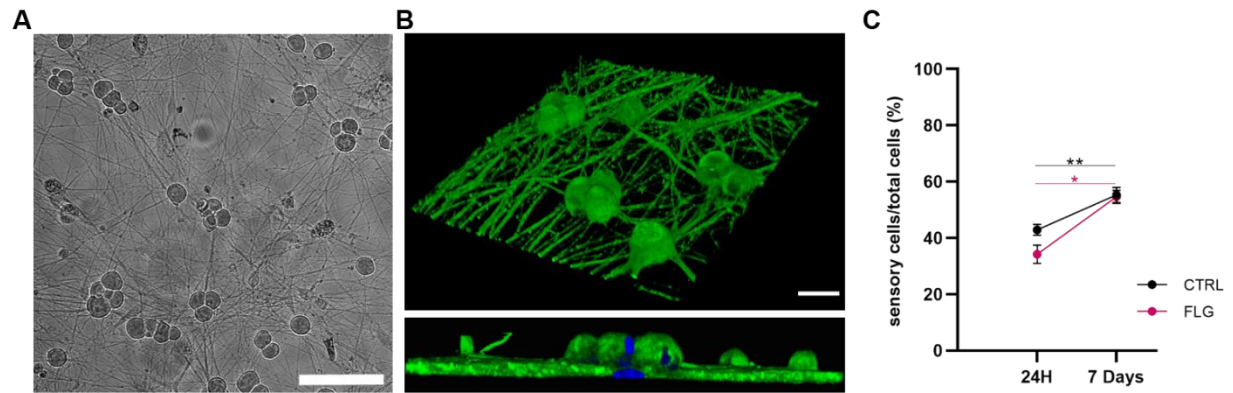


Supporting Information for

Few-layered graphene increases the response of nociceptive neurons to irritant stimuli

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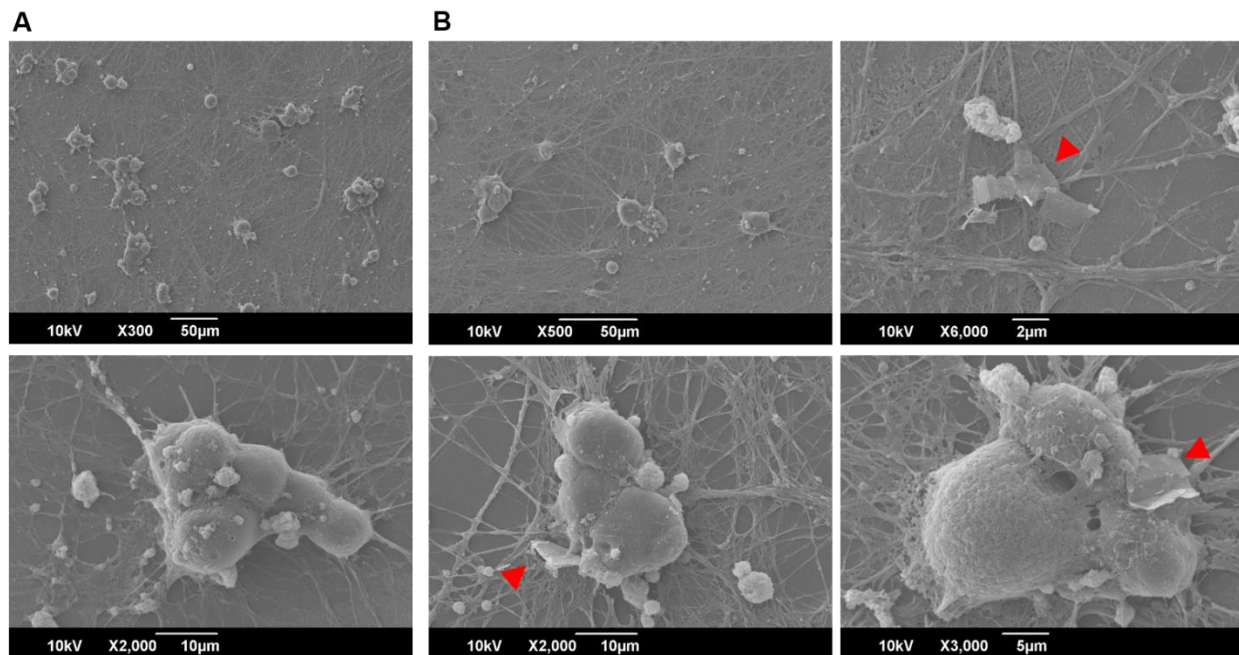


Figure S2. Morphology of DRG sensory neurons exposed to FLG. Representative SEM micrographs of DRG neuron cultures at different magnification. **A)** Untreated control neurons and **B)** neurons treated with FLG for 24 h. Extracellular GRM flakes are highlighted with red arrows.

