

Supporting Information

Collagenase Motors in Gelatine-Based Hydrogels

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Table S1. Overview of the silica particle properties

Silica Particle	Mass (mg) ^{a)}	Surface area (μm ²) ^{a)}	Diameter (μm) ^{a)}
^{0.5} M	1 × 10 ⁻¹⁰	0.8	0.5
^{0.8} M	6 × 10 ⁻¹⁰	2.0	0.8
¹ M	1 × 10 ⁻⁹	3.1	1.0
⁴ M	9 × 10 ⁻⁹	12.6	2.0
^R M	2 × 10 ⁻¹⁰	1.4	1.5 (width) 0.3 (length) 5.6 (aspect ratio)

^{a)} of a single particle

Table S2. Overview of the collagenase immobilization characteristics

Motor	Silica particle stock solution (mg mL ⁻¹)	Particle V (polymer brush coated) (μL) ^{a)}	Mass of Coll for immobilization (mg) ^{b)}	Total mass of Coll/Total area of motor (μg μm ⁻²) ^{c)}
^R M ^{Coll}			1.5	4 × 10 ⁻⁸
^{0.5} M ^{Coll}	30	66	1.4	5 × 10 ⁻⁸
^{0.8} M ^{Coll}	50	60	1.6	9 × 10 ⁻⁸
¹ M ^{Coll}	50	80	3.2	3 × 10 ⁻⁸
² M ^{Coll}	50	180	2.4	8 × 10 ⁻⁸
^{0.5} M ^{Coll} (low)	30	45	0.4	NA ^{d)}

^{a)}particle volume used for Coll coating to have comparable total surface area of 1 × 10¹⁰ μm²; ^{b)}from a 4 mg ml⁻¹ Coll stock solution in HEPES buffer; ^{c)}experimentally determined from a BCA analysis (n=2); ^{d)}below the detection limit of the BCA analysis.

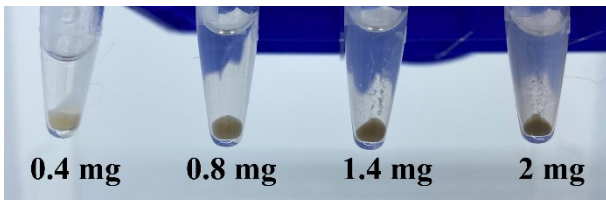
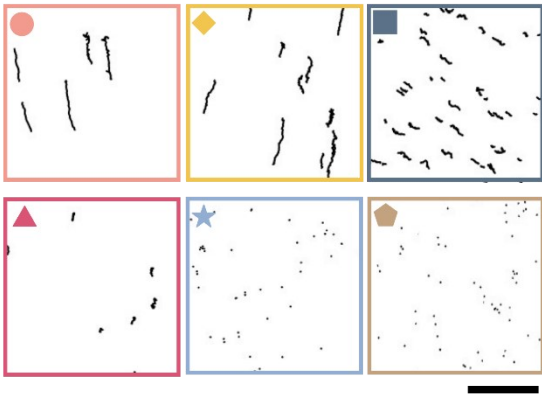
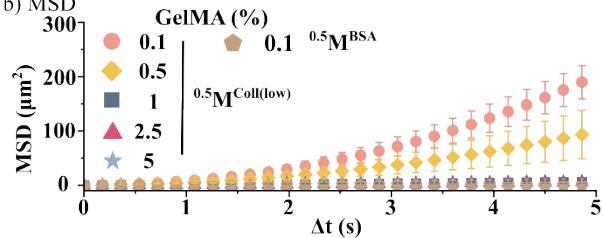


Figure S1. Photo of $0.5\text{M}^{\text{Coll}}$ after being coated with different amounts of collagenase. $0.5\text{M}^{\text{Coll}(\text{low})}$ and $0.5\text{M}^{\text{Coll}}$ correspond to the first tube (0.4 mg collagenase) and third tube (1.4 mg collagenase).

a) Trajectories



b) MSD



c) Velocity

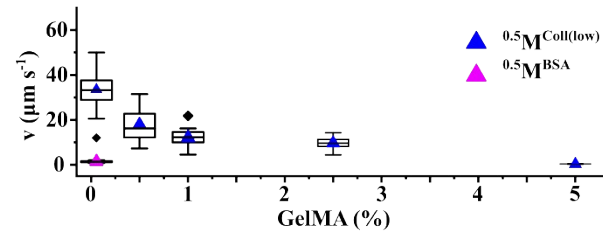
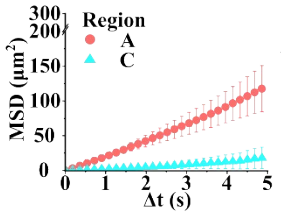
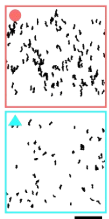


