

Supporting Information

**Smart salt-responsive thread for highly sensitive
microfluidic glucose detection in sweat**

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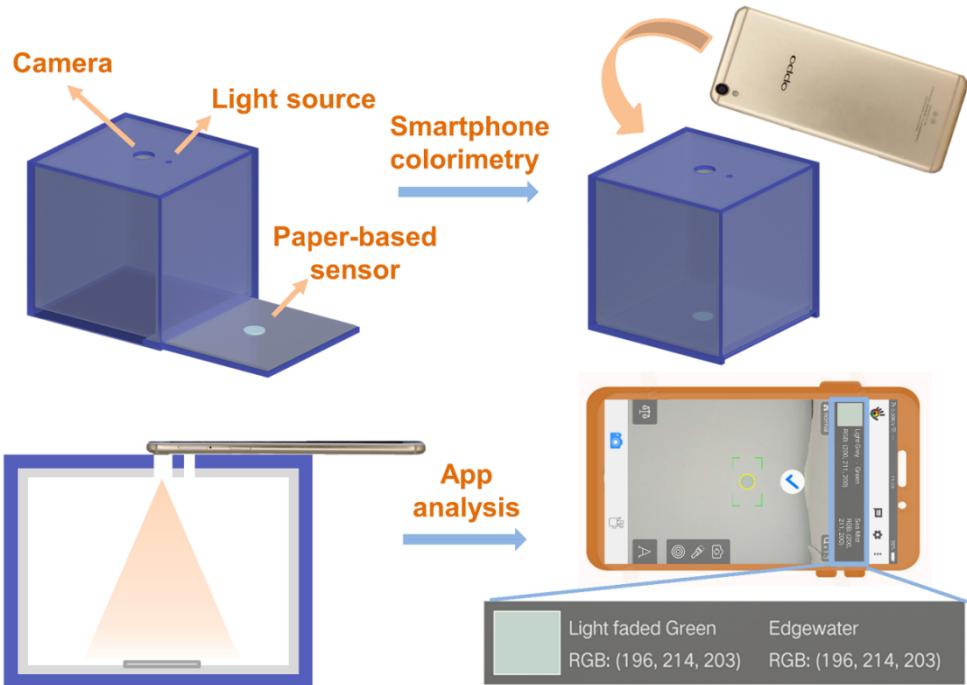


Figure S1. Smartphone-based data collecting system.

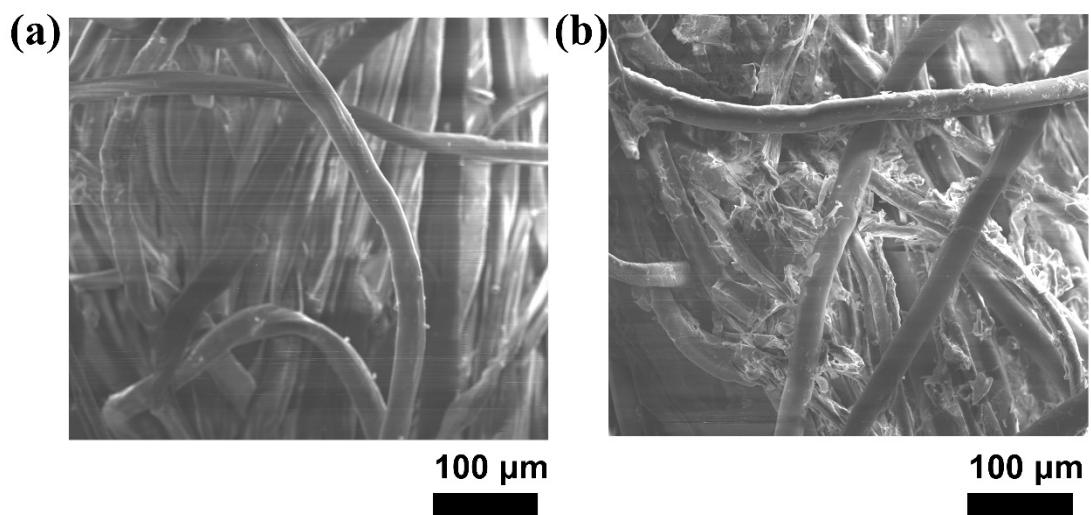


Figure S2. SEM images of the pristine (a) and PSBMA-modified (b) cotton threads.

