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# **Supplementary Information**

# Visible-light-promoted C(sp<sup>3</sup>)-H thiolation of aliphatic ethers with

# thiosulfonates

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### 1. General information of reagents and instruments

All reagents and solvents were purchased from commercial suppliers without further purification. All reactions were carried out in borosilicate glass vessels in a photo reactor manufactured by Beijing Roger Technology Co., Ltd. And, all reactions were performed under the irradiation of 5 W blue LED light without using filters (Figure S1, left). The blue light's energy peak wavelength is 452.6 nm, peak width at half-height is 21.6 nm and the irradiance@5W is 146.2 mW/cm<sup>2</sup> (Figure S1, right). LED irradiate through a high-reflection channel (path length is 2 cm) to the test tube. The progress of the reactions were monitored by TLC (thin-layer chromatography) under 254 nm UV light. Products were purified by chromatography on 200-300 mesh silica gels. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded in CDCl<sub>3</sub> at room temperature ( $20 \pm 2$  °C) with Bruker Avance 400 MHz spectrometer operating at 400 MHz and 100 MHz, respectively. Chemical shifts are given as  $\delta$  value (ppm) with reference to tetramethylsilane (TMS) as an internal standard. The peak patterns are indicated as follows: s, singlet; d, doublet; t, triplet; m, multiplet; q, quartet. The coupling constants, J, are reported in Hertz (Hz). High-resolution mass spectra (HRMS) were obtained on Agilent Technologies 6530 Accurate mass Q-TOF LC/MS with ESI as ion source and Agilent Technologies 7250 Accurate-Mass Quadrupole Time-of-Flight (Q-TOF) GC/MS with EI as ion source. GC-MS analysis of the products were performed on Thermo Fisher DSQ II with EI ion source. Injection port was set at 220 °C, ion source temperature and AUX temperature zone were set at 250 °C, and injection volume was 1.0 µL. Fluorescence quenching experiments were performed by Hitachi F7000 fluorescence spectrometer.



Figure S1 The photo reactor (left) and the blue LED light source test report (right).

# 2. General synthetic procedure for thiosulfonates and selenosulfonates

**2.1 Synthesis of thiosulfonates.** Thiosulfonates were synthesized according to literature (*Org. Lett.*, 2020, **22**, 4908) with minor modifications using disulfides, sodium benzenesulfonates and iodine in  $CH_2Cl_2$ . After completion of the reaction, sodium thiosulfate was added to remove the excess iodine. Then, the mixture was washed by brine; the organic phase were combined and dried over anhydrous  $Na_2SO_4$ . After filtration and evaporation of the solvent, the crude product was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 10/1, v/v) to give the corresponding desired products (Scheme S1).



Scheme S1 Structure of the synthesized thiosulfonates 1a-o

**2.2** Synthesis of selenosulfonates. *Se*-phenyl benzenesulfonoselenoate and *Se*-benzyl benzenesulfonoselenoate were synthesized according to literature (*J. Org. Chem.*, 2019, **84**, 8100.) by the use of 1,2-diphenyldiselane or 1,2-dibenzyldiselane and sodium benzenesulfonate with [bis(trifluoroacetoxy)iodo]benzene (PIFA) in dichloromethane at 0 °C to room temperature for 3 h. The desired product was purified by silica gel column chromatography (petroleum ether : ethyl acetate = 10 : 1, v/v).

$$R^{-Se}Se^{-R} + O^{Na} \xrightarrow{CH_2Cl_2, PIFA} O^{S-Se}Se^{-R}$$

$$R = Phenyl, Benzyl$$

Scheme S2 Synthesis of selenosulfonates

### 3. General synthetic procedure for products 3

Thiosulfonates (1, 0.4 mmol), TBHP (0.4 mmol, 70% aqueous solution), Na<sub>2</sub>-eosin Y (2.5 mol%) and K<sub>2</sub>CO<sub>3</sub> (1.0 equiv.) were dissolved in corresponding ethers (2, 2.0 mL) in a 25 mL reaction tube, and then the mixture was stirred with the irradiation of 5 W blue LED light under N<sub>2</sub> at room temperature for 12 h. After reaction, the mixture was diluted with brine and extracted with CH<sub>2</sub>Cl<sub>2</sub> (15 mL × 3). The organic layers were combined and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. The residue was

purified by silica gel column chromatography (petroleum ether/ethyl acetate =  $10\sim 20/1$ , v/v) to afford the desired products **3**.



