

## **SUPPORTING INFORMATION**

### **Exigent carbon nanodots for trapping 6-thioguanine to resist fire blight caused by *Erwinia amylovora* in an orchard**

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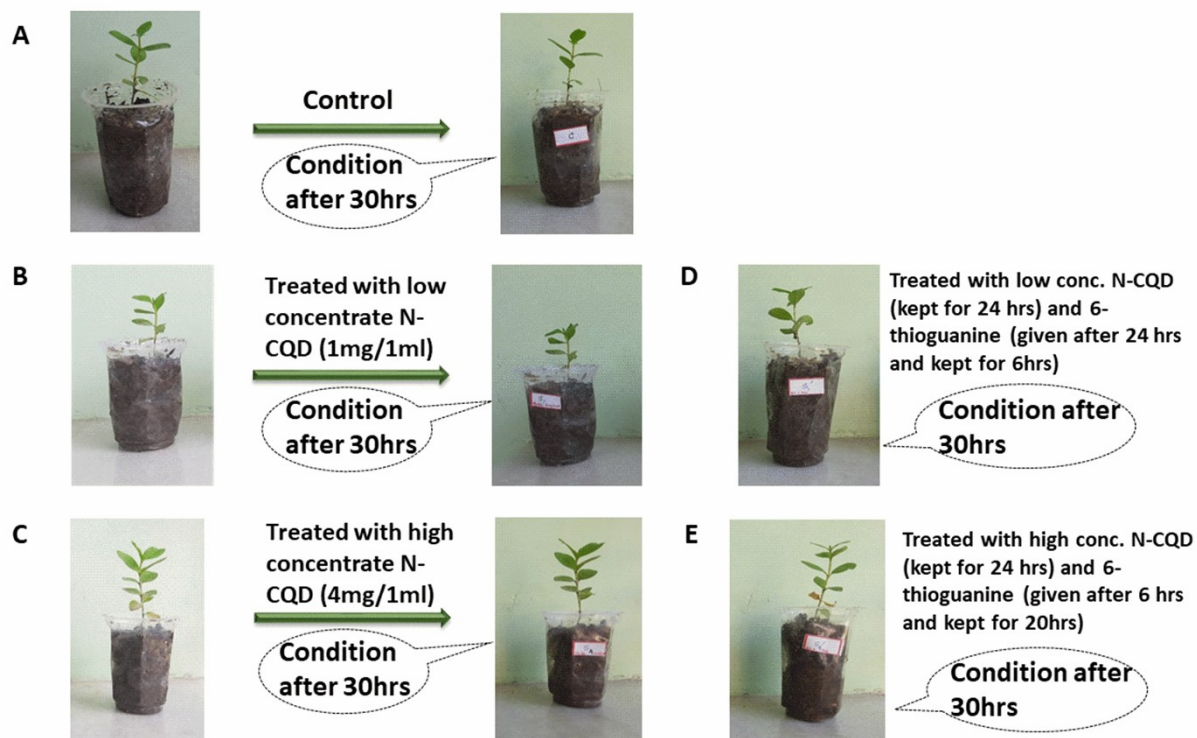
Number of Tables: 3

Number of Figures: 10

**Table S1. Performance comparison of existing methods and present method for detection of 6-Thioguanine using nanosensors**

