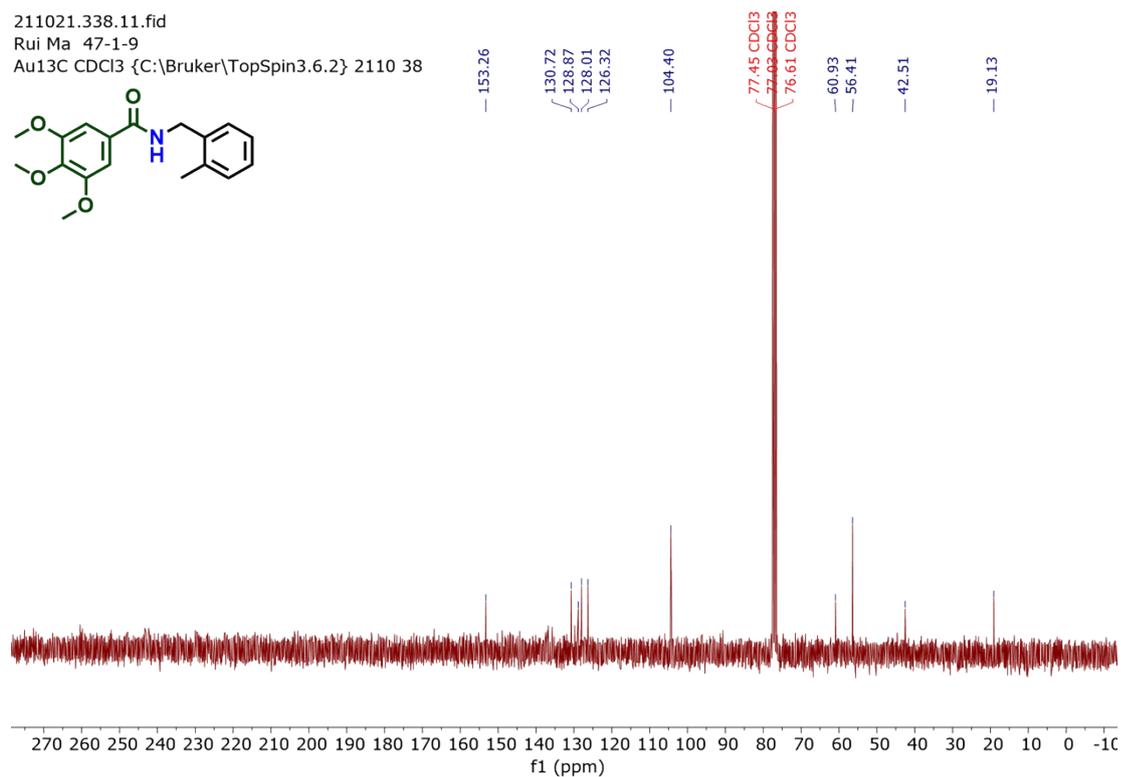
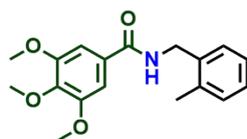
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Jie Gao 170-3-15
Au13C DMSO (C:\Bruker\TopSpin3.5pl6) 2106 58



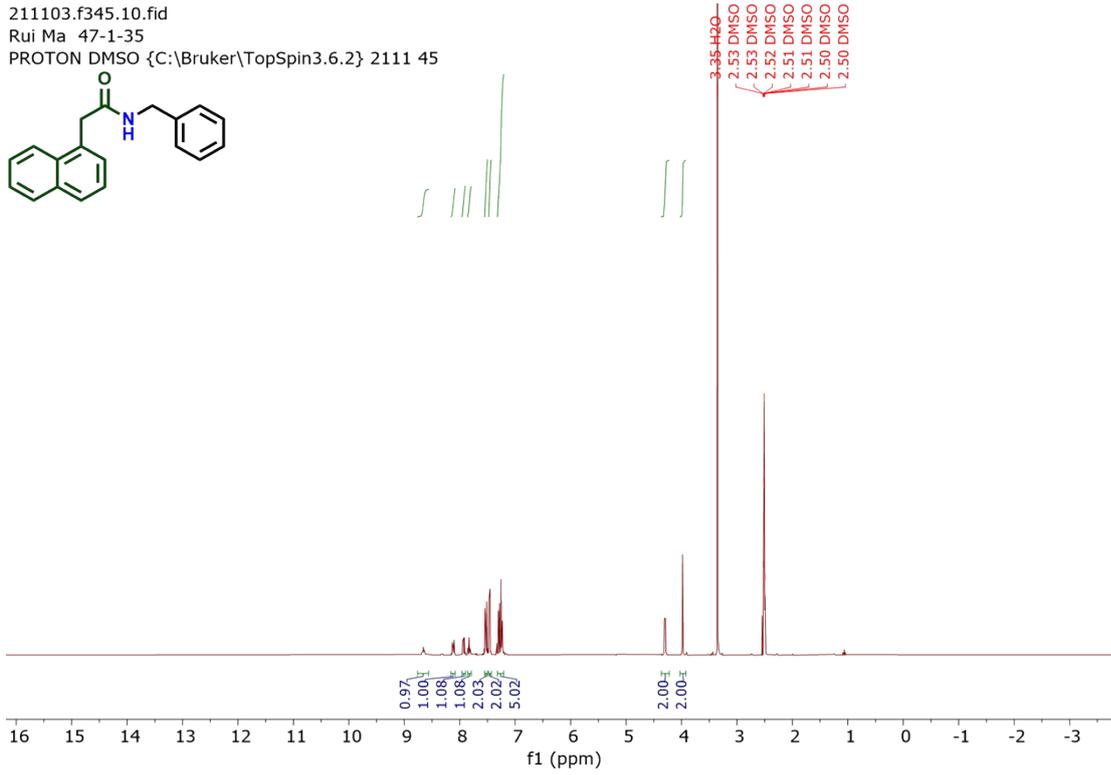
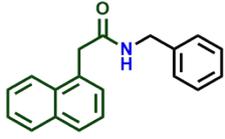
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Rui Ma 47-1-9
Au1H CDCl3 {C:\Bruker\TopSpin3.6.2} 2110 38



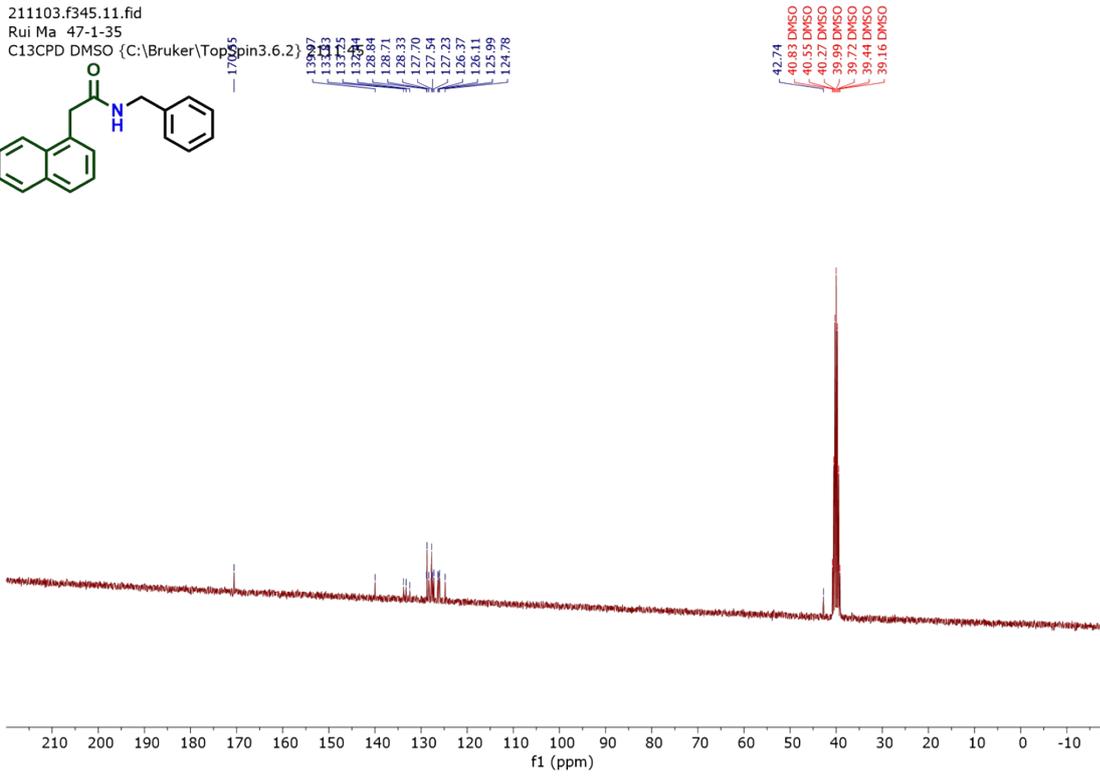
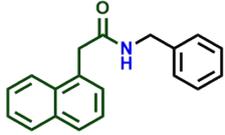
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Rui Ma 47-1-9
Au13C CDCl3 {C:\Bruker\TopSpin3.6.2} 2110 38



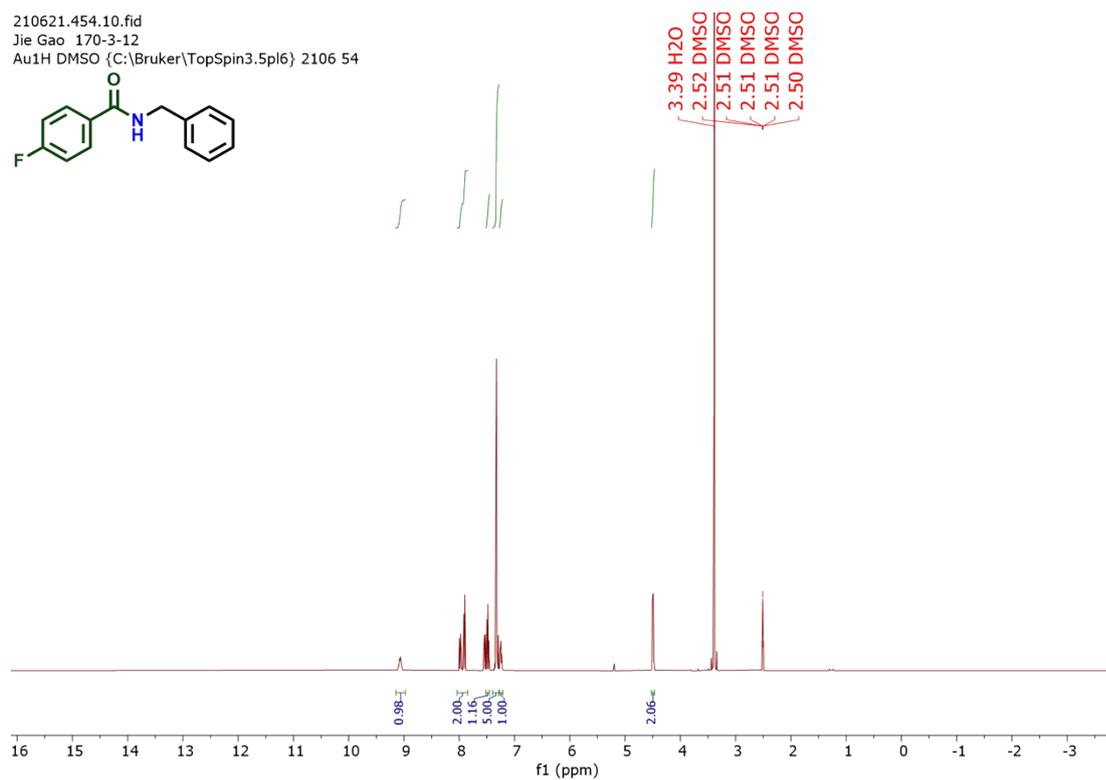
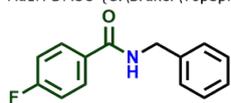
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Rui Ma 47-1-35
PROTON DMSO {C:\Bruker\TopSpin3.6.2} 2111 45