Scheme S3 Synthesis of products 3

### 4. Investigations of photocatalyst, oxidant and base on the model reaction

Table S1 Optimization of reaction conditions.<sup>a</sup>

	$ \begin{array}{c} & \bigcirc \\ & & \bigcirc \\ & & \\ & $				
	1a		2a	3a	
Entry	1	Ox.	PC.	Base	Yield $(\%)^b$
1	1a	TBHP	RhB	K <sub>2</sub> CO <sub>3</sub>	63
2	1a	TBHP	EB	$K_2CO_3$	77
3	1a	TBHP	MB	$K_2CO_3$	73
4	1a	TBHP	RB	$K_2CO_3$	62
5	1a	TBHP	FL	$K_2CO_3$	61
6	1a	TBHP	$EYH_2$	$K_2CO_3$	67
7	1a	TBHP	Na <sub>2</sub> -EY	$K_2CO_3$	83
8	1a	TBHP	Ru(bpy) <sub>3</sub> Cl <sub>2</sub>	$K_2CO_3$	43
9	1a	DTBP	Na <sub>2</sub> -EY	$K_2CO_3$	25
10	1a	$K_2S_2O_8$	Na <sub>2</sub> -EY	$K_2CO_3$	17
11	1a	TBPB	Na <sub>2</sub> -EY	$K_2CO_3$	trace
12	1a	BPO	Na <sub>2</sub> -EY	K <sub>2</sub> CO <sub>3</sub>	trace
13	1a	$H_2O_2$	Na <sub>2</sub> -EY	$K_2CO_3$	33
14	1a	<i>m</i> -CPBA	Na <sub>2</sub> -EY	$K_2CO_3$	trace
15	1a	TBHP	Na <sub>2</sub> -EY	KOH	44
16	1a	TBHP	Na <sub>2</sub> -EY	NEt <sub>3</sub>	38
17	1a	TBHP	Na <sub>2</sub> -EY	NaHCO <sub>3</sub>	21
18	1a	-	Na <sub>2</sub> -EY	$K_2CO_3$	n.r.
19	1a	TBHP	Na <sub>2</sub> -EY-	-	n.r.

<sup>*a*</sup> Reaction conditions: **1a** (0.4 mmol), oxidant (1.0 equiv.), photocatalyst (2.5 mol%) and base (1.0 equiv.) were mixed in THF (**2a**, 2.0 mL) with the irradiation of 5 W blue LED light under N<sub>2</sub> atmosphere at room temperature for 12 h. <sup>*b*</sup> Isolated yield based on **1** were provided. RhB = rhodamine B, EB = eosin B, MB = methylene blue, RB = rose bengal, FL = fluorescein, EYH<sub>2</sub> = eosin Y, Na<sub>2</sub>-EY = Na<sub>2</sub>-eosin Y, TBHP = *tert*-butyl hydroperoxide (70% aqueous solution), DTBP = di-*tert*-butyl peroxide, TBPB = *tert*-butyl peroxybenzoate, BPO = benzoyl peroxide, H<sub>2</sub>O<sub>2</sub> was 30% aqueous solution, *m*-CPBA = *m*-chloroperoxybenzoic acid. n.r. = no reaction.

# 5. Mechanism Study

### 5.1 Experiment interfered with radical scavenger

In a 25 mL reaction tube, *S*-phenyl benzenethiosulfonate (**1a**, 0.4 mmol), TBHP (0.4 mmol, 70% aqueous solution), Na<sub>2</sub>-eosin Y (2.5 mol%), K<sub>2</sub>CO<sub>3</sub> (1.0 equiv.) and 2.0 equiv. of (2,2,6,6-tetramethylpiperidin-1-yl)oxidanyl (TEMPO) or 1,1-diphenylethylene (ASYM) were dissolved in THF (2.0 mL), respectively. The mixtures were stirred under standard reaction conditions for 12 h and then detected by HRMS.



