**Supporting Information for** 

# From Oximes to Tertiary Alcohols in Water, at Room Temperature in Air: Hybrid One-pot Tandem Assembly of Enzymatic Deoximation and RLi/RMgX Reagents

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# Table of Contents

Abbreviations list	<b>S</b> 3
Experimental details	S4
Deoximation of propiophenone oxime 1a into ketone 2a promoted by the system	
laccase/TEMPO/O <sub>2</sub> in aqueous media at room temperature after 24 hours	S5
Hybrid one-pot tandem transformation of ketoxime 1a into tertiary alcohols 3a-b promoted by	
combination of the laccase/TEMPO/O $_2$ system with the chemoselective addition of RLi reagents	
(R = Ph  or  n-Bu) in aqueous media, at room temperature and in the presence of air	S16
<sup>1</sup> H and <sup>13</sup> C NMR spectra of compounds <b>3a-b</b>	S21
Hybrid one-pot tandem transformation of ketoxime 1a into tertiary alcohols 3c-i promoted by	
combination of the laccase/TEMPO/O <sub>2</sub> system with the chemoselective addition of RLi/RMgX	
in aqueous media, at room temperature and in the presence of air	S23
<sup>1</sup> H and <sup>13</sup> C NMR spectra of compounds <b>3c-i</b>	S30
Hybrid one-pot tandem transformation of ketoximes <b>1a-j</b> into tertiary alcohols <b>3c,h,j-x</b> promoted	
by combination of the laccase/TEMPO/O2 system with the chemoselective addition of	
MeLi/AllylMgBr in aqueous media, at room temperature and in the presence of air	S37
<sup>1</sup> H and <sup>13</sup> C NMR spectra of compounds <b>3j-x</b>	S43
References	<b>S</b> 58

# Abbreviations list

AZADO	2-azaadamantane-N-oxyl
BnMgCl	benzylmagnesium chloride
n-BuLi	normal-butyllithium
s-BuLi	sec-butyllithium
t-BuLi	<i>tert</i> -butyllithium
CDCl <sub>3</sub>	deuterated chloroform
CHCl <sub>3</sub>	chloroform
CPME	cyclopentyl methyl ether
DCM	dichloromethane
Et <sub>2</sub> O	diethyl ether
EtOAc	ethyl acetate
GC-FID	gas chromatography-flame ionization detector
CH <sub>3</sub> CN	acetonitrile
MeLi	methyllithium
NMR	nuclear magnetic resonance
PhLi	phenyllithium
$R_{f}$	retention factor
TEMPO	2,2,6,6-tetramethylpiperidine-1-oxyl
THF	tetrahydrofuran
TLC	thin layer chromatography
UV	ultraviolet

#### **Experimental Details**

**Materials and methods**. All reagents were obtained from commercial suppliers and used without further purification. Laccase from *Trametes Versicolor* or *Rhus Vernicifera*, TEMPO and AZADO were purchased from Sigma Aldrich. Organometallic reagents were purchased from Sigma Aldrich: i) 2.5 M solution of *n*-BuLi in hexanes; ii) 1.6 M solution of MeLi in Et<sub>2</sub>O; iii) 1.4 M solution of *s*-BuLi in cyclohexane; iv) 1.7 M solution of *t*-BuLi in pentane; v) 1.9 M solution of PhLi in dibutyl ether; vi) 2.0 M solution of BnMgCl in THF; vii) 1.0 M solution of 2-thienyllithium in THF/hexanes; viii) 1.0 M solution of allylmagnesium bromide in Et<sub>2</sub>O. Concentrations of all organolithium reagents were determined by titration with L-menthol,<sup>1</sup> and for the Grignard reagents titration against iodine was employed.<sup>2</sup> All the rest of reagents and solvents were of the highest quality available. Ketoximes **1a-j** were synthesized according to the procedure reported in the literature.<sup>3</sup> Reactions were monitored by GC-FID analysis or by thin-layer chromatography (TLC) carried out on 0.25 mm silica gel coated aluminum plates (60 Merck F<sub>254</sub>) with UV light (254 nm) or *p*-anisaldehyde indicator<sup>4</sup> as visualizing agents. *R<sub>f</sub>* values refer to TLC carried out on silica gel plates. Chromatographic separations were carried out under pressure on silica gel (40-63 µm, 230-400 mesh) using flash-column techniques. Full characterization data, including copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra, have been presented for the known compounds.

**Instrumentation**. <sup>1</sup>H NMR (600 MHz) and <sup>13</sup>C{<sup>1</sup>H} (150 MHz) NMR spectra were recorded on a Jeol ECZR600 spectrometer at room temperature. <sup>1</sup>H NMR (300 MHz) and <sup>13</sup>C{<sup>1</sup>H} (75 MHz) NMR spectra were recorded on a Bruker DPX-300 spectrometer at room temperature. Calibration was made on the signal of the residual solvent (<sup>1</sup>H CHCl<sub>3</sub>: 7.26 ppm; <sup>13</sup>C{<sup>1</sup>H} CDCl<sub>3</sub>: 77.16 ppm). Chemical shifts ( $\delta$ ) are given in parts per million (ppm) and coupling constants (*J*) in Hertz (Hz). Multiplicities are reported as follows: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), br (broad). Gas chromatography (GC) analyses were performed on an Agilent Technologies 7820A chromatographic system equipped with a HP-5 (30 m x 0.32 mm x 0.25 µm) column. Melting points were determined on a Stuart Scientific SMP3 melting point apparatus.

# Deoximation of propiophenone oxime 1a into ketone 2a promoted by the system laccase/TEMPO/O<sub>2</sub> in aqueous media at room temperature after 24 hours

All reactions were performed at room temperature. In an 8 mL vial equipped with a magnetic stirrer Laccase and co-catalyst (eq.) were added to a 0.73 mmol (109 mg) suspension of propiophenone oxime **1a** in water (1 mL) and the mixture was stirred under the selected atmosphere for 24 h. Then, the reaction mixture was extracted with dichloromethane (3 x 5 mL), the organic layers were combined, washed with brine (1 x 5 mL), dried with Na<sub>2</sub>SO<sub>4</sub> and the solvent removed *in vacuo*. Conversion of **1a** into propiophenone **2a** was determined by GC-FID analysis of the crude reaction mixtures (presented in Figures S8-S17). A sample of **1a** was synthesized according to the procedure reported in the literature<sup>3</sup> and used as reference for GC-FID analyses (<sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR presented in Figure S1 and S2, GC-FID presented in Figure S5). A sample of commercial **2a** (Sigma-Aldrich 99%) was analyzed and used as reference for GC-FID analyses of the reaction crudes (<sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR presented in Figure S3 and S4, GC-FID presented in Figure S6).

**Table S1**. Deoximation of propiophenone oxime **1a** into ketone **2a** promoted by the system laccase/TEMPO/O<sub>2</sub> in aqueous media at room temperature after 24 hours.<sup>*a*</sup>

OH	Laccase / Air or O <sub>2</sub>		
N <sup>*</sup>	Co-catalyst (10-33 mol%)	_	O II
Ph	H <sub>2</sub> O / rt / 1800 rpm / 24 h		Ph
( <b>1</b> a)			(2a)

Entry	Laccase <sup>b, c</sup>	Co-catalyst	Oxidant	Conversion <sup>d</sup> (%)
1	T. Versicolor	TEMPO (33 mol%)	Air	47 <sup>e</sup>
2	T. Versicolor	TEMPO (33 mol%)	Air	84
3	T. Versicolor	TEMPO (10 mol%)	Air	40
4	T. Versicolor	TEMPO (33 mol%)	$O_2$	>99
5	Rhus Vernicifera	TEMPO (33 mol%)	$O_2$	1
6	CuCl <sub>2</sub> ·2H <sub>2</sub> O/TMEDA	TEMPO (33 mol%)	$O_2$	1
7	T. Versicolor	AZADO (33 mol%)	$O_2$	>99
8	T. Versicolor	TEMPO (33 mol%)	$O_2$	>99 <sup>f</sup>
9	-	TEMPO (33 mol%)	$O_2$	0
10	T. Versicolor	-	$O_2$	2

<sup>*a*</sup> General conditions: 24 h of reaction at room temperature and at 1800 rpm, using 0.73 mmol of **1a** in 1 mL of water. <sup>*b*</sup> 280 mg of *T. Versicolor* (0.5 U/mg); 2.8 mg of *Rhus Vernicifera* (50 U/mg) were employed <sup>*c*</sup> U/mg = Units of activity per mg of enzyme. <sup>*d*</sup> Determined by GC-FID, no significant amount of by-products was detected [110 °C; 4 min; 10 °C/min; 220 °C; 2 min]. <sup>*e*</sup> Stirring speed 800 rpm. <sup>*f*</sup> 100 µL of CH<sub>3</sub>CN were added as co-solvent.



**1-phenylpropan-1-one oxime (1a):** white solid ( $R_f = 0.23$  hexane/EtOAc 8/2 v/v), mp 50.2–51.6 °C (hexane). Mixture of *E* and *Z* stereoisomers (E/Z = 10/1). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, mixture of *E* and *Z* stereoisomers):  $\delta$  7.88 (br s, 2H, *E* + *Z*), 7.68-7.51 (m, 4H, *E* + *Z*), 7.50-7.28 (m, 6H, *E* + *Z*), 2.84 (q, *J* = 7.6 Hz, 2H, *E*), 2.63 (q, *J* = 7.4 Hz, 2H, *Z*), 1.19 (t, *J* = 7.6 Hz, 3H, *E*) superimposed to 1.12 (t, *J* = 7.6 Hz, 3H, *Z*). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, mixture of *E* and *Z* stereoisomers):  $\delta$  160.9 (*E*), 159.7 (*Z*), 135.7 (*E*), 133.7 (*Z*), 129.3 (*E*), 129.0 (*Z*), 128.7 (*E*), 128.4 (*Z*), 127.9 (*Z*), 126.4 (*E*), 29.1 (*Z*), 19.9 (*E*), 11.3 (*Z*), 11.0 (*E*).<sup>5</sup>



**1-phenylpropan-1-one (2a):** colorless liquid ( $R_f = 0.25$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.97 (d, J = 7.4 Hz, 2H), 7.55 (t, J = 7.3 Hz, 1H) superimposed to 7.46 (t, J = 7.3 Hz, 2H), 3.01 (q, J = 7.2 Hz, 2H), 1.23 (t, J = 7.2 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  200.9, 137.1, 133.0, 128.7, 128.1, 31.9, 8.4.<sup>6</sup>

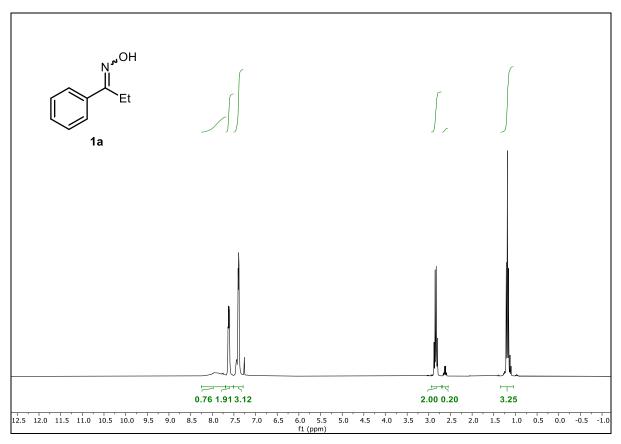


Figure S1. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) spectrum of propiophenone oxime 1a.

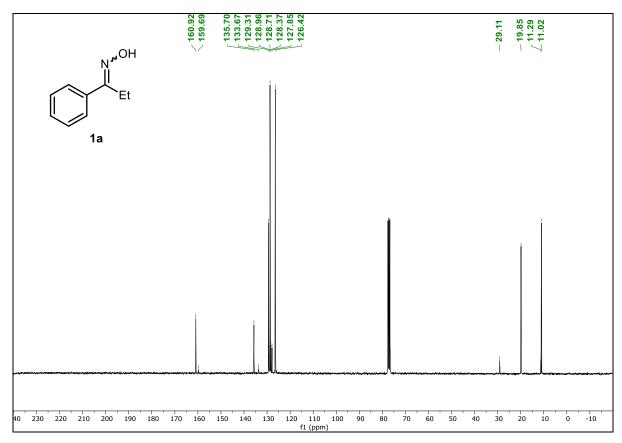


Figure S2. <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>) spectrum of propiophenone oxime 1a.

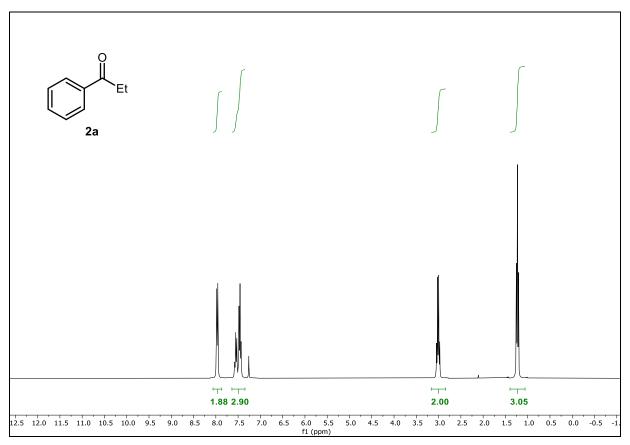


Figure S3. <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) spectrum of propiophenone 2a.

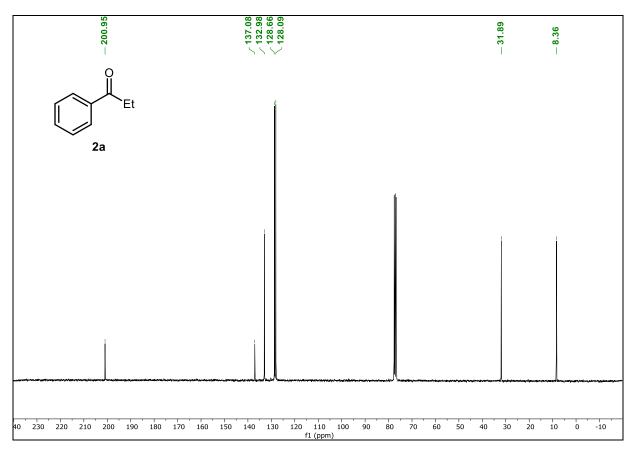


Figure S4. <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>) spectrum of propiophenone 2a.