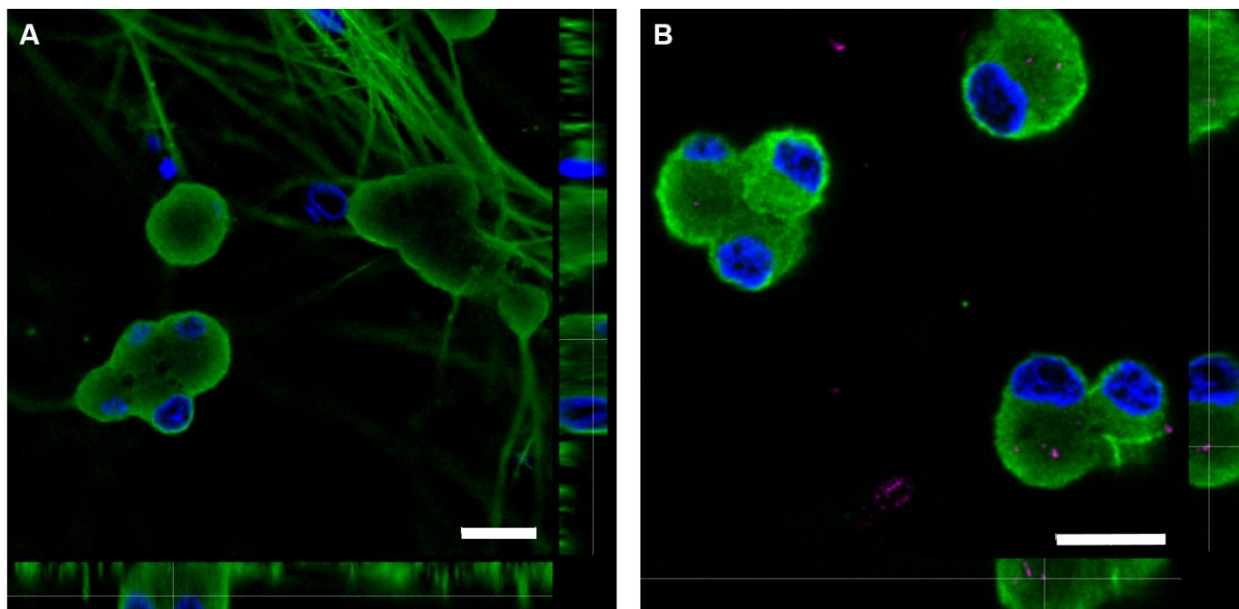


Figure S3. FLG internalization visualized through reflective light at the confocal microscope. A) CTRL and **B)** FLG. β 3-tubulin III (green); DAPI (blue); Graphene (Pink). Pictures are taken at one focal plane of a z-stack at 63x magnification. Since the sensory cell bodies pop out from the planar plane, not all axons are visible in this picture (see Figure S1). Scale bars, 20 μ m.