Figure S2 HRMS spectrum of the benzenesulfenyl radical/TEMPO adduct 5



Figure S3 HRMS spectrum of the benzenesulfonyl radical/TEMPO adduct 6



Figure S4 HRMS spectrum of the  $\alpha$ -alkoxyalkyl radical /TEMPO adduct 7



Figure S5 HRMS Spectrum of the benzenesulfenyl radical /ASYM adduct 8



Figure S6 HRMS spectrum of the benzenesulfonyl radical / ASYM adduct 9



Figure S7 HRMS spectrum of the  $\alpha$ -alkoxyalkyl radical / ASYM adduct 10

### 5.2 Fluorescence quenching experiments

A stock solution of Na<sub>2</sub>-eosin Y (5 mM in THF) was prepared for the quenching experiment. 200  $\mu$ L Na<sub>2</sub>-eosin Y stock solution was diluted to 2.0 mL with THF in a quartz cuvette (1 cm × 1 cm). The fluorescence excitation and emission spectra were firstly recorded as shown below. The maximum excitation/emission wavelength were detected as 461/559 nm. Then, quenching experiments were performed with addition of TBHP (70% aqueous solution) or **1a**, respectively.



**Figure S8** Fluorescence excitation (left) and emission (right) spectra of Na<sub>2</sub>-eosin Y ( $5 \times 10^{-4}$  M) in



Figure S9 Fluorescence emission spectra of Na<sub>2</sub>-eosin Y ( $5 \times 10^{-4}$  M) in THF with TBHP (1.0 - 5.0 mM)



Figure S10 The linear relationship between  $I_0/I$  ( $I_0$  and I are the fluorescence intensities before and after the addition of TBHP) and the concentration of TBHP



Figure S11 Fluorescence emission spectra of Na<sub>2</sub>-eosin Y ( $5 \times 10^{-4}$  M) in THF with 1a (1.0 - 5.0 mM)



Figure S12 The linear relationship between  $I_0/I$  ( $I_0$  and I are the fluorescence intensities before and after the addition of 1a) and the concentration of 1a

### 6. Characterization data of compounds 3a-aa and 4a-c

### 2-(phenylthio)tetrahydrofuran (3a)



Pale oil, yield: 83%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.51 (m, 2H), 7.29 (m, 2H), 7.22 (m, 1H), 5.65 (m, 1H), 4.06-3.94 (m, 2H), 2.39-2.34 (m, 1H), 2.04-1.90 (m, 2H), 1.90-1.87 (m, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 135.7, 131.1, 128.8, 126.8, 87.2, 67.3, 32.7, 24.8. GC-HRMS (EI) for C<sub>10</sub>H<sub>12</sub>OS (m/z): calcd. 180.0609, found 180.0583.

#### 2-(p-tolylthio)tetrahydrofuran (3b)



Pale oil, yield: 69%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.40 (m, 2H), 7.11 (m, 2H), 5.57 (m, 1H), 4.03-3.93 (m, 2H), 2.36-2.30 (m, 4H), 2.03-1.84 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 137.0, 131.9, 131.7, 129.6, 87.6, 67.2, 32.6, 24.8, 21.1. GC-MS (EI) for C<sub>11</sub>H<sub>14</sub>OS (m/z): calcd. 194.08, found 194.10.

#### 2-((4-methoxyphenyl)thio)tetrahydrofuran (3c)



Pale oil, yield: 73%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.46 (m, 2H), 6.85 (m, 2H), 5.47 (m, 1H), 4.02-3.90 (m, 2H), 3.79 (s, 3H), 2.34-2.27 (m, 1H), 2.00-1.83 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 159.4, 134.6, 125.6, 114.4, 88.2, 67.2, 55.3, 32.5, 24.8. GC-MS (EI) for C<sub>11</sub>H<sub>14</sub>O<sub>2</sub>S (m/z): calcd. 210.07, found 210.08.

