

Electronical Supporting Information

Chemistry and bioactivity of lindenane sesquiterpenoids and their oligomers

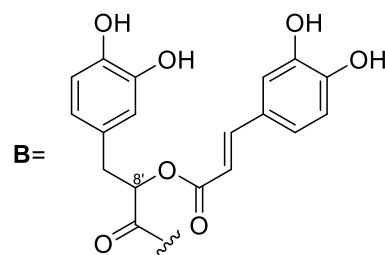
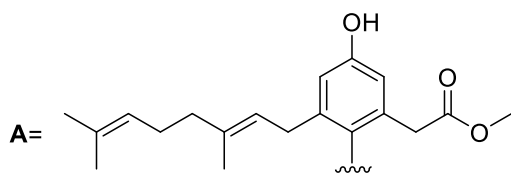
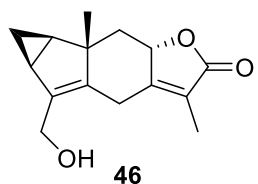
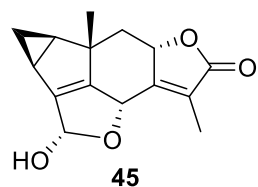
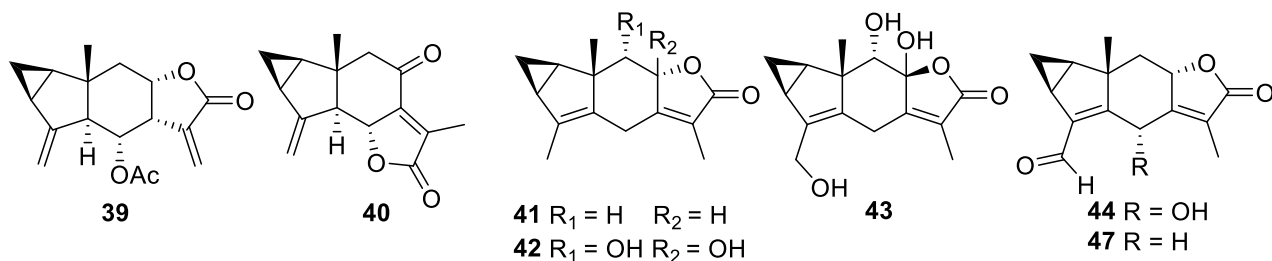
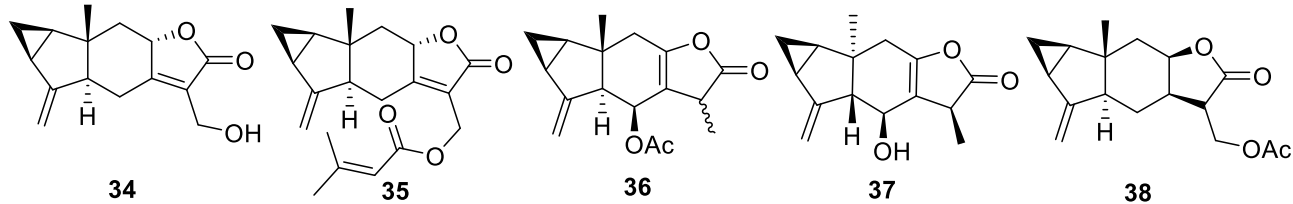
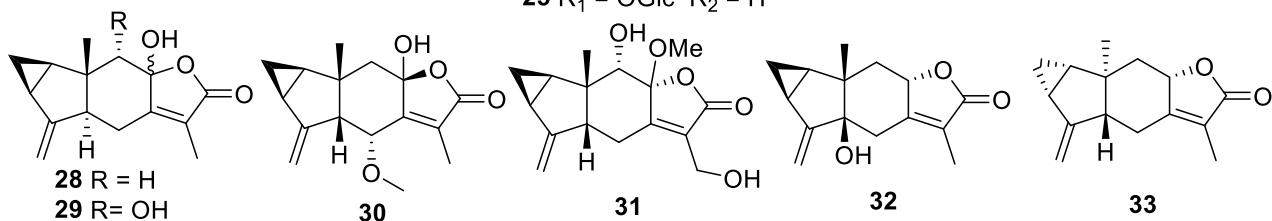
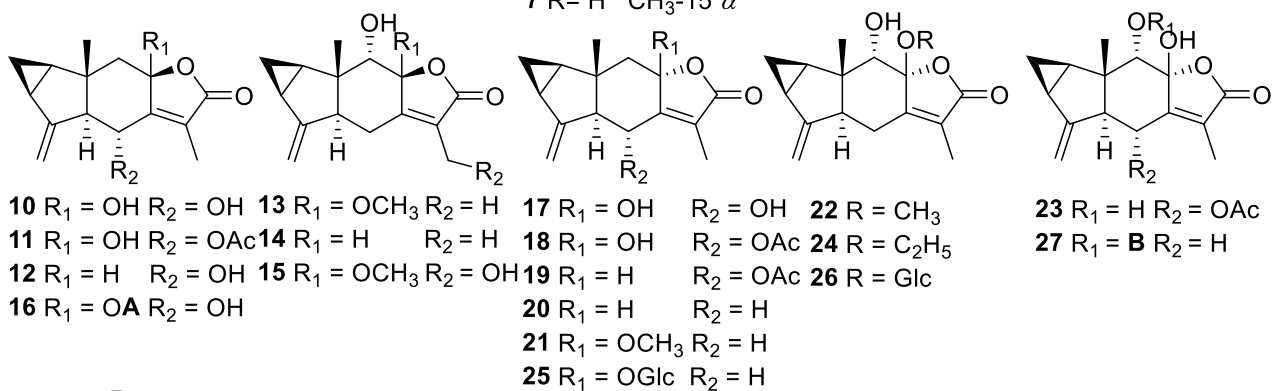
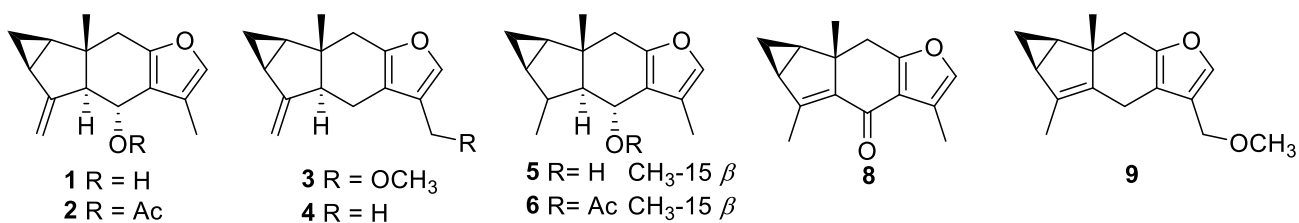
Jun Luo, Danyang Zhang, Pengfei Tang, Nan Wang, Shuai Zhao, Lingyi Kong*

Jiangsu Key Laboratory of Bioactive Natural Product Research and State Key Laboratory of Natural Medicines, School of Traditional Chinese Pharmacy, China Pharmaceutical University, Nanjing 210009, People's Republic of China

*Corresponding author: cpu_lykong@126.com (Lingyi Kong)

Contents

Table S1-1 Structure names, sources and bioactivity of ring- <i>intact</i> type LS monomers (1-27).....	S5
Table S1-2 Structure names, sources and bioactivity of ring- <i>intact</i> type LS monomers (28-55).....	S6
Table S1-3 Structure names, sources and bioactivity of ring- <i>intact</i> type LS monomers (56-81).....	S7
Table S1-4 Structure names, sources and bioactivity of ring- <i>intact</i> type LS monomers (82-95).....	S8
Table S2 Structure names, sources and bioactivity of ring- <i>seco</i> type LS monomers (96-110).....	S9
Table S3-1 Structure names, sources and bioactivity of <i>homo</i> -[4 + 2] cycloaddition type LS dimers (111-136).....	S12
Table S3-2 Structure names, sources and bioactivity of <i>homo</i> -[4 + 2] cycloaddition type LS dimers (137-163).....	S13
Table S3-3 Structure names, sources and bioactivity of <i>homo</i> -[4 + 2] cycloaddition type LS dimers (164-188).....	S15
Table S3-4 Structure names, sources and bioactivity of <i>homo</i> -[4 + 2] cycloaddition type LS dimers (189-215).....	S17
Table S3-5 Structure names, sources and bioactivity of <i>homo</i> -[4 + 2] cycloaddition type LS dimers (216-240).....	S19
Table S3-6 Structure names, sources and bioactivity of <i>homo</i> -[4 + 2] cycloaddition type LS dimers (241-254).....	S21
Table S4 Structure names, sources and bioactivity of <i>homo</i> -[2 + 2] and [6 + 6] cycloaddition type LS dimers (255-271).....	S23
Table S5-1 Structure names, sources and bioactivity of <i>homo</i> -linear linkage type LS dimers (272-298).....	S25
Table S5-2 Structure names, sources and bioactivity of <i>homo</i> -linear linkage type LS dimers (299-304).....	S26
Table S6 Structure names, sources and bioactivity of <i>hetero</i> -[4 + 2] cycloaddition type LS dimers (305-311).....	S26
Table S7 Structure names, sources and bioactivity of <i>hetero</i> -[2 + 2 + 2] cycloaddition type LS dimers (312-323).....	S27
Table S8 Structure names, sources and bioactivity of <i>hetero</i> -linear linkage type LS dimers (324-335).....	S28
Table S9 Structure names, sources and bioactivity of <i>homo</i> -[4 + 2] cycloaddition type LS trimers (336-345).....	S29
Table S10 Structure names, sources and bioactivity of <i>homo</i> -linear linkage type LS trimers (346-348).....	S30
Table S11 Structure names, sources and bioactivity of <i>hetero</i> -LS trimers (349-354).....	S30
Reference	S31



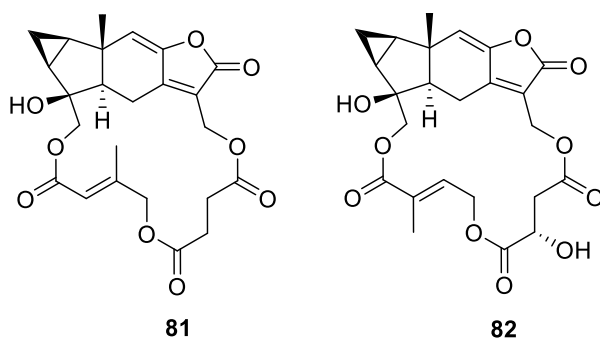
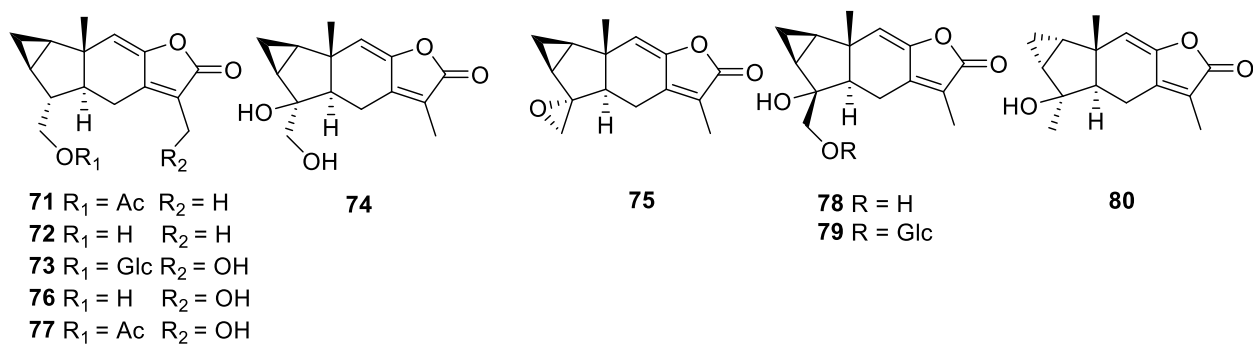
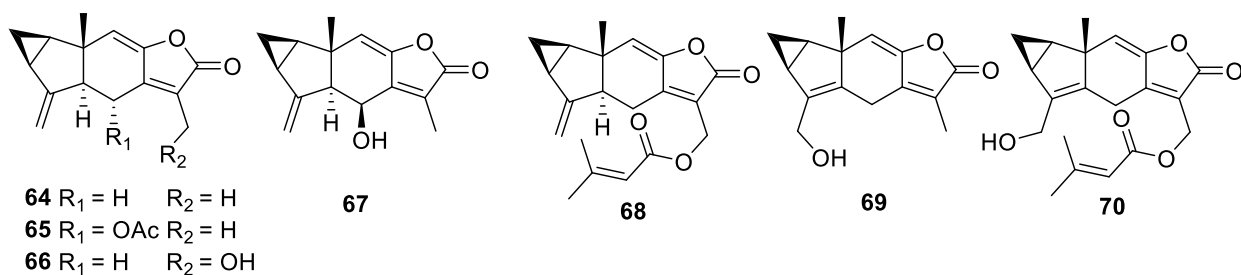
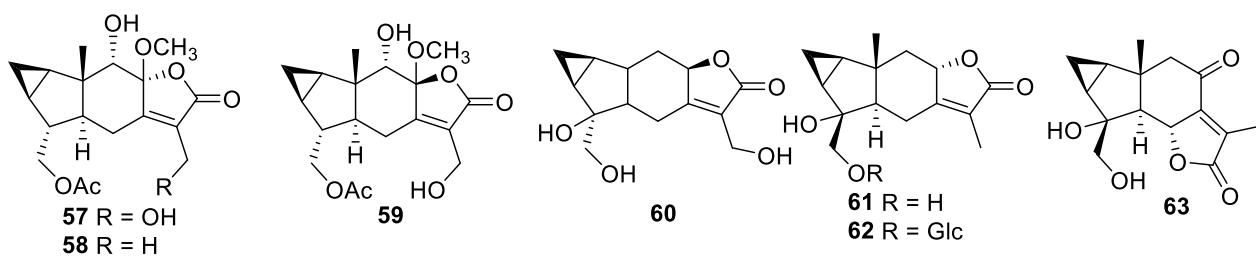
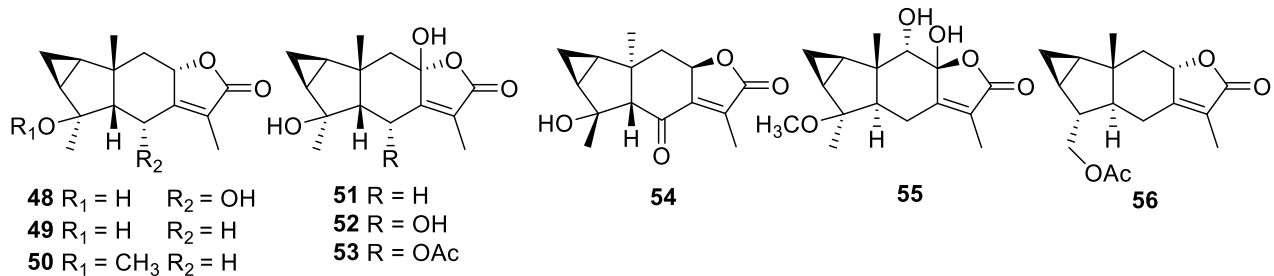


Table S1-1 Structure names, sources and bioactivity of ring-intact type LS monomers (1-27).

No	Compound	Sources	Bioactivity
1	Lindenenol	<i>Lindera strychnifolia</i> , ¹⁻⁴ <i>L. chunii</i> , ⁵ <i>L. aggregata</i> ^{6,7}	
2	Lindenyl acetate	<i>L. strychnifolia</i> , ² <i>L. chunii</i> , ⁵ <i>L. aggregata</i> ⁶	
3	Linderoxide	<i>L. strychnifolia</i> ⁸	
4	Lindenene	<i>L. strychnifolia</i> ^{4,9}	
5	Linderene	<i>L. strychnifolia</i> ¹⁰	
6	Linderene acetate	<i>L. strychnifolia</i> ¹⁰	
7	Isodihydrolinderene	<i>L. strychnifolia</i> ¹⁰	
8	Lindenone	<i>L. strychnifolia</i> ¹⁰	
9	Isolinderoxide	<i>L. strychnifolia</i> ¹¹	
10	Strychnistenolide A	<i>L. strychnifolia</i> ¹²	
11	6 α -acetyl-lindenanolide A (Strychnistenolide 6-O- acetate B1)	<i>L. aggregata</i> , ¹³ <i>L. chunii</i> , ⁵ <i>L. strychnifolia</i> ¹²	
12	Strychnilactone 2,6- dihydroxyxanthone	<i>L. strychnifolia</i> ^{3,13}	
13	Chlorajapolide F	<i>Chloranthus japonicus</i> ¹⁴	Cytotoxicity ¹⁴
14	Chlojaponilactone G	<i>C. japonicus</i> ¹⁵	
15	Decorone A	<i>Gochmatia decora</i> ¹⁶	
16	Linderin A	<i>L. aggregata</i> ⁶	
17	Strychnistenolide B	<i>L. strychnifolia</i> ¹²	
18	6 α -acetyl-lindenanolides B (Strychnistenolide 6-O- acetate B2)	<i>L. aggregata</i> , ¹² <i>L. chunii</i> , ⁵ <i>L. strychnifolia</i> ¹³	
19	Lindenanolide H	<i>L. chunii</i> , ⁵ <i>L. strychnifolia</i> ⁴	
20	Shizukanolide (Shizukanolide A)	<i>C. japonicus</i> , ^{14,17,18} <i>L. strychnifolia</i> ⁴	Cytotoxicity ¹⁴
21	Heterogorgiolide	<i>Heterogorgia uatumani</i> ¹⁹	
22	9-hydroxy heterogorgiolide (Chlorajapolide Fa)	<i>C. japonicus</i> , ^{14,20} <i>S. glabra</i> ²¹	Collagen inhibitory ²⁰
23	Chlojaponilactone F	<i>C. japonicus</i> ¹⁵	
24	Chlojaponilactone H	<i>C. japonicus</i> ¹⁵	Antifungal ¹⁵
25	Chlorajaposide	<i>C. japonicus</i> ²²	
26	Chloranthalactone E 8-O- β - D-glucopyranoside	<i>Sarcandra glabra</i> ²³	Hepatoprotective ²³
27	Rosmarylchloranthalactone E	<i>C. japonicus</i> ²⁴	PDE4 inhibitory ²⁴

Table S1-2 Structure names, sources and bioactivity of ring-intact type LS monomers (27-55).

