

## Supplementary information

### Triblock copolymer micelles enhance solubility, permeability and activity of a quorum sensing inhibitor against *Pseudomonas aeruginosa* biofilms

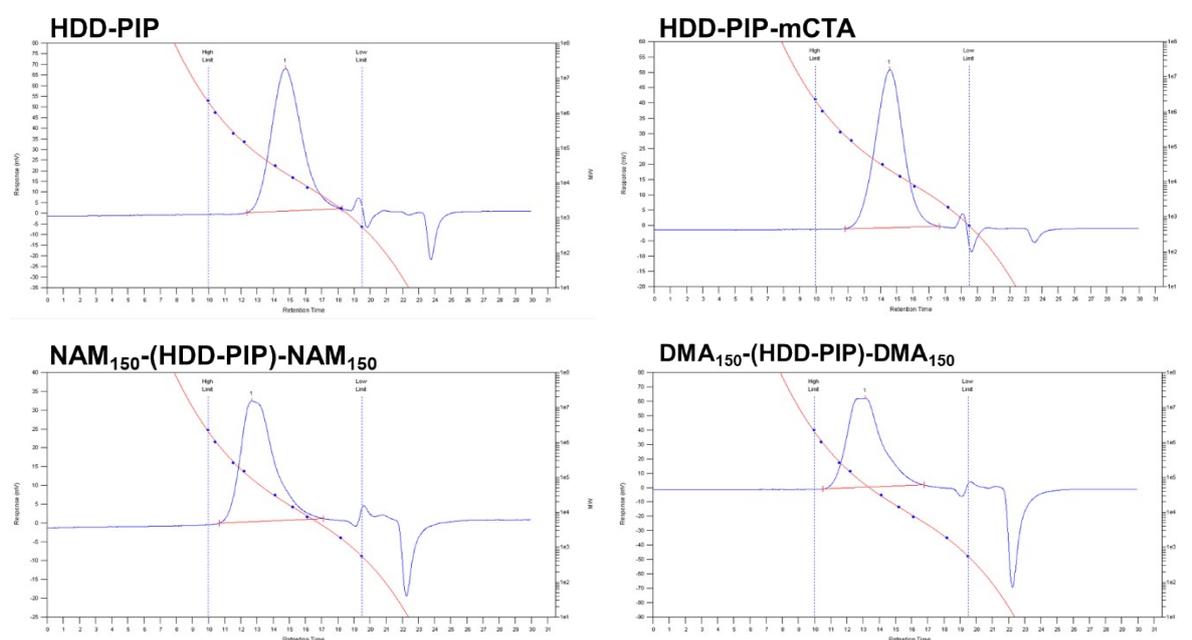