| Methods  | Methods/Material applied  | Linear Range                              | Limit of Detection      | Application   | References                    |
|--|---|---|-------------------------|---|-------------------------------|
| <b>Droplet-based microfluidic SERS technique (Surface-enhanced Raman scattering)</b> | Au nanoparticles<br>QY= Not available                           | 0.010 $\mu\text{M}$ -<br>10 $\mu\text{M}$ | 0.032 $\mu\text{M}$     | Detection in human serum                                      | Zhang <i>et. al.</i> 2019 [1] |
| <b>Fluorimetry</b>   | CQD–AgNP<br>CQD–AuNP<br>QY= ~ 6%                                | 0.03-<br>1.0 $\mu\text{M}$                | 0.01 $\mu\text{M}$      | Detection in human plasma samples                             | Amjadi <i>et.al.</i> 2017 [2] |
| <b>Free-standing liquid membrane SERS substrate</b>                                  | Ag <sub>core</sub> Au <sub>shell</sub> NPs<br>QY= Not available | 10 ~<br>100 $\mu\text{g kg}^{-1}$         | 5 $\mu\text{g kg}^{-1}$ | Detection in human body fluids                                | Liu <i>et.al.</i> 2022 [3]    |
| <b>Fluorimetry</b>   | SNCQDs<br>QY= 23%   | 0.005-<br>80 $\mu\text{M}$                | 0.0016 $\mu\text{M}$    | Detection in plasma and urine of leukemia patient             | Yu <i>et.al.</i> 2019 [4]     |
| <b>Fluorimetry</b>   | NCQDs<br>QY= 19.45%   | 4.8-<br>55.2 $\mu\text{M}$                | 0.0113 $\mu\text{M}$    | Detection and trapping of 6TG in plants to resist fire blight | <b>This Work</b>              |

## Plant Treatment:



**Fig. S1** (A). Control: No treatment was given to the plant and monitor for 30hrs. (B).  $S_1$ : Treated with low concentrate N-CQD (1mg/1ml) and kept in observation for 30hrs. (C).  $S_4$ : Treated with high concentrate N-CQD (4mg/1ml) and kept in observation for 30hrs. (D)  $S_1'$ : Treated another plant with low concentrate N-CQD (1mg/1ml) as  $S_1$  (monitor for 24hrs) and after 24hrs, again treated with 6-thioguanine (monitor for 6hrs). Kept under surveillance for a total of 30 hours. (E)  $S_4'$ : Treated another plant with high concentrate N-CQD (4mg/1ml) as  $S_4$  (monitor for 24hrs) and after 24hrs, again treated with 6-thioguanine (monitor for 6hrs). Kept under surveillance for a total of 30 hours.

## TEM and HRTEM images of the N-CQDs:

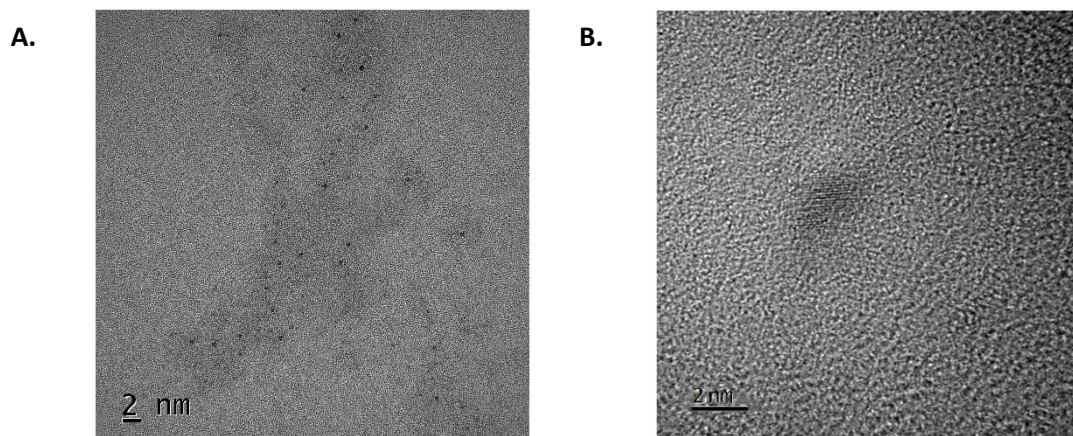


Fig. S2 A) TEM and B) HRTEM images of the N-CQDs.

