

Electronic Supporting Information

Preparation of Reusable Copper-Based Biomass-Carbon Aerogel Catalysts and Application in Highly Selective Reduction of Maleimides to Succinimides with Hydrosilane as Hydrogen Source

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I. General remarks

All reagents were obtained from commercial sources and used as received. Technical grade petroleum ether (40-60°C bp.) and ethyl acetate were used for chromatography column. Chitin, potassium hydroxide, urea, tert-butanol, and copper trifluoromethane sulfonate ($\text{Cu}(\text{OTf})_2$) were purchased from Aladdin Biochemical Technology Co., Ltd. (Shanghai, China).

SEM images of $\text{Cu}(\text{OTf})_2@\text{CAC}$ were taken with an FE-SEM (Sigma 500, Zeiss, Germany) operating at an acceleration voltage of 5 kV. Optical microscope picture was recorded on a polarizing microscope (DM2700P, Leica, Germany). TEM images of the super-thin sliced $\text{Cu}(\text{OTf})_2@\text{CAC}$ were taken with a transmission electron microscopy (JEM-2100, JEOL, Japan) operating at an accelerating voltage of 200 kV. The partial carbonization process on the chitin/ Cu^{2+} composite aerogels was carried out on a thermogravimetric analysis system (Q50, TA, America) at a heating rate of $10\text{ }^\circ\text{C min}^{-1}$ from 25 to 250 $^\circ\text{C}$, followed by a constant temperature at 250 $^\circ\text{C}$ for 4 h under the nitrogen atmosphere. The Brunauer-Emmett-Teller (BET) surface areas were measured and calculated by the nitrogen sorption-desorption isotherms using an adsorption analyzer (ASAP 2020, Micromeritics, USA). 2D Raman imaging was performed using a laser confocal Raman imaging microscope (inVia, Renishaw, UK). The wavelength of the excitation laser was 532 nm. Raman maps (scan range, $50\text{ }\mu\text{m} \times 50\text{ }\mu\text{m}$) were collected using a spatial resolution of 200 nm. The direct classical least squares (DCLS) method developed by accessory software was applied for imaging the chitin and the amorphous carbon domains. X-ray photoelectron spectra was conducted on an X-ray photoelectron spectrometer (ESCALAB250Xi, Thermo Fisher Scientific, America) with monochromatic Al target test, and full spectrum passing energy 100 eV.

^1H NMR spectra were recorded in CDCl_3 at ambient temperature on Bruker AVANCE I 500 spectrometers at 500.1 MHz, using the solvent as internal standard (7.26 ppm). ^{13}C NMR spectra were obtained at 125 MHz and referenced to the internal solvent signals (central peak is 77.2 ppm). Chemical shift (δ) and coupling constants (J) are given in ppm and in Hz, respectively. The peak patterns are indicated as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet, and br. for broad.

GC analyses were performed with GC-7890A (Agilent) equipped with a 30-m capillary column (HP-5ms, fused silica capillary column, 30 M*0.25 mm*0.25 mm film thickness), was used with N_2/air as vector gas. GCMS were measured by GCMS-7890A-5975C (Agilent) with GC-7890A equipped with a 30-m capillary column (HP-5ms, fused silica capillary column, 30 M*0.25 mm*0.25 mm film thickness), was used with helium as vector gas. HRMS were measured by MAT 95XP (Termol) (LCMS-IT-TOF).

II. Supplementary information of $\text{Cu}(\text{OTf})_2@ \text{CAC}$

2.1 Chitin powder purification

First, 400 g crab shells were crushed, dispersed in the 4 L, 4 wt.% NaOH solution, and stirred for 8h to remove proteins. Then, the substance was successively dispersed in 4 L, 0.3 wt.% Na_2ClO (8 h, stirring, 80 °C) and the pH level of the mixture was constantly maintained at by adding 7 wt.% HCl to remove pigments and minerals. After repeating the above steps twice, the chitin powders were obtained by drying at 50 °C (Fig S1).



Figure S1. The crab shells after purification are pale yellow chitin powder.

2.2 Chitin hydrogel and chitin aerogel preparation

First, 5.26 g chitin powders were added slowly to the 100 g 20 wt.% KOH/4 wt.% Urea solution while stirring in the trap at -40 °C for 2h. the turbid mixture turned into yellow viscous chitin solution after centrifugation at 10000 rpm, 0 °C (Fig S2). Then, the chitin solution was poured and cast into the homemade mold. The chitin gel was formed by immersing the mold in 1 L absolute alcohol at -25 °C for 24h (Fig S3). Finally, the transparent chitin hydrogel was cut into 2×2 cm chitin gel pieces.

The chitin aerogels were fabricated by freezing-drying. Specifically, the chitin hydrogel was replaced solvent with 50 mL tert-butanol twice. Then, the gels were immersed in a box filled with liquid nitrogen for 15min and displace to freeze-drying (Christ Alpha 2-4 LDplus, -80 °C, 0.045 atm) for 12h to obtain the white and lightweight chitin aerogel.

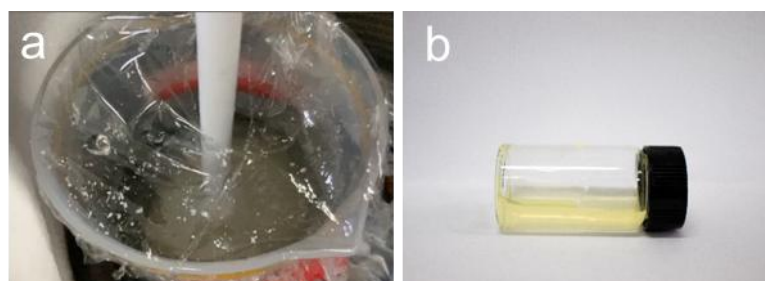


Figure S2. a) the process of the crab shells dissolution. b) the chitin solution with excellent fluidity.

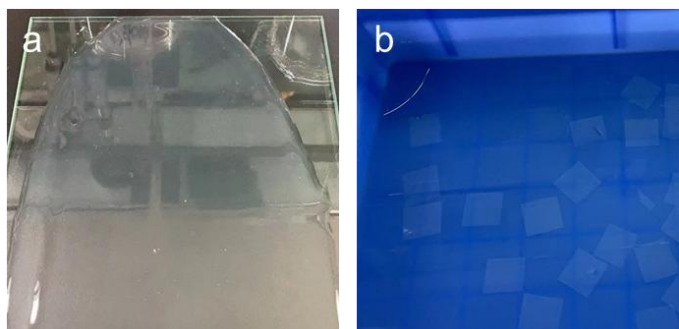


Figure S3. a) the chitin solution was poured into the mold through casting method. b) the chitin hydrogel with 3D nanofibrous network.

2.3 Cu(OTf)₂@CAC with different pyrolysis times preparation

First, 16 pieces of the chitin gel were replaced solvent with 50 mL acetone to obtain chitin organogels and immersed into 50 mL, 5 wt.% Cu(OTf)₂ acetone solution and stirred for 5 h (Fig S4). Subsequently, the chitin/Cu gels were washed and replaced with 50 mL acetone and 50 mL tert-butanol for 24 h, respectively. Then the chitin/Cu gels were immersed in a box filled with liquid nitrogen and the chitin/Cu aerogels were obtained by freeze-drying (Christ Alpha 2-4 LDplus, -80 °C, 0.045 atm). Finally, 120 mg of the chitin/Cu aerogels were carbonized under a nitrogen atmosphere at 250 °C (heating rate is 5 °C/min and cooling rate is 10 °C/min) for 1h to obtain Cu(OTf)₂@CAC-1h. Whatmore, 120 mg of the chitin/Cu aerogels were carbonized under a nitrogen atmosphere at 250 °C (heating rate is 5 °C/min and cooling rate is 10 °C/min) for 3h to obtain Cu(OTf)₂@CAC-3h, and 120 mg of the chitin/Cu aerogels were carbonized for 4h as the same procedure to obtain Cu(OTf)₂@CAC-4h. All Cu(OTf)₂@CAC-1h, Cu(OTf)₂@CAC-3h and Cu(OTf)₂@CAC-4h were washed by 10 mL acetone for 12h and dried at 50 °C to remove impurities.



Figure S4. the process of the chitin organogel chelated with Cu²⁺ in the Cu(OTf)₂-containing acetone solution

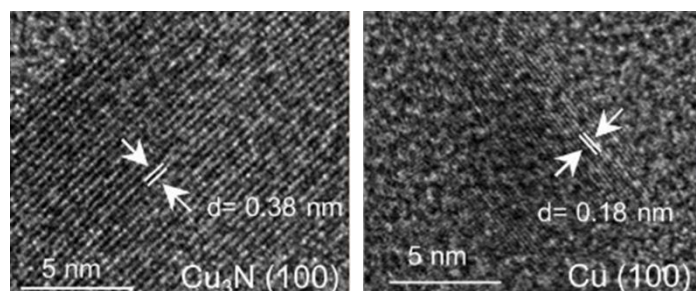


Figure S5. HRTEM image of $\text{Cu}(\text{OTf})_2@CAC$ with obvious lattice spacing.

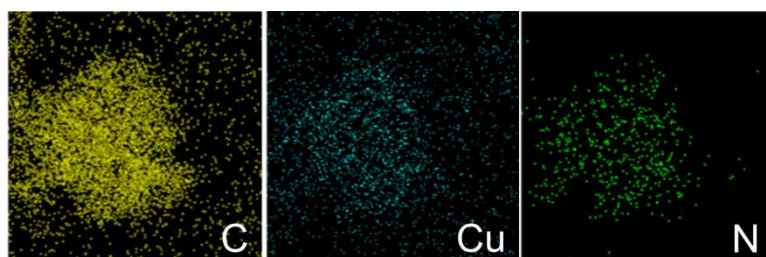


Figure S6. EDS test of the C, N, and Cu elements inside the $\text{Cu}(\text{OTf})_2@CAC$.

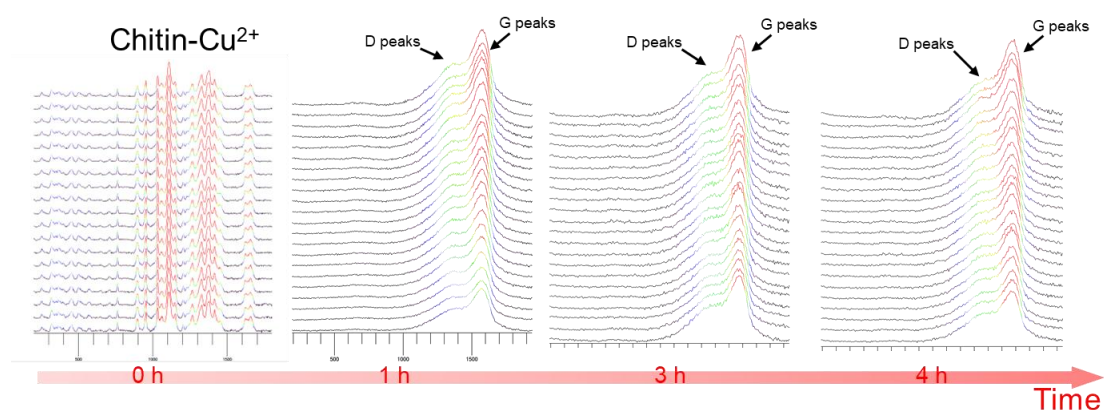


Figure S7. Representative Raman spectra of the composite aerogel treated with different pyrolysis times.

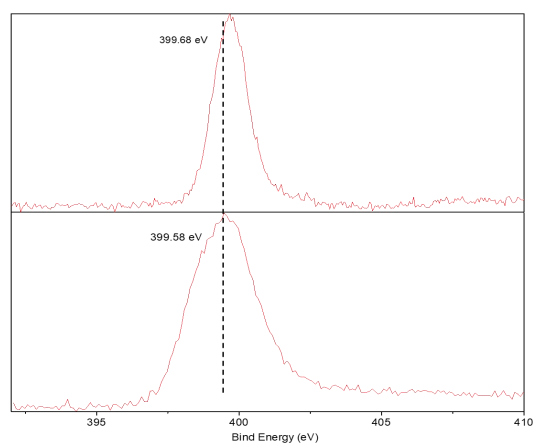


Figure S8. N 1s spectra of composite carbon aerogels, chitin/Cu aerogels

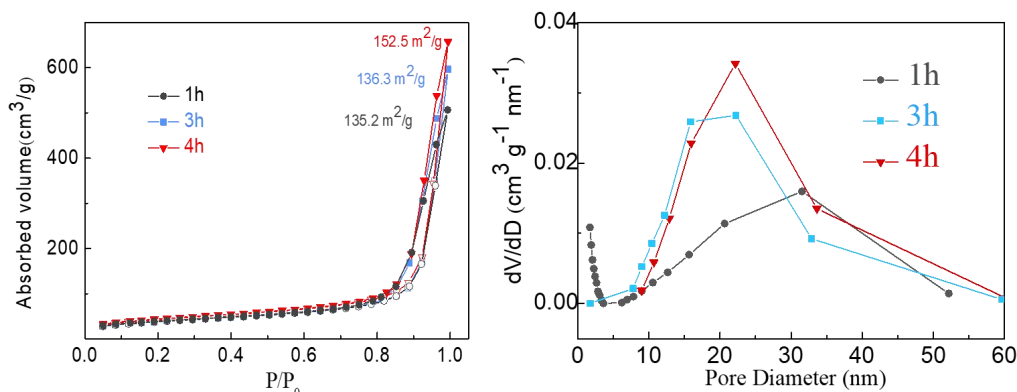


Figure S9. Nitrogen adsorption-desorption isotherms and pore size distributions of aerogel with different carbonization treatment times

2.4 Cu⁽⁰⁾@CAC and Cu^(I)(OTf)^(I)@CAC with different pyrolysis times preparation

First, 1 g chitin aerogels were carbonized under a nitrogen atmosphere at 250 °C (heating rate is 5 °C/min and cooling rate is 10 °C/min) for 1h to obtain chitin carbon aerogel (CAC)-1h, and 1 g chitin aerogels were carbonized under a nitrogen atmosphere at 250 °C (heating rate is 5 °C/min and cooling rate is 10 °C/min) for 3h to obtain CAC-3h, then, at the same procedure, chitin aerogels were carbonized for 4h to obtain CAC-4h.

Subsequently, 500 mg CAC-1h was mixed with 5 mg Cu⁽⁰⁾ powder and Cu^(I)(OTf) powder by grinding thoroughly to fabricate Cu⁽⁰⁾@chitin carbon aerogel catalyst (Cu⁽⁰⁾@CAC-1h) and Cu^(I)@CAC-1h. Whatmore, CAC-3h and CAC-4h were treated as the same procedure to obtain Cu⁽⁰⁾@CAC-3h, Cu^(I)(OTf)^(I)@CAC-3h, and Cu⁽⁰⁾@CAC-4h, Cu^(I)(OTf)^(I)@CAC-4h, respectively.

2.5 RuCl₃@CAC preparation

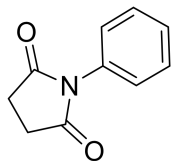
First, 500 mg of the chitin gel were replaced solvent with 50 mL acetone to obtain chitin organogels which immersed into 50 mL acetone solution with 8.93 mg of the RuCl₃ powder, while stirring for 5h. Subsequently, the chitin/Ru gels were washed and replaced with 50 mL acetone and 50 mL tert-butanol for 24h, respectively. Then the chitin/Ru gels were immersed in a box filled with liquid nitrogen and the chitin/Ru aerogels were obtained by freeze-drying (Christ Alpha 2-4 LDplus, -80 °C, 0.045 atm). Finally, 120 mg of the chitin/Cu aerogels were carbonized under a nitrogen atmosphere at 250 °C (heating rate is 5 °C/min and cooling rate is 10 °C/min) for 4h to obtain RuCl₃@chitin carbon aerogel catalyst (RuCl₃@CAC).

III. General procedures for Cu(OTf)₂@CAC-catalyzed reduction of maleimides

Cu(OTf)₂@CAC (1.2 mol%, 20 mg), maleimide (0.25 mmol), PhSiH₃ (0.5 mmol, 62 μ L), and THF (1 mL) were introduced in tube under air, equipped with magnetic stirring bar and was stirred at 60 °C. After 12 h, the conversion of the reaction was analyzed by gas chromatography. The solvent was then evaporated under vacuum and the desired product was purified by using a silica gel column chromatography and a mixture of petrol ether/ethyl acetate as eluent.

IV. Characterization data for all compounds

1-phenylpyrrolidine-2,5-dione¹ (3a)

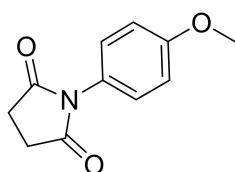


White solid, yield = 82%, 36 mg

¹H NMR (500 MHz, CDCl₃): δ = 7.521-7.516 (m, 2H), 7.44-7.41 (m, 1H), 7.32-7.29 (m, 2H), 2.91 (s, 4H).

¹³C NMR (125 MHz, CDCl₃): δ = 176.3, 131.9, 129.3, 128.7, 126.5, 28.5.

1-(4-methoxyphenyl)pyrrolidine-2,5-dione¹ (3b)

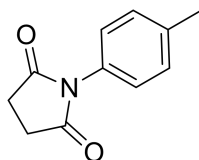


Light yellow solid, yield = 75%, 38 mg

¹H NMR (500 MHz, CDCl₃): δ = 7.20 (d, 2H, *J* = 9.0 Hz), 7.00 (d, 2H, *J* = 9.0 Hz), 3.83 (s, 3H), 2.88 (s, 4H).

¹³C NMR (125 MHz, CDCl₃): δ = 176.6, 159.5, 127.7, 124.5, 114.6, 55.5, 28.4.

1-(p-tolyl)pyrrolidine-2,5-dione¹ (3c)

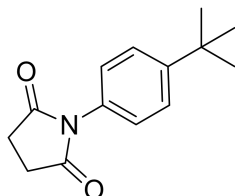


White solid, yield = 80%, 38 mg.

¹H NMR (500 MHz, DMSO): δ = 7.29 (d, 2H, *J* = 8.0 Hz), 7.14 (d, 2H, *J* = 8.0 Hz), 2.77 (s, 4H), 2.35 (s, 3H).

¹³C NMR (125 MHz, DMSO): δ = 177.5, 138.1, 130.6, 129.7, 127.4, 28.9, 21.2.

1-(4-(tert-butyl)phenyl)pyrrolidine-2,5-dione² (3d)

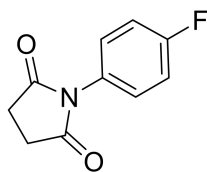


White solid, yield = 86%, 50 mg.

¹H NMR (500 MHz, CDCl₃): δ = 7.51 (d, 2H, *J* = 8.5 Hz), 7.22 (d, 2H, *J* = 8.5 Hz), 2.89 (s, 4H), 1.35 (s, 9H).

¹³C NMR (125 MHz, CDCl₃): δ = 176.5, 151.7, 129.1, 126.3, 125.9, 34.8, 31.3, 28.4.

1-(4-fluorophenyl)pyrrolidine-2,5-dione³ (3e)



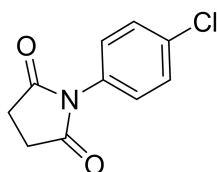
White solid, yield = 92%, 44 mg.

¹H NMR (500 MHz, CDCl₃): δ = 7.32-7.29 (m, 2H), 7.21-7.17 (m, 2H), 2.93 (s, 4H).

¹³C NMR (125 MHz, CDCl₃): δ = 176.1, 162.2 (d, *J*_{CF} = 247.50 Hz), 128.3 (d, *J*_{CF} = 8.75 Hz), 127.8 (d, *J*_{CF} = 3.38 Hz), 116.3 (d, *J*_{CF} = 22.75 Hz), 28.4.

¹⁹F NMR (470 MHz, CDCl₃): δ = -112.2.

1-(4-chlorophenyl)pyrrolidine-2,5-dione¹ (3f)

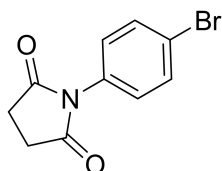


White solid, yield = 75%, 39 mg.

¹H NMR (500 MHz, DMSO): δ = 7.57 (d, 2H, *J* = 8.5 Hz), 7.31 (d, 2H, *J* = 9.0 Hz), 2.92 (s, 4H).

¹³C NMR (125 MHz, DMSO): δ = 177.2, 133.0, 132.0, 129.35, 129.33, 29.0.

1-(4-bromophenyl)pyrrolidine-2,5-dione¹ (3g)

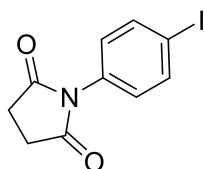


White solid, yield = 74%, 47 mg.

¹H NMR (500 MHz, CDCl₃): δ = 7.62 (d, 2H, *J* = 8.5 Hz), 7.22 (d, 2H, *J* = 8.5 Hz), 2.92-2.90 (m, 4H).

¹³C NMR (125 MHz, CDCl₃): δ = 169.1, 134.3, 132.3, 130.3, 127.4, 121.6.

1-(4-iodophenyl)pyrrolidine-2,5-dione⁴ (3h)

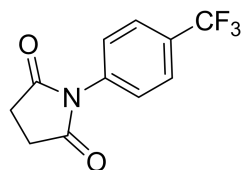


White solid, yield = 55%, 42 mg.

¹H NMR (500 MHz, CDCl₃): δ = 7.83 (d, 2H, *J* = 8.5 Hz), 7.09 (d, 2H, *J* = 8.5 Hz), 2.92 (s, 4H).

^{13}C NMR (125 MHz, CDCl_3): $\delta = 175.8, 138.4, 131.6, 128.1, 94.1, 28.4$.

1-(4-(trifluoromethyl)phenyl)pyrrolidine-2,5-dione⁴ (3i)

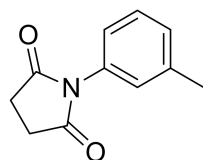


White solid, yield = 40%, 25 mg.