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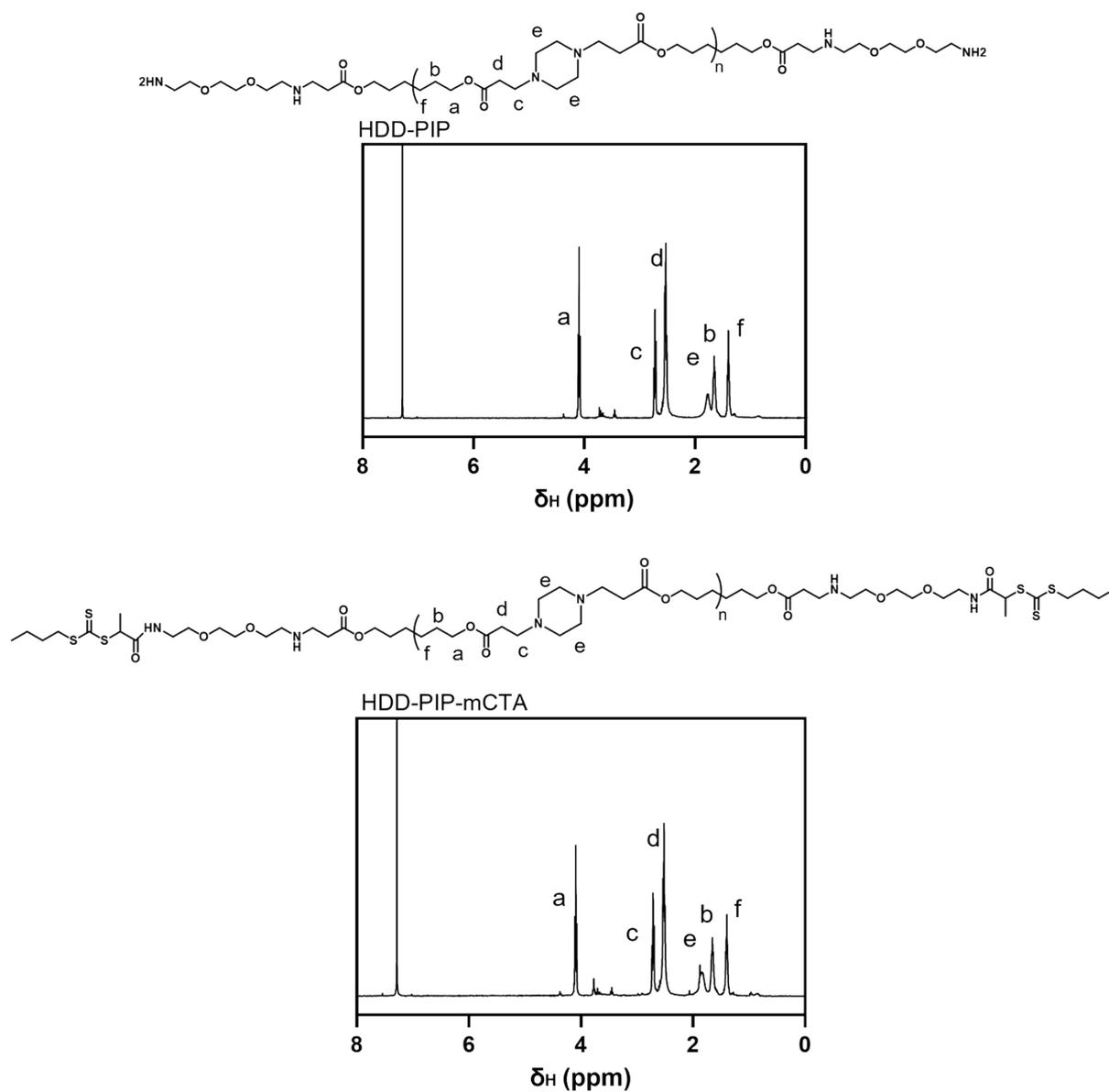
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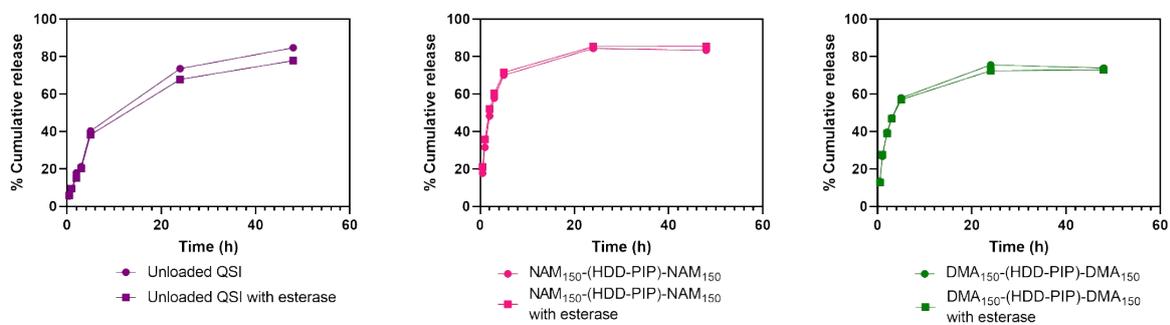
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**Figure S1:** DMF-SEC chromatograms of prepared polymers.