## EI (electronic image) and EDX images of the N-CQDs:

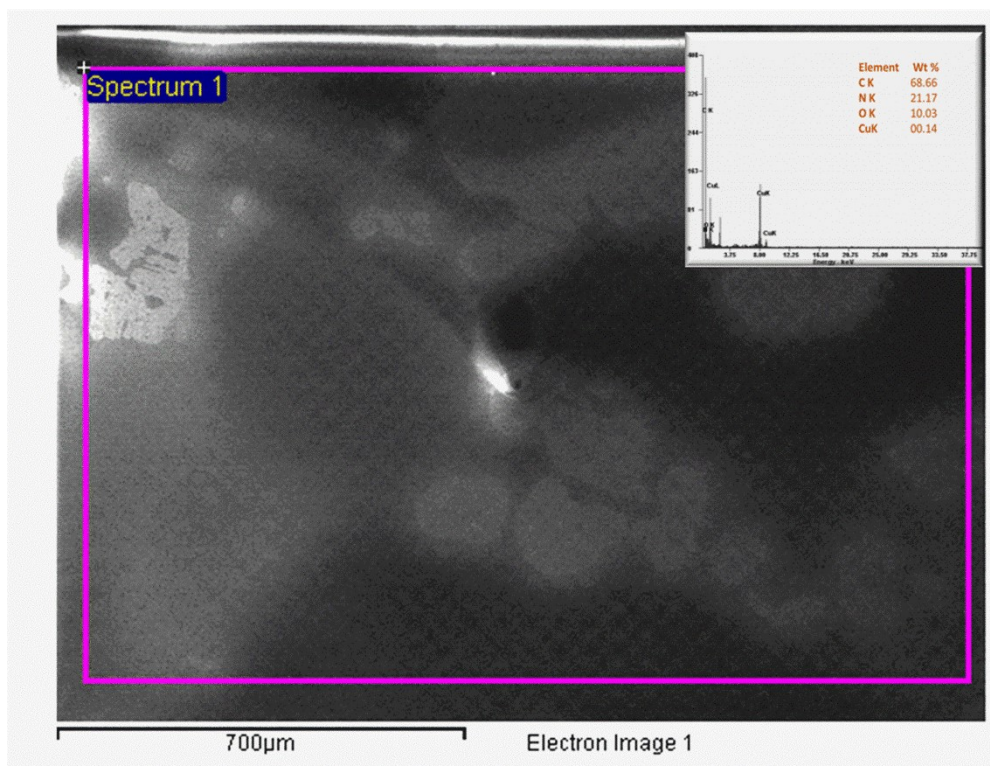


Fig. S3 EDX pattern of the N-CQDs.

## XPS image of the N-CQDs:

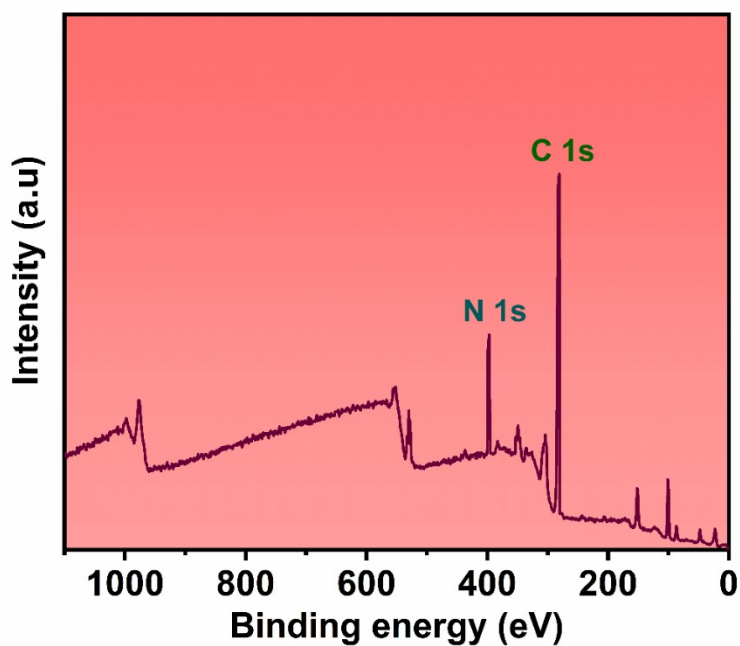


Fig. S4 XPS spectrum of N-CQDs.