^1H NMR (500 MHz, CDCl_3): $\delta = 7.75$ (d, 2H, $J = 8.5$ Hz), 7.48 (d, 2H, $J = 8.0$ Hz), 2.94 (s, 4H).

^{13}C NMR (125 MHz, CDCl_3): $\delta = 175.8, 126.8, 126.5, 126.4, 28.6$.

1-(*m*-tolyl)pyrrolidine-2,5-dione⁵ (3j)

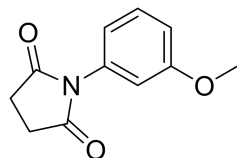


White solid, yield = 90%, 43 mg.

^1H NMR (500 MHz, CDCl_3): $\delta = 7.39$ - 7.36 (m, 1H), 7.23 (d, 1H, $J = 7.5$ Hz), 7.10 - 7.07 (m, 2H), 2.88 (s, 4H), 2.41 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3): $\delta = 176.4, 139.3, 129.6, 129.1, 127.2, 123.6, 28.5, 21.4$.

1-(3-methoxyphenyl)pyrrolidine-2,5-dione⁶ (3k)

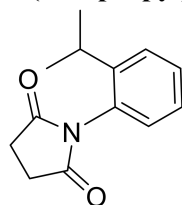


White solid, yield = 79%, 41 mg.

^1H NMR (500 MHz, CDCl_3): $\delta = 7.40$ (t, 1H, $J = 8.0$ Hz), 6.97 - 6.83 (m, 3H), 3.83 (s, 3H), 2.90 (s, 4H).

^{13}C NMR (125 MHz, CDCl_3): $\delta = 176.2, 160.1, 132.9, 130.0, 118.8, 114.6, 112.4, 55.4, 28.5$.

1-(2-isopropylphenyl)pyrrolidine-2,5-dione⁷ (3l)

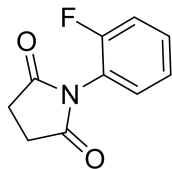


White solid, yield = 72%, 39 mg.

^1H NMR (500 MHz, CDCl_3): $\delta = 7.47$ - 7.45 (m, 2H), 7.33 - 7.30 (m, 1H), 7.04 - 7.02 (m, 1H), 2.95 - 2.94 (m, 4H), 2.71 - 2.65 (m, 1H), 1.22 (s, 3H), 1.21 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3): δ = 176.8, 146.2, 130.1, 129.7, 128.2, 127.0, 126.8, 28.7, 28.6, 23.7.

1-(2-fluorophenyl)pyrrolidine-2,5-dione (3m)



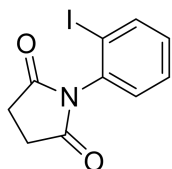
White solid, yield = 85%, 41 mg.

^1H NMR (500 MHz, CDCl_3): δ = 7.46-7.43 (m, 1H), 7.29-7.23 (m, 3H), 2.95 (d, 4H, J = 9.0 Hz).

^{13}C NMR (125 MHz, CDCl_3): δ = 175.4, 157.4 (d, J_{CF} = 250.88 Hz), 131.1 (d, J_{CF} = 8.13 Hz), 129.3, 124.7 (d, J_{CF} = 4.00 Hz), 119.7 (d, J_{CF} = 13.00 Hz), 116.8 (d, J_{CF} = 19.63 Hz), 28.6.

^{19}F NMR (470 MHz, CDCl_3): δ = -119.4.

1-(2-iodophenyl)pyrrolidine-2,5-dione³ (3n)

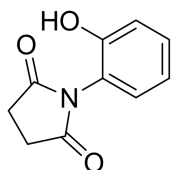


White solid, yield = 66%, 50 mg.

^1H NMR (500 MHz, CDCl_3): δ = 7.97 (dd, 1H, J = 8.0, 1.5 Hz), 7.51-7.48 (m, 1H), 7.22-7.18 (m, 2H), 3.03-2.91 (m, 4H).

^{13}C NMR (125 MHz, CDCl_3): δ = 175.4, 140.0, 135.4, 131.1, 129.6, 129.4, 97.9, 28.9.

1-(2-hydroxyphenyl)pyrrolidine-2,5-dione⁸ (3o)

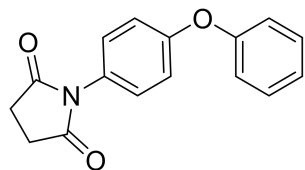


White solid, yield = 87%, 42 mg.

^1H NMR (500 MHz, CDCl_3): δ = 7.31-7.29 (m, 1H), 7.15 (dd, 1H, J = 8.0, 1.5 Hz), 7.05-7.02 (m, 1H), 6.99 (dd, 1H, J = 8.0, 1.5 Hz), 2.88 (s, 4H), 1.28 (s, 1H).

^{13}C NMR (125 MHz, CDCl_3): δ = 177.2, 151.2, 130.7, 128.3, 121.2, 119.8, 118.6, 28.6.

1-(4-phenoxyphenyl)pyrrolidine-2,5-dione (3p)

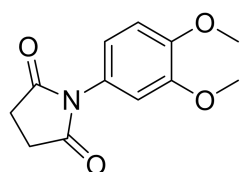


White solid, yield = 67%, 45 mg.

^1H NMR (500 MHz, CDCl_3): δ = 7.41-7.36 (m, 2H), 7.25 (d, 2H, J = 9.0 Hz), 7.17 (t, 1H, J = 7.5 Hz), 7.11-7.07 (m, 4H), 2.78 (s, 4H).

^{13}C NMR (125 MHz, CDCl_3): δ = 176.4, 157.7, 156.3, 129.9, 127.9, 126.4, 124.0, 119.7, 118.8, 28.4.

1-(3,4-dimethoxyphenyl)pyrrolidine-2,5-dione (3q)

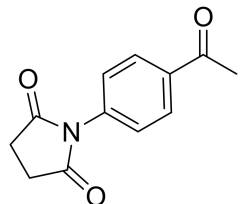


White solid, yield = 85%, 50 mg.

^1H NMR (500 MHz, CDCl_3): δ = 6.95 (d, 1H, J = 8.5 Hz), 6.84 (dd, 1H, J = 8.5, 2.5 Hz), 6.78 (d, 1H, J = 2.5 Hz), 3.91 (s, 3H), 3.88 (s, 3H), 2.90 (s, 4H).

^{13}C NMR (125 MHz, CDCl_3): δ = 176.5, 149.3, 149.2, 124.6, 119.1, 111.2, 109.9, 56.1, 56.0, 28.4.

1-(4-acetylphenyl)pyrrolidine-2,5-dione⁹ (3r)

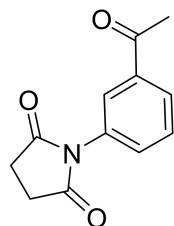


White solid, yield = 83%, 45 mg.

^1H NMR (500 MHz, CDCl_3): δ = 8.08 (d, 2H, J = 9.0 Hz), 7.47 (d, 2H, J = 8.5 Hz), 2.95 (s, 4H), 2.64 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3): δ = 197.1, 175.7, 136.7, 135.9, 129.2, 126.4, 28.5, 26.7.

1-(3-acetylphenyl)pyrrolidine-2,5-dione (3s)

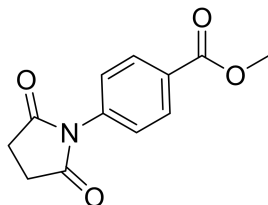


White solid, yield = 80%, 43 mg.

^1H NMR (500 MHz, CDCl_3): δ = 8.01 (d, 1H, J = 7.5 Hz), 7.92 (t, 1H, J = 2.0 Hz), 7.62 (t, 1H, J = 8.0 Hz), 7.54-7.52 (m, 1H), 2.96 (s, 4H), 2.64 (s, 3H).

^{13}C NMR (125 MHz, CDCl_3): δ = 196.9, 175.9, 138.1, 132.4, 131.0, 129.6, 128.4, 126.5, 28.5, 26.7.

methyl 4-(2,5-dioxopyrrolidin-1-yl)benzoate¹⁰ (3t)

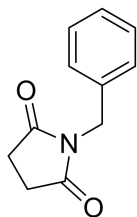


White solid, yield = 69%, 40 mg.

^1H NMR (500 MHz, CDCl_3): δ = 8.17 (d, 2H, J = 8.5 Hz), 7.44 (d, 2H, J = 8.5 Hz), 3.95 (s, 3H), 2.94 (s, 4H).

^{13}C NMR (125 MHz, CDCl_3): δ = 175.7, 166.2, 135.9, 130.49, 130.04, 126.2, 52.4, 28.4.

1-benzylpyrrolidine-2,5-dione³ (3u)

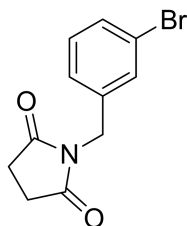


White solid, yield = 74%, 35 mg.

^1H NMR (500 MHz, CDCl_3): δ = 7.42-7.40 (m, 2H), 7.35-7.30 (m, 3H), 4.68 (s, 2H), 2.72 (s, 4H).

^{13}C NMR (125 MHz, CDCl_3): δ = 176.9, 135.8, 128.9, 128.7, 128.0, 42.4, 28.2.

1-(3-bromobenzyl)pyrrolidine-2,5-dione¹¹ (3v)

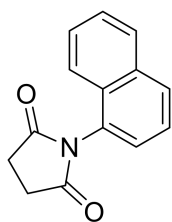


White solid, yield = 83%, 56 mg.

^1H NMR (500 MHz, CDCl_3): δ = 7.55 (s, 1H), 7.43 (d, 1H, J = 8.5 Hz), 7.35 (d, 1H, J = 7.5 Hz), 7.22-7.19 (m, 1H), 4.64 (s, 2H), 2.75 (s, 4H).

^{13}C NMR (125 MHz, CDCl_3): δ = 176.7, 137.8, 131.8, 131.2, 130.2, 127.6, 122.7, 41.8, 28.2.

1-(naphthalen-1-yl)pyrrolidine-2,5-dione (3w)

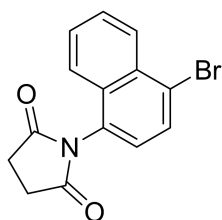


White solid, yield = 71%, 40 mg.

^1H NMR (500 MHz, CDCl_3): δ = 8.00-7.95 (m, 2H), 7.61-7.53 (m, 4H), 7.36 (dd, 1H, J = 7.0, 1.0 Hz), 3.12-2.99 (m, 4H).

^{13}C NMR (125 MHz, CDCl_3): δ = 176.6, 134.5, 130.1, 129.3, 128.8, 128.6, 127.3, 126.7, 126.3, 125.4, 121.8, 28.8.

1-(4-bromonaphthalen-1-yl)pyrrolidine-2,5-dione (3x)

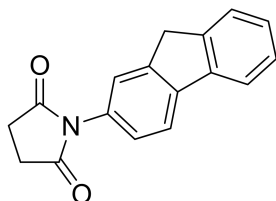


White solid, yield = 51%, 39 mg.

^1H NMR (500 MHz, CDCl_3): δ = 8.35 (d, 1H, J = 8.5 Hz), 7.91 (d, 1H, J = 8.0 Hz), 7.68 (t, 1H, J = 7.5 Hz), 7.60 (t, 1H, J = 7.0 Hz), 7.52 (d, 1H, J = 8.0 Hz), 7.44 (d, 1H, J = 8.0 Hz), 3.14-3.01 (m, 4H).

^{13}C NMR (125 MHz, CDCl_3): δ = 176.3, 132.9, 130.4, 129.5, 128.5, 128.2, 128.1, 128.0, 126.7, 125.0, 122.4, 28.9.

1-(9H-fluoren-2-yl)pyrrolidine-2,5-dione (3y)

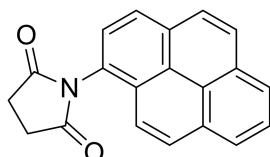


White solid, yield = 79%, 52 mg.

^1H NMR (500 MHz, CDCl_3): δ = 7.89 (d, 1H, J = 8.0 Hz), 7.82 (d, 1H, J = 7.5 Hz), 7.58 (d, 1H, J = 7.5 Hz), 7.48 (s, 1H), 7.43-7.30 (m, 3H), 3.97 (s, 2H), 2.96 (s, 4H).

^{13}C NMR (125 MHz, CDCl_3): δ = 176.5, 144.1, 143.6, 142.3, 140.7, 130.2, 127.3, 126.9, 125.3, 125.1, 123.3, 120.4, 120.3, 37.0, 28.5.

1-(pyren-1-yl)pyrrolidine-2,5-dione (3z)

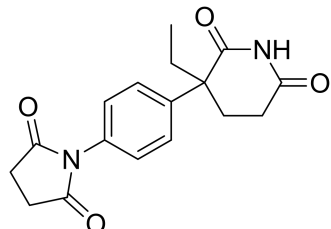


White solid, yield = 61%, 46 mg.

^1H NMR (500 MHz, CDCl_3): δ = 8.30-8.24 (m, 3H), 8.18-8.06 (m, 4H), 7.84 (d, 1H, J = 8.0 Hz), 7.74 (d, 1H, J = 9.5 Hz), 3.22-3.10 (m, 4H).

^{13}C NMR (125 MHz, CDCl_3): δ = 176.7, 132.2, 131.0, 130.7, 129.1, 128.6, 127.4, 127.1, 126.5, 126.2, 126.0, 125.6, 125.4, 125.2, 124.4, 121.1, 28.9.

3-(4-(2,5-dioxopyrrolidin-1-yl)phenyl)-3-ethylpiperidine-2,6-dione (3aa)

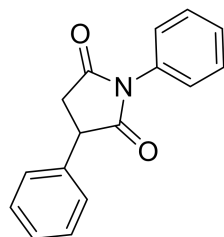


White solid, yield = 54%, 42 mg.

^1H NMR (500 MHz, CDCl_3): δ = 7.42 (d, 2H, J = 8.5 Hz), 7.56 (d, 2H, J = 8.5 Hz), 2.93 (s, 4H), 2.66-2.61 (m, 1H), 2.49-2.39 (m, 2H), 2.31-2.25 (m, 1H), 2.14-2.06 (m, 1H), 1.97-1.90 (m, 1H), 0.91 (t, 3H, J = 7.5 Hz).

^{13}C NMR (125 MHz, CDCl_3): δ = 176.1, 174.1, 171.2, 139.2, 131.3, 127.1, 126.8, 51.0, 33.0, 29.3, 28.4, 26.9, 9.1.

1,3-diphenylpyrrolidine-2,5-dione (3ab)

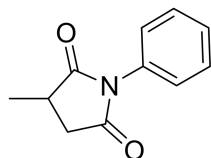


White solid, yield = 61%, 38 mg.

^1H NMR (500 MHz, CDCl_3): δ = 7.53-7.49 (m, 2H), 7.45-7.42 (m, 3H), 7.39-7.33 (m, 5H), 4.23 (dd, 1H, J = 9.5, 5.0 Hz), 3.44-3.38 (m, 1H), 3.04 (dd, 1H, J = 18.5, 5.0 Hz).

^{13}C NMR (125 MHz, CDCl_3): δ = 176.7, 175.2, 137.2, 131.9, 129.3, 129.2, 128.8, 128.1, 127.4, 126.5, 46.0, 37.3.

3-methyl-1-phenylpyrrolidine-2,5-dione (3ac)



White solid, yield = 93%, 44 mg.

^1H NMR (500 MHz, CDCl_3): δ = 7.51-7.48 (m, 2H), 7.43-7.40 (m, 1H), 7.35-7.30 (m, 2H), 3.14-3.02 (m, 2H), 2.55-2.51 (m, 1H), 1.475 (d, 3H, J = 7.0 Hz).

^{13}C NMR (125 MHz, CDCl_3): δ = 179.6, 175.5, 132.0, 129.2, 128.6, 126.5, 36.7, 34.9, 16.9.

V. Reference

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VI. ¹H and ¹³C NMR Spectra for all compounds

1-phenylpyrrolidine-2,5-dione (3a)

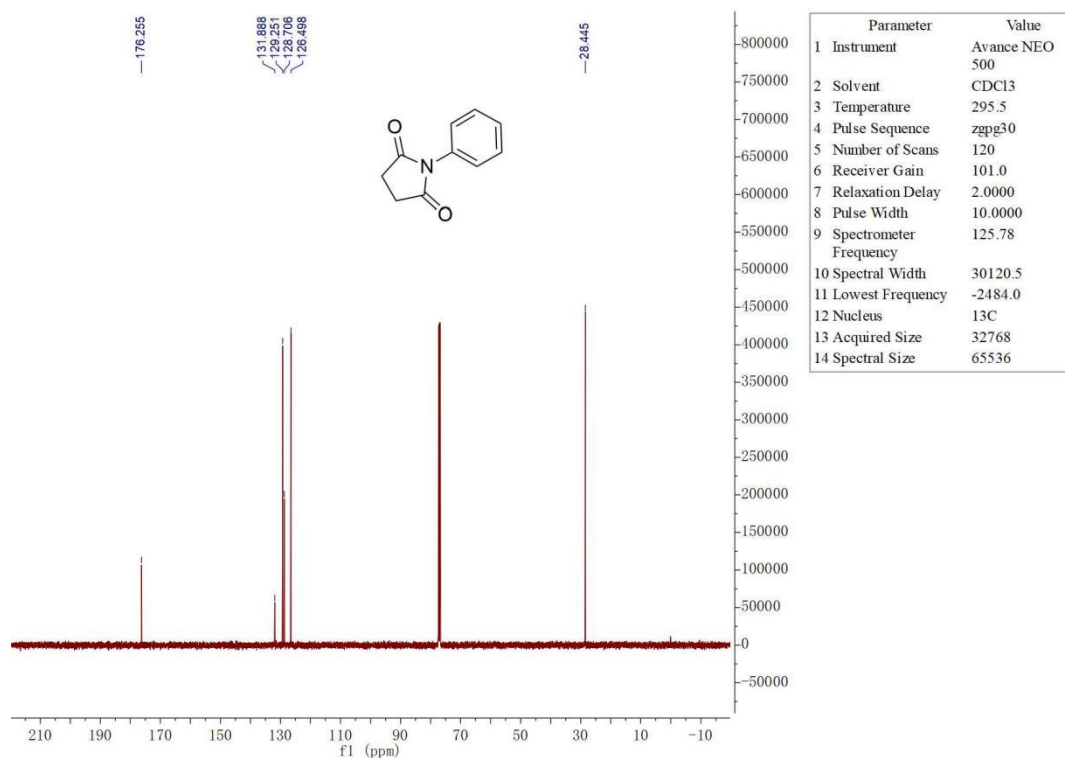
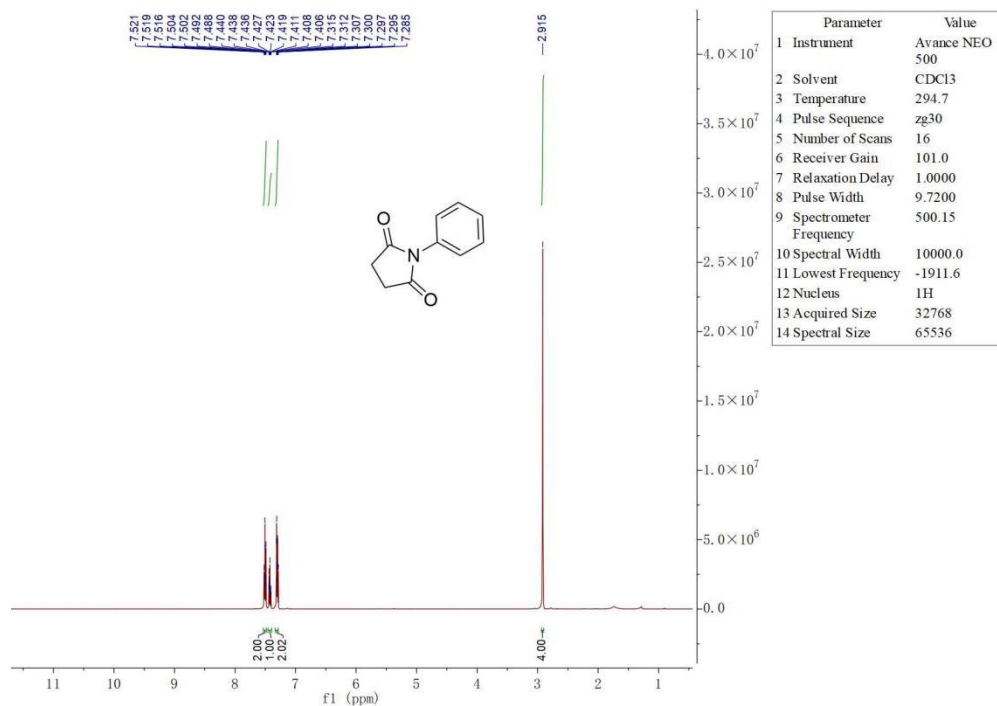
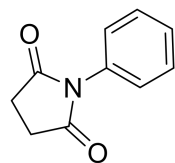


Figure S10. ¹H and ¹³C spectrum of compound 3a.

