Supporting Information

Engineering the substrate preference of glucose oxidase for the

enzymatic oxidation of xylose

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Supplemental data

Coding sequence of wild-type *Aspergillus niger* glucose oxidase used in this study (mature protein, codon optimized for *Pichia pastoris*):

TCTAATGGTATTGAAGCTTCTTTGTTGACTGATCCAAAAGATGTTTCTGGTAGAACTGTT GATTATATTATTGCTGGTGGAGGTTTGACTGGTTTGACTACTGCTGCTAGATTGACTGAA AATCCAAATATTTCTGTTTTGGTTATTGAATCTGGTTCTTATGAATCTGATAGAGGTCCA ATTATTGAAGATTTGAATGCTTATGGTGATATTTTTGGTTCTTCTGTTGATCATGCTTATG AAACTGTTGAATTGGCTACTAATAATCAAACTGCTTTGATTAGATCTGGTAATGGTTTGG GTGGTTCTACTTTGGTTAATGGTGGTACTTGGACTAGACCACATAAAGCTCAAGTTGAT TCTTGGGAAACTGTTTTTGGTAATGAAGGTTGGAATTGGGATAATGTTGCTGCTTATTCT TTGCAAGCTGAAAGAGCTAGAGCTCCAAATGCTAAACAAATTGCTGCTGGTCATTATTT TAATGCTTCTTGTCATGGTGTTAATGGTACTGTTCATGCTGGTCCAAGAGATACTGGTGA TGATTATTCTCCAATTGTTAAAGCTTTGATGTCTGCTGTTGAAGATAGAGGTGTTCCAAC TAAAAAGATTTTGGTTGTGGTGATCCACATGGTGTTTCTATGTTTCCAAATACTTTGCA TGAAGATCAAGTTAGATCTGATGCTGCTAGAGAATGGTTGTTGCCAAATTATCAAAGAC CAAATTTGCAAGTTTTGACTGGTCAATATGTTGGTAAAGTTTTGTTGTCTCAAAATGGT ACTACTCCAAGAGCTGTTGGTGTTGAATTTGGTACTCATAAAGGTAATACTCATAATGTT TATGCTAAACATGAAGTTTTGTTGGCTGCTGGTTCTGCTGTTTCTCCAACTATTTTGGAA TATTCTGGTATTGGTATGAAATCTATTTTGGAACCATTGGGTATTGATACTGTTGTTGATT TGCCTGTTGGTTTGAATTTGCAAGATCAAACTACTGCTACTGTTAGATCTAGAATTACTT CTGCTGGTGCTGGTCAAGGACAAGCTGCTTGGTTTGCTACTTTCAATGAAACTTTTGG AGACTATTCTGAAAAAGCTCATGAATTGTTGAATACTAAATTGGAACAATGGGCTGAAG AAGCTGTTGCTAGAGGAGGATTTCACAATACTACTGCTTTGTTGATTCAATATGAAAAT TATAGAGATTGGATCGTCAATCACAATGTTGCTTATTCTGAATTGTTTTTGGATACTGCT GGTGTTGCTTCTTTTGATGTTTGGGATTTGTTGCCATTTACTAGAGGTTATGTTCATATTT GGATTTGTTGGGTCAAGCTGCAGCTACTCAATTGGCTAGAAATATTTCTAATTCTGGTG CTATGCAAACTTATTTTGCTGGTGAAACTATTCCTGGTGATAATTTGGCTTATGATGCTG ATTTGTCTGCTTGGACTGAATATATTCCATATCATTTTAGACCAAATTATCATGGTGTTGG TACTTGTTCTATGATGCCAAAAGAAATGGGTGGTGTTGTTGATAATGCAGCAAGAGTTT ATGGTGTTCAAGGTTTGAGAGTTATTGATGGTTCTATTCCACCAACTCAAATGTCTTCTC ATGTTATGACTGTTTTTATGCTATGGCTTTGAAAATTTCTGATGCTATTTTGGAAGATTA TGCTTCTATGCAATAA