## Photostability:

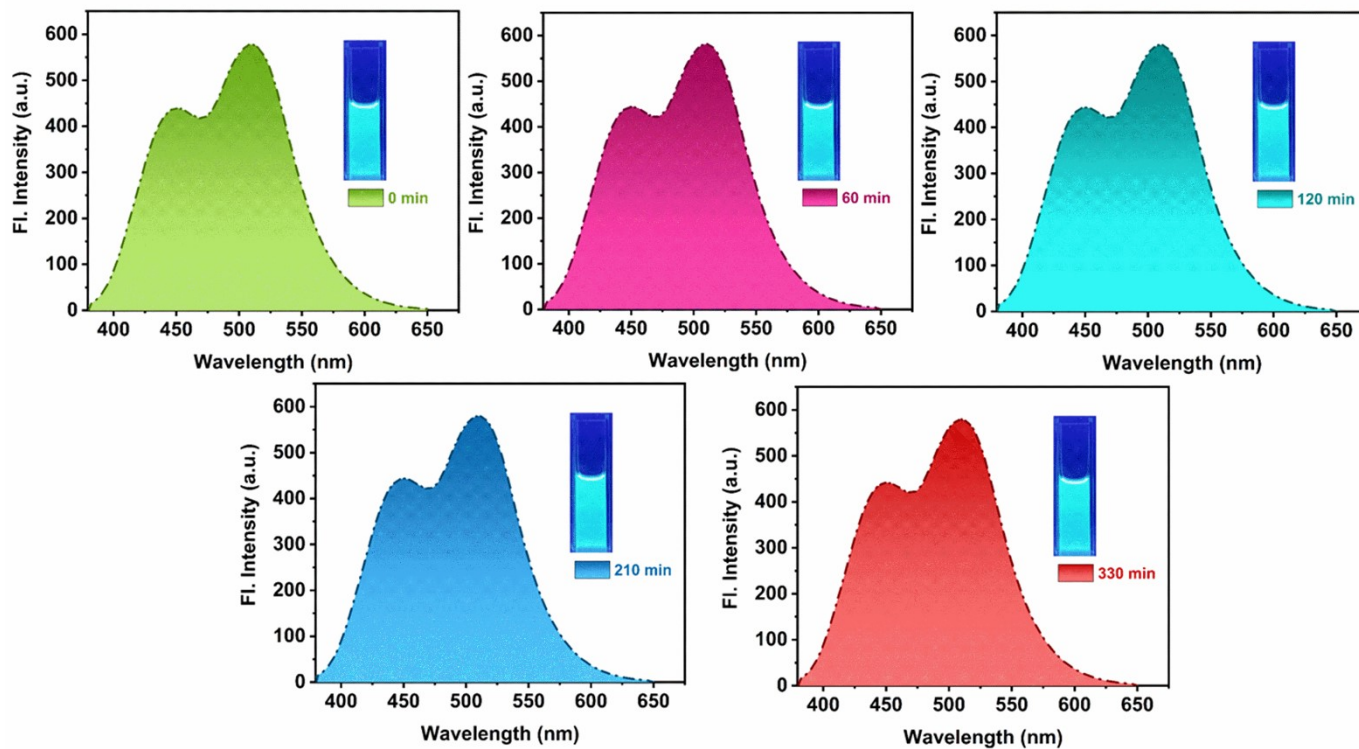
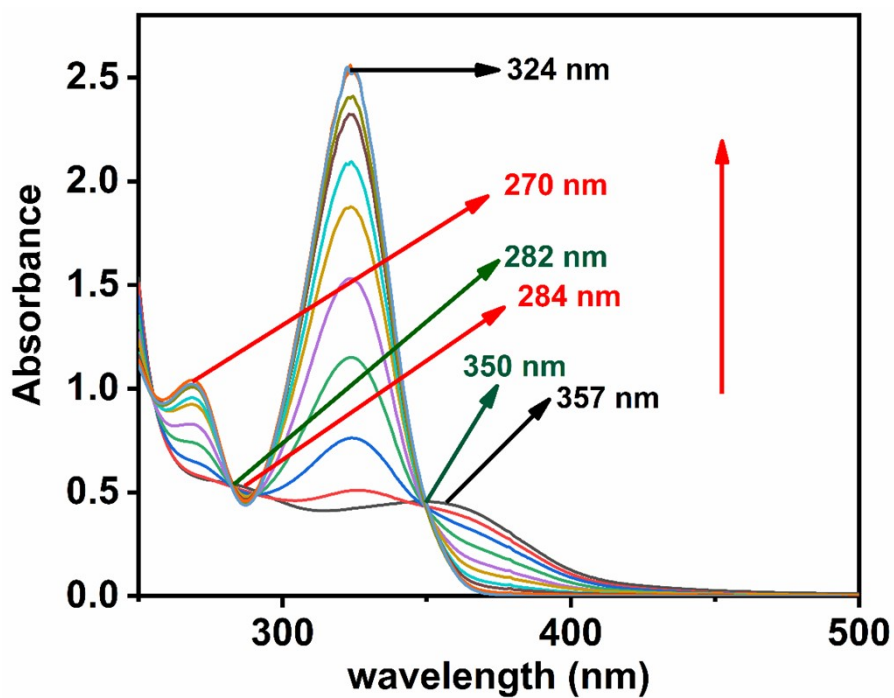