1-(4-methoxyphenyl)pyrrolidine-2,5-dione (3b)

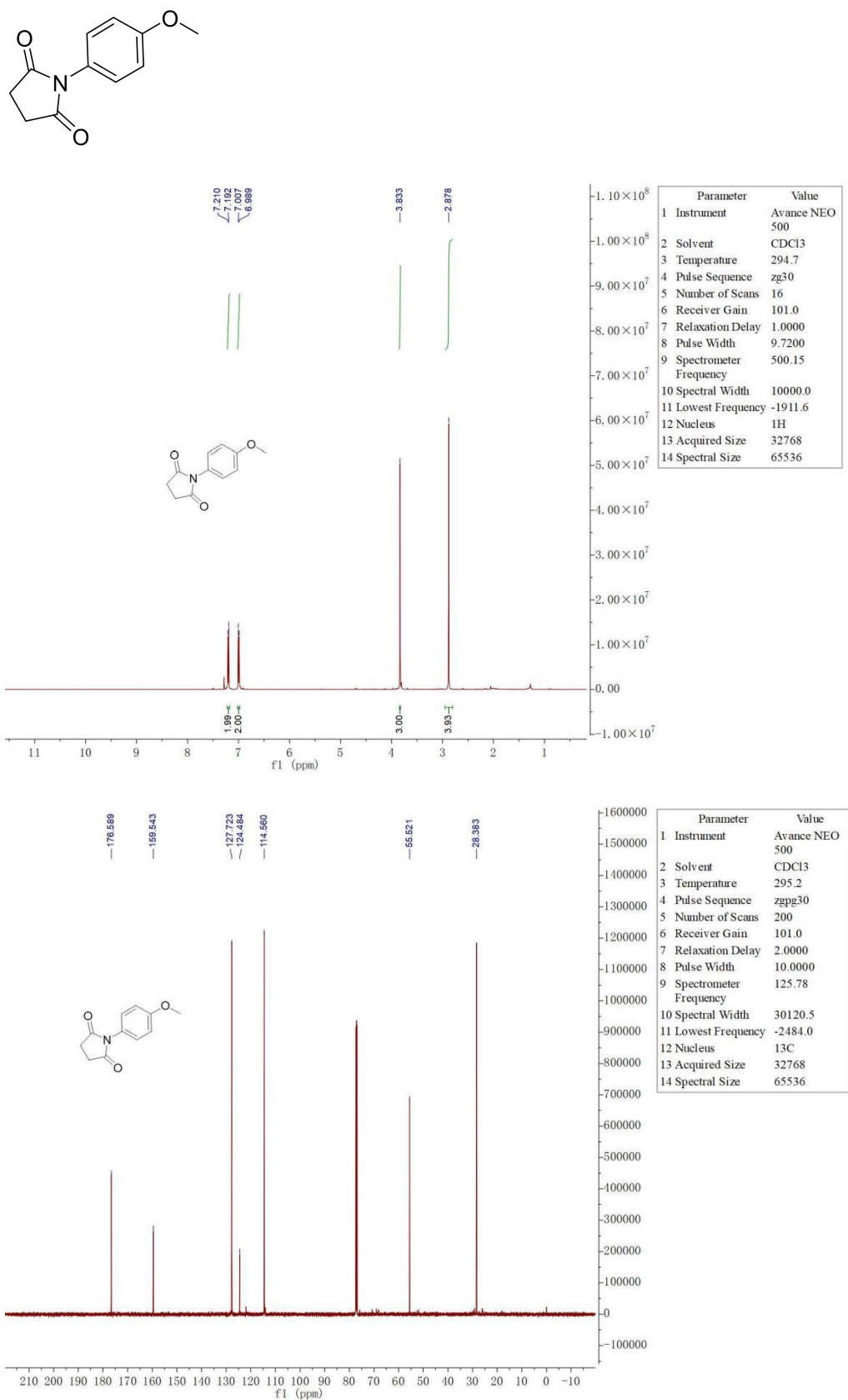


Figure S11. ¹H and ¹³C spectrum of compound 3b.

1-(p-tolyl)pyrrolidine-2,5-dione (3c)

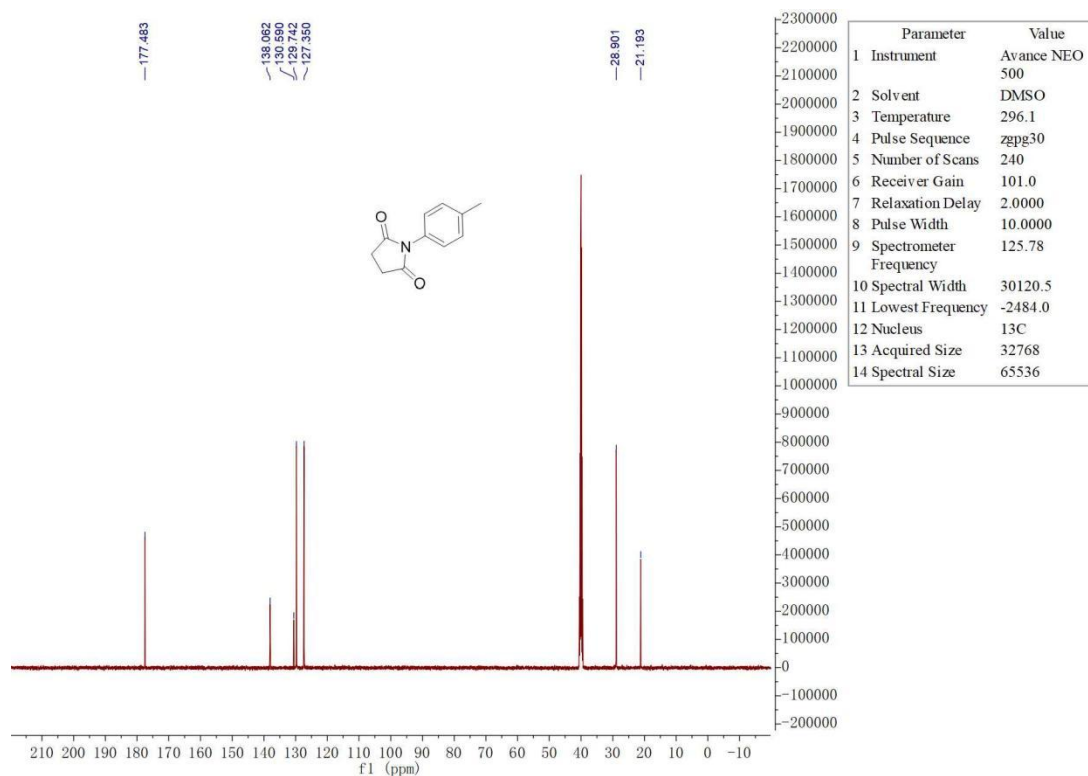
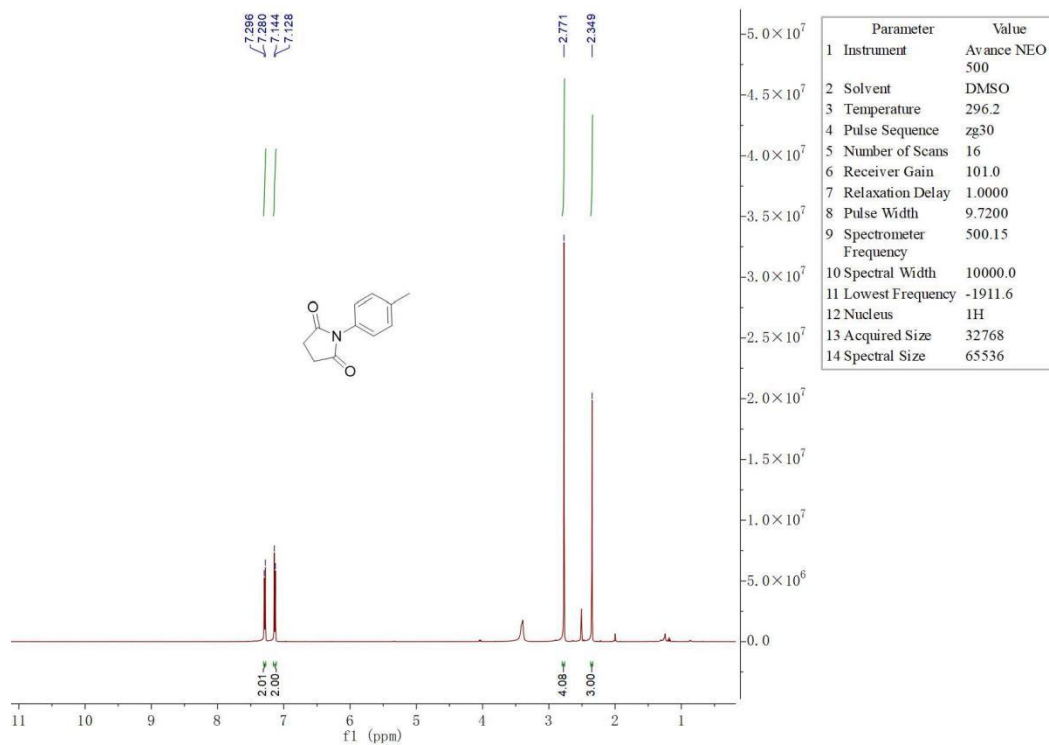
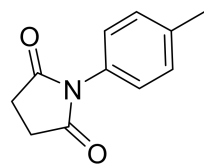


Figure S12. ¹H and ¹³C spectrum of compound 3c.

1-(4-(tert-butyl)phenyl)pyrrolidine-2,5-dione (3d)

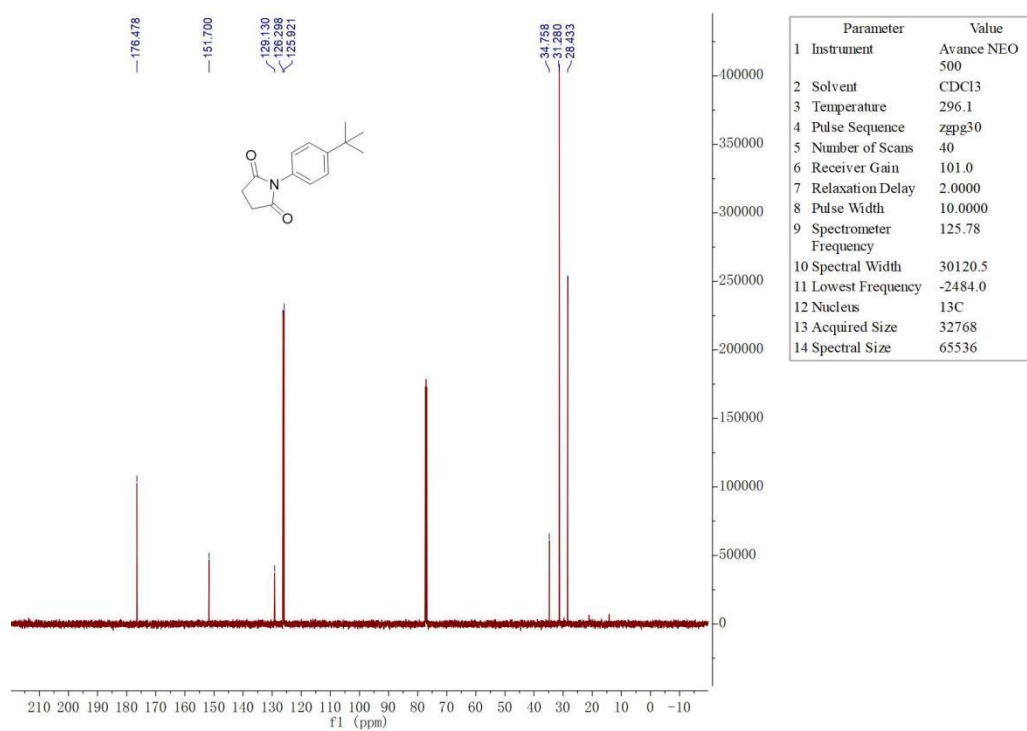
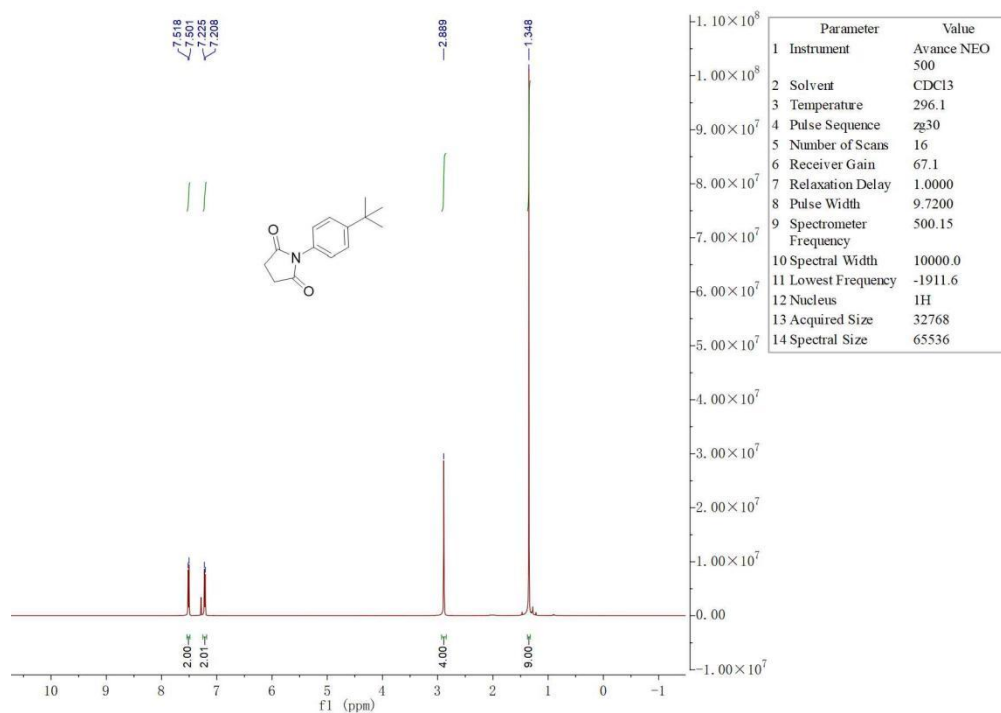
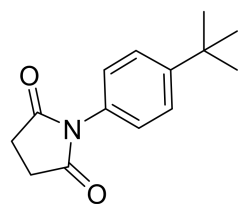


Figure S13. ¹H and ¹³C spectrum of compound 3d.

1-(4-fluorophenyl)pyrrolidine-2,5-dione (3e)

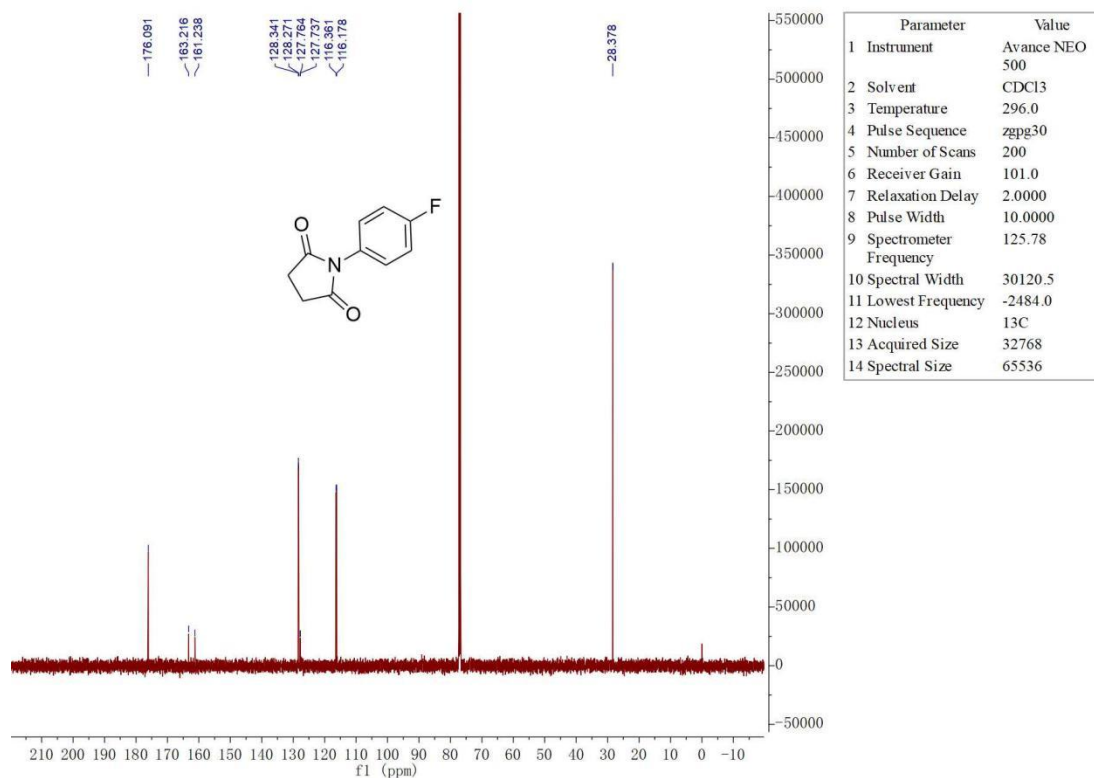
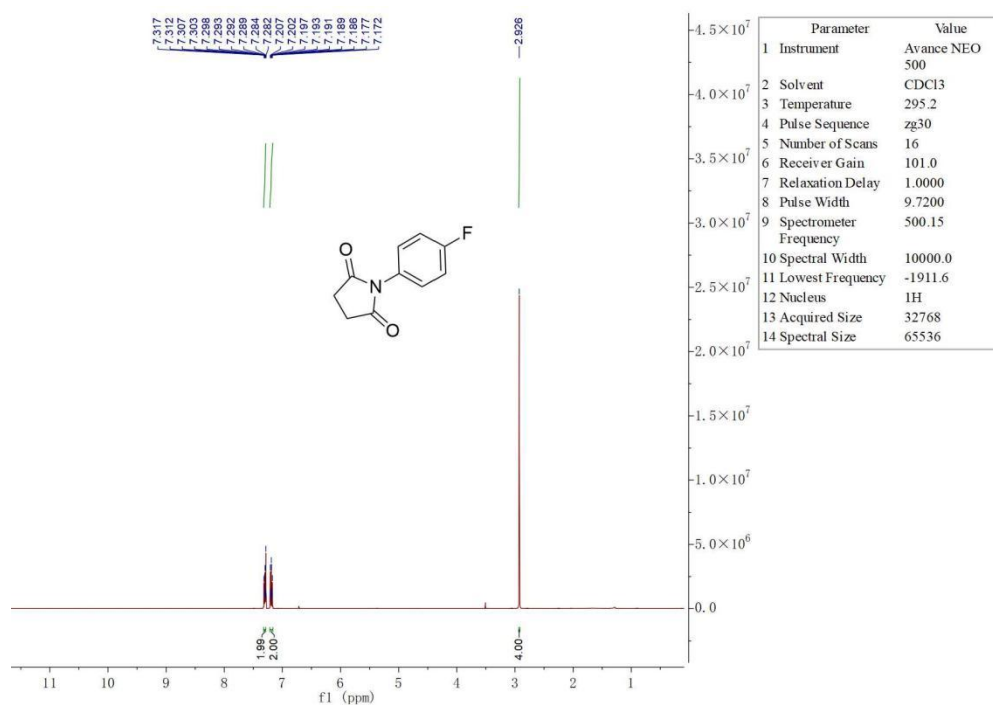
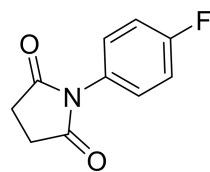


Figure S14. ¹H and ¹³C spectrum of compound 3e.

1-(4-chlorophenyl)pyrrolidine-2,5-dione (3f)

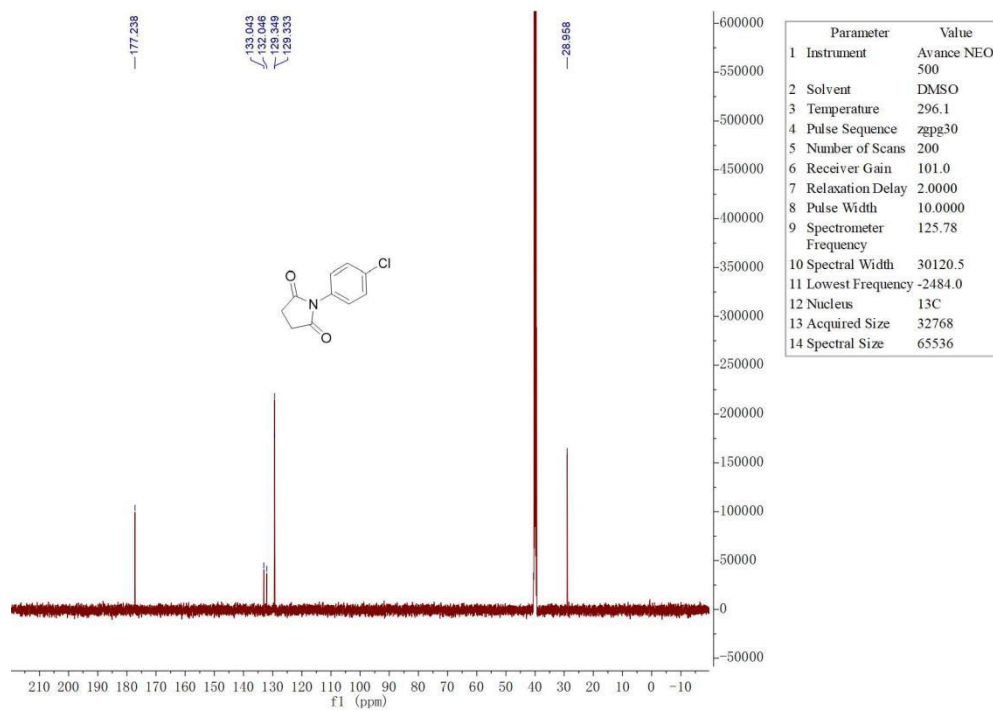
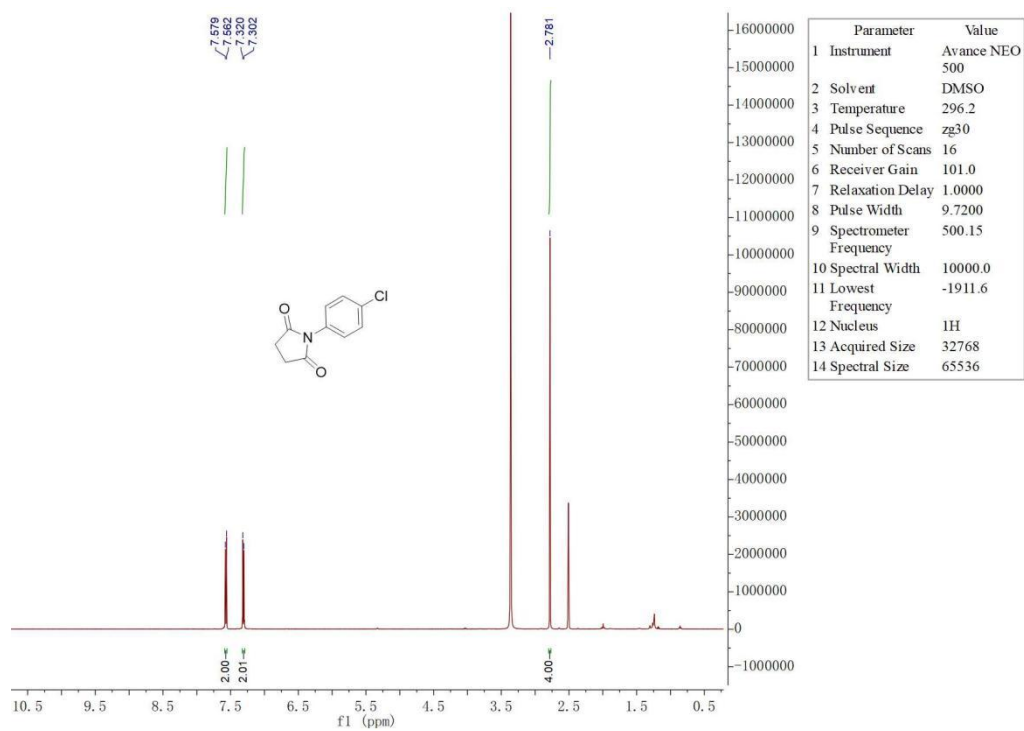
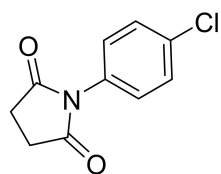


Figure S15. ¹H and ¹³C spectrum of compound 3f.

