

Electronic Supplementary Information

Identification of a Highly Promiscuous Glucosyltransferase from *Penstemon barbatus* for natural product glycodiversification

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1. Supplementary Tables

Table S1. Information of 10 candidates and 6His-MBP tag

Name.	ORF (bp)	amino acids	Molecular weight(kDa)	enzyme activity
45566	1374	457	50.9	β 1 \rightarrow 3 rhamnosyltransferase
43462	1455	484	54.2	C-4'' OH glycosyltransferase
51909	1338	445	50.4	No activity
54198	1428	475	54.4	No activity
43107	1371	456	50.8	No activity
57483	1365	454	51.3	No activity
56249	1371	456	51.5	No activity
56575	1425	474	52.1	No activity
49774	1377	458	51.8	No activity
50252	1284	427	47.3	No activity
6His-Mbp	1188	396	43.5	No activity

Table S2. Primers used in this study

Name	Sequences (5'-3')
MBP_45566_F	AATCCAATATTGGAAGTGAATGCCTGAATTACAAAAGTCATCAAAC
MBP_45566_R	GGCGCTCGAATTCGGATCCGCTACAATTCATACAAATTCTGAATAAACTATCAATG
MBP_43462_F	AATCCAATATTGGAAGTGAATGACCGCTGCAGAGAAAAACAC
MBP_43462_R	GGCGCTCGAATTCGGATCCGCTAGATTATTGTATGATCGGTTCTATCCGG
MBP_51909_F	AATCCAATATTGGAAGTGAATGGAATCAAATCTATTTTCAGAGTTCTAATG
MBP_51909_R	GGCGCTCGAATTCGGATCCGTCATTCTTCATACAAATGTTTGACAACCTGC
MBP_54198_F	AATCCAATATTGGAAGTGAATGTTGATAATTGTAAGAAAGAAACTAGCAAAG
MBP_54198_R	GGCGCTCGAATTCGGATCCGCTACTTGTCTTCATACAAAGTTTCATTAGC
MBP_43107_F	AATCCAATATTGGAAGTGGATGAATTTGATGATGAAGAAGAAGAAAATAATTTAATACC
MBP_43107_R	GGCGCTCGAATTCGGATCCGTCAAATAAATTGAGAGAAAGTAGACATTGCTGTAG
MBP_57483_F	AATCCAATATTGGAAGTGAATGGATAATCAAGGTACAACATCTGTTCTGC
MBP_57483_R	GGCGCTCGAATTCGGATCCGTTAAGCAGTAGAAGATCTAAATGATGTAATAAAATCAACC
MBP_56249_F	AATCCAATATTGGAAGTGAATGGATACGGTGATAAAGAACCTACCC
MBP_56249_R	GGCGCTCGAATTCGGATCCGTCATTAATGATGAAATATCTTTCAAAAAGAATCCAAG
MBP_56575_F	AATCCAATATTGGAAGTGAATGTCAGACTCCGGCGCACACATC
MBP_56575_R	GGCGCTCGAATTCGGATCCGTCACATTAAACCTTTGTCCAATAAATCTTTGGTG
MBP_49774_F	AATCCAATATTGGAAGTGAATGGACTACCATATTCTCAAGTAACATTCCAG
MBP_49774_R	GGCGCTCGAATTCGGATCCGTTACCCTTGACAATGCCACCAAC
MBP_50252_F	AATCCAATATTGGAAGTGAATGGGGATGGAATTGGCTCGAGGG
MBP_50252_R	GGCGCTCGAATTCGGATCCGTTACGCCTCAACCAGTTCAACCTTTC
T7	TAATACGACTCACTATAGG
T7t	CAAAAACCCCTCAAGACCCGTTTAGAGGCCCAAGGGTTATGCTAG

Table S3. Information of all tested compounds catalyzed by UGT84A95.

Type	No.	Name	Molecular formula	Molecular weight	CAS number
PhGs	1	Osmanthuside B	C ₂₉ H ₃₆ O ₁₃	592.588	94492-23-6
	2	Verbascoside	C ₂₉ H ₃₆ O ₁₅	624.587	61276-17-3
	3	Forsythoside A	C ₂₉ H ₃₆ O ₁₅	624.587	79916-77-1
	4	Forsythoside B	C ₃₄ H ₄₄ O ₁₉	756.702	81525-13-5
	5	Calceolarioside B	C ₂₃ H ₂₆ O ₁₁	478.446	105471-98-5
	6	Calceolarioside A	C ₂₃ H ₂₆ O ₁₁	478.446	84744-28-5
	7	Echinacoside	C ₃₅ H ₄₆ O ₂₀	786.728	82854-37-3
flavonoids	8	Rutin	C ₂₇ H ₃₀ O ₁₆	610.518	153-18-4
	9	Quercetin	C ₁₅ H ₁₀ O ₇	302.236	117-39-5
	10	Icariin	C ₃₃ H ₄₀ O ₁₅	676.662	489-32-7
	11	Orientin	C ₂₁ H ₂₀ O ₁₁	448.377	28608-75-5
	12	Kaempferol	C ₁₅ H ₁₀ O ₆	286.236	520-18-3
	13	Eriodictyol	C ₁₅ H ₁₂ O ₆	288.252	552-58-9
terpenoids	14	Picroside II	C ₂₃ H ₂₈ O ₁₃	512.461	39012-20-9
	15	Oleuropein	C ₂₅ H ₃₂ O ₁₃	540.514	32619-42-4
	16	Ginsenoside Re	C ₄₈ H ₈₂ O ₁₈	947.154	52286-59-6
	17	Ginsenoside Rg1	C ₄₂ H ₇₂ O ₁₄	801.013	22427-39-0
Stilbene glycosides	18	Paclitaxel	C ₄₇ H ₅₁ NO ₁₄	853.906	33069-62-4
	19	2,3,5,4'-Tetrahydroxy stilbene 2-O-β-D-glucoside	C ₂₀ H ₂₂ O ₉	406.383	82373-94-2
coumarins	20	alloysoimperatorin	C ₁₆ H ₁₄ O ₄	270.280	35214-83-6
	21	Umbelliferone	C ₉ H ₆ O ₃	162.142	93-35-6
	22	Scopoletin	C ₁₀ H ₈ O ₄	192.168	92-61-5
	23	<u>Isoscopoletin</u>	C ₁₀ H ₈ O ₄	192.168	776-86-3
	24	Esculetin	C ₉ H ₆ O ₄	178.141	305-01-1
	25	7-demethylsuberosin	C ₁₄ H ₁₄ O ₃	230.259	21422-04-8
simple polyphenols	26	Caffeic acid	C ₉ H ₈ O ₄	180.157	331-39-5
	27	ferulic acid	C ₁₀ H ₁₀ O ₄	194.184	1135-24-6
	28	Resveratrol	C ₁₄ H ₁₂ O ₃	228.243	501-36-0

Table S4. HPLC conditions used in this study.

Method	Solvent A	Solvent B	Gradient Analysis	Substrates
A	H ₂ O containing 0.1% formic acid	Acetonitrile	5% B, 5min; 5-13.2%B, 13min; 18-21.6%B, 25min; 100%B, 7min; 5%B, 7min.	1, 2, 6, 7
B	H ₂ O containing 0.1% formic acid	Acetonitrile	5% B, 5min; 5-52.5%B, 20min; 100%B, 7min; 5%B, 7min.	3, 4, 5, 8-28

Table S5. Information of large-scale reactions and purification.

Substrate (No)	reaction volumes	Reaction time(h)	gradient conditions	UV (nm)	product No. – Weight (mg)
1	1mL×15	5	18% B, 5 min; 18-19.5% B, 15 min; 100 B, 10 min; 18% B, 10min;	312	1a –5.5 1b –3.2
2	1mL×20	5	17.5% B, 5 min; 17.5-19% B, 15 min; 100 B, 10 min; 17.5% B, 10min;	312	2a –4.5 2b –1.5
3	1mL×15	5	15% B, 12 min; 100% B, 8 min; 15% B, 8 min;	312	3a –3.7
4	1mL×20	24	15% B, 12 min; 100% B, 8 min; 15% B, 8 min;	312	4a –3.3
5	1mL×15	5	16% B, 10 min; 16-16.5% B, 5 min; 100% B, 8 min; 16% B, 8 min;	312	5a –3.9
8	1mL×15	5	15% B, 12 min; 100% B, 8 min; 15% B, 8 min;	254	8a –2.8
9	1mL×30	7	16% B, 7 min; 16-24% B, 20 min; 100% B, 8 min; 16% B, 8 min;	254	9b –4.5 9c –1.2 9d –3.4
14	1mL×15	5	14% B, 10 min; 100% B, 8 min; 14% B, 8 min;	254	14a –6.5
15	1mL×40	24	18.5% B, 8min; 19% B, 13 min; 100% B, 8 min; 18.5% B, 8 min;	254	15a –6.0

Table S6. The sugar donor specificity of UGT84A95. Different substrates (**2**, **8**, **14**, **21**) were selected as acceptor substrates. “√” means that the enzymatic reactions could be catalyzed by UGT84A95.

	UDP-Glc	UDP-GlcA	UDP-Xyl	UDP-Rha	UDP-GlcNAc	UDP-Gal
2	√					
8	√		√			√
14	√					√
21	√		√			√

Table S7. Information of reported plant UGTs with characterized functions that are used for the phylogenetic analysis in this study.

Name	Source	Genbank	Reference
FaGT2	<i>Fragaria anana</i>	Q66PF4.1	1
VLR5gt	<i>Vitis labrusca</i>	ABH03018.1	2
UGT84A77	<i>Canarium album</i>	QZM06937.1	3
UGT84A23	<i>Punica granatum</i>	ANN02875.1	4
UGT84A34	<i>Nemophila menziesii</i>	BBA68563.1	5
DcUSAGT1	<i>Daucus carota</i>	AMC33112.1	6
UGT84A33	<i>Carthamus tinctorius</i>	AYW01718.1	7
UGT84A9a	<i>Gentiana triflora</i>	BAQ19550 .1	8
UGT73A4	<i>Beta vulgaris</i>	AAS94329	9
UGT84F1	<i>Medicago truncatula</i>	ABI94023.1	10
UGT84F6	<i>Glycyrrhiza uralensis</i>	QDM38904.1	11
UGT84F9	<i>Medicago truncatula</i>	XP_013470035.1	12
GgSGT	<i>Gomphrena globosa</i>	BAG14302.1	13
UGT84A9	<i>Brassica napus</i> var. <i>napus</i>	CAS03354 .1	14
UGT84A2	<i>Arabidopsis thaliana</i>	NP_188793.1	15
UGT84A1	<i>Arabidopsis thaliana</i>	Q5XF20	15
UGT84A57	<i>Eutrema japonicum</i>	BBI55602.1	16
UGT84B1	<i>Arabidopsis thaliana</i>	NP_179907.1	17
GAGT	<i>Solanum lycopersicum</i>	CAI62049.1	18
UGT74M1	<i>Nicotiana tabacum</i>	AAF61647.1	19
UGT74F2	<i>Arabidopsis thaliana</i>	NP_181910.1	20
PgUGT8	<i>Panax ginseng</i>	AIE12488.1	21
UGT74M1	<i>Gypsophila vaccaria</i>	ABK76266	22
LOC542250	<i>Zea mays</i>	AAA59054	23
UGT74H5	<i>Avena strigosa</i>	ACD03250	24
UGT74F11	<i>Oryza sativa</i> Japonica Group	5TME	25
HvUGT13248	<i>Hordeum vulgare</i> subsp. <i>vulgare</i>	ADC92550	26
Bradi5g03300	<i>Brachypodium distachyon</i>	PNT60691.1	27
UGT723F9	<i>Crocus sativus</i>	ACM66950.1	28
UGT75C1	<i>Arabidopsis thaliana</i>	AAL69494.1	29
UGT75B1	<i>Arabidopsis thaliana</i>	AEE27854	30
UGT75D1	<i>Arabidopsis thaliana</i>	AAB58497.1	15

2. Supplementary Figures

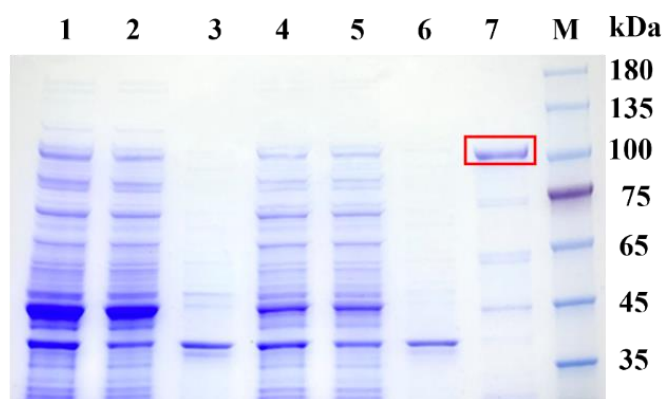


Figure S1. SDS-PAGE analysis of expression of His-MBP-UGT84A95 fusion protein.