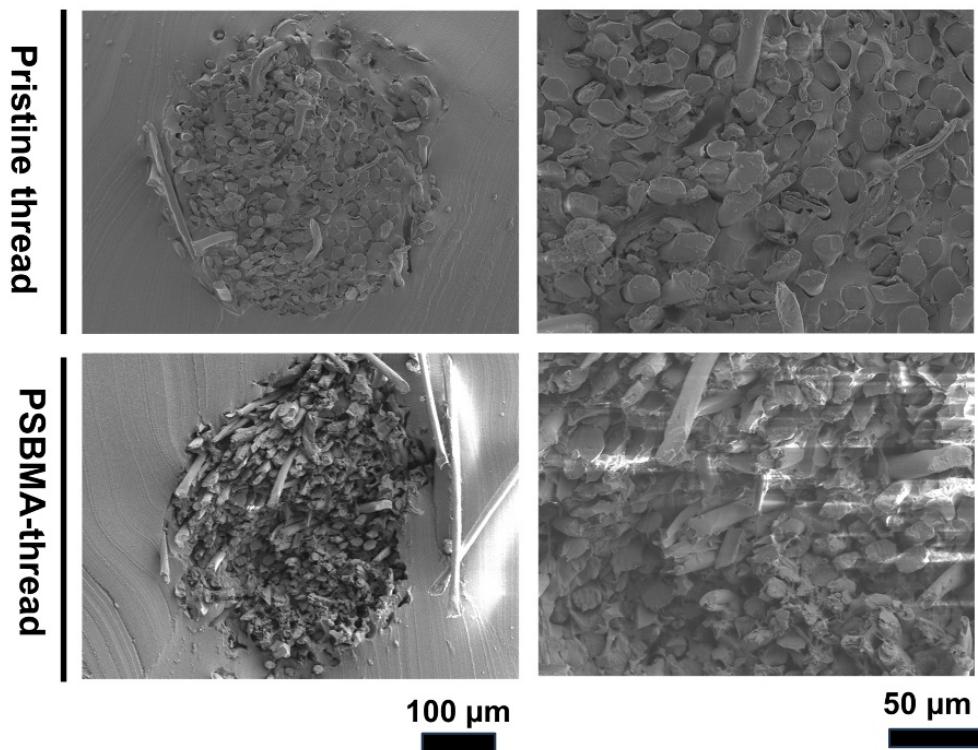


Figure S3. Cross-sectional SEM images of the pristine thread and PSBMA-thread.

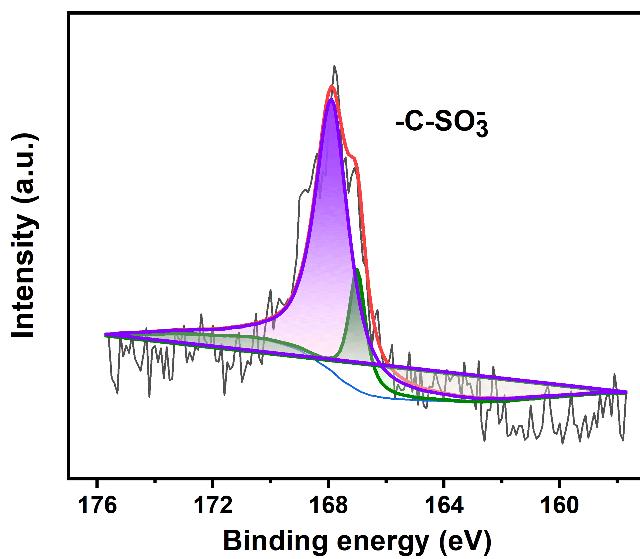


Figure S4. XPS S2p spectra of PSBMA-thread.