1-(4-bromophenyl)pyrrolidine-2,5-dione (3g)

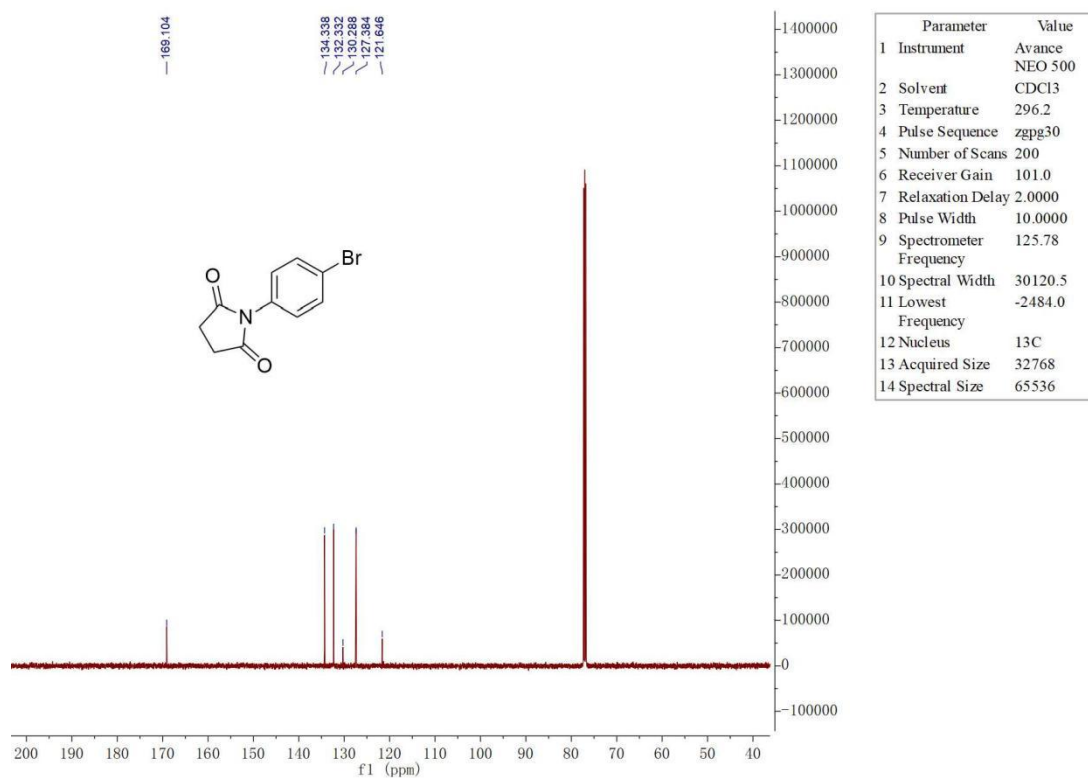
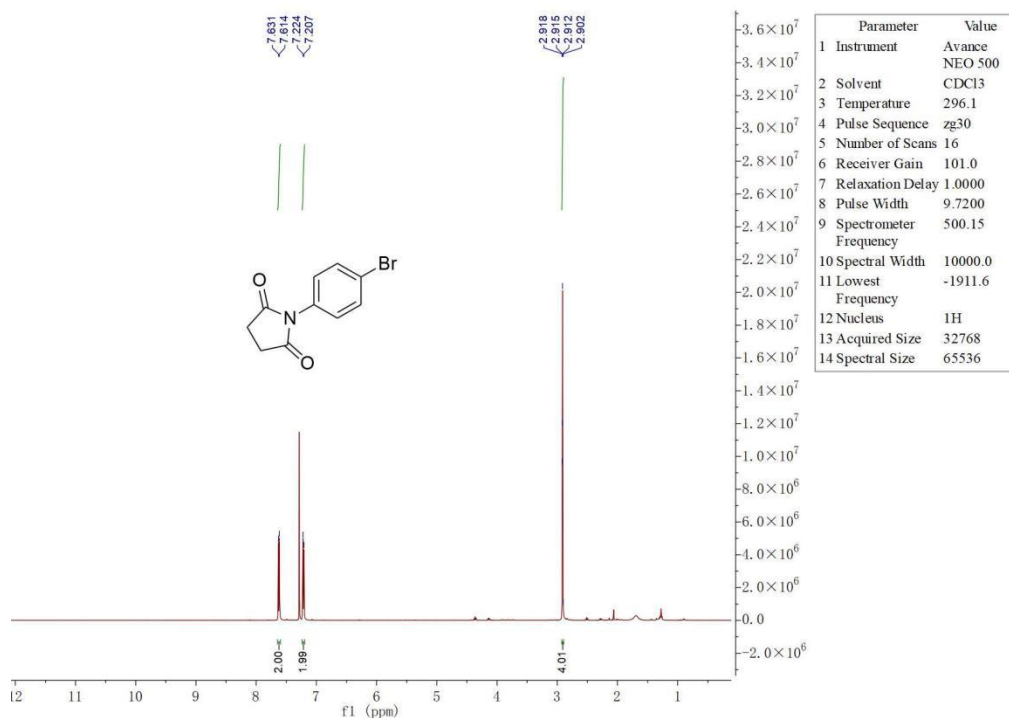
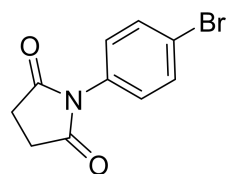


Figure S16. ¹H and ¹³C spectrum of compound 3g.

1-(4-iodophenyl)pyrrolidine-2,5-dione (3h)

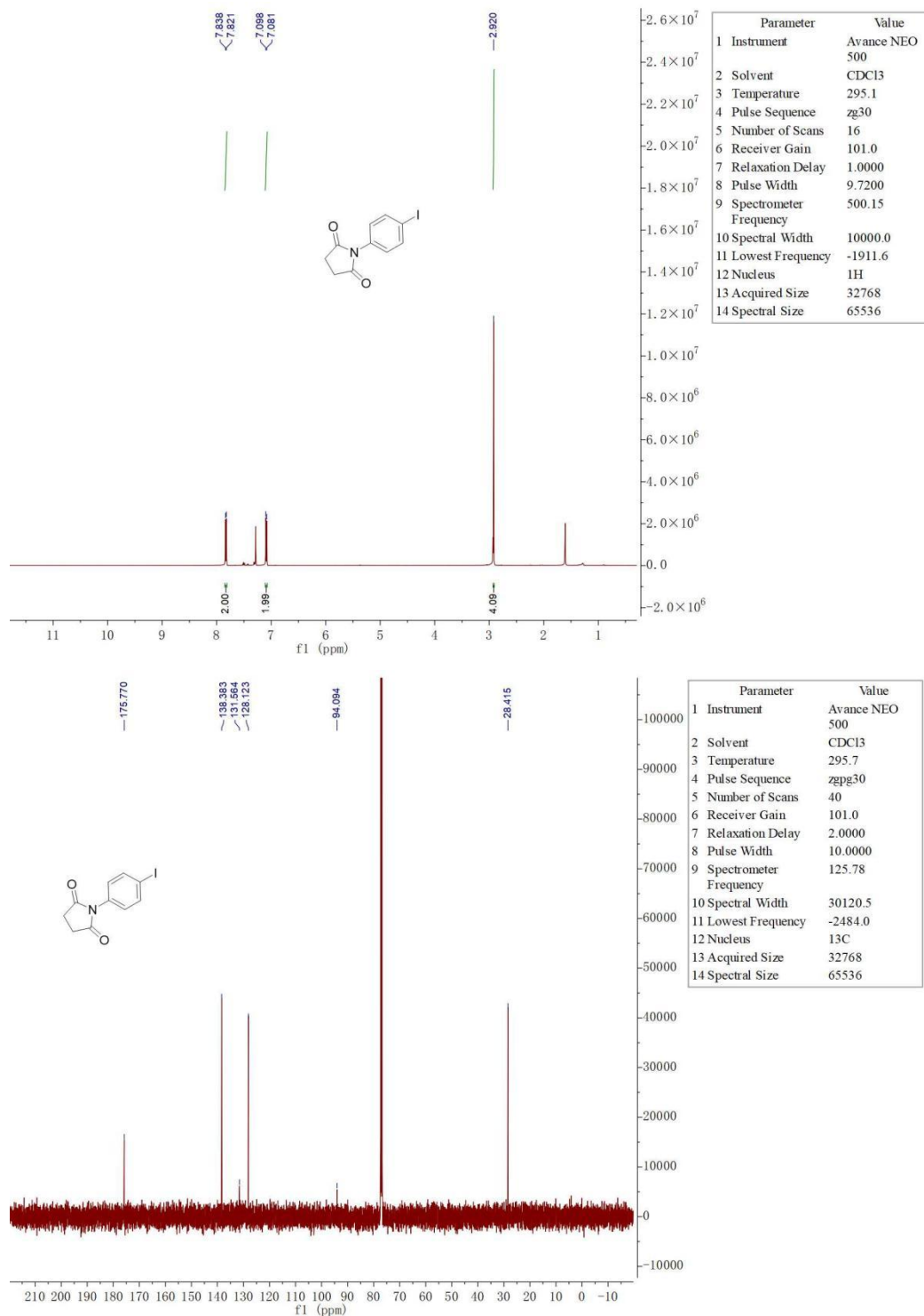
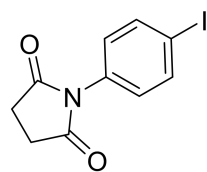


Figure S17. ¹H and ¹³C spectrum of compound 3h.

1-(4-(trifluoromethyl)phenyl)pyrrolidine-2,5-dione (3i)

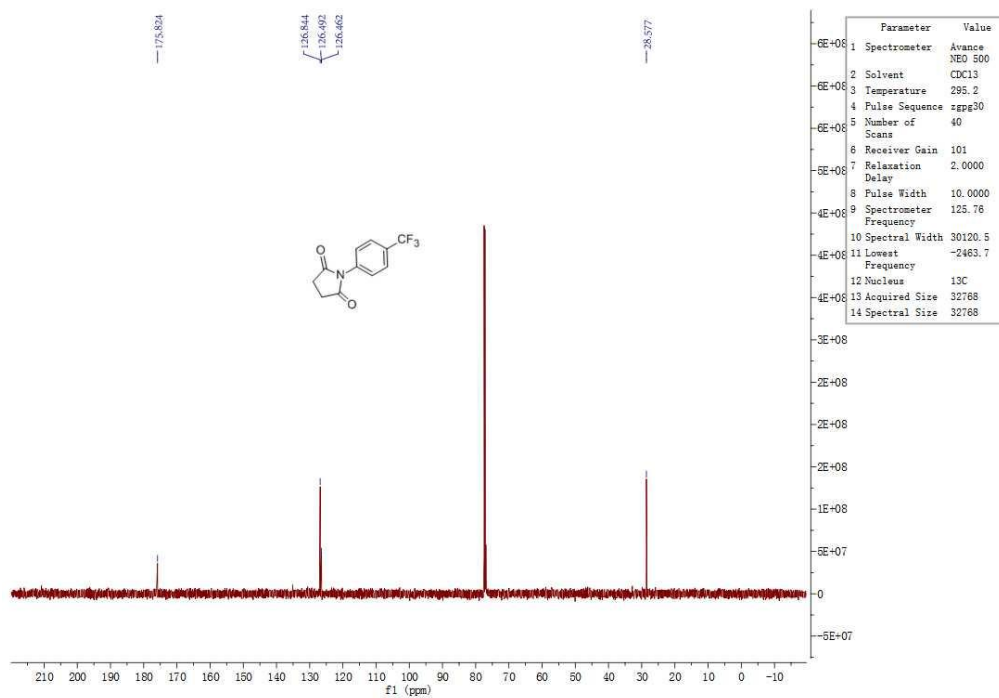
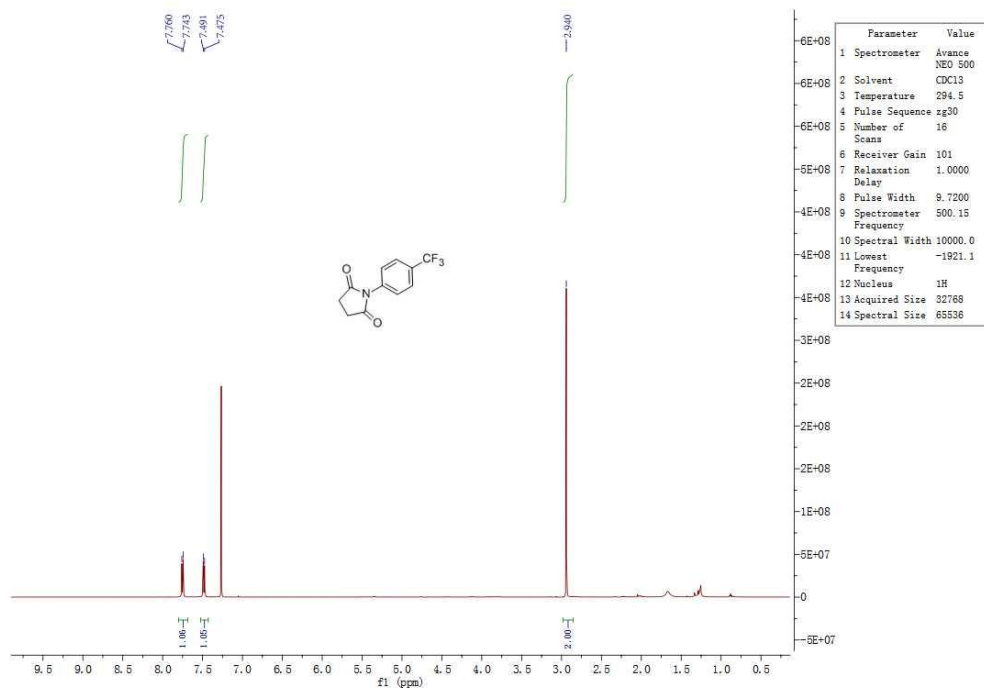
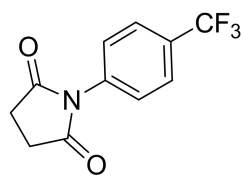


Figure S18. ^1H and ^{13}C spectrum of compound 3i.

1-(*m*-tolyl)pyrrolidine-2,5-dione (3j)

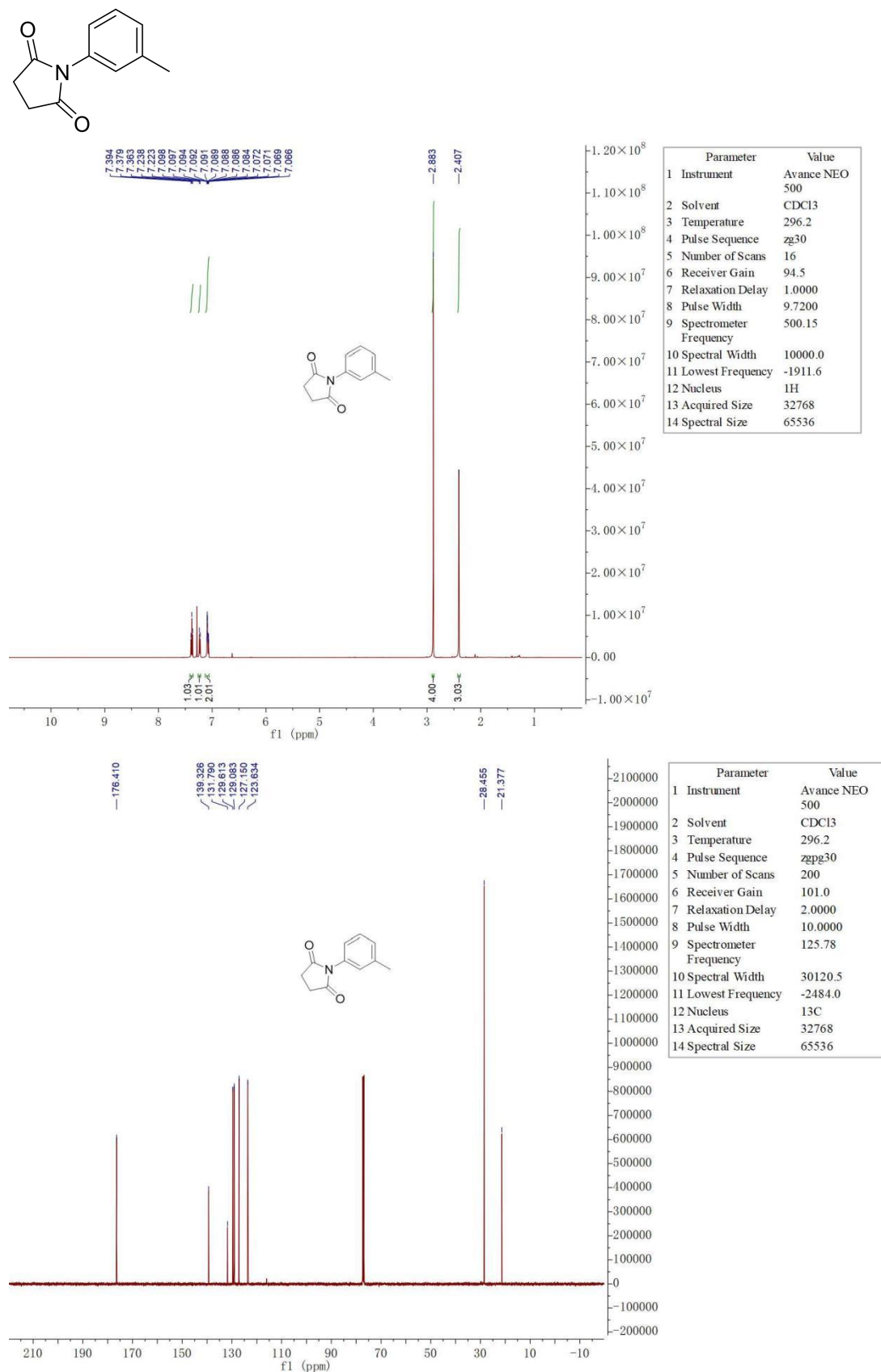


Figure S19. ¹H and ¹³C spectrum of compound **3j**.