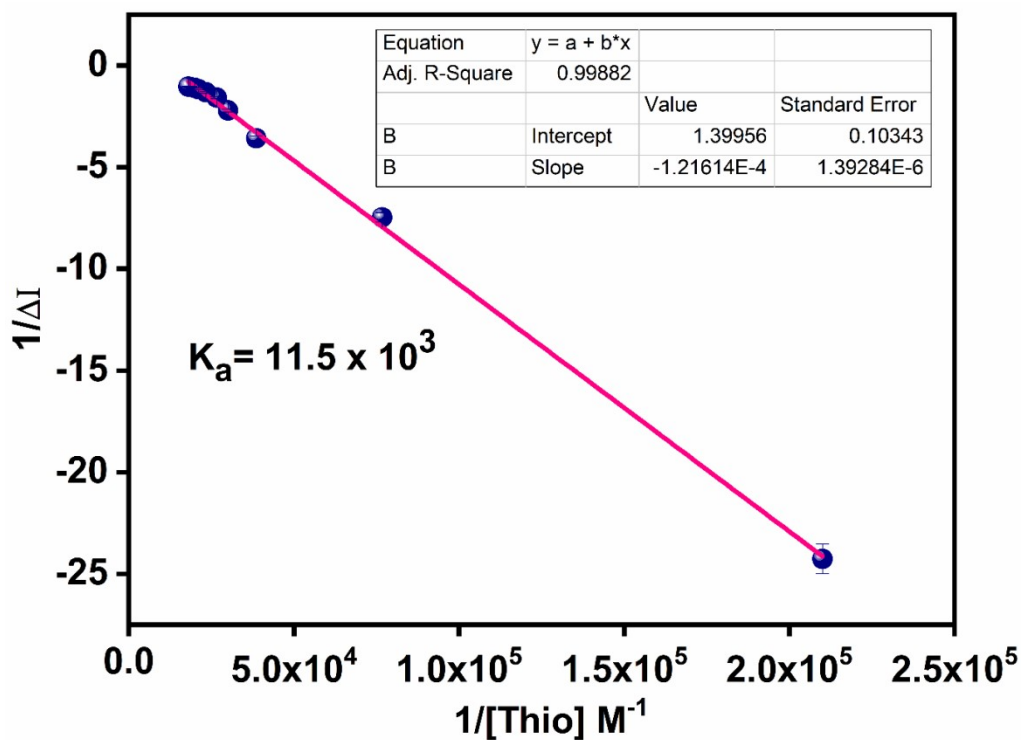
Fig. S5 Photostability test and fluorescence response of N-CQDs under continuous 365 nm UV light irradiation.