No	Compound	Sources	Bioactivity
28	Chloranthalactone D	<i>C. japonicus</i> , ²⁵ <i>L. strychnifolia</i> ^{4, 13}	
29	Chloranthalactone E	<i>C. japonicus</i> , ^{14, 15, 25} <i>C. glaber</i> , ²⁶ <i>S. glabra</i> ²⁷⁻²⁹	Cytotoxicity ¹⁴
30	Linderolide T	<i>L. strychnifolia</i> ¹³	
31	Decorone B	<i>G. decora</i> ¹⁶	
32	Sarcandralactone A	<i>S. glabra</i> ²⁸	
33	Menelloide C	<i>Gorgonian Coral Menella sp</i> ³⁰ <i>L. strychnifolia</i> ¹³	
34	Onoseriolide A*	<i>Leontopodium leontopodioides</i> ³¹	
35	8 α ,9-dihydroonoseriolide senecioute	<i>L. leontopodioides</i> ³¹	
36	Lindenanolide A	<i>L. strychnifolia</i> ^{4, 32}	
37	Linderolide V	<i>L. strychnifolia</i> ³³	ARE-Inducing activity ³³
38	Onoseriolide acetate	<i>Hyalis argentea var. latisquama</i> ³⁴	
39	Wunderoild	<i>Wunderlichia Mirabilis</i> ³⁵	
40	Sibirolide A	<i>Xanthium sibiricum</i> ³⁶	
41	Isoshizukanolide	<i>C. fortunei</i> ³⁷	
42	8 β ,9 α -dihydroxylindan-4(5),7(11)-dien-8 α ,12-olide	<i>S. glabra</i> ²⁷	
43	Glabranol A	<i>S. glabra</i> , ³⁸ <i>C. japonicus</i> ³⁹	
44	Chlorajapolide A	<i>C. japonicus</i> ²²	
45	Chlorajapolide B	<i>C. japonicus</i> ²²	
46	Chlorajapolide C	<i>C. japonicus</i> ²²	Cytotoxicity ²²
47	Chlojaponilacton D	<i>C. japonicus</i> ¹⁸	
48	Linderolide K	<i>L. strychnifolia</i> ⁴	
49	Linderolide N	<i>L. strychnifolia</i> ¹³	
50	Linderolide O	<i>L. strychnifolia</i> ¹³	Anti-Inflammatory ¹³
51	Linderolide P	<i>L. strychnifolia</i> ¹³	Anti-Inflammatory ¹³
52	Linderolide Q	<i>L. strychnifolia</i> ¹³	
53	Linderolide R	<i>L. strychnifolia</i> ¹³	
54	Linderolide S	<i>L. strychnifolia</i> ¹³	
55	Sarcandralactone C	<i>S. glabra</i> ²⁹	

*Compound 34 and compound 66 possessing the same name, in order to distinguish them, we named 34 Onoseriolide A.

Table S1-3 Structure names, sources and bioactivity of ring-intact type LS monomers (**56-81**).

No	Compound	Sources	Bioactivity
56	Chlojaponilactone E	<i>C. japonicus</i> ^{14, 18}	Cytotoxicity ¹⁴
57	Chlorajapolide E	<i>C. japonicus</i> ²²	
58	Chlorajapolide H	<i>C. japonicus</i> ¹⁴	Cytotoxicity ¹⁴
59	Chlojaponilactone I	<i>C. japonicus</i> ¹⁵	
60	Decorone D	<i>G. decora</i> ¹⁶	
61	Chlorajapolide D	<i>C. japonicus</i> , ²² <i>C. sessilifolius</i> ⁴⁰	
62	Yinxiancaoside A	<i>C. japonicus</i> , ⁴¹ <i>S. glabra</i> ²¹	Cytotoxicity ⁴¹
63	Myrrhalindenane B	<i>L. myrrha</i> ⁴²	
64	Chloranthalactone A (Shizukanolide B)	<i>S. glabra</i> , ⁴³ <i>C. glaber</i> , ^{26, 44} <i>C. japonicus</i> , ^{15, 17, 25} <i>C. tianmushanensis</i> , ⁴⁵ <i>Hedyosmum angustifolium</i> ⁴⁶	Cytotoxicity ²⁵
65	Chlojaponilactone B	<i>C. japonicus</i> ¹⁸	
66	Onoseriolid B*	<i>H. angustifolium</i> ⁴⁶	
67	Sibirolide B	<i>X. sibiricum</i> ³⁶	
68	Onoseriolide senecioate	<i>L. leontopodioides</i> , ³¹ <i>H. argentea</i> var. <i>latisquama</i> ³⁴	
69	Sarcandralactone D	<i>S. glabra</i> ²⁹	
70	15-hydroxyisoonoseriolide senecioate	<i>L. leontopodioides</i>	
71	Chloranthalactone C	<i>C. japonicus</i> , ^{18, 25, 47, 48} <i>C. fortunei</i> , ⁴⁹ <i>S. glabra</i> ²⁸	
72	Shizukanolide C	<i>C. japonicus</i> , ^{18, 47, 48} <i>C. fortunei</i> , ⁴⁹	
73	Chloranoside B	<i>C. glaber</i> ⁵⁰	
74	Chlorajaponol F	<i>C. japonicus</i> ³⁹	
75	Chloranthalactone G	<i>S. glabra</i> ⁴³	
76	Shizukanolide F	<i>C. japonicus</i> , ⁴⁷ <i>C. fortunei</i> ⁴⁹	
77	Shizukanolide H	<i>C. fortunei</i> ⁴⁹	
78	Shizukanolide E	<i>C. japonicus</i> ⁴⁷	
79	Chloranoside A	<i>C. glaber</i> , ⁵⁰ <i>S. glabra</i> , ^{21, 23} <i>C. japonicus</i> , ⁴¹ <i>C. fortunei</i> ⁴⁹	Hepatoprotective ²³ Cytotoxicity ⁴¹
80	Linderaggregin B	<i>L. aggregata</i> ⁵¹	
81	Chlorafortulide	<i>C. fortunei</i> ⁵²	
82	Sarglabolide L	<i>S. glabra</i> ⁵³	Anti-Inflammatory ⁵³

*Compound 66 and compound 34 possessing the same name, in order to distinguish them, we named 66 Onoseriolide B.

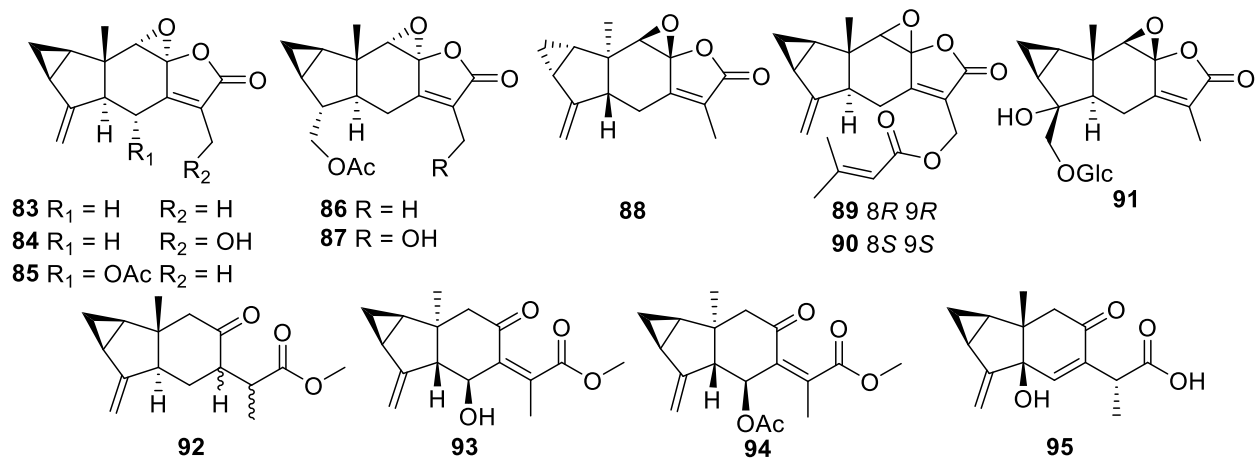


Table S1-4 Structure names, sources and bioactivity of ring-intact type LS monomers (**82-95**).

No	Compound	Sources	Bioactivity
83	Chloranthalactone B	<i>C. glaber</i> , ^{26, 44} <i>S. glabra</i> , ²¹ <i>C. japonicus</i> ^{14, 15, 25}	Cytotoxicity ^{14, 25}
84	Oxyonoseriolide	<i>H. angustifolium</i> ⁴⁶	Cytotoxicity ⁴⁶
85	Chlojaponilactone C	<i>C. japonicus</i> ¹⁸	
86	Shizukanolide D	<i>C. japonicus</i> ⁴⁷	
87	Shizukanolide G	<i>C. fortunei</i> ⁴⁹	
88	(+)-chloranthalactone B	<i>Menella sp</i> ⁵⁴	
89	13-desoxyisoonoseriolide	<i>H. argentea var. latisquama</i> ³⁴	
90	8β,9β-epoxyonoseriolide senecioate	<i>L. leontopodioides</i>	
91	Sarcaglaboside F	<i>S. glabra</i> ²¹	
92	Hedyosmone	<i>H. angustifolium</i> ⁴⁶	
93	Linderolide U	<i>L. aggregata</i> , ⁷ <i>L. strychnifolia</i> ³³	
94	Linderolide L	<i>L. strychnifolia</i> ⁴	
95	Myrrhalindenane A	<i>L. myrrha</i> ⁴²	

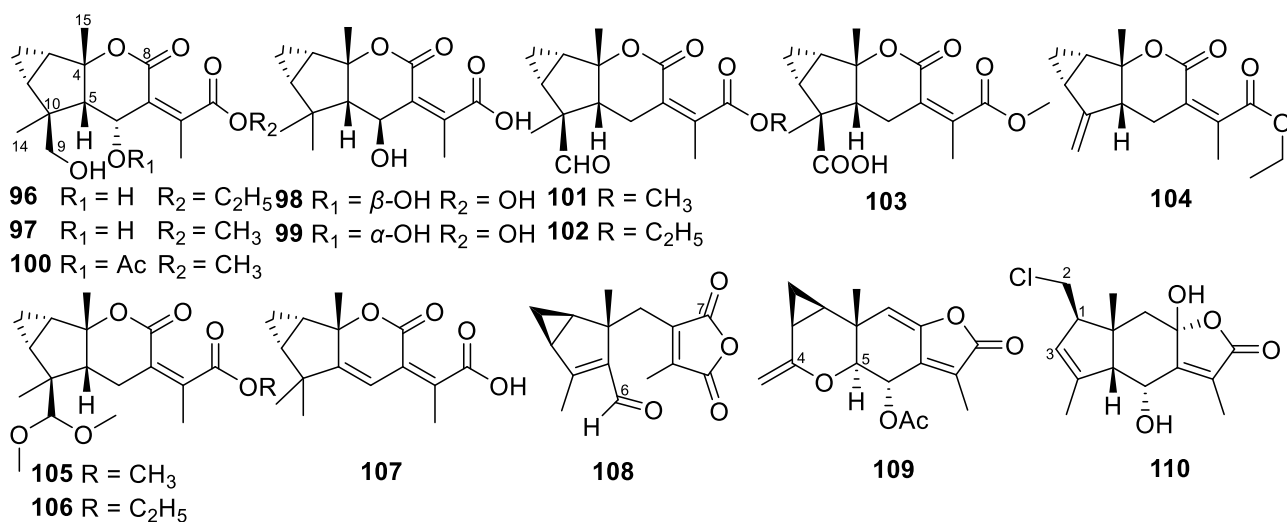
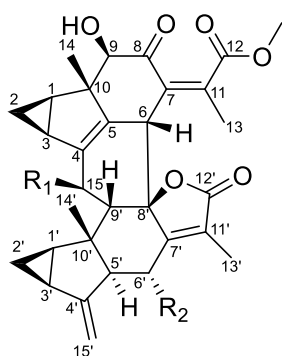
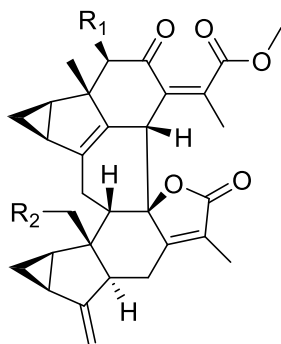


Table S2 Structure names, sources and bioactivity of ring-*seco* type LS monomers (**96-110**).

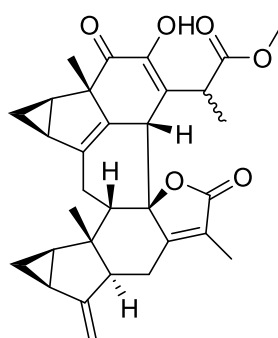
No	Compound	Sources	Bioactivity
96	Strychnilactone	<i>L. strychnifolia</i> ¹²	
97	Lindenanolide G	<i>L. chunii</i> , ⁵ <i>L. strychnifolia</i> ¹³	
98	Linderagalactone B	<i>L. aggregata</i> ⁵⁵	
99	Linderolide M	<i>L. strychnifolia</i> ⁴	
100	Rotundusolide A	<i>Cyperus rotundus</i> ⁵⁶	
101	Chloranerectuslactone V	<i>C. erecyus</i> ⁵⁷	
102	Sarglactone I	<i>S. glabra</i> ⁵⁸	Cytotoxicity ⁵⁸
103	Sarglactone J	<i>S. glabra</i> ⁵⁸	
104	Sarglactone K	<i>S. glabra</i> ⁵⁸	
105	Sarglactone L	<i>S. glabra</i> ⁵⁸	
106	Sarglactone M	<i>S. glabra</i> ⁵⁸	
107	Linderagalactone C	<i>L. aggregata</i> ⁵⁵	
108	Lindenanolide E	<i>L. chunii</i> ⁵	
109	Linderaggregedin C	<i>L. aggregata</i> ⁵¹	
110	Linderagalactone A	<i>L. aggregata</i> ⁵⁵	



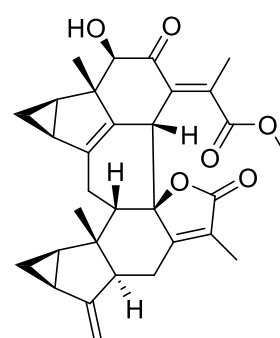
111 $R_1 = H$ $R_2 = H$
 113 $R_1 = H$ $R_2 = OAc$
 114 $R_1 = OH$ $R_2 = H$



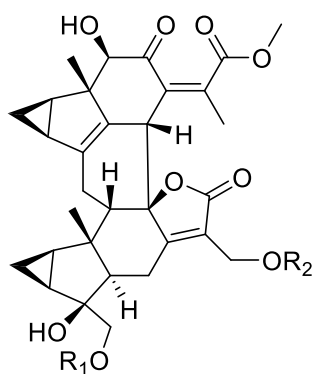
112 $R_1 = OAc$ $R_2 = H$
 115 $R_1 = H$ $R_2 = H$
 117 $R_1 = H$ $R_2 = OAc$



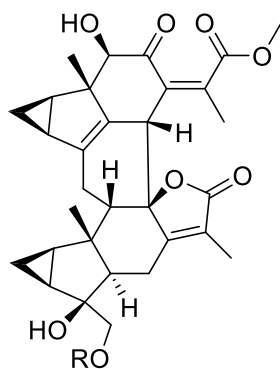
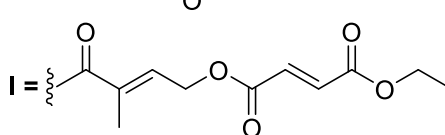
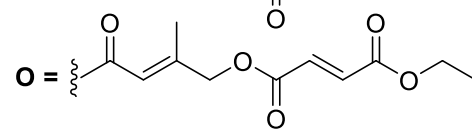
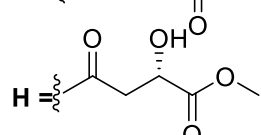
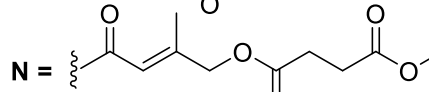
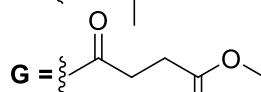
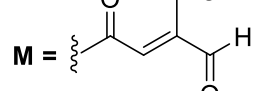
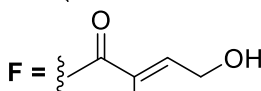
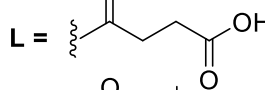
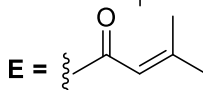
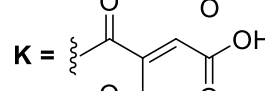
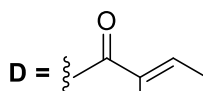
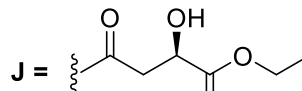
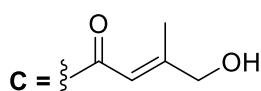
116