1-(3-methoxyphenyl)pyrrolidine-2,5-dione (3k)

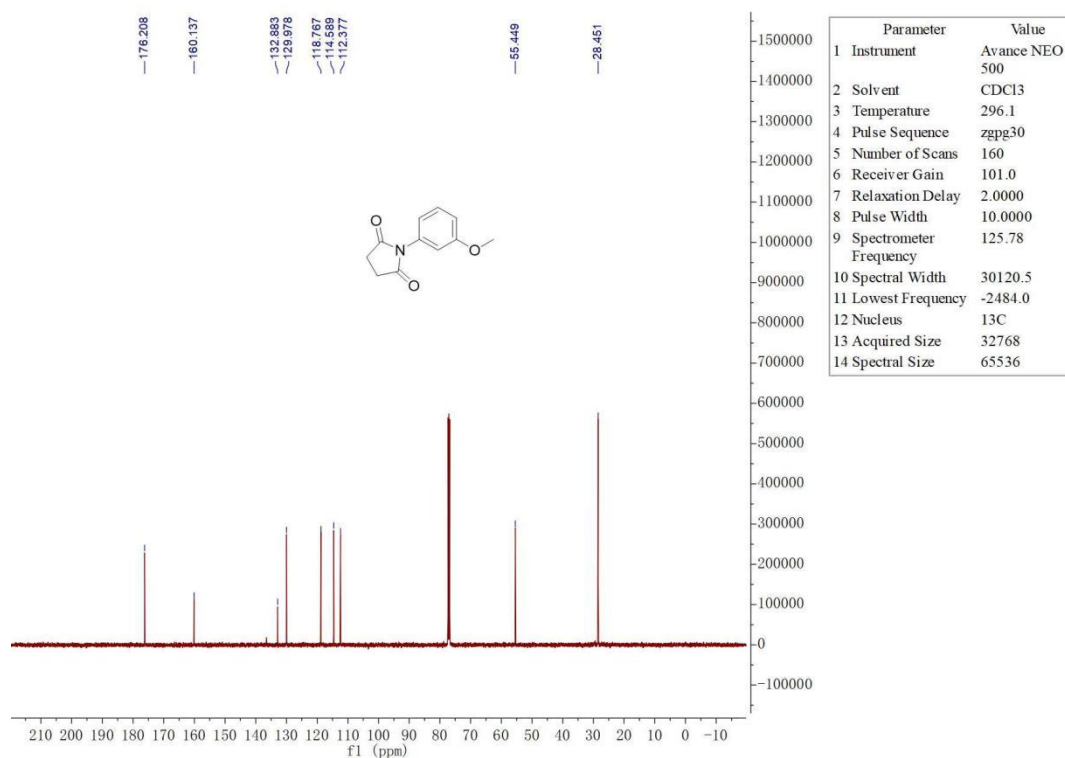
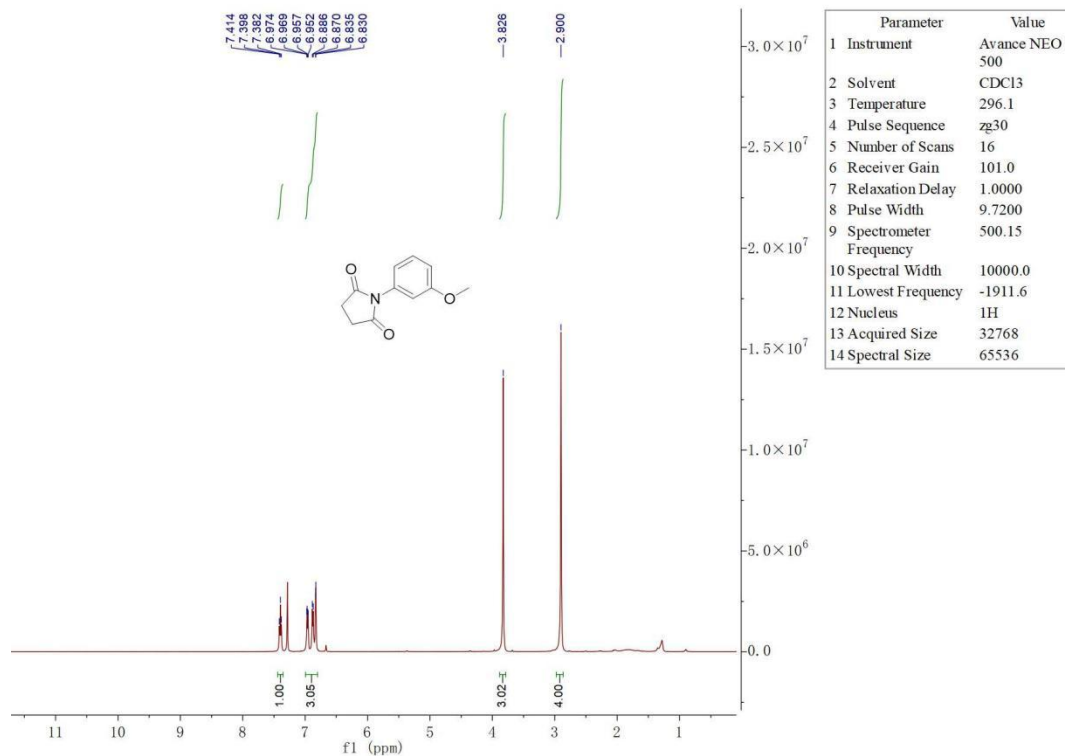
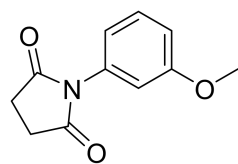


Figure S20. ^1H , ^{13}C and ^{19}F spectrum of compound **3k**.

1-(2-isopropylphenyl)pyrrolidine-2,5-dione (31)

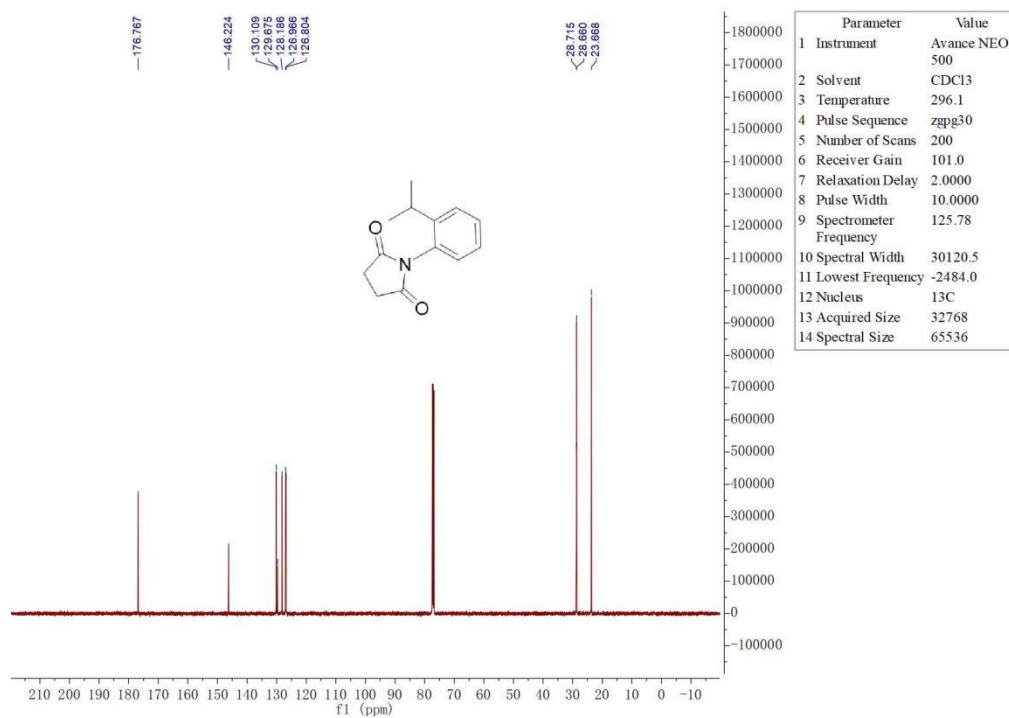
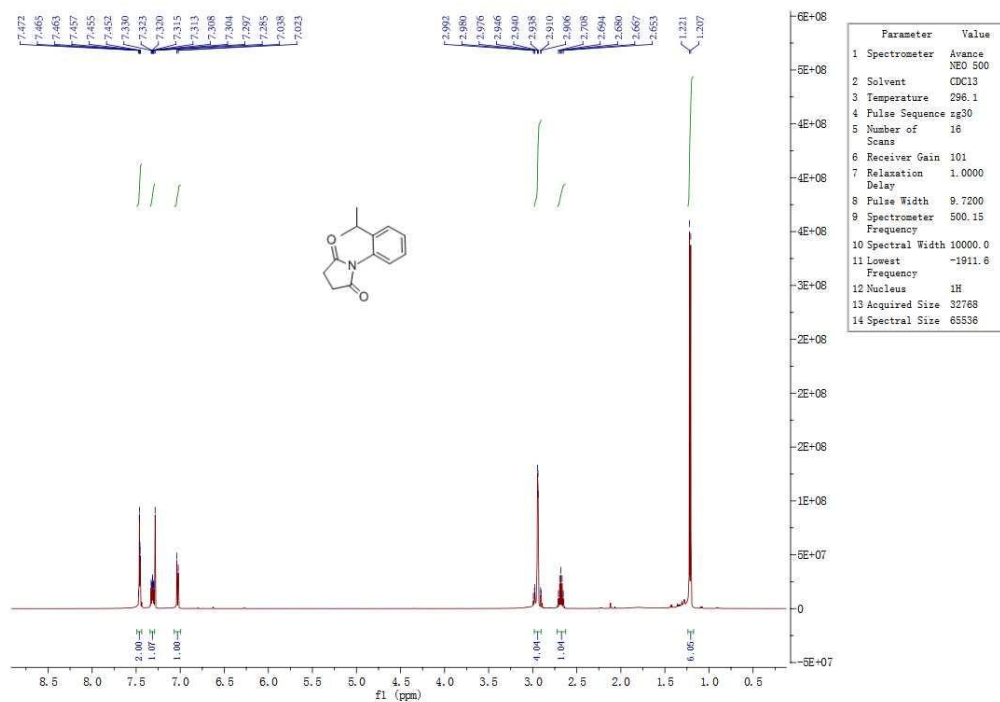
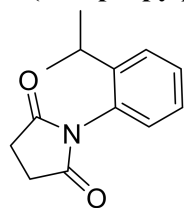


Figure S21. ¹H, ¹³C and ¹⁹F spectrum of compound 31.

1-(2-fluorophenyl)pyrrolidine-2,5-dione (3m)

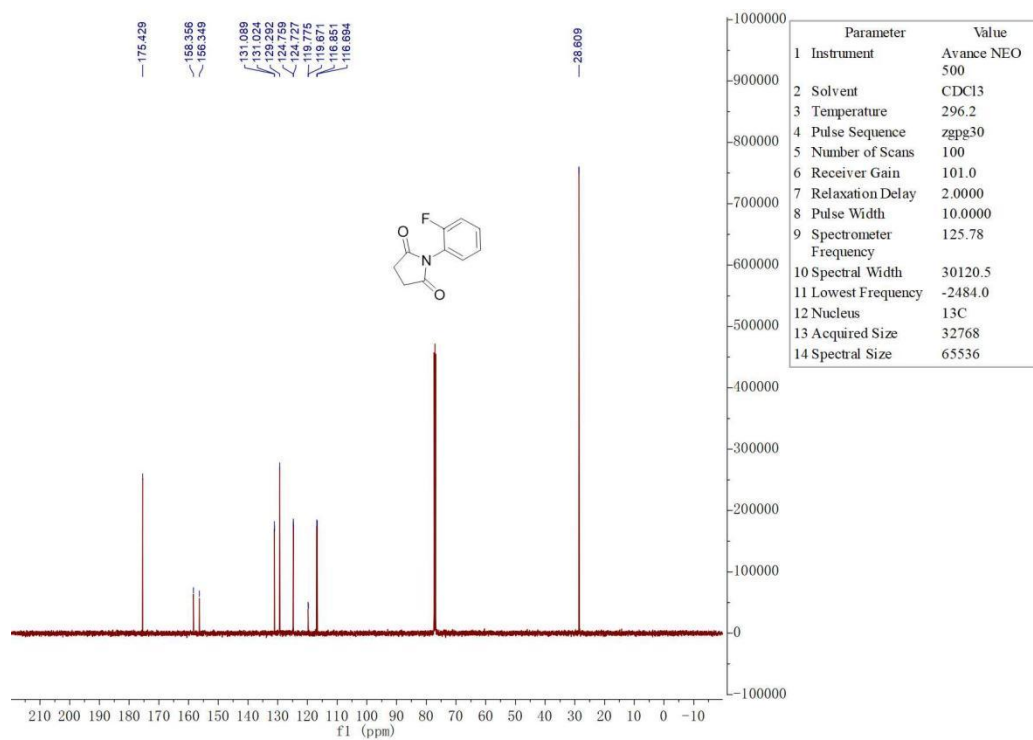
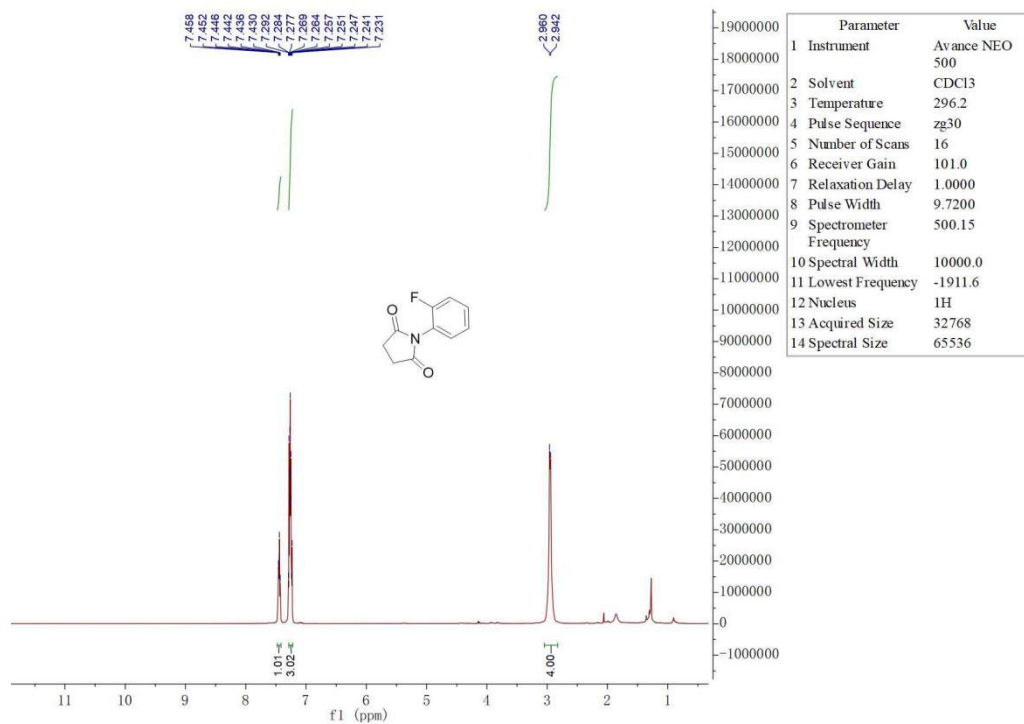


Figure S22. ¹H and ¹³C spectrum of compound 3m.

1-(2-iodophenyl)pyrrolidine-2,5-dione (3n)

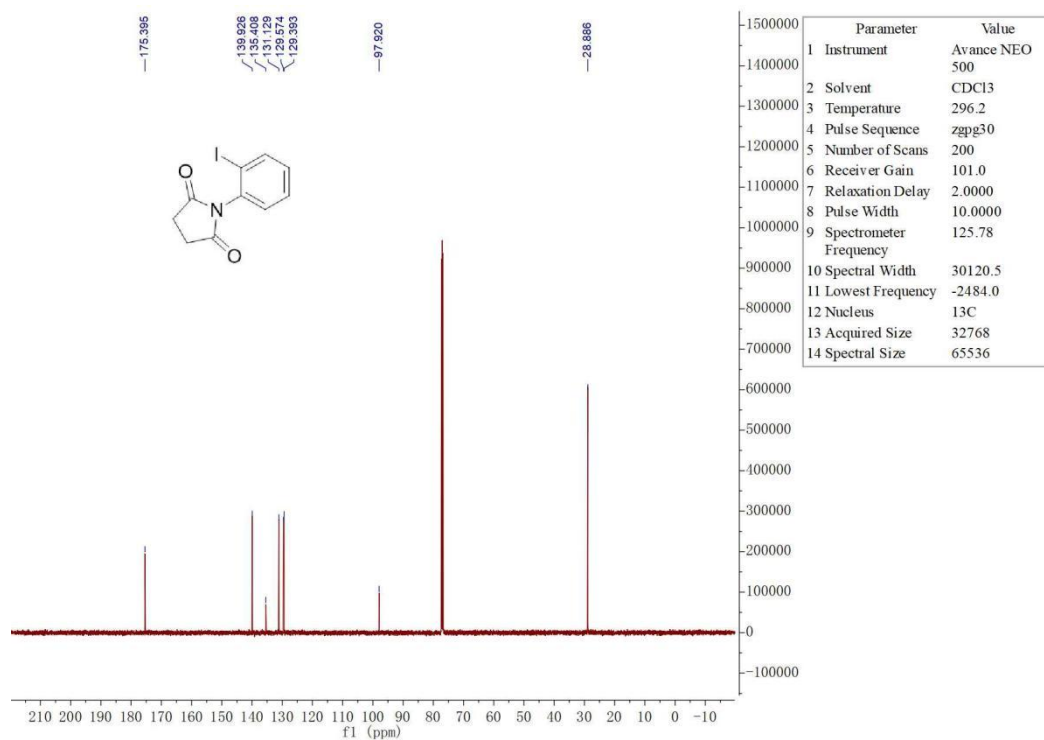
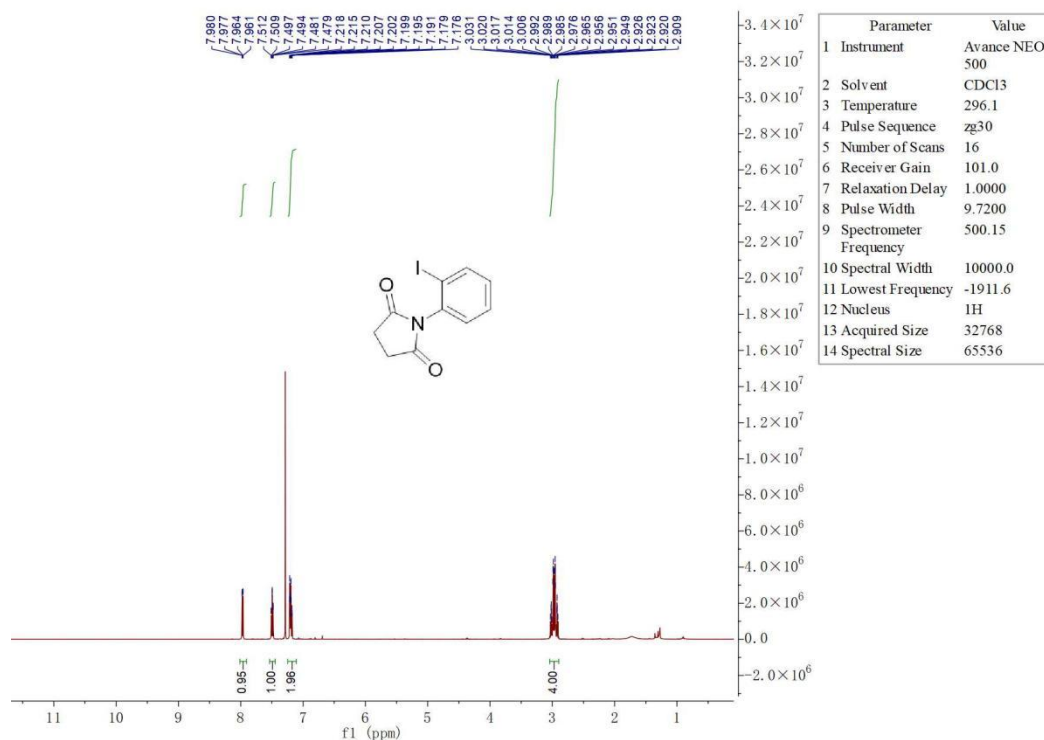
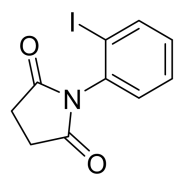


Figure S23. ¹H and ¹³C spectrum of compound 3n.

1-(2-hydroxyphenyl)pyrrolidine-2,5-dione (3o)