Lane 1, 2 and 3: Cell lysate, Supernatant and Precipitation of His-MBP.

Lane 4, 5 and 6: Cell lysate, Supernatant and Precipitation of His-MBP-UGT84A95.

Lane 7: His-MBP-UGT84A95 purified by Ni-NTA affinity chromatography.

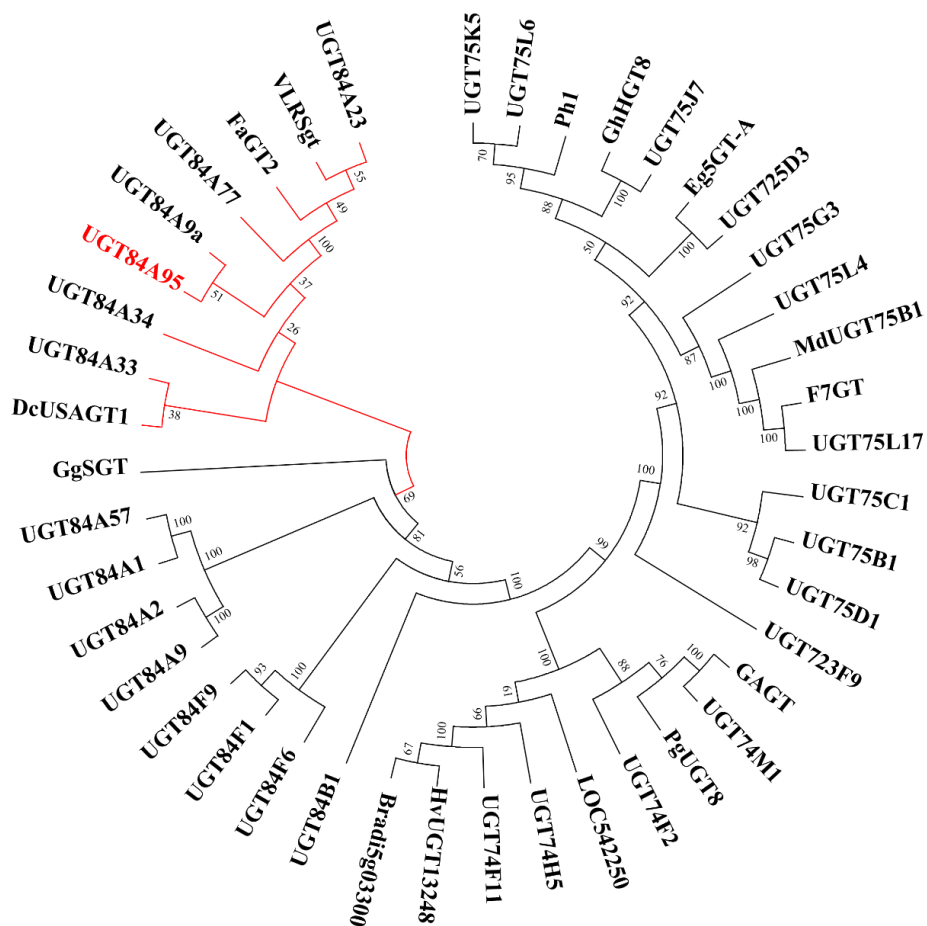


Figure S2. The phylogenetic relationships between UGT84A95 and the reported plant UGTs with characterized functions. The accession numbers of the sequences used in this study are shown in Table S6. The protein sequences were aligned using ClustalW. The neighbour-joining phylogenetic tree was drawn using MEGA 7. The bootstrap value was 1000, and the branch lengths represent the relative genetic distances.

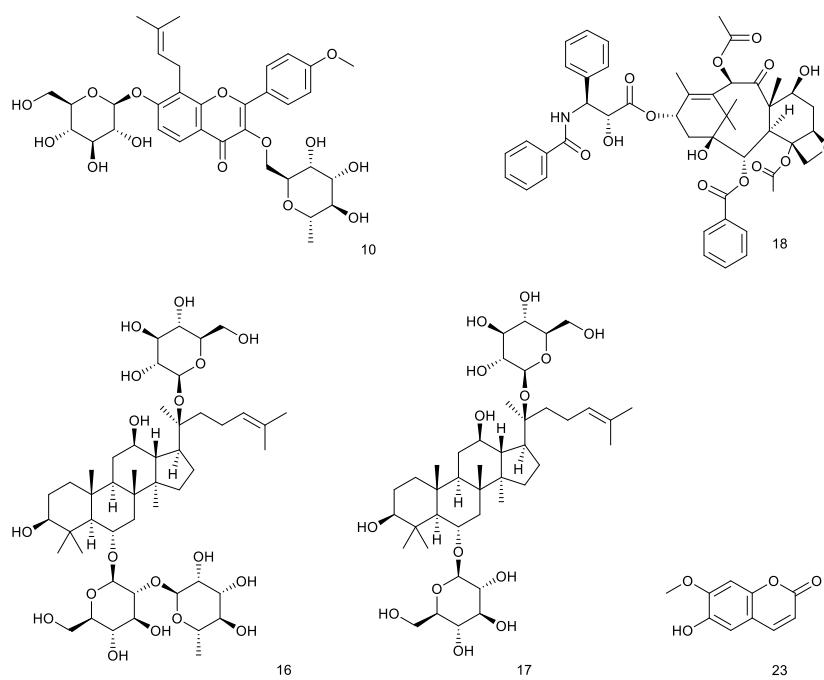


Figure S3. The compounds are not accepted by UGT84A95

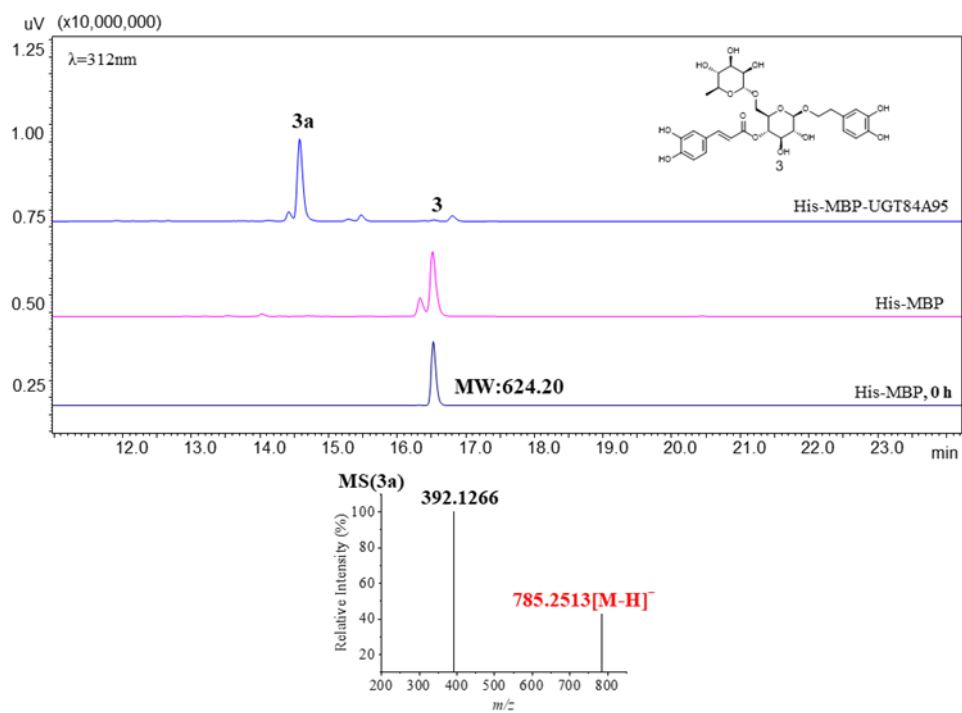


Figure S4. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound 3.

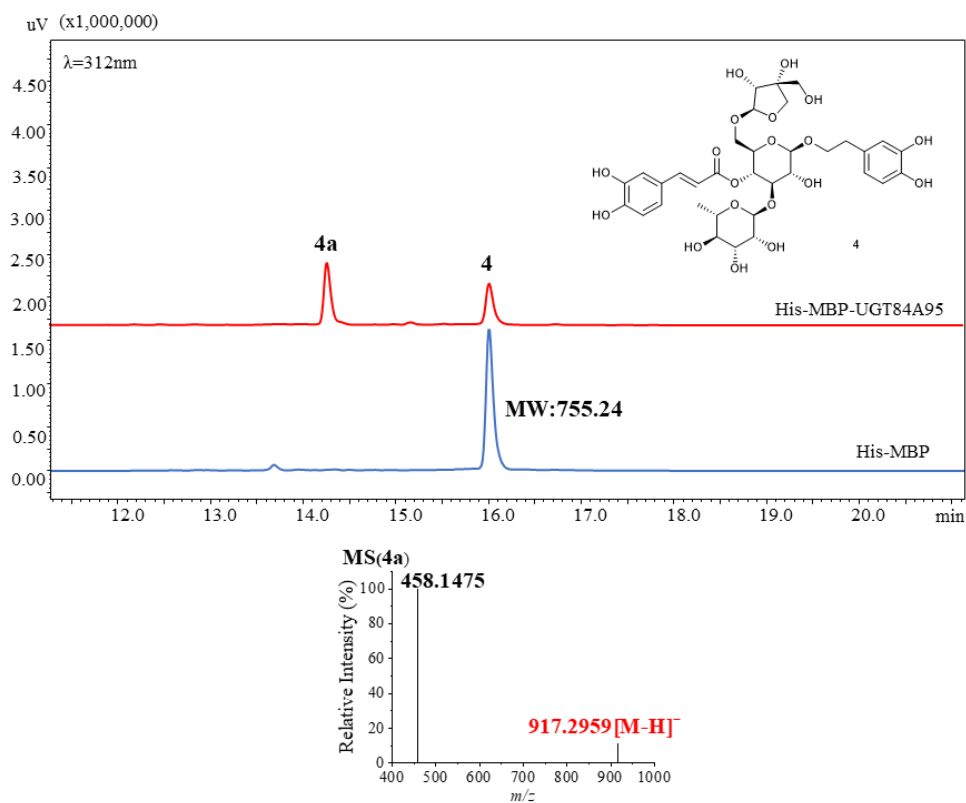


Figure S5. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound 4.

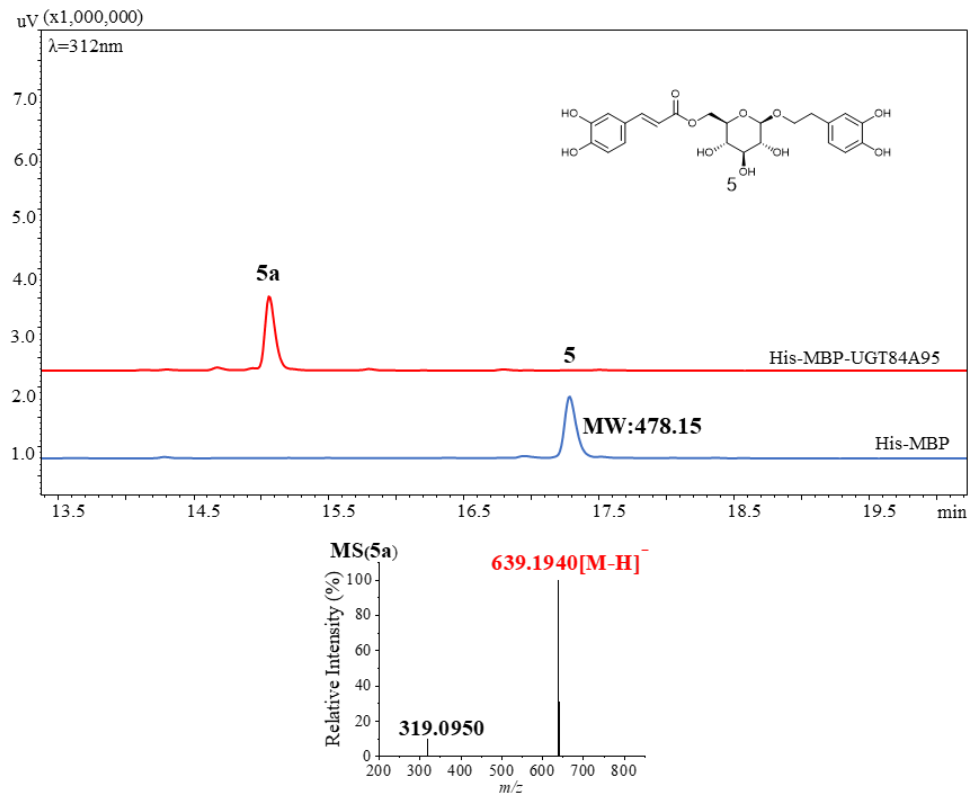


Figure S6. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound 5.