**UV-Titration:**

**Fig. S6** UV-vis absorption spectra of N-CQDs upon addition of 6-thioguanine ( $10^{-4}$  M) at  $p^H$  7.4 PBS buffer.

**Binding constant calculation graph (Fluorescence method):**



**Fig. S7** Linear regression analysis for the calculation of association constant value by fluorescence titration method.

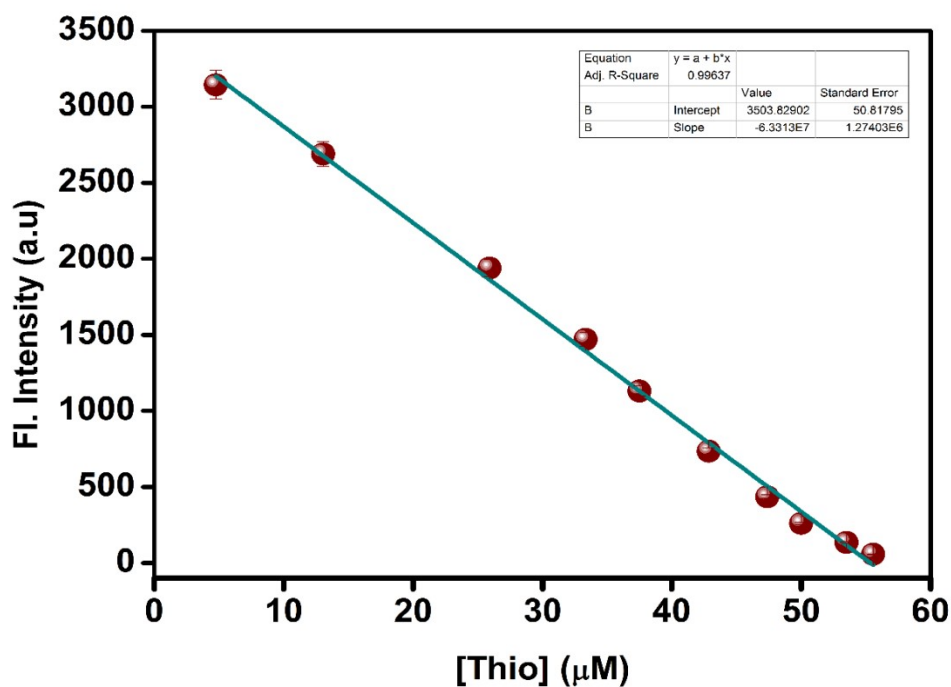
The association const. ( $K_a$ ) of N-CQDs for sensing 6-thioguanine was determined from the equation:  $K_a = \text{intercept/slope}$ . From the linear fit graph, we get intercept= $1.39956$ , slope= $1.21614 \times 10^{-4}$ . Thus, we get  $K_a = (1.39956) / (1.21614 \times 10^{-4}) = 11.5 \times 10^3 \text{ M}^{-1}$