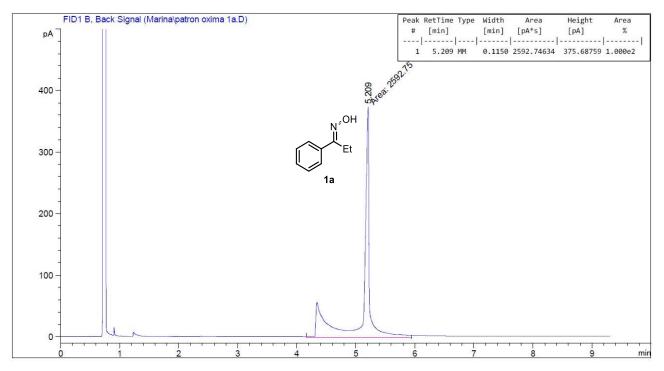


Figure S5. GC-FID chromatogram of propiophenone oxime (1a).

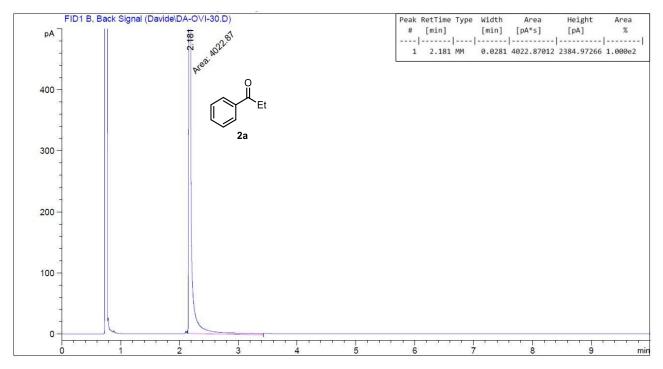


Figure S6. GC-FID chromatogram of propiophenone (2a).

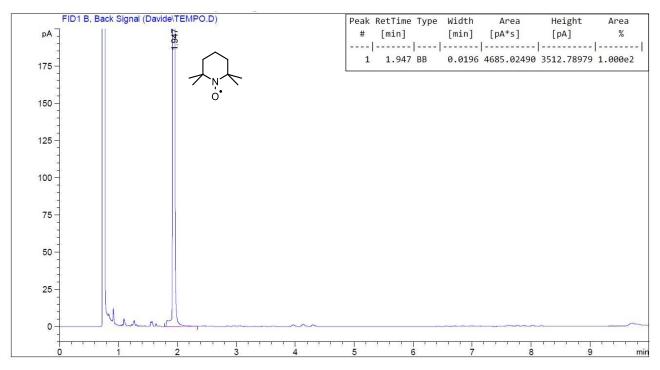


Figure S7. GC-FID chromatogram of TEMPO.

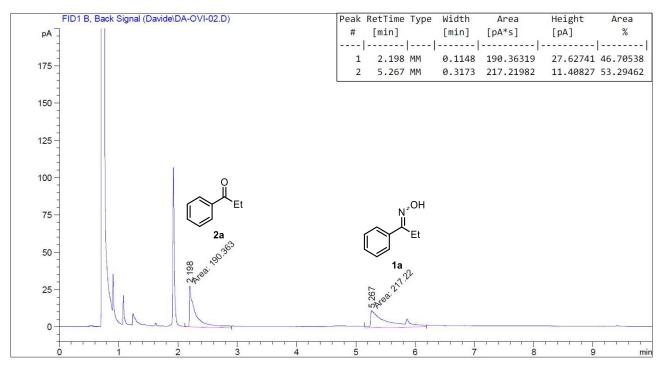


Figure S8. Table S1, entry 1: GC-FID chromatogram of the reaction crude.

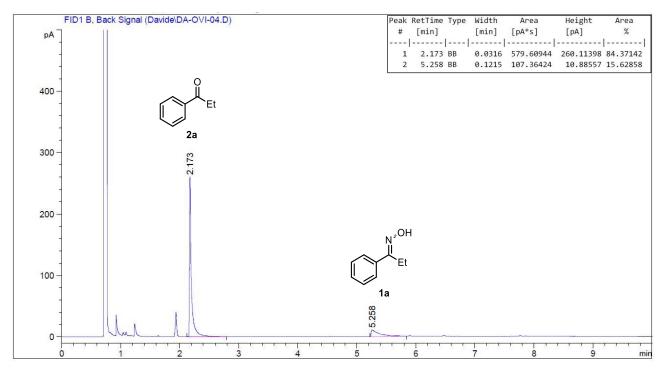


Figure S9. Table S1, entry 2: GC-FID chromatogram of the reaction crude.

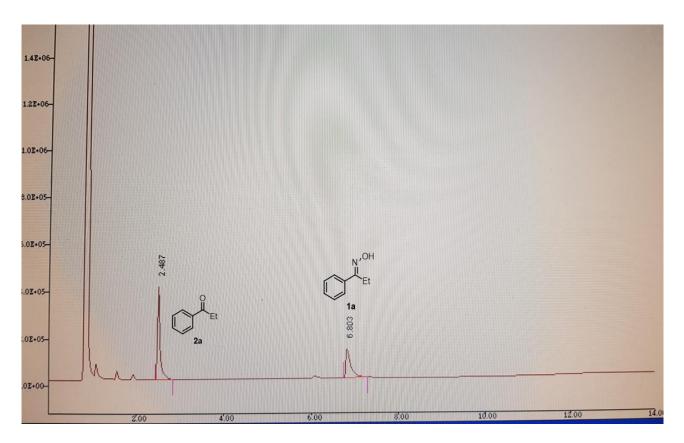


Figure S10. Table S1, entry 3: GC-FID chromatogram of the reaction crude.

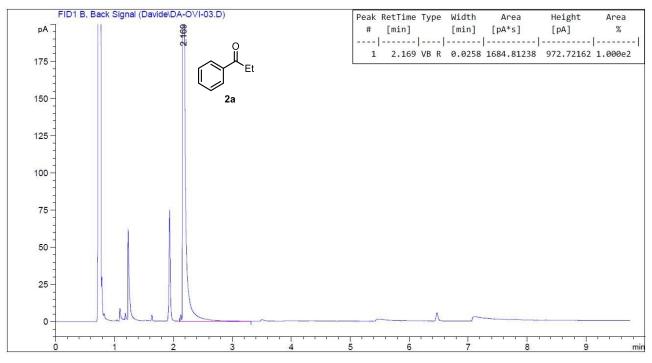


Figure S11. Table S1, entry 4: GC-FID chromatogram of the reaction crude.

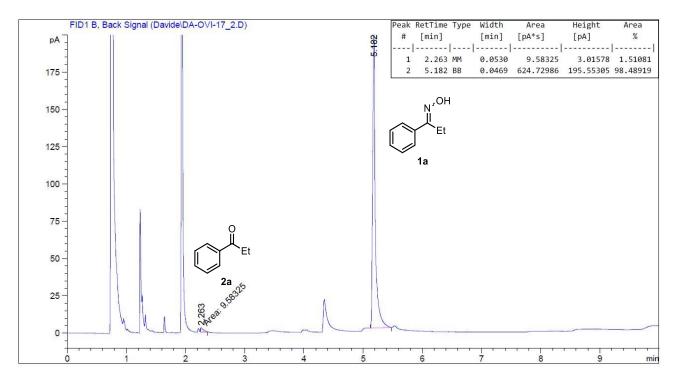


Figure S12. Table S1, entry 5: GC-FID chromatogram of the reaction crude.

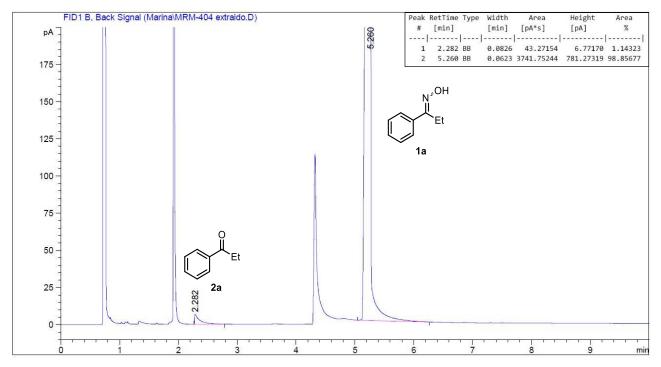


Figure S13. Table S1, entry 6: GC-FID chromatogram of the reaction crude.

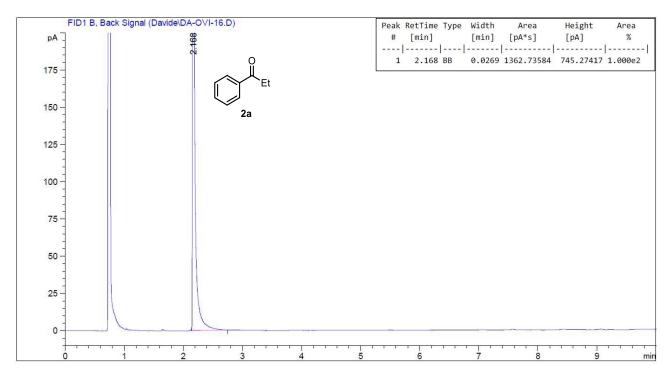


Figure S14. Table S1, entry 7: GC-FID chromatogram of the reaction crude.

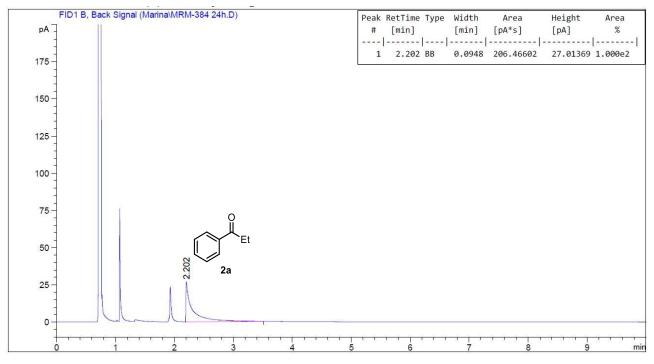


Figure S15. Table S1, entry 8: GC-FID chromatogram of the reaction crude.

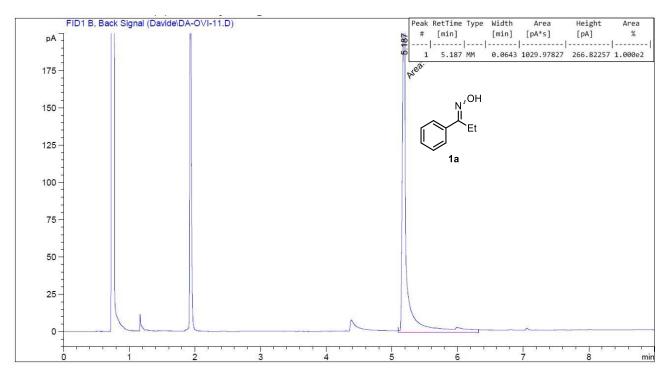


Figure S16. Table S1, entry 9: GC-FID chromatogram of the reaction crude.

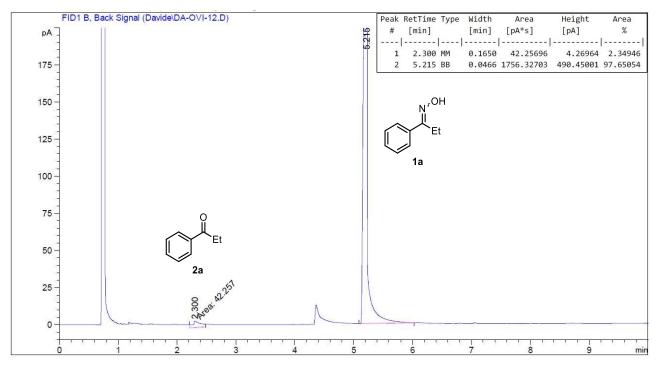


Figure S17. Table S1, entry 10: GC-FID chromatogram of the reaction crude.

Hybrid one-pot tandem transformation of ketoxime **1a** into tertiary alcohols **3a-b** promoted by combination of the laccase/TEMPO/O<sub>2</sub> system with the chemoselective addition of RLi reagents (R = Ph or *n*-Bu) in aqueous media, at room temperature and in the presence of air

*T. versicolor* laccase (280 mg, 0.5 U/mg) and TEMPO (38 mg, 33 mol%) were added to a 0.73 mmol (109 mg) suspension of propiophenone oxime **1a** in water (1 mL) and the mixture was stirred vigorously (1800 rpm) in an 8 mL vial under oxygen atmosphere for 24 h. Once the biodeoximation reaction was completed (GC-FID analysis, 24 h), 1 mL of ethereal co-solvent was added to form a biphasic reaction medium (apart from Table S2, entry 1). Next, the corresponding organolithium reagent (RLi, selected equivalents) was rapidly spreaded over the reaction mixture at room temperature, under air. After 10 s, a saturated solution of NH<sub>4</sub>Cl<sub>aq</sub> (2.5 mL) was added, and the mixture was extracted with dichloromethane (3 x 5 mL). The combined organic phases were washed with brine (1 x 5 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and the solvents were removed *in vacuo*. Conversion of **1a** into tertiary alchols **3a-b** was determined by GC-FID analysis of the crude reaction mixtures (presented in Figures S18-S24). The crude products obtained with the optimized conditions (Table S2, entry 2 for **3a**; Table S2, entry 6 for **3b**) were purified by flash column chromatography and characterized by <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR.