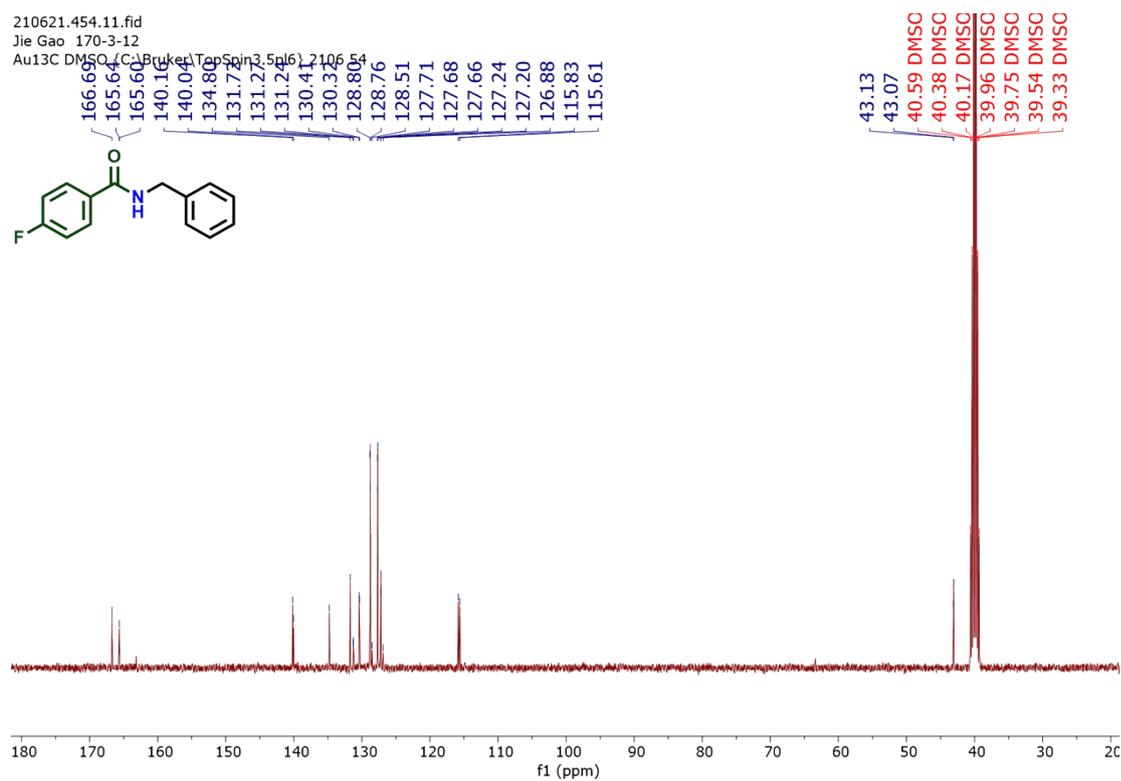
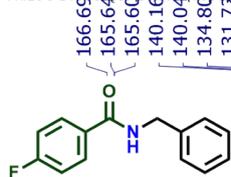
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C13CPD DMSO {C:\Bruker\TopSpin3.6.2}



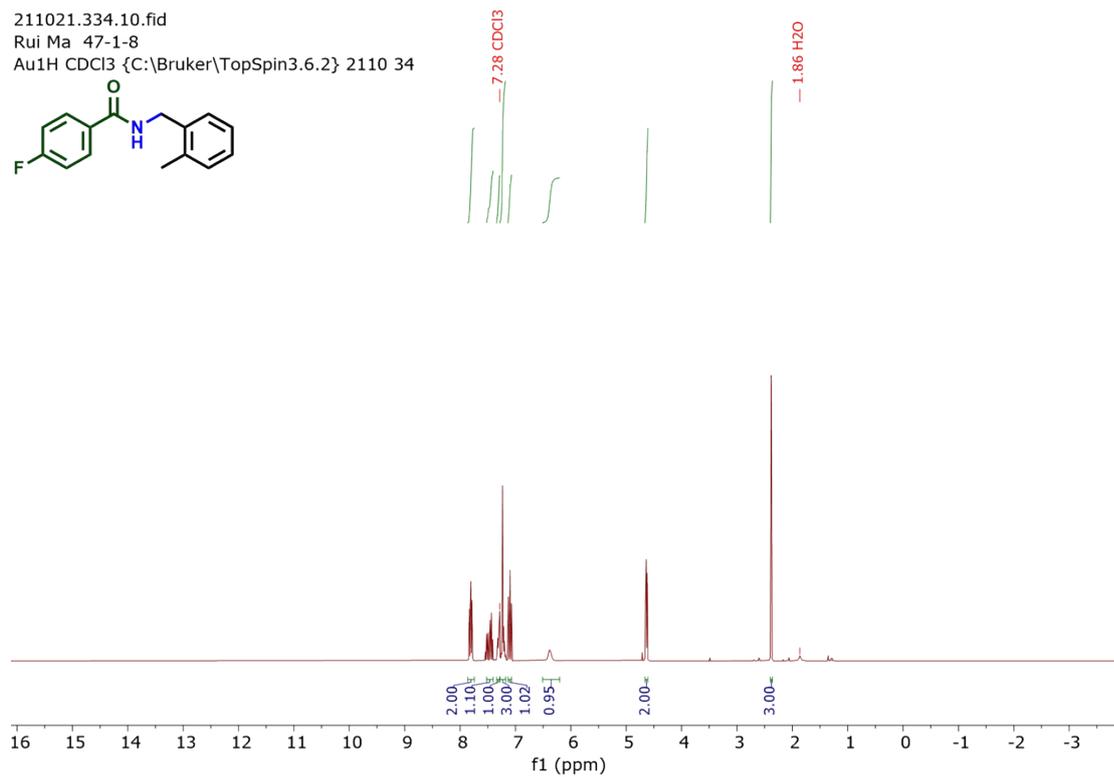
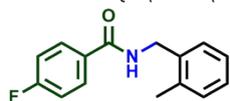
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Jie Gao 170-3-12
Au1H DMSO (C:\Bruker\TopSpin3.5pl6) 2106 54



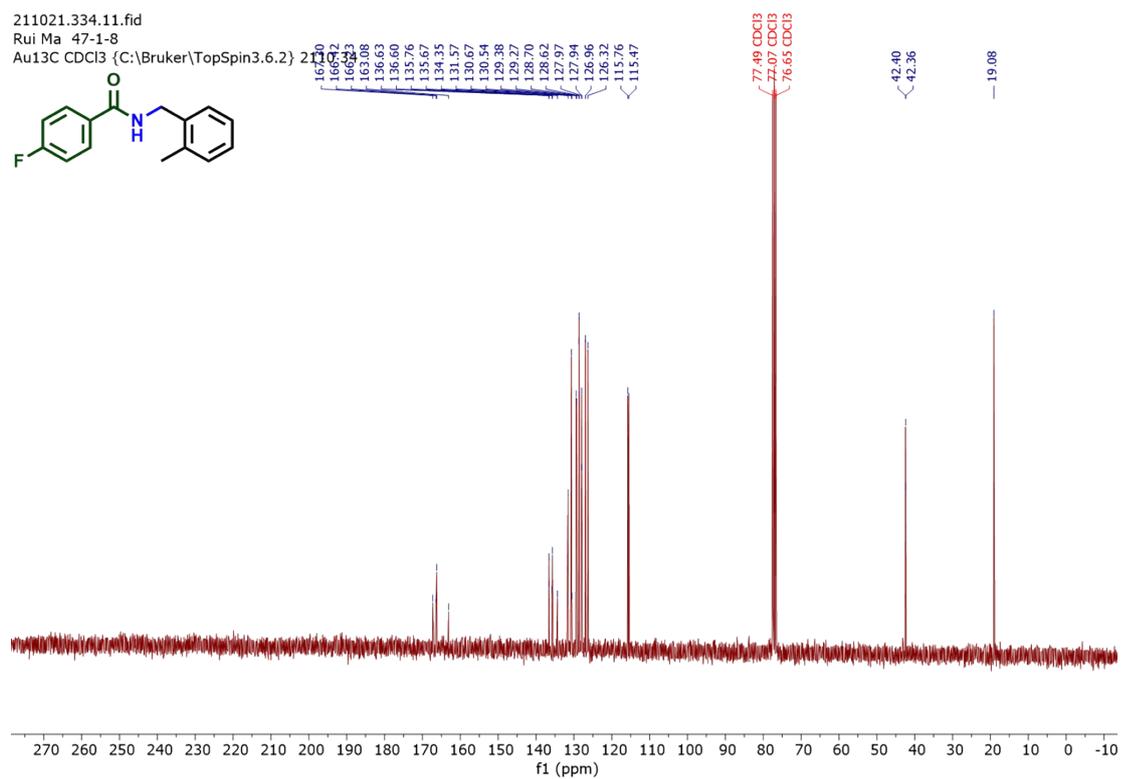
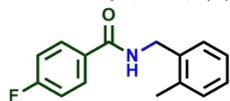
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Jie Gao 170-3-12
Au13C DMSO (C:\Bruker\TopSpin3.5pl6) 2106 54



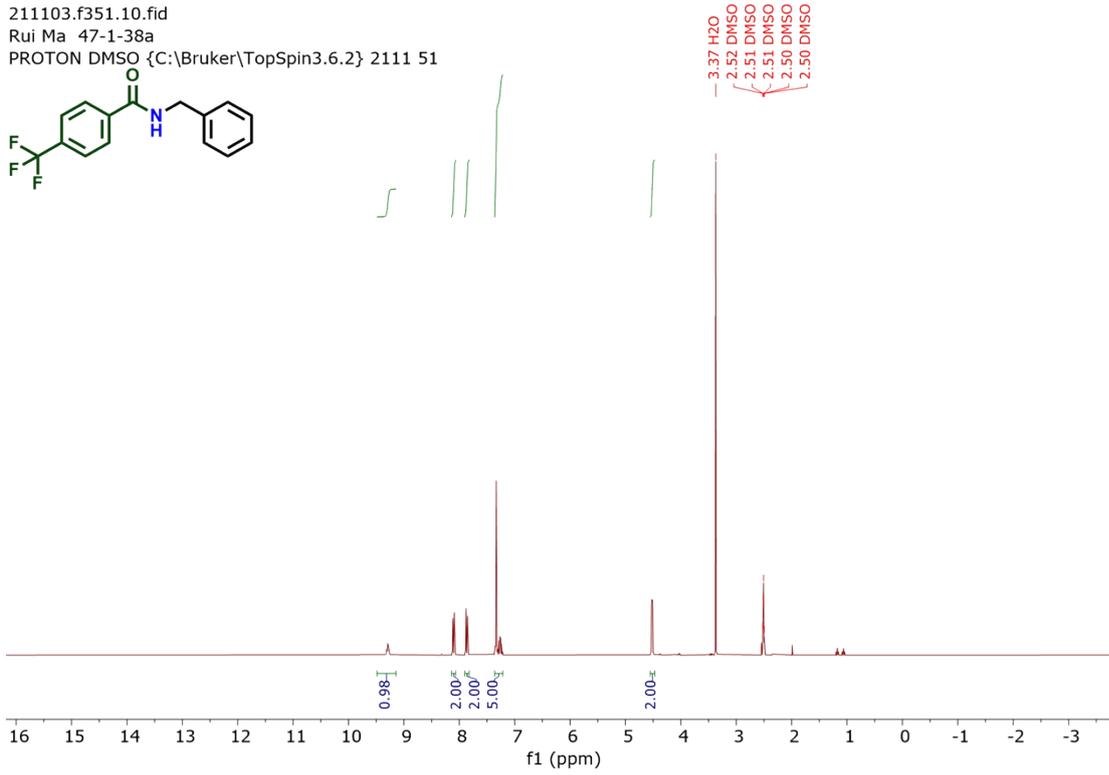
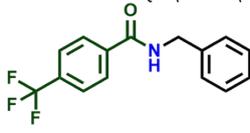
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Rui Ma 47-1-8
Au1H CDCl3 {C:\Bruker\TopSpin3.6.2} 2110 34