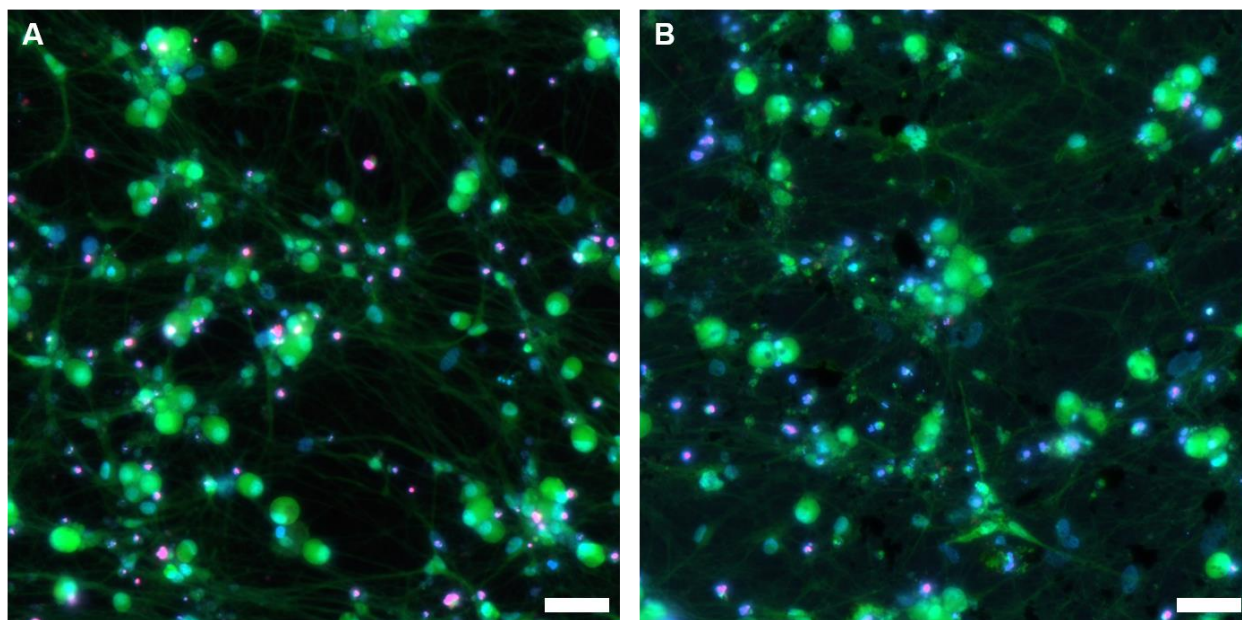


Figure S4. Viability of DRG sensory neurons upon FLG exposure. Calcein-AM (green) and PI (red) staining of DRG neuron cultures at DIV4, exposed for 24 h to $100 \mu\text{g mL}^{-1}$ of colloidal FLG. Nuclei are visualized by Hoechst staining (blue). **A)** CTRL and **B)** FLG-treated samples at 10x magnification. Scale bars, 50 μm .

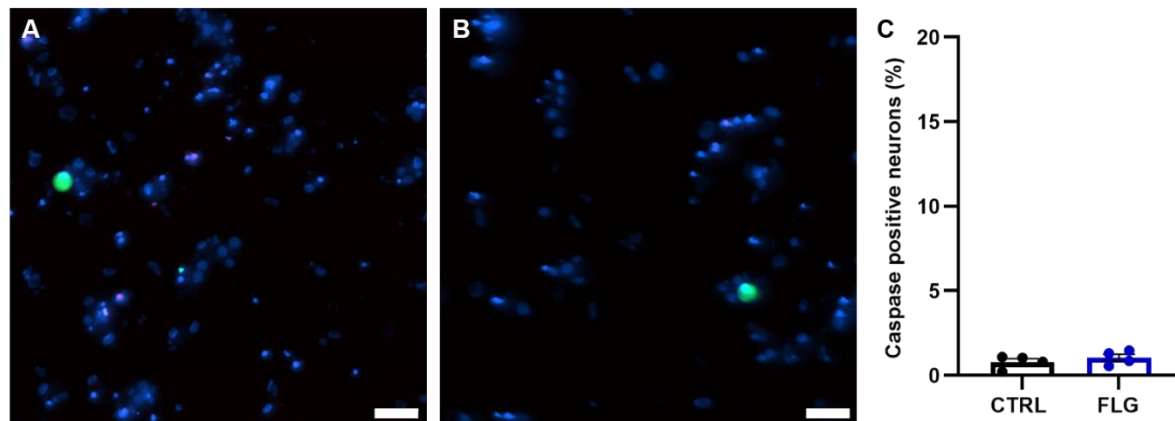


Figure S5. Evaluation of apoptosis cascade activation upon FLG exposure. Caspase 3/7 staining of DRG neuron cultures at DIV4, exposed for 24 h to $100 \mu\text{g mL}^{-1}$ of colloidal FLG. **A)** CTRL, **B)** FLG and **C)** Percentage of sensory neurons expressing active Caspase over total number of sensory neurons. All data are expressed as means \pm SEM; $N=3$ (4 replicates). Statistical analysis was performed using the Student's *t*-test. Caspase (green), DAPI (blue) and PI (red) at 10x magnification. Scale bars, 50 μm .