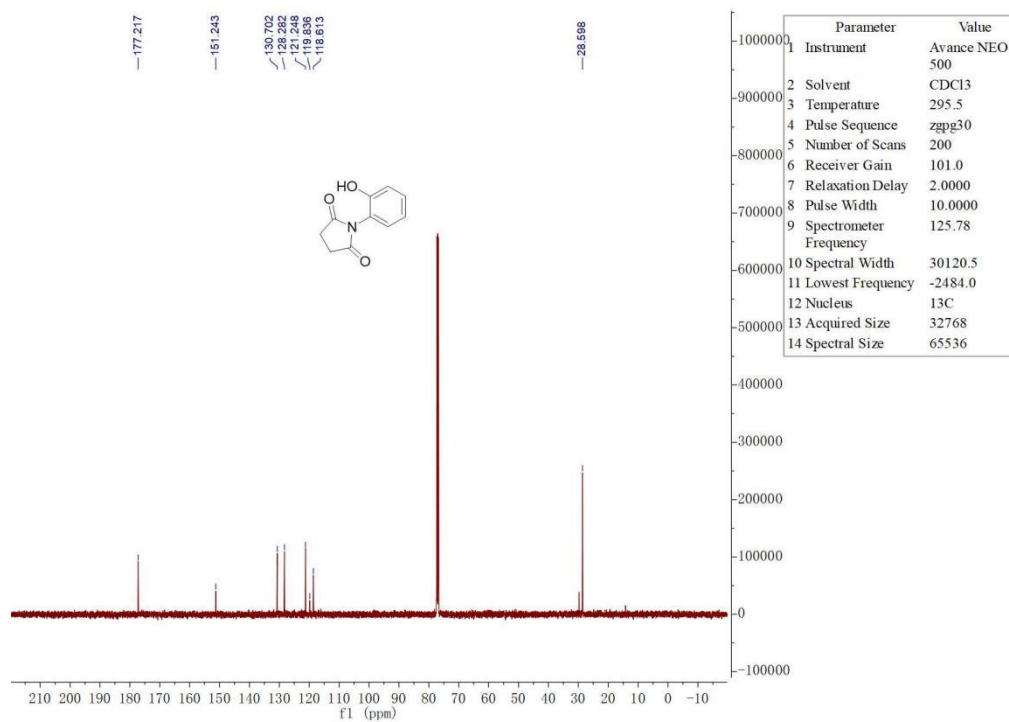
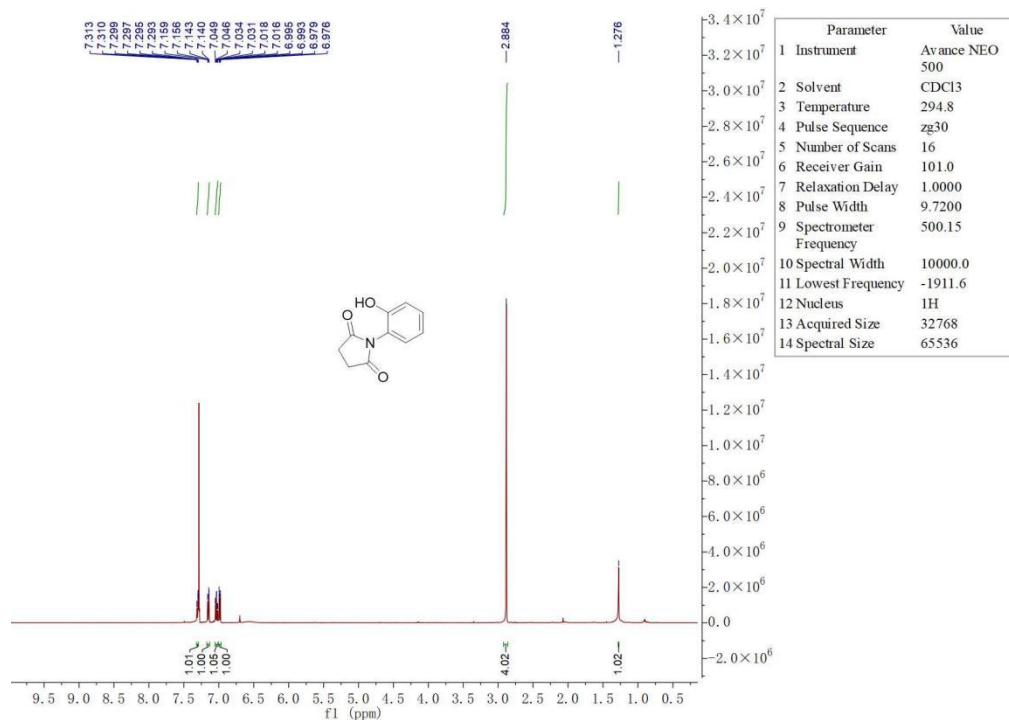
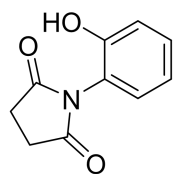
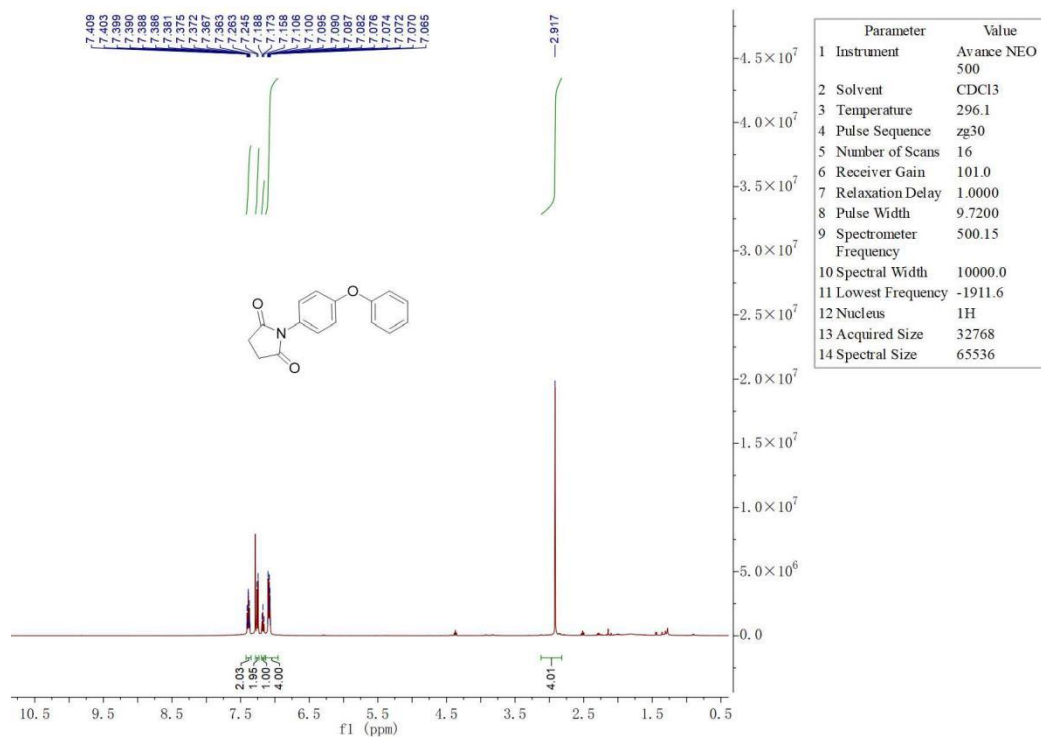
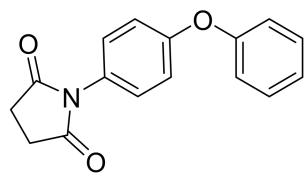
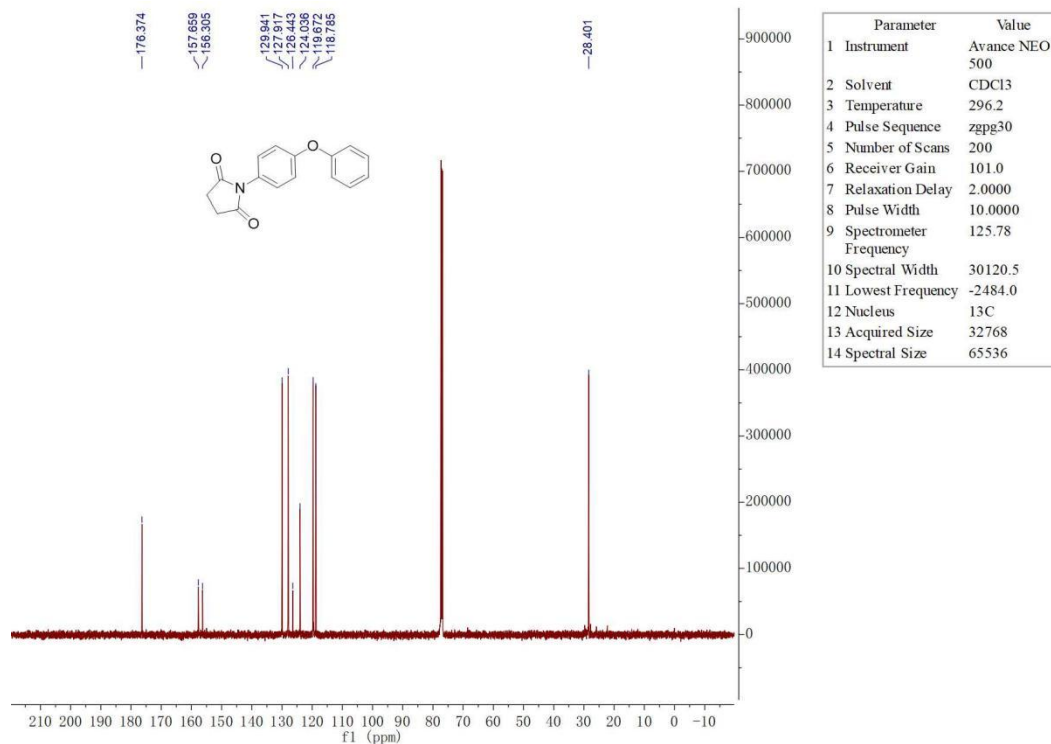


Figure S24. ¹H and ¹³C spectrum of compound 3o.

1-(4-phenoxyphenyl)pyrrolidine-2,5-dione (3p)



Parameter	Value
1 Instrument	Avance NEO
2 Solvent	CDCl3
3 Temperature	296.1
4 Pulse Sequence	zg30
5 Number of Scans	16
6 Receiver Gain	101.0
7 Relaxation Delay	1.0000
8 Pulse Width	9.7200
9 Spectrometer	500.15
Frequency	
10 Spectral Width	10000.0
11 Lowest Frequency	-1911.6
12 Nucleus	1H
13 Acquired Size	32768
14 Spectral Size	65536



Parameter	Value
1 Instrument	Avance NEO
2 Solvent	CDCl3
3 Temperature	296.2
4 Pulse Sequence	zgpg30
5 Number of Scans	200
6 Receiver Gain	101.0
7 Relaxation Delay	2.0000
8 Pulse Width	10.0000
9 Spectrometer	125.78
Frequency	
10 Spectral Width	30120.5
11 Lowest Frequency	-2484.0
12 Nucleus	13C
13 Acquired Size	32768
14 Spectral Size	65536

Figure S25. ¹H and ¹³C spectrum of compound 3p.

1-(3,4-dimethoxyphenyl)pyrrolidine-2,5-dione (3q)

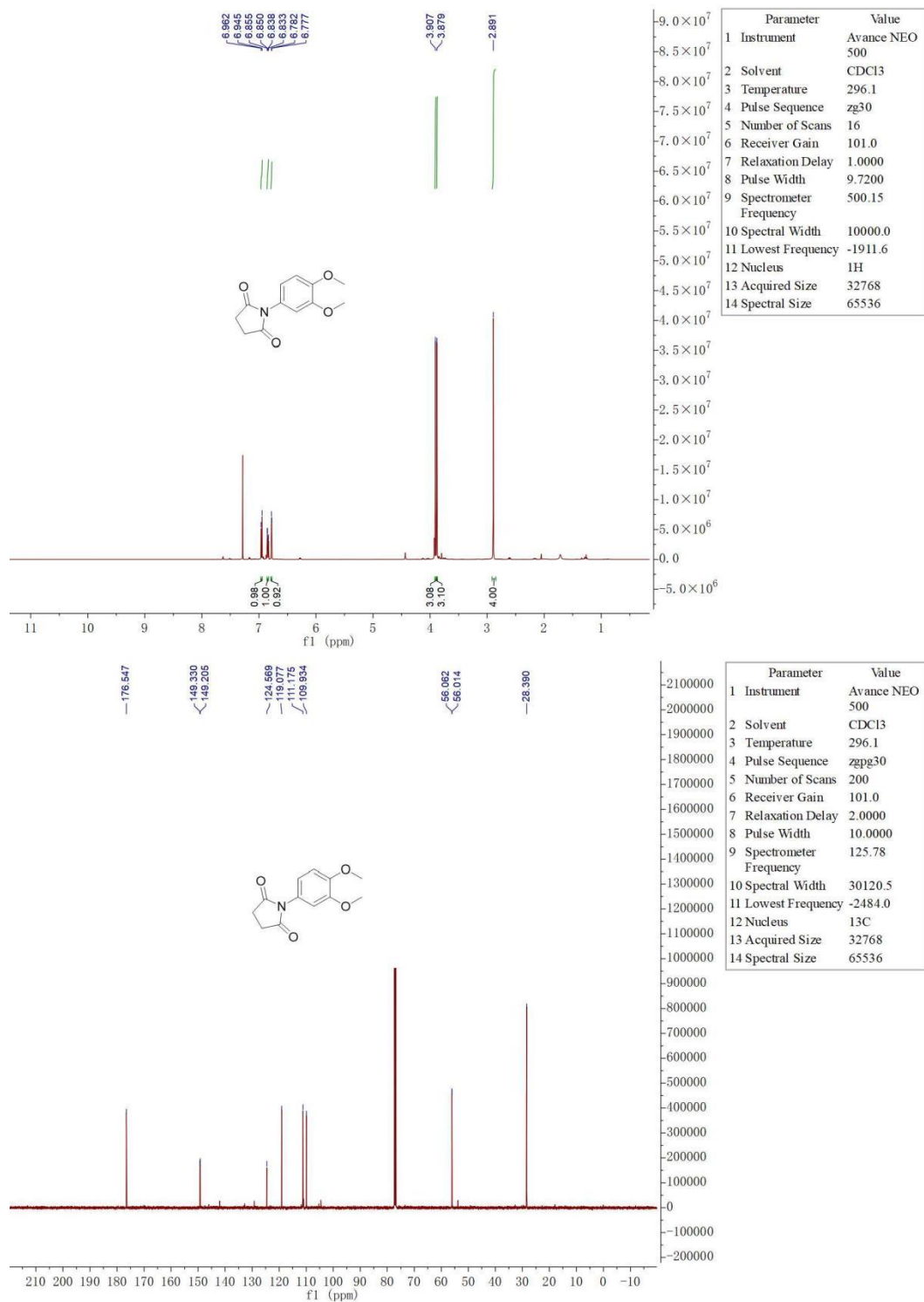
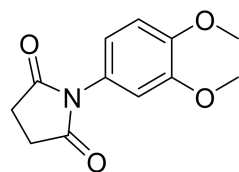


Figure S26. ¹H and ¹³C spectrum of compound 3q.

1-(4-acetylphenyl)pyrrolidine-2,5-dione (3r)

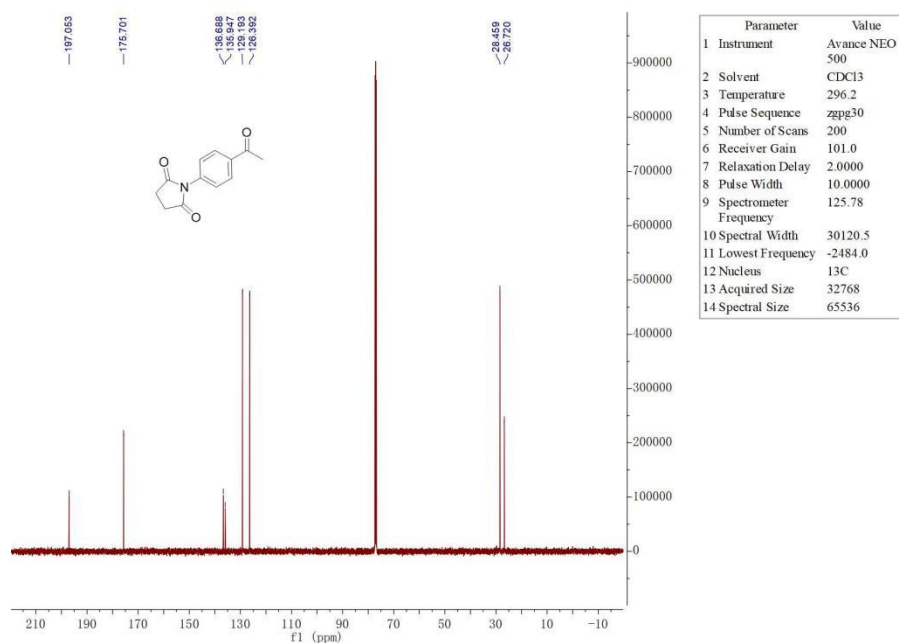
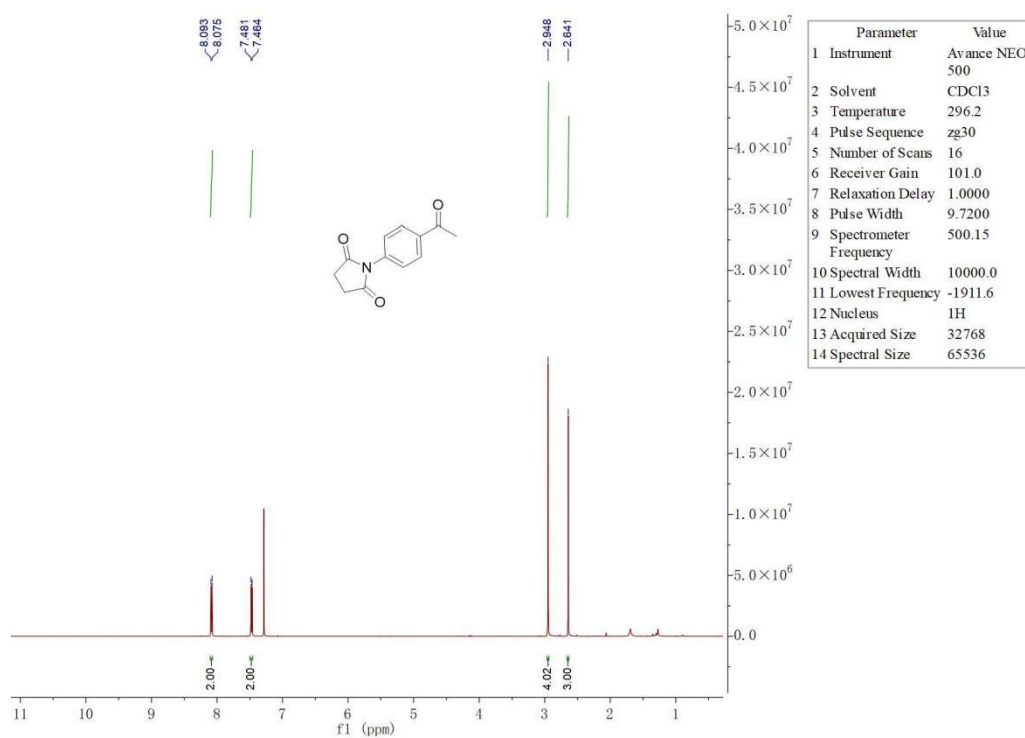
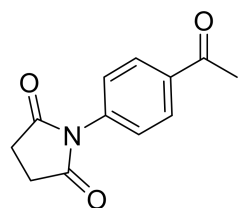


Figure S27. ¹H and ¹³C spectrum of compound 3r.

1-(3-acetylphenyl)pyrrolidine-2,5-dione (3s)

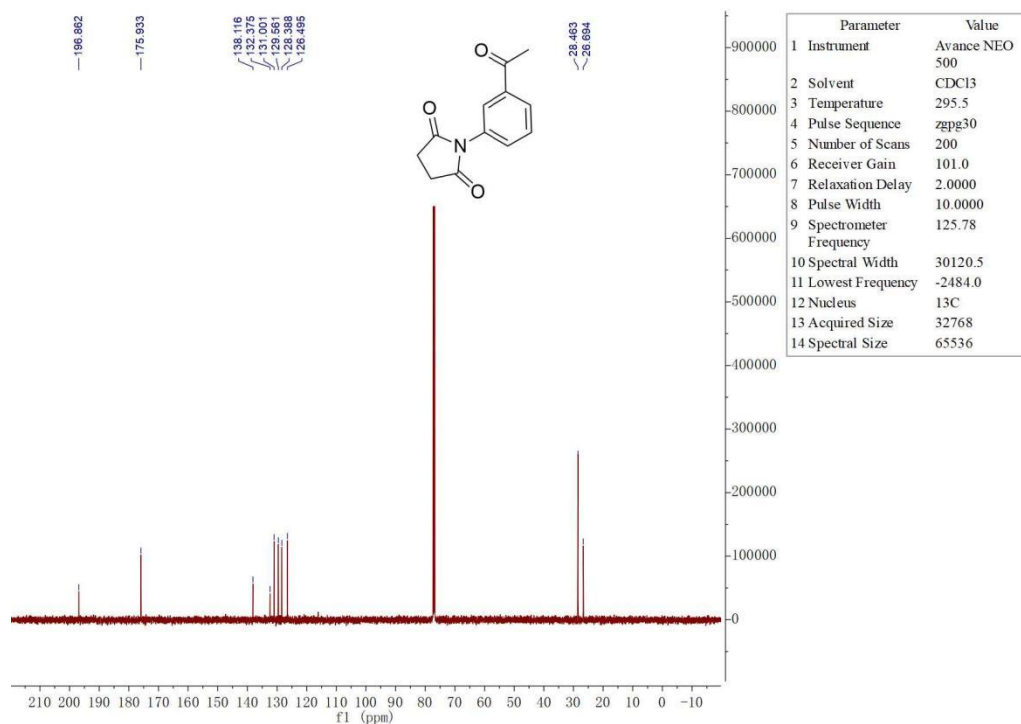
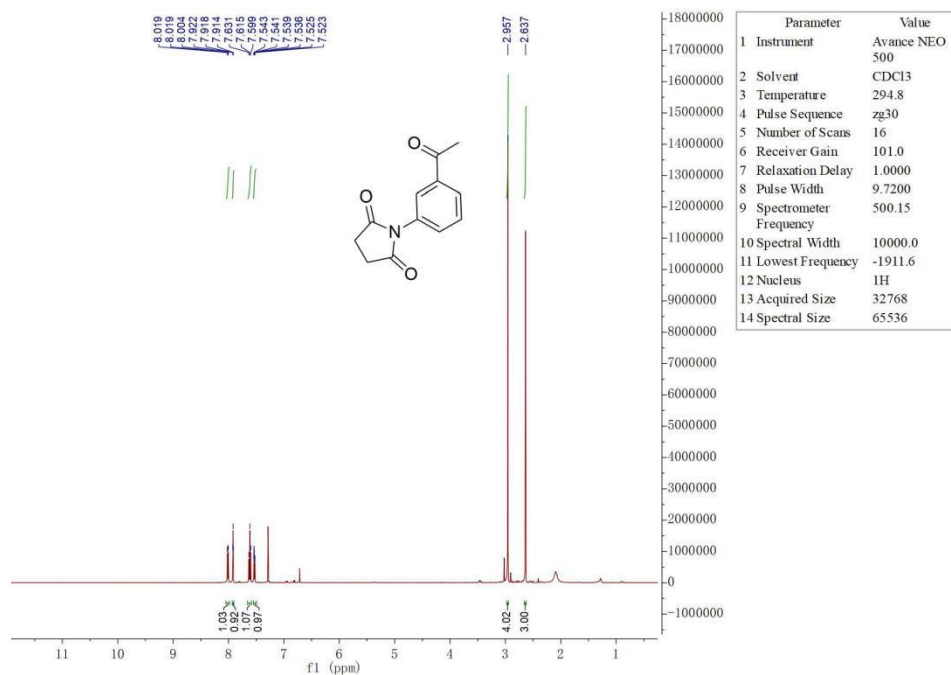
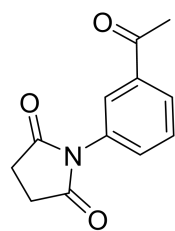


Figure S28. ¹H and ¹³C spectrum of compound 3s.

methyl 4-(2,5-dioxopyrrolidin-1-yl)benzoate (**3t**)

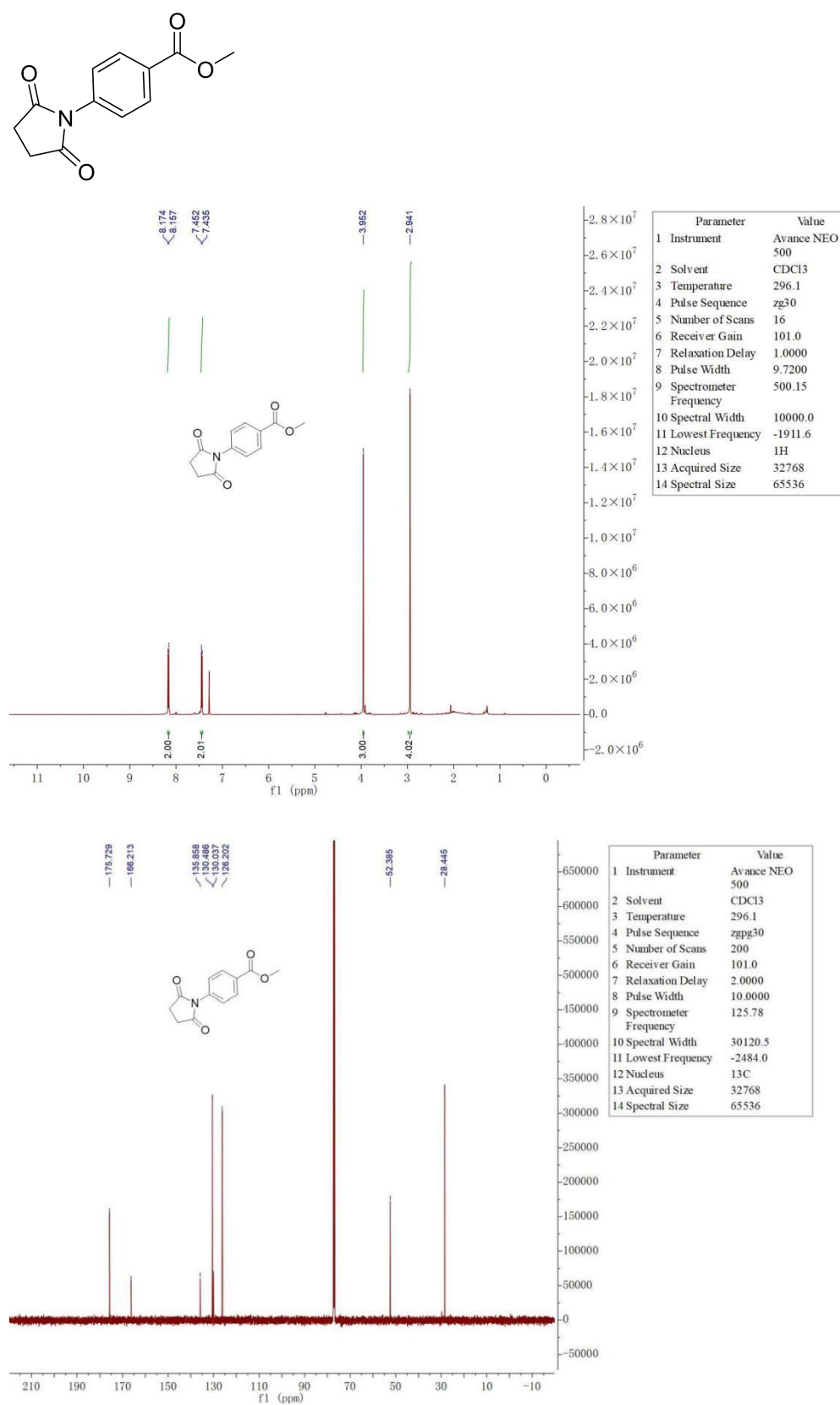


Figure S29. ¹H and ¹³C spectrum of compound **3t**.