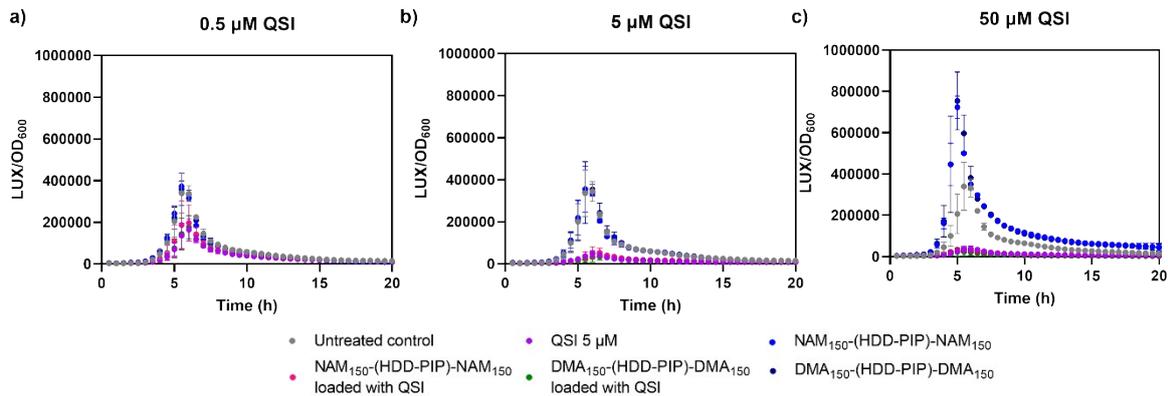


**Figure S2:**  $^1\text{H}$  NMR spectra of polymers in chloroform-d recorded at 25°C with a Bruker NMR400 MHz.

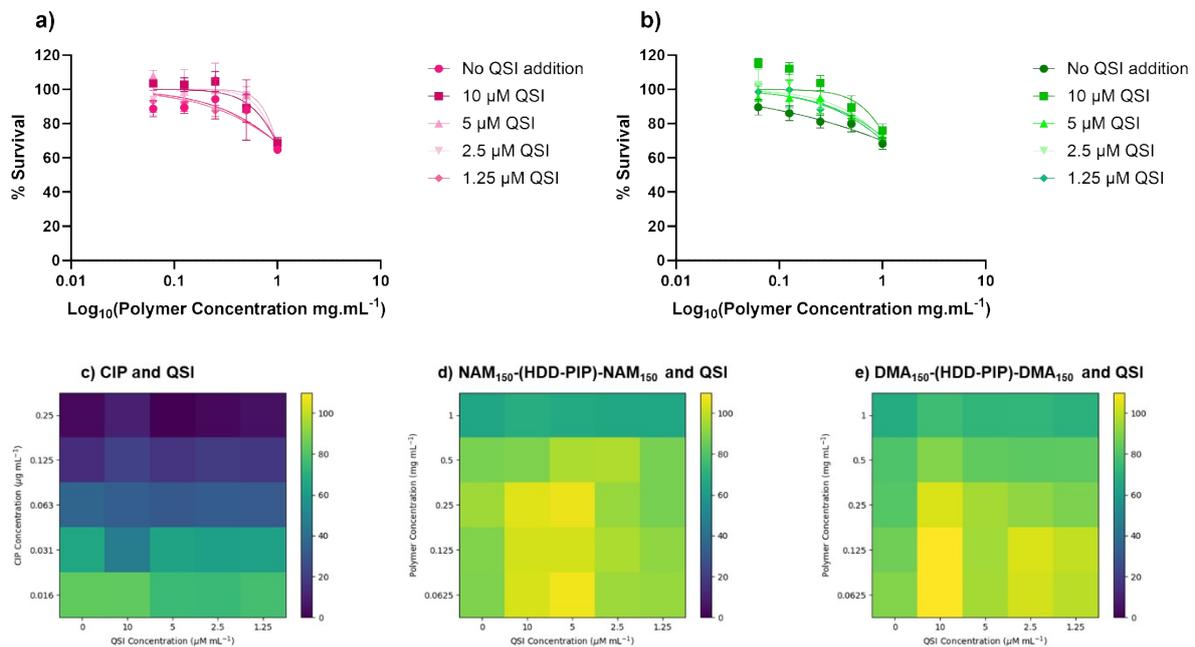


**Figure S3:** QSI release from  $\text{NAM}_{150}$ -(HDD-PIP)- $\text{NAM}_{150}$  (pink) and  $\text{DMA}_{150}$ -(HDD-PIP)- $\text{DMA}_{150}$  (green) particles and unloaded QSI (purple) in phosphate buffer pH 7.4, containing

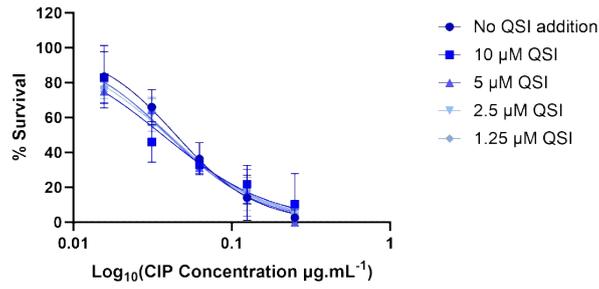
0.01% Tween 20, at 37°C, across 48 h, with and without esterase presence samples analysed by HPLC.



**Figure S4:** Effect of QSI and QSI loaded polymers on a  $P_{pqSA}$ -*lux* transcriptional fusion, which reports the PQS-dependent activation of the *pqs* operon mediated by PqsR. **a)** QSI concentrations at 0.5 μM QSI and non-loaded particles at corresponding concentrations; **b)** QSI concentrations at 5 μM QSI and non-loaded particles at corresponding concentrations; **c)** QSI concentrations at 50 