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## **Supplemental Materials**



**Fig. S1** Possible targets for disulfide engineering as predicted by the web-server Yosshi. The distances between the amino acids in loops C and D are indicated by dashed lines. (A) T49-T78, (B) K55-I75, and (C) A51-T78.



**Fig. S2** Overlay of the simulated structure of A51C/G77C hCyt c (blue) with the solution structure of WT hCyt c (PDB code 2N9J, gray) showing the conformational changes upon formation of the disulfide bond Cys51-Cys77.



**Fig. S3** (A) SDS-PAGE of the purified WT and A51C/G77C hCyt c. The protein marker was shown for comparison. ESI-MS spectra of the purified (B) WT hCyt c, (C) A51C/G77C hCyt c and (D) treated with reducing agent TCEP.



**Fig. S4** Fluorescence spectra of WT hCyt c varied with the concentration of guanidine hydrochloride (0-6 M).



**Fig. S5** (A) pH-dependent UV-vis spectra changes of the WT hCyt c in acidic unfolding studies; (B) Plot of 695 nm versus pH for acid unfolding of WT and A51C/G77C hCyt c; (C) pH-dependent UV-vis spectra changes of the WT hCyt c in alkaline transition studies.



Fig. S6 Time-dependent UV-vis spectra of WT hCyt c in a reaction with 100 mM H<sub>2</sub>O<sub>2</sub>. The spectral change of the Soret band was shown as an inset.



**Fig. S7** Peroxidase activity assay using ABTS as a substrate. Stopped-flow kinetic of ABTS (0.1 mM) catalyzed by (A) A51C/G77C and (B) WT *h*Cyt *c* in the presence of H<sub>2</sub>O<sub>2</sub>. (C) Michaelis–Menten plots vs the concentrations of ABTS for WT *h*Cyt *c* at pH 5–8.