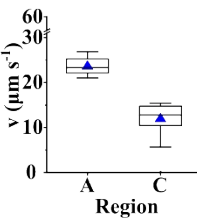
Figure S2. Locomotion properties of $0.5\text{M}^{\text{Coll}(\text{low})}$. Trajectories maps (a) and MSD plots (b) of $0.5\text{M}^{\text{Coll}(\text{low})}$ moving in different viscosity gelatine-based environments when triggered with 2 mM calcium. Scale bar is 50 μm . c) Velocities of $0.5\text{M}^{\text{Coll}(\text{low})}$ represented as whisker plots. 0.5M^{BSA} is used as control.

a) Locomotion properties of 0.5M and $0.5\text{M}^{\text{Coll}}$

i) Trajectories ii) MSD

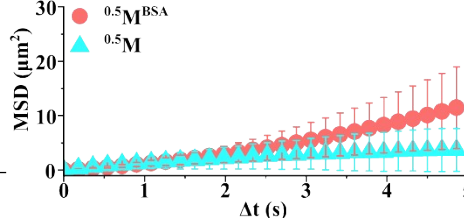


iii) Velocity



b) Locomotion properties of 0.5M^{BSA} and 0.5M

i) MSD



ii) Velocity

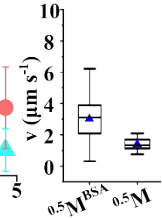


Figure S3. a) Mixture of motors 0.5M and $0.5\text{M}^{\text{Coll}}$ in the regions A (close to the inlet) and C (at the other end of the microfluidic channel). Trajectories maps (i) and MSD plots (ii) of $0.5\text{M}^{\text{Coll}}$ and 0.5M moving in an environment made of 1% GelMA when triggered with 2 mM calcium. Scale bar is 100 μm . iii) Velocities of $0.5\text{M}^{\text{Coll}}$ and 0.5M represented as whisker plots. b) i) MSD plots of 0.5M^{BSA} and 0.5M in 1% GelMA using 2 mM calcium as the trigger (not as a mixed population). ii) Velocities of 0.5M^{BSA} and 0.5M represented as whisker plots.

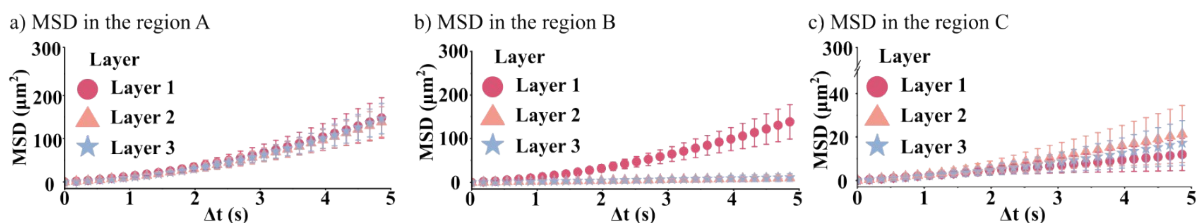


Figure S4. MSD plots of $^{0.5}M^{Coll}$ in the region A (a), the region B (b) and the region C (c) in 1 % GelMA using 2 mM calcium as the trigger.