**Table S2. Calculation of Standard Deviation and Limit of Detection (LOD) for 6-thioguanine**

| Blank Reading (N-CQDs) | Fluorescence Intensities at 430 nm (X) | Mean ( $\bar{X}$ ) | Standard Deviation ( $\sigma$ ) = $\sqrt{\frac{\sum  X - \bar{x} ^2}{N}}$ |
|------------------------|--|--------------------|---|
| Reading 1              | 3106.00                                | 3105.95            | 0.2391  |
| Reading 2              | 3105.82                                |                    |   |
| Reading 3              | 3106.14                                |                    |   |
| Reading 4              | 3105.56                                |                    |   |
| Reading 5              | 3106.23                                |                    |   |

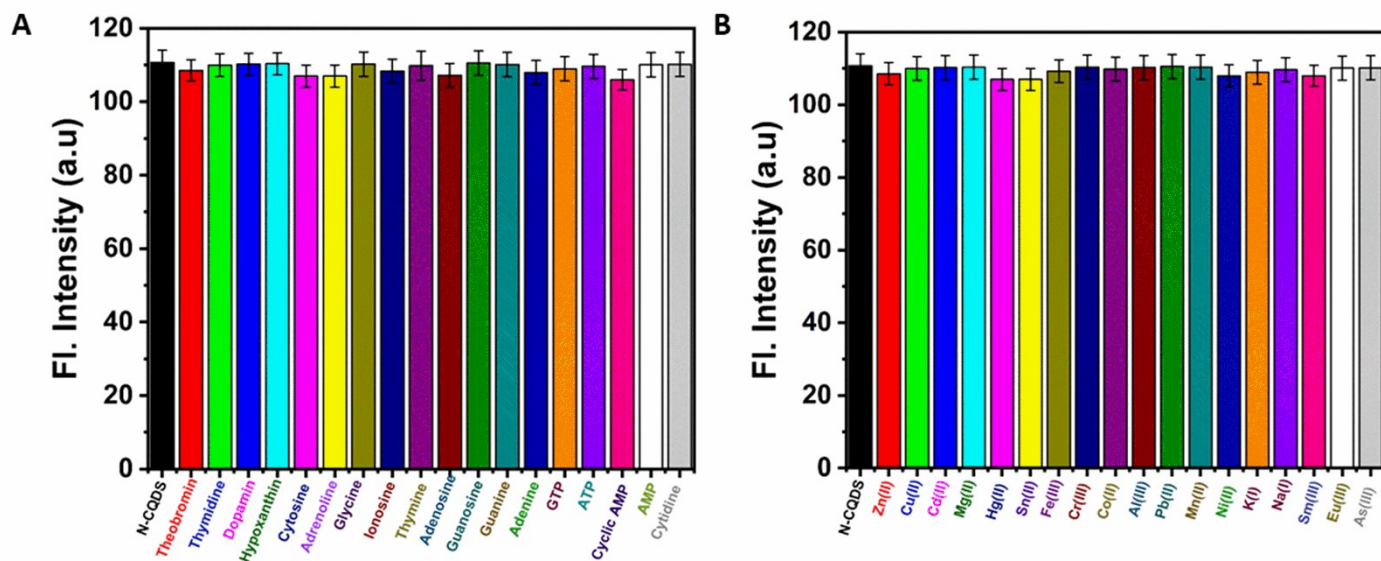
Slope, m for 6-thioguanine =  $6.3313 \times 10^7$

LOD for 6-thioguanine =  $3\sigma/m = (3 \times 0.2391) / (6.3313 \times 10^7) = 0.0113 \times 10^{-6} \text{ M} = 0.0113 \mu\text{M} = 11.29 \text{ nM}$