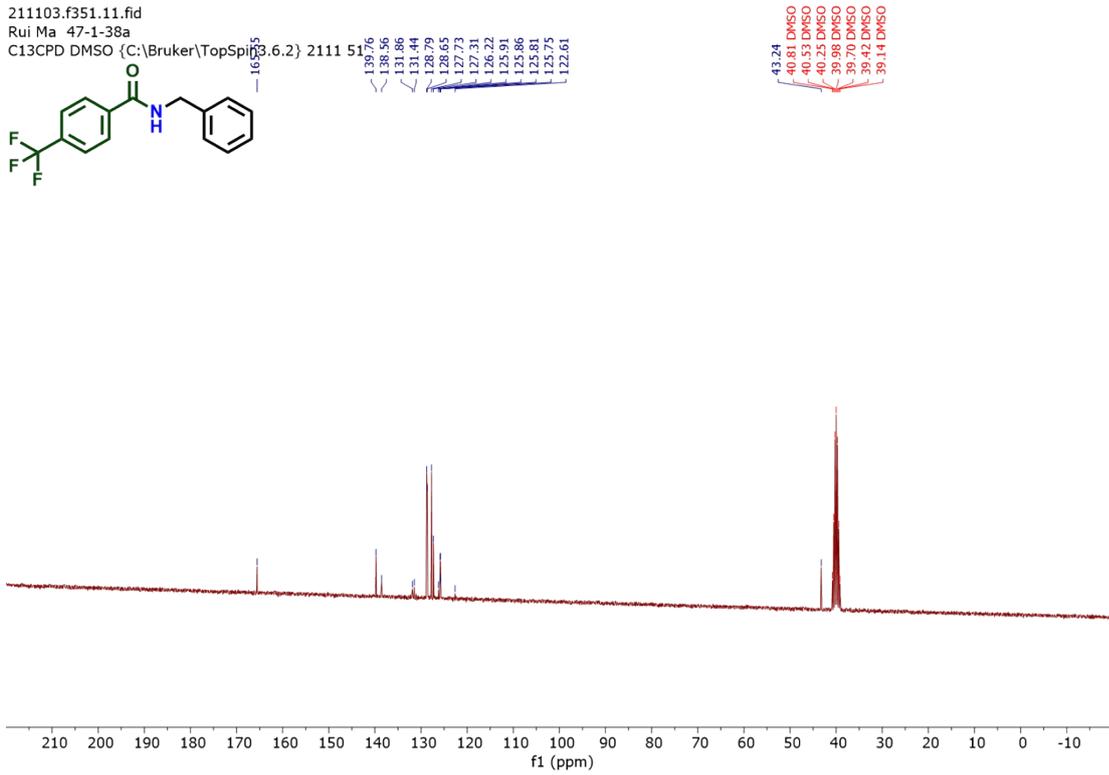
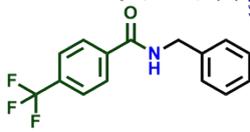
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Rui Ma 47-1-8
Au13C CDCl3 {C:\Bruker\TopSpin3.6.2} 2110 34



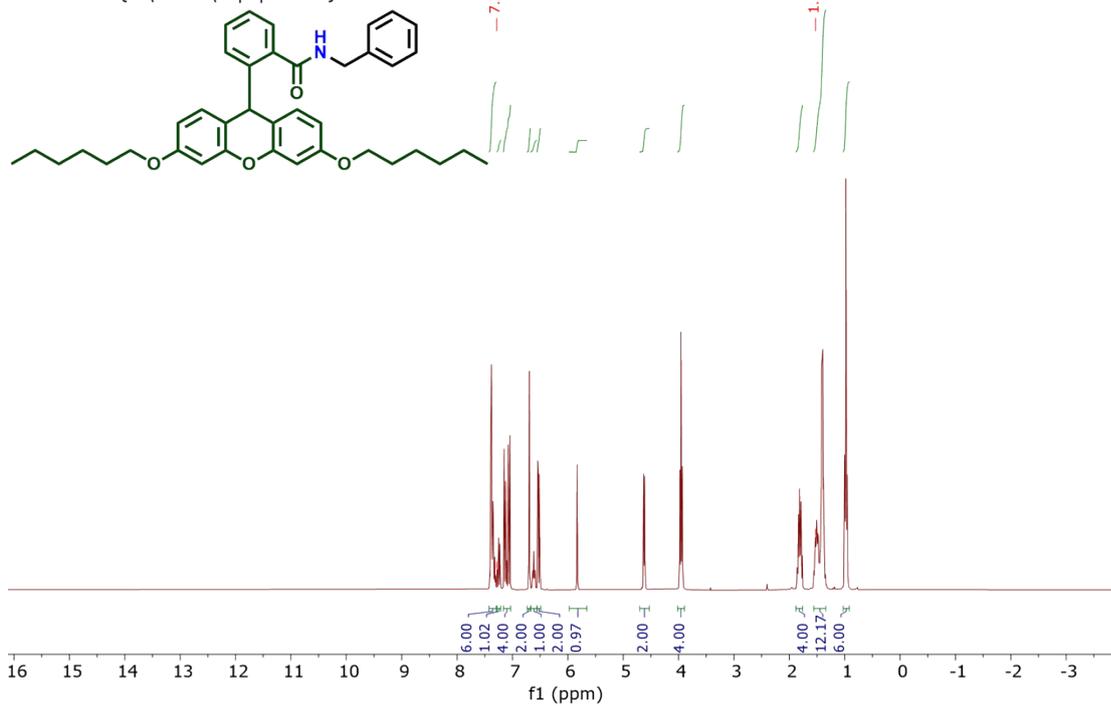
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Rui Ma 47-1-38a
PROTON DMSO {C:\Bruker\TopSpin3.6.2} 2111 51



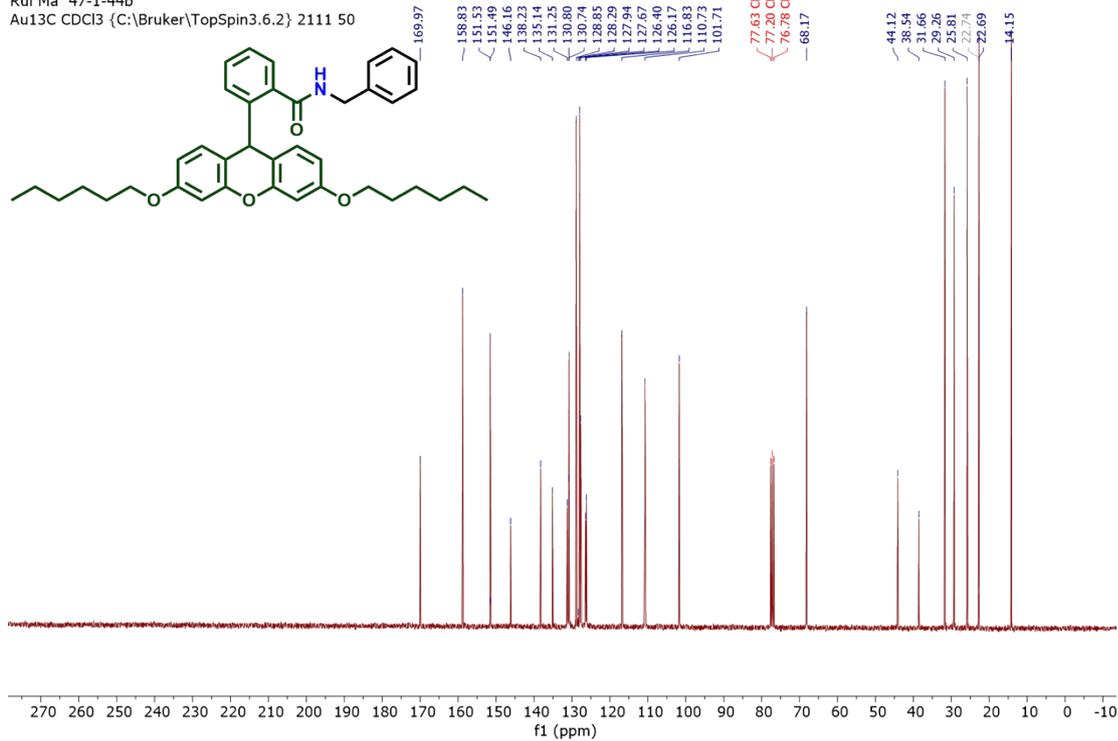
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C13CPD DMSO {C:\Bruker\TopSpin3.6.2} 2111 51



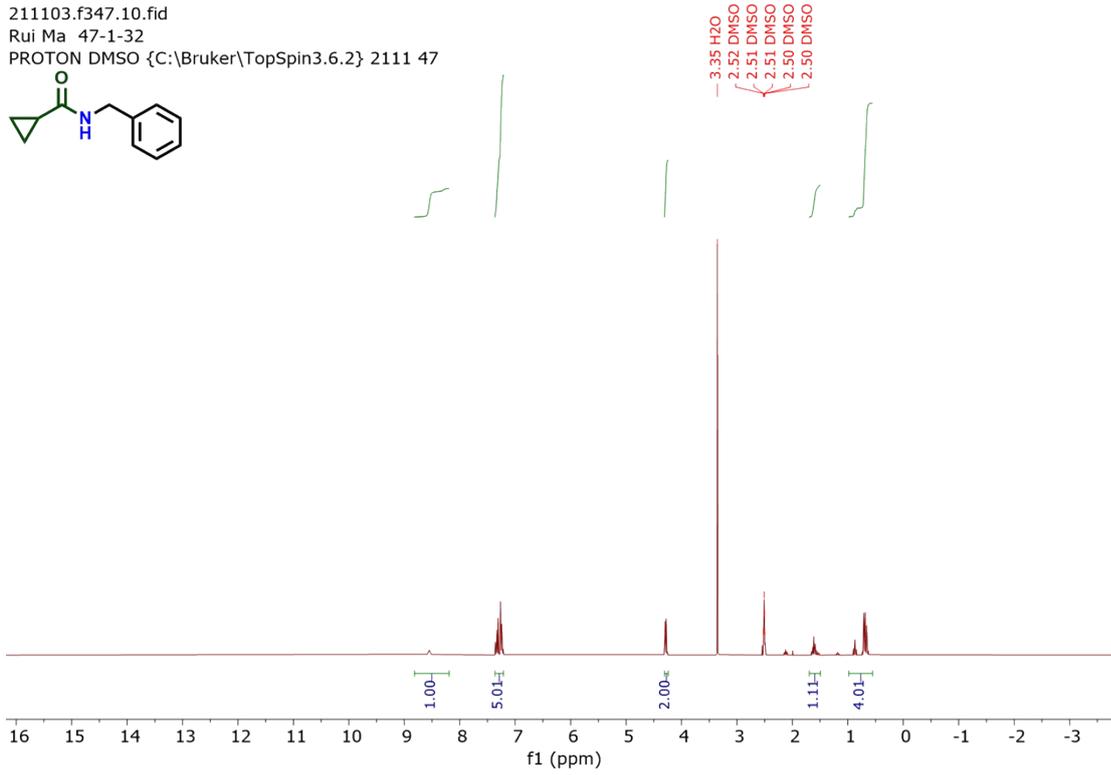
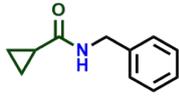
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Au1H CDCl3 {C:\Bruker\TopSpin3.6.2} 2111 50



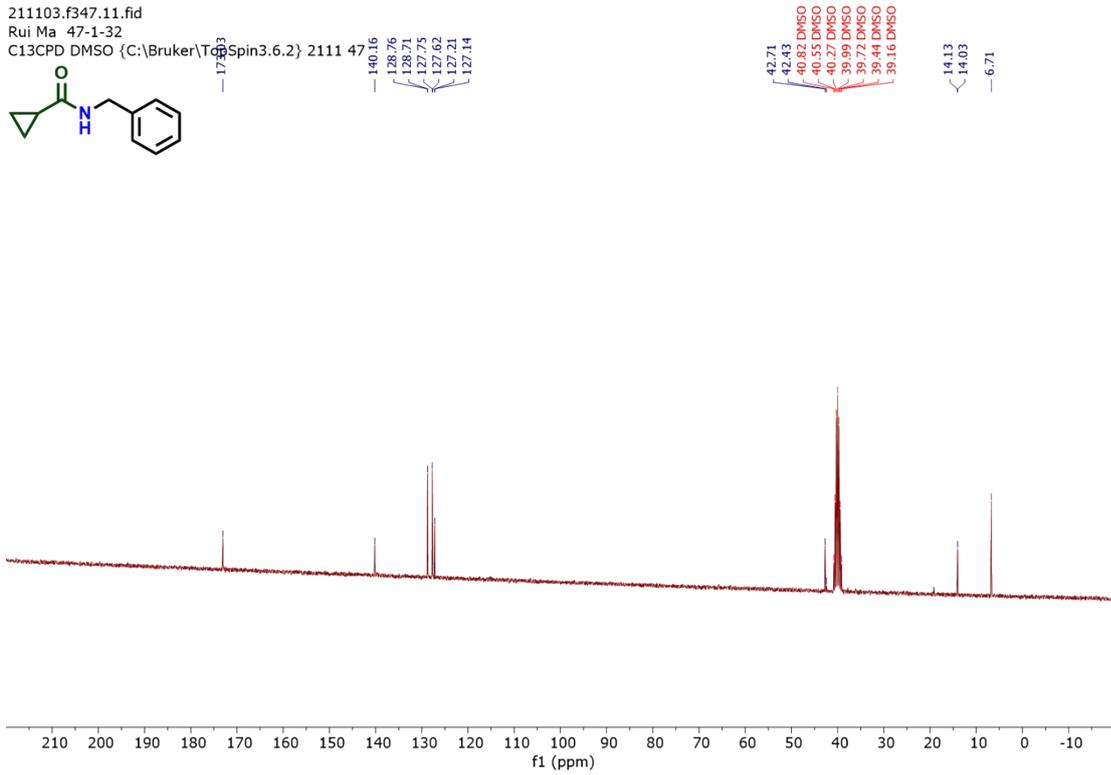
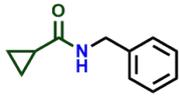
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Au13C CDCl3 {C:\Bruker\TopSpin3.6.2} 2111 50