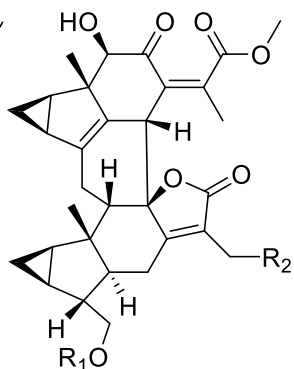
118



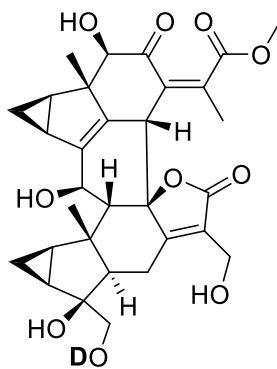
119 $R_1 = Ac$ $R_2 = H$ 133 $R_1 = H$ $R_2 = H$
 121 $R_1 = C$ $R_2 = H$ 134 $R_1 = F$ $R_2 = H$
 122 $R_1 = D$ $R_2 = Ac$ 135 $R_1 = F$ $R_2 = J$
 123 $R_1 = E$ $R_2 = Ac$ 136 $R_1 = O$ $R_2 = H$
 124 $R_1 = Ac$ $R_2 = H$ 137 $R_1 = D$ $R_2 = L$
 126 $R_1 = D$ $R_2 = H$ 138 $R_1 = M$ $R_2 = G$
 128 $R_1 = F$ $R_2 = H$ 139 $R_1 = N$ $R_2 = H$
 129 $R_1 = D$ $R_2 = G$ 140 $R_1 = E$ $R_2 = H$
 130 $R_1 = C$ $R_2 = L$ 141 $R_1 = C$ $R_2 = H$
 131 $R_1 = C$ $R_2 = G$ 142 $R_1 = C$ $R_2 = L$
 132 $R_1 = I$ $R_2 = H$ 145 $R_1 = Ac$ $R_2 = Ac$



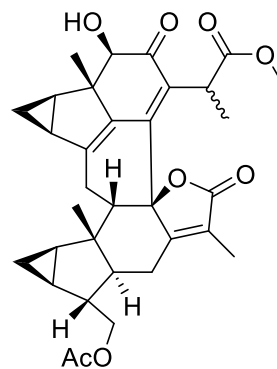
120 $R = C$
 125 $R = D$
 127 $R = K$



143 $R_1 = H$ $R_2 = OH$
 146 $R_1 = Ac$ $R_2 = OH$
 147 $R_1 = Ac$ $R_2 = H$



144



148

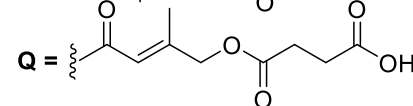
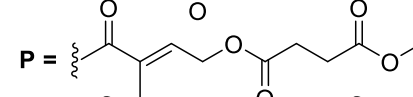
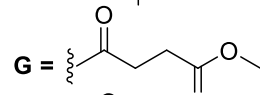
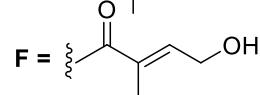
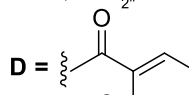
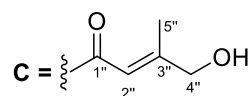
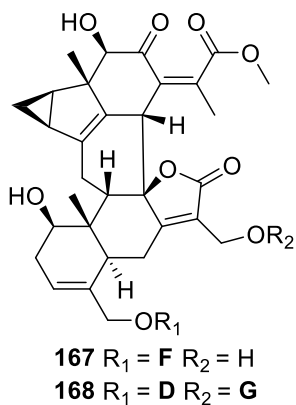
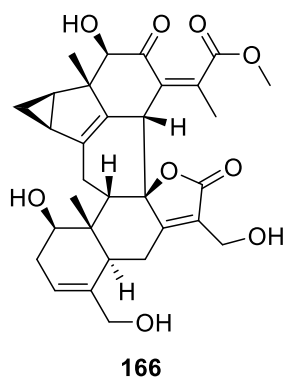
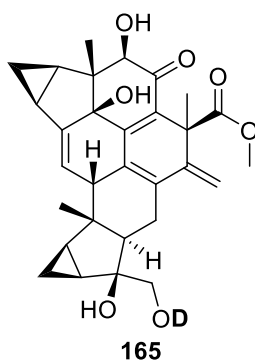
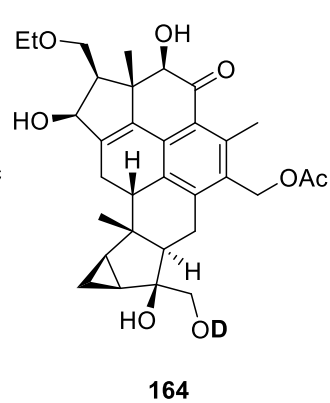
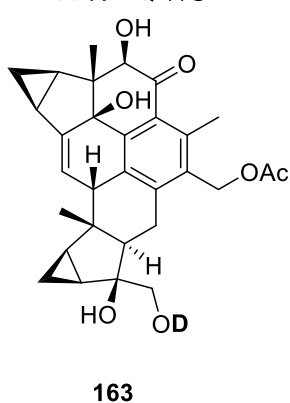
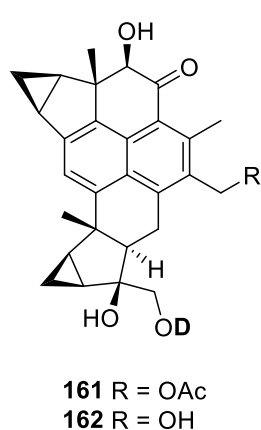
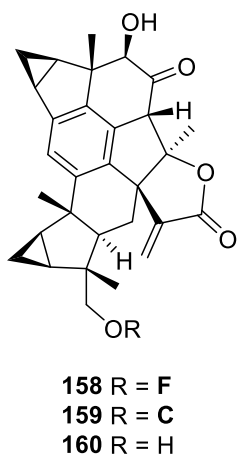
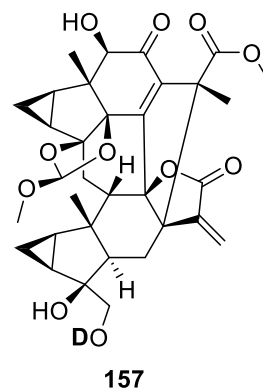
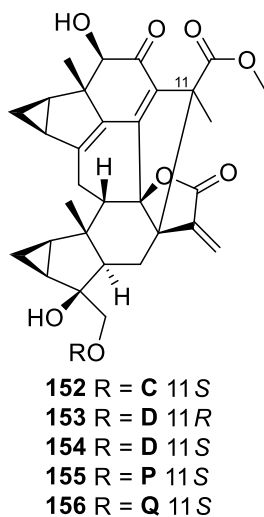
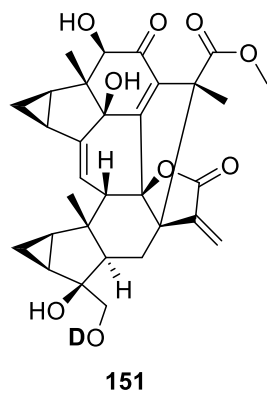
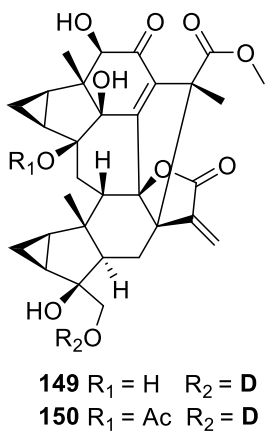


Table S3-1 Structure names, sources and bioactivity of *homo*-[4 + 2] cycloaddition type LS dimers (111-136).

No	Compound	Sources	Bioactivity
111	Shizukaol A	<i>C. japonicus</i> , ⁵⁹⁻⁶¹	
112	Shizukaol A acetate	<i>C. serratus</i> , ⁶² <i>C. fortunei</i> ⁶³ <i>C. japonicus</i> ⁶⁴	
113	Chlojapolide D	<i>C. japonicus</i> ⁵⁹	
114	Chlojapolide E	<i>C. japonicus</i> ⁵⁹	
115	Chlojapolide F	<i>C. japonicus</i> ⁵⁹	
116	Chlojapolide B	<i>C. japonicus</i> ⁵⁹	
117	Chololactone D	<i>C. holostegius</i> ⁶⁵	
118	Chlojapolide C	<i>C. japonicus</i> ⁵⁹	
119	Shizukaol N	<i>C. fortunei</i> , ⁶³ <i>S. glabra</i> ⁶⁶	
120	Shizukaol M	<i>C. fortunei</i> ^{37, 52, 63}	Anti-Inflammatory ⁵²
121	Shizukaol I	<i>C. japonicus</i> , ⁶⁷ <i>C. fortunei</i> ^{37, 63}	
122	Chlorahololide D (henriol D)	<i>C. holostegius</i> , ⁶⁸ <i>C. henryi</i> , ⁶⁹ <i>C. fortunei</i> , ^{37, 63} <i>C. spicatus</i> , ⁷⁰ <i>S. glabra</i> , ²⁹ <i>C. serratus</i> ⁷¹	Antimalarial ³⁷
123	Shizukaol K	<i>C. fortunei</i> ^{37, 63}	
124	Multistalide B	<i>C. multistachys</i> ⁷²	
125	Sarcandrolide A	<i>S. glabra</i> ²⁸	Cytotoxicity ²⁸
126	Shizukaol C	<i>C. serratus</i> , ⁶² <i>S. glabra</i> , ^{28, 29, 66, 73} <i>C. multistachys</i> , ^{74, 75} <i>C. japonicus</i> , ^{14,} ^{60, 76} <i>C. fortunei</i> , ^{37, 52} <i>C. holostegius</i> ⁶⁵	Anti-HIV ⁶⁰ Cytotoxicity ^{52, 60, 73, 76}
127	Shizukaol L	<i>C. fortunei</i> ⁶³	
128	Sarcandrolide B	<i>S. glabra</i> ²⁸	Cytotoxicity ²⁸
129	13'- <i>O</i> -Methyl succinylshizukaol C *	<i>C. japonicus</i> ²²	
130	Chololactone E	<i>C. holostegius</i> ⁶⁵	Anti-Inflammatory ⁶⁵
131	Chololactone F	<i>C. japonicus</i> , ⁶⁰ <i>C. holostegius</i> ⁶⁵	Anti-Inflammatory ⁶⁵
132	Sarglabolide H	<i>S. glabra</i> ⁶⁶	
133	Sarglabolide I	<i>S. glabra</i> , ⁶⁶ <i>C. fortunei</i> ³⁷	
134	Sarglabolide J	<i>S. glabra</i> , ⁶⁶ <i>C. fortunei</i> ³⁷	Antimalarial ³⁷
135	Sarglabolide K	<i>S. glabra</i> ⁶⁶	
136	Fortunilide M	<i>C. fortunei</i> ⁷⁷	

The structure of Chlorajaponol (129) with $\Delta^{4,5}$ double bonds should be of *E*-geometry, revised to 3'-*O*-Methyl succinylshizukaol C.

Table S3-2 Structure names, sources and bioactivity of *homo*-[4 + 2] cycloaddition type LS dimers (137-163).

No	Compound	Sources	Bioactivity
137	Chlojapolide A	<i>C. japonicus</i> ⁵⁹	Anti-Inflammatory ⁵⁹
138	Fortunilide A	<i>C. fortunei</i> ³⁷	
139	Fortunilide B	<i>C. fortunei</i> ³⁷	Antimalarial ³⁷
140	Fortunilide C	<i>C. fortunei</i> ³⁷	
141	Chlorahupetol F	<i>Chloranthus henryi</i> var. <i>hupehensis</i> ⁷⁸	
142	Shizukaol O	<i>C. fortunei</i> ^{52, 63} , <i>C. japonicus</i> ¹⁴	Anti-Inflammatory ⁵² Cytotoxicity ¹⁴
143	Sarcandrolide J	<i>S. glabra</i> , ²⁹ <i>C. fortunei</i> ³⁷	
144	Chlomultiol A	<i>C. multistachys</i> ⁷⁴	Anti-Inflammatory ⁷⁴
145	Sarcanolide E	<i>S. glabra</i> ⁷⁹	Anti-Inflammatory ⁷⁹
146	Shizukaol D	<i>C. serratus</i> , ^{62, 71, 80} <i>C. fortunei</i> , ⁵² <i>C. spicatus</i> , ⁷⁰ <i>C. multistachys</i> , ⁷⁵ <i>S. glabra</i> , ^{29, 73} <i>C. japonicus</i> ^{14, 60, 81}	Cytotoxicity ¹⁴ Anti-Inflammatory ⁵²
147	Shizukaol E	<i>C. japonicus</i> , ^{67, 82, 83} <i>C. fortunei</i> , ^{52, 63} <i>S. glabra</i> ²⁸	Anti-Inflammatory ⁵² Anti-HIV ⁸³ Anti-HCV ⁸³
148	Chololactone G	<i>C. holostegius</i> ⁶⁵	Anti-Inflammatory ⁶⁵
149	Sarcanolide A	<i>S. hainanensis</i> ⁸⁴	Cytotoxicity ⁸⁴
150	Sarcanolide D	<i>S. glabra</i> ⁷⁹	Anti-Inflammatory ⁷⁹
151	Sarcanolide B	<i>S. hainanensis</i> ⁸⁴	Cytotoxicity ⁸⁴
152	Fortunilide N	<i>C. fortunei</i> ⁷⁷	
153	Chololactone A	<i>C. holostegius</i> ⁶⁵	Anti-Inflammatory ⁶⁵
154	Fortunilide K	<i>C. fortunei</i> ³⁷	
155	Fortunilide L	<i>C. fortunei</i> ³⁷	
156	Fortunilide O	<i>C. fortunei</i> ⁷⁷	
157	Sarcanolide C	<i>S. glabra</i> ⁷⁹	Anti-Inflammatory ⁷⁹
158	Chlorahupetone G	<i>C. henryi</i> var. <i>hupehensis</i> ⁸⁵	Cytotoxicity ⁸⁵
159	Chlorahupetone H	<i>C. henryi</i> var. <i>hupehensis</i> ⁸⁵	Cytotoxicity ⁸⁵
160	Chlorahupetone I	<i>C. henryi</i> var. <i>hupehensis</i> ⁸⁵	Cytotoxicity ⁸⁵
161	Sarglaromatic A	<i>S. glabra</i> ⁸⁶	Anti-Nonalcoholic ⁸⁶
162	Sarglaromatic B	<i>S. glabra</i> ⁸⁶	Anti-Nonalcoholic ⁸⁶
163	Sarglaromatic C	<i>S. glabra</i> ⁸⁶	Anti-Nonalcoholic ⁸⁶