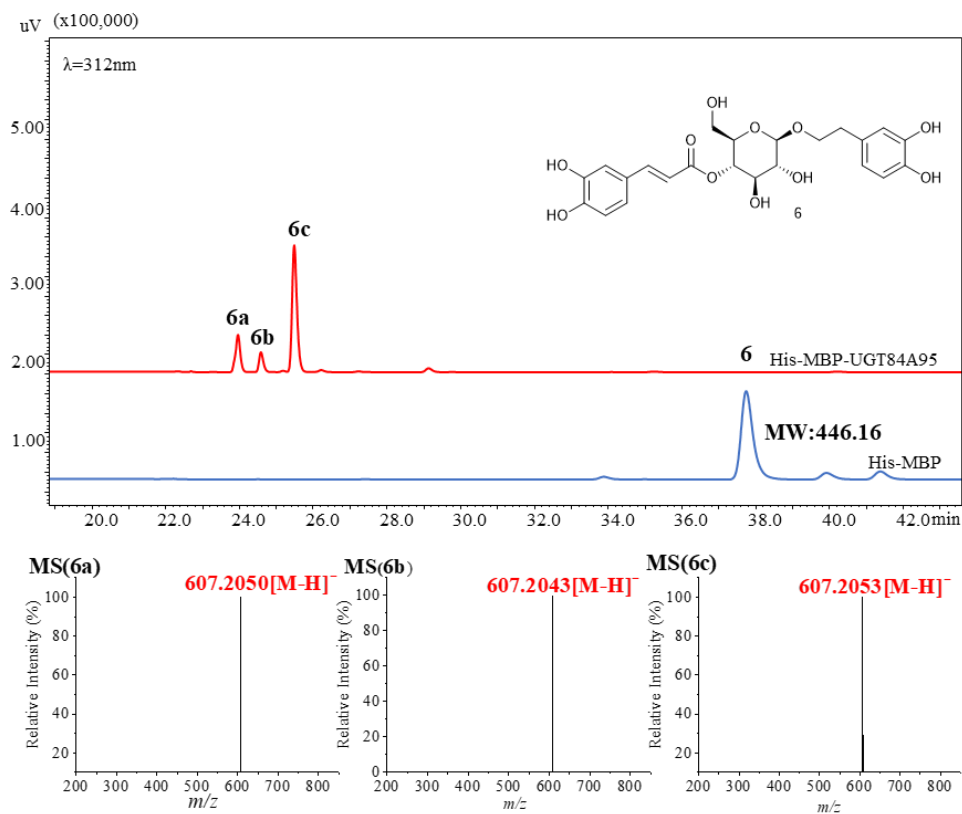


Figure S7. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound 6.

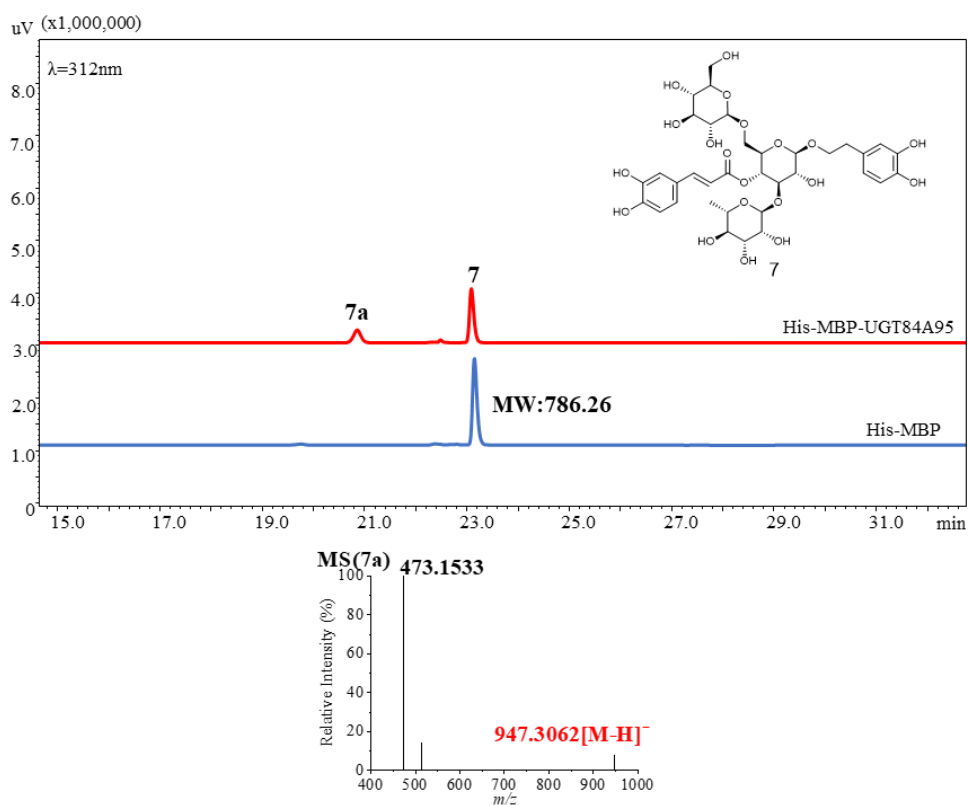


Figure S8. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound 7.

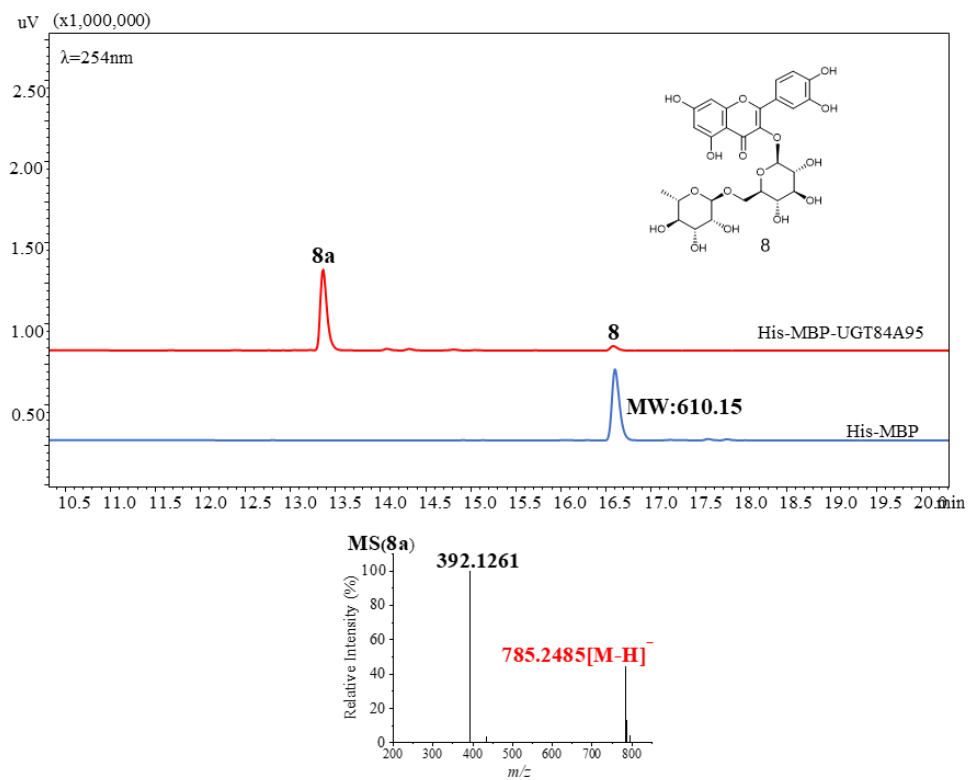


Figure S9. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **8**.

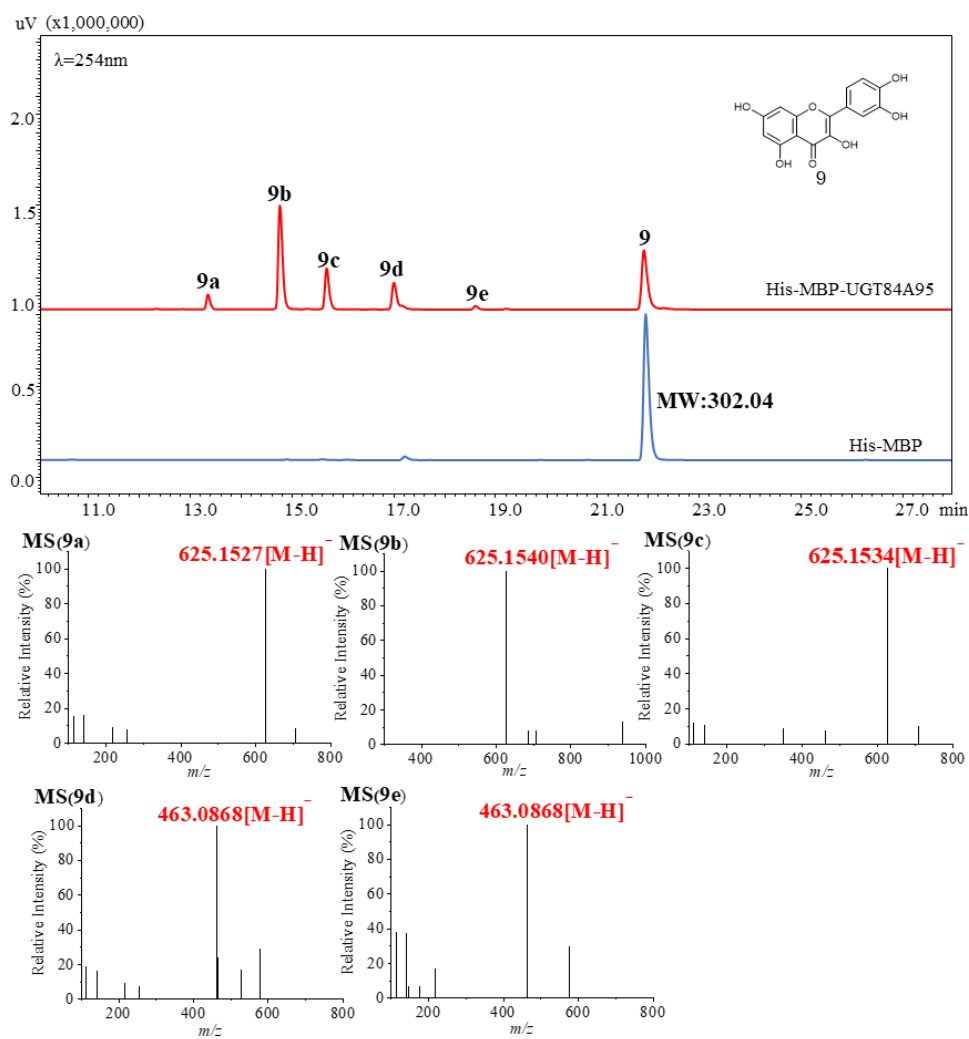


Figure S10. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **9**.

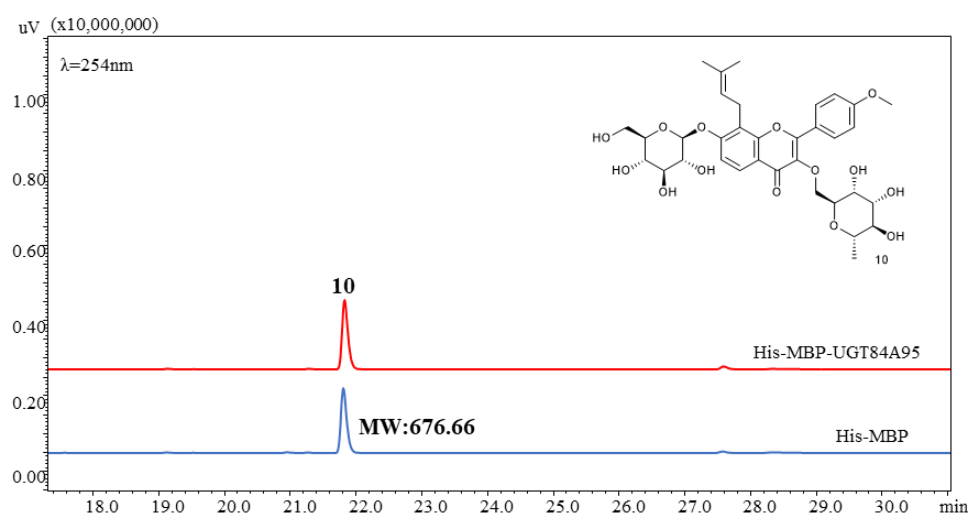


Figure S11. HPLC analysis of UGT84A95 catalytic reaction mixtures for compound **10**.