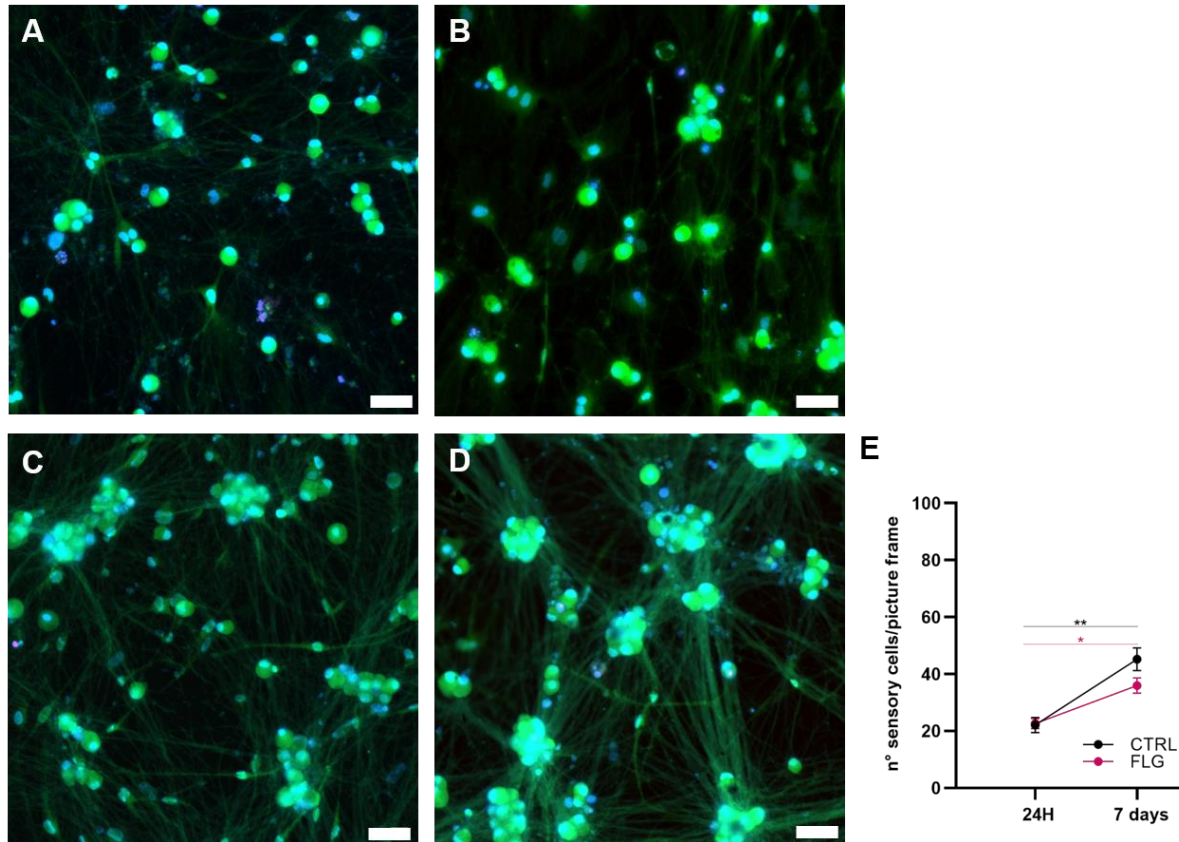


Figure S6. Sensory neuron clustering over time. DRG neurons live stained with Calcein AM (green), DAPI (blue) and PI (red) at 10x magnification from **A)** CTRL and **B)** FLG 10 $\mu\text{g mL}^{-1}$ at DIV5 (24 h treatment); **C)** CTRL and **D)** FLG at DIV11 (7-day treatment). **E)** Number of sensory neurons per picture frame. All data are expressed as means \pm SEM; $N=3$ (3/4 replicates). ** $p=0.0091$; * $p=0.0149$, unpaired Student's t -test. Scale bars, 50 μm .

