

Supporting Information

DNAzyme-powered AuNRs-DNA walker for Visual Detection of *Escherichia coli*

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Table of content:

1. **Table S1.** The sequences of oligonucleotides used in this work.
2. **Table S2.** Comparison of different methods for *E. coli* determination.
3. **Figure S1.** Feasibility of polyacrylamide gel electrophoresis analysis.

Table S1 The sequences of oligonucleotides used in this work.

| Oligonucleotides name | Sequence (5' to 3') description |
|--------------------------|--|
| Apt | CCGGACGCTTATGCCTGCCATCTACAGAGCAGGTGTGACGG |
| T | TCCGTCACACCTTTAAGCGTCCGG |
| HAP | SH-polyT(10)-TGGGTAGGGCGGGTTGGGACTAT/rA/GGAAGAGA AAAGCCCTAC |
| I | AAGAGACCGGACGCTTAAAAAGGTGTGACGGA |
| Dz | SH-polyT(55)-ACCTTTTAAGCGTCCGGTCTCTCTCCGAGCCGG TCGAAATAGT |

Table S2. Comparison of different methods for *E. coli* determination.

| Signal read-out methods | LOD (cfu/mL) | Linear range (cfu/mL) | References |
|--|----------------------|-------------------------------------|------------|
| Surface Enhanced Raman Spectroscopy | 1.5×10^2 | $1.5 \times 10^2 - 3.0 \times 10^5$ | [1] |
| Colorimetric Assay | 4.1×10^1 | $1.2 \times 10^2 - 3.6 \times 10^3$ | [2] |
| Electrochemical | 1.0×10^1 | $1.0 \times 10^1 - 1.0 \times 10^7$ | [3] |
| Electrochemical | 7.0×10^1 | $1.0 \times 10^1 - 1.0 \times 10^5$ | [4] |
| Electrochemiluminescence | 2.5×10^{-4} | $5.0 \times 10^2 - 5.0 \times 10^5$ | [5] |
| Colorimetric Assay | 1.0×10^2 | $0 - 1.0 \times 10^5$ | [6] |
| Colorimetric Assay | 5.8×10^0 | $1.0 \times 10^1 - 1.0 \times 10^5$ | This work |

Figure S1. Feasibility of polyacrylamide gel electrophoresis analysis.

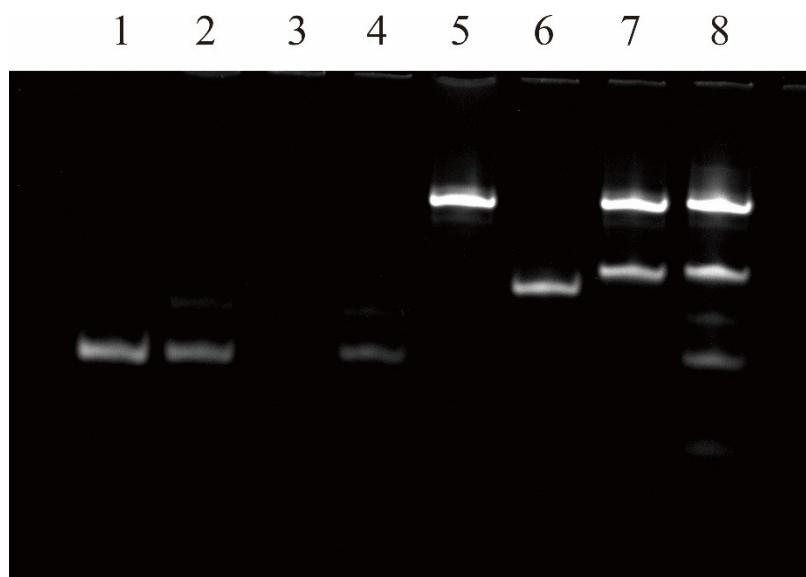


Figure. S1 Feasibility of polyacrylamide gel electrophoresis analysis. Lane 1, Apt; Lane 2, HAP
Lane 3, T; Lane 4, I; Lane 5, Dz; Lane 6, Apt-T; Lane 7, Mix T, I and Dz; Lane 8, probe incubated
with *E. coli*.

References

- [1] S.K. Srivastava, H.B. Hamo, A. Kushmaro, R.S. Marks, C. Gruner, B. Rauschenbach, I. Abdulhalim, Highly sensitive and specific detection of *E. coli* by a SERS nanobiosensor chip utilizing metallic nanosculptured thin films, *Analyst* 140(9) (2015) 3201-9.
- [2] Y. Xie, Y. Huang, J. Li, J. Wu, A trigger-based aggregation of aptamer-functionalized gold nanoparticles for colorimetry: An example on detection of *Escherichia coli* O157:H7, *Sensors Actuators B: Chem.* 339 (2021).
- [3] W. Zhang, C. Luo, L. Zhong, S. Nie, W. Cheng, D. Zhao, S. Ding, Sensitive detection of enteropathogenic *E. coli* using a bfpA gene-based electrochemical sensor, *Microchimica Acta* 180(13-14) (2013) 1233-1240.
- [4] E.B. Setterington, E.C. Alocilja, Rapid electrochemical detection of polyaniline-labeled *Escherichia coli* O157:H7, *Biosens. Bioelectron.* 26(5) (2011) 2208-14.
- [5] H. Yang, Y. Wang, H. Qi, Q. Gao, C. Zhang, Electrogenerated chemiluminescence biosensor incorporating ruthenium complex-labelled Concanavalin A as a probe for the detection of *Escherichia coli*, *Biosens. Bioelectron.* 35(1) (2012) 376-381.
- [6] W. Ren, W. Liu, J. Irudayaraj, A net fishing enrichment strategy for colorimetric detection of *E. coli* O157:H7, *Sensors Actuators B: Chem.* 247 (2017) 923-929.