**Table S2**. Hybrid one-pot tandem transformation of ketoxime **1a** into tertiary alcohols **3a-b** promoted by combination of the laccase/TEMPO/O<sub>2</sub> system with the chemoselective addition of RLi reagents (R = Ph or n-Bu) in aqueous media, at room temperature and in the presence of air.<sup>*a*</sup>

	Ph (1a)	Laccase / O <sub>2</sub> TEMPO (33 mol%) H <sub>2</sub> O / rt 1800 rpm / 24 h	$- \begin{bmatrix} 0 \\ Ph \\ (2a) \end{bmatrix}$	$R^{1}-Li$ $R^{1} = Ph, n-Bu$ $rt / 10 sec.$ aqueous solvent	OH Ph R <sup>1</sup> ( <b>3a-b</b> )
Entry	R <sup>1</sup> -Li	Equiv.	Solvent	Product	Conversion <sup><math>b</math></sup> (%)
1	PhLi	3.0	H <sub>2</sub> O	<b>3</b> a	67
2	PhLi	3.0	H <sub>2</sub> O/CPME	<b>3</b> a	79
3	PhLi	2.0	H <sub>2</sub> O/CPME	<b>3</b> a	60
4	PhLi	3.0	H <sub>2</sub> O/2-MeTHF	3a	65
5	<i>n</i> -BuLi	2.0	H <sub>2</sub> O/CPME	3b	28
6	<i>n</i> -BuLi	3.0	H <sub>2</sub> O/CPME	3b	62
7	<i>n</i> -BuLi	3.0	H <sub>2</sub> O/2-MeTHF	<b>3</b> b	37

<sup>*a*</sup> General conditions: 24 h of reaction at room temperature and at 1800 rpm; Laccase from *T. Versicolor* (0.5 U/mg, 280 mg) per 0.73 mmol of **1a**, 0.33 eq. TEMPO in 1 mL of water were used. Then 1 mL of the co-solvent and the RLi reagent [R = Ph (1.9 M in *n*-Bu<sub>2</sub>O) or *n*-Bu (2.5 M in hexanes)] were added without any isolation/purification. <sup>*b*</sup> Determined by GC-FID, no significant amount of by-products was detected [110 °C; 4 min; 10 °C/min; 220 °C; 2 min].

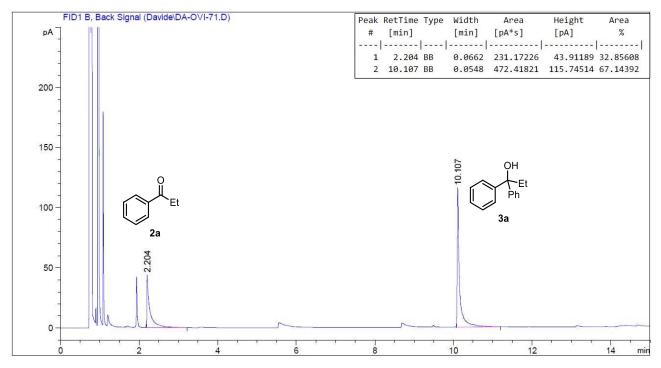


Figure S18. Table S2, entry 1: GC-FID chromatogram of the reaction crude.

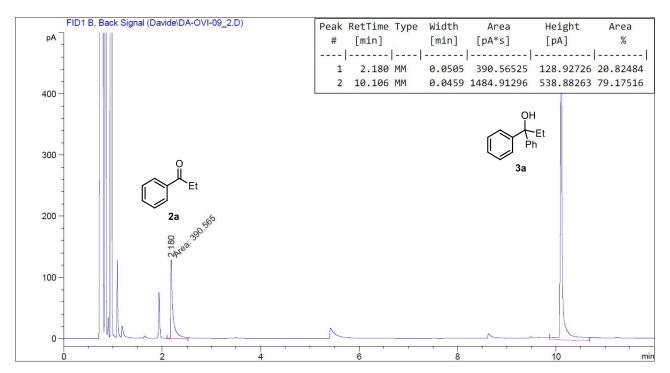


Figure S19. Table S2, entry 2: GC-FID chromatogram of the reaction crude.

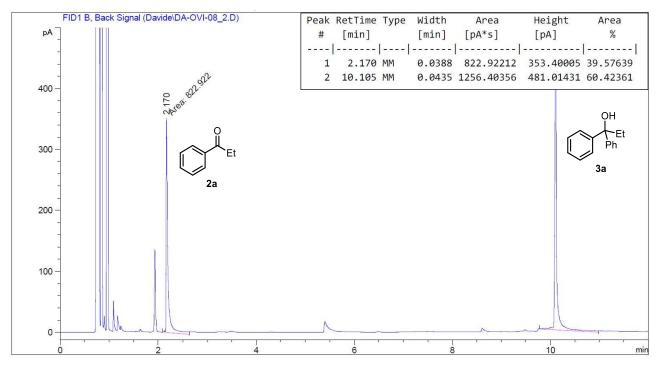


Figure S20. Table S2, entry 3: GC-FID chromatogram of the reaction crude.

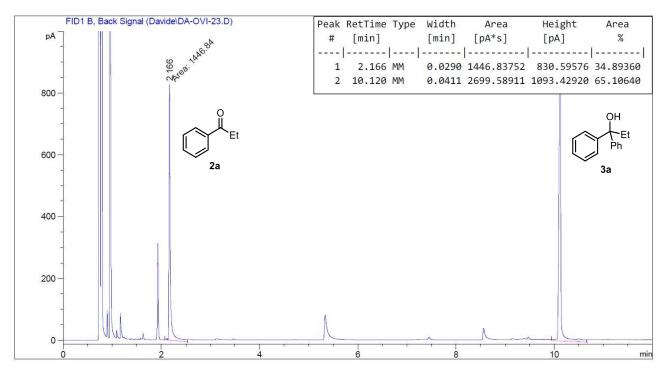


Figure S21. Table S2, entry 4: GC-FID chromatogram of the reaction crude.

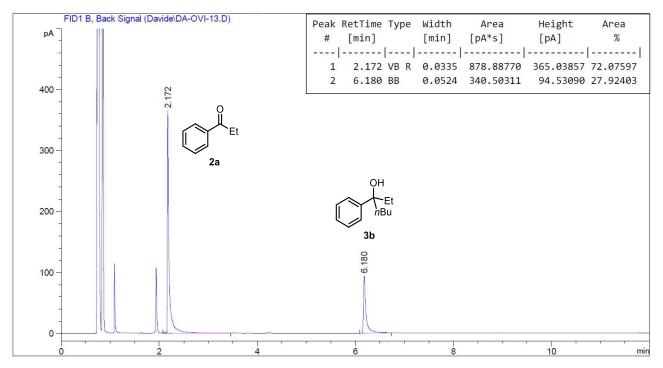


Figure S22. Table S2, entry 5: GC-FID chromatogram of the reaction crude.

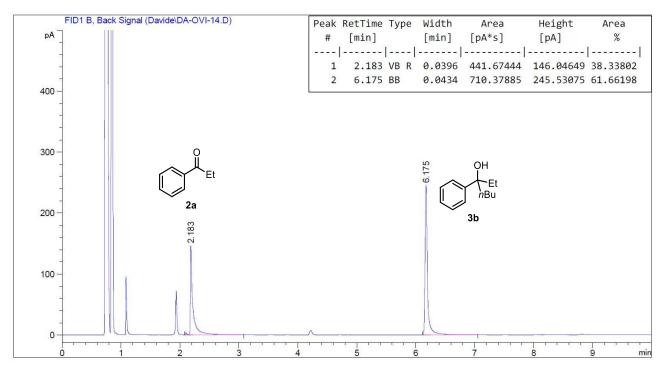


Figure S23. Table S2, entry 6: GC-FID chromatogram of the reaction crude.

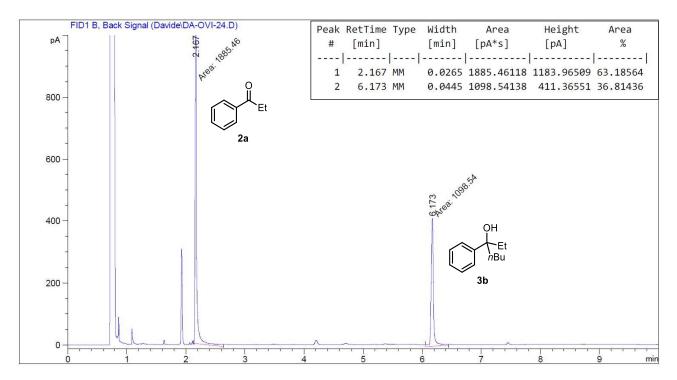


Figure S24. Table S2, entry 7: GC-FID chromatogram of the reaction crude.



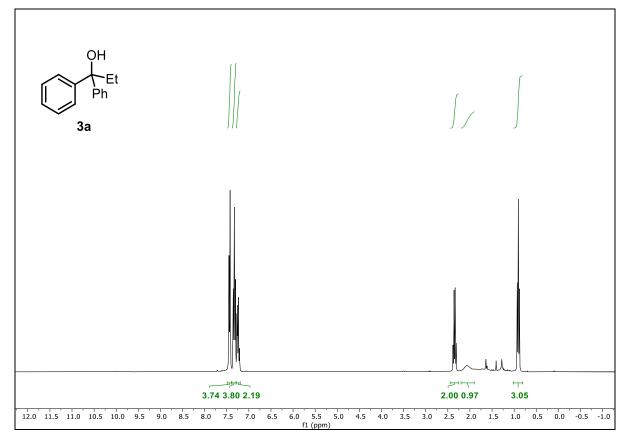
**1,1-diphenylpropan-1-ol (3a):** flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product **3a** as a white solid (70%,  $R_f = 0.19$  hexane/Et<sub>2</sub>O 9/1 v/v), mp 93.2–94.6 °C (hexane). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.44 (d, J = 8.0 Hz, 4H), 7.33 (t, J = 7.6 Hz, 4H), 7.28-7.18 (m, 2H), 2.35 (q, J = 7.3 Hz, 2H), 2.06 (br s, 1H), 0.91 (t, J = 7.3 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  147.0, 128.2, 126.9, 126.2, 78.6, 34.6, 8.2.<sup>7</sup>



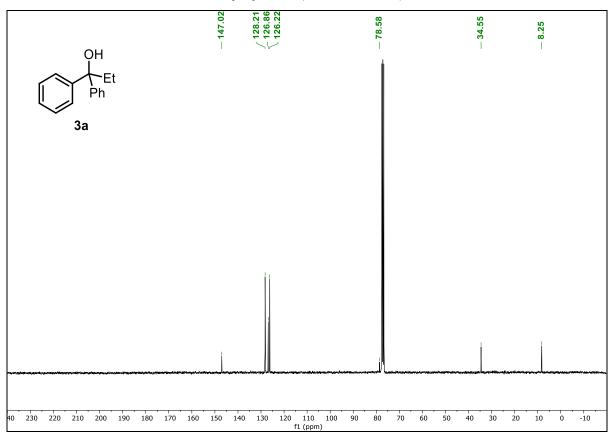
**3-phenylheptan-3-ol (3b):** flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product **3b** as a colorless oil (57%,  $R_f = 0.29$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.48-7.34 (m, 4H), 7.33-7.23 (m, 1H), 2.02-1.76 (m, 4H), 1.67 (br s, 1H), 1.40-1.22 (m, 3H), 1.18-1.00 (m, 1H), 0.90 (t, *J* = 6.9 Hz, 3H), 0.82 (t, *J* = 7.3 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  145.7, 127.6, 125.8, 124.9, 41.9, 35.0, 25.2, 22.7, 13.6, 7.4.<sup>7</sup>

# 1,1-diphenylpropan-1-ol (3a)

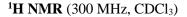


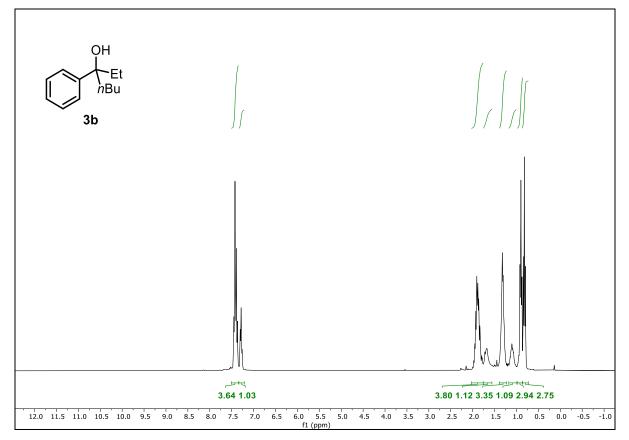


<sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>)

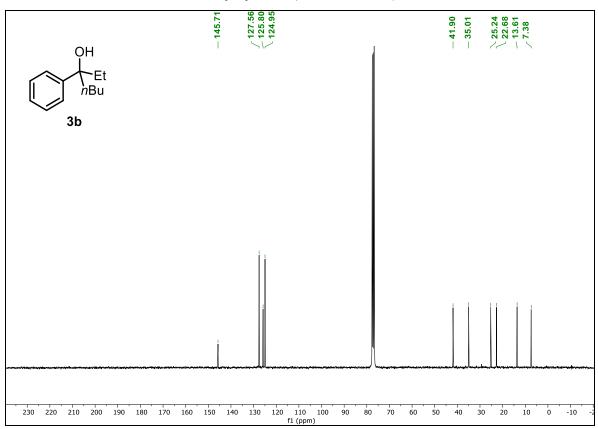


#### 3-phenylheptan-3-ol (3b)





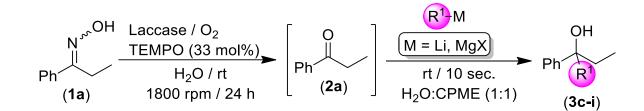
<sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>)



Hybrid one-pot tandem transformation of ketoxime **1a** into tertiary alcohols **3c-i** promoted by combination of the laccase/TEMPO/O<sub>2</sub> system with the chemoselective addition of RLi/RMgX in aqueous media, at room temperature and in the presence of air

*T. versicolor* laccase (280 mg, 0.5 U/mg) and TEMPO (38 mg, 33 mol%) were added to a 0.73 mmol (109 mg) suspension of propiophenone oxime **1a** in water (1 mL) and the mixture was stirred vigorously (1800 rpm) in an 8 mL vial under oxygen atmosphere for 24 h. Once the biodeoximation reaction was completed (GC-FID analysis, 24 h), 1 mL of CPME was added as co-solvent to form a biphasic reaction medium. Next, the corresponding organolithium (RLi, 3.0 eq) or Grignard (RMgX, 3.0 eq) reagent was rapidly spreaded over the reaction mixture at room temperature, under air. After 10 s, a saturated solution of NH<sub>4</sub>Cl<sub>aq</sub> (2.5 mL) (2.5 mL) was added, and the mixture was extracted with dichloromethane (3 x 5 mL). The combined organic phases were washed with brine (1 x 5 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and the solvents were removed *in vacuo*. Conversion of **1a** into tertiary alchols **3c-i** was determined by GC-FID analysis of the crude reaction mixtures (presented in Figures S25-S31). The crude products were purified by flash column chromatography and characterized by <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR.