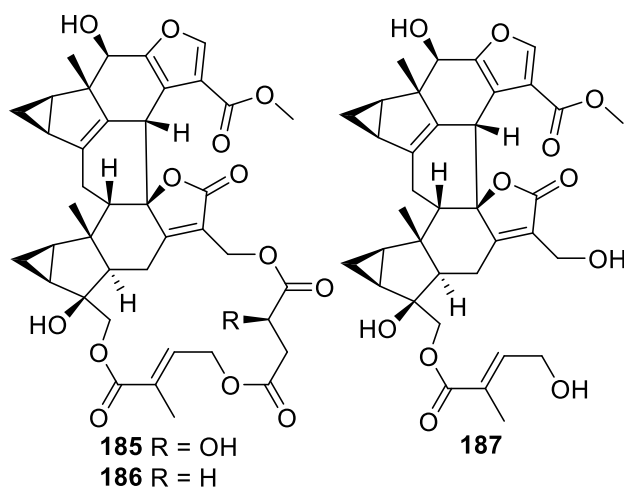
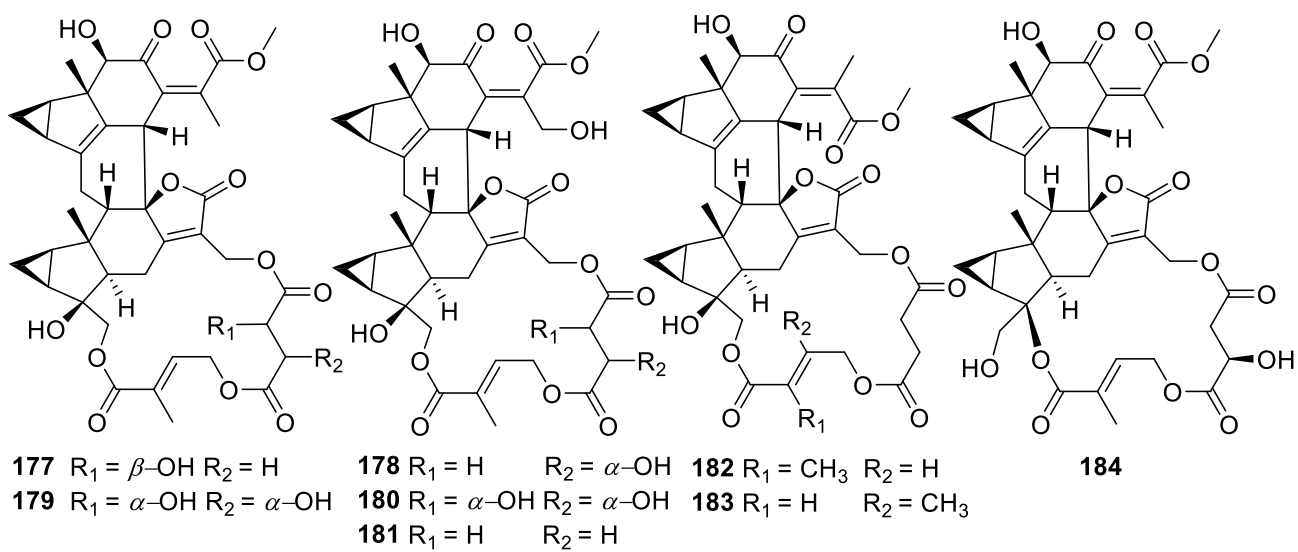
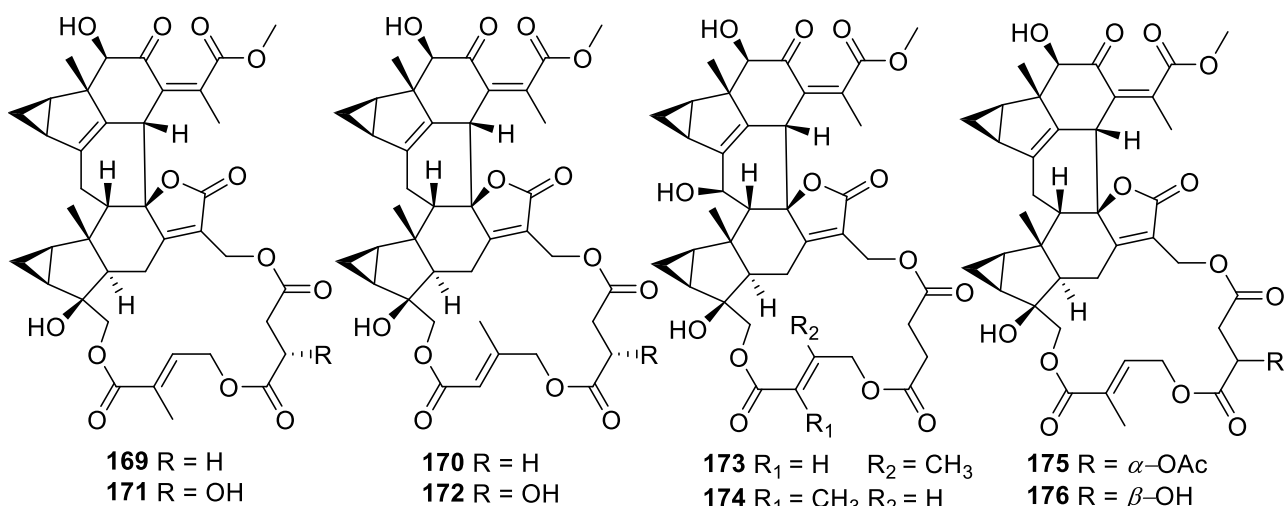
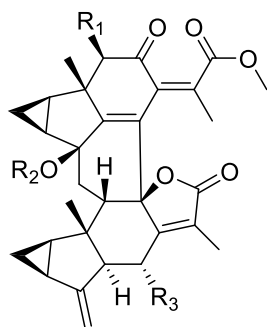
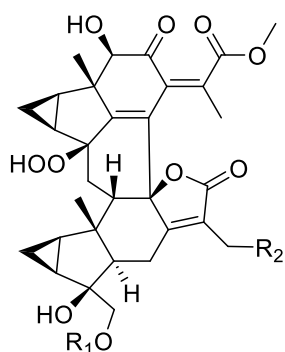


Table S3-3 Structure names, sources and bioactivity of *homo*-[4 + 2] cycloaddition type LS dimers (164-188).

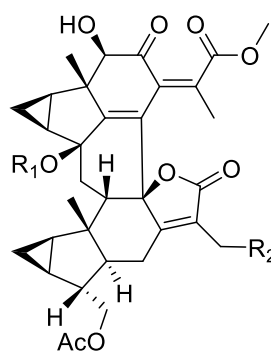
No	Compound	Sources	Bioactivity
164	Sarglaromatic D	<i>S. glabra</i> ⁸⁶	
165	Sarglaromatic E	<i>S. glabra</i> ⁸⁶	
166	Fortunoid C	<i>C. fortunei</i> ³⁹	Antimalarial ³⁹
167	15'- <i>O</i> -(4-Hydroxytigloyl)fortunoid C	<i>C. fortunei</i> ⁸⁷	
168	13'- <i>O</i> -Methyl succinyl-15'- <i>O</i> -tigloylfortunoid C	<i>S. glabra</i> ⁸⁷	
169	Shizukaol B	<i>C. serratus</i> , ^{62, 71, 80} <i>C. fortunei</i> , ^{37, 52, 63} <i>C. japonicus</i> , ^{59, 76} <i>S. glabra</i> , ^{28, 66} <i>C. spicatus</i> ⁷⁰ , <i>C. tianmushanensis</i> ⁴⁵	Anti-HIV ^{60, 70} Cytotoxicity ^{52, 60, 70} Anti-Inflammatory ^{14, 66, 76, 88}
170	Shizukaol F	<i>C. japonicus</i> , ^{59, 60, 67, 76} <i>C. fortunei</i> , ^{37, 49, 63} <i>C. spicatus</i> ⁷⁰	Antimalarial ³⁷ Anti-HIV ⁷⁰ Cytotoxicity ⁷⁰ Antimalarial ³⁷
171	Shizukaol G	<i>C. japonicus</i> , ^{67, 76, 83} <i>S. glabra</i> , ^{28, 66} <i>C. fortunei</i> ^{37, 52}	Anti-Inflammatory ^{52, 66, 88} Antimalarial ³⁷
172	Shizukaol H	<i>C. japonicus</i> , ^{60, 67} <i>S. glabra</i> ²⁹	Anti-HIV ⁶⁰ Cytotoxicity ⁶⁰
173	Shizukaol P	<i>C. fortunei</i> , ⁴⁹ <i>C. spicatus</i> ⁷⁰	
174	Chloramultiol A	<i>C. multistachys</i> ⁷⁵	Cytotoxicity ⁷⁵
175	Sarcandrolide C	<i>S. glabra</i> ²⁸	
176	Sarglabolide B	<i>S. glabra</i> ⁶⁶	
177	Sarglabolide C	<i>S. glabra</i> ⁶⁶	Cytotoxicity ⁶⁶
178	Sarglabolide D	<i>S. glabra</i> ⁶⁶	
179	Sarglabolide E	<i>S. glabra</i> ⁶⁶	
180	Sarglabolide F	<i>S. glabra</i> ⁶⁶	
181	Sarglabolide G	<i>S. glabra</i> ⁶⁶	
182	Henriol C	<i>C. henryi</i> , ⁶⁹ <i>C. serratus</i> ⁷¹	Cytotoxicity ⁶⁹
183	Fortunilide G	<i>C. fortunei</i> , ³⁷ <i>C. holostegius</i> ⁶⁵	
184	Sarglabolide A	<i>S. glabra</i> ⁶⁶	Anti-Inflammatory ⁶⁶
185	Sarglafuran A	<i>S. glabra</i> ⁸⁹	
186	Chlorahupetol C	<i>C. henryi</i> var. <i>hupehensis</i> ⁷⁸	
187	Chlorahupetol D	<i>C. henryi</i> var. <i>hupehensis</i> ⁷⁸	
188	Chlorahololide A	<i>C. holostegius</i> ⁹⁰	Voltagegated potassium activity ⁹⁰



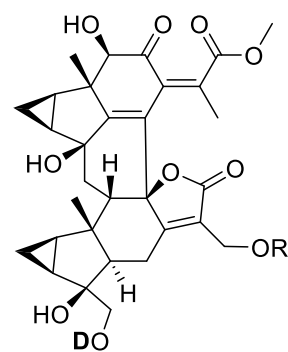
188 R₁ = H R₂ = H R₃ = OAc
189 R₁ = OH R₂ = H R₃ = OAc
190 R₁ = OH R₂ = OH R₃ = H



191 R₁ = D R₂ = OAc
194 R₁ = D R₂ = OH
197 R₁ = N R₂ = OH
198 R₁ = C R₂ = OG
199 R₁ = H R₂ = H



192 R₁ = OH R₂ = OH
195 R₁ = H R₂ = OH
200 R₁ = OH R₂ = H



193 R = Ac
196 R = H

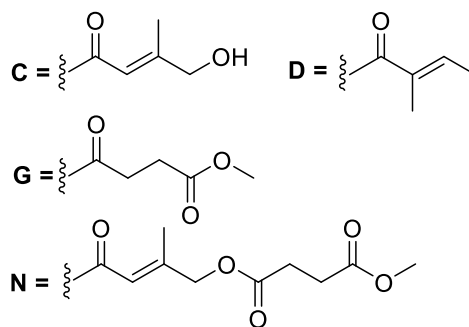
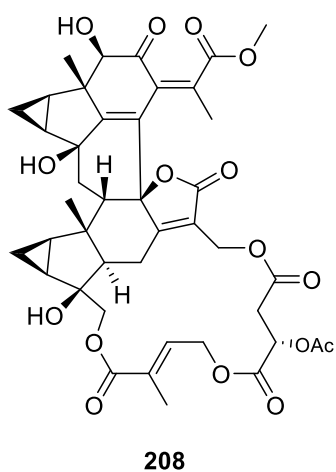
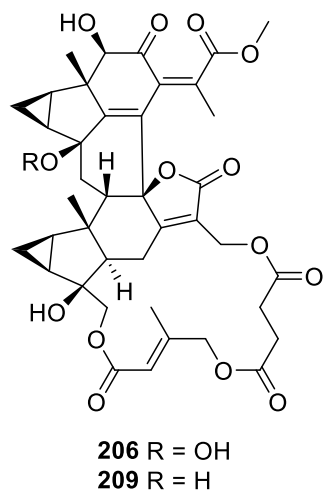
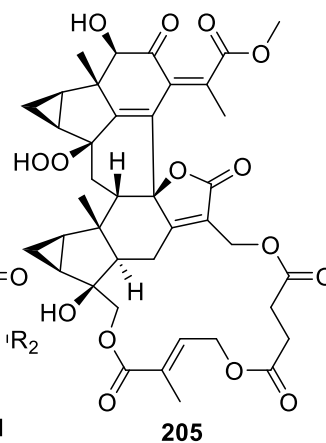
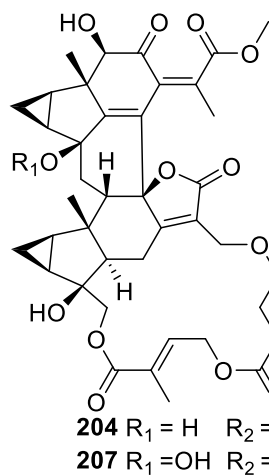
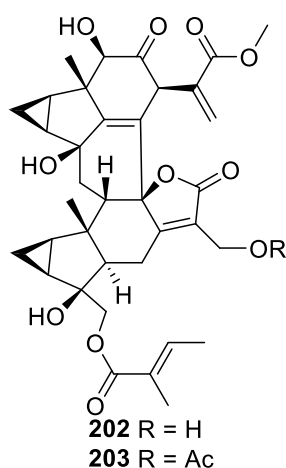
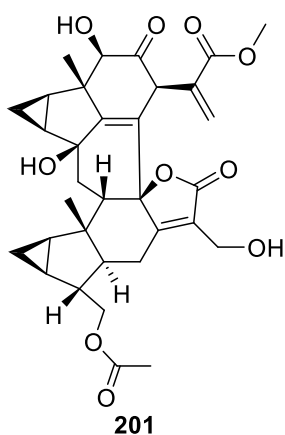


Table S3-4 Structure names, sources and bioactivity of *homo*-[4 + 2] cycloaddition type LS dimers (189-215).

No	Compound	Sources	Bioactivity
189	Chlorahololide C	<i>C. holostegius</i> , ⁶⁸ <i>C. japonicus</i> ⁵⁹	
190	Spicachlorantin J	<i>C. spicatus</i> , ⁷⁰ <i>C. japonicus</i> ⁵⁹	
191	Spicachlorantin E	<i>C. spicatus</i> ⁹¹	
192	Spicachlorantin F	<i>C. spicatus</i> , ⁹¹ <i>S. glabra</i> ²⁹	
193	Sarcandrolide E	<i>S. glabra</i> ^{28, 29, 73}	Cytotoxicity ⁷³
194	Chlorajaponilide E	<i>C. japonicus</i> , ⁶⁰ <i>S. glabra</i> ²⁹	
195	Spicachlorantin G	<i>C. spicatus</i> ⁷⁰	
196	Chlorasessilifol A	<i>C. sessilifolius</i> ⁴⁰	
197	Fortunilide D	<i>C. fortunei</i> ³⁷	
198	Fortunilide E	<i>C. fortunei</i> ³⁷	
199	Fortunilide F	<i>C. fortunei</i> ³⁷	
200	Chololactone B	<i>C. holostegius</i> ⁶⁵	Anti-Inflammatory ⁶⁵
201	Multistalide A	<i>C. multistachys</i> ⁷²	
202	Chlomultiol B	<i>C. multistachys</i> ⁷⁴	Anti-Inflammatory ⁷⁴
203	Sarcaglabrin B	<i>S. glabra</i> ⁷³	
204	Chloramultilide A	<i>C. multistachys</i> , ⁹² <i>C. spicatus</i> ^{70, 93}	
205	Spicachlorantin C	<i>C. spicatus</i> ⁹¹	
206	Spicachlorantin D	<i>C. spicatus</i> , ⁹¹ <i>C. fortunei</i> , ³⁷ <i>C. japonicus</i> ⁹⁴	
207	Sarcandrolide G	<i>S. glabra</i> , ²⁹ <i>C. japonicus</i> ⁸³	
208	Sarcandrolide H	<i>S. glabra</i> ²⁹	Cytotoxicity ²⁹
209	Chlorajaponilide H	<i>C. japonicus</i> ⁸³	
210	Sarcandrolide F	<i>S. glabra</i> , ²⁹ <i>C. japonicus</i> ⁸³	Cytotoxicity ²⁹ Anti-HIV ⁸³ Anti-HCV ⁸³
211	Chlorajaponilide F	<i>C. japonicus</i> ⁸³	Anti-HIV ⁸³ Anti-HCV ⁸³
212	Chlorajaponilide I	<i>C. japonicus</i> ⁹⁴	
213	Chlorajaponilide G	<i>C. japonicus</i> ⁸³	
214	Chlorahololide E	<i>C. holostegius</i> ⁶⁸	Voltagegated potassium activity ⁶⁸
215	Spicachlorantin H	<i>C. spicatus</i> , ⁷⁰ <i>C. japonicus</i> ^{59, 83}	