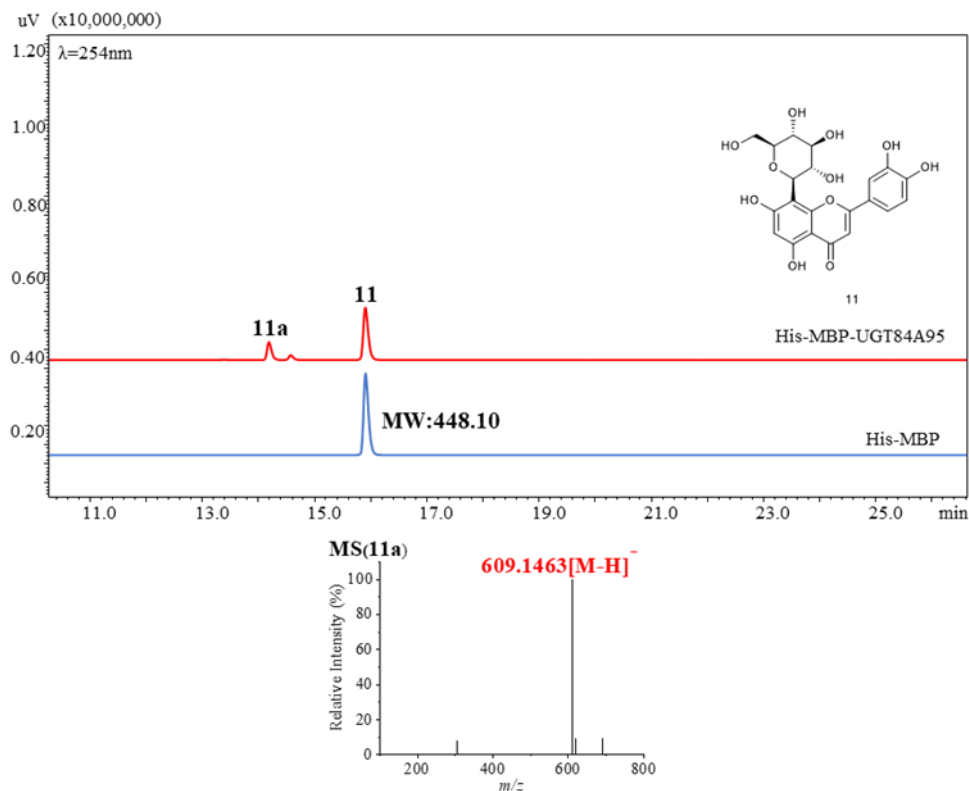


Figure S12. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **11**.

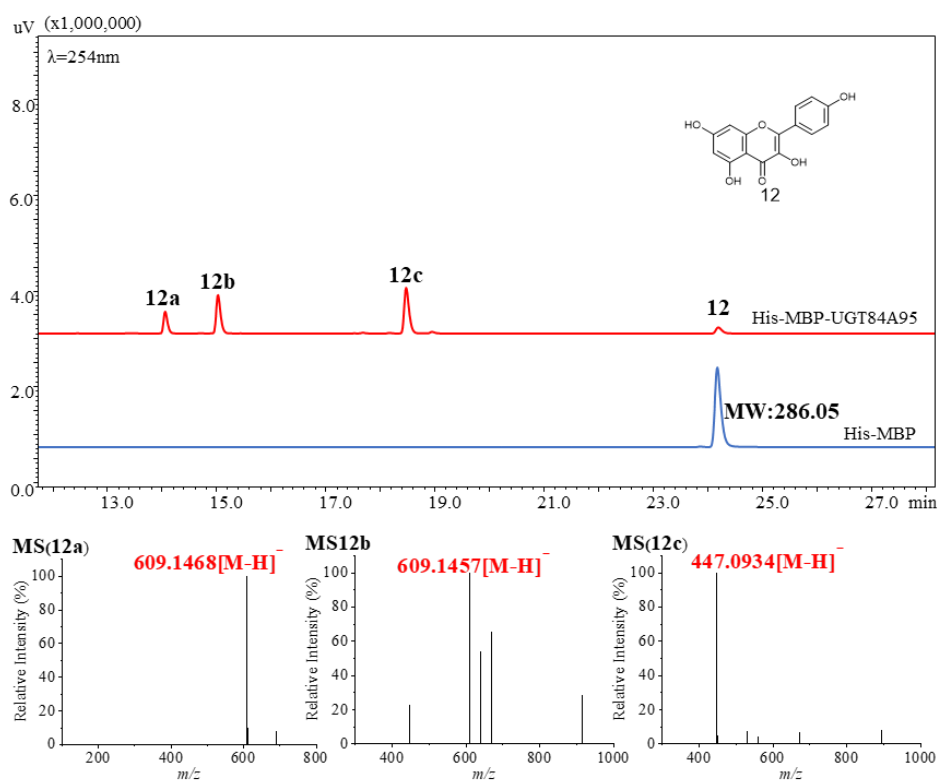


Figure S13. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **12**.

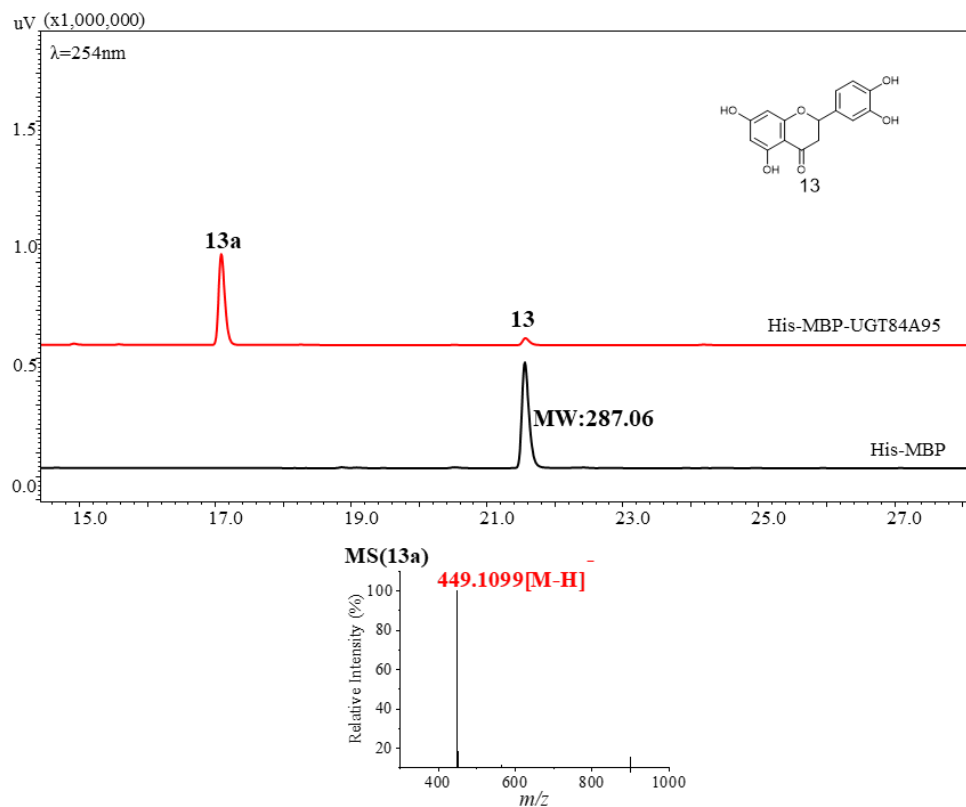


Figure S14. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **13**.

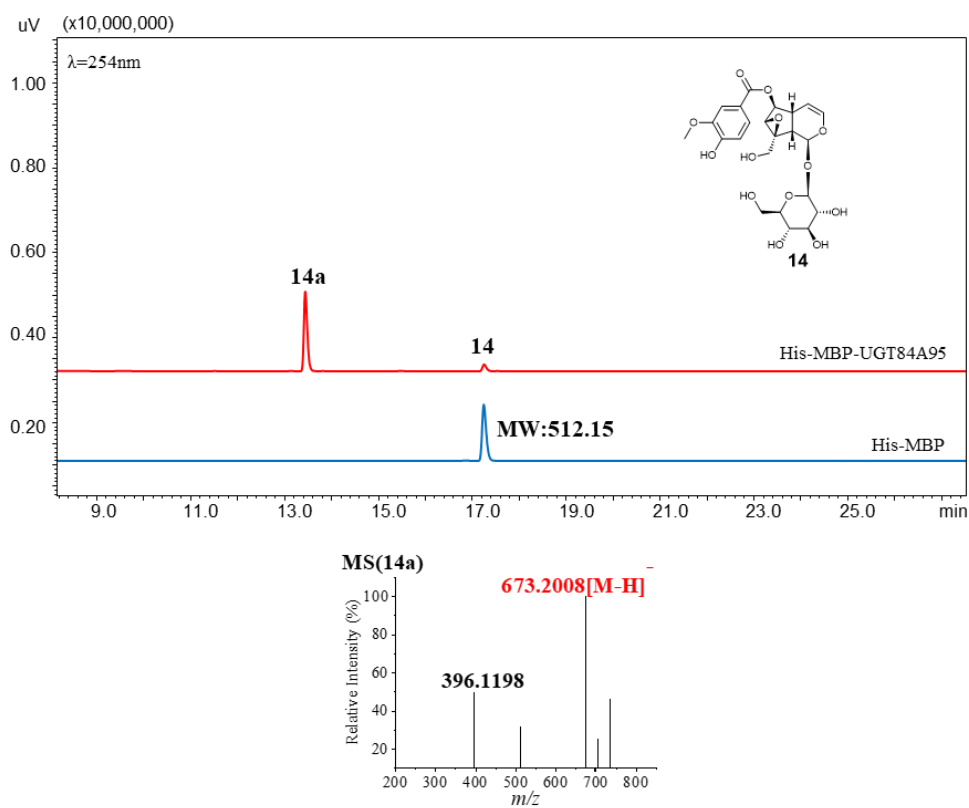


Figure S15. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **14**.

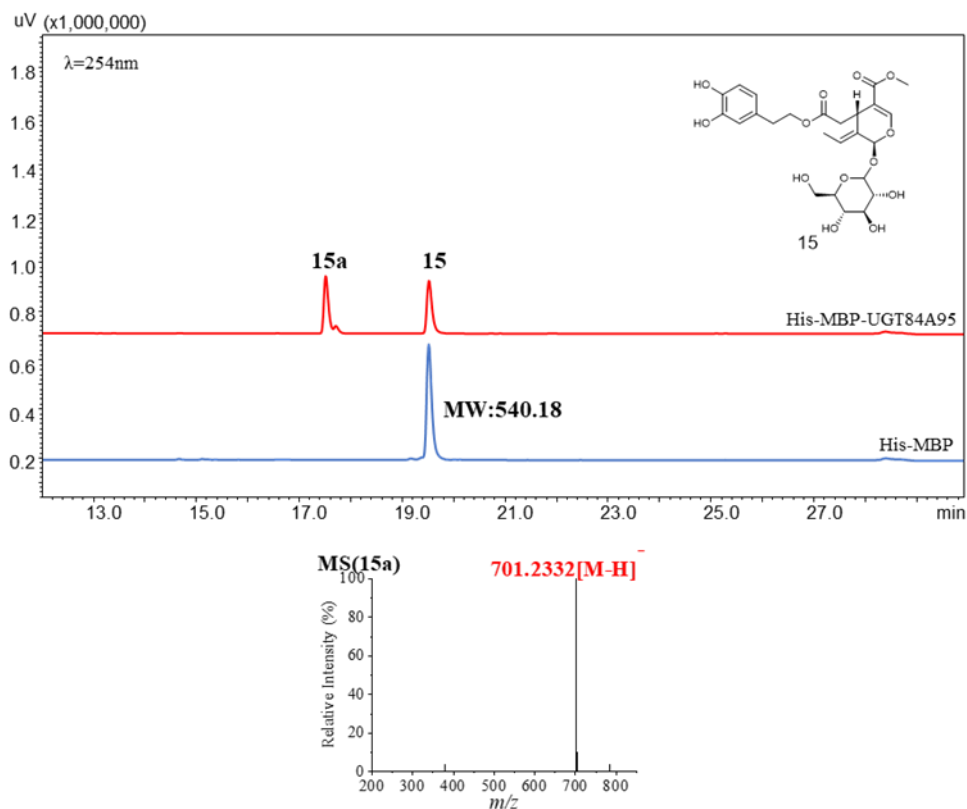


Figure S16. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound 15.