**Table S3**. Hybrid one-pot tandem transformation of ketoxime **1a** into tertiary alcohols **3c-i** promoted by combination of the laccase/TEMPO/O<sub>2</sub> system with the chemoselective addition of RLi/RMgX in aqueous media, at room temperature and in the presence of air.<sup>*a*</sup>



Entry	$R^{1}-M^{b}$ (3.0 eq)	Product	Conversion <sup>c</sup> (%)	Yield <sup><math>d</math></sup> (%)
1	MeLi	3c	91	82
2	EtLi	3d	73	66
3	s-BuLi	3e	69	64
4	t-BuLi	<b>3f</b>	53	40
5	2-ThienLi	3g	55	46
6	AllylMgBr	3h	62	50
7	BenzylMgCl	<b>3i</b>	46	33

<sup>&</sup>lt;sup>*a*</sup> General conditions: 24 h of reaction at room temperature and at 1800 rpm; Laccase from *T. Versicolor* (0.5 U/mg, 280 mg) per 0.73 mmol of **1a**, 0.33 eq. TEMPO in 1 mL of water were used. <sup>*b*</sup> Then 1 mL of CPME and the RLi [R = Me (1.6 M in Et<sub>2</sub>O); Et (0.5 M in benzene/cyclohexane); *s*-Bu (1.4 M in cyclohexane); *t*-Bu (1.7 M in pentane); 2-thienyl (1.0 M in THF/hexanes)] or RMgX [R = allyl (1.0 M in Et<sub>2</sub>O); benzyl (2.0 M in THF)] reagents were added without any isolation/purification. <sup>*c*</sup> Determined by GC-FID, no significant amount of by-products was detected [110 °C; 4 min; 10 °C/min; 220 °C; 2 min]. <sup>*d*</sup> Isolated yield.

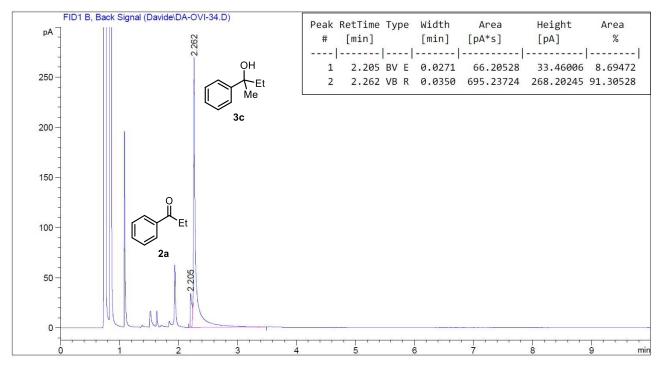


Figure S25. Table S3, entry 1 (R-M: MeLi): GC-FID chromatogram of the reaction crude.

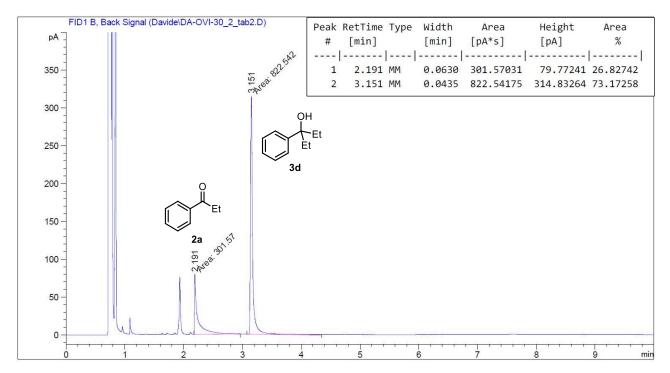


Figure S26. Table S3, entry 2 (R-M: EtLi): GC-FID chromatogram of the reaction crude.

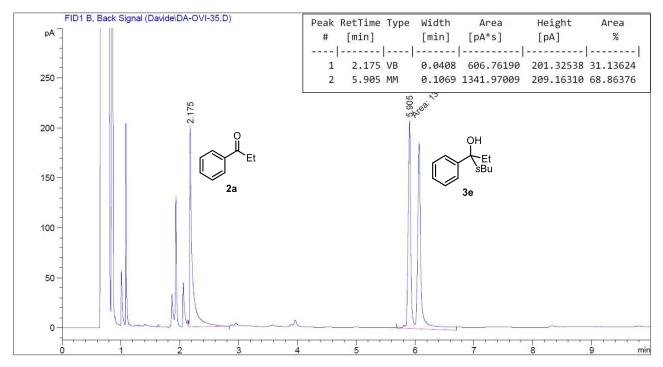


Figure S27. Table S3, entry 3 (R-M: *s*-BuLi): GC-FID chromatogram of the reaction crude.

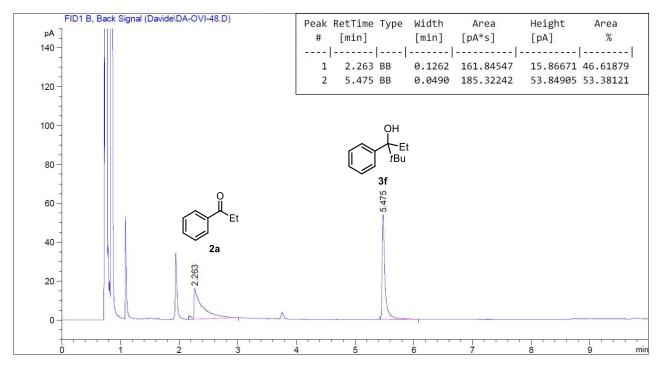


Figure S28. Table S3, entry 4 (R-M: t-BuLi): GC-FID chromatogram of the reaction crude.

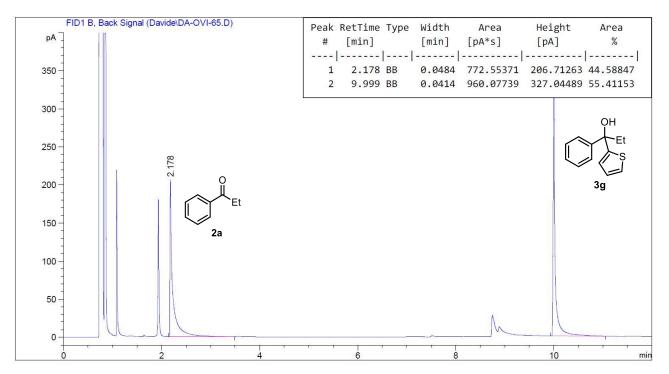


Figure S29. Table S3, entry 5 (R-M: 2-ThienLi): GC-FID chromatogram of the reaction crude.

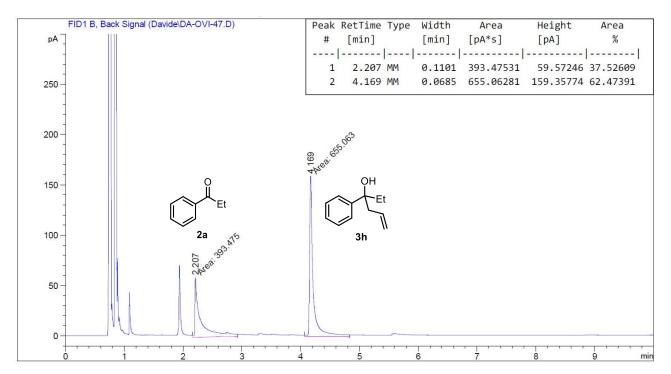


Figure S30. Table S3, entry 6 (R-M: AllylMgBr): GC-FID chromatogram of the reaction crude.

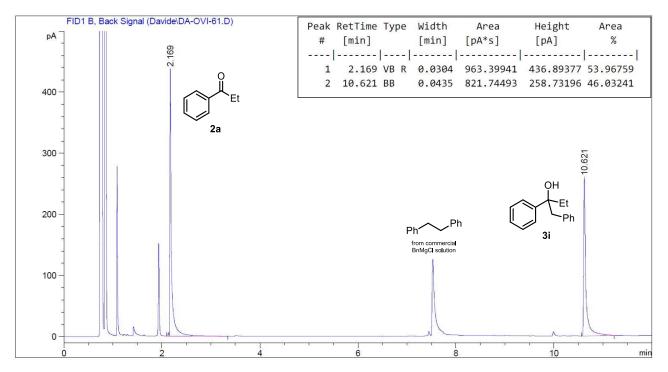


Figure S31. Table S3, entry 7 (R-M: BnMgCl): GC-FID chromatogram of the reaction crude.



**2-phenylbutan-2-ol (3c):** flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product **3c** as a colorless oil (82%,  $R_f = 0.16$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.51-7.42 (m, 2H), 7.41-7.32 (m, 2H), 7.30-7.22 (m, 1H), 1.87 (qd, J = 7.3, 3.8 Hz, 2H), 1.77 (br s, 1H), 1.58 (s, 3H), 0.82 (t, J = 7.4 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  147.9, 128.2, 126.6, 125.0, 75.1, 36.8, 29.8, 8.4.<sup>7</sup>



**3-phenylpentan-3-ol (3d):** flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product **3d** as a colorless oil (66%,  $R_f = 0.22$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.47-7.32 (m, 4H), 7.30-7.20 (m, 1H), 2.00-1.77 (m, 4H), 1.69 (br s, 1H), 0.80 (t, J = 7.4 Hz, 6H). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  145.7, 127.9, 126.2, 125.4, 77.3, 34.9, 7.7.<sup>8</sup>



**4-methyl-3-phenylhexan-3-ol (3e):** flash column chromatography (hexane/Et<sub>2</sub>O 95/5 v/v) gave product **3e** as a colorless oil (64%,  $R_f = 0.21$  hexane/Et<sub>2</sub>O 95/5 v/v). Mixture of diastereomers ( $d_r = 1:1$ ). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>, mixture of diastereoisomers):  $\delta$  7.46-7.31 (m, 8H), 7.30-7.20 (m, 2H), 1.95 (dq, J = 14.4, 7.4 Hz, 4H), 1.86-1.68 (m, 2H), 1.59 (br s, 2H), 1.44-1.23 (m, 2H), 1.05-0.64 (m, 18H). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>, mixture of diastereoisomers):  $\delta$  144.7, 144.4, 127.0, 126.9, 125.3, 125.2, 125.1, 79.1, 78.9, 44.0, 43.8, 31.3, 31.0, 23.3, 22.4, 12.9, 11.9, 11.8, 7.1.<sup>7</sup>



**2,2-dimethyl-3-phenylpentan-3-ol (3f):** flash column chromatography (hexane/Et<sub>2</sub>O 95/5 v/v) gave product **3f** as a colorless oil (40%,  $R_f = 0.26$  hexane/Et<sub>2</sub>O 95/5 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.38 (d, J = 7.7 Hz, 2H), 7.33-7.28 (m, 2H), 7.24-7.19 (m, 1H), 2.23 (dq, J = 14.8, 7.4 Hz, 1H), 1.87 (dq, J = 14.5, 7.3 Hz, 1H), 1.68 (s, 1H), 0.91 (s, 9H), 0.68 (t, J = 7.4 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  143.0, 127.9, 127.2, 126.3, 81.4, 38.5, 27.0, 26.0, 8.3.<sup>9</sup>



**1-phenyl-1-(thiophen-2-yl)propan-1-ol (3g):** flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product **3g** as a pale yellow oil (46%,  $R_f = 0.23$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.51 (d, J = 7.0 Hz, 2H), 7.42-7.21 (m, 4H), 7.00-6.89 (m, 2H), 2.38 (q, J = 7.5 Hz, 2H) superimposed to 2.33 (s, 1H), 0.94 (t, J = 7.3 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  153.2, 145.7, 128.2, 127.2, 126.7, 125.9, 124.8, 124.1, 77.5, 36.5, 8.4.<sup>10</sup>



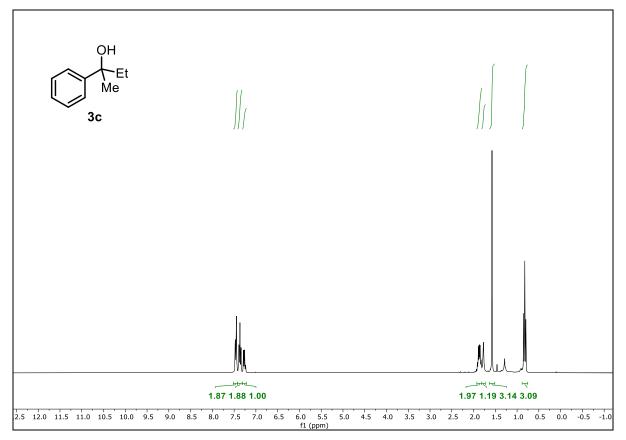
**3-phenylhex-5-en-3-ol (3h):** flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product **3h** as a colorless oil (50%,  $R_f = 0.23$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.41 (d, J = 7.1 Hz, 2H), 7.36 (t, J = 7.6 Hz, 2H), 7.27-7.22 (m, 1H), 5.65-5.53 (m, 1H), 5.19-5.07 (m, 2H), 2.74 (dd, J = 13.9, 6.0 Hz, 1H), 2.52 (dd, J = 13.8, 8.6 Hz, 1H), 1.99 (br s, 1H), 1.92-1.80 (m, 2H), 0.79 (t, J = 7.4 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  145.3, 133.2, 127.6, 126.0, 125.0, 119.1, 75.5, 46.5, 34.8, 7.4.<sup>11</sup>



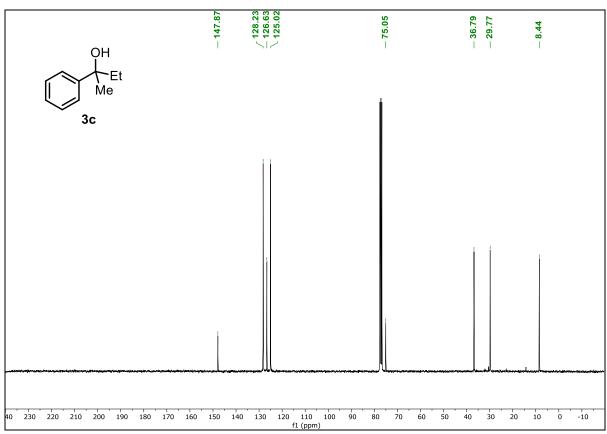
**1,2-diphenylbutan-2-ol (3i):** flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product **3i** as a colorless oil (33%,  $R_f = 0.25$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.37-7.31 (m, 4H), 7.27-7.23 (m, 1H), 7.22-7.19 (m, 3H), 7.00-6.95 (m, 2H), 3.18 (d, J = 13.4 Hz, 1H), 3.08 (d, J = 13.3 Hz, 1H), 2.01 (dq, J = 14.8, 7.5 Hz, 1H), 1.85 (dq, J = 14.5, 7.3 Hz, 1H), 1.76 (br s, 1H), 0.79 (t, J = 7.4 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  145.1, 136.1, 130.3, 127.7, 127.6, 126.3, 126.1, 125.2, 76.6, 49.1, 34.1, 7.5.<sup>12</sup>