| Primer | Sequence (5'-3') | Note |
|-------------------------|---|--|
| Sc Alpha-m+GOx- Pp F | TCTAGAACTAGTGGATCCCCCATGA GATTTCCTTCAATTTTTAC | Amplification of <i>Angox</i> sequence upstream of T110 codon |
| T110-R | ACCACCATTAACCAAAGTAGAACC ACCCAAACCATTACCAG | |
| T110-F | CTACTTTGGTTAATGGTGGT <u>NNK</u> TG GACTAGACCACATAAAGC | Amplification of <i>Angox</i> sequence downstream of T110 codon |
| Sc GOx-Pp+Alpha- m R | ACTAATTACATGAGGGCCCCCCTT ATTGCATAGAAGCATAATC | |
| F414-R | CAATTCAGAATAAGCAACATTGTGA TTGACGATCCAATCTCTA | Amplification of <i>Angox</i> together with Sc Alpha- m+GOx-Pp F |
| F414-F | CAATGTTGCTTATTCTGAATTG <u>NNK</u> TTGGATACTGCTGGTGTTG | Amplification of <i>Angox</i> together with Sc GOx- Pp+Alpha-m R |
| Pp-GOx-Pp F | AGGCTGAAGCTTACGTAGAATTCTC TAATGGTATTGAAGCTTCTTTG | Amplification of <i>Angox</i> sequence upstream of T110V codon |
| T110V R | CTTTATGTGGTCTAGTCCA <u>AAC</u> ACC ACCATTAACCAAAGTAG | |
| T110V F | CTACTTTGGTTAATGGTGGT <u>GTT</u> TG GACTAGACCACATAAAG | Amplification of <i>Angox</i> sequence downstream of T110V codon |
| Pp-GOx-Pp R | TTAATTCGCGGCCGCCCTAGGGTTA TTGCATAGAAGCATAATC | |
| F414L R | GCAACACCAGCAGTATCCAACAA <u>C</u> <u>AA</u> TTCAGAATAAGCAAC | Amplification of <i>Angox</i> together with Pp-GOx-Pp F |
| F414L F | GTTGCTTATTCTGAATTGTTG <u>TTG</u> G ATACTGCTGGTGTTGC | Amplification of <i>Angox</i> together withPp-GOx-Pp R |

Table S1. Primers used in this study.



Fig. S1. Mutagenesis of AnGOx based on the comparison with AfGDH. (A) Amino acid variations between AnGOX (cyan) and AfGDH (green) around the substratebinding pocket. (B) $\Delta OD_{500 \text{ nm}}$ values in the D-xylose oxidase assay of the culture supernatants of *Saccharomyces cerevisiae* strains expressing AnGOx mutants. Data represent the mean \pm SD from duplicate cultivations.



Fig. S2. SDS-PAGE analysis of purified enzymes.



Fig. S3. Plots for the determination of kinetic parameters of enzymes. The Hill equation was used to fit the plots. (A–C) WT, T110V and T110V/F414L on D-xylose, respectively. (D–F) WT, T110V and T110V/F414L on D-glucose, respectively.



Fig. S4. Thermostabilities of AnGOx (WT) and the T110V/F414L mutant. The purified proteins were incubated at different temperatures for the indicated time, and the activities towards D-xylose (A, B) and D-glucose (C, D) were measured at pH 6.0 and 35 °C. Data represent the mean \pm SD from duplicate measurements.



Fig. S5. Time evolution of RMSD for AnGOx, T110V, and T110V/F414L with D-xylose.



Fig. S6. Changes in the RMSF values of all residues (A) and active-site residues (B) of T110V and T110V/F414L relative to the wild-type AnGOx. The data were analyzed using the simulations with D-xylose.



Fig. S7. Front view of the substrate pocket of AnGOX. The D-glucose oxidation product LGC was placed in the model by superimposing the structure of AnGOX (PDB: 1CF3) onto that of the AfGDH-LGC complex (PDB: 4YNU). Sites A (cyan) and B (pink) defined in the text are shown. (A) Stick representation. (B) Surface representation.



Fig. S8. Distribution of distances between the O1 atom of D-glucose and catalytic histidine residues of AnGOx (A), T110V (B), and T110V/F414L (C), during MD simulations.



Fig. S9. ¹H NMR spectra of commercial lithium D-xylonate as a standard (A) and purified D-xylonate product (B).