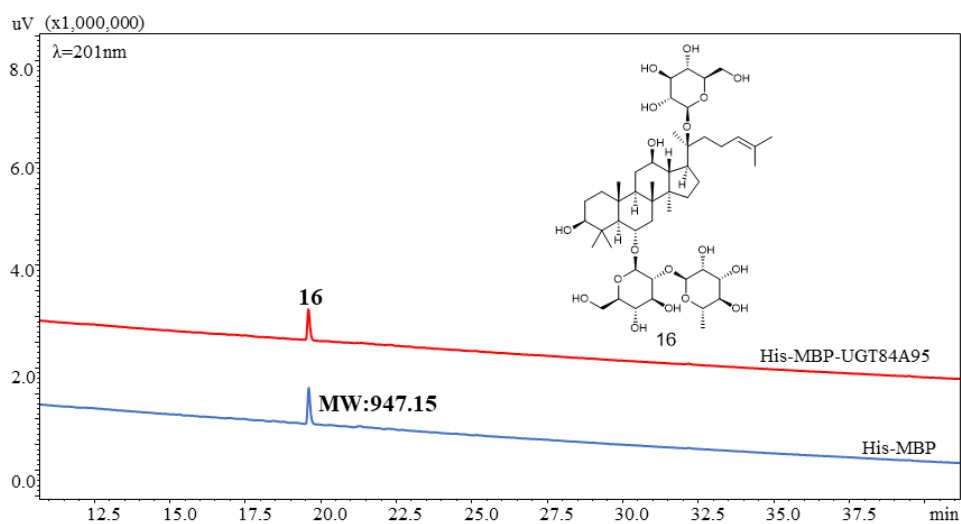


Figure S17. HPLC analysis of UGT84A95 catalytic reaction mixtures for compound 16.

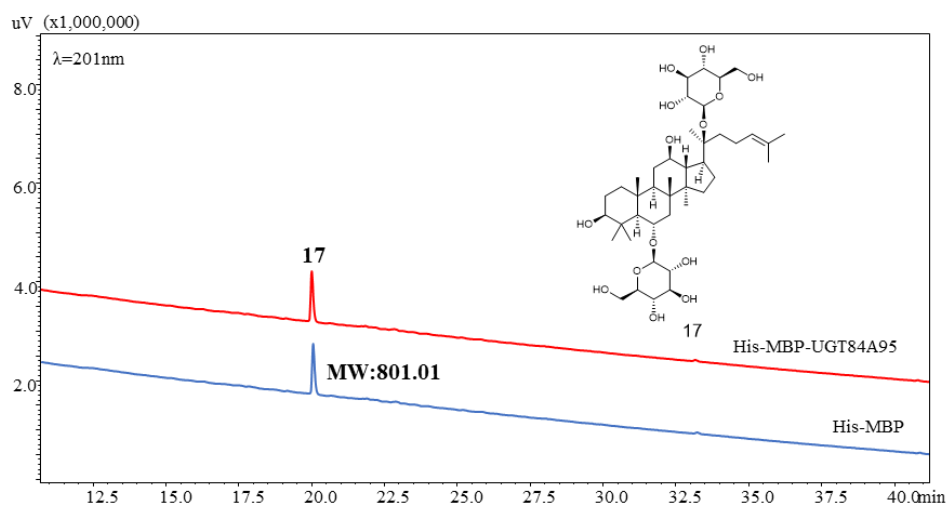


Figure S18. HPLC analysis of UGT84A95 catalytic reaction mixtures for compound **17**.

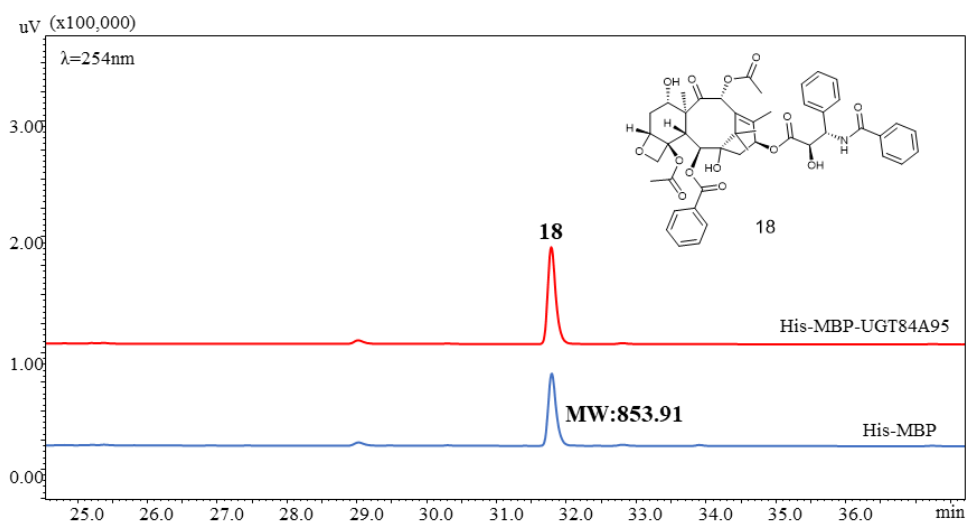


Figure S19. HPLC analysis of UGT84A95 catalytic reaction mixtures for compound **18**.

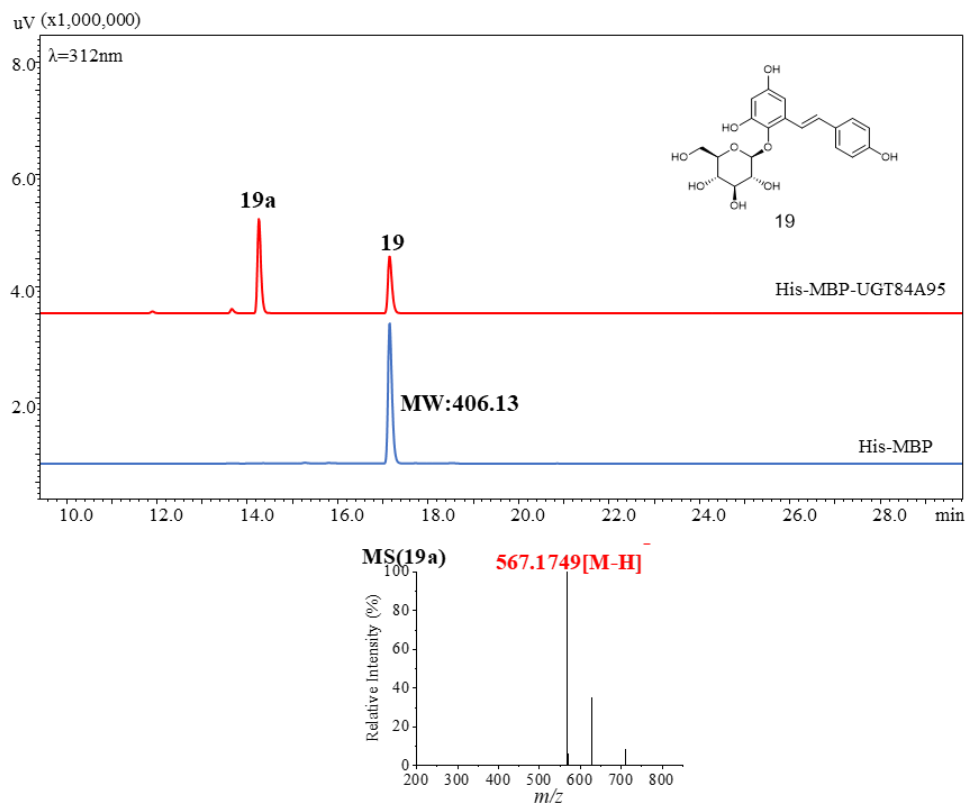


Figure S20. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **19**.

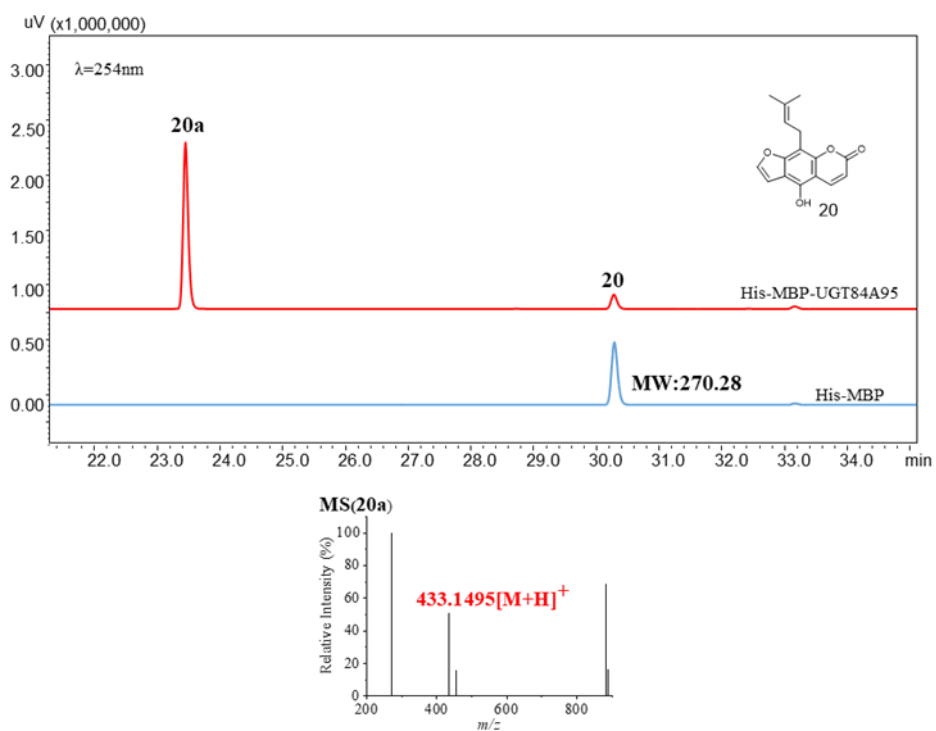


Figure S21. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **20**.

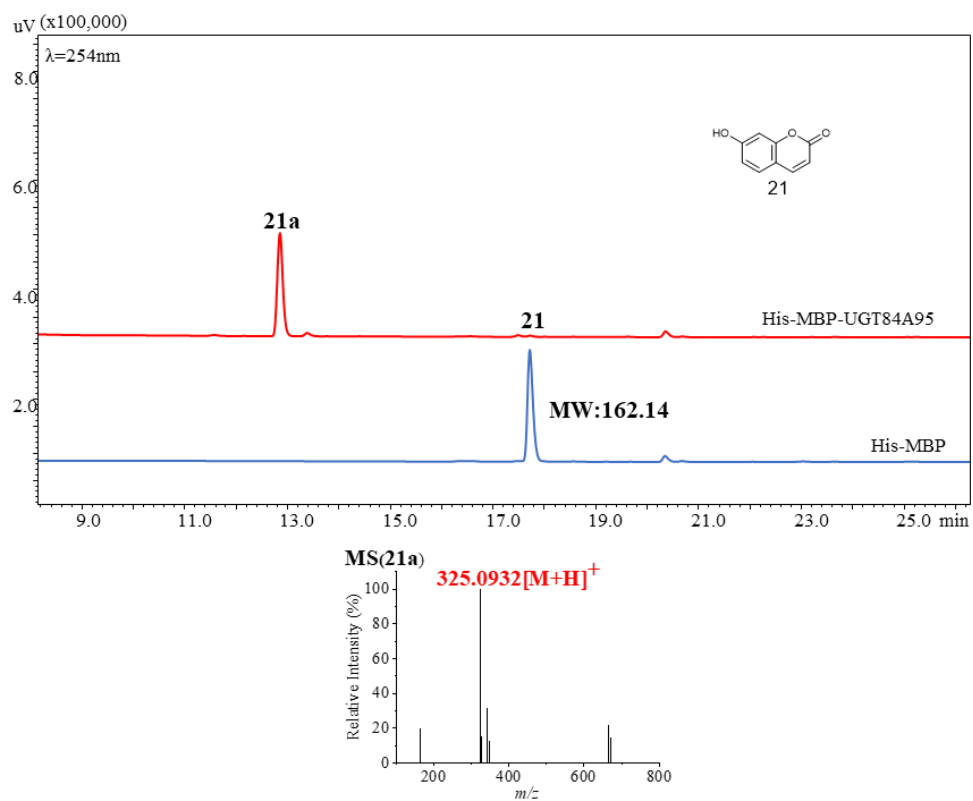


Figure S22. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **21**.