# 2-phenylbutan-2-ol (3c)

<sup>1</sup>**H NMR** (300 MHz, CDCl<sub>3</sub>)

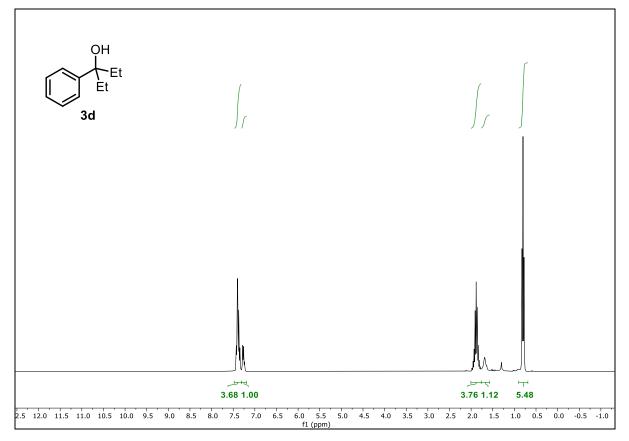


<sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>)

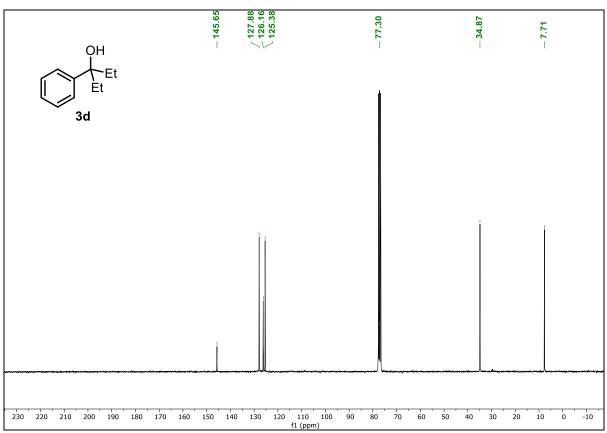


# 3-phenylpentan-3-ol (3d)

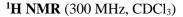


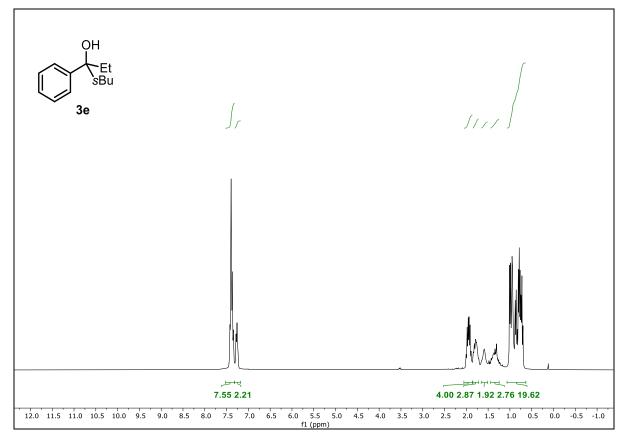


<sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>)

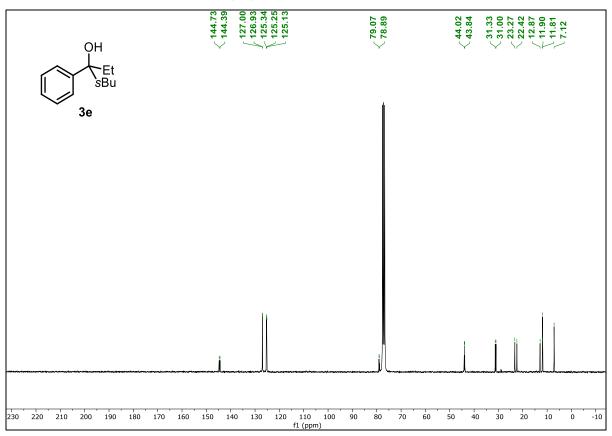


# 4-methyl-3-phenylhexan-3-ol (3e)

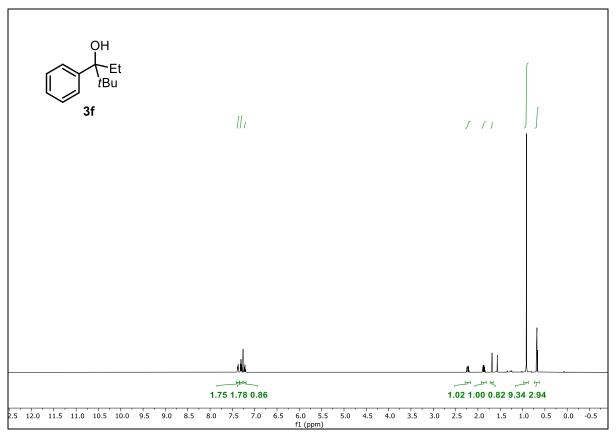




<sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>)

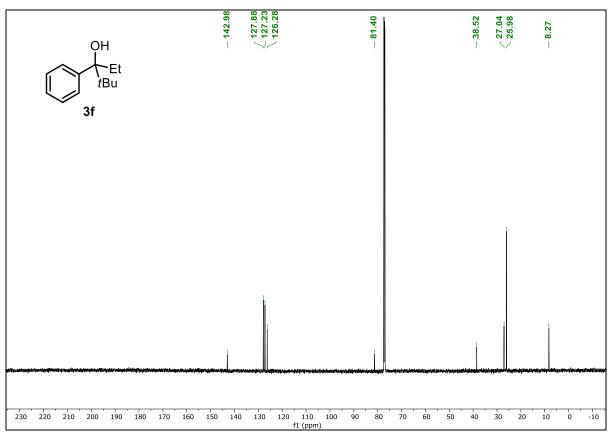


### 2,2-dimethyl-3-phenylpentan-3-ol (3f)

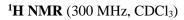


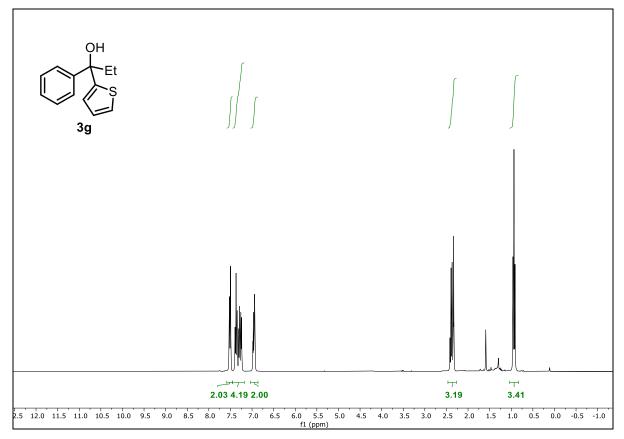
#### <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)

<sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)

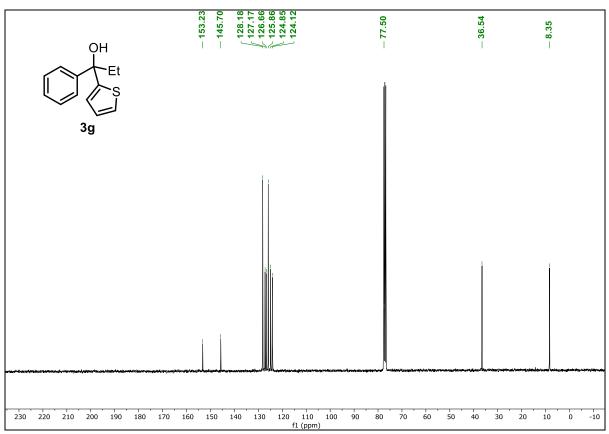


## 1-phenyl-1-(thiophen-2-yl)propan-1-ol (3g)



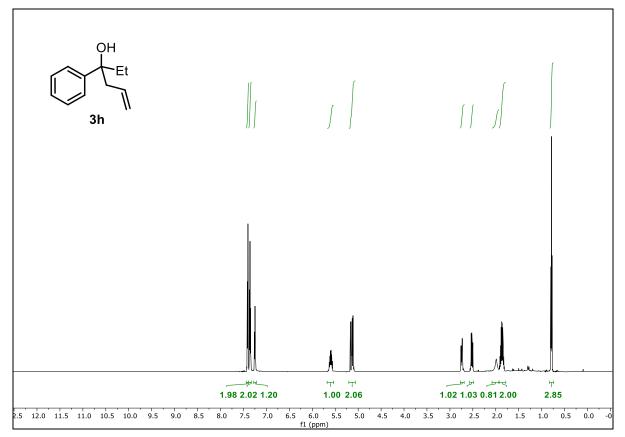


<sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>)

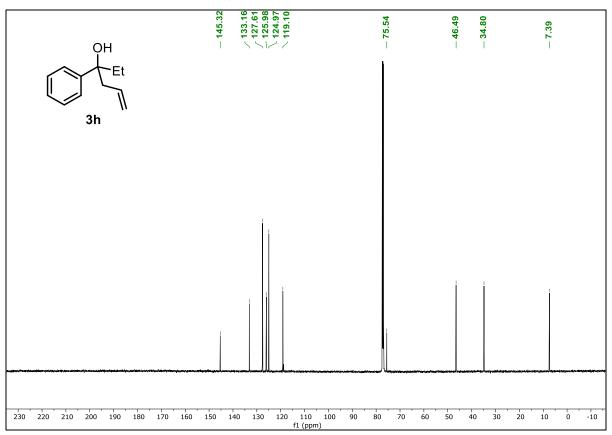


#### 3-phenylhex-5-en-3-ol (3h)



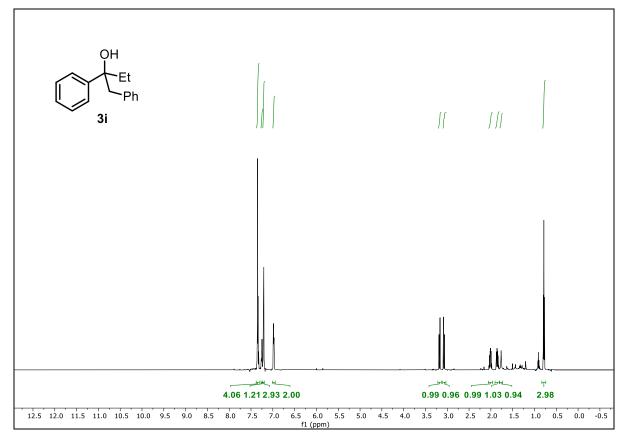


<sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)

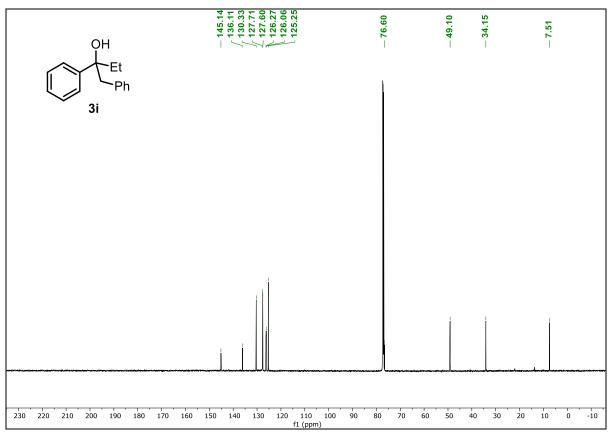


#### 1,2-diphenylbutan-2-ol (3i)





#### <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)



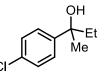
Hybrid one-pot tandem transformation of ketoximes **1a-j** into tertiary alcohols **3c,h,j-x** promoted by combination of the laccase/TEMPO/O<sub>2</sub> system with the chemoselective addition of MeLi/AllylMgBr in aqueous media, at room temperature and in the presence of air

*T. versicolor* laccase (280 mg, 0.5 U/mg) and TEMPO (38 mg, 33 mol%) were added to a 0.73 mmol (109 mg) suspension of the corresponding ketoxime **1a-j** in water (1 mL) and the mixture was stirred vigorously (1200 rpm) in an 8 mL vial under oxygen atmosphere for 24 h. Then 1 mL of CPME was added as co-solvent to form a biphasic reaction medium. Next, MeLi (3.0 eq, 1.6 M in Et<sub>2</sub>O) or AllylMgBr (3.0 eq, 1.0 M in Et<sub>2</sub>O) was rapidly spreaded over the reaction mixture at room temperature, under air. After 3 s, a saturated solution of NH<sub>4</sub>Cl<sub>aq</sub>(2.5 mL) was added, and the mixture was extracted with dichloromethane (3 x 5 mL). The combined organic phases were washed with brine (1 x 5 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and the solvents were removed *in vacuo*. The crude tertiary alcohols **3c,h,j-x** products obtained were purified by flash column chromatography and characterized by <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR.