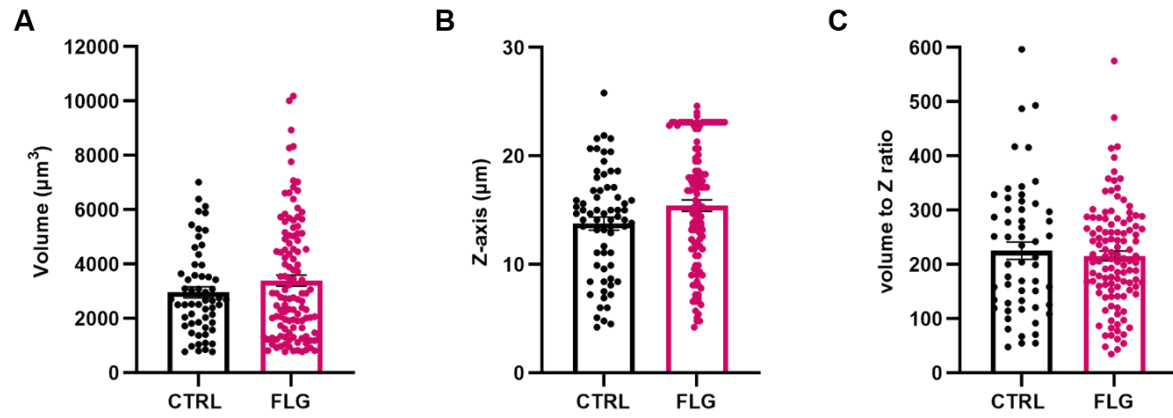


Figure S7. 3D morphological analysis. **A)** Volume of neuron soma, **B)** length of Z-axis and **C)** soma volume/soma height for DRG neuron cultures at DIV4, exposed for 24 h to $10 \mu\text{g mL}^{-1}$ of FLG. Each point represents a single neuron. All data are expressed as means \pm SEM, $N=3$ (6 different cultures). Unpaired Student's *t*-test/Mann-Whitney *U*-test.