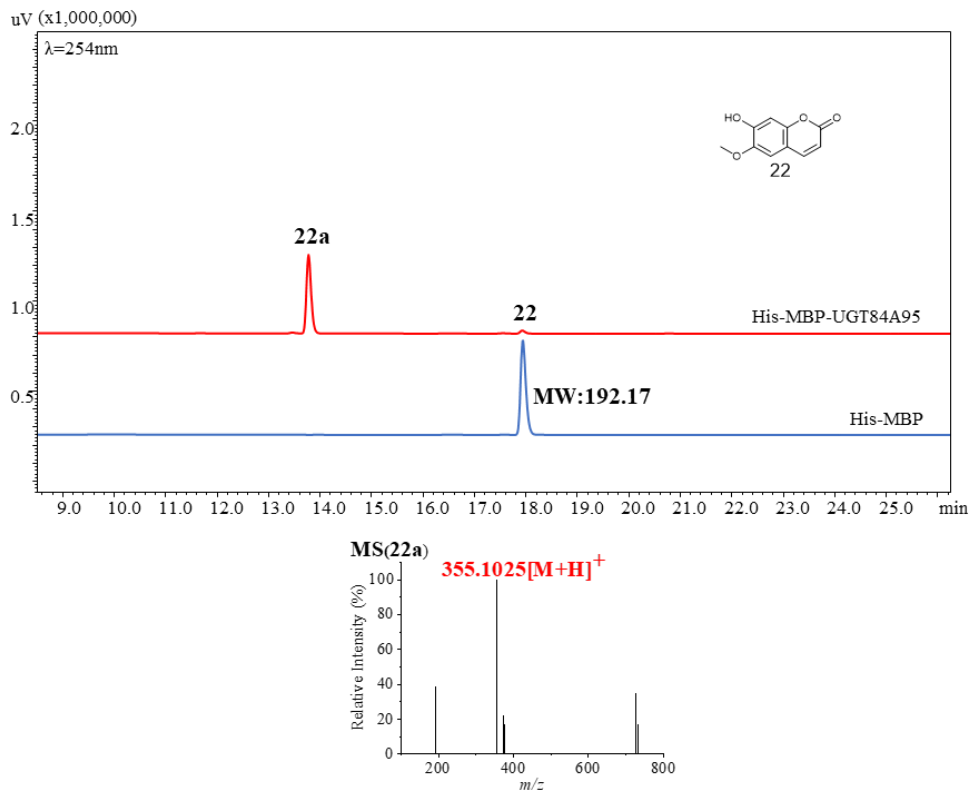


Figure S23. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **22**.

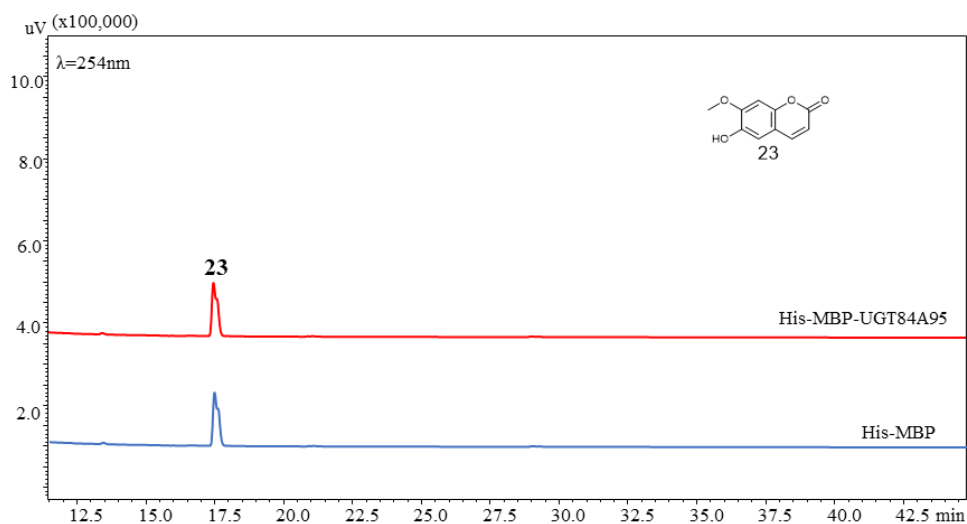


Figure S24. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **23**.

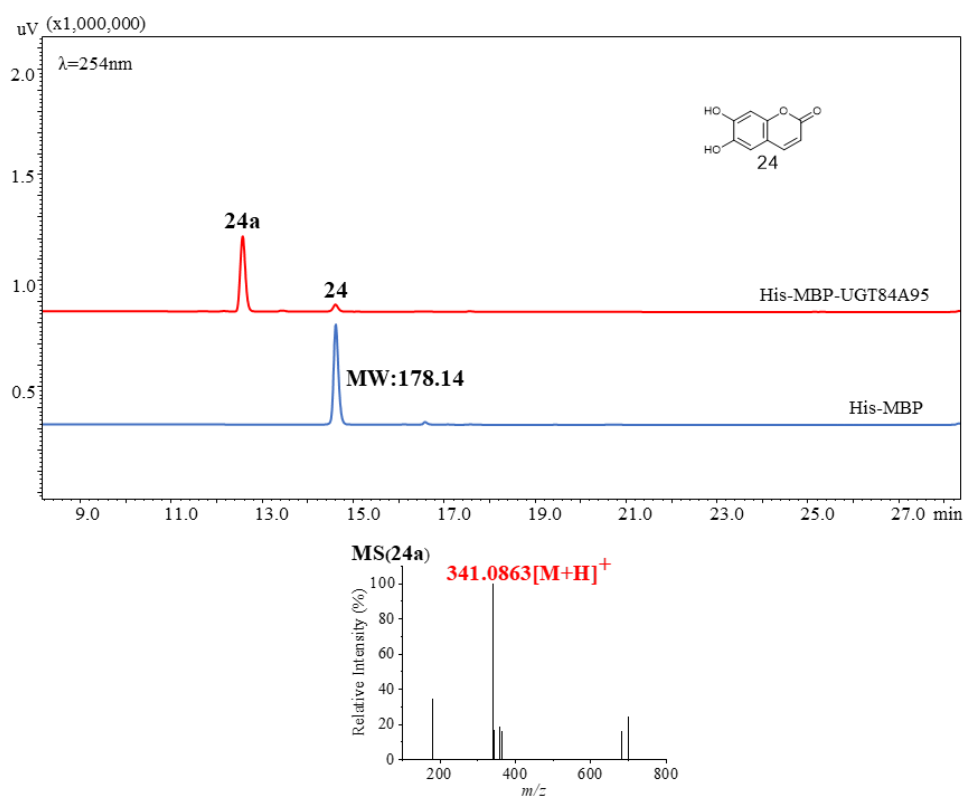


Figure S25. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **24**.

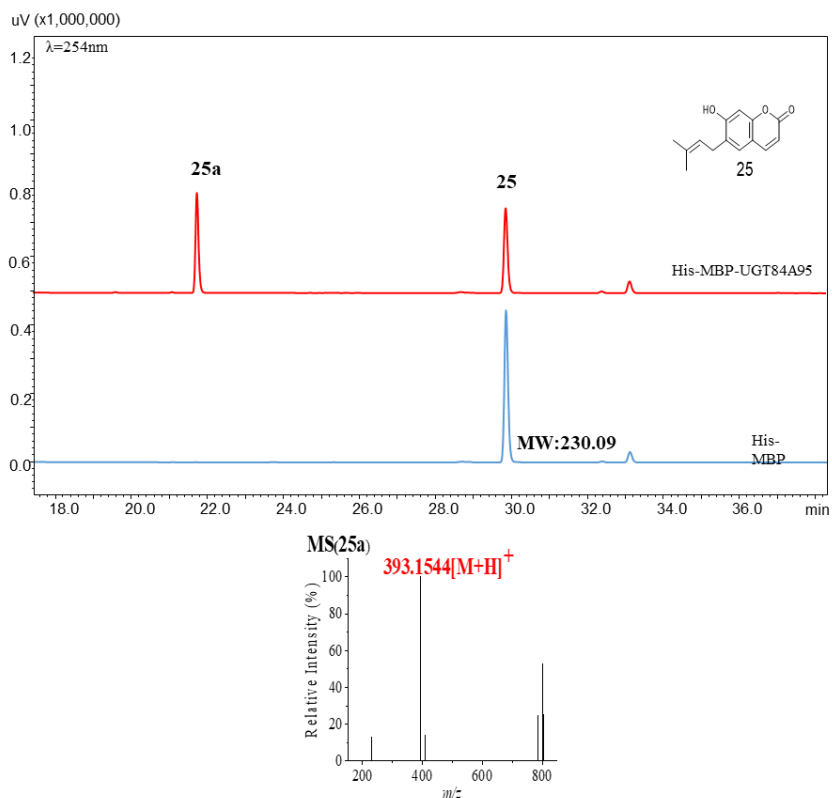


Figure S26. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound 25.

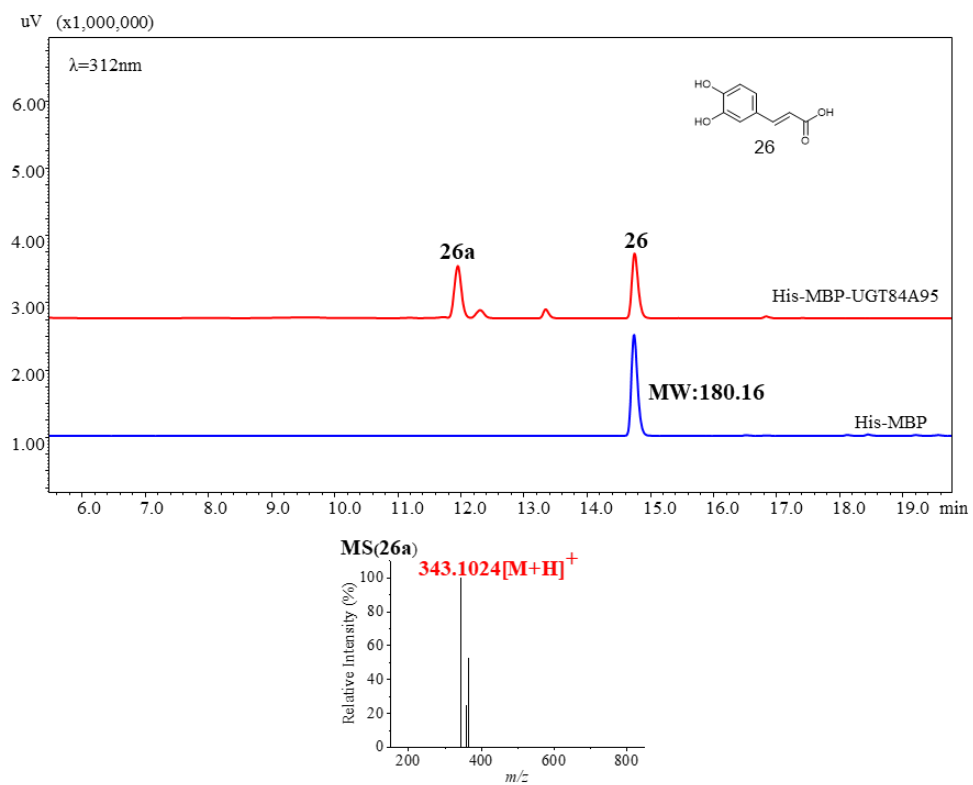


Figure S27. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound 26.

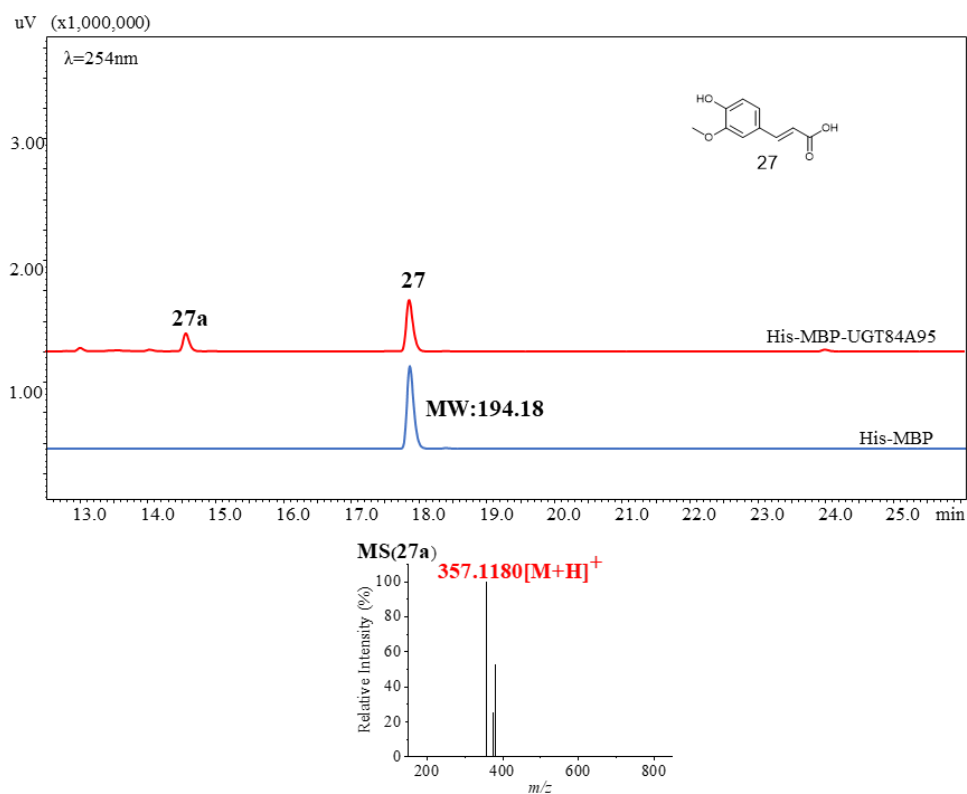


Figure S28. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **27**.