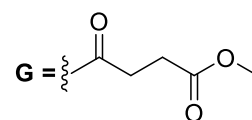
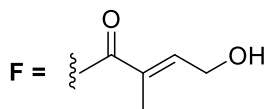
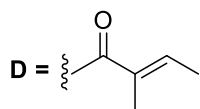
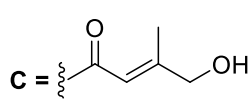
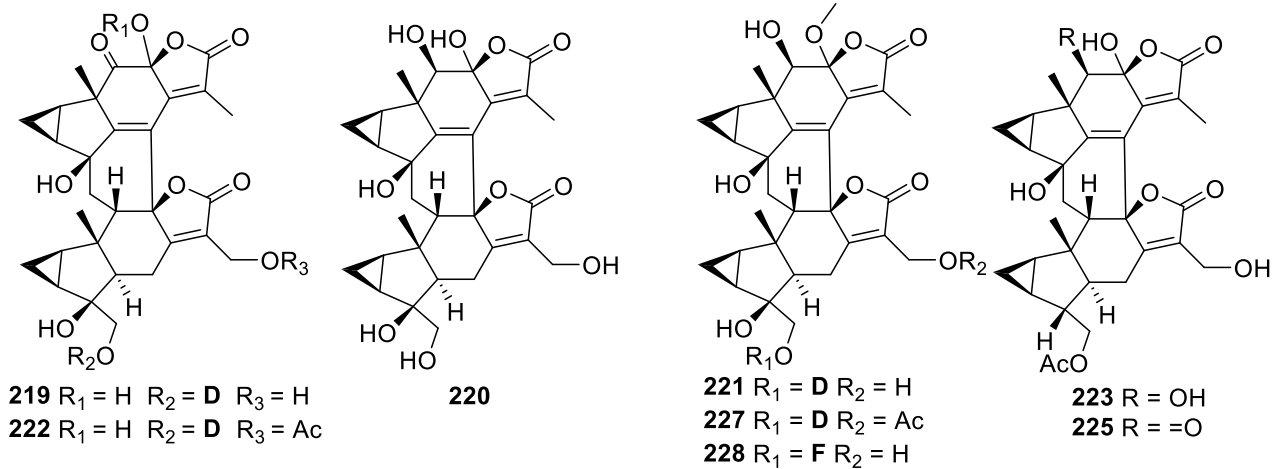
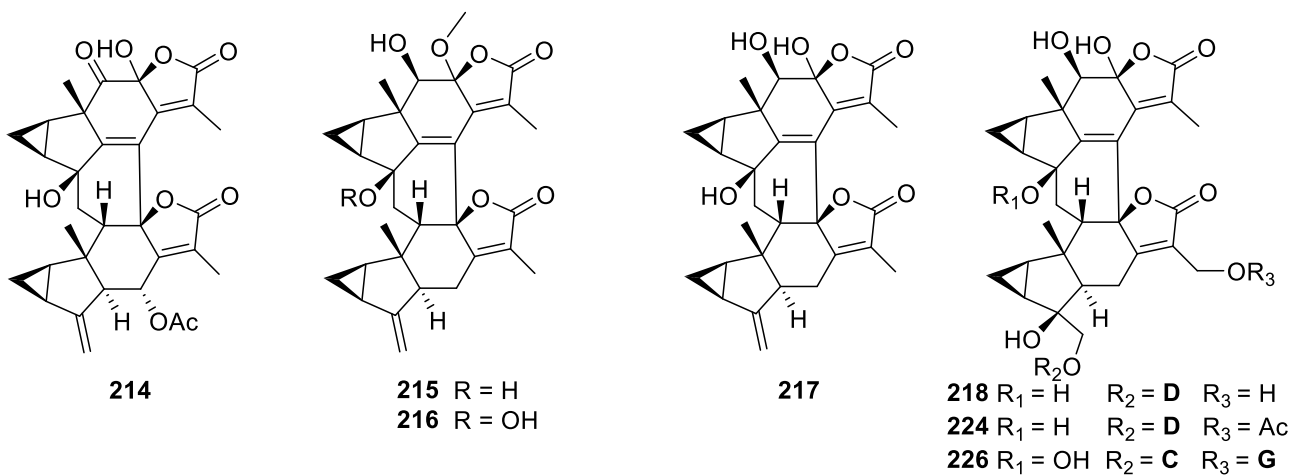
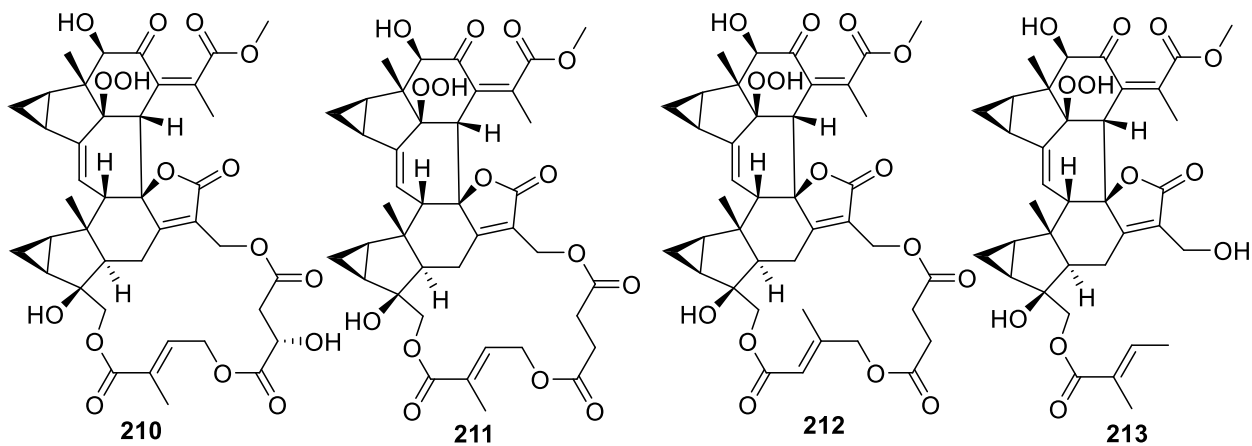
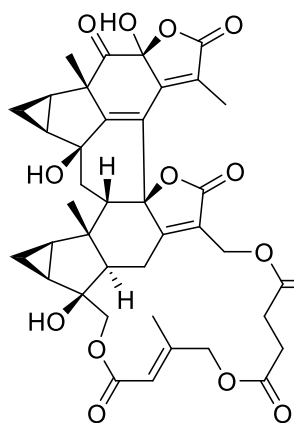
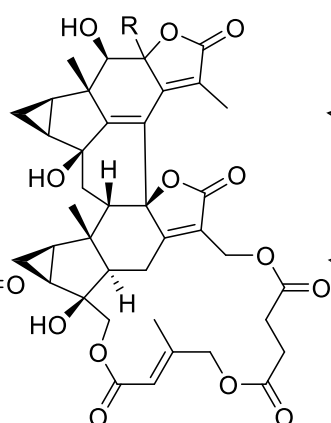


Table S3-5 Structure names, sources and bioactivity of *homo*-[4 + 2] cycloaddition type LS dimers (216-240).

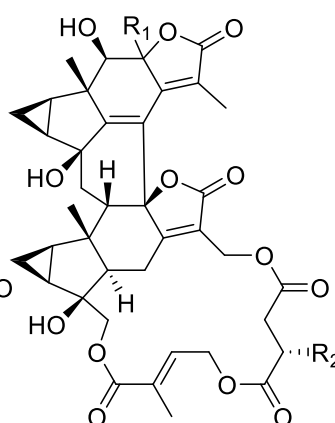
No	Compound	Sources	Bioactivity
216	Spicachlorantin I	<i>C. spicatus</i> ⁷⁰	
217	Chlojapolide G	<i>C. japonicus</i> ⁸³	
218	Chloramultilide D	<i>C. spicatus</i> ⁹⁵	
219	Chloramultiol D	<i>C. multistachys</i> ⁷⁵	Cytotoxicity ⁷⁵
220	Chloramultiol B	<i>C. multistachys</i> ⁷⁵	Cytotoxicity ⁷⁵
221	Chloramultiol C	<i>C. multistachys</i> ⁷⁵	Cytotoxicity ⁷⁵
222	Chlorahololide F	<i>C. holostegius</i> , ⁶⁸ <i>S. glabra</i> ²⁸	
223	Chloramultiol E	<i>C. multistachys</i> ⁷⁵	Cytotoxicity ⁷⁵
224	Sarcandrolide D	<i>S. glabra</i> , ^{28, 29, 73} <i>C. serratus</i> ⁷¹	Cytotoxicity
225	Chlorasessilifol B	<i>C. sessilifolius</i> ⁴⁰	
226	Fortunilide I	<i>C. fortunei</i> ³⁷	
227	Sarcaglabrin C	<i>S. glabra</i> ⁷³	
228	chlorahupetol E	<i>C. henryi</i> var. <i>hupehensis</i> ⁷⁸	
229	Chlorahololide B	<i>C. holostegius</i> , ⁹⁰ <i>C. japonicus</i> ⁶⁰	Voltagegated potassium activity ⁹⁰
230	Chloramultilide B	<i>C. spicatus</i> , ⁹⁵ <i>C. fortunei</i> , ⁴⁹ <i>C. japonicus</i> , ^{59, 83, 94} <i>C. serratus</i> ⁷¹	Antifungal ⁹⁵
231	Chloramultilide C	<i>C. spicatus</i> , ⁹⁵ <i>C. henryi</i> , ⁶⁹ <i>C. multistachys</i> , ⁷⁵ <i>C. japonicus</i> , ^{60, 94} <i>C. fortunei</i> ³⁷	
232	Tianmushanol	<i>C. tianmushanensis</i> , ⁴⁵ <i>C. multistachys</i> ⁷⁴	
233	8- <i>O</i> -methyltianmushanol	<i>C. tianmushanensis</i> , ⁴⁵ <i>C. spicatus</i> , ⁹³ <i>C. multistachys</i> , ⁷⁵ <i>C. japonicus</i> , ^{60, 88} <i>C. serratus</i> ⁷¹	Tyrosinase Inhibitory ⁴⁵
234	Yinxiancaol	<i>C. japonicus</i> , ^{60, 82} <i>C. fortunei</i> ⁴⁹	
235	Spicachlorantin A	<i>C. spicatus</i> , ⁹³ <i>C. japonicus</i> , ^{59, 60} <i>C. serratus</i> , ⁷¹ <i>C. tianmushanensis</i> , ⁴⁵ <i>C. multistachys</i> ⁷⁵	
236	Chlorajaponilide D	<i>C. japonicus</i> ⁶⁰	
237	chloraserratolide A	<i>C. serratus</i> ⁸⁰	
238	Sarcandrolide I	<i>S. glabra</i> ²⁹	
239	Chlomultiol C	<i>C. multistachys</i> ⁷⁴	Anti-Inflammatory ⁷⁴
240	chloraserratolide B	<i>C. serratus</i> ⁸⁰	Anti-Inflammatory ⁸⁰



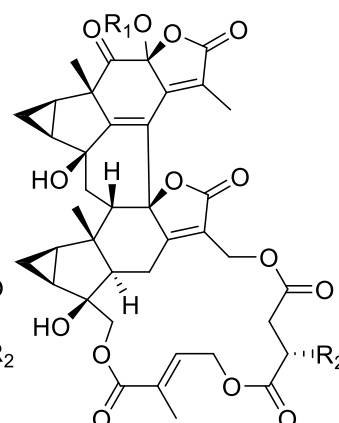
229



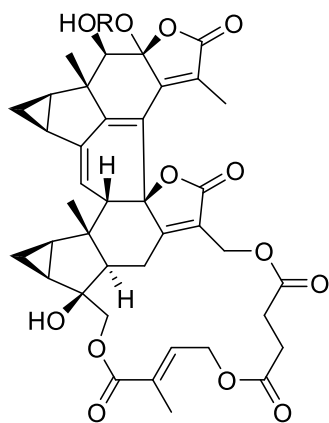
230 R = α -OH
234 R = β -OCH₃



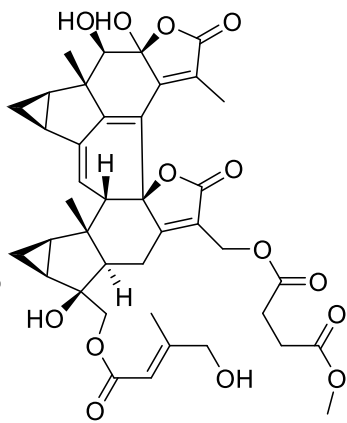
231 R₁ = α -OH R₂ = H
232 R₁ = β -OH R₂ = H
233 R₁ = β -OCH₃ R₂ = H
239 R₁ = α -OH R₂ = OH



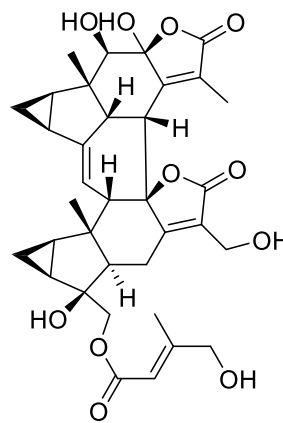
235 R₁ = H R₂ = H
236 R₁ = CH₃ R₂ = H
237 R₁ = C₂H₅ R₂ = H
238 R₁ = H R₂ = OAc



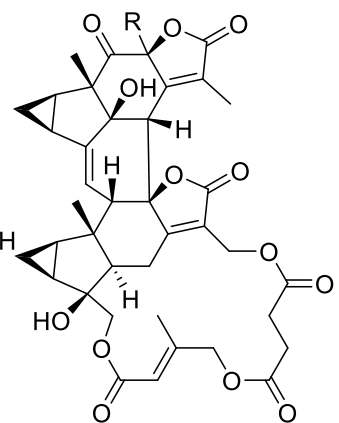
240 R = H
241 R = CH₃



242



243



244 R = H
245 R = OH

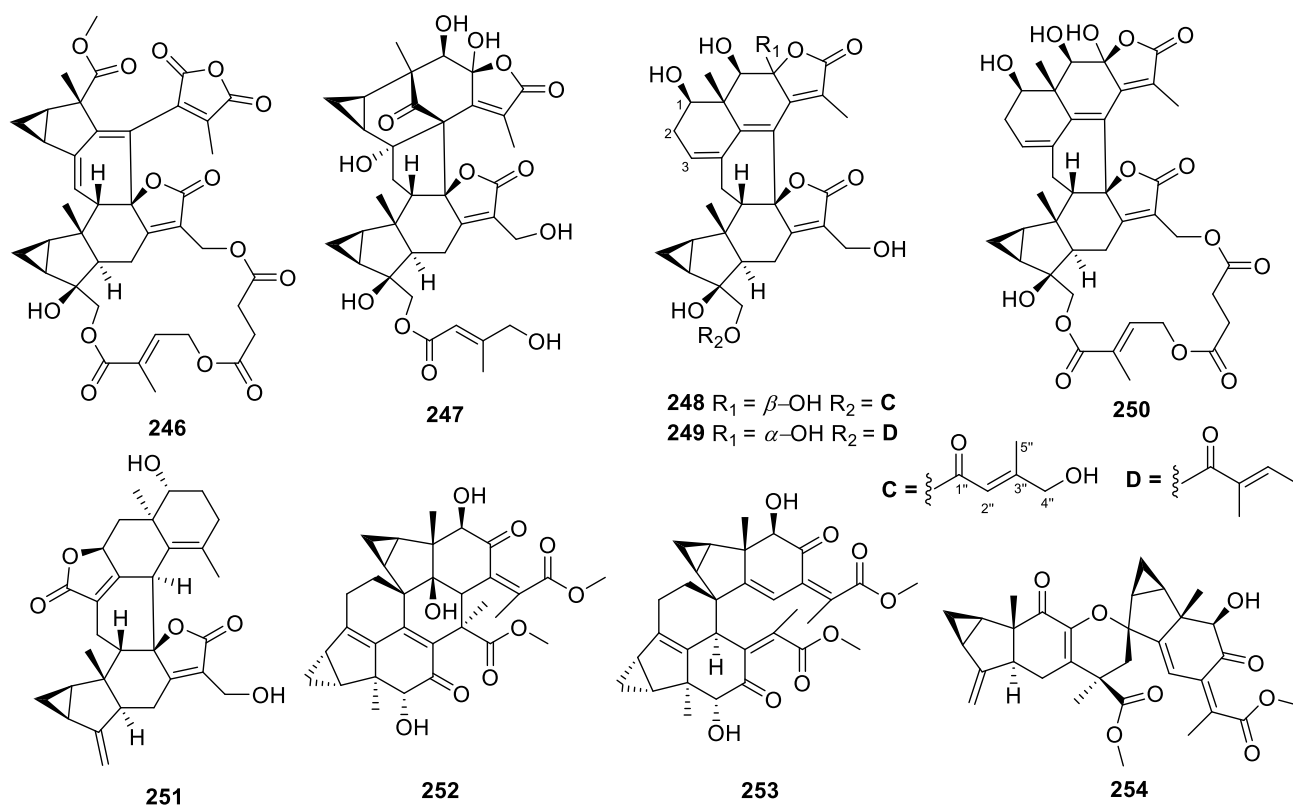
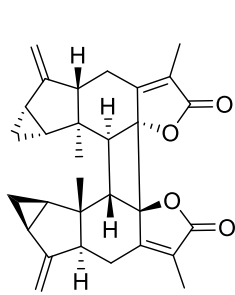
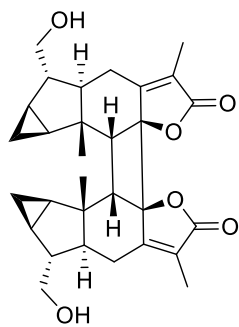


Table S3-6 Structure names, sources and bioactivity of *homo*-[4 + 2] cycloaddition type LS dimers (241-254).