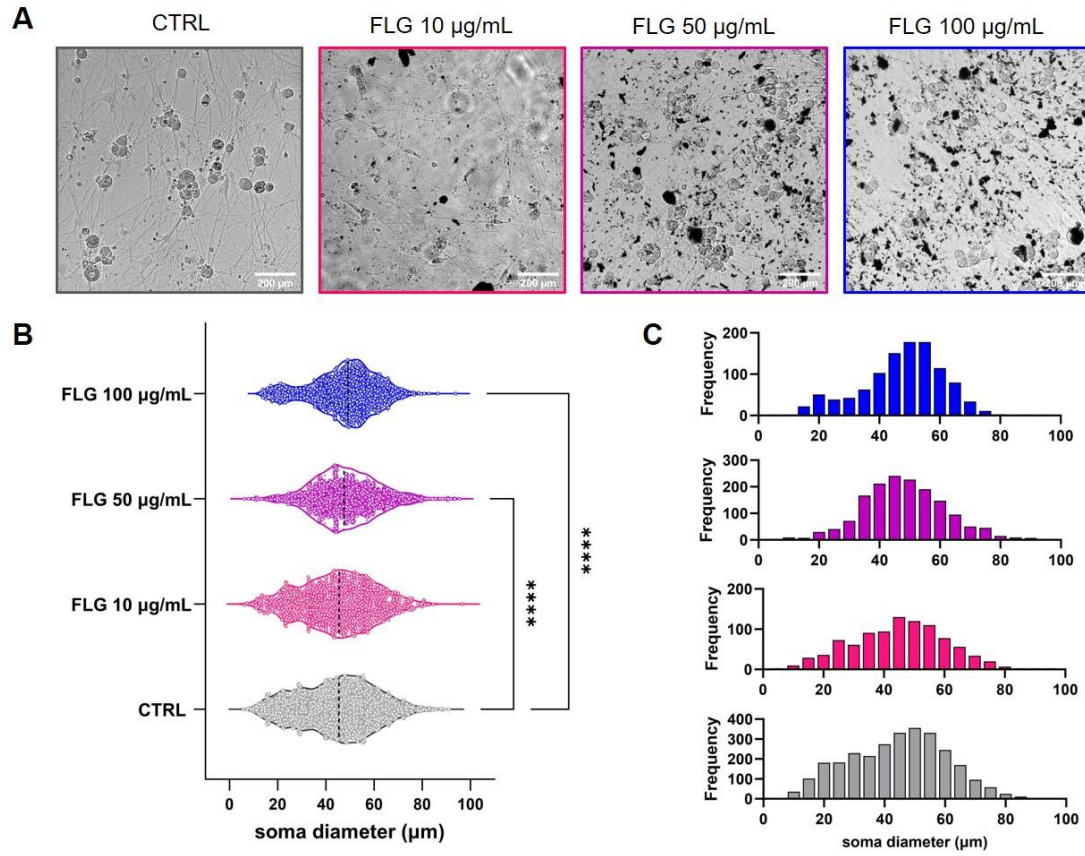


Figure S8. DRG size distribution analysis. **A)** Representative bright field (BF) images used for the 2D size distribution analysis. For increasing concentrations of FLG, dark agglomerates of increasing entities can be noticed. Scale bars: 200 µm. **B)** Violin plots representing the soma diameters when incubated with the different FLG doses (10 µg/mL: 955 cells; 50 µg/mL: 1573 cells; 100 µg/mL: 1074 cells) compared to untreated (CTRL: 2851 cells) conditions. Median value is represented by the black dotted line. Statistical analysis is one-way ANOVA with Tukey's post hoc test, ****= $p < 0.0001$. **C)** Statistical size distributions of the soma diameters when incubated with the different FLG doses compared to untreated (CTRL) conditions. Bin size = 5.