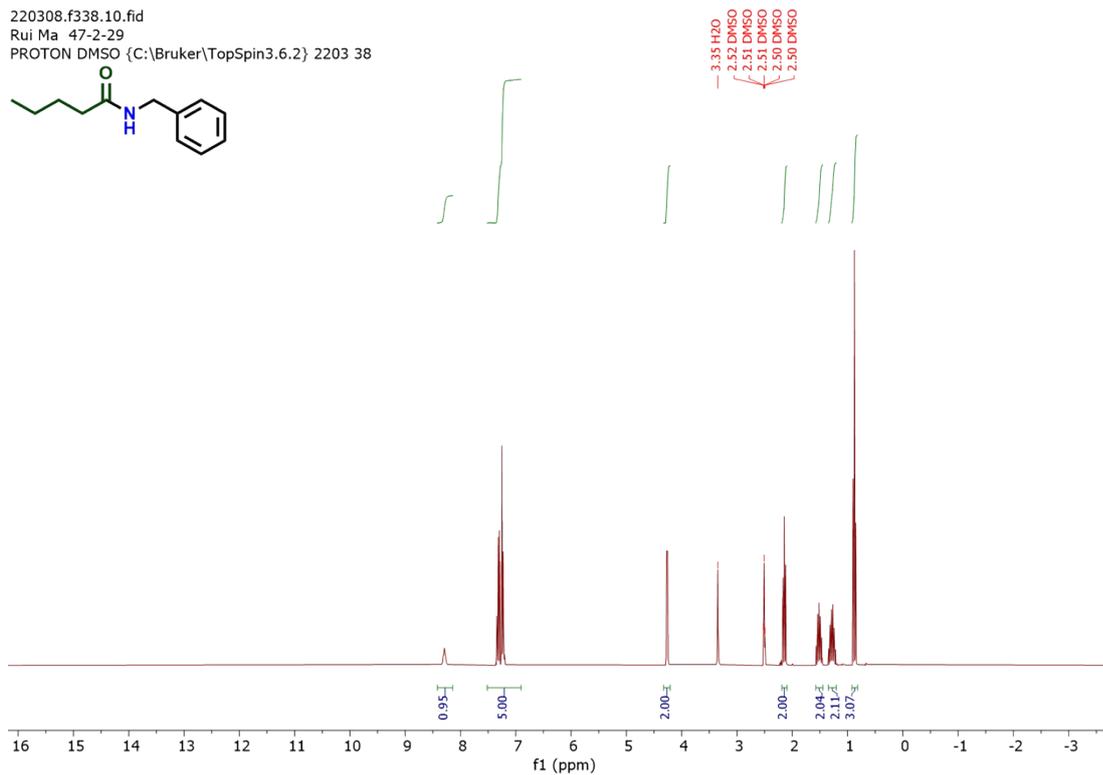
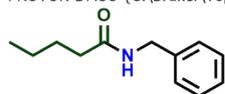
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Rui Ma 47-1-32
PROTON DMSO {C:\Bruker\TopSpin3.6.2} 2111 47



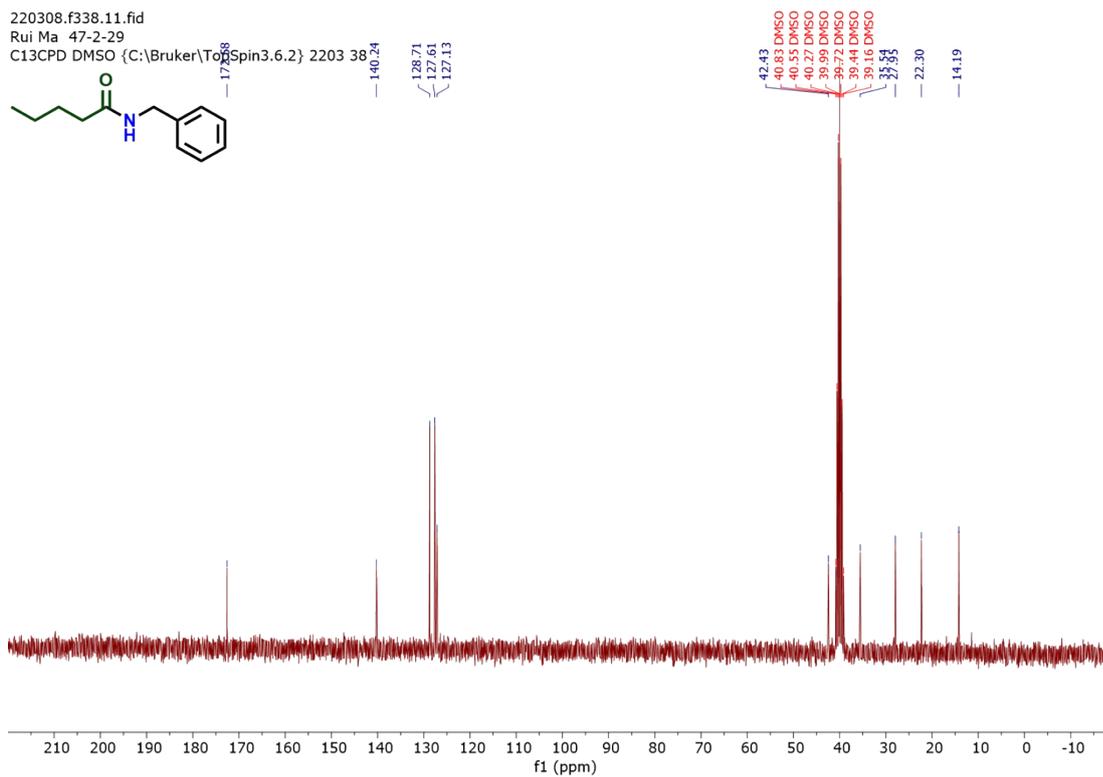
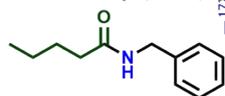
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C13CPD DMSO {C:\Bruker\TopSpin3.6.2} 2111 47



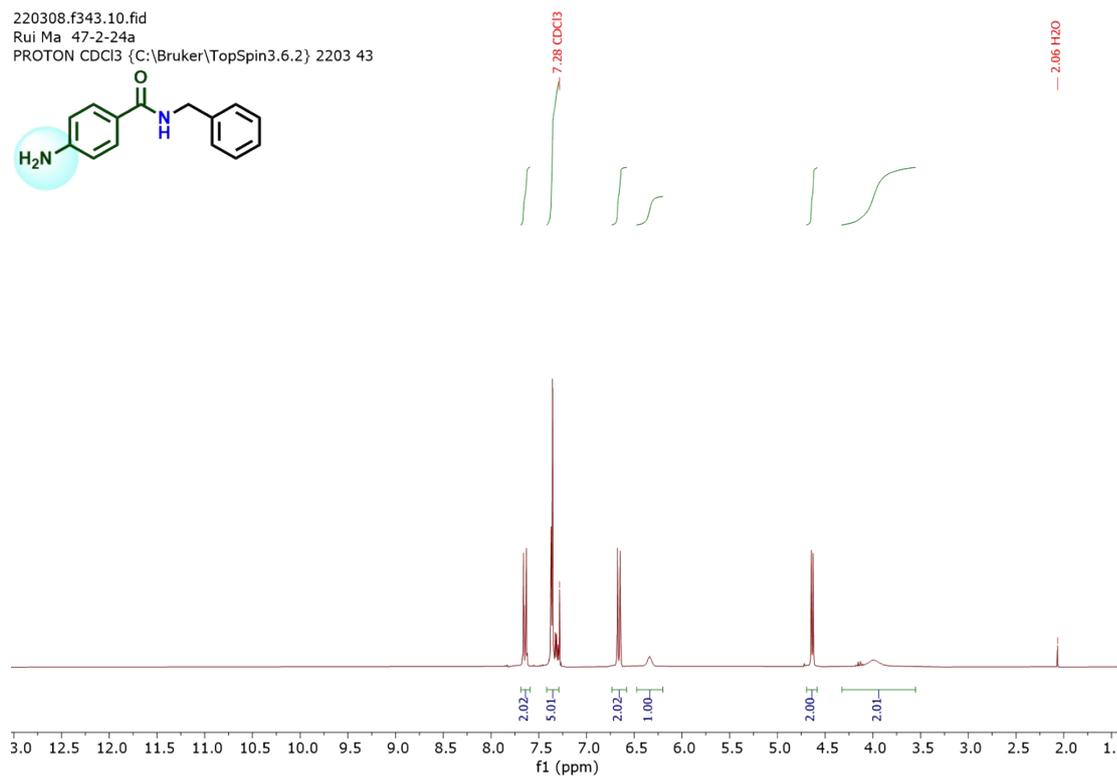
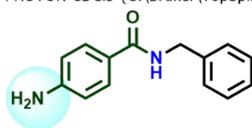
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Rui Ma 47-2-29
PROTON DMSO (C:\Bruker\TopSpin3.6.2) 2203 38



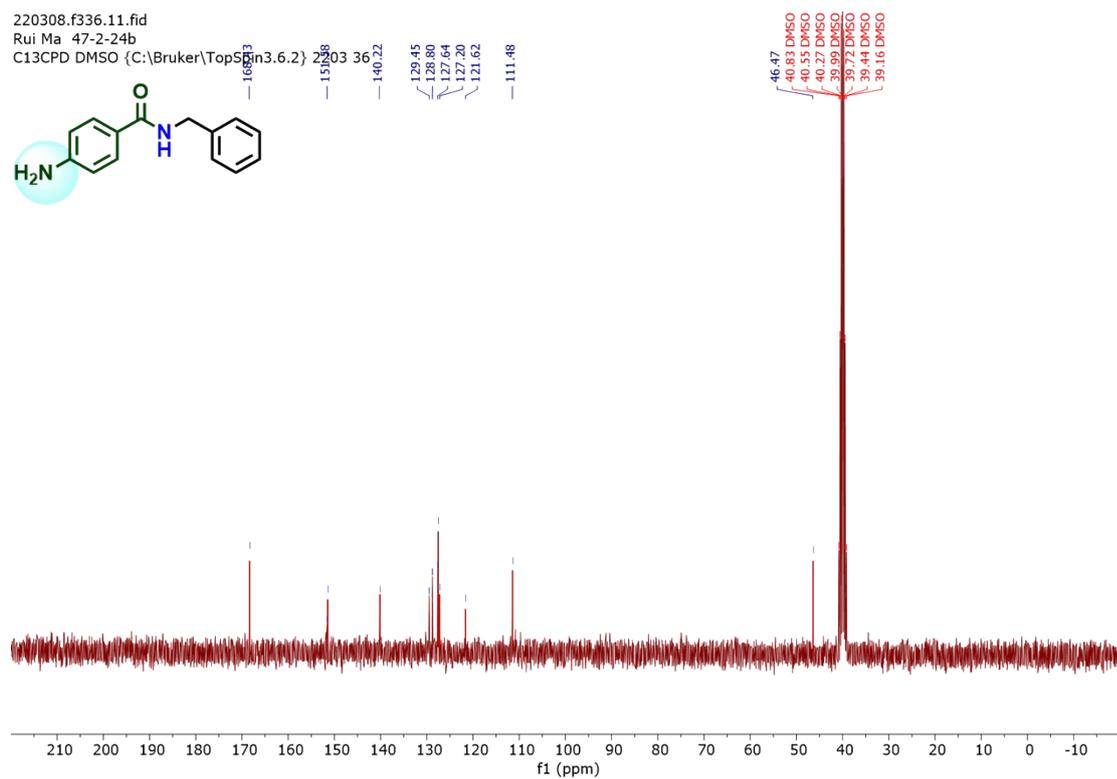
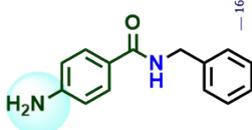
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C13CPD DMSO (C:\Bruker\TopSpin3.6.2) 2203 38