### 2-((2-fluorophenyl)thio)tetrahydrofuran (3d)



Pale oil, yield: 81%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.60 (m, 1H), 7.25 (m, 1H), 7.12-7.04 (m, 2H), 5.70 (m, 1H), 4.07-3.93 (m, 2H), 2.40-2.33 (m, 1H), 2.08-2.00 (m, 2H), 1.92-1.87 (m, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 162.7 and 160.3, 133.9, 129.0 and 128.9, 124.52 and 124.48, 122.4 and 122.2, 115.7 and 115.4, 86.44 and 86.43, 67.4, 32.7, 24.6. GC-MS (EI) for C<sub>10</sub>H<sub>11</sub>FOS (m/z): calcd. 198.05, found 198.04.

#### 2-((4-chlorophenyl)thio)tetrahydrofuran (3e)



Pale oil, yield: 85%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.43 (m, 2H), 7.26 (m, 2H), 5.60 (m, 1H), 4.04-3.93 (m, 2H), 2.39-2.34 (m, 1H), 2.04-1.86 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 134.2, 133.0, 132.4, 128.9, 87.3, 67.3, 32.6, 24.8. GC-MS (EI) for C<sub>10</sub>H<sub>11</sub>ClOS (m/z): calcd. 214.02, found 214.03.

### 2-((tetrahydrofuran-2-yl)thio)phenyl acetate (3f)

Pale oil, yield: 77%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.52 (m, 2H), 7.02 (m, 2H), 5.60 (m, 1H), 4.03-3.93 (m, 2H), 2.38-2.33 (m, 1H), 2.28 (s, 3H), 2.03-1.87 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 169.3, 149.8, 132.5, 122.0, 87.5, 67.2, 32.6, 24.8, 21.1. GC-MS (EI) for C<sub>12</sub>H<sub>14</sub>O<sub>3</sub>S (m/z): calcd. 238.07, found 238.08.

# 2-(thiophen-2-ylthio)tetrahydrofuran (3g)



Pale oil, yield: 75%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.36 (m, 1H), 7.17 (m, 1H), 7.00 (m, 1H), 5.43 (m, 1H), 4.06-4.00 (m, 1H), 3.98-3.93 (m, 1H), 2.32-2.26 (m, 1H), 2.01-1.97 (m, 2H), 1.95-1.84 (m, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 134.3, 132.4, 129.9, 127.5, 89.5, 67.5, 32.1, 24.6. GC-MS (EI) for C<sub>8</sub>H<sub>10</sub>OS<sub>2</sub> (m/z): calcd. 186.02, found 186.10.

# 2-methyl-3-((tetrahydrofuran-2-yl)thio)furan (3h)



Pale oil, yield: 69%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.27 (m, 1H), 6.41 (m, 1H), 5.32 (m, 1H), 4.02-3.93 (m, 1H), 3.92-3.88 (m, 1H), 2.35 (s, 3H), 2.28-2.22 (m, 1H), 2.01-1.84 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 155.4, 140.4, 115.5, 109.3, 87.7, 67.2, 32.3, 24.8, 11.9. GC-MS (EI) for C<sub>9</sub>H<sub>12</sub>O<sub>2</sub>S (m/z): calcd. 184.06, found 184.05.

# 2-(benzylthio)tetrahydrofuran (3i)



Pale oil, yield: 78%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.36-7.29 (m, 4H), 7.23 (m, 1H), 5.23 (m, 1H), 3.98-3.88 (m, 2H), 3.77-3.73 (m, 2H), 2.22-2.18 (m, 1H), 2.02-1.97 (m, 1H), 1.97-1.77 (m, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 138.6, 129.0, 128.4, 126.8, 83.0, 66.8, 32.0, 32.1, 24.8. GC-MS (EI) for C<sub>11</sub>H<sub>14</sub>OS (m/z): calcd. 194.08, found 194.10.

### 2-(phenylthio)-1,4-dioxane (3l)



Pale oil, yield: 80%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.50 (m, 2H), 7.32-7.26 (m, 3H), 5.12 (m, 1H), 4.25-4.20 (m, 1H), 4.00-3.96 (m, 1H), 3.74-3.65 (m, 4H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 134.0, 131.6, 129.0, 127.4, 83.3, 70.0, 66.5, 63.8. GC-MS (EI) for C<sub>10</sub>H<sub>12</sub>O<sub>2</sub>S (m/z): calcd. 196.06, found 196.07.