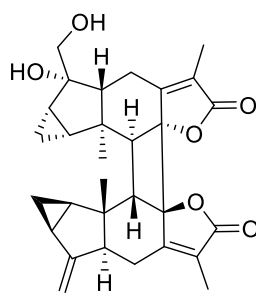
No	Compound	Sources	Bioactivity
241	Chloramultiol F	<i>C. multistachys</i> ⁷⁵	Cytotoxicity ⁷⁵
242	Fortunilide H	<i>C. fortunei</i> ³⁷	
243	Fortunilide J	<i>C. fortunei</i> ³⁷	
244	Chlorajaponilide A	<i>C. japonicus</i> ⁶⁰	
245	Chlorajaponilide B	<i>C. japonicus</i> ⁶⁰	
246	Chloramultiol G	<i>C. multistachys</i> ⁶⁰	
247	Fortunoid A	<i>C. fortunei</i> ³⁹	Antimalarial ³⁹
248	Fortunoid B	<i>C. fortunei</i> ³⁹	Antimalarial ³⁹
249	Chlorahupetol B	<i>C. henryi</i> var. <i>hupehensis</i> ⁷⁸	
250	Chlorahupetol A	<i>C. henryi</i> var. <i>hupehensis</i> ⁷⁸	
251	Horienoid B	<i>H. orientale</i> ⁹⁶	Anti-Inflammatory ⁹⁶
252	Chlotrichene A	<i>C. holostegius</i> ⁹⁷	
253	Chlotrichene B	<i>C. holostegius</i> ⁹⁷	Cytotoxicity ⁹⁷
254	Spirolindemer A	<i>C. henryi</i> ⁹⁸	Anti-Inflammatory ⁹⁸



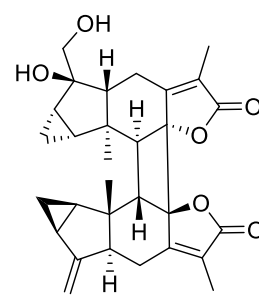
255



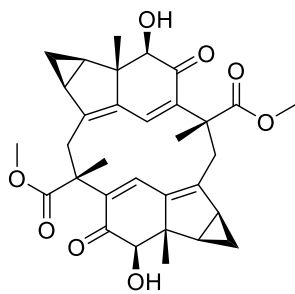
256



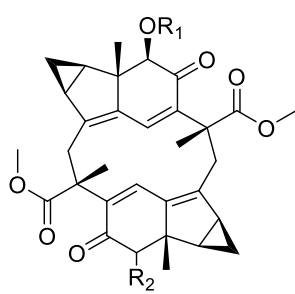
257



258

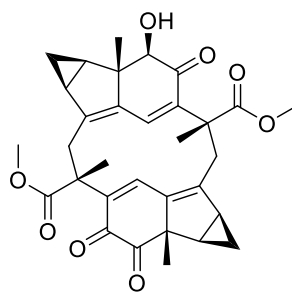


259

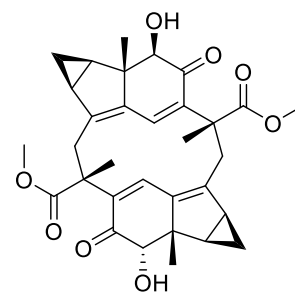


260 R₁ = Glu R₂ = β-OH

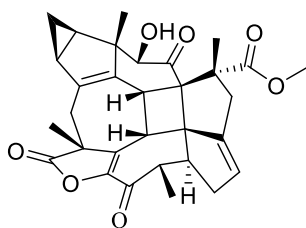
261 R₁ = H R₂ = H



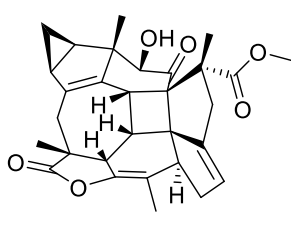
262



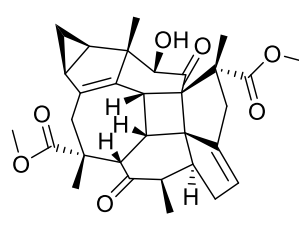
263



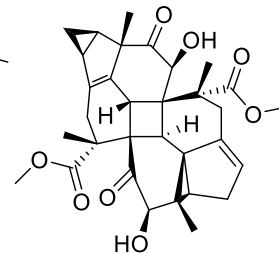
264



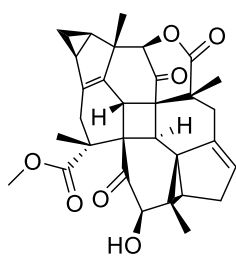
265



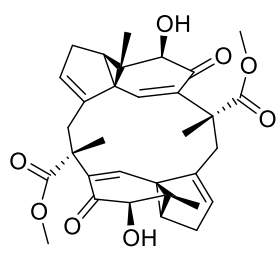
266



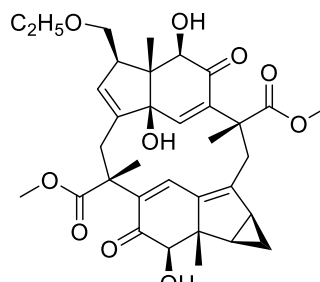
267



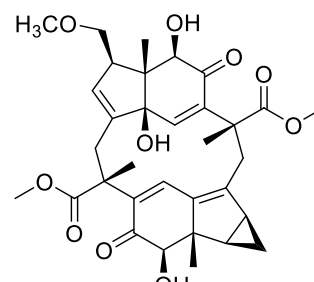
268



269



270



271

Table S4 Structure names, sources and bioactivity of *homo*-[2 + 2] and [6 + 6] cycloaddition type LS dimers (255-271).

No	Compound	Sources	Bioactivity
255	Chloranthalactone F	<i>C. japonicus</i> ⁶⁴	
256	Chololactone H	<i>C. holostegius</i> ⁶⁵	Anti-Inflammatory ⁶⁵
257	Sarglactone N	<i>S. glabra</i> ⁸⁹	
258	Sarglactone O	<i>S. glabra</i> ⁸⁹	
259	Cycloshizukaol A	<i>C. serratus</i> , ⁹⁹ <i>C. fortunei</i> , ⁴⁹ <i>C. multistachys</i> , ⁷⁵ <i>S. glabra</i> , ²⁸ <i>C.</i> <i>spicatus</i> , ⁷⁰ <i>C. japonicus</i> , ⁶⁰ <i>C.</i> <i>sessilifolius</i> ⁴⁰	
260	9- <i>O</i> - β - glucopyranosylcycloshizukaol A	<i>C. fortunei</i> , ⁴⁹ <i>C. japonicus</i> ⁷⁶	
261	Japonicone A	<i>C. japonicus</i> ¹⁰⁰	
262	Japonicone B	<i>C. japonicus</i> ¹⁰⁰	Cytotoxicity ¹⁰⁰
263	Japonicone C	<i>C. japonicus</i> ¹⁰⁰	
264	Chlorahupetone A	<i>C. henryi</i> var. <i>hupehensis</i> ⁸⁵	Cytotoxicity ⁸⁵
265	Chlorahupetone B	<i>C. henryi</i> var. <i>hupehensis</i> ⁸⁵	
266	Chlorahupetone C	<i>C. henryi</i> var. <i>hupehensis</i> ⁸⁵	
267	Chlorahupetone D	<i>C. henryi</i> var. <i>hupehensis</i> ⁸⁵	
268	Chlorahupetone E	<i>C. henryi</i> var. <i>hupehensis</i> ⁸⁵	
269	Chlorahupetone F	<i>C. henryi</i> var. <i>hupehensis</i> ⁸⁵	
270	Chlospicene A	<i>C. henryi</i> ¹⁰¹	Anti-nonalcoholic ¹⁰¹
271	Chlospicene B	<i>C. henryi</i> ¹⁰¹	Anti-nonalcoholic ¹⁰¹

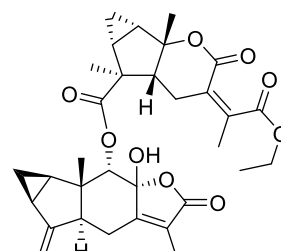
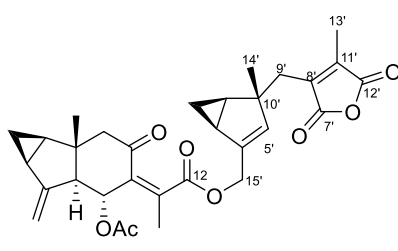
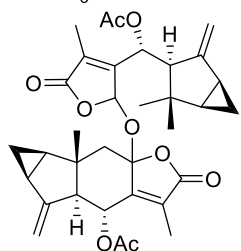
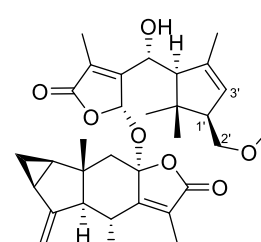
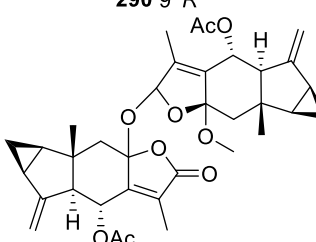
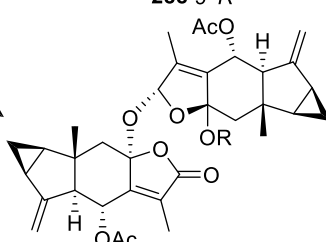
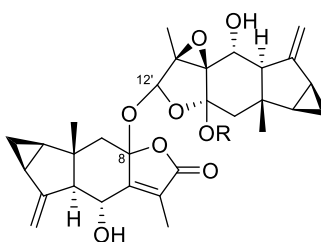
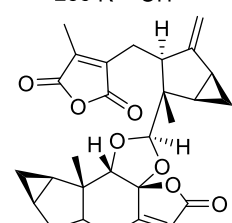
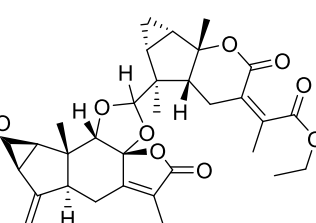
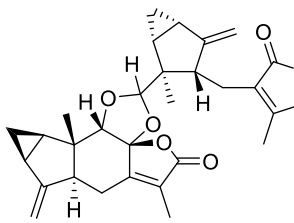
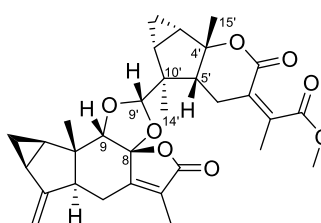
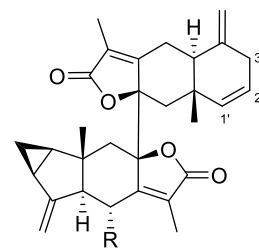
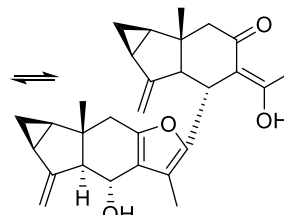
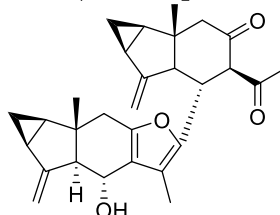
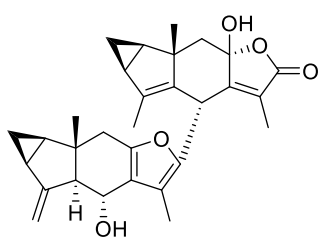
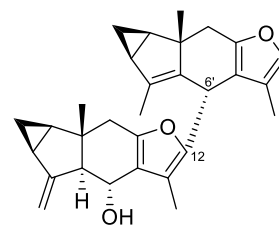
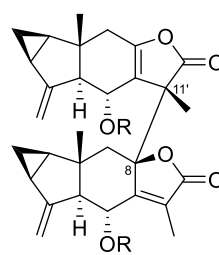
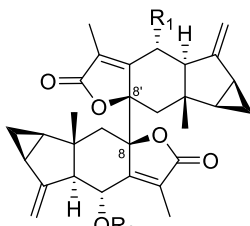
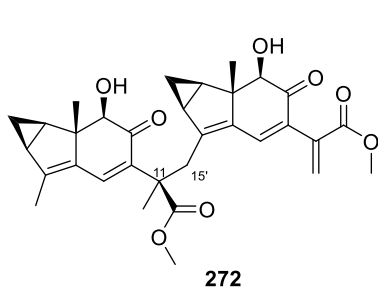


Table S5-1 Structure names, sources and bioactivity of *homo*-linear linkage type LS dimers (272-298).

No	Compound	Sources	Bioactivity
272	Shizukaol J	<i>C. japonicus</i> , ¹⁰² <i>C. fortunei</i> ⁵⁶³	
273	Lindenanolide F	<i>L. aggregata</i> , ¹⁰³ <i>L. chunii</i> ¹⁰⁴	
274	Linderanoid L	<i>L. aggregata</i> ¹⁰³	
275	Linderanoid M	<i>L. aggregata</i> ¹⁰³	
276	Linderanoid N	<i>L. aggregata</i> ¹⁰³	
277	Linderanoid O	<i>L. aggregata</i> ¹⁰³	
278	Linderanoid J	<i>L. aggregata</i> ¹⁰³	
279	Lindenanolide I	<i>L. chunii</i> ¹⁰⁴	
280	Linderanoid H	<i>L. aggregata</i> ¹⁰³	
281	Linderanoid I	<i>L. aggregata</i> ¹⁰³	
282	Linderanoid Ka	<i>L. aggregata</i> ¹⁰³	
283	Linderanoid Kb	<i>L. aggregata</i> ¹⁰³	
284	Linderanoid D	<i>L. aggregata</i> ¹⁰³	
285	Linderanoid E	<i>L. aggregata</i> ¹⁰³	
286	Chlojapolactone A	<i>C. japonicus</i> ⁸⁸	Anti-Inflammatory ⁸⁸
287	Sarglactone D	<i>S. glabra</i> ⁵⁸	Cytotoxicity ⁵⁸
288	Sarglactone E	<i>S. glabra</i> ⁵⁸	Cytotoxicity ⁵⁸
289	Sarglactone F	<i>S. glabra</i> ⁵⁸	Cytotoxicity ⁵⁸
290	Sarglactone G	<i>S. glabra</i> ⁵⁸	Cytotoxicity ⁵⁸
291	Chlojapolactone B	<i>C. japonicus</i> ¹⁰⁵	
292	Linderaggrenolide A	<i>L. aggregata</i> ¹⁰⁶	
293	Linderaggrenolide B	<i>L. aggregata</i> ¹⁰⁶	
294	Linderaggrenolide C	<i>L. aggregata</i> ¹⁰⁶	
295	Linderaggrenolide D	<i>L. aggregata</i> ¹⁰⁶	
296	Linderaggrenolide E	<i>L. aggregata</i> ¹⁰⁶	
297	Linderaggrenolide F	<i>L. aggregata</i> ¹⁰⁶	
298	Linderaggrenolide G	<i>L. aggregata</i> ¹⁰⁶	

Table S5-2 Structure names, sources and bioactivity of *homo*-linear linkage type LS dimers (**299-304**).

No	Compound	Sources	Bioactivity
299	Linderaggrenolide J	<i>L. aggregata</i> ¹⁰⁶	
300	Linderaggrenolide K	<i>L. aggregata</i> ¹⁰⁶	
301	Linderaggrenolide L	<i>L. aggregata</i> ¹⁰⁶	
302	Linderaggrenolide M	<i>L. aggregata</i> ¹⁰⁶	
303	Linderaggrenolide N	<i>L. aggregata</i> ¹⁰⁶	
304	Sarglactone H	<i>S. glabra</i> ⁵⁸	Cytotoxicity ⁵⁸

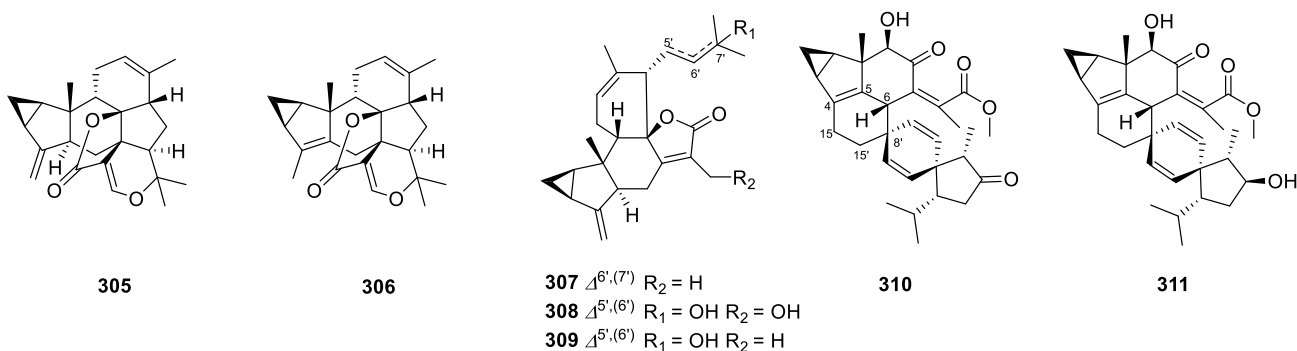


Table S6 Structure names, sources and bioactivity of *hetero*-[4 + 2] cycloaddition type LS dimers (**305-311**).

No	Compound	Sources	Bioactivity
305	Bolivianine	<i>H. angustifolium</i> ¹⁰⁷	
306	Isobolivianine	<i>H. angustifolium</i> ¹⁰⁷	
307	Sarcaglabrin A	<i>S. glabra</i> ⁷³	
308	Dyosmunoid A	<i>H. orientale</i> ¹⁰⁸	
309	7'-oxyisosarcaglabrin A	<i>S. glabra</i> ¹⁰⁹	
310	Chlorfortunone A	<i>C. fortunei</i> ¹¹⁰	TGF- β Inhibitory ¹¹⁰
311	Chlorfortunone B	<i>C. fortunei</i> ¹¹⁰	

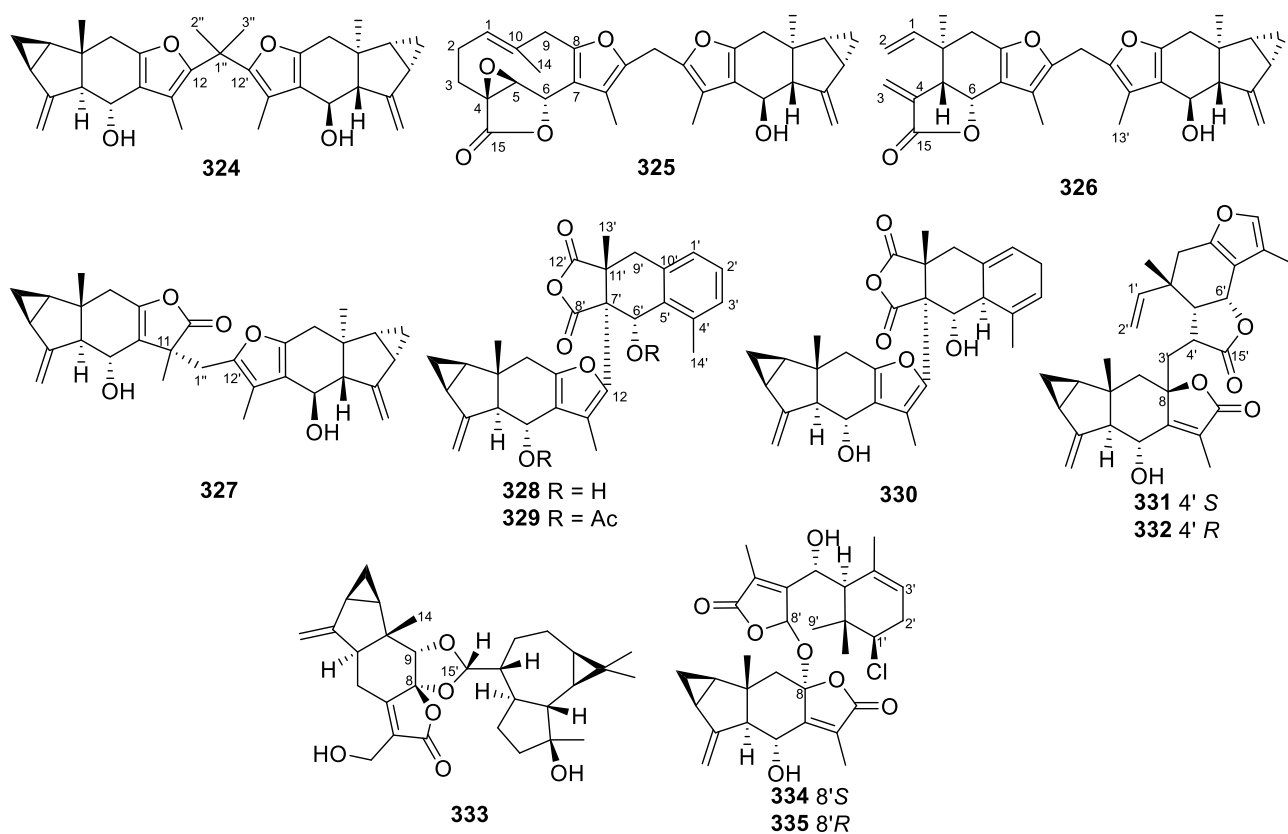


Table S8 Structure names, sources and bioactivity of *hetero-linear linkage type LS dimers (324-335)*.