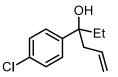
**Table S4**. Hybrid one-pot tandem transformation of ketoximes **1a-i** into tertiary alcohols **3c,h,j-x** promoted by combination of the laccase/TEMPO/O<sub>2</sub> system with the chemoselective addition of MeLi/AllylMgBr in aqueous media, at room temperature and in the presence of air.<sup>*a*</sup>

(1a-j)	N <sup>°</sup> OH    33 mol% TEMPC    R <sup>2</sup> H <sub>2</sub> O / rt / 24 h		(2a-j)	R <sup>3</sup> ·M O:CPME / 10 sec. , R <sup>3</sup> = Me; M = I	(3c,h,j-x)
Entry	$\mathbb{R}^1$	R <sup>2</sup>	R <sup>3</sup> -M	Product	Yield <sup>b</sup> (%)
1	H ( <b>1a</b> )	Et	MeLi	3c	82
2	H ( <b>1a</b> )	Et	AllylMgBr	3h	50
3	<i>p</i> -Cl ( <b>1b</b> )	Et	MeLi	3ј	83
4	<i>p</i> -Cl ( <b>1b</b> )	Et	AllylMgBr	3k	58
5	<i>p</i> -OMe ( <b>1c</b> )	Et	MeLi	31	72
6	<i>p</i> -OMe ( <b>1c</b> )	Et	AllylMgBr	3m	52
7	<i>p</i> -Me (1d)	Et	MeLi	3n	54
8	<i>p</i> -Me (1d)	Et	AllylMgBr	30	36
9	H ( <b>1e</b> )	Me	MeLi	3p	74
10	H ( <b>1e</b> )	Me	AllylMgBr	3q	43
11	<i>p</i> -Cl ( <b>1f</b> )	Me	MeLi	3r	78
12	<i>m</i> -Cl ( <b>1g</b> )	Me	MeLi	<b>3</b> s	29
13	<i>o</i> -Cl ( <b>1h</b> )	Me	MeLi	3t	11
14	<i>p</i> -Cl ( <b>1f</b> )	Me	AllylMgBr	3u	16
13	<i>m</i> -OMe (1i)	Me	MeLi	3v	40
14	<i>m</i> -OMe (1i)	Me	AllylMgBr	3w	28
15	<i>p</i> -OMe ( <b>1i</b> )	Me	MeLi	3x	88

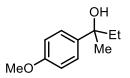
<sup>*a*</sup> General conditions: 24 h of reaction at room temperature and at 1800 rpm; Laccase from *T. Versicolor* (0.5 U/mg, 280 mg) per 0.73 mmol of **1a-i**, 0.33 eq. TEMPO in 1 mL of water were used. Then 1 mL of CPME and MeLi (3.0 eq, 1.6 M in Et<sub>2</sub>O) or AllylMgBr (3.0 eq, 1.0 M in Et<sub>2</sub>O) reagents were added without any isolation/purification. <sup>*b*</sup> Isolated yield.



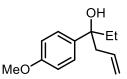
**2-(4-chlorophenyl)butan-2-ol (3j):** flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product **3j** as a colorless oil (83%,  $R_f = 0.21$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.37 (d, J = 8.6 Hz, 2H) superimposed to 7.30 (d, J = 8.6 Hz, 2H), 1.90-1.75 (m, 2H) superimposed to 1.72 (br s, 1H), 1.54 (s, 3H), 0.80 (t, J = 7.4 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  146.2, 132.2, 128.1, 126.4, 74.6, 36.6, 29.7, 8.2.<sup>13</sup>



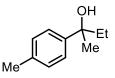
**3-(4-chlorophenyl)hex-5-en-3-ol (3k):** flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product **3k** as a colorless oil (58%,  $R_f = 0.20$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.40 (d, J = 8.7 Hz, 2H) superimposed to 7.37 (d, J = 8.7 Hz, 2H), 5.70-5.56 (m, 1H), 5.25-5.14 (m, 2H), 2.75 (dd, J = 13.8, 5.9 Hz, 1H), 2.55 (dd, J = 13.7, 8.4 Hz, 1H), 2.00-1.81 (m, 3H), 0.83 (t, J = 7.2 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  143.2, 132.0, 131.1, 127.0, 125.9, 118.8, 74.6, 45.8, 34.1, 6.6.<sup>14</sup>



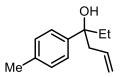
**2-(4-methoxyphenyl)butan-2-ol (3l):** flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product **3l** as a colorless oil (72%,  $R_f = 0.19$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.35 (d, J = 8.8 Hz, 2H), 6.87 (d, J = 8.8 Hz, 2H), 3.81 (s, 3H), 1.82 (qd, J = 7.3, 2.4 Hz, 2H), 1.70 (br s, 1H), 1.53 (s, 3H), 0.79 (t, J = 7.4 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  158.3, 140.0, 126.2, 113.5, 74.7, 55.3, 36.8, 29.7, 8.5.<sup>15</sup>



**3-(4-methoxyphenyl)hex-5-en-3-ol (3m):** flash column chromatography (hexane/EtOAc 95/5 v/v) gave product **3m** as a colorless oil (35%, Rf = 0.11 hexane/EtOAc 95/5 v/v). <sup>1</sup>H NMR (300 MHz, CDCl3):  $\delta$  7.32 (d, *J* = 8.7 Hz, 2H), 6.89 (d, *J* = 8.7 Hz, 2H), 5.68-5.54 (m, 1H), 5.19-5.08 (m, 2H), 3.83 (s, 3H), 2.72 (dd, *J* = 13.5, 6.0 Hz, 1H), 2.49 (dd, *J* = 13.5, 6.0 Hz, 1H), 1.90-1.77 (m, 2H), 1.61 (br s, 1H), 0.78 (t, *J* = 7.5 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl3):  $\delta$  158.1, 137.9, 133.8, 126.6, 119.4, 113.3, 75.8, 55.2, 46.9, 35.3, 7.9.<sup>16</sup>



**2-**(*p*-tolyl)butan-2-ol (3n): flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product 3n as a colorless oil (54%,  $R_f = 0.18$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  7.32 (d, *J* = 8.1 Hz, 2H), 7.15 (d, *J* = 7.9 Hz, 2H), 2.34 (s, 3H), 1.83 (qd, *J* = 7.2, 3.2 Hz, 2H), 1.69 (br s, 1H), 1.54 (s, 3H), 0.80 (t, *J* = 7.4 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  145.0, 136.2, 128.9, 125.0, 74.9, 36.8, 29.8, 21.1, 8.5.<sup>14</sup>



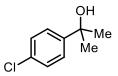
**3-**(*p*-tolyl)hex-5-en-3-ol (30): flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product 30 as a colorless oil (36%,  $R_f = 0.24$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.29-7.24 (m, 2H), 7.15 (d, J = 7.8 Hz, 2H), 5.66-5.51 (m, 1H), 5.17-5.04 (m, 2H), 2.74-2.64 (m, 1H), 2.53-2.44 (m, 1H), 2.34 (s, 3H), 1.97 (br s, 1H), 1.89-1.76 (m, 2H), 0.77 (t, J = 7.4 Hz, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  142.9, 136.0, 133.9, 128.9, 125.5, 119.5, 76.0, 47.0, 35.4, 21.1, 8.0.<sup>17</sup>



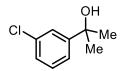
**2-phenylpropan-2-ol (3p):** flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product **3p** as a colorless oil (74%,  $R_f = 0.21$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.51-7.48 (m, 2H), 7.37-7.32 (m, 2H), 7.27-7.23 (m, 1H), 1.79 (br s, 1H), 1.60 (s, 6H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  149.2, 128.4, 126.8, 124.5, 72.7, 31.9.<sup>18</sup>



**2-phenylpent-4-en-2-ol (3q):** flash column chromatography (hexane/Et<sub>2</sub>O 9/1 v/v) gave product **3q** as a colorless oil (43%,  $R_f = 0.25$  hexane/Et<sub>2</sub>O 9/1 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.47-7.42 (m, 2H), 7.37-7.32 (m, 2H), 7.27-7.22 (m, 1H), 5.63 (dddd, J = 16.9, 10.3, 8.3, 6.4 Hz, 1H), 5.17-5.08 (m, 2H), 2.69 (dd, J = 13.7, 6.5 Hz, 1H), 2.51 (dd, J = 13.6, 8.4 Hz, 1H), 2.06 (br s, 1H), 1.56 (s, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  147.7, 133.8, 128.3, 126.7, 124.9, 119.6, 73.7, 48.6, 30.0.<sup>19</sup>



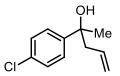
**2-(4-chlorophenyl)propan-2-ol (3r):** flash column chromatography (hexane/EtOAc 9/1 v/v) gave product **3r** as a colorless oil (78%,  $R_f = 0.25$  hexane/EtOAc 9/1 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.41 (d, J = 8.0 Hz, 2H), 7.29 (d, J = 8.0 Hz, 2H), 1.56 (s, 6H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  147.7, 132.5, 128.3, 125.9, 72.2, 31.8.<sup>15</sup>



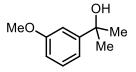
**2-(3-chlorophenyl)propan-2-ol (3s):** flash column chromatography (hexane/EtOAc 9/1 v/v) gave product **3s** as a colorless oil (29%,  $R_f = 0.25$  hexane/EtOAc 9/1 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.49 (t, J = 1.6 Hz, 20 1H), 7.36 (d, J = 8.0 Hz, 1H), 7.27 (t, J = 8.0 Hz, 1H), 7.22 (d, J = 8.0 Hz, 1H), 1.57 (s, 6H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  151.4, 134.9, 129.4, 126.7, 124.9, 122.7, 72.2, 31.6.<sup>15</sup>



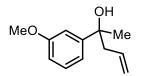
**2-(2-chlorophenyl)propan-2-ol (3t):** flash column chromatography (hexane/EtOAc 9/1 v/v) gave product **3t** as a colorless oil (11%,  $R_f = 0.25$  hexane/EtOAc 9/1 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.69 (d, J = 7.5 Hz, 1H), 7.38 (d, J = 7.5 Hz, 1H), 7.28 (t, J = 7.5 Hz, 1H), 7.21 (t, J = 7.5 Hz, 1H), 2.68 (brs, 1H), 1.76 (s, 6H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  144.8, 131.3, 131.2, 128.1, 126.8, 72.9, 29.3.<sup>15</sup>



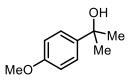
**2-(4-chlorophenyl)pent-4-en-2-ol (3u):** flash column chromatography (hexane/EtOAc 8/2 v/v) gave product **3u** as a pale yellow oil (16%,  $R_f = 0.26$  hexane/EtOAc 8/2 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.40–7.34 (m, 2H), 7.32–7.27 (m, 2H), 5.62–5.58 (m, 1H), 5.18–5.08 (m, 2H), 2.63 (dt, J = 18.6, 9.3 Hz, 1H), 2.48 (dd, J = 13.8, 8.2 Hz, 1H), 2.23–2.08 (m, 1H), 1.52 (s, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  146.2, 133.3, 132.2, 128.3, 126.3, 119.8, 73.5, 48.5, 30.1.<sup>20</sup>



**2-(3-methoxyphenyl)propan-2-ol (3v):** flash column chromatography (hexane/EtOAc 8/2 v/v) gave product **3v** as a colorless oil (40%,  $R_f = 0.21$  hexane/EtOAc 8/2 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.27 (d, J = 8.0 Hz, 1H), 7.07 (t, J = 8.0 Hz, 2H), 6.80 (dd, J = 8.0, 2.4 Hz, 1H), 3.82 (s, 3H), 1.58 (s, 6H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  159.6, 150.9, 129.2, 116.8, 111.8, 110.6, 72.4, 55.2, 31.6.<sup>15</sup>

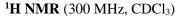


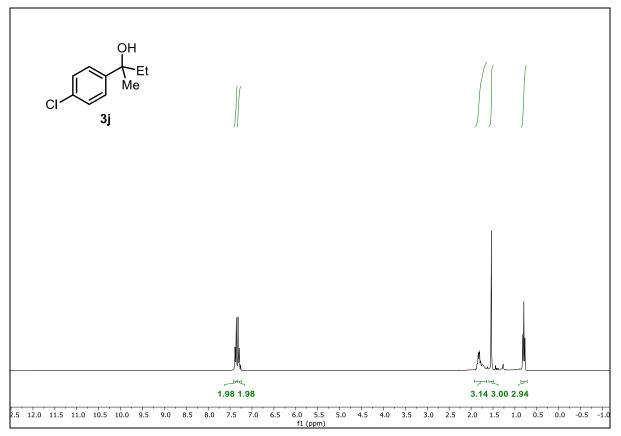
**2-(3-methoxy-phenyl)-pent-4-en-2-ol (3w):** flash column chromatography (hexane/EtOAc 8/2 v/v) gave product **3w** as a colorless oil (28%,  $R_f = 0.21$  hexane/EtOAc 8/2 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.26 (t, J = 8.0 Hz, 1H), 7.01 (t, J = 8.0 Hz, 2H), 6.78 (t, J = 4.0 Hz, 1H), 5.67–5.57 (m, 1H), 5.16–5.11 (m, 2H), 3.83 (s, 3H), 2.69 (q, J = 6.8 Hz, 1H), 2.48 (q, J = 6.8 Hz, 1H), 2.05 (s, br, 1H), 1.53 (s, 3H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>):  $\delta$  159.9, 149.5, 133.6, 129.1, 119.6, 117.2, 111.7, 110.5, 73.5, 55.3, 48.4, 29.9.<sup>20</sup>



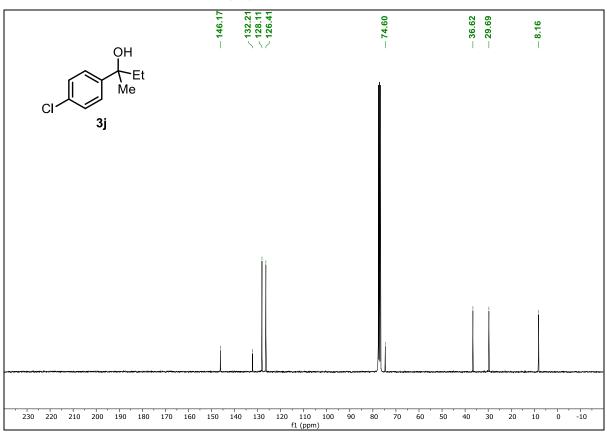
**2-(4-Methoxyphenyl)propan-2-ol (3x):** colorless oil (88%,  $R_f = 0.26$  hexane/AcOEt 8/2 v/v). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  7.41 (d, J = 6.0 Hz, 2H), 6.87 (d, J = 6.0 Hz, 2H), 3.80 (s, 3H), 1.56 (s, 6H). <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>): 154.4, 137.4, 121.6, 109.5, 68.2, 51.3, 27.9, 27.8.<sup>8</sup>