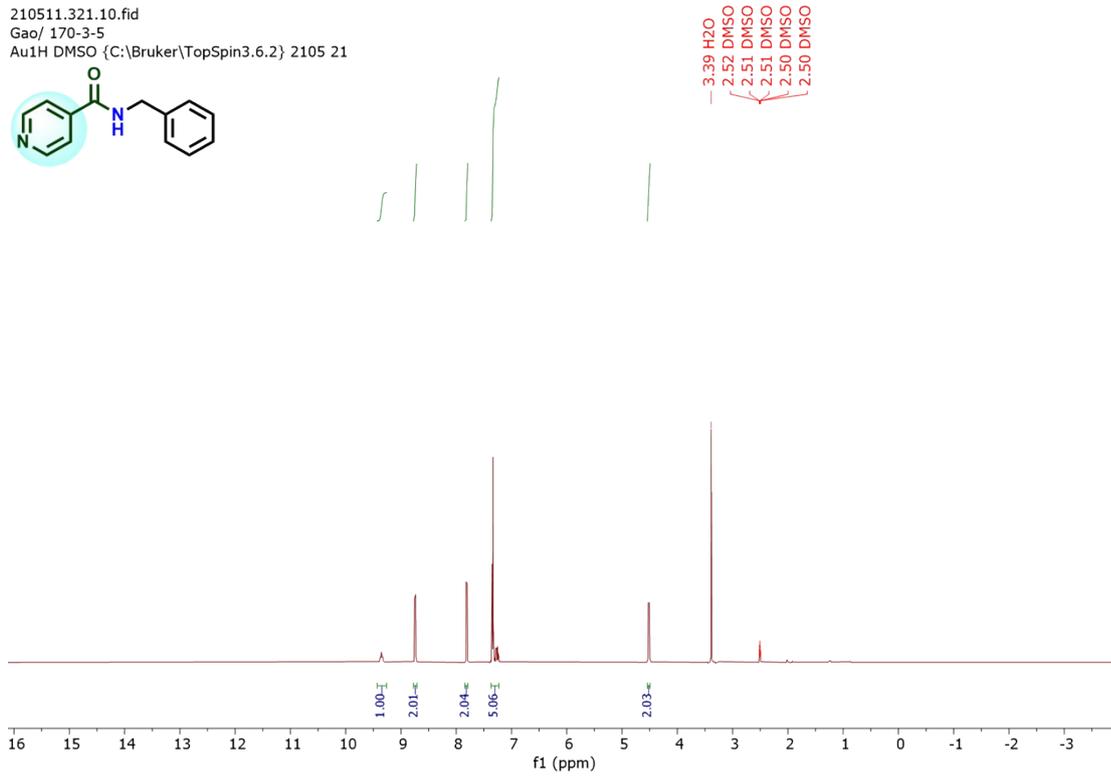
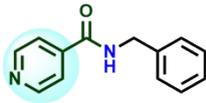
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PROTON CDCl3 {C:\Bruker\TopSpin3.6.2} 2203 43



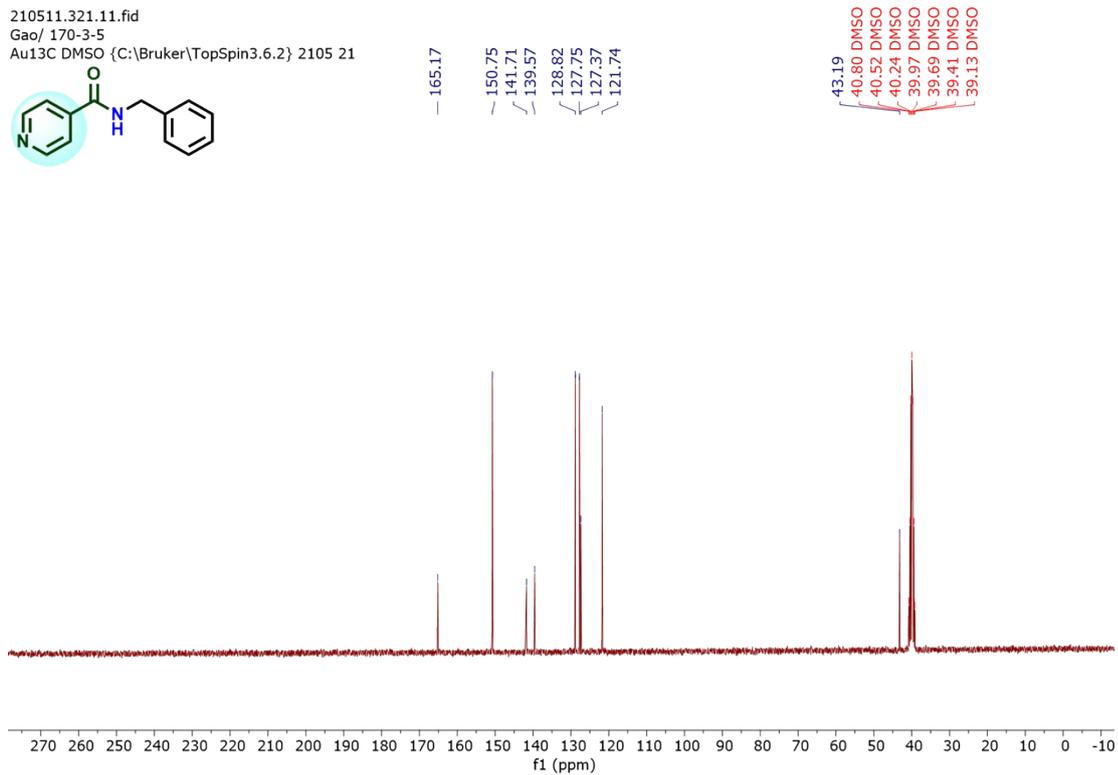
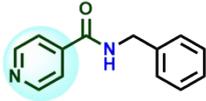
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C13CPD DMSO {C:\Bruker\TopSpin3.6.2} 2203 36



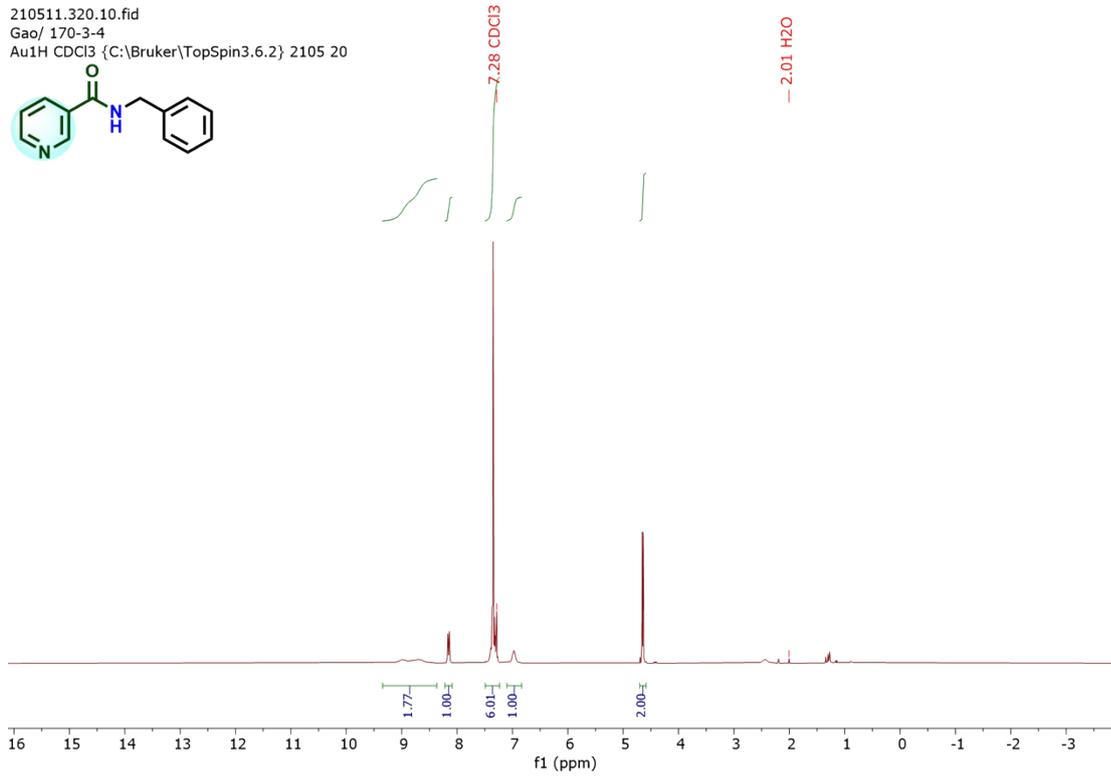
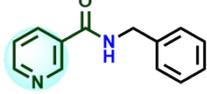
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Gao/ 170-3-5
Au1H DMSO {C:\Bruker\TopSpin3.6.2} 2105 21



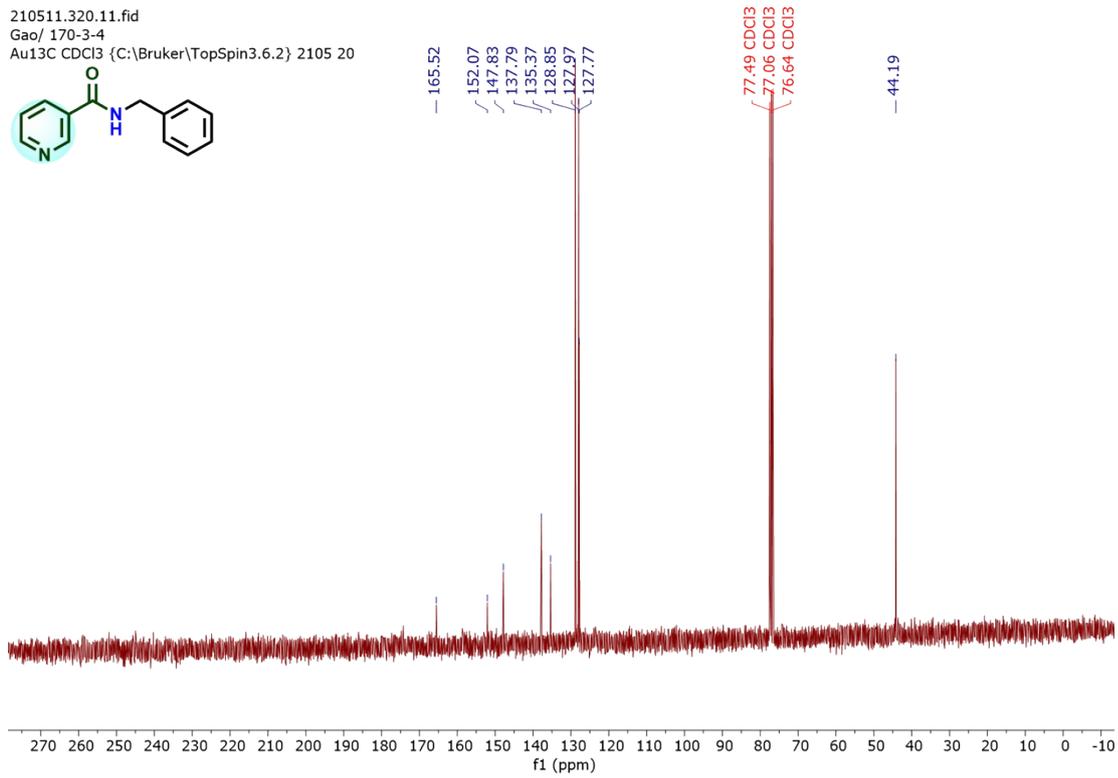
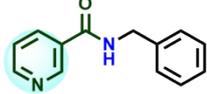
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Gao/ 170-3-5
Au13C DMSO {C:\Bruker\TopSpin3.6.2} 2105 21