No	Compound	Sources	Bioactivity
324	Aggreganoid C	<i>L. aggregata</i> ¹¹⁴	
325	Aggreganoid D	<i>L. aggregata</i> ¹¹⁴	
326	Aggreganoid E	<i>L. aggregata</i> ¹¹⁴	
327	Aggreganoid F	<i>L. aggregata</i> ¹¹⁴	
328	Linderanoid A	<i>L. aggregata</i> ¹⁰³	
329	Linderanoid B	<i>L. aggregata</i> ¹⁰³	
330	Linderanoid C	<i>L. aggregata</i> ¹⁰³	
331	Linderanoid F	<i>L. aggregata</i> ¹⁰³	TGF- β Inhibitory ¹⁰³
332	Linderanoid G	<i>L. aggregata</i> ¹⁰³	
333	Hedyorienoid A	<i>H. orientale</i> ¹¹⁵	
334	Linderaggrenolide H	<i>L. aggregata</i> ¹⁰⁶	
335	Linderaggrenolide I	<i>L. aggregata</i> ¹⁰⁶	

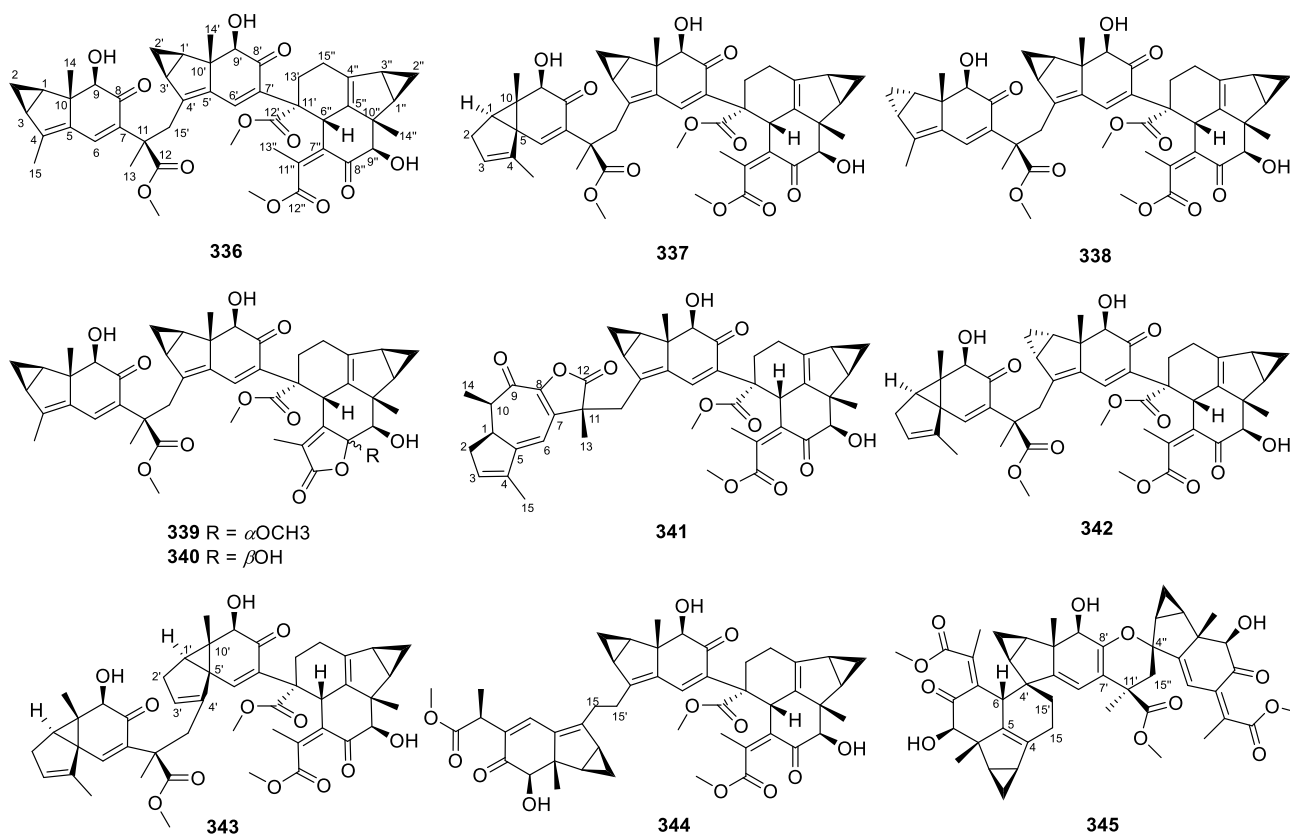


Table S9 Structure names, sources and bioactivity of *homo*-[4 + 2] cycloaddition type LS trimers (336-345).

No	Compound	Sources	Bioactivity
336	Trishizukaol A	<i>C. japonicus</i> , ¹⁰² <i>C. spicatus</i> , ¹¹⁶ <i>C. fortunei</i> ¹¹⁷	Antimalarial ¹¹⁶
337	Trichloranoid A	<i>C. spicatus</i> ¹¹⁶	Antimalarial ¹¹⁶ Anti-Inflammatory ¹¹⁷
338	Trichloranoid B	<i>C. spicatus</i> ¹¹⁶	Anti-Inflammatory ¹¹⁷
339	Trichloranoid C	<i>C. spicatus</i> ¹¹⁶	
340	Trichloranoid D	<i>C. spicatus</i> ¹¹⁶	Antimalarial ¹¹⁶
341	Chlofortunin B	<i>C. fortunei</i> ¹¹⁷	Anti-Inflammatory ¹¹⁷
342	Chlofortunin C	<i>C. fortunei</i> ¹¹⁷	Anti-Inflammatory ¹¹⁷
343	Chlofortunin D	<i>C. fortunei</i> ¹¹⁷	Anti-Inflammatory ¹¹⁷
344	Chlofortunin A	<i>C. fortunei</i> ¹¹⁷	
345	Spirolindemer B	<i>C. henryi</i> ⁹⁸	

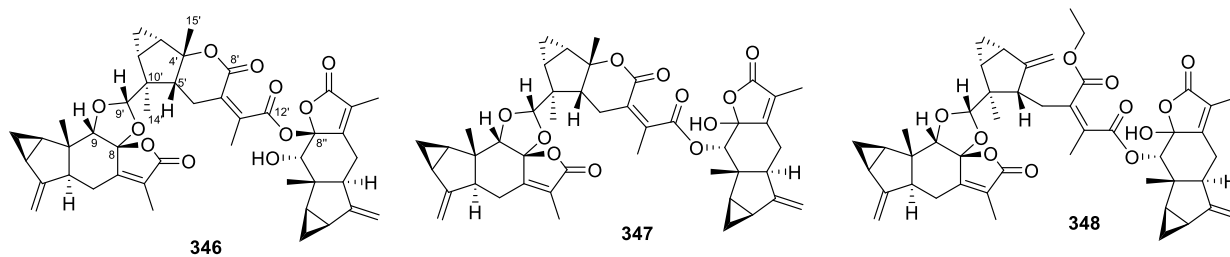


Table S10 Structure names, sources and bioactivity of *homo*-linear linkage type LS trimers (346-348).

No	Compound	Sources	Bioactivity
346	Sarglactone A	<i>S. glabra</i> ⁵⁸	Cytotoxicity ⁵⁸
347	Sarglactone B	<i>S. glabra</i> ⁵⁸	Cytotoxicity ⁵⁸
348	Sarglactone C	<i>S. glabra</i> ⁵⁸	Cytotoxicity ⁵⁸

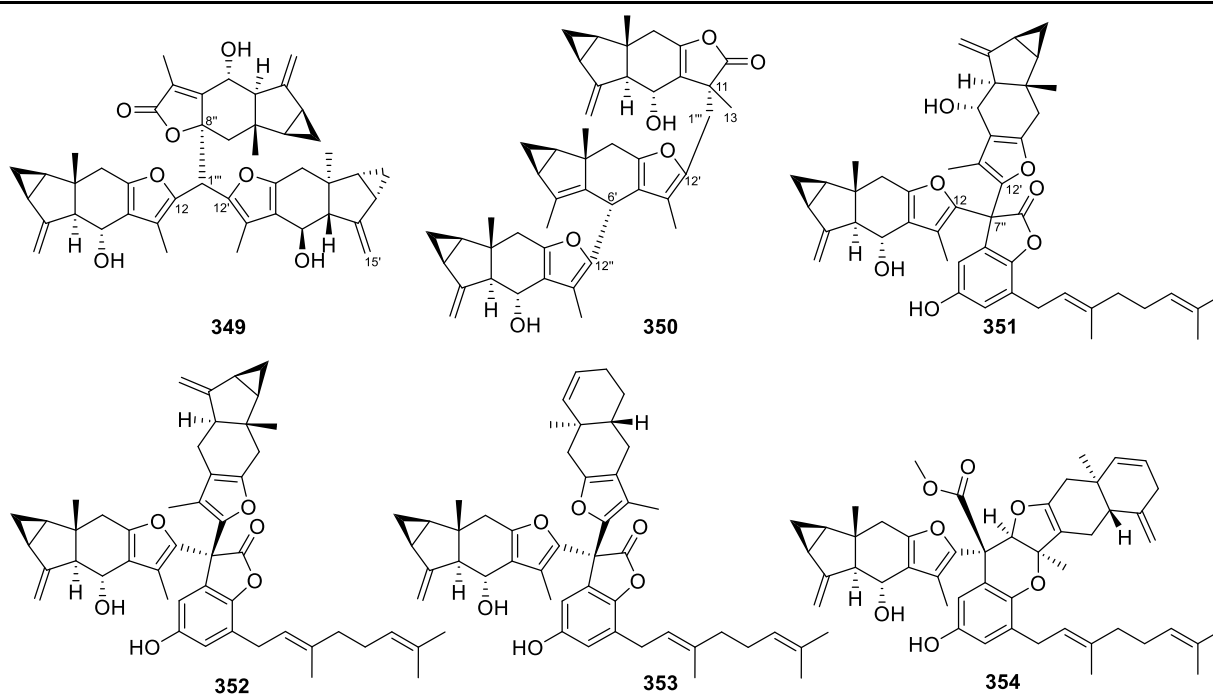


Table S11 Structure names, sources and bioactivity of *hetero*-LS trimers (349-354).

No	Compound	Sources	Bioactivity
349	Aggreganoid A	<i>L. aggregata</i> ¹¹⁴	TGF- β Inhibitory ¹¹⁴
350	Aggreganoid B	<i>L. aggregata</i> ¹¹⁴	
351	Linalide A	<i>L. aggregata</i> ¹¹⁸	
352	Linalide B	<i>L. aggregata</i> ¹¹⁸	
353	Linalide C	<i>L. aggregata</i> ¹¹⁸	
354	Linalide D	<i>L. aggregata</i> ¹¹⁸	Anti-Inflammatory ¹¹⁸

Reference

1. H. Kondo and T. Sanada, *Yakugaku Zasshi*, 1925, **526**, 1047–1057.
2. K. Takeda, M. Ikuta and M. Miyawaki, *Tetrahedron*, 1964, **20**, 2991–2997.
3. J. B. Li, Y. Ding and W. M. Li, *Chin. Chem. Lett.*, 2002, **13**, 965–967.
4. Q. Liu, J. H. Ahn, S. B. Kim, C. Lee, B. Y. Hwang and M. K. Lee, *Phytochemistry*, 2013, **87**, 112–118.
5. C. F. Zhang, N. Nakamura, S. Tewtrakul, M. Hattori, Q. S. Sun, Z. T. Wang and T. Fujiwara, *Chem. Pharm. Bull.*, 2002, **50**, 1195–1200.
6. S. S. Wen, Y. Wang, J. P. Xu, Q. Liu, L. Zhang, J. Zheng, L. Li, N. Zhang, X. Liu, Y. W. Xu and Z. L. Sun, *Nat. Prod. Res.*, 2022, **36**, 5407–5415.
7. H. J. Yang, E. B. Kwon and W. Li, *Nat. Prod. Res.*, 2022, **36**, 1914–1918.
8. H. Ishii, T. Tozyo, M. Nakamura and K. Takeda, *Tetrahedron*, 1968, **24**, 625–631.
9. K. Takeda, H. Ishii, T. Tozyo and H. Minato, *J. Chem. Soc. C*, 1969, 1920–1921.
10. Y. Cao, B. F. Xuan, B. Peng, C. Li, X. Y. Chai and P. F. Tu, *Phytochem. Rev.*, 2015, **15**, 869–906.
11. K. Takeda, H. Minato, I. Horibe and M. Miyawaki, *J. Chem. Soc., Perkin Trans. 1*, 1967, **7**, 631–634.
12. I. Kouno, A. Hirai, A. Fukushima, Z. H. Jiang and T. Tanaka, *J. Nat. Prod.*, 2001, **64**, 286–288.
13. Q. Liu, Y. H. Jo, S. B. Kim, Q. Jin, B. Y. Hwang and M. K. Lee, *Bioorg. Med. Chem. Lett.*, 2016, **26**, 4950–4954.
14. M. Zhang, J. S. Wang, P. R. Wang, M. Oyama, J. Luo, T. Ito, M. Iinuma and L. Y. Kong, *Fitoterapia*, 2012, **83**, 1604–1609.
15. X. H. Li, H. Y. Li, W. Ni, X. J. Qin, Q. Zhao, Z. Q. Ji and H. Y. Liu, *Phytochem. Lett.*, 2016, **15**, 199–203.
16. Q. H. Cao, W. F. Dai, B. C. Li and M. Zhang, *Phytochem. Lett.*, 2019, **30**, 6–9.
17. J. Kawabata, S. Tahara, J. Mizutani, A. Furusaki, N. Hashiba and T. Matsumoto, *Agric. Biol. Chem.*, 1979, **43**, 885–887.
18. H. Yan, X. H. Li, X. F. Zheng, C. L. Sun and H. Y. Liu, *Helv. Chim. Acta.*, 2013, **96**, 1386–1391.
19. L. F. Maia, R. d. A. Epifanio, T. Eve and W. Fenical, *J. Nat. Prod.*, 1999, **62**, 1322–1324.
20. J. E. Heo, G. L. Jin, Y. Y. Lee and H. S. Y. Choi, *Nat. Prod. Sci.*, 2005, **11**, 41–44.
21. X. R. Hu, J. S. Yang and X. D. Xu, *Chem. Pharm. Bull.*, 2009, **57**, 418–420.
22. Q. H. Wang, H. X. Kuang, B. Y. Yang, Y. G. Xia, J. S. Wang and L. Y. Kong, *J. Nat. Prod.*, 2011, **74**, 16–20.
23. Y. Li, D. M. Zhang, J. B. Li, S. S. Yu, Y. Li and Y. M. Luo, *J. Nat. Prod.*, 2006, **69**, 616–620.
24. C. S. Jiang, Y. Q. Guo, S. Yin, H. Zhang and G. H. Tang, *J. Asian. Nat. Prod. Res.*, 2019, **21**, 377–383.
25. M. Uchida, Y. Koike, G. Kusano, Y. Kondo, S. Nozoe, C. Kabuto and T. Takemoto, *Chem. Pharm. Bull.*, 1980, **28**, 92–102.
26. Y. Takeda, H. Yamashita, T. Matsumoto and H. Terao, *Phytochemistry*, 1993, **33**, 713–715.
27. L. P. Zhu, Y. Li, J. Z. Yang, L. Zuo and D. M. Zhang, *J. Asian. Nat. Prod. Res.*, 2008, **10**, 541–545.
28. X. F. He, S. Yin, Y. C. Ji, Z. S. Su, M. Y. Geng and J. M. Yue, *J. Nat. Prod.*, 2010, **73**, 45–50.
29. G. Ni, H. Zhang, H. C. Liu, S. P. Yang, M. Y. Geng and J. M. Yue, *Tetrahedron*, 2013, **69**, 564–569.
30. S. Y. Kao, J. H. Su, T. L. Hwang, J. H. Sheu, Z. H. Wen, Y. C. Wu and P. J. Sung, *Mar. Drugs*, 2011, **9**, 1534–1542.
31. M. Bittner, J. Jakupovic, F. Bohlmann and M. Silva, *Phytochemistry*, 1989, **28**, 271–273.
32. G. X. Chou, N. Norio, C. M. Ma, Z. T. Wang, H. Masao, L. S. Xu and G. J. Xu, *Zhongguo Yaoke Daxue Xuebao*, 2000, **31**, 339.
33. J. H. Kim, J. S. Jeon, J. H. Kim, E. J. Jung, Y. J. Lee, E. M. Gao, A. S. Syed, R. H. Son and C. Y. Kim, *Molecules*, 2021, **26**, 5269.
34. L. R. Fernandez, E. Butassi, L. Svetaz, S. A. Zacchino, J. A. Palermo and M. Sanchez, *J. Nat. Prod.*, 2014, **77**, 1579–1585.