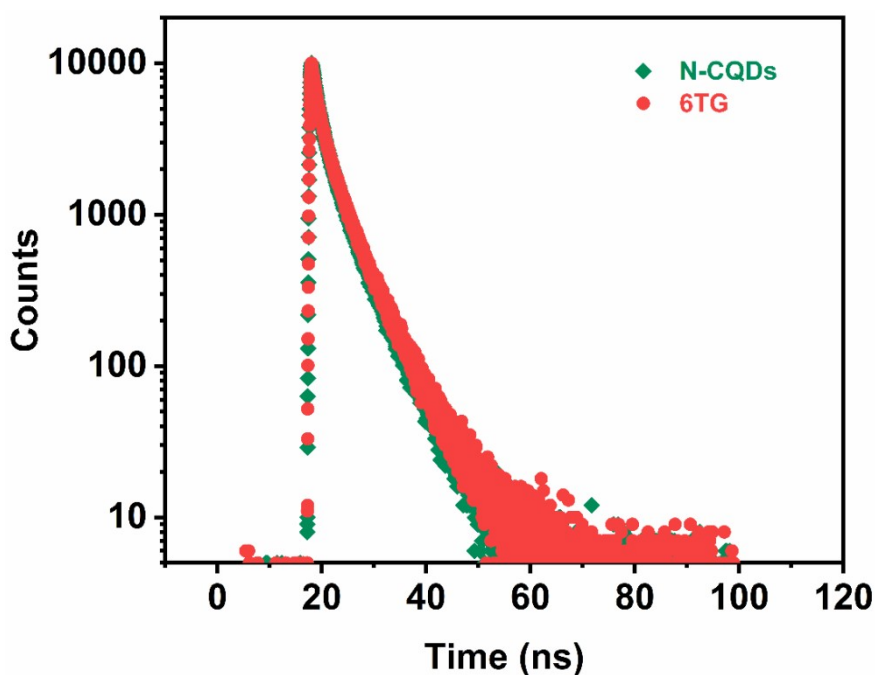
**Fig. S8** Linear fit curve of N-CQDs at 450 nm with respect to 6-thioguanine concentration.

**Selectivity:**



**Fig. S9** (A) Competitive fluorescence spectra of N-CQDs with different biomolecules and (B) metals at 450 nm ( $\lambda_{ex} = 360$  nm).

**Fluorescence lifetime decay:**



**Fig. S10** The fluorescence lifetime of the N-CQDs before (green) and after the addition of 6TG (red).



**Table S3. Decay time components of N-CQDs, N-CQDs + 6TG**

| Syst em       | b <sub>1</sub>   | τ <sub>1</sub>   | b <sub>2</sub>   | τ <sub>2</sub>   | b <sub>3</sub>  | τ <sub>3</sub>   | b <sub>4</sub>   | τ <sub>4</sub>   | $\langle\tau\rangle=b_1\tau_1+b_2\tau_2+b_3\tau_3+b_4\tau_4$ |
|---------------|------------------|------------------|------------------|------------------|-----------------|------------------|------------------|------------------|--|
| N-CQ Ds       | 2.1892<br>36E-02 | 1.2222<br>63E-09 | 1.9427<br>66E-02 | 3.6329<br>71E-09 | 3.5005<br>2E-03 | 8.0617<br>95E-09 | 4.4366<br>11E-02 | 2.6429<br>87E-10 | 0.14ns   |
| N-CQ Ds + 6TG | 2.4452<br>79E-02 | 8.8547<br>46E-10 | 1.9274<br>59E-02 | 3.0727<br>05E-09 | 7.7199<br>4E-03 | 6.7318<br>14E-09 | 9.5881<br>96E-02 | 1.0365<br>29E-10 | 0.14ns   |

**REFERENCES**

1. W. S. Zhang, Y. N. Wang, Y. Wang, Z. R. Xu *Sens. Actuators B Chem.* 2019, **283**, 532–537.
2. M. Amjadi, R. Shokri, T. Hallaj. *Luminescence*, 2017, **32(3)**, 292-297.
3. W. Liu, S. Zhou, J. Liu, X. Zhao, Z. Feng, D. Wang, Z. Gong, M. Fan. *Anal Bioanal Chem.* 2022, **414(4)**, 1663-1670.
4. C. Yu, X. Jiang, D. Qin, G. Mo. *ACS Sustainable Chem. Eng.* 2019, **7 (19)**, 16112–16120.