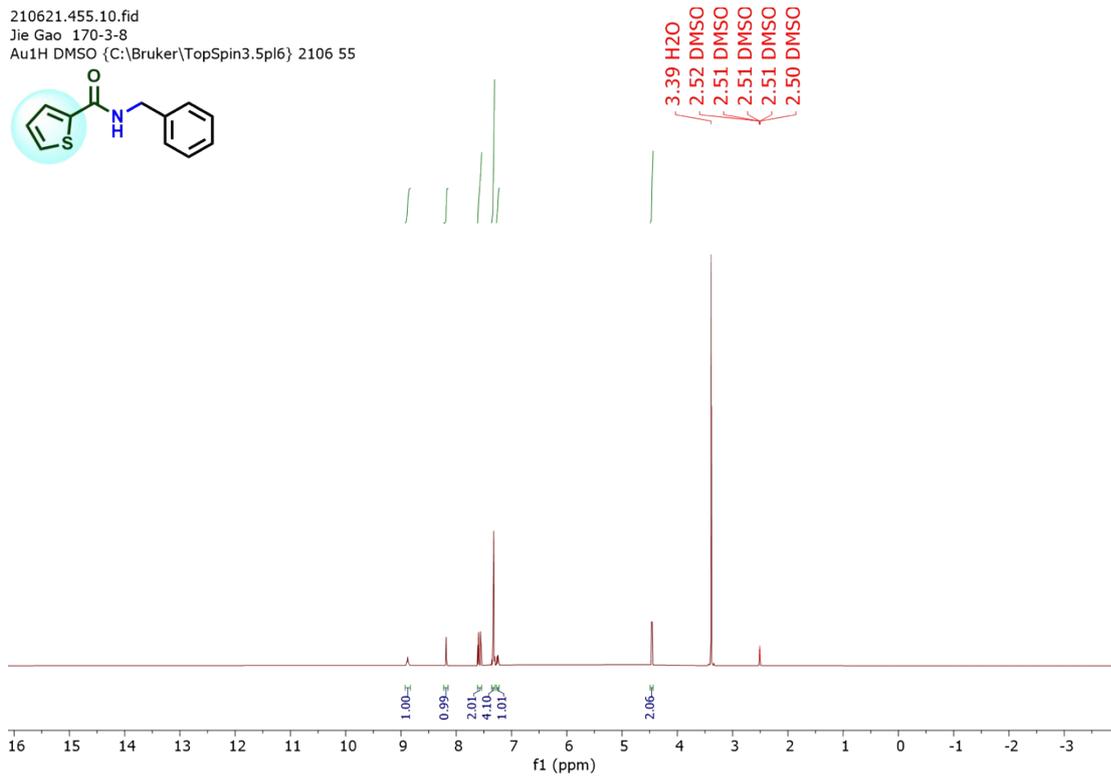
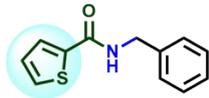
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Gao/ 170-3-4
Au1H CDCl3 {C:\Bruker\TopSpin3.6.2} 2105 20



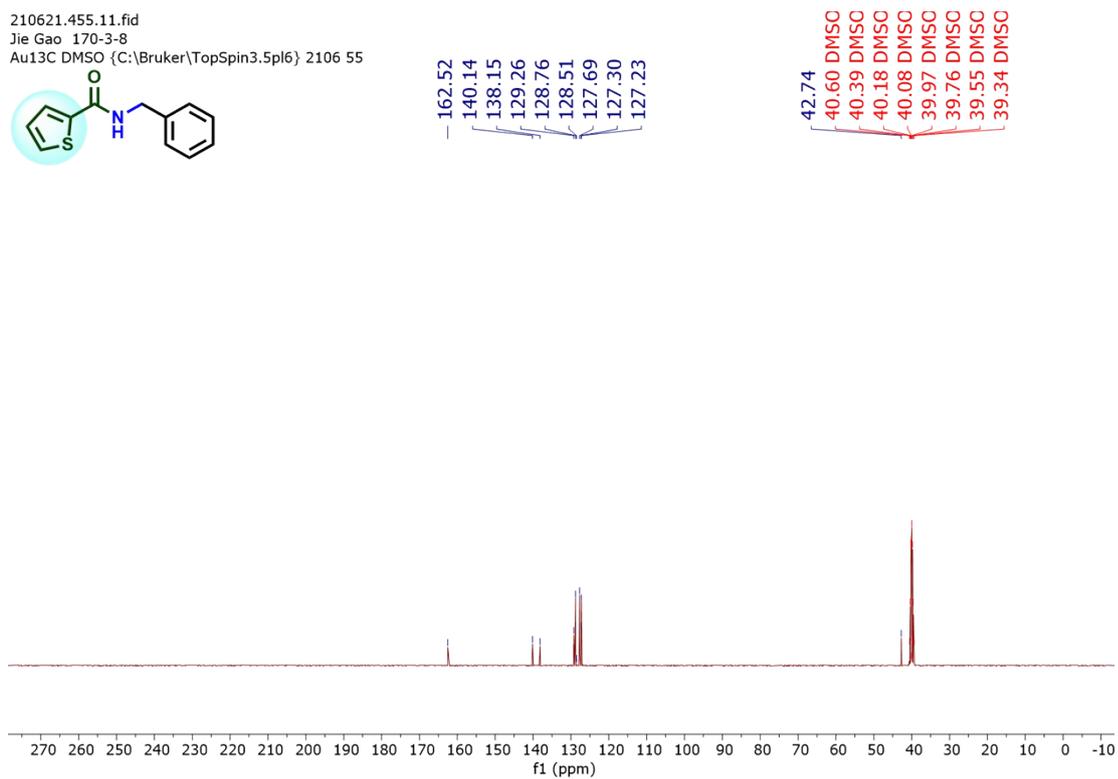
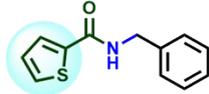
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Au13C CDCl3 {C:\Bruker\TopSpin3.6.2} 2105 20



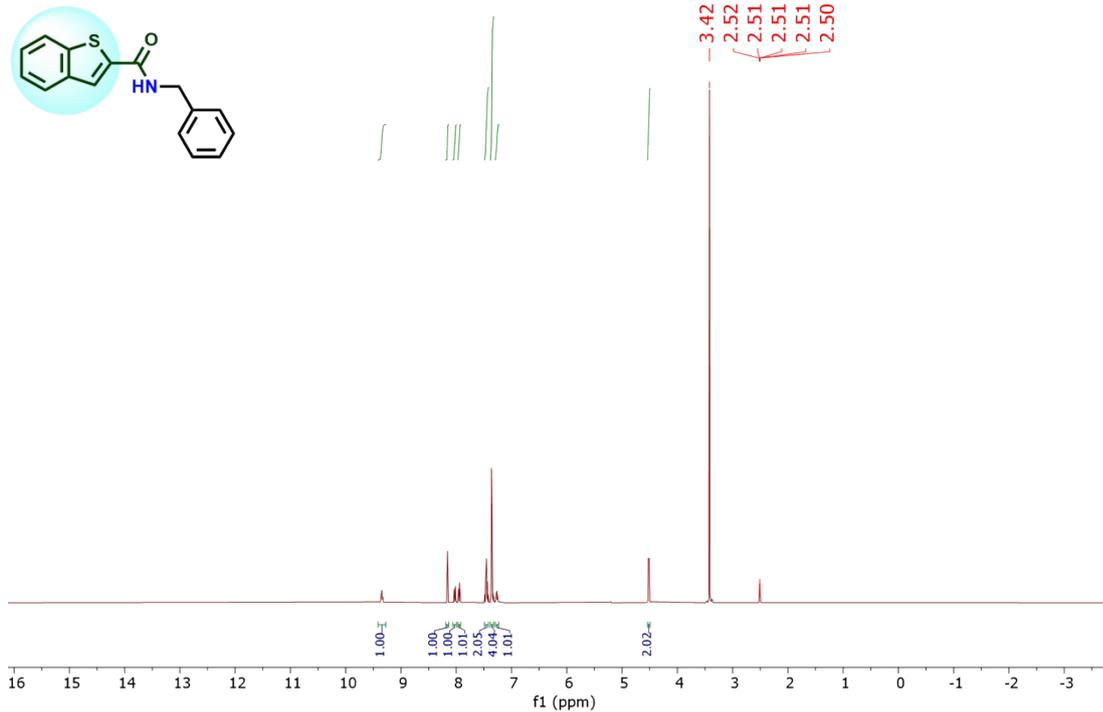
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Jie Gao 170-3-8
Au1H DMSO {C:\Bruker\TopSpin3.5pl6} 2106 55



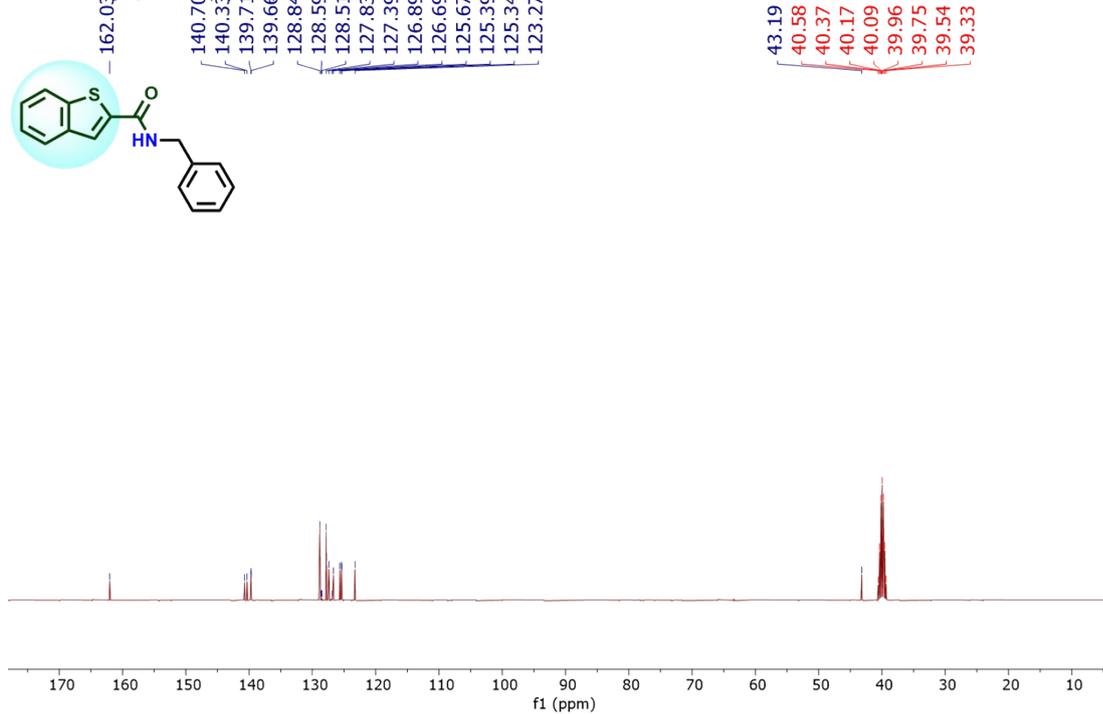
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Au13C DMSO {C:\Bruker\TopSpin3.5pl6} 2106 55



