

1 Supplementary Information for

2 **A Low-temperature Digital Microfluidic System Used for**  
3 **Protein-Protein Interaction Detection**

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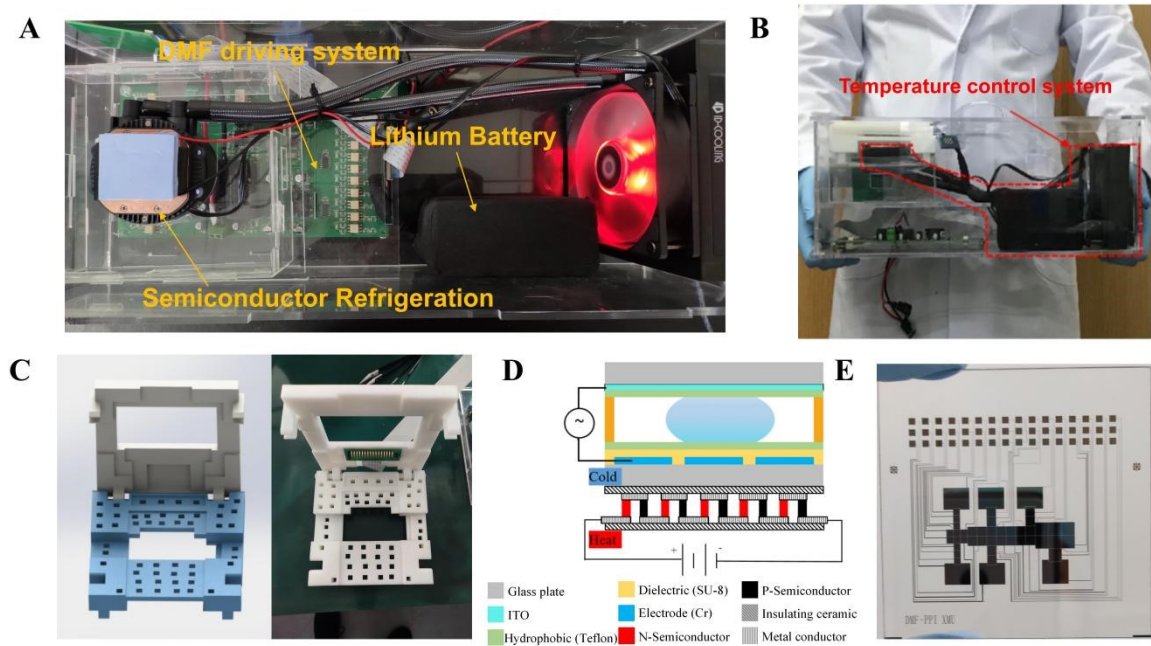
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32 **1. LTDMF-PPI-Box**

33 LTDMF-PPI-Box is mainly composed of two parts, digital microfluidic system and  
34 temperature control system. The digital microfluidic system includes: DMF driven  
35 system, magnet, LTDMF-PPI chip and holder (Fig. S1). Table. S1 presents the  
36 performance parameters of TEC. Fig. S2 shows the structure and dimensions of  
37 LTDMF-PPI-Box.



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39 Fig. S1 (A) DMF driving system. (B) The overview of temperature control system.

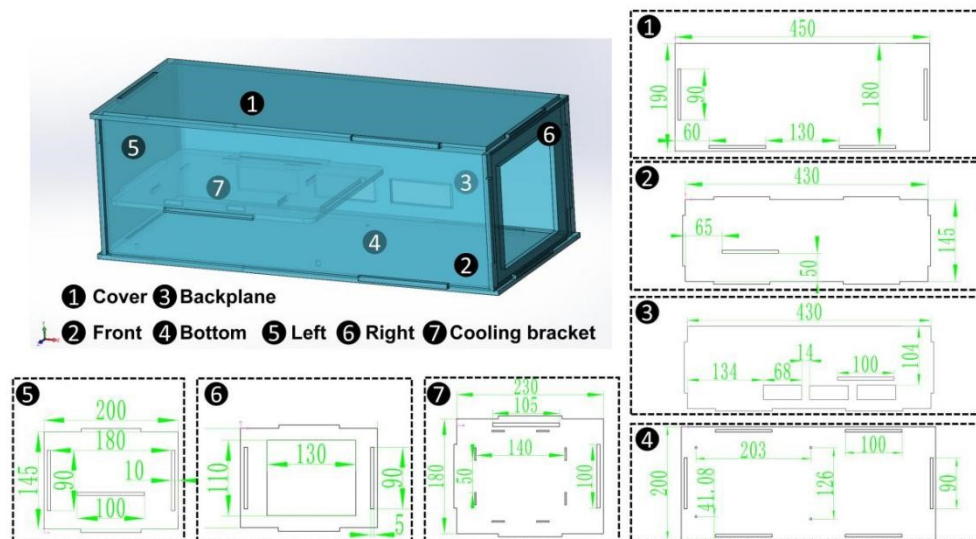
40 (C) The holder for LTDMF-PPI chip. (D) The structure diagram of the realization of