μM QSI and non-loaded particles at corresponding concentrations. Values given are averages from three different cultures for QSI activity testing and two different cultures for the testing of non-loaded polymeric particles and correspond to the relative light units normalized to culture density (Lux/OD<sub>600</sub>) over time (20 h).

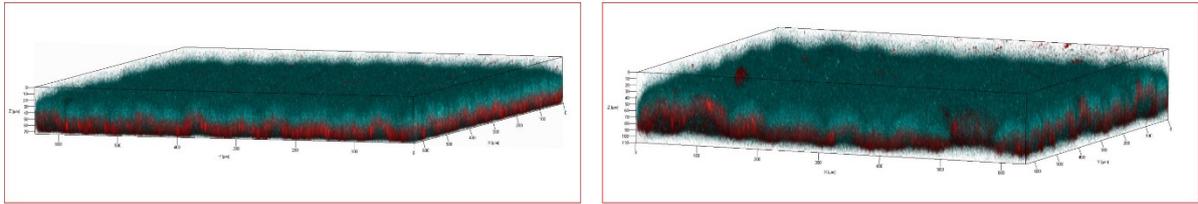


**Figure S5:** Antimicrobial activity of polymeric particles with and without QSI61 addition. **a)** Concentration curves of NAM<sub>150</sub>-(HDD-PIP)-NAM<sub>150</sub> (pink) with and without QSI61 addition against planktonic *P. aeruginosa*; **b)** Concentration-activity curves of DMA<sub>150</sub>-(HDD-PIP)-DMA<sub>150</sub> (green) with and without QSI61 addition against planktonic *P. aeruginosa*. Heat maps to show effects of combinations of **c)** CIP + QSI, **d)** NAM<sub>150</sub>-(HDD-PIP)-NAM<sub>150</sub> and QSI, and **e)** DMA<sub>150</sub>-(HDD-PIP)-DMA<sub>150</sub> and QSI.