1-benzylpyrrolidine-2,5-dione (3u)

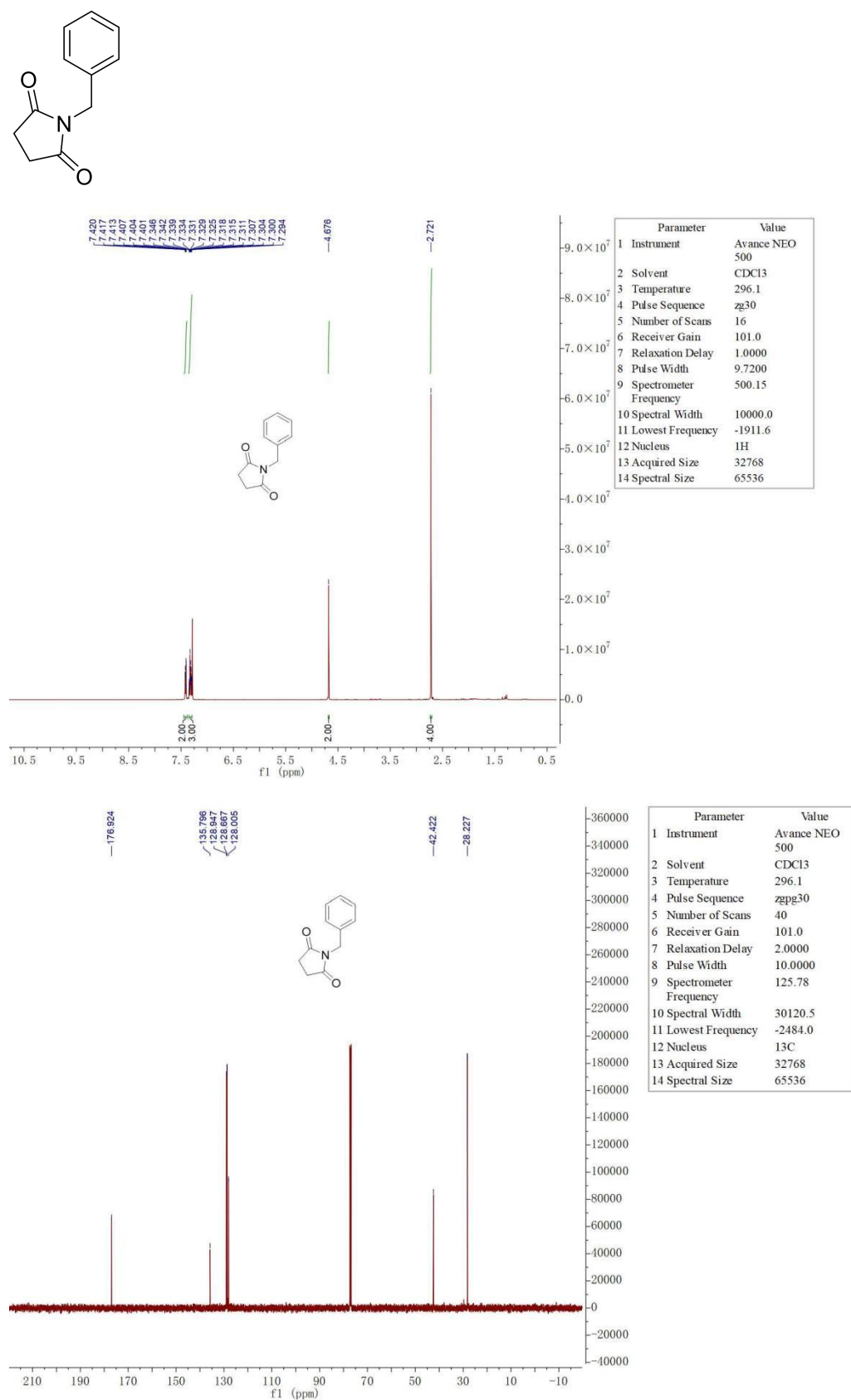


Figure S30. ¹H and ¹³C spectrum of compound 3u.

1-(3-bromobenzyl)pyrrolidine-2,5-dione (3v)

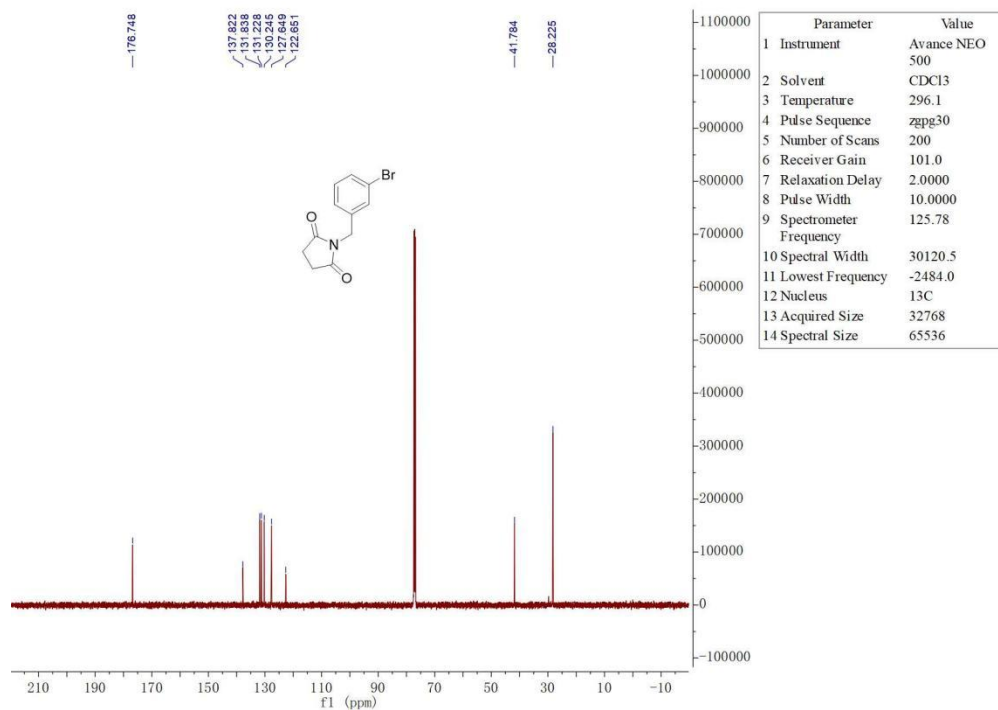
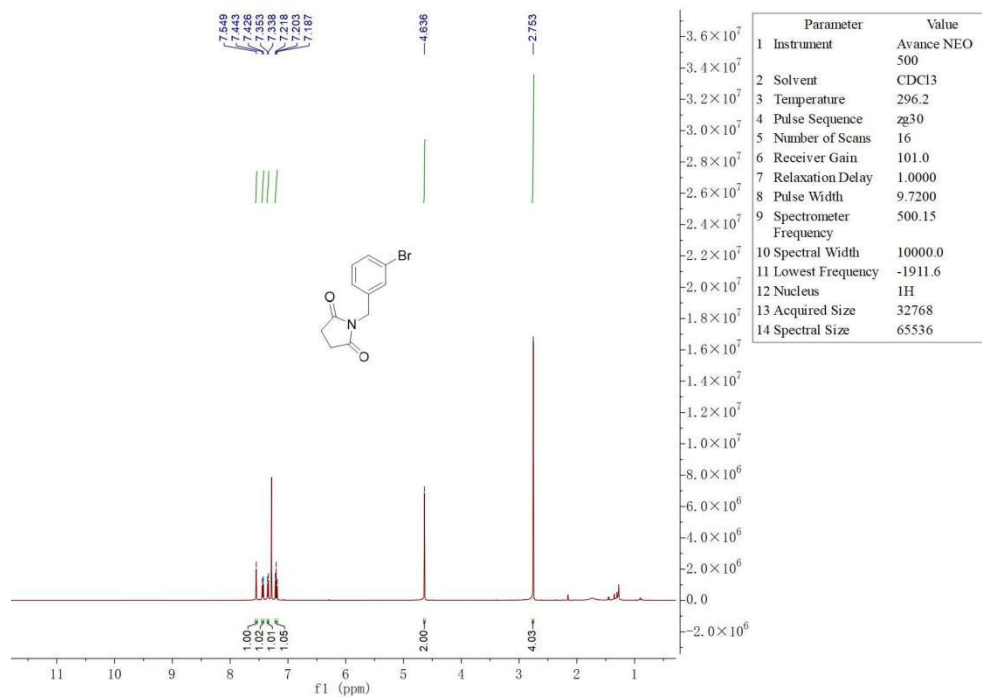
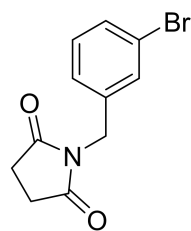


Figure S31. ¹H and ¹³C spectrum of compound 3v.

1-(naphthalen-1-yl)pyrrolidine-2,5-dione (3w)

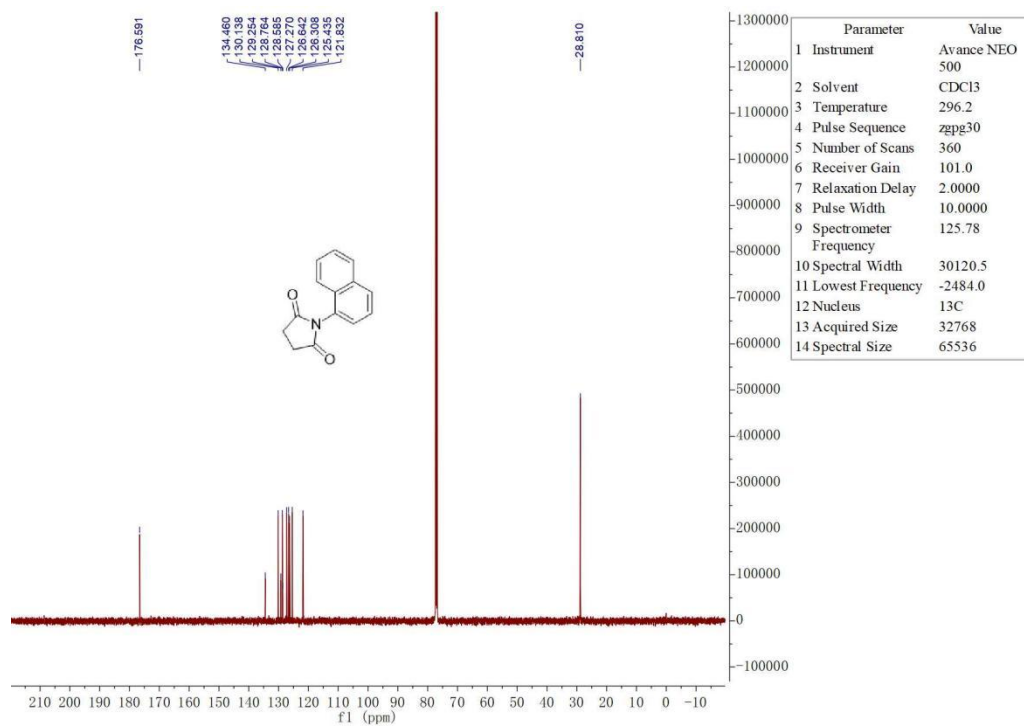
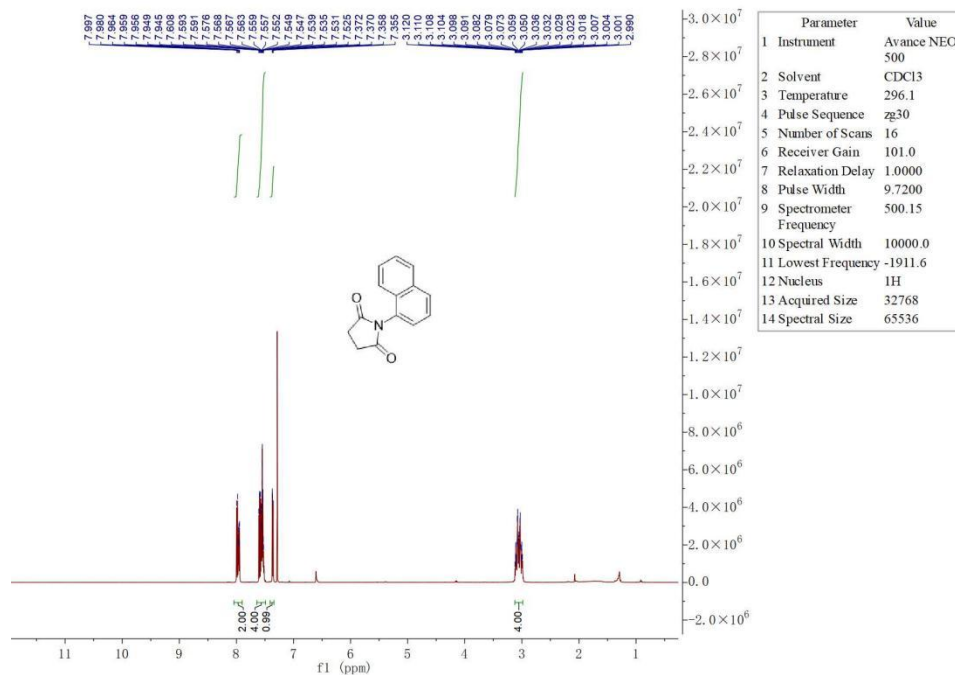
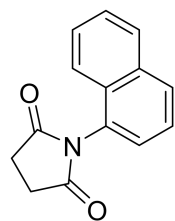


Figure S32. ¹H and ¹³C spectrum of compound 3w.

1-(4-bromonaphthalen-1-yl)pyrrolidine-2,5-dione (3x)

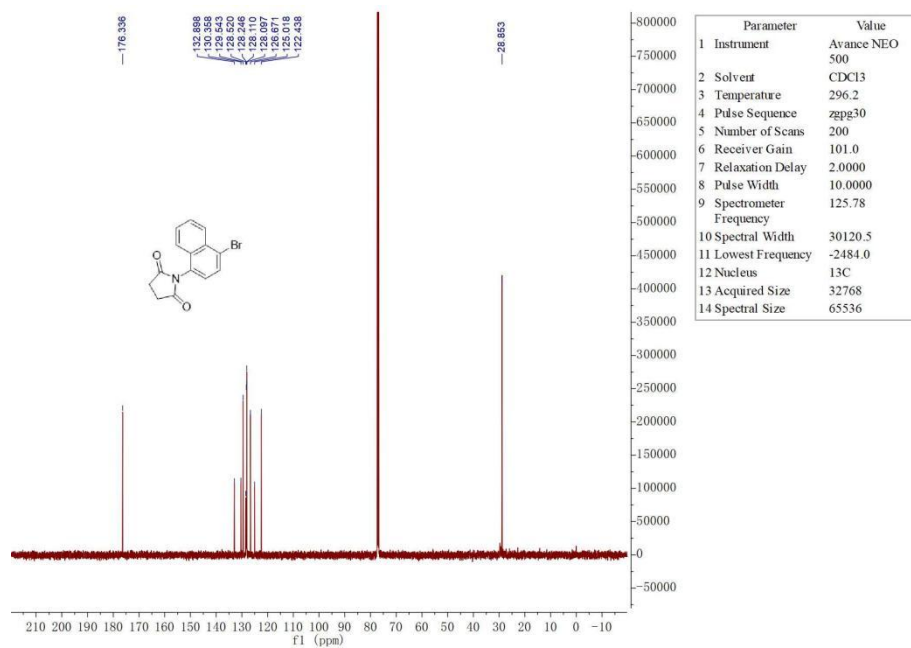
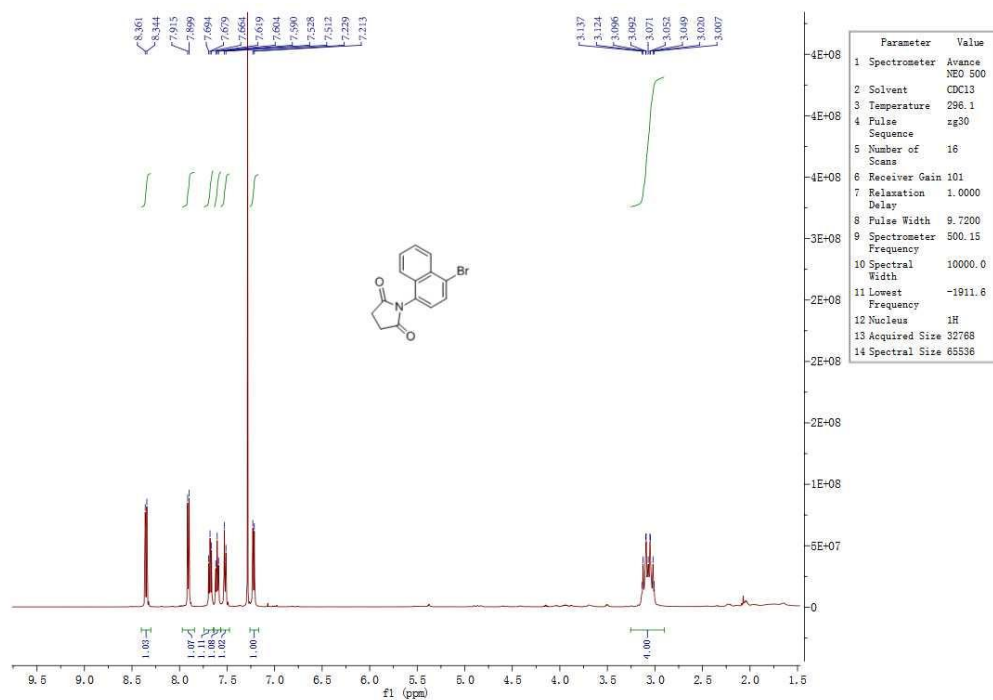
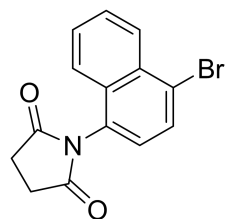


Figure S33. ¹H and ¹³C spectrum of compound 3x.