41 LTDMF-PPI chip. (E) LTDMF-PPI chip.

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performance parameters	Values
Model number	TEC1-12706
Operating temperature range	-50 °C ~ 80 °C
Maximum working current / $I_{\max}$	6.0 A
Maximum temperature difference/ $\Delta T_{\max}$ ( $Q_c = 0$ )	68 °C
Maximum operating voltage / $V_{\max}$	15.4 V
Maximum refrigerating power / $Q_{\max}$	59.1 W
Internal resistance/ $R_i$	1.0 ~ 1.1 $\Omega$



## 48 2. Fabrication of LTDMF-PPI chip

49 Fig. S3 shows the manufacturing flow chart for the bottom plate of LTDMF-PPI  
50 chip. The detailed process is described as follows:

51 a. A piranha solution (concentrated sulfuric acid:hydrogen peroxide = 3:1) was used  
52 to clean glass substrate.

53 b. A chromium layer (400 nm in depth) was grown on the glass substrate by  
54 magnetron sputtering.

55 c. A photoresist (AZ5214E) was coated on the chromium layer, exposed and  
56 developed.

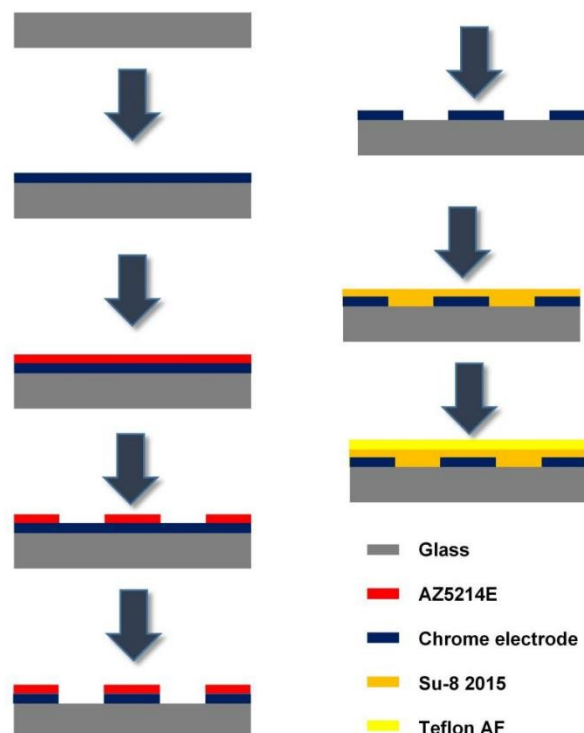
57 d. The exposed chromium was etched by the etching solution.

58 e. After etching, the photoresist was removed by the acetone.

59 f. A photoresist (SU-8 2015) was coated on the substrate, exposed and post-baked.

60 g. The Teflon AF (~200 nm in depth) was coated on the substrate.

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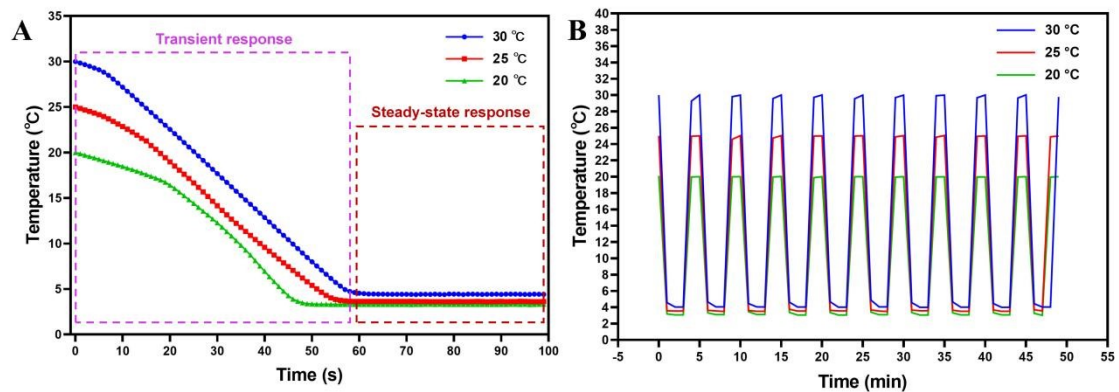
63 Fig. S3 Manufacturing flow chart for the bottom plate of LTDMF-PPI chip.

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### 66 3. The low-temperature capability of LTDMF-PPI-Box

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69 Fig. S4 The temperature curves of low-temperature control system. (A) Transient  
70 response and steady-state response, (B) Stability curve.

71

### 72 4. Cell transfection

73 Cell transfection experiments were carried out by using 293T human renal  
74 epithelial cells with high transfection efficiency. All operations were completed in a  
75 cell room. The process is described as follows:

76 (1) The condition of cells was observed under a microscope.

77 (2) 100  $\mu$ L of cell culture medium without serum and 1  $\mu$ g of plasmid vector were  
78 added to the centrifuge tube and shaken to mix.