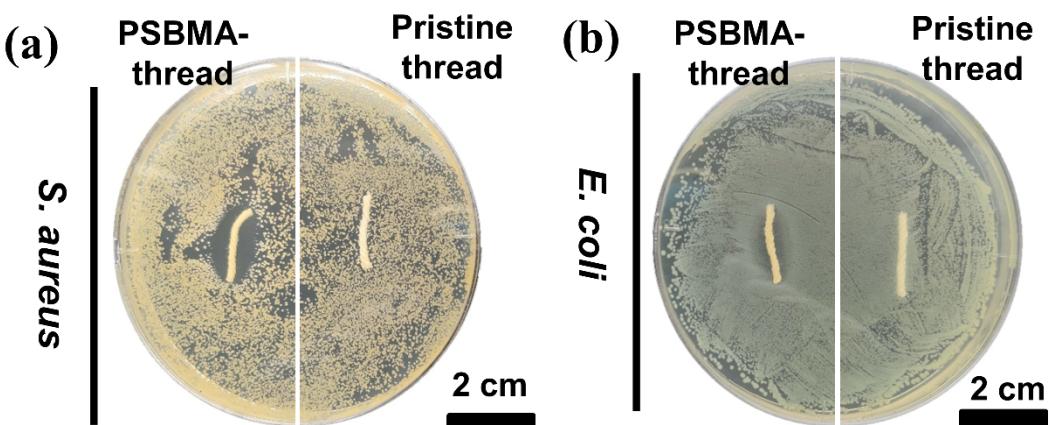


Figure S5. Antibacterial effects of pristine cotton thread and PSBMA-modified thread.

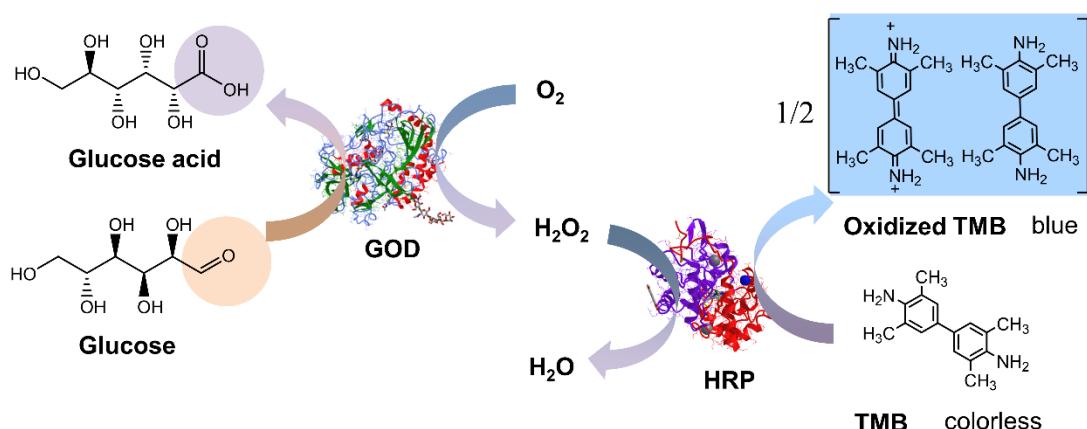


Figure S6. Working principle of the GOD/HRP/TMB-based colorimetric sensors.