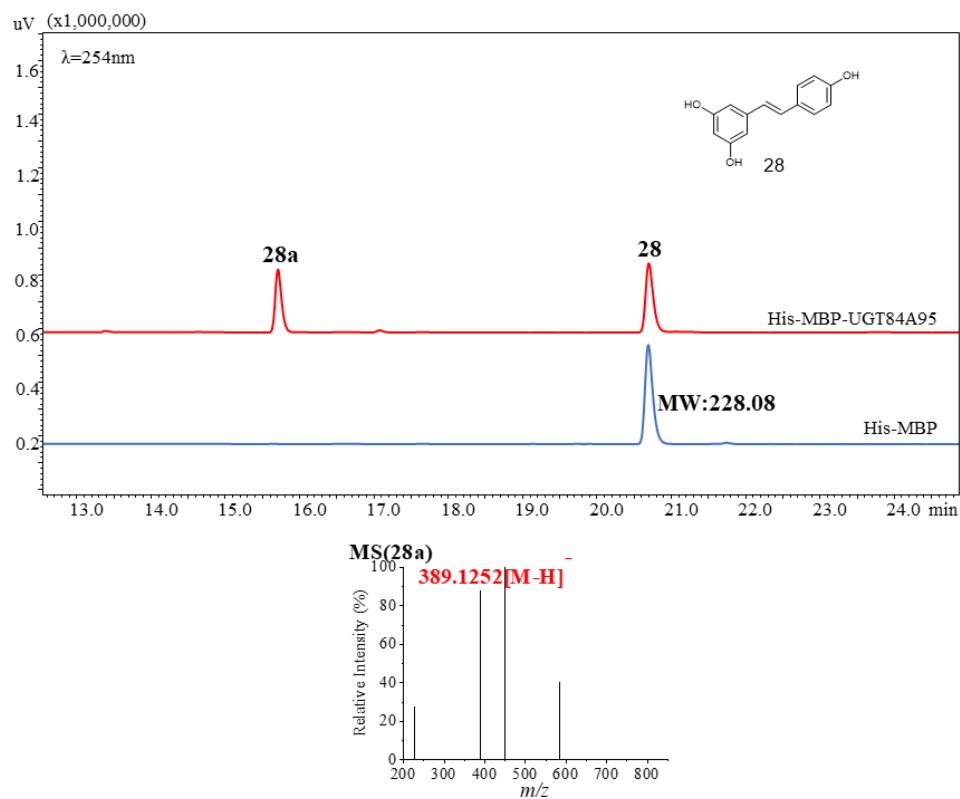


Figure S29. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures for compound **28**.

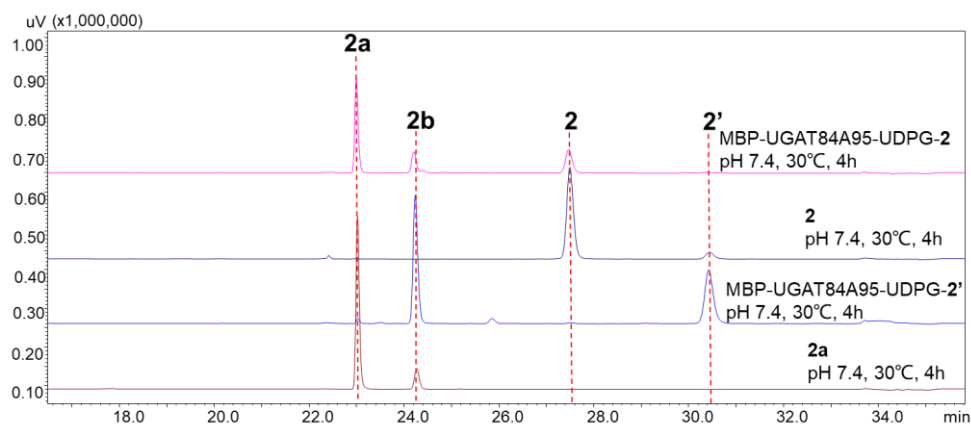


Figure 30. HPLC analysis of UGT84A95 catalytic reactions for compound **2** and **2'**.

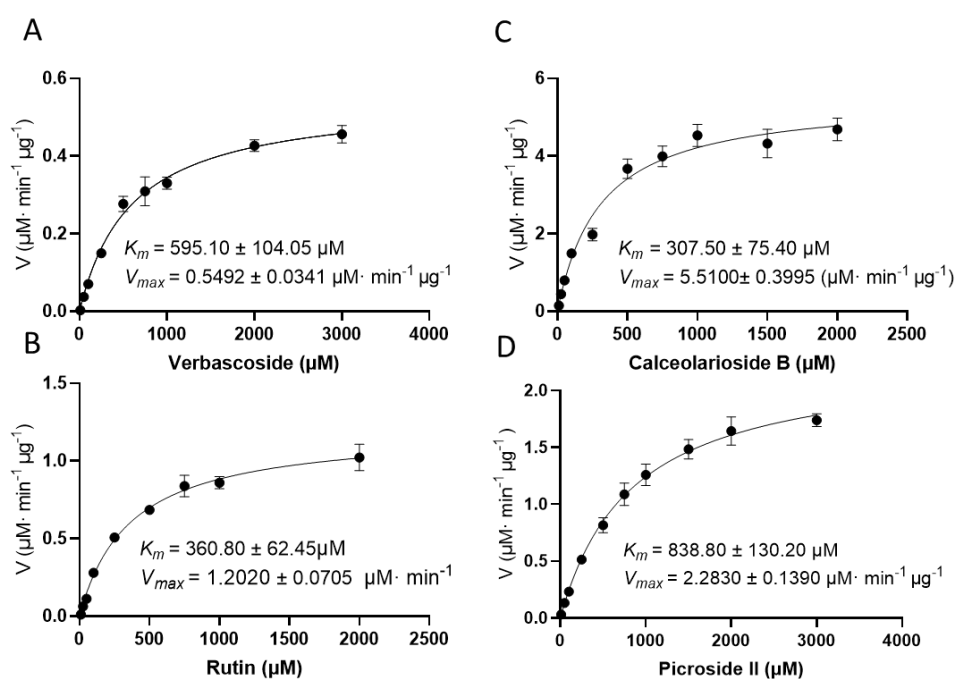


Figure S31. Determination of kinetic parameters (K_m and V_{max} values) for purified UGT71BD1 using UDP-Glc as a sugar donor and **2**, **5**, **8** and **14** as sugar acceptors, respectively.

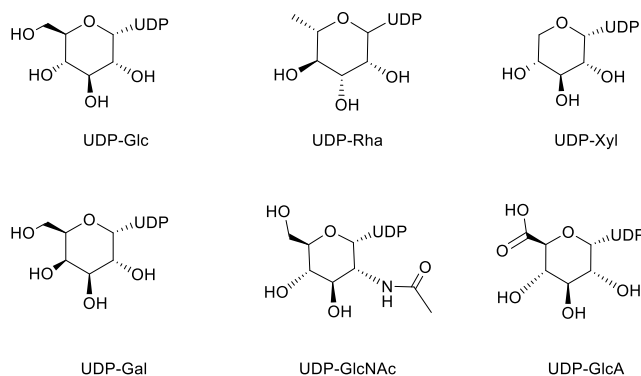


Figure S32. Structures of sugar donor in this study.

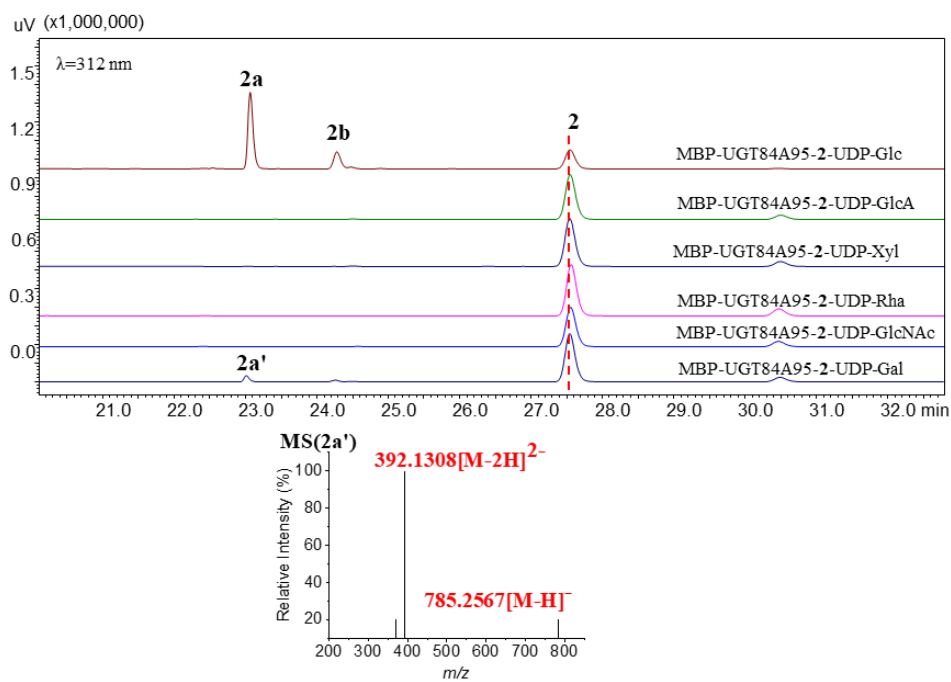


Figure S33. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures using different sugar donors and using **2** as the acceptor.

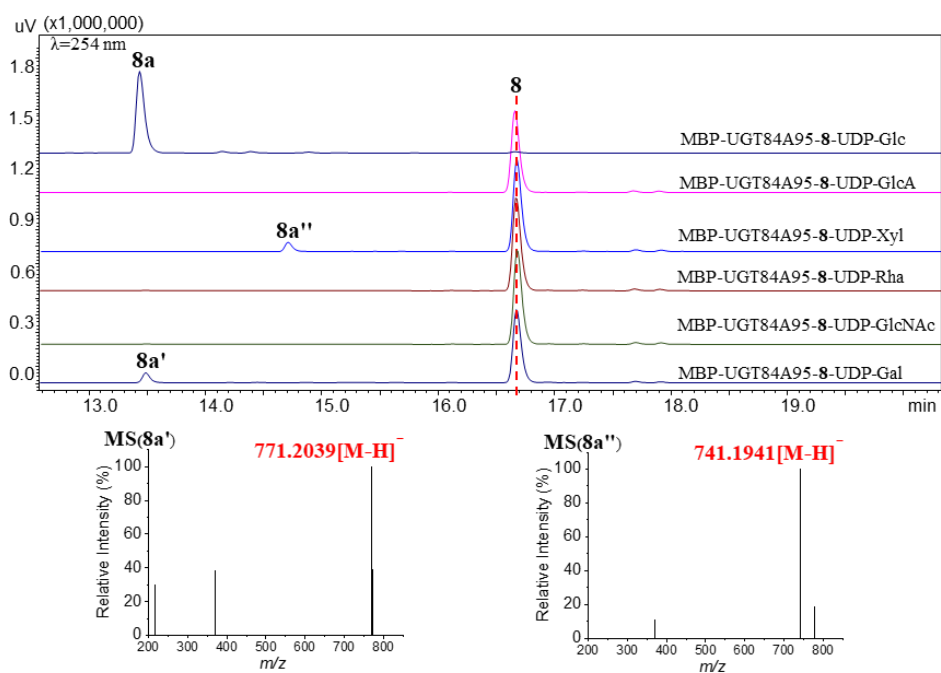


Figure S34. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures using different sugar donors and using **8** as the acceptor.