1-(9H-fluoren-2-yl)pyrrolidine-2,5-dione (3y)

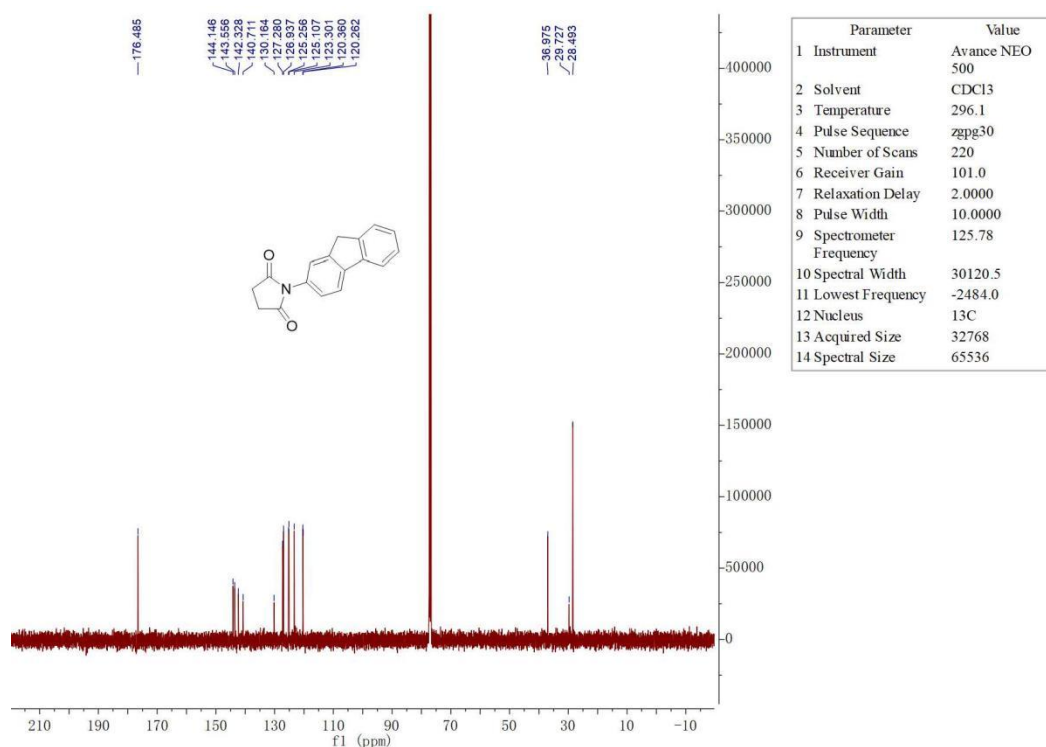
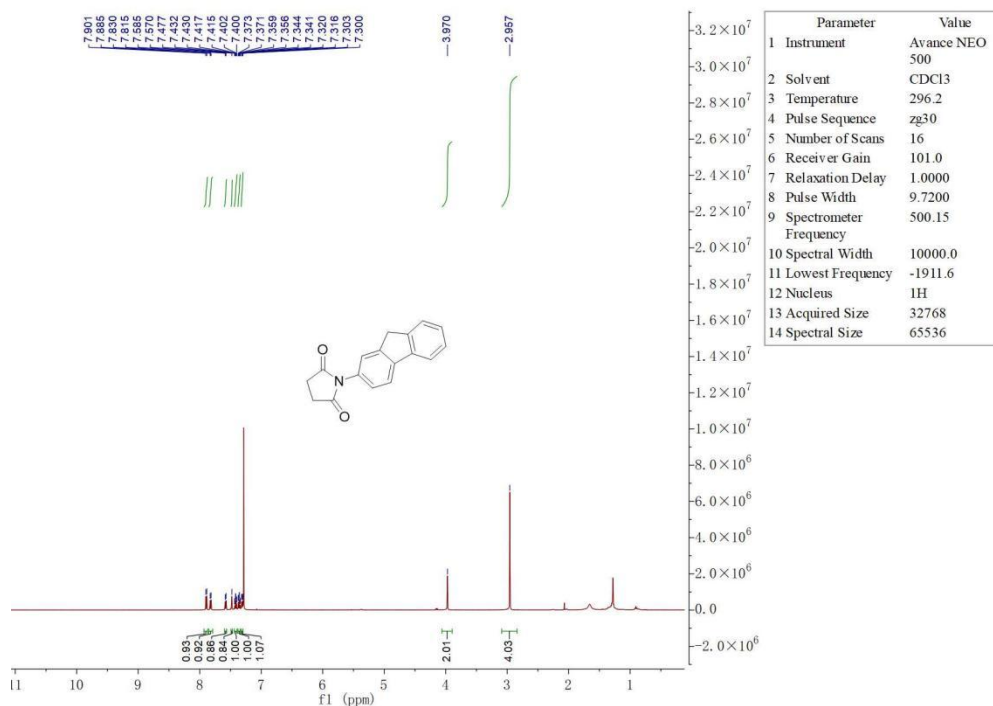
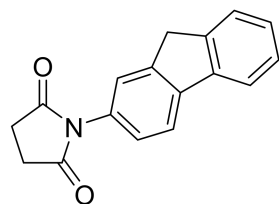


Figure S34. ¹H and ¹³C spectrum of compound 3y.

1-(pyren-1-yl)pyrrolidine-2,5-dione (3z)

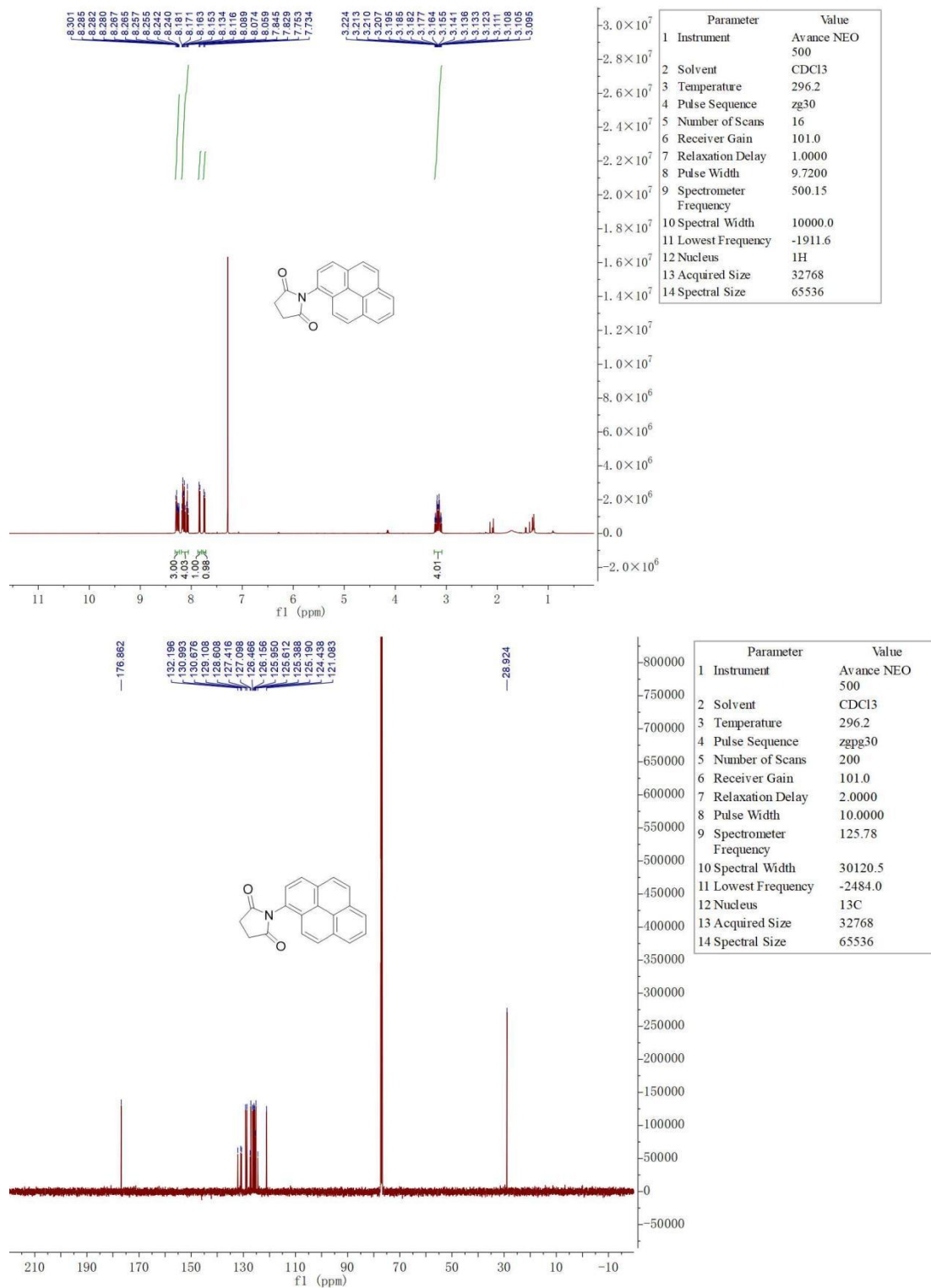
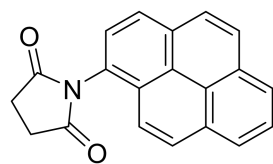


Figure S35. ¹H and ¹³C spectrum of compound 3z.

3-(4-(2,5-dioxopyrrolidin-1-yl)phenyl)-3-ethylpiperidine-2,6-dione (3aa)

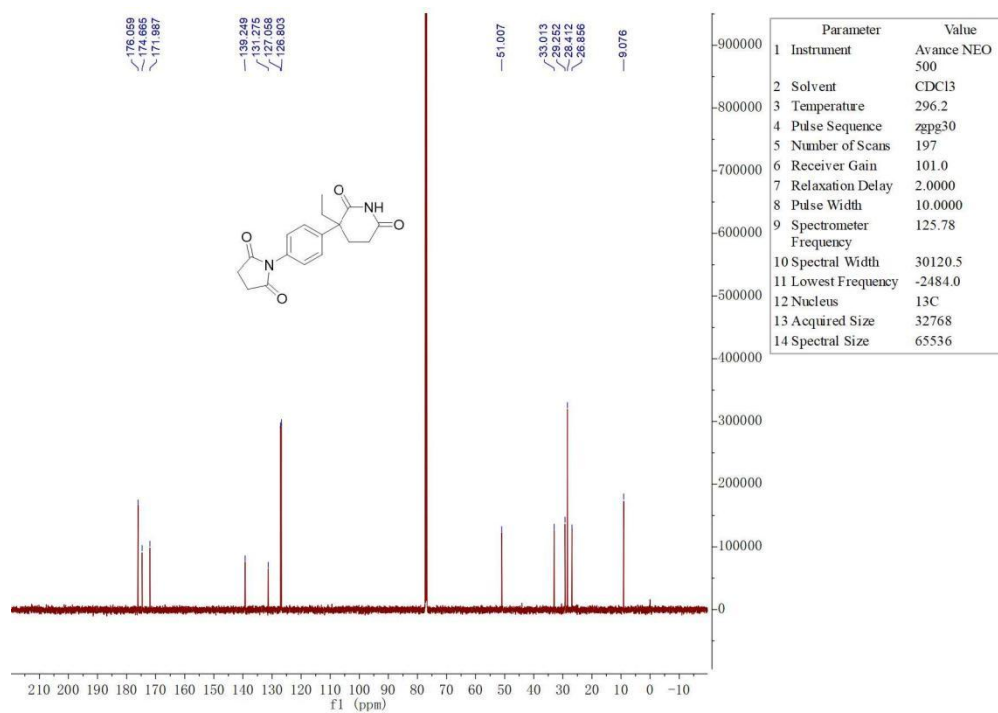
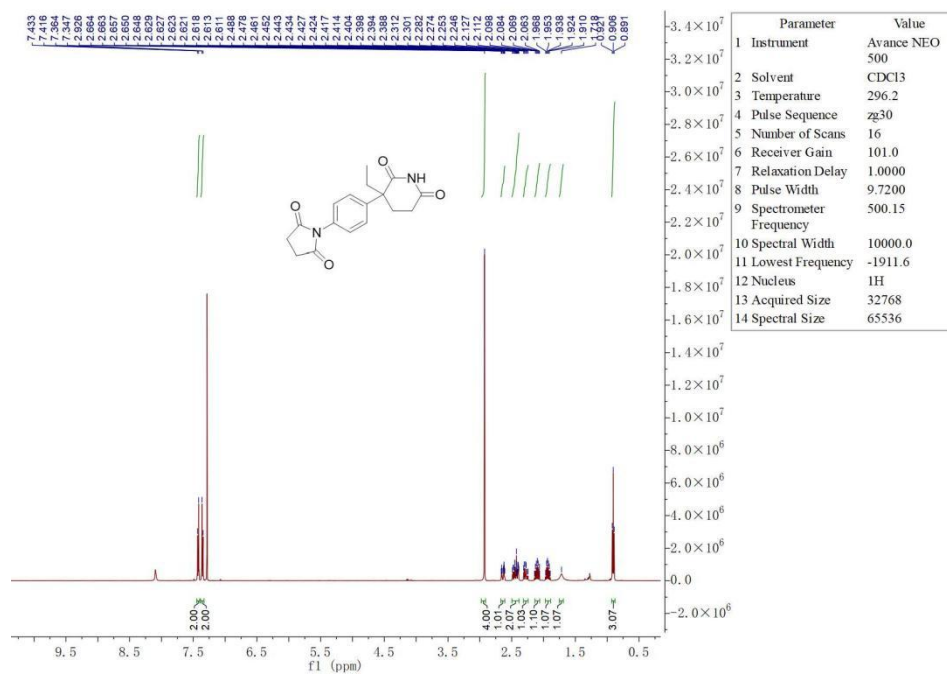
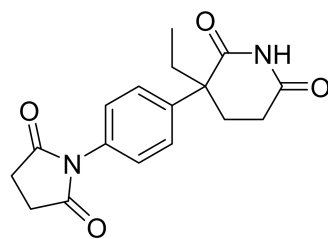


Figure S36. ¹H and ¹³C spectrum of compound **3aa**.

1,3-diphenylpyrrolidine-2,5-dione (3ab)

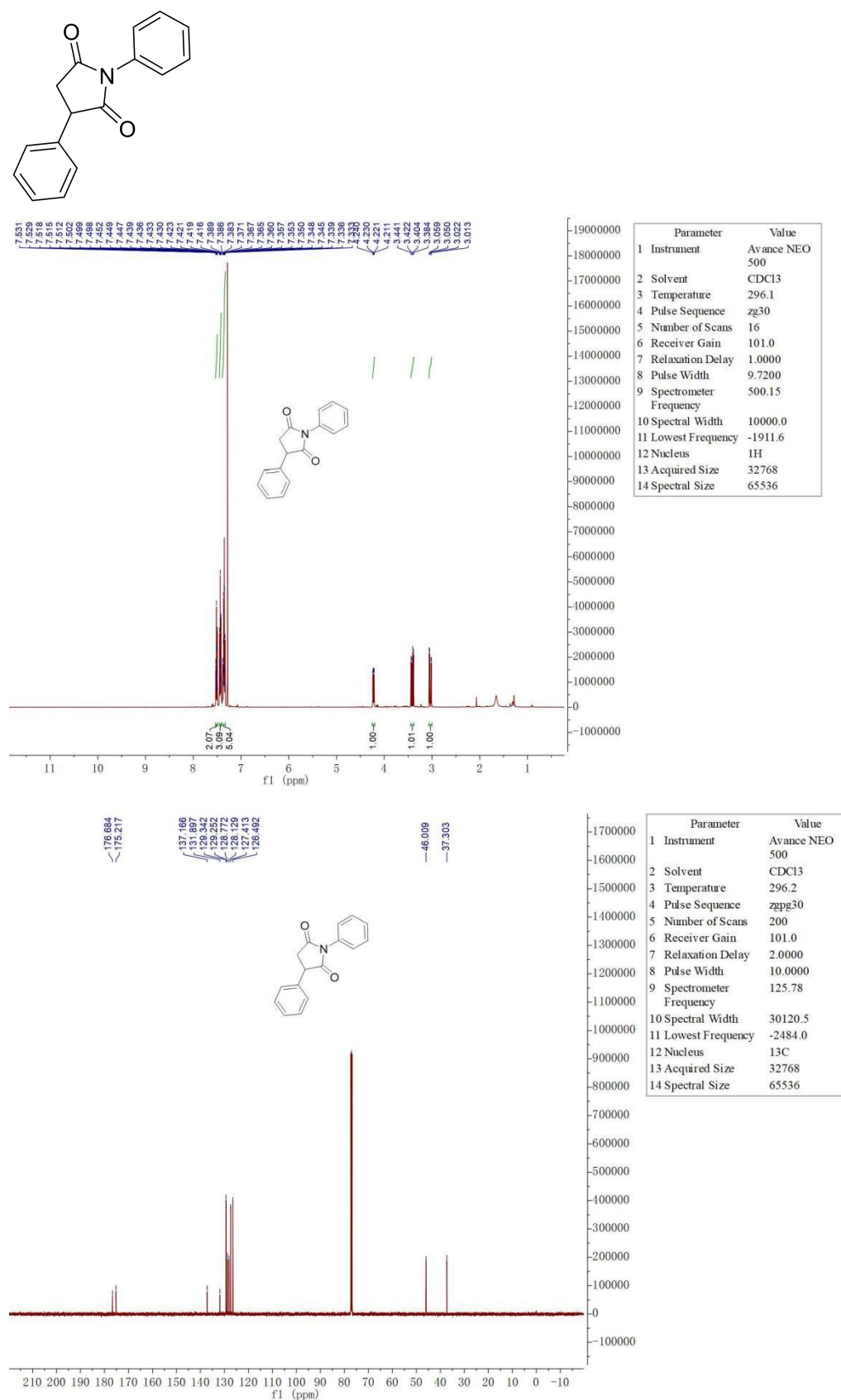


Figure S37. ¹H and ¹³C spectrum of compound **3ab**.

3-methyl-1-phenylpyrrolidine-2,5-dione (3ac)

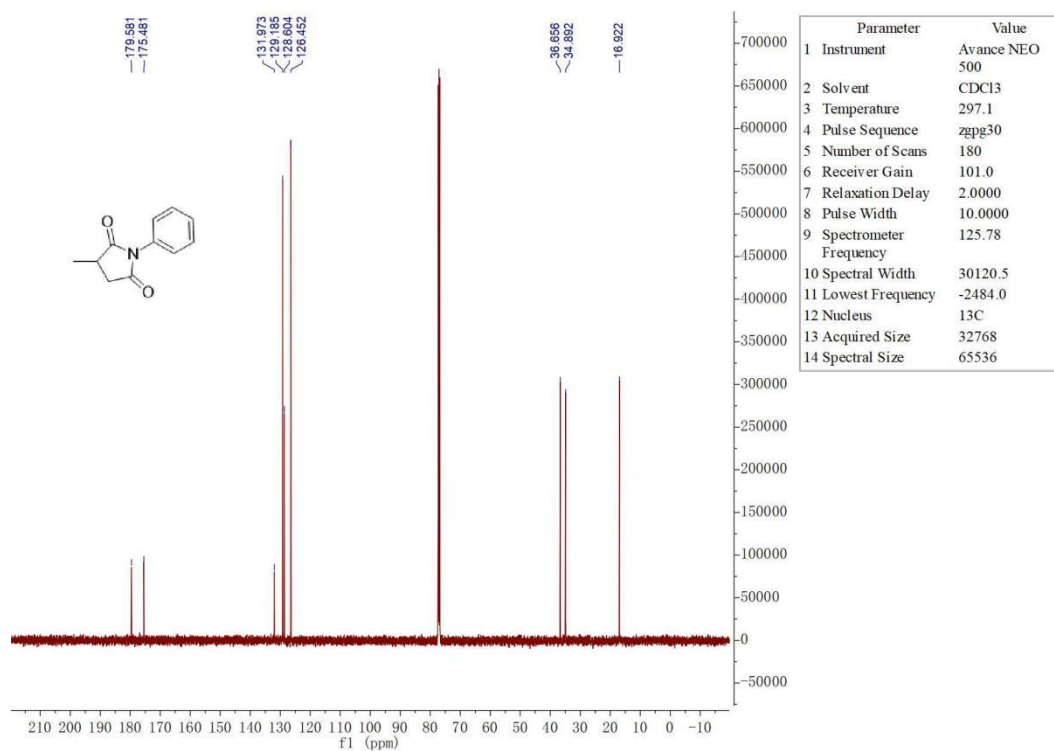
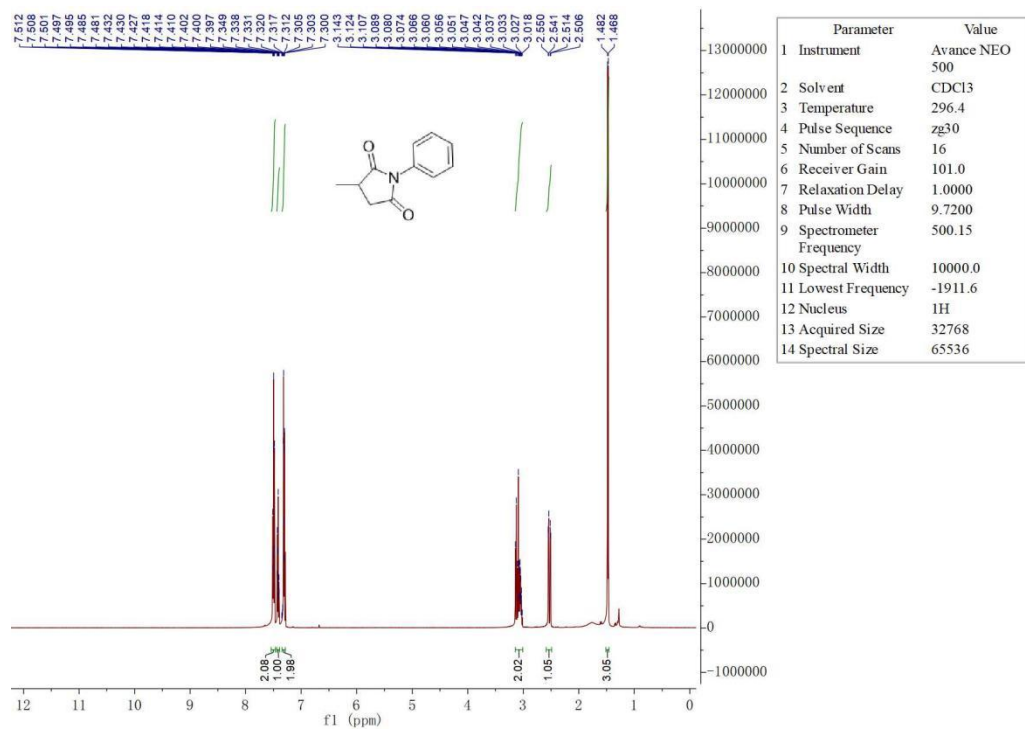
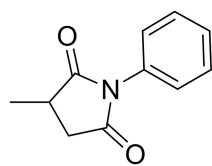


Figure S38. ¹H and ¹³C spectrum of compound 3ac.