### 2-(p-tolylthio)-1,4-dioxane (3m)



Pale oil, yield: 76%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.41 (m, 2H), 7.11 (m, 2H), 5.02 (m, 1H), 4.22-4.18 (m, 1H), 3.98-3.94 (m, 1H), 3.72-3.65 (m, 4H), 2.33 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 137.7, 136.5, 132.4, 129.7, 83.5, 69.9, 66.4, 64.0, 21.1. GC-MS (EI) for C<sub>11</sub>H<sub>14</sub>O<sub>2</sub>S (m/z): calcd. 210.07, found 210.07.

#### 2-((4-chlorophenyl)thio)-1,4-dioxane (3n)



Pale oil, yield: 83%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.43 (m, 2H), 7.27 (m, 2H), 5.08 (m, 1H), 4.26-4.21 (m, 1H), 3.99-3.95 (m, 1H), 3.74-3.64 (m, 4H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 133.6, 132.9, 132.5, 129.1, 83.3, 69.8, 66.5, 63.6. GC-MS (EI) for C<sub>10</sub>H<sub>11</sub>ClO<sub>2</sub>S (m/z): calcd. 230.02, found 230.02.

### 2-(thiophen-2-ylthio)-1,4-dioxane (30)



Pale oil, yield: 67%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.39 (m, 1H), 7.19 (m, 1H), 7.00 (m, 1H), 4.91 (m, 1H), 4.28-4.23 (m, 1H), 3.96-3.93 (m, 1H), 3.72-3.66 (m, 4H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 135.0, 130.4, 130.3, 127.5, 84.5, 69.4, 66.5, 63.7. GC-MS (EI) for C<sub>8</sub>H<sub>10</sub>O<sub>2</sub>S<sub>2</sub> (m/z): calcd. 202.01, found 202.01.

### 2-(phenylthio)tetrahydro-2H-pyran (3p)



Pale oil, yield: 71%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ: 7.48 (m, 2H), 7.29 (m, 2H), 7.26-7.19 (m, 1H), 5.21 (m, 1H), 4.20-4.15 (m, 1H), 3.61-3.56 (m, 1H), 2.04-1.99 (m, 1H), 1.86-1.81 (m, 2H), 1.67-1.61 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ: 135.4, 130.9, 128.8, 126.7, 85.3, 64.5, 31.6, 25.5,

### 21.7. GC-MS (EI) for C<sub>11</sub>H<sub>14</sub>OS (m/z): calcd. 194.08, found 194.08.

### 2-(p-tolylthio)tetrahydro-2H-pyran (3q)



Pale oil, yield: 67%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ: 7.38 (m, 2H), 7.10 (m, 2H), 5.12 (m, 1H), 4.20-4.14 (m, 1H), 3.59-3.53 (m, 1H), 2.32 (s, 3H), 2.04-1.98 (m, 1H), 187-1.78 (m, 2H), 1.78-1.60 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ: 136.9, 131.7, 131.4, 129.6, 85.7, 64.6, 31.6, 25.5, 21.7, 21.1.

#### 2-((4-chlorophenyl)thio)tetrahydro-2H-pyran (3r)



Pale oil, yield: 75%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.40 (m, 2H), 7.25 (m, 2H), 5.17 (m, 1H), 4.18-4.12 (m, 1H), 3.61-3.55 (m, 1H), 2.03-2.00 (m, 1H), 1.85-1.81 (m, 2H), 1.66-1.61 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 133.9, 132.8, 132.2, 128.9, 85.4, 64.4, 31.5, 25.5, 21.5. GC-MS (EI) for C<sub>11</sub>H<sub>13</sub>ClOS (m/z): calcd. 228.04, found 228.04.

#### 2-(thiophen-2-ylthio)tetrahydro-2H-pyran (3s)



Pale oil, yield: 63%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ: 7.36 (m, 1H), 7.15 (m, 1H), 6.99 (m, 1H), 5.00 (m, 1H), 4.22-4.16 (m, 1H), 3.61-3.56 (m, 1H), 2.02-1.95 (m, 1H), 1.86-1.77 (m, 2H), 1.64-1.60 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ: 134.1, 132.0, 129.6, 127.4, 87.3, 64.4, 30.9, 25.5, 21.3. GC-MS (EI) for C<sub>9</sub>H<sub>12</sub>OS<sub>2</sub> (m/z): calcd. 200.03, found 200.04.