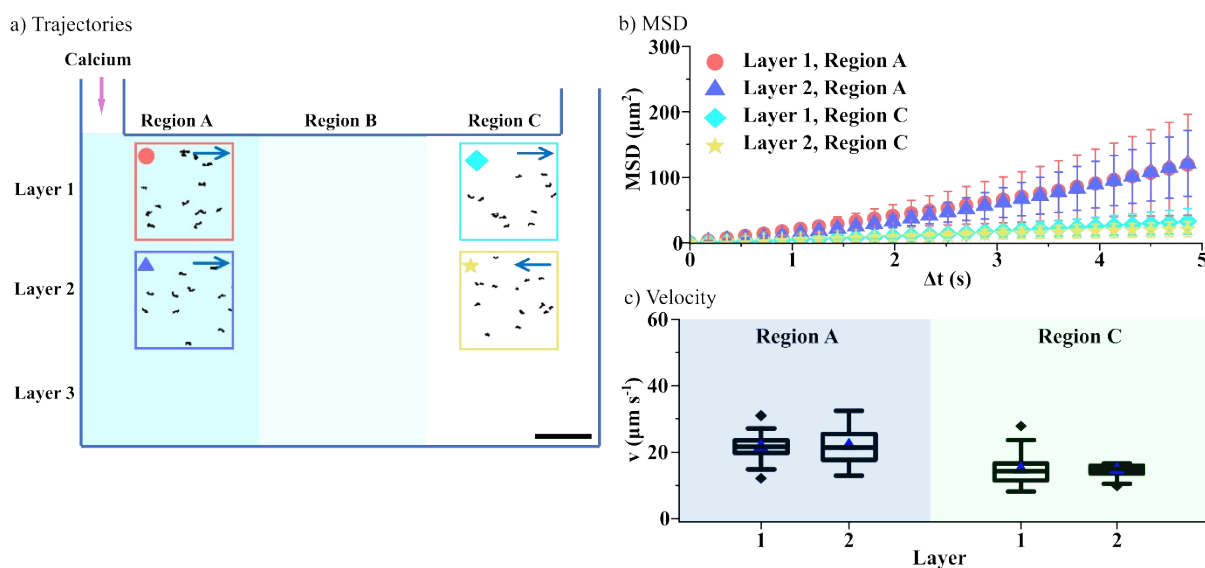


Figure S5. Motors $^{0.5}M$ and $^{0.5}M^{Coll}$ in different positions. Trajectories maps (a) and MSD plots (b) of $^{0.5}M^{Coll}$ and $^{0.5}M$ moving in an environment made of 1% GelMA when triggered with 2 mM calcium. Scale bar is 50 μm . c) Velocities of $^{0.5}M^{Coll}$ and $^{0.5}M$ represented as whisker plots.

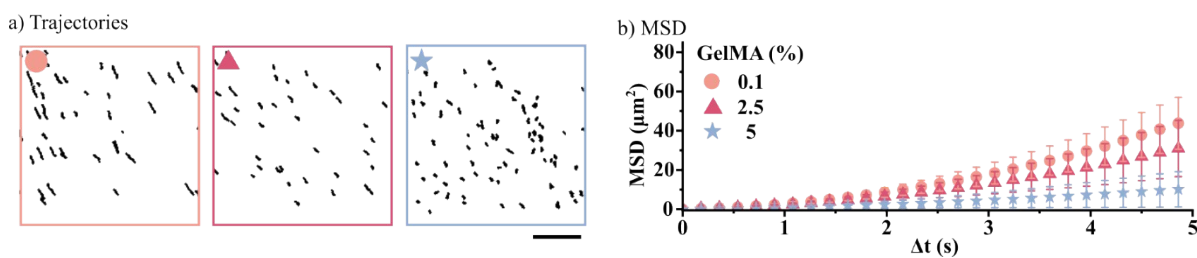


Figure S6. Trajectory maps (a) and MSD plots (b) of $^{0.8}M^{Coll}$ in hydrogels made from 0.1, 2.5, or 5 % GelMA using 2 mM calcium as the trigger. Scale bar is 100 μm .

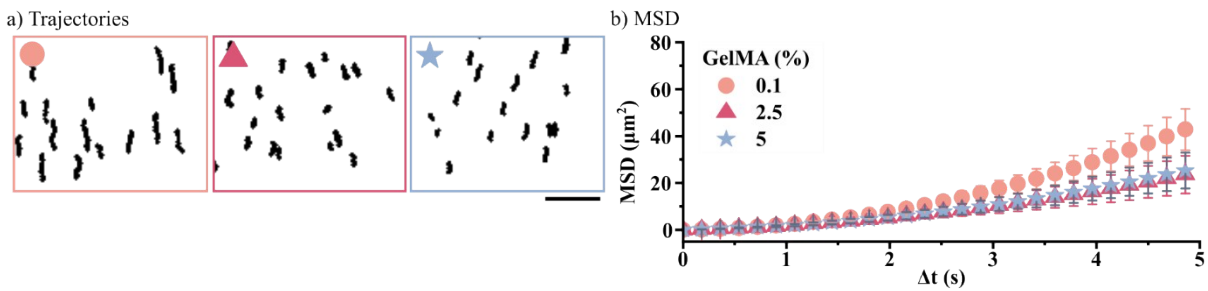


Figure S7. Trajectory maps (a) and MSD plots (b) of $^1M^{Coll}$ in hydrogels made from 0.1, 2.5, or 5 % GelMA using 2 mM calcium as the trigger. Scale bar is 100 μm .