## 2-(4-chlorophenyl)butan-2-ol (3j)



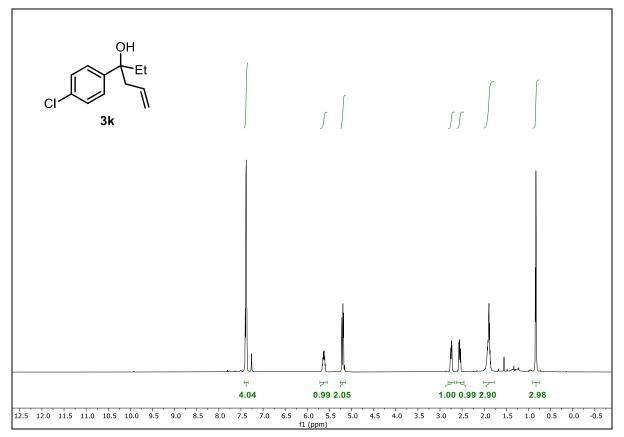


<sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>)

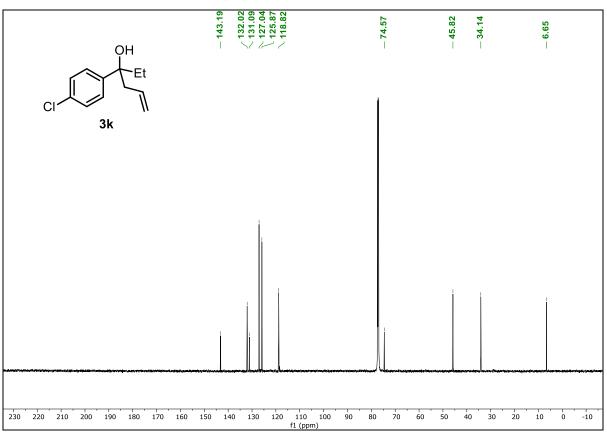


#### 3-(4-chlorophenyl)hex-5-en-3-ol (3k)

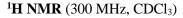


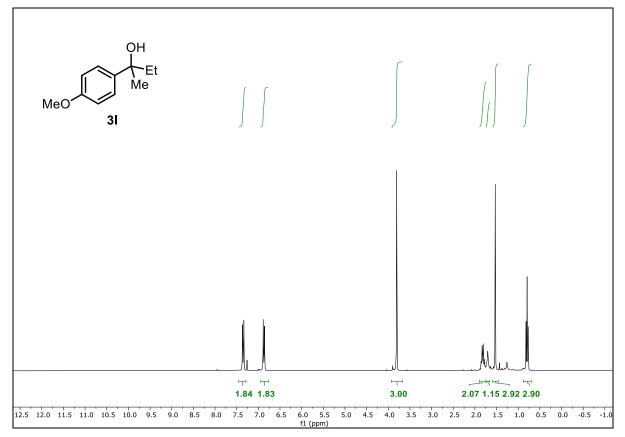


<sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)

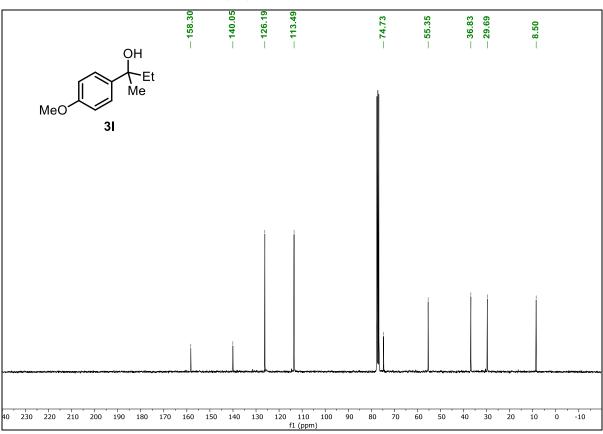


## 2-(4-methoxyphenyl)butan-2-ol (3l)

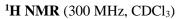


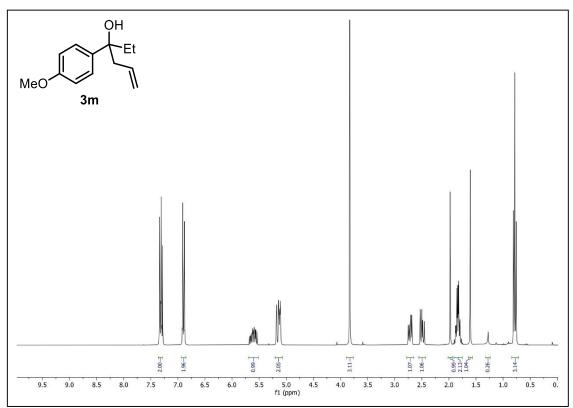


<sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>)

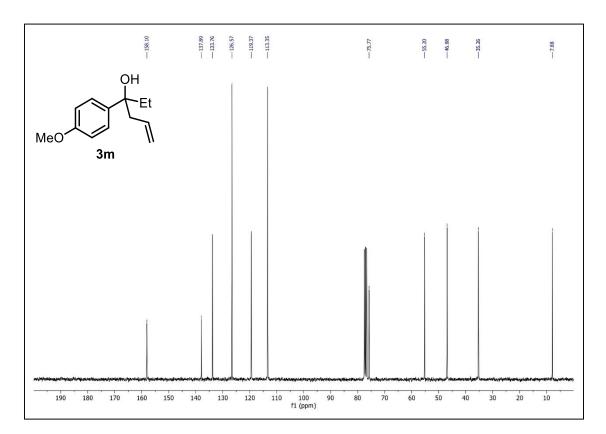


## 3-(4-methoxyphenyl)hex-5-en-3-ol (3m)



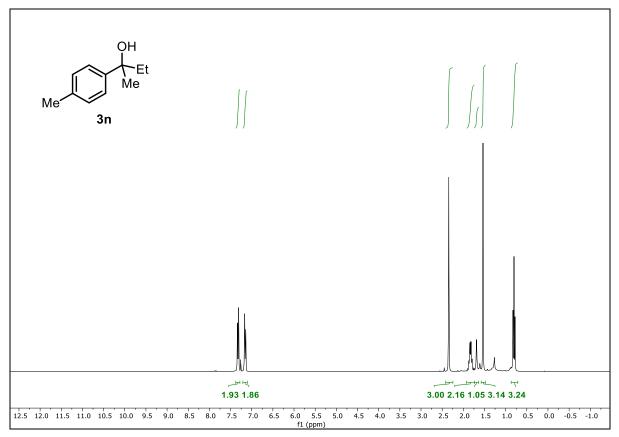


<sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>)

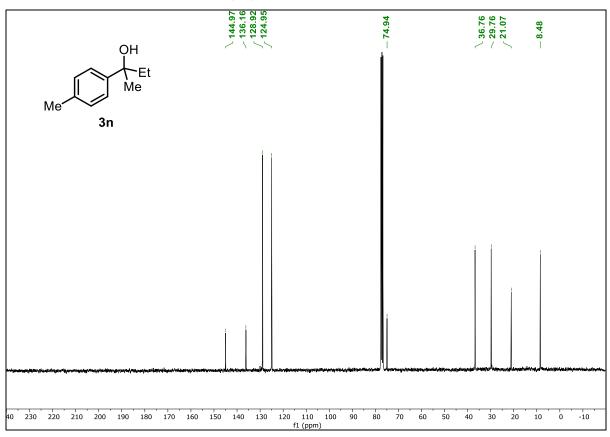


#### 2-(p-tolyl)butan-2-ol (3n)



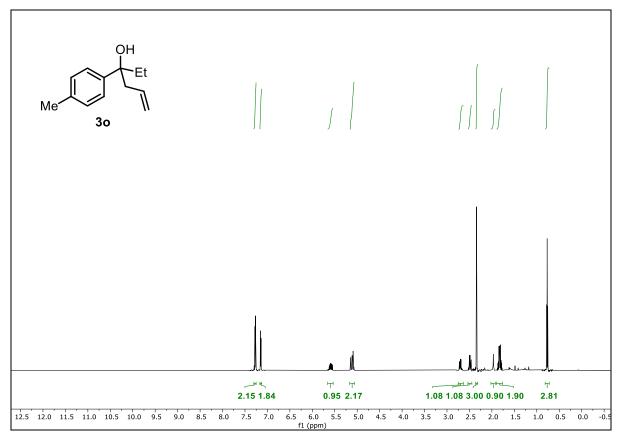


## <sup>13</sup>C{<sup>1</sup>H} NMR (75 MHz, CDCl<sub>3</sub>)

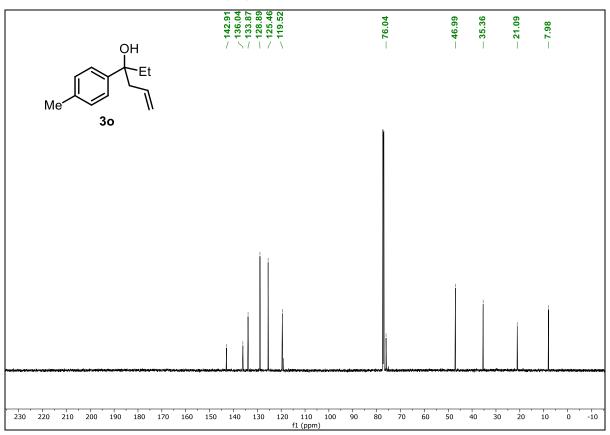


#### 3-(*p*-tolyl)hex-5-en-3-ol (30)

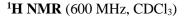
<sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)

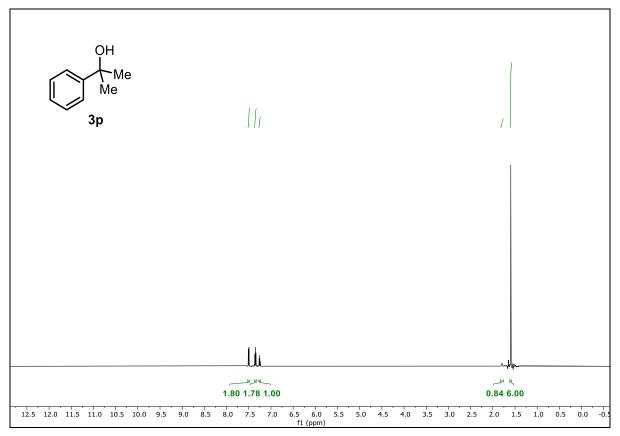


<sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)

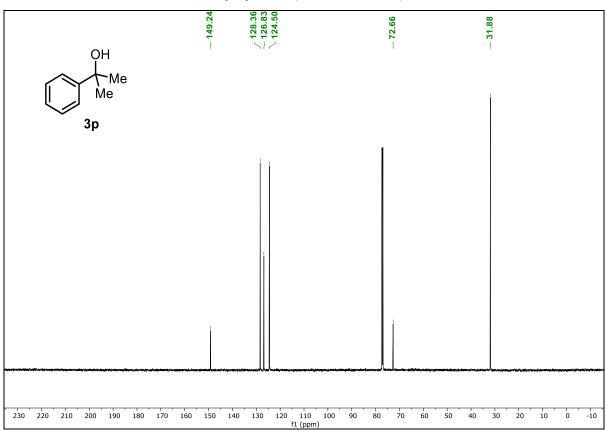


## 2-phenylpropan-2-ol (3p)

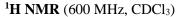


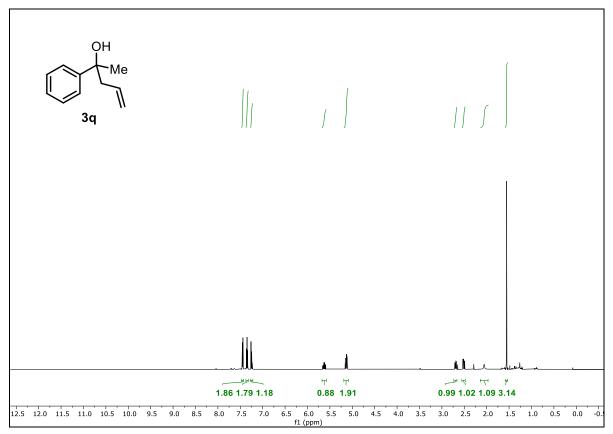


<sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)

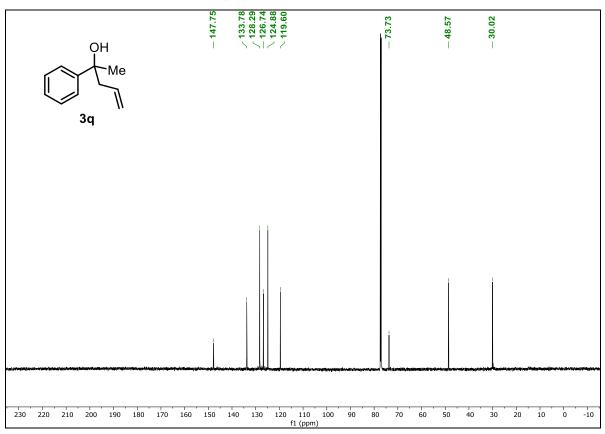


#### 2-phenylpent-4-en-2-ol (3q)

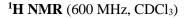


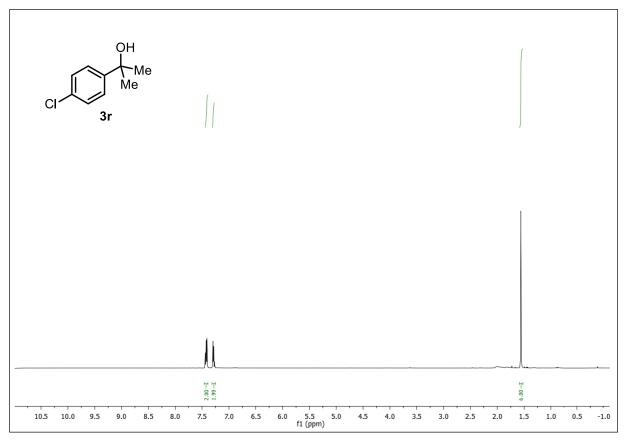


## <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)

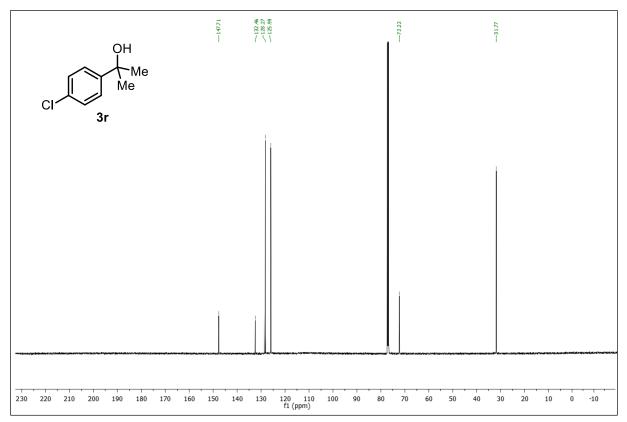


## 2-(4-chlorophenyl)propan-2-ol (3r)

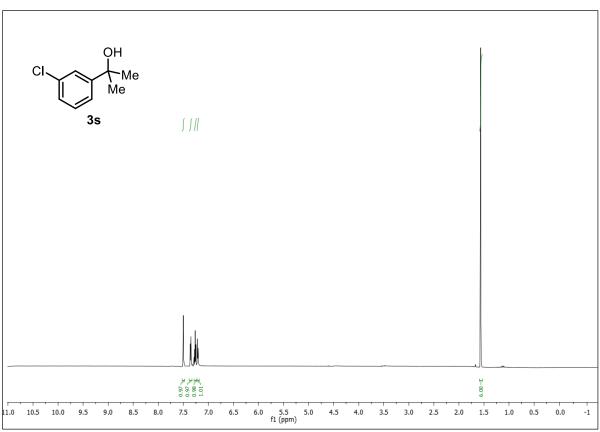




<sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)