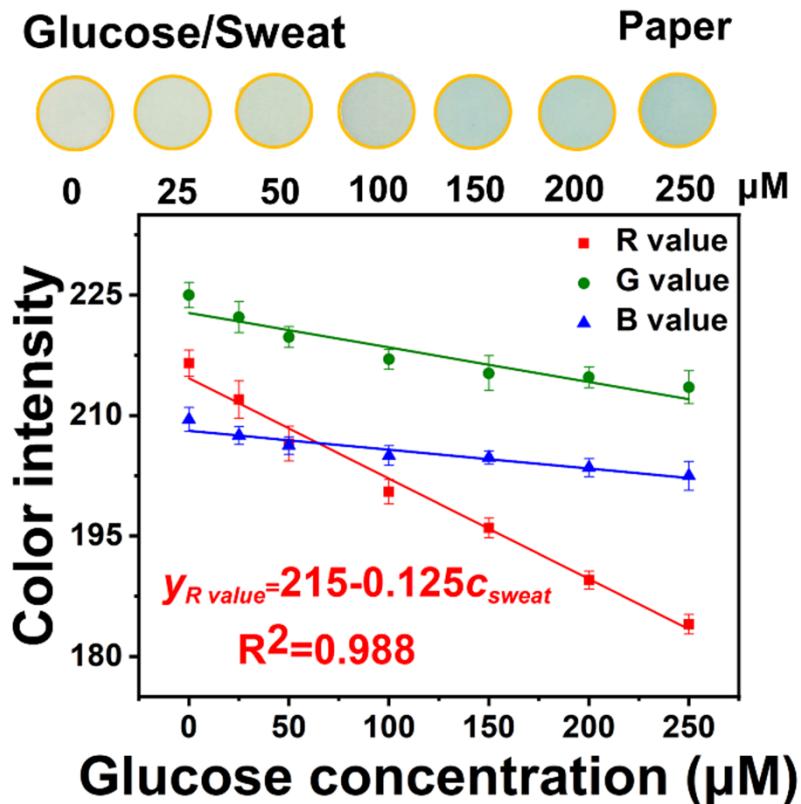


Figure S7. Performance of the paper-based colorimetric glucose sensor.

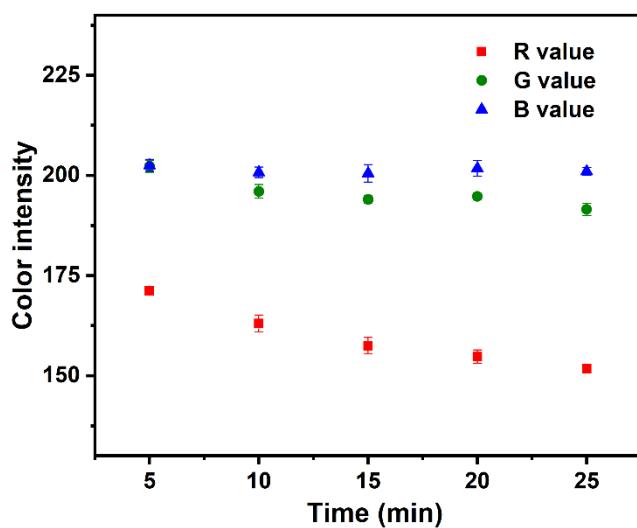


Figure S8. Effects of the reaction duration on the color intensity of the paper-based sensors.

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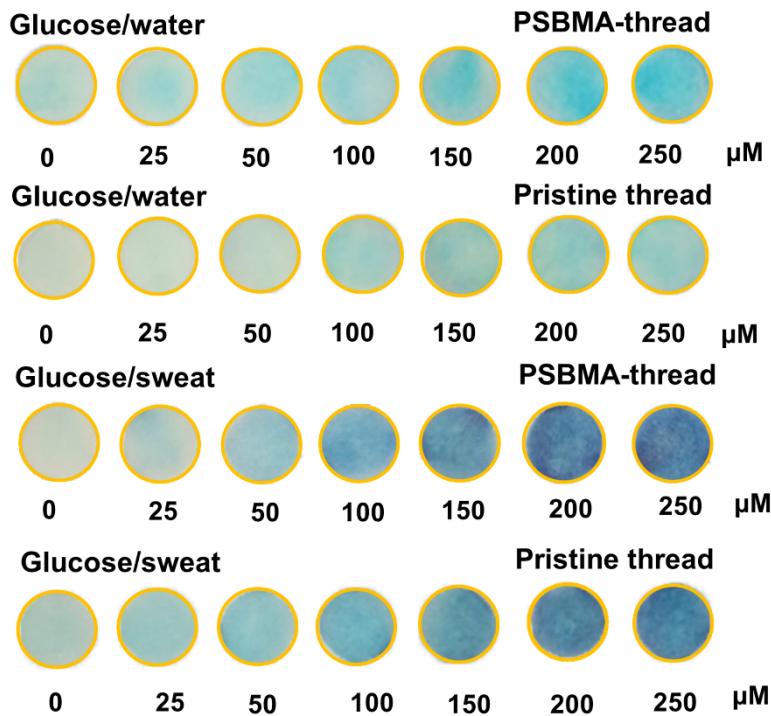


Figure S9. Photos of the corresponding paper sensing units after glucose sensing.