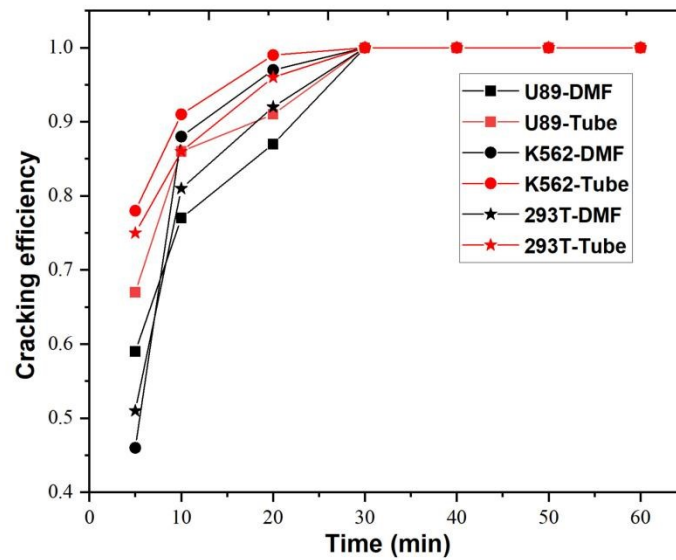
79 (3) 100  $\mu$ L of cell culture medium without serum and 5  $\mu$ L of PEI transfection reagent  
80 were added to another centrifuge tube, and shaken to mix.

81 (4) After standing the above two centrifuge tubes for 5 min, added the PEI suspension  
82 into the plasmid carrier suspension and let it stand for 20 min.

83 (5) Replaced the medium of the transfected cells with a serum-free medium, and then  
84 added the transfection mixture into the cell culture dish.

85 (6) The cells were cultured in an incubator for 8 h and then replaced with the  
86 complete medium. Then confirmed the transfection efficiency of the cell under the  
87 fluorescence microscopy.

## 88 5. Optimization of cell lysis time for various cell types



89

90 Fig. S5 The lysis efficiency of U89, K562 and 293T on LTDMF-PPI

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## 92 6. Co-immunoprecipitation

93 Co-immunoprecipitation is a classic method to study PPI, which bases on the  
94 specific binding between antibodies and antigens. It is also the most convincing  
95 method in routine experiments. Beads used in Co-immunoprecipitation are usually  
96 protein A/G, which are not magnetic and cannot be used in LTDMF-PPI chips. The  
97 specific process of the improved immunoprecipitation method is described as follows:

98 (1) The fusion protein Myc-Rab26 and GFP-RILP were expressed in 293T cells by  
99 transfection experiment.

100 (2) The cells were lysed to obtain Myc-Rab26 protein and GFP-RILP protein, and  
101 then the 5% lysis mixture was used as control.

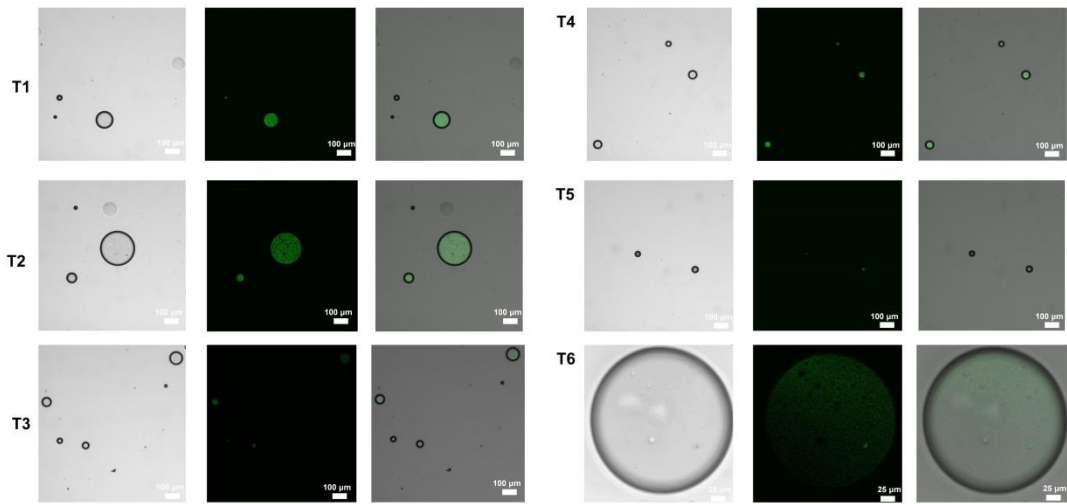
102 (3) The lysis mixture containing the fusion protein Myc-Rab26 and GFP-RILP were  
103 mixed with anti-Myc coated magnetic beads at 4 °C for 8 hours.

104 (4) The magnetic beads were washed 3 times with washing buffer.

105 (5) 20  $\mu$ L SDS loading buffer were added in the above magnetic beads, and mixed for  
106 5 minutes at 95 °C. Then western blot was carried to confirm whether there was an  
107 interaction between the two protein molecules.

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109 **7. Repeated experimental results**



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111 Fig. S6 Images on the interaction between Myc-Rab26 and GFP-RILP protein on  
112 LTDMF-PPI chip. (Left column: images from bright field, Middle column:  
113 fluorescent images, Right column: overlapped images)