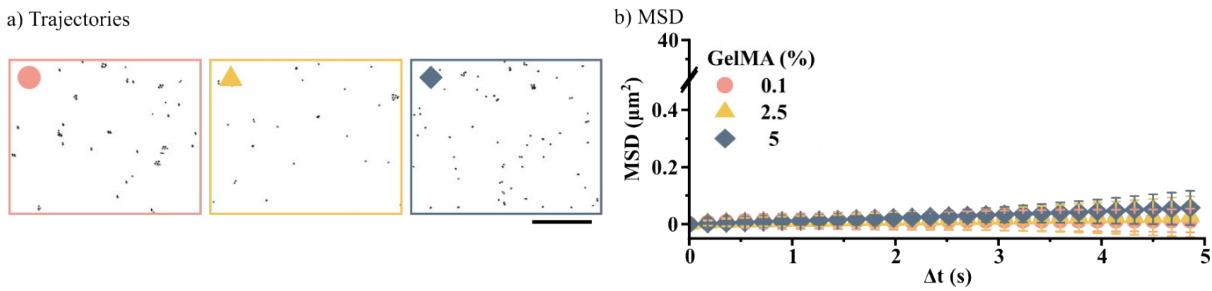


Figure S8. Trajectory maps (a) and MSD plots (b) of $^2M^{Coll}$ in hydrogels made from 0.1, 2.5, or 5 % GelMA using 2 mM calcium as the trigger. Scale bar is 50 μm .

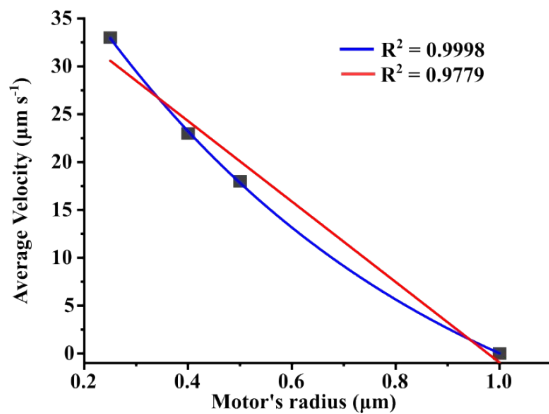


Figure S9. Plot of velocity vs. motor's radius comparing a linear (red) and an exponential fit (blue). Theory predicts a linear dependence of velocity and particle radius for slow particles moving in continuous fluids, while a non-linear dependence is expected for fast particles and complex environments.

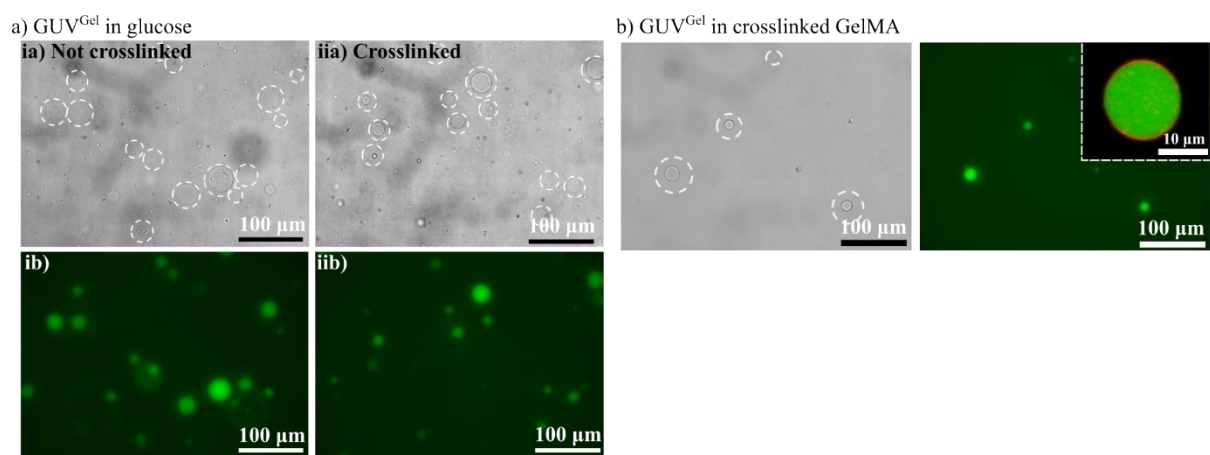


Figure S10. a) Representative fluorescence inverted microscope image of GUV^{Gel} in glucose buffer before and after UV crosslinking. b) Representative fluorescence inverted microscope image of GUV^{Gel} in crosslinked gelatine.