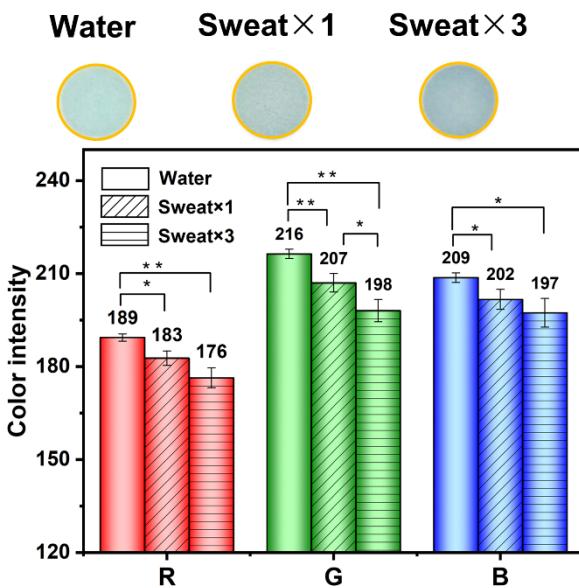


Figure S10. Effects of the ionic strength on the color intensity of the paper-based sensors.

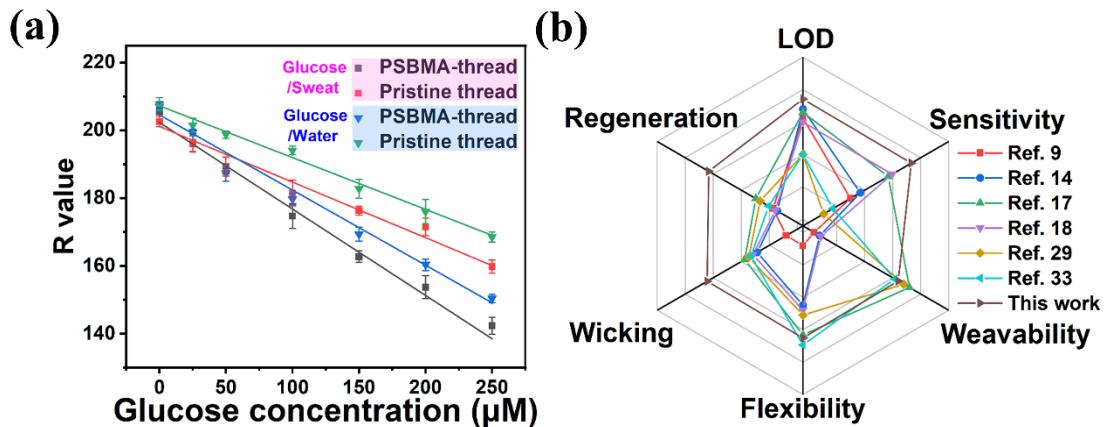


Figure S11. Performance of the microfluidic thread/paper-based glucose sensing systems in different solutions (a). A comparison of the performance of glucose sensors for body fluid (b).