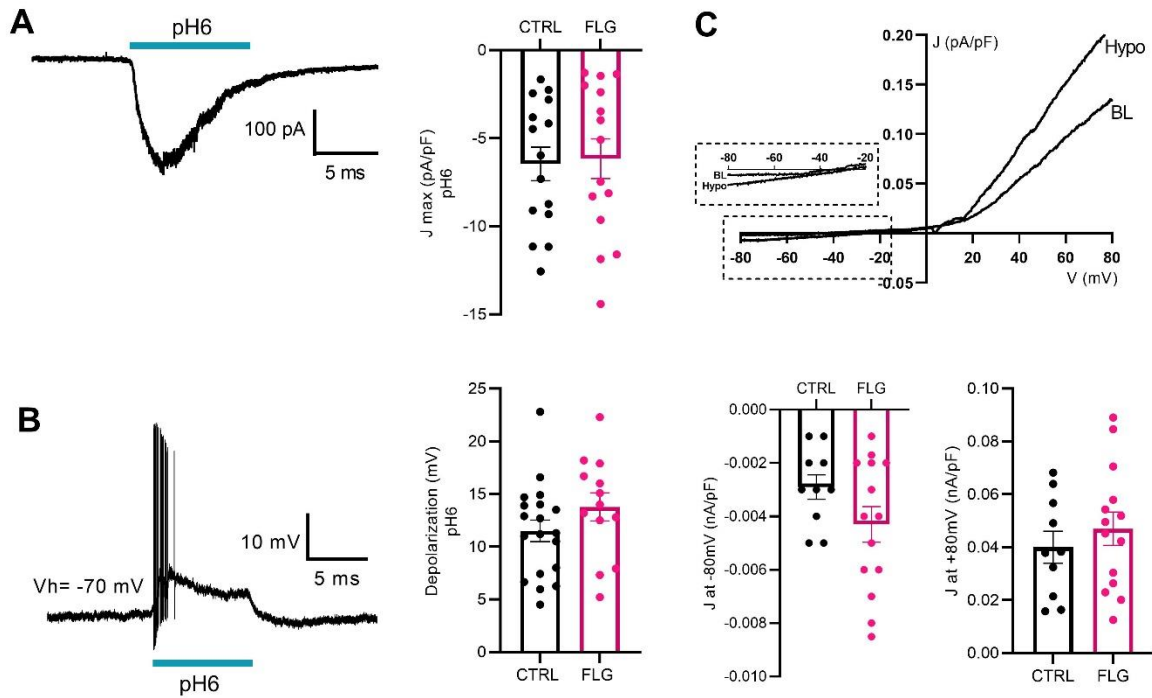


Figure S9. Response of DRG sensory neurons to pH and hypotonic solution. **A) Left:** Representative pH 6-elicited inward currents recorded at a holding potential of -70 mV in control and FLG-treated neurons. **Right:** Quantitative evaluation of current density after pH 6 stimulus under both experimental conditions (CTRL, n=15; FLG, n=15). **B) Left:** Representative current-clamp recordings of membrane depolarization evoked by pH 6 application in CTRL and FLG-treated neurons. **Right:** Quantitative evaluation of pH 6-dependent depolarization studied under both experimental conditions (CTRL, n=19; FLG, n=13). **C) Top:** Representative currents elicited by standard solution (baseline, BL) and hypotonic solution (Hypo) using a voltage ramp protocol. In the inset, magnification of the currents recorded in the -20 to -80 mV range. **Bottom:** Quantitative evaluation of whole-cell currents recorded at 80 mV and -80 mV in CTRL and FLG-treated neurons (CTRL, n=10; FLG, n=14). Data are expressed as means \pm SEM. Statistical analysis was performed using the unpaired Student's *t*-test.