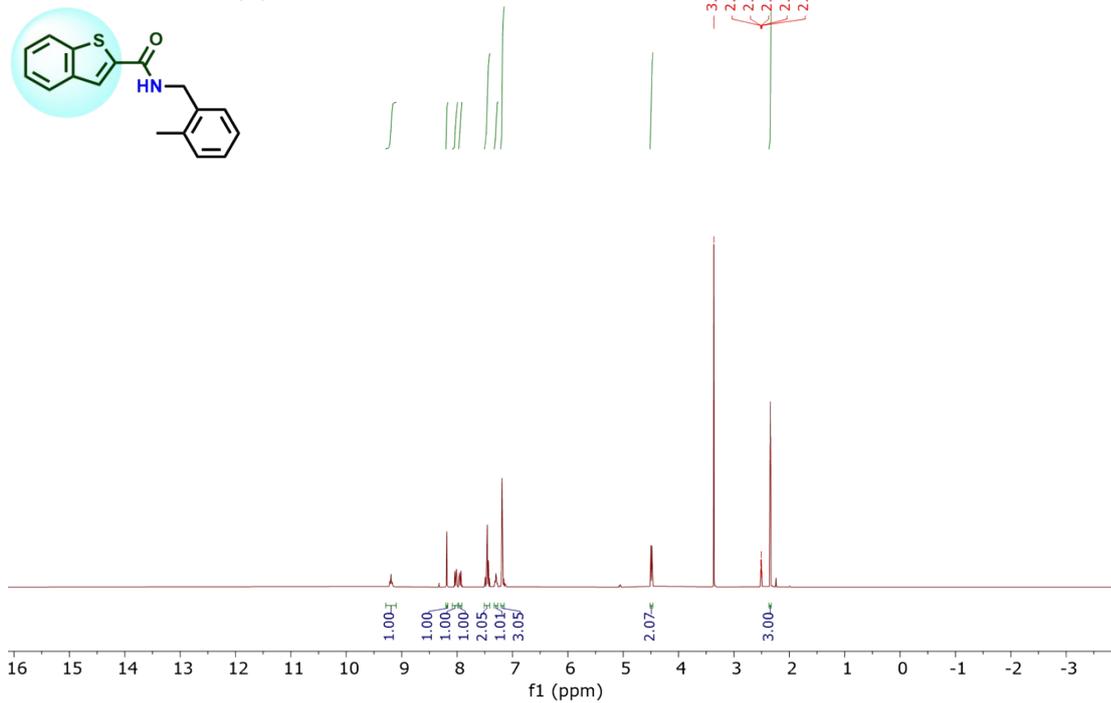
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Au1H DMSO (C:\Bruker\TopSpin3.5pl6) 2106 59



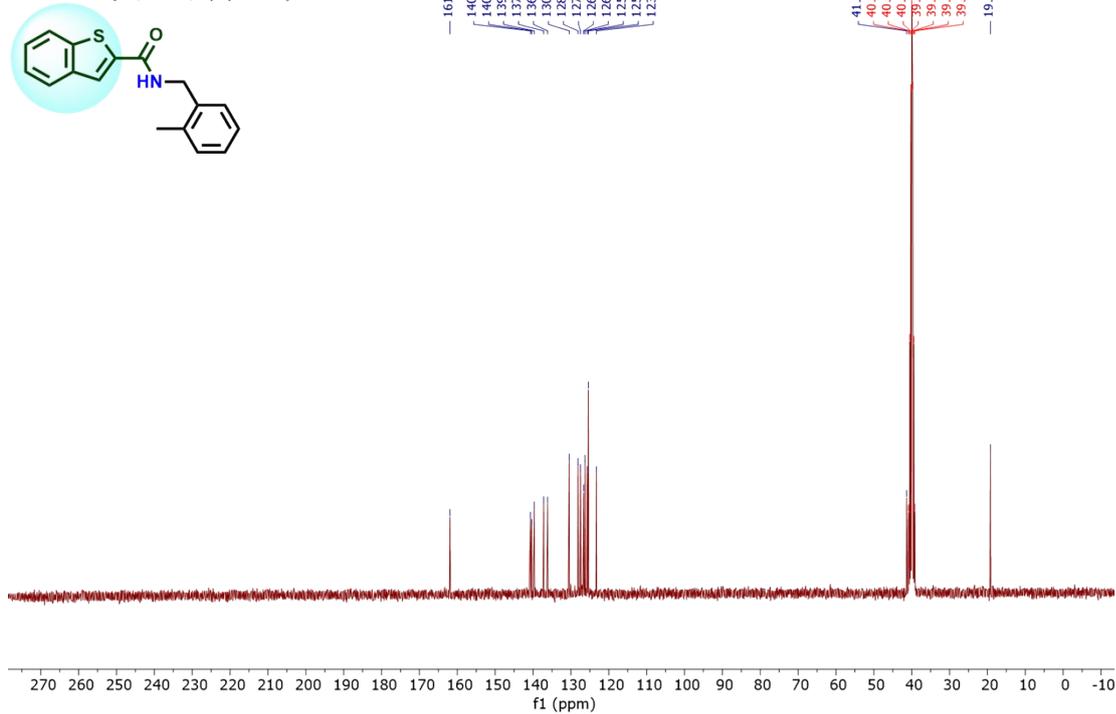
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Au13C DMSO (C:\Bruker\TopSpin3.5pl6) 2106 59



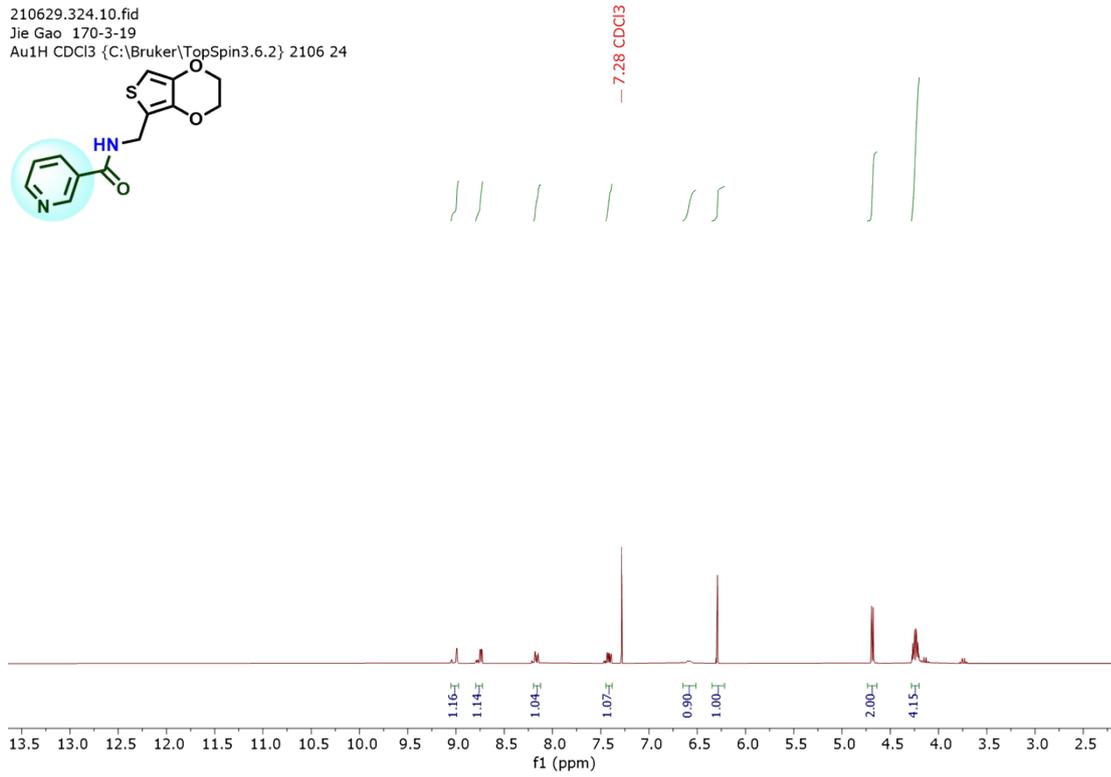
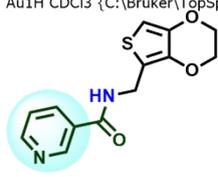
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Rui Ma 47-1-10
Au1H DMSO {C:\Bruker\TopSpin3.6.2} 2110 9



