Electronic Supplementary Material (ESI) for Materials Advances. This journal is © The Royal Society of Chemistry 2024

## Supplementary material

\*: <sup>1</sup>H peak from N-hydroxysuccinimide

2.5

Fig. S1. Synthesis scheme of TCDA-NHS and corresponding <sup>1</sup>H NMR analysis results

ppm

2.0

<u>--</u> 0.5

Composition (mole ratio)	CR (%) 70°C@20min	Photo	note
PCDA:PCDA-NHS:DMPC (0.9:0.1:0.6)	20.1%		
TCDA:TCDA-NHS:DMPC (0.9:0.1:0.6)	37.9%		N
TCDA:TCDA-NHS:DMPC (0.9:0.1:0.3)	25.2%		No aptamer
TCDA:TCDA-NHS:DMPE (0.9:0.1:0.6)	28.0%		
Он			
PCDA			TCDA
0-N	~~~	~~~**	
PCDA-NHS		-	ГCDA-NHS
O HO O P	^	~~~~	O H
DMPE			DMPC

**Table S1.** Heat Stimulated colorimetric response of different liposome composition and chemical structure of each components.

Liposomes of different compositions were prepared by the bath method to the same final concentration of 1 mM, and UV polymerization was performed without aptamer attachment. The sensitivity of each liposomal material was evaluated through a thermal colorimetric response after a 20-minute incubation at 70 °C. The extent of color alteration was quantified as CR (%), and the specific calculation method is explained in the manuscript.

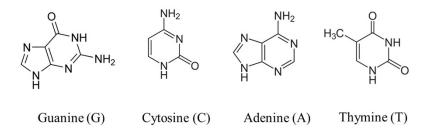
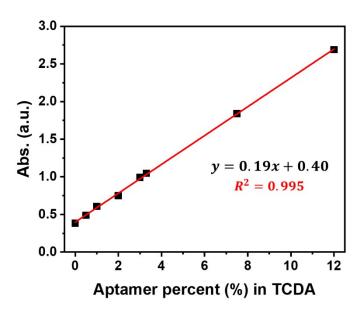
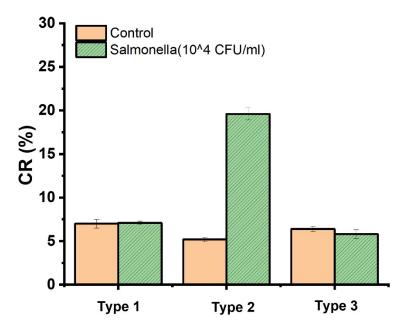


Fig. S2. Chemical structure of Guanine, Cytosine, Adenine, and Thymine



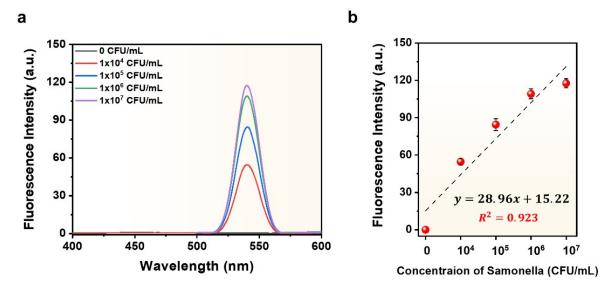
Abs.	Aptamer feeding concentration (%)			n (%)
(260 nm)	1	3	7.5	12
(a) Aptamer conc. In TCDA liposome solution	0.6084	0.9810	1.8399	2.690
(b) After dialysis TCDA liposome solution	0.4830	0.7722	1.5195	2.155
TCDA liposome conjugated aptamer %	79.4	78.8	82.6	80.1

**Fig. S3.** To generate a master curve, aptamer conjugation onto TCDA liposomes is performed across different concentrations (1%, 3%, 7.5%, and 12%). Using 100-500 Dalton dialysis membrane tubing, the mixture is subjected to water exchange and stirring over 3 days, ensuring that only TCDA liposomes remain. After verifying the absorbance of the obtained solution through UV-vis spectroscopy, aptamer efficiency is determined by comparing it with the absorbance of the TCDA liposome-aptamer solution.



Sensor	Aptamer	Salmonella (CFU/mL)	CR (%)
Type 1	N	0	7.0
	No aptamer	104	7.1
Type 2 Salmonella Specific aptamer	Salmonella	0	5.2
	104	19.6	
Type 3 Non-specific aptamer	Non-specific	0	6.4
	aptamer	$10^{4}$	5.8

Fig. S4. Salmonella -specific sensing properties of PDA liposome sensors depending on aptamer type



**Fig. S5.** (a) Fluorescence spectra for different *Salmonella* concentrations from  $1x10^4$ ,  $1x10^5$ ,  $1x10^6$ ,  $1x10^7$  CFU/mL in eggs. (b) Calibration curve and linearity results for fluorescence intensity after 15 minutes of exposure to different *Salmonella* concentrations. (ranging from 500 to 600 nm)