35. F. Bohlmann, G. W. Ludwig, J. Jakupovic, R. M. King and H. Robinson, *Chemischer Informationsdienst*, 1984, **15**, 304.
36. Y. S. Shi, Y. B. Liu, S. G. Ma, Y. Li, J. Qu, L. Li, S. P. Yuan, Q. Hou, Y. H. Li, J. D. Jiang and S. S. Yu, *J. Nat. Prod.*, 2015, **78**, 1526–1535.
37. B. Zhou, Y. Wu, S. Dalal, E. F. Merino, Q. F. Liu, C. H. Xu, T. Yuan, J. Ding, D. G. Kingston, M. B. Cassera and J. M. Yue, *J. Nat. Prod.*, 2017, **80**, 96–107.
38. T. O. Do, T. K. Pham, T. B. H. Nguyen, H. Y. Pham, H. H. Tran, X. C. Nguyen, V. L. Dang, V. M. Chau and V. K. Phan, *Nat. Prod. Commun.*, 2010, **5**, 1717–1720.
39. B. Zhou, Q. F. Liu, S. Dalal, M. B. Cassera and J. M. Yue, *Org. Lett.*, 2017, **19**, 734–737.
40. L. J. Wang, J. Xiong, C. Lau, L. L. Pan and J. F. Hu, *J. Asian. Nat. Prod. Res.*, 2015, **17**, 1220–1230.
41. H. X. Kuang, Y. G. Xia, B. Y. Yang, Q. H. Wang and S. W. Lu, *Chem. Biodivers.*, 2008, **5**, 1736–1742.
42. T. H. Duong, M. A. Beniddir, N. T. Trung, C. D. Phan, V. G. Vo, V. K. Nguyen, Q. L. Le, H. D. Nguyen and P. L. Pogam, *Molecules*, 2020, **25**, 1830.
43. W. Y. Tsui, *Phytochemistry*, 1996, **43**, 819–821.
44. M. Uchida, G. Kusano, Y. Kondo and S. Nozoe, *Heterocycles*, 1978, **9**, 139–144.
45. B. Wu, J. Chen, H. B. Qu and Y. Y. Cheng, *J. Nat. Prod.*, 2008, **71**, 877–880.
46. L. Acebey, V. Jullian, D. Sereno, S. Chevalley, Y. Estevez, C. Moulis, S. Beck, A. Valentin, A. Gimenez and M. Sauvain, *Planta Med.*, 2010, **76**, 365–368.
47. J. Kawabata and J. Mizutani, *Agric. Biol. Chem.*, 1989, **53**, 203–207.
48. S. Tahara, Y. Fukushi, J. Kawabata and J. Mizutani, *Agric. Biol. Chem.*, 2014, **45**, 1511–1512.
49. X. C. Wang, L. L. Wang, X. W. Ouyang, S. P. Ma, J. H. Liu and L. H. Hu, *Helv. Chim. Acta.*, 2009, **92**, 313–320.
50. H. Okamura, N. Nakashima, T. Iwagawa, N. Nakayama and M. Nakatani, *Chem. Lett.*, 1994, 1541–1542.
51. P. C. Kuo, Y. H. Wu, H. Y. Hung, S. H. Lam, G. H. Ma, L. M. Kuo, T. L. Hwang, D. H. Kuo and T. S. Wu, *Bioorg. Med. Chem. Lett.*, 2020, **30**, 127224.
52. M. Zhang, J. S. Wang, M. Oyama, J. Luo, C. Guo, T. Ito, M. Inuma and L. Y. Kong, *J. Asian. Nat. Prod. Res.*, 2012, **14**, 708–712.
53. S. Yaermainaiti, P. Wang, J. Luo, R. J. Li and L. Y. Kong, *Fitoterapia*, 2016, **111**, 7–11.
54. S. Y. Kao, J. H. Su, T. L. Hwang, J. H. Sheu, Y. D. Su, C. S. Lin, Y. C. Chang, W. H. Wang, L. S. Fang and P. J. Sung, *Tetrahedron*, 2011, **67**, 7311–7315.
55. L. S. Gan, Y. L. Zheng, J. X. Mo, X. Liu, X. H. Li and C. X. Zhou, *J. Nat. Prod.*, 2009, **72**, 1497–1501.
56. J. L. Yang and Y. P. Shi, *Planta. Med.*, 2012, **78**, 59–64.
57. T. T. Huong, N. V. Thong, T. T. Minh, L. H. Tram, N. T. Anh, H. D. Cuong, P. V. Cuong and D. V. Ca, *Lett. Org. Chem.*, 2014, **11**, 639–642.
58. J. Chi, S. S. Wei, H. L. Gao, D. Xu, L. Zhang, L. Yang, W. J. Xu, J. Luo and L. Y. Kong, *J. Org. Chem.*, 2019, **84**, 9117–9126.
59. Y. Q. Guo, J. J. Zhao, Z. Z. Li, G. H. Tang, Z. M. Zhao and S. Yin, *Bioorg. Med. Chem. Lett.*, 2016, **26**, 3163–3166.
60. P. L. Fang, Y. L. Cao, H. Yan, L. L. Pan, S. C. Liu, N. B. Gong, Y. Lu, C. X. Chen, H. M. Zhong, Y. Guo and H. Y. Liu, *J. Nat. Prod.*, 2011, **74**, 1408–1413.
61. J. Kawabata, Y. Fukushi, S. Tahara and J. Mizutani, *Phytochemistry*, 1990, **29**, 2332–2334.
62. J. Kawabata and J. Mizutani, *Phytochemistry*, 1992, **31**, 1293–1296.
63. X. C. Wang, Y. N. Zhang, L. L. Wang, S. P. Ma, J. H. Liu and L. H. Hu, *J. Nat. Prod.*, 2008, **71**, 674–677.
64. G. Kusano, M. Abe, Y. Koike, M. Uchida, S. Nozoe and Z. Taira, *Yakugaku Zasshi*, 1991, **111**, 756–764.
65. C. P. Shen, J. G. Luo, M. H. Yang and L. Y. Kong, *Phytochemistry*, 2017, **137**, 117–122.

66. P. Wang, J. Luo, Y. M. Zhang and L. Y. Kong, *Tetrahedron*, 2015, **71**, 5362–5370.
67. J. Kawabata, E. Fukushi and J. Mizutani, *Phytochemistry*, 1995, **39**, 121–125.
68. S. P. Yang, Z. B. Gao, Y. Wu, G. Y. Hu and J. M. Yue, *Tetrahedron*, 2008, **64**, 2027–2034.
69. C. J. Li, D. M. Zhang, Y. M. Luo, S. S. Yu, Y. Li and Y. Lu, *Phytochemistry*, 2008, **69**, 2867–2874.
70. S. Y. Kim, Y. Kashiwada, K. Kawazoe, K. Murakami, H. D. Sun, S. L. Li and Y. Takaishi, *Chem. Pharm. Bull.*, 2011, **59**, 1281–1284.
71. B. Bai, S. X. Ye, D. P. Yang, L. P. Zhu, G. H. Tang, Y. Y. Chen, G. Q. Li and Z. M. Zhao, *J. Nat. Prod.*, 2019, **82**, 407–411.
72. S. Zhang, S. P. Yang, T. Yuan, B. D. Lin, Y. Wu and J. M. Yue, *Tetrahedron Lett.*, 2010, **51**, 764–766.
73. X. R. Yang, N. Tanaka, D. Tsuji, F. L. Lu, X. J. Yan, K. Itoh, D. P. Li and Y. Kashiwada, *Tetrahedron Lett.*, 2020, **61**, 151916.
74. W. M. Huang, Y. T. Bian, F. Y. Chen, T. J. Ning, Z. Y. Zhu, Z. C. Chen and Y. M. Luo, *Phytochemistry*, 2022, **193**, 113001.
75. X. H. Ran, F. Teng, C. X. Chen, G. Wei, X. J. Hao and H. Y. Liu, *J. Nat. Prod.*, 2010, **73**, 972–975.
76. Z. G. Zhuo, G. Z. Wu, X. Fang, X. H. Tian, H. Y. Dong, X. K. Xu, H. L. Li, N. Xie, W. D. Zhang and Y. H. Shen, *Fitoterapia*, 2017, **119**, 90–99.
77. S. Y. Wang, Y. Q. Li, X. L. Wang, X. Q. Zhang, F. Xu, P. Ying, L. Y. Kong and J. Luo, *Fitoterapia*, 2023, **168**, 105547.
78. D. Y. Zhang, Y. R. Hu, H. J. Yang, Y. N. Wang, S. Liu, Y. H. Zou, J. J. Zhou, P. Y. Zhuang, X. X. Wang and H. Liu, *Chin. J. Chem.*, 2023, **41**, 1209–1225.
79. L. G. Xiao, P. Li, H. Yan, W. Ni, L. He and H. Y. Liu, *Org. Biomol. Chem.*, 2022, **20**, 1320–1326.
80. M. Zhang, M. Iinuma, J. S. Wang, M. Oyama, T. Ito and L. Y. Kong, *J. Nat. Prod.*, 2012, **75**, 694–698.
81. X. W. Shi, Q. Q. Lu, G. Pescitelli, T. Ivsic, J. H. Zhou and J. M. Gao, *Chirality*, 2016, **28**, 158–163.
82. B. Wu, H. B. Qu and Y. Y. Cheng, *Helv. Chim. Acta.*, 2008, **91**, 725–733.
83. H. Yan, M. Y. Ba, X. H. Li, J. M. Guo, X. J. Qin, L. He, Z. Q. Zhang, Y. Guo and H. Y. Liu, *Fitoterapia*, 2016, **115**, 64–68.
84. X. F. He, S. Zhang, R. X. Zhu, S. P. Yang, T. Yuan and J. M. Yue, *Tetrahedron*, 2011, **67**, 3170–3174.
85. D. Y. Zhang, X. X. Wang, Y. N. Wang, M. Wang, P. Y. Zhuang, Y. Jin and H. Liu, *Org. Chem. Front.*, 2021, **8**, 4374–4386.
86. Y. P. Sun, J. Chi, L. Zhang, S. Y. Wang, Z. H. Chen, H. Zhang, L. Y. Kong and J. Luo, *J. Org. Chem.*, 2022, **87**, 4323–4332.
87. B. Zhou, F. M. Zimbres, J. H. Butler, C. H. Xu, R. S. Haney, Y. Wu, M. B. Cassera and J. M. Yue, *Sci. China: Chem.*, 2021, **65**, 82–86.
88. Y. Q. Guo, G. H. Tang, Z. Z. Li, S. L. Lin and S. Yin, *RSC Adv.*, 2015, **5**, 103047–103051.
89. Y. Y. Wang, Z. H. Chen, Q. R. Li, L. T. Cui, J. Chi, J. X. Li, Y. P. Sun, L. Y. Kong and J. Luo, *Tetrahedron Lett.*, 2022, **98**, 153834.
90. S. P. Yang, Z. B. Gao, F. D. Wang, S. G. Liao, H. D. Chen, C. R. Zhang, G. Y. Hu and J. M. Yue, *Org. Lett.*, 2007, **9**, 903–906.
91. S. Y. Kim, Y. Kashiwada, K. Kawazoe, K. Murakami, H. D. Sun, S. L. Li and Y. Takaishi, *Tetrahedron Lett.*, 2009, **50**, 6032–6035.
92. S. P. Yang and J. M. Yue, *Tetrahedron Lett.*, 2006, **47**, 1129–1132.
93. S. Y. Kim, Y. Kashiwada, K. Kawazoe, K. Murakami, H. D. Sun, S. L. Li and Y. Takaishi, *Phytochem. Lett.*, 2009, **2**, 110–113.
94. Z. G. Zhuo, Z. Y. Cheng, J. Ye, H. L. Li, X. K. Xu, N. Xie, W. D. Zhang and Y. H. Shen, *Phytochem. Lett.*, 2017, **20**, 133–138.

95. Y. J. Xu, C. P. Tang, C. Q. Ke, J. B. Zhang, H. C. Weiss, E. R. Gesing and Y. Ye, *J. Nat. Prod.*, 2007, **70**, 1987–1990.
96. Y. Y. Fan, L. S. Gan, S. X. Chen, Q. Gong, H. Y. Zhang and J. M. Yue, *J. Org. Chem.*, 2021, **86**, 11277–11283.
97. J. Chi, W. J. Xu, S. S. Wei, X. B. Wang, J. X. Li, H. L. Gao, L. Y. Kong and J. Luo, *Org. Lett.*, 2019, **21**, 789–792.
98. J. X. Li, J. Chi, P. F. Tang, Y. P. Sun, W. J. Lu, W. J. Xu, Y. Y. Wang, J. Luo and L. Y. Kong, *Chin. J. Chem.*, 2022, **40**, 603–608.
99. J. Kawabata, E. Fukushi and J. Mizutani, *Phytochemistry*, 1993, **32**, 1347–1349.
100. H. Yan, X. J. Qin, X. H. Li, Q. Yu, W. Ni, L. He and H. Y. Liu, *Tetrahedron Lett.*, 2019, **60**, 713–717.
101. J. X. Li, Z. R. Cui, Y. Y. Li, C. H. Han, Y. Q. Zhang, P. F. Tang, L. T. Cui, H. Zhang, J. Luo and L. Y. Kong, *Chin. Chem. Lett.*, 2022, **33**, 4257–4260.
102. J. Kawabata, E. Fukushi and J. Mizutani, *Phytochemistry*, 1998, **47**, 231–235.
103. X. Liu, J. Fu, R. S. Shen, X. J. Wu, J. Yang, L. P. Bai, Z. H. Jiang and G. Y. Zhu, *Phytochemistry*, 2021, **191**, 112924.
104. Y. X. Li, S. S. Wen, H. Yang, Y. Wang, Y. Wu and Z. L. Sun, *Chem. Nat. Compd.*, 2019, **55**, 1069–1072.
105. Q. Y. Li, Y. Wang, S. S. Wen, Y. Wu, L. H. Xu and Z. L. Sun, *Rec. Nat. Prod.*, 2019, **13**, 483–490.
106. X. Liu, J. Fu, J. Yang, A. C. Huang, R. F. Li, L. P. Bai, L. Liu, Z. H. Jiang and G. Y. Zhu, *ACS. Omega.*, 2021, **6**, 5898–5909.
107. L. Acebey, M. Sauvain, S. Beck, C. Moulis, A. Gimenez and V. Jullian, *Org. Lett.*, 2007, **9**, 4693–4696.
108. Y. Y. Fan, C. Y. Zheng, B. Zhou, F. M. Zimbres, M. B. Cassera and J. M. Yue, *Chin. J. Chem.*, 2022, **41**, 392–398.
109. Y. P. Sun, Y. Y. Wang, Y. Y. Li, S. Y. Wang, D. Y. Zhang, L. Y. Kong and J. Luo, *Org. Biomol. Chem.*, 2022, **20**, 9222–9227.
110. X. J. Wu, D. Cao, F. L. Chen, R. S. Shen, J. Gao, L. P. Bai, W. Zhang, Z. H. Jiang and G. Y. Zhu, *ACS. Omega.*, 2022, **7**, 35063–35068.
111. P. Wang, R. J. Li, R. H. Liu, K. L. Jian, M. H. Yang, L. Yang, L. Y. Kong and J. Luo, *Org. Lett.*, 2016, **18**, 832–835.
112. Y. Y. Wang, Z. R. Cui, J. Chi, P. F. Tang, M. H. Zhang, J. X. Li, Y. Y. Li, H. Zhang, J. Luo and L. Y. Kong, *Chin. J. Chem.*, 2020, **39**, 129–136.
113. Z. R. Cui, Y. Y. Wang, J. X. Li, J. Chi, P. P. Zhang, L. Y. Kong and J. Luo, *Org. Lett.*, 2022, **24**, 9107–9111.
114. X. Liu, J. Yang, J. Fu, X. J. Yao, J. R. Wang, L. Liu, Z. H. Jiang and G. Y. Zhu, *Org. Lett.*, 2019, **21**, 5753–5756.
115. Y. Y. Fan, Y. L. Sun, B. Zhou, J. X. Zhao, L. Sheng, J. Y. Li and J. M. Yue, *Org. Lett.*, 2018, **20**, 5435–5438.
116. J. S. Zhou, Q. F. Liu, F. M. Zimbres, J. H. Butler, M. B. Cassera, B. Zhou and J. M. Yue, *Org. Chem. Front.*, 2021, **8**, 1795–1801.
117. S. Y. Wang, Y. P. Sun, Y. Q. Li, W. J. Xu, Q. Q. Li, Y. B. Mu, L. Y. Kong and J. Luo, *J. Org. Chem.*, 2023, **88**, 347–354.
118. X. Liu, J. Yang, X. J. Yao, X. Yang, J. Fu, L. P. Bai, L. Liu, Z. H. Jiang and G. Y. Zhu, *J. Org. Chem.*, 2019, **84**, 8242–8247.