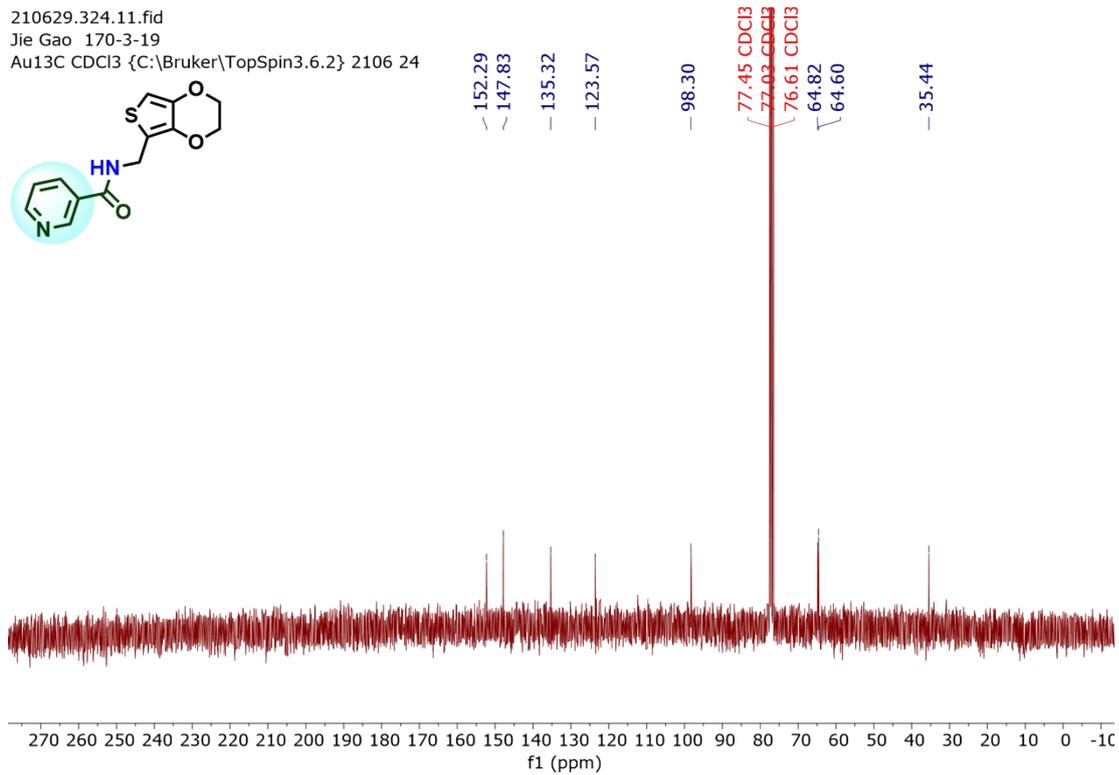
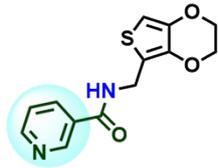
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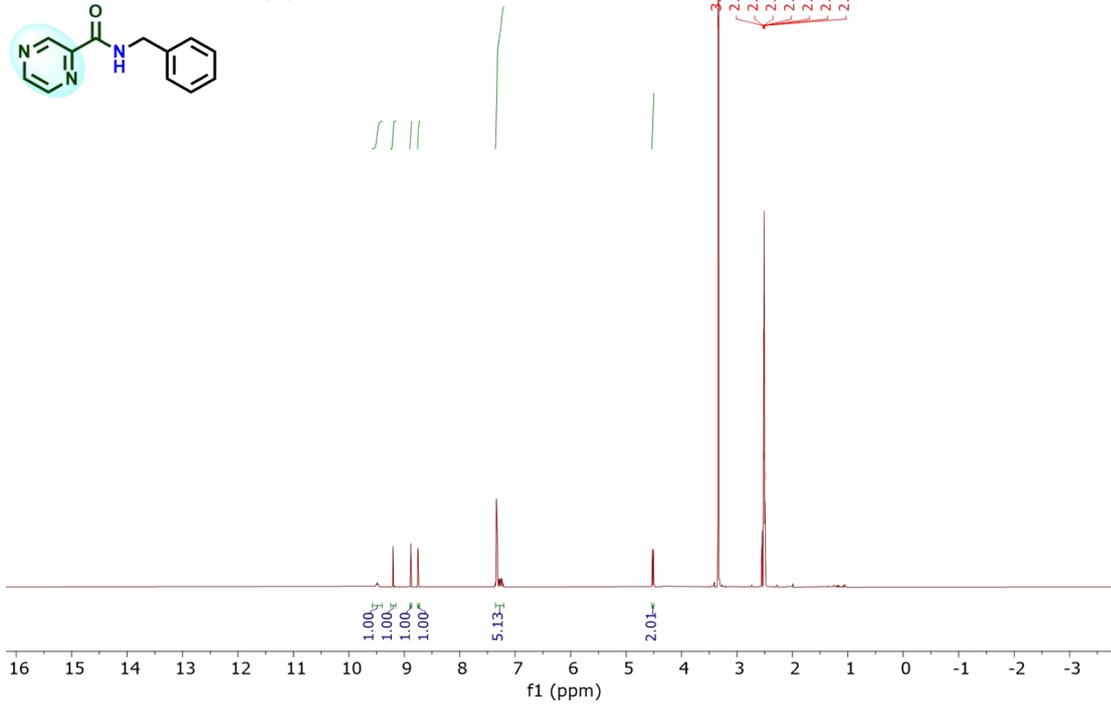
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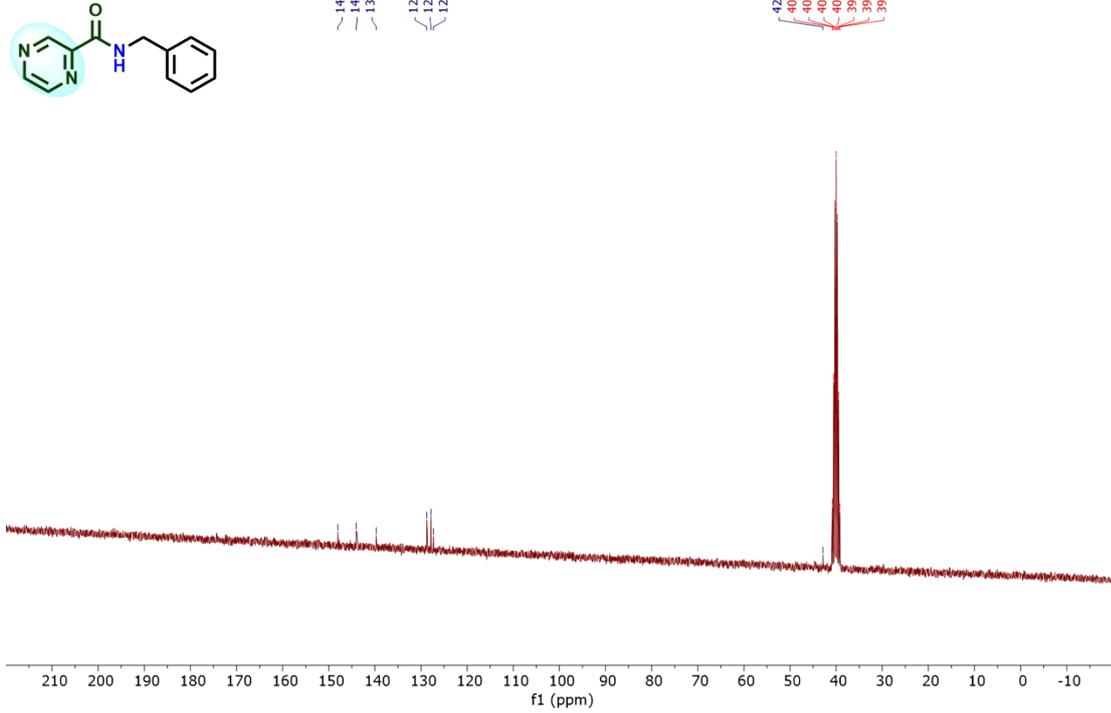
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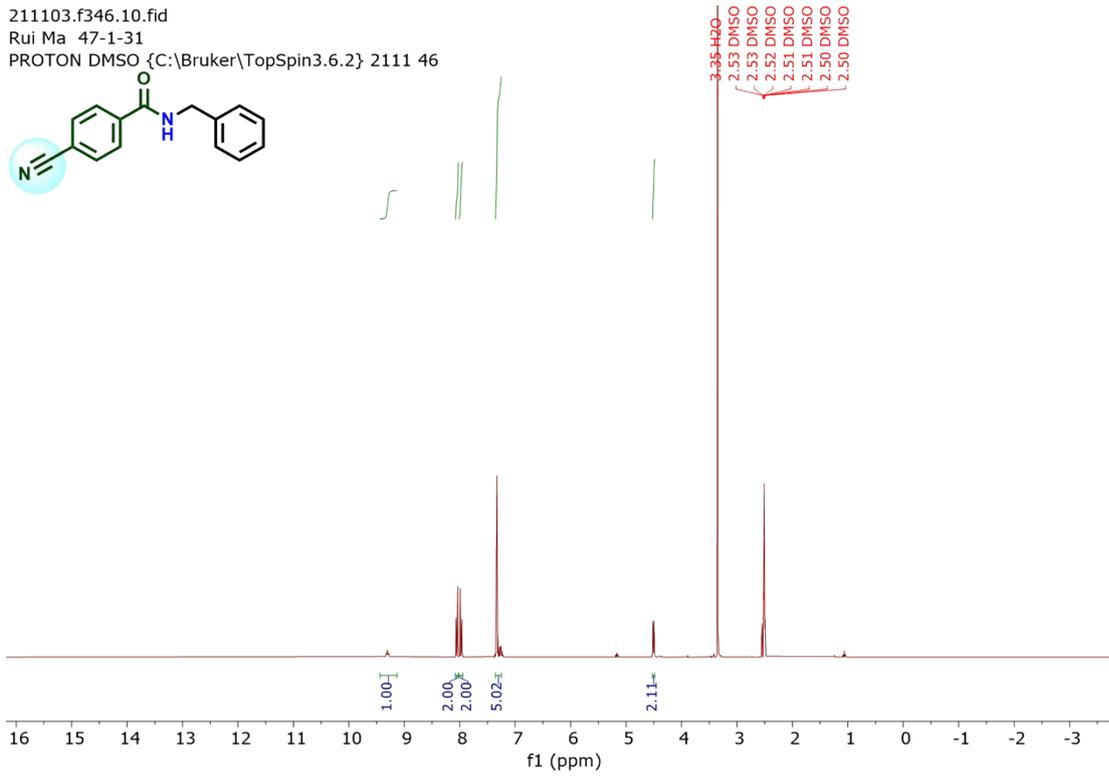
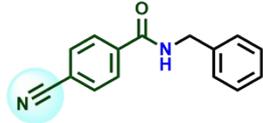
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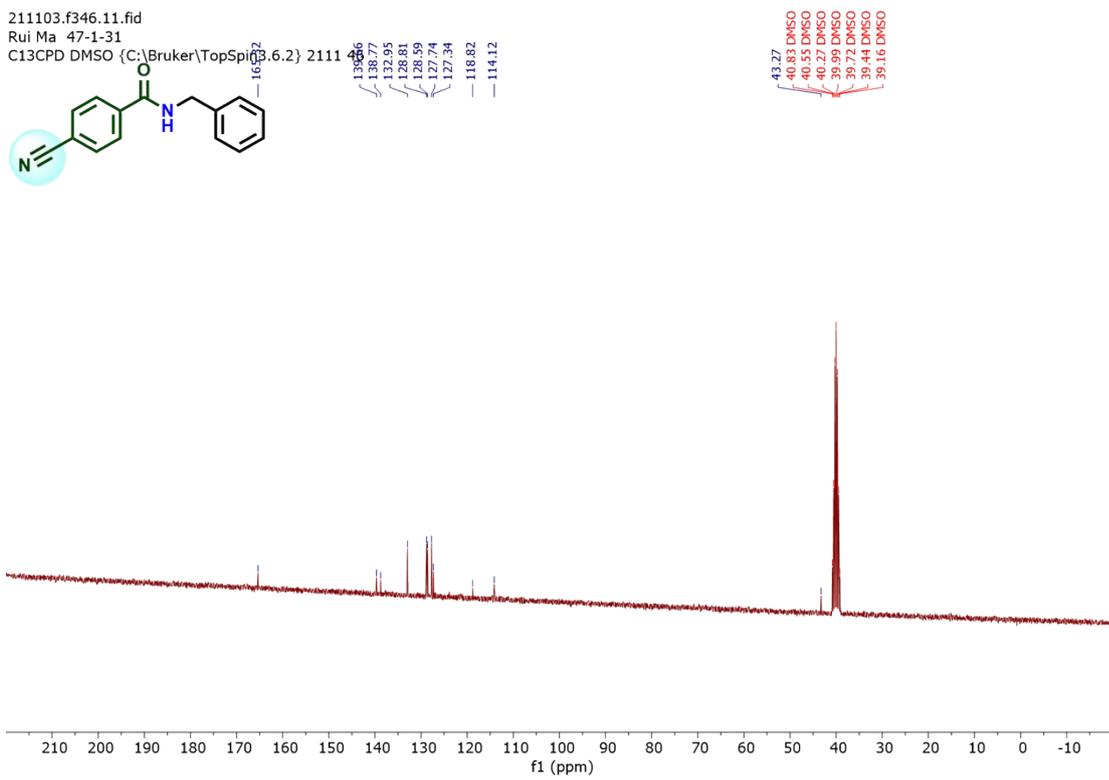
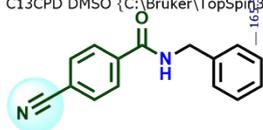
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C13CPD DMSO {C:\Bruker\TopSpin3.6.2} 2111 48



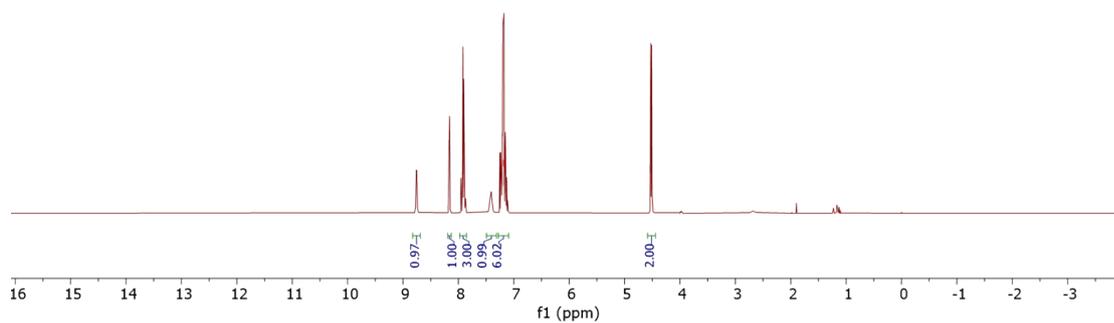
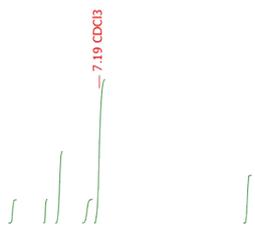
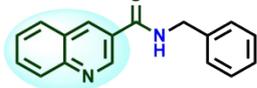
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PROTON DMSO {C:\Bruker\TopSpin3.6.2} 2111 46



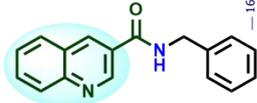
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PROTON CDCl3 {C:\Bruker\TopSpin3.6.2} 2206 33



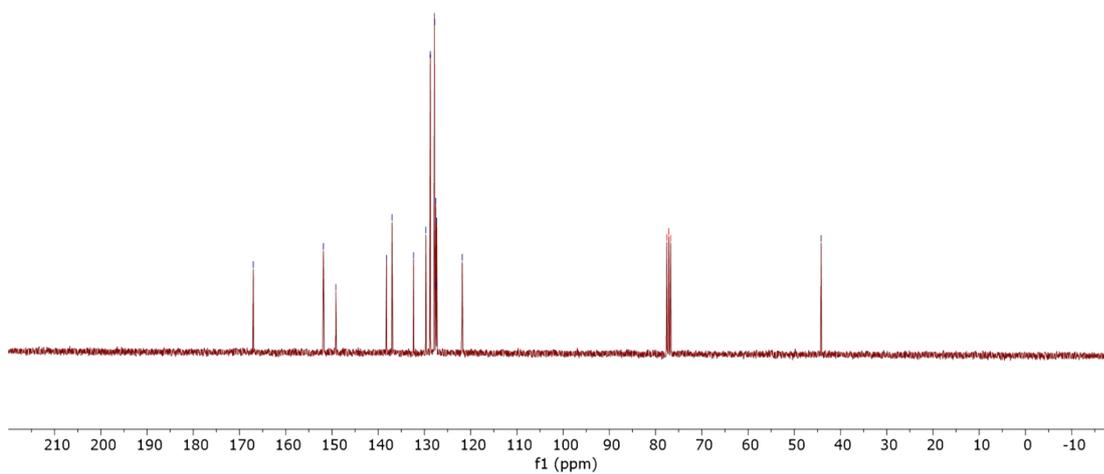
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14.68
13.80
13.58
132.35
129.71
128.71
127.81
127.54
127.48
127.35
121.82

77.58 CDCl3
77.16 CDCl3
76.74 CDCl3

44.19



8. References

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