#### (tert-butoxymethyl)(phenyl)sulfane (3t)



Pale oil, yield: 57%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ: 7.49 (m, 2H), 7.29-7.26 (m, 2H), 7.20 (m, 1H), 4.90 (s, 2H), 1.26 (s, 9H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ: 136.6, 130.0, 128.8, 126.4, 75.3, 68.2, 27.8. GC-MS (EI) for C<sub>11</sub>H<sub>16</sub>OS (m/z): calcd. 196.09, found 196.09.

### (tert-butoxymethyl)(4-chlorophenyl)sulfane (3u)

n C

Pale oil, yield: 62%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ: 7.42 (m, 2H), 7.25 (m, 2H), 4.87 (s, 2H), 1.55 (s, 9H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ: 135.1, 132.6, 131.4, 128.9, 75.5, 68.3, 27.8. GC-MS (EI)

for  $C_{10}H_{13}ClO_2S$  (m/z): calcd. 230.05, found 230.05.

(4-chlorophenyl)(1-methoxycyclopentyl)sulfane (3x-i) and (4-chlorophenyl)((cyclopentyloxy)methyl)sulfane (3x-ii) (3x)



Pale oil, yield: 75% (**3x-i** : **3x-ii** = 2.5 : 1.0). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.40 (m, 3H), 7.25 (m, 3H), 4.93-4.33 (m, 1H), 3.46 (s, 3H), 1.87-1.68 (m, 11H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 134.8 and 134.7, 133.6 and 132.9, 132.6 and 131.4, 129.0 and 128.7, 100.4, 79.1 and 73.9, 51.4, 38.0, 32.0, 23.5 and 23.2. GC-MS (EI) for C<sub>12</sub>H<sub>15</sub>ClOS (m/z): calcd. 242.05, found 242.04.

2-((4-chlorophenyl)thio)-2-methyltetrahydrofuran (3y-i) and 2-((4-chlorophenyl)thio)-5methyltetrahydrofuran (3y-ii) (3y)



Pale oil, yield: 81% (**3y-i** : **3y-ii** = 3.0 : 1.0). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 8.36 (m, 2H), 7.86 (m, 2H), 1.58 (s, 9H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 152.9, 149.8, 126.3, 124.2, 84.3, 29.9. GC-MS (EI) for C<sub>11</sub>H<sub>13</sub>ClOS (m/z): calcd. 228.04, found 228.02.

### (4-chlorophenyl)(1,2-dimethoxyethyl)sulfane (3z-i)



Pale oil, yield: 46%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.42 (m, 2H), 7.27 (m, 2H), 4.72 (m, 1H), 3.56-3.49 (m, 5H), 3.36 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 135.2, 134.2, 130.6, 129.0, 88.8, 74.5, 59.1, 56.3. GC-MS (EI) for C<sub>10</sub>H<sub>13</sub>ClO<sub>2</sub>S (m/z): calcd. 232.03, found 232.06.

#### (4-chlorophenyl)((2-methoxyethoxy)methyl)sulfane (3z-ii)



Pale oil, yield: 31%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.41 (m, 2H), 7.25 (m, 2H), 5.03 (s, 2H), 3.79-3.76 (m, 2H), 3.59-3.55 (m, 2H), 3.38 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 134.4, 132.8, 131.5, 129.0, 76.5, 71.5, 67.5, 59.0. GC-MS (EI) for C<sub>10</sub>H<sub>13</sub>ClO<sub>2</sub>S (m/z): calcd. 232.03, found 232.06.

2-((4-chlorophenyl)thio)-1,3-dioxolane (3aa-i) and 4-((4-chlorophenyl)thio)-1,3-dioxolane (3aa-ii) (3aa)



Pale oil, yield: 68% (**3aa-i** : **3aa-ii** = 2.7 : 1.0). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$ : 7.69-7.67 (m, 2H), 7.52-7.50 (m, 3H), 1.56 (s, 9H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 146.6, 131.5, 128.9, 124.9, 82.8, 29.9. GC-MS (EI) for C<sub>9</sub>H<sub>9</sub>ClO<sub>2</sub>S (m/z): calcd. 216.00, found 216.00.