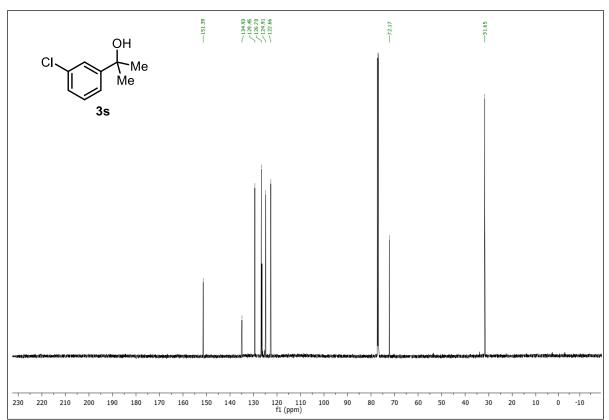


## 2-(3-chlorophenyl)propan-2-ol (3s)

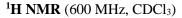


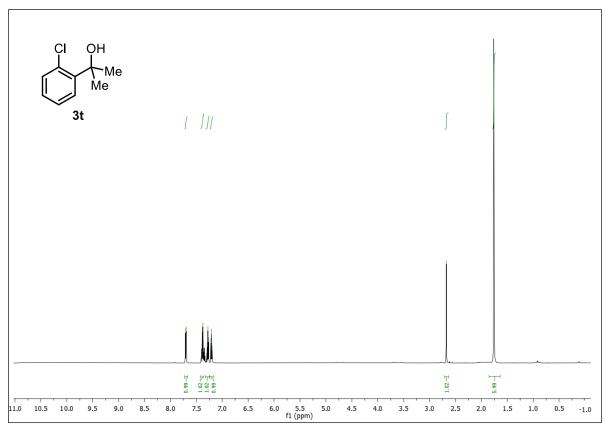
#### <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)

#### <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)

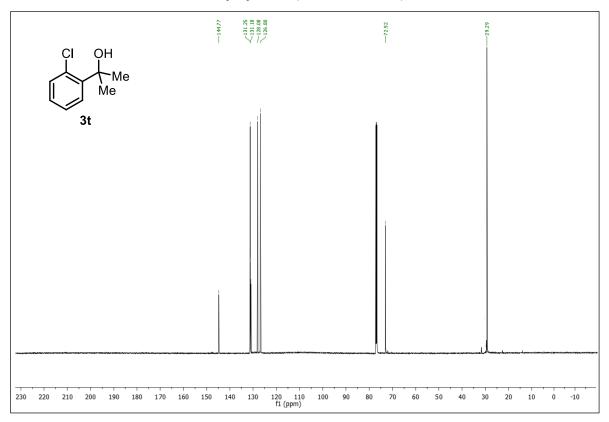


## 2-(2-chlorophenyl)propan-2-ol (3t)

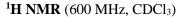


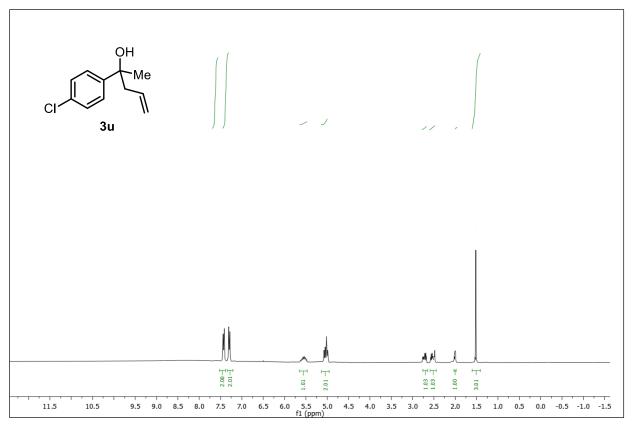


# <sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)

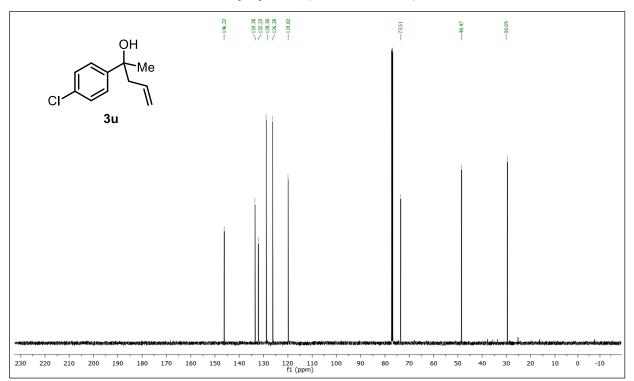


#### 2-(4-chlorophenyl)pent-4-en-2-ol (3u)

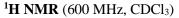


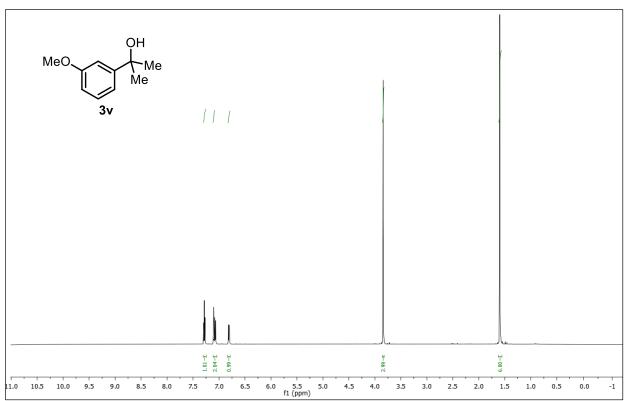


<sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)

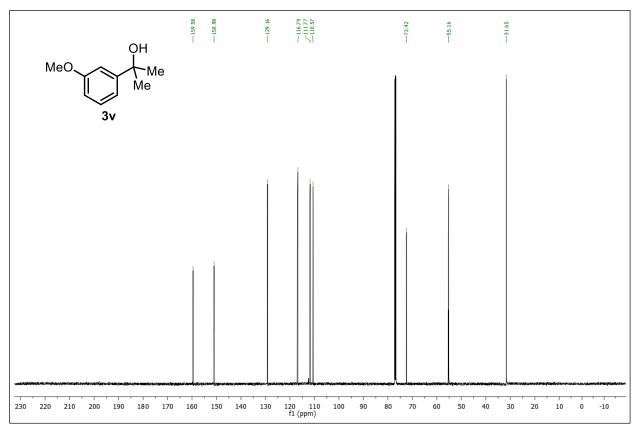


## 2-(3-methoxyphenyl)propan-2-ol (3v)

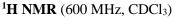


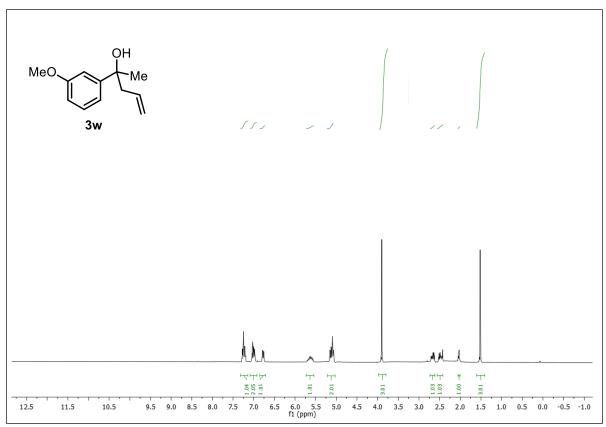


<sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)

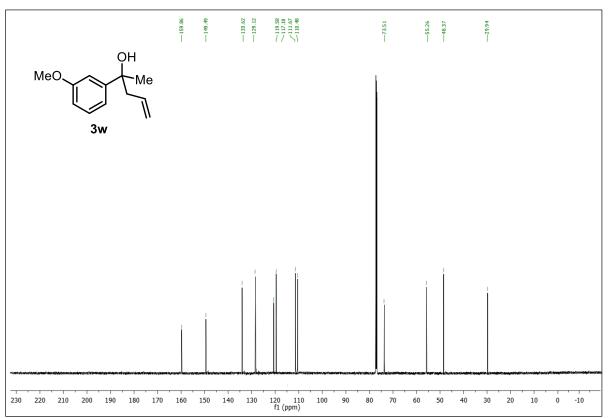


#### 2-(3-methoxy-phenyl)-pent-4-en-2-ol (3w)

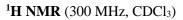


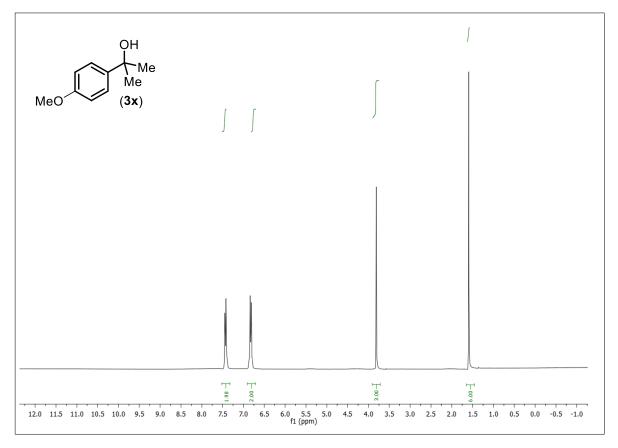


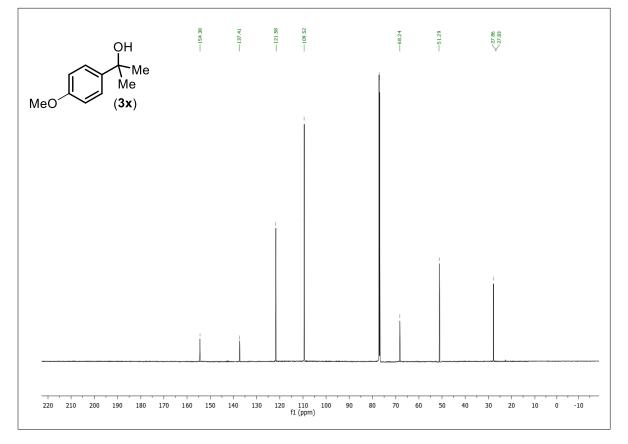
<sup>13</sup>C{<sup>1</sup>H} NMR (150 MHz, CDCl<sub>3</sub>)



## 2-(4-methoxyphenyl)propan-2-ol (3x)







#### References

[1] (a) S. C. Watson and J. F. Eastham, *J. Organomet. Chem.*, 1967, **9**, 165; (b) H.-S. Lin and L. A. Paquette, *Synth. Commun.*, 2007, **24**, 2503.

[2] A. Krasovskiy and P. Knochel, Synthesis, 2006, 5, 890.

[3] C. Ramalingan and Y.-T. Park, J. Org. Chem., 2007, 72 (12), 4536.

[4] C.L.F. Meyers and D.J. Meyers, *Current Protocols in Nucleic Acid Chemistry*, Thin-Layer Chromatography, John Wiley & Sons, Inc., Hoboken, 2008.

[5] S. Abedi, B. Karimi, F. Kazemi, M. Bostina and H. Vali, *Org. Biomol. Chem.*, 2013, **11** (3), 416.

[6] J.-C. Wu, L.-B. Gong, Y. Xia, R.-J. Song, Y.-X. Xie and J.-H. Li, *Angew. Chem. Int. Ed.*, 2012, **51** (39), 9909.

[7] D. Elorriaga, M. J. Rodríguez-Álvarez, N. Ríos-Lombardía, F. Morís, A. Presa Soto, J. González-Sabín, E. Hevia and J. García-Álvarez, *Chem. Commun.*, 2020, **56**, 8932.

[8] A. F. Quivelli, G. D'Addato, P. Vitale, J. García-Alvarez, F. M. Perna and V. Capriati *Tetrahedron*, 2021, **81**, 131898.

[9] S. Andersson and T. Drakenberg, Org. Magn. Reson., 1983, 21, 730.

[10] W. E. Noland et al, J. Heterocycl. Chem., 2018, 55 (12), 2698.

[11] J. A. Read and Y. Yang, Org. Lett., 2017, 19 (13), 3346.

[12] X. Peng, Y. Hirao, S. Yabu, H. Sato, M. Higashi, T. Akai, S. Masaoka, H. Mitsunuma and M. Kanai, *J. Org. Chem.*, 2022, doi: 10.1021/acs.joc.2c00603.

[13] M. Hatano, T. Mizuno and K. Ishihara, Tetrahedron, 2011, 67 (24), 4417.

[14] D. G. Gilheany and S.E. Kavanagh, Org. Lett., 2020, 22 (21), 8198.

[15] D. Hu and X. Jiang, Green Chem., 2022, 24 (1), 124.

[16] U. Schneider and S. Kobayashi, Angew. Chem. Int. Ed., 2007, 46 (31), 5909.

[17] Y. Zhang, X. Jia and J.-X. Wang, Eur. J. Org. Chem., 2009, 18, 2983.

[18] G.-L. Chai, B. Zhu and J. Chang, J. Org. Chem., 2019, 84 (1), 120.

[19] R. Masuda, T. Yasukawa, Y. Yamashita and S. Kobayashi, J. Org. Chem., 2022, 87 (5), 3453.

[20] J. Pogula, S. Laha, B. Sreedhar, P. R. Linkhar., Adv. Synth. Catal., 2020, 362 (5), 1176.