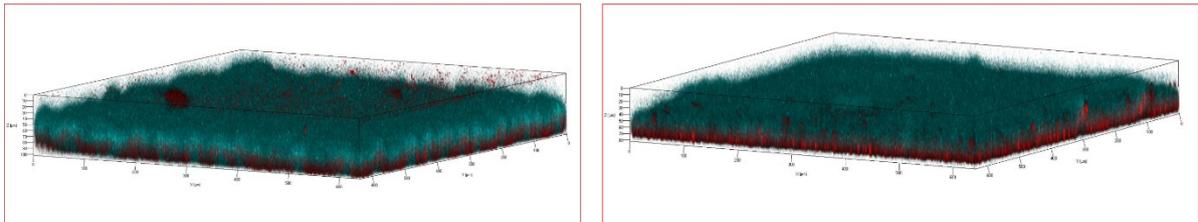


**Figure S6:** CIP concentration-activity curves with and without QSI61 addition against planktonic *P. aeruginosa*.

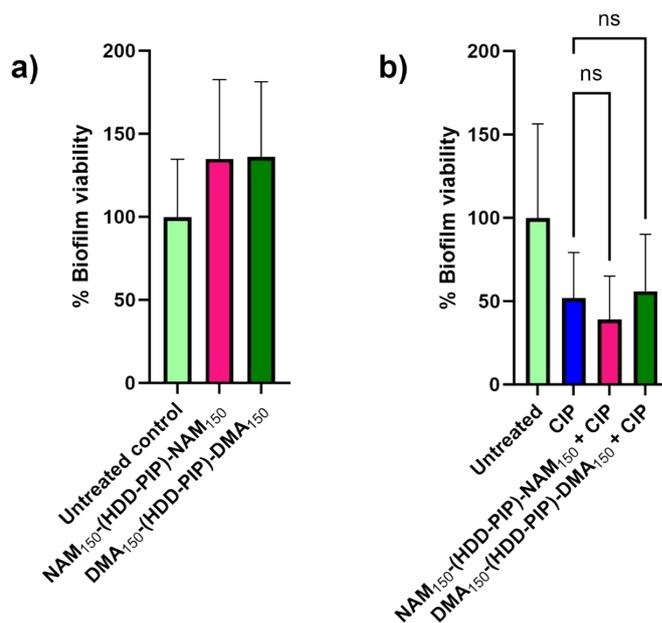
a)



b)



**Figure S7:** Confocal laser scanning microscopy (CLSM) images the biofilm penetration of rhodamine-tagged DMA<sub>150</sub>-(HDD-PIP)-DMA<sub>150</sub> (a) and NAM<sub>150</sub>-(HDD-PIP)-NAM<sub>150</sub> particles (b) (red) in mature *P. aeruginosa* PA01L biofilms (1-day old) stained with SYTO9 (blue) following a 4 h incubation.



**Figure S8:** Antimicrobial activity of polymeric particles in *P. aeruginosa* biofilms; **a)** Bar charts showing viability in mature *P. aeruginosa* biofilms, quantified after treatment with NAM<sub>150</sub>-(HDD-PIP)-NAM<sub>150</sub> (pink) and DMA<sub>150</sub>-(HDD-PIP)-DMA<sub>150</sub> (green) particles with no encapsulated QSI; measurements were performed in duplicate, using biologically independent replicates and the error bars represent the mean  $\pm$  standard deviation; **b)** Bar charts showing viability in mature *P. aeruginosa* biofilms, quantified after treatment with CIP (60  $\mu\text{g mL}^{-1}$ ) and NAM<sub>150</sub>-(HDD-PIP)-NAM<sub>150</sub> (pink) and DMA<sub>150</sub>-(HDD-PIP)-DMA<sub>150</sub> (green) particles with no encapsulated QSI administered in combination with CIP (60  $\mu\text{g mL}^{-1}$ ); measurements are an average of 15 coverslip measurements using one biological replicate and the error bars represent the mean  $\pm$  standard deviation. Statistical testing was performed with a one-way ANOVA followed by a post-hoc Tukey test to identify individual comparisons. Statistical significance is represented as \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ , \*\*\*\* $p < 0.0001$ .