### tert-butyl 4-nitrobenzenesulfinate (4a)

O<sub>2</sub>N

Yellow solid, yield: 75%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ: 8.36 (m, 2H), 7.86 (m, 2H), 1.58 (s, 9H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ: 152.9, 149.8, 126.3, 124.2, 84.3, 29.9.

# tert-butyl benzenesulfinate (4b)

White oil, yield: 78%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ: 7.69-7.67 (m, 2H), 7.52-7.50 (m, 3H), 1.56 (s, 9H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ: 146.6, 131.5, 128.9, 124.9, 82.8, 29.9.

# *tert*-butyl 4-chlorobenzenesulfinate (4c)

CI

White oil, yield: 81%. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ: 7.62-7.60 (m, 2H), 7.50-7.48 (m, 2H), 1.55 (s, 9H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ: 145.2, 137.9, 129.2, 126.4, 83.3, 29.9.

# 7. <sup>1</sup>H NMR, <sup>13</sup>C NMR and HRMS/MS copies of compounds 3a-aa and 4a-c



<sup>13</sup>C NMR spectrum of compound **3a** 



HRMS spectrum of compound 3a



<sup>1</sup>H NMR spectrum of compound **3b** 



 $^{13}\mathrm{C}$  NMR spectrum of compound  $\mathbf{3b}$ 



MS spectrum of compound 3b



<sup>13</sup>C NMR spectrum of compound **3**c







<sup>13</sup>C NMR spectrum of compound **3d** 



MS spectrum of compound 3d





H NMR spectrum of compound 3e



<sup>13</sup>C NMR spectrum of compound **3e** 







 $^{13}\mathrm{C}$  NMR spectrum of compound 3f



MS spectrum of compound  $\mathbf{3f}$ 



<sup>13</sup>C NMR spectrum of compound **3g** 



<sup>1</sup>H NMR spectrum of compound **3h** 



<sup>13</sup>C NMR spectrum of compound **3h** 



MS spectrum of compound 3h



<sup>13</sup>C NMR spectrum of compound **3i** 







<sup>13</sup>C NMR spectrum of compound **3**l



MS spectrum of compound 31



<sup>13</sup>C NMR spectrum of compound **3m** 



<sup>1</sup>H NMR spectrum of compound **3n** 



<sup>13</sup>C NMR spectrum of compound **3n** 



MS spectrum of compound **3n** 



<sup>13</sup>C NMR spectrum of compound **30** 







<sup>13</sup>C NMR spectrum of compound **3p** 



MS spectrum of compound **3p** 



<sup>13</sup>C NMR spectrum of compound **3**q



<sup>13</sup>C NMR spectrum of compound **3r** 



<sup>1</sup>H NMR spectrum of compound **3s** 



<sup>13</sup>C NMR spectrum of compound **3s** 



MS spectrum of compound 3s



<sup>13</sup>C NMR spectrum of compound **3**t



<sup>1</sup>H NMR spectrum of compound **3u** 



<sup>13</sup>C NMR spectrum of compound **3u** 



MS spectrum of compound  $\mathbf{3u}$ 



<sup>13</sup>C NMR spectrum of compound 3x



MS spectrum of compound 3x



<sup>1</sup>H NMR spectrum of compound 3y



 $^{13}\text{C}$  NMR spectrum of compound 3y







<sup>13</sup>C NMR spectrum of compound **3z-i** 



<sup>1</sup>H NMR spectrum of compound **3z-ii** 



<sup>13</sup>C NMR spectrum of compound **3z-ii** 



MS spectrum of compound 3z-ii



<sup>13</sup>C NMR spectrum of compound **3aa** 



<sup>1</sup>H NMR spectrum of compound 4a



<sup>13</sup>C NMR spectrum of compound **4a** 



<sup>1</sup>H NMR spectrum of compound **4b** 



<sup>13</sup>C NMR spectrum of compound **4b** 



<sup>1</sup>H NMR spectrum of compound 4c



<sup>13</sup>C NMR spectrum of compound **4c**