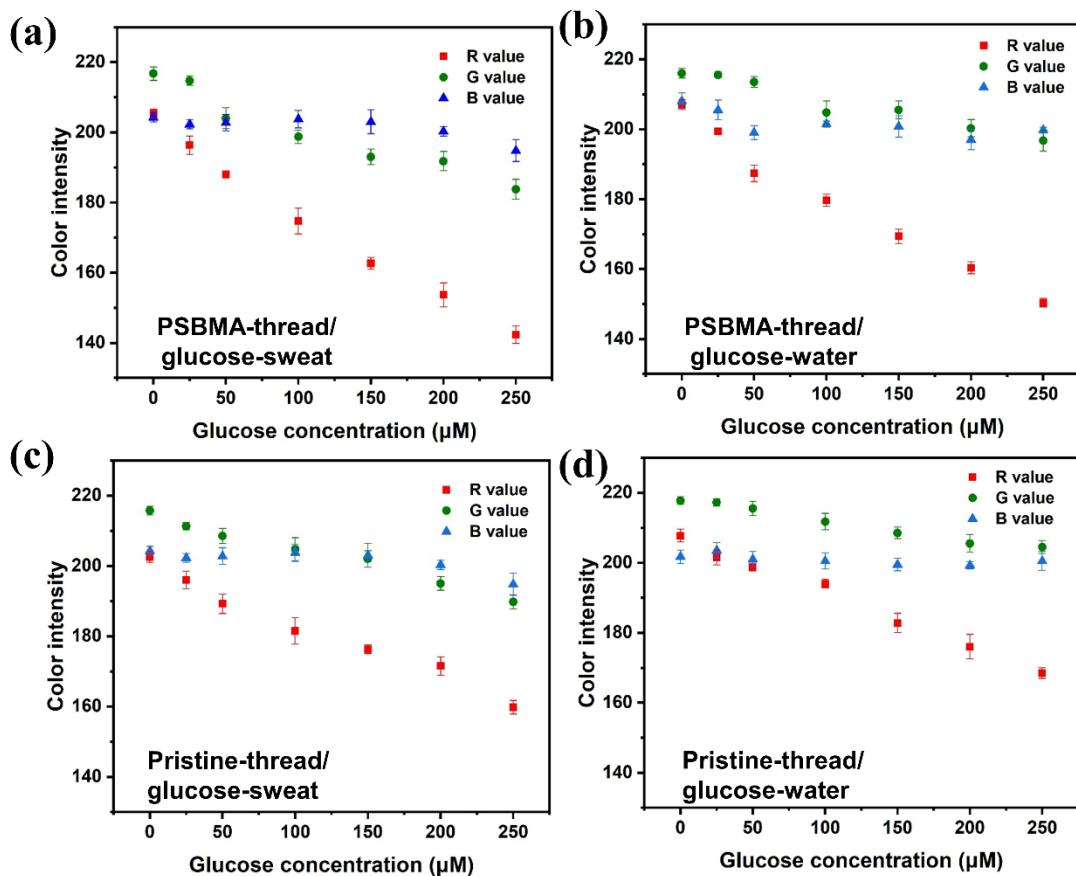


Figure S12. Color intensity against glucose concentration in terms of the R-, G-, and B-value.

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Table S1. Parameters for the calibration curves.

	PSBMA-thread/glucose-water	Pristine-thread/glucose-water	PSBMA-thread/glucose-sweat	Pristine-thread/glucose-sweat
Intercept	205	207	202	201
Adj. R-Square	0.981	0.984	0.989	0.986
LOD/ μM	19.1	35.1	14.7	27.4
Sensitivity/ μM^{-1}	-0.222	-0.153	-0.255	-0.164
¹				

Table S2. Recovery ratio of the thread-based microfluidic sensors.

Thread	Spiked concentration/ μM (glucose)	μTPAD assay (μM) (mean \pm SD, n=3)	Recovery/% (mean \pm SD, n=3)	Coefficient of variation/% (CV)
PSBMA-thread	50	48.7 \pm 0.4	97.4 \pm 0.7	0.8
	150	150.3 \pm 1.5	100.2 \pm 1.0	1.0
	250	247.8 \pm 1.8	99.1 \pm 0.7	0.7
Pristine thread	50	49.5 \pm 0.6	99.0 \pm 1.1	1.1
	150	148.4 \pm 1.2	98.9 \pm 0.8	0.8
	250	246.3 \pm 1.5	98.5 \pm 0.6	0.6

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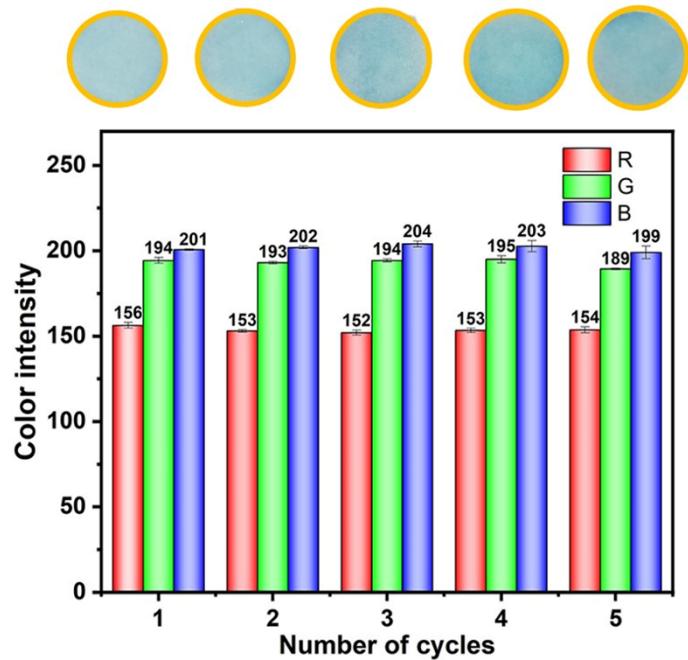