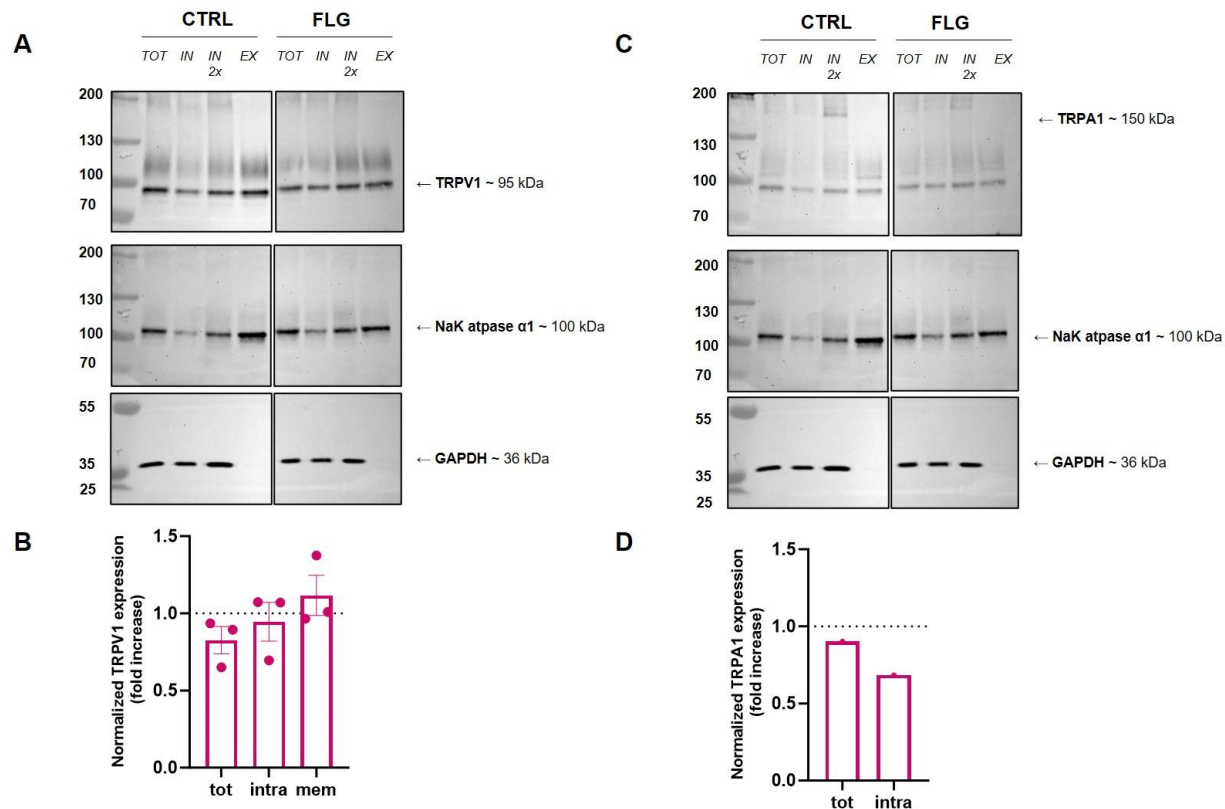


Figure S10. TRPV1 and TRPA1 expression and translocation. Western blotting (WB) was performed on DRG lysates at DIV4, after 24 h exposure to $10 \mu\text{g mL}^{-1}$ of FLG. Lysine biotinylation was performed to separate membrane and intracellular compartments to quantify the plasma membrane and intracellular fractions of the channels. **A)** Representative WB stained with antibodies to TRPV1 (95 kDa), the intracellular protein GAPDH (36 kDa), and the plasma membrane-associated protein Na^+/K^+ ATPase (100 kDa). **B)** Quantification of TRPV1 expression in FLG-treated ($10 \mu\text{g/mL}$, 24 h) samples (total, intracellular, and membrane compartments) normalized over untreated control conditions. Data are expressed as means \pm SEM, $N=3$. **C)** Representative WB stained with antibodies to TRPA1, GAPDH, and Na^+/K^+ ATPase. The extracellular lane is empty because TRPA1 does not contain lysine residues in the extracellular loops. **D)** Quantification of TRPA1 expression (total, intracellular) normalized over untreated control conditions ($N=1$).

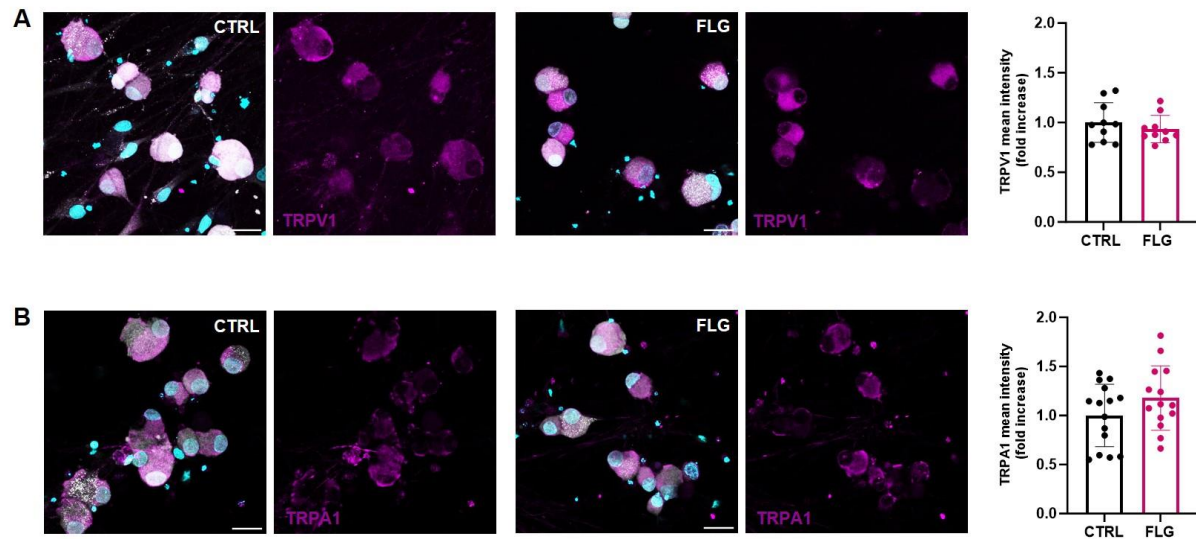


Figure S11. Extracellular TRPV1 and TRPA1. Representative confocal images (left panels) and fluorescence intensity quantification (right panels) of the extracellular exposure of TRPV1 (**A**) and TRPA1 (**B**) labeled by specific antibodies in non-permeabilized control and FLG-treated (10 μ g/mL, 24 h) DRG neurons. Nuclei (cyan), intracellular compartment (gray), and channels (purple). Scale bars, 25 μ m.