Figure S13. Regeneration of the PBMSA-modified threads in the microfluidic system.

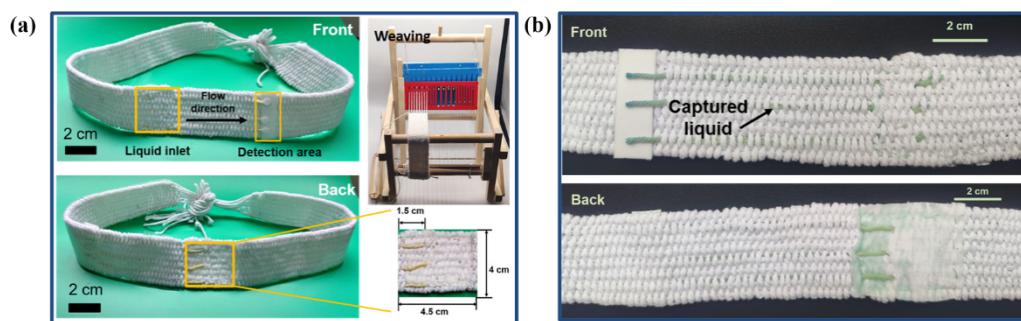


Figure S14. (a) Preparation of the glucose sensing headband; (b) Photos of the headband worn on the forehead during exercise.

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Table S3. Summary of glucose colorimetric sensors in body-fluid analysis

Materials and Substrate	Enzyme and chromogenic reagent	LOD	Detection range	Sensitivity	Test sample	Reference
PDMS	GOD/HRP/o-dianisidine	30 μM	0.1–0.5 mM	111.23 mM^{-1}	Sweat	1
PDMS	GOD/AuNCs/ODA	21 μM	0.05–1.6 mM	0.469 dec(mM) $^{-1}$	Sweat	2
BC/CMC hydrogel	GOD/HRP/KI	25 μM	0.025–0.5 mM	Not given	Sweat	3
PVA/sucrose hydrogel	GOD/HRP/4-aminoantipyrine	Not given	0–2 mM	Not given	Sweat	4
TiO ₂ nanotubes/alginate hydrogel	GOD/HRP/TMB	44 μM	0.1–0.8 mM	83.678 mM^{-1}	Sweat	5
Cotton cloth	GOD/HRP/TMB	1700 μM	3.0–15.0 mM	53.963 dec(mM) $^{-1}$	Serum	6
Filter paper	GOD/HRP/TBHBA/4-AAP	300 μM	1.0–11.0 mM	0.5238 mM^{-1}	Serum	7
Filter paper	GOD/HRP/TMB	14 μM	0.02–4.0 mM	108.06 dec(mM) $^{-1}$	Serum /Tear	8
Filter paper	GOD/HRP/4-AAP/DHBS	50 μM	0.05–0.3 mM	200.9 mM^{-1}	Sweat	9
Cotton thread	GOD/HRP/KI	100 μM	0.1–5.0 mM	8.28 mM^{-1}	Tear	10
Cotton thread	GOD/HRP/KI	100 μM	0.1–3 mM	34.455 mM^{-1}	Sweat	11
Cotton thread/paper	GOD/HRP/TMB	35 μM	0.05–0.25 mM	190 mM^{-1}	Sweat	12
PSBMA thread/paper	GOD/HRP/TMB	14.7 μM	0.025–0.25 mM	255 mM^{-1}	Sweat	This work

Reference

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