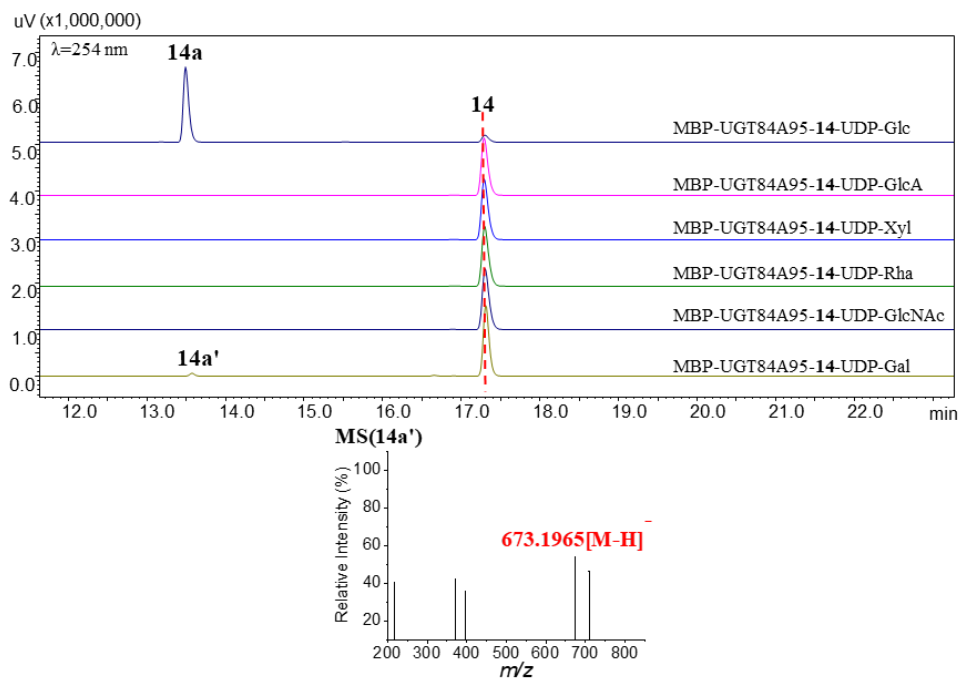


Figure S35. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures using different sugar donors and using **14** as the acceptor

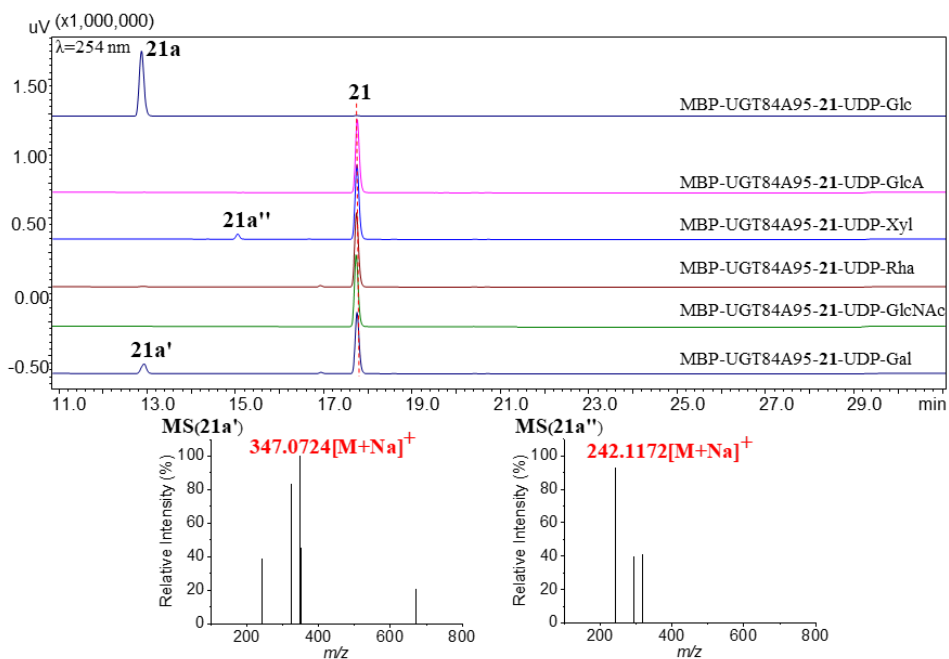
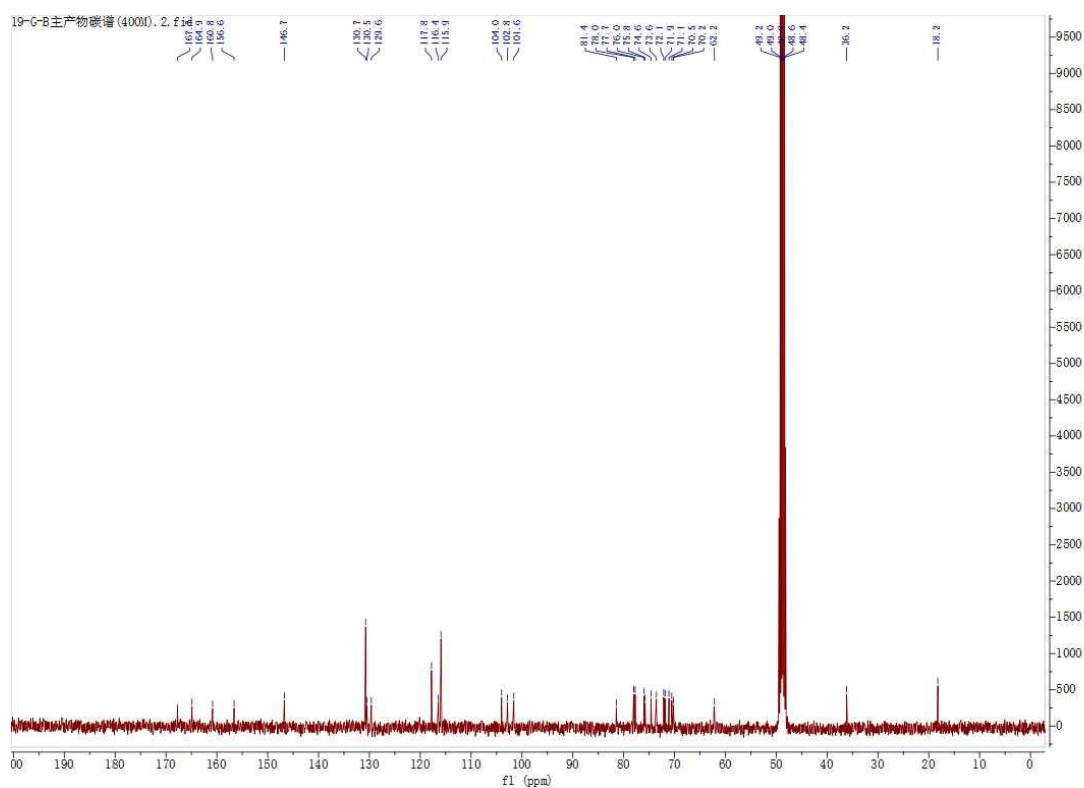
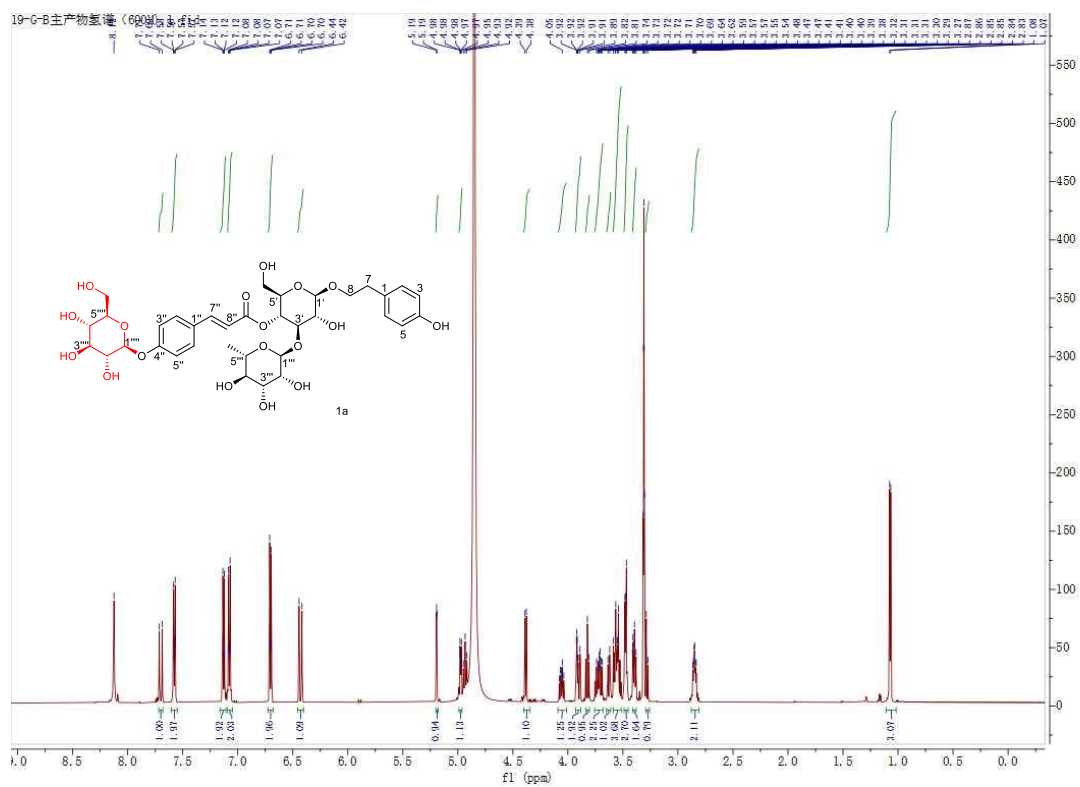


Figure S36. HPLC and LC-HRMS analysis of UGT84A95 catalytic reaction mixtures using different sugar donors and using **21** as the acceptor.



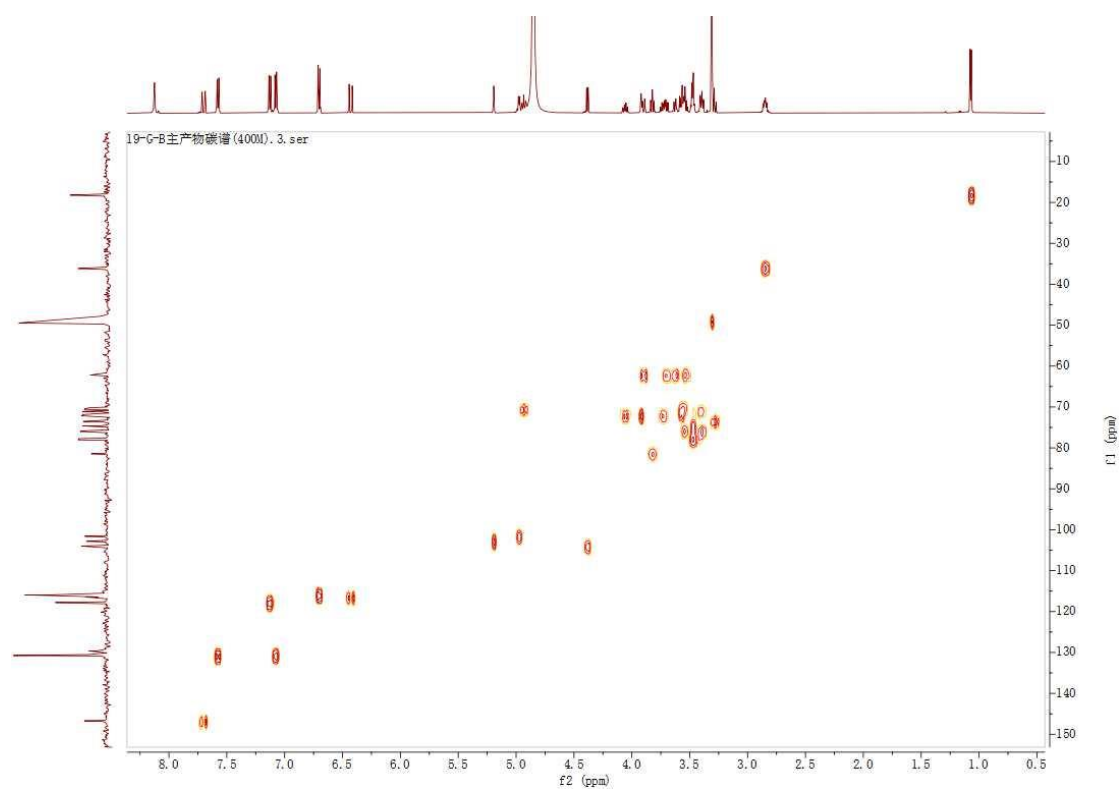


Figure S39. HSQC spectrum of glycosylated product **1a** (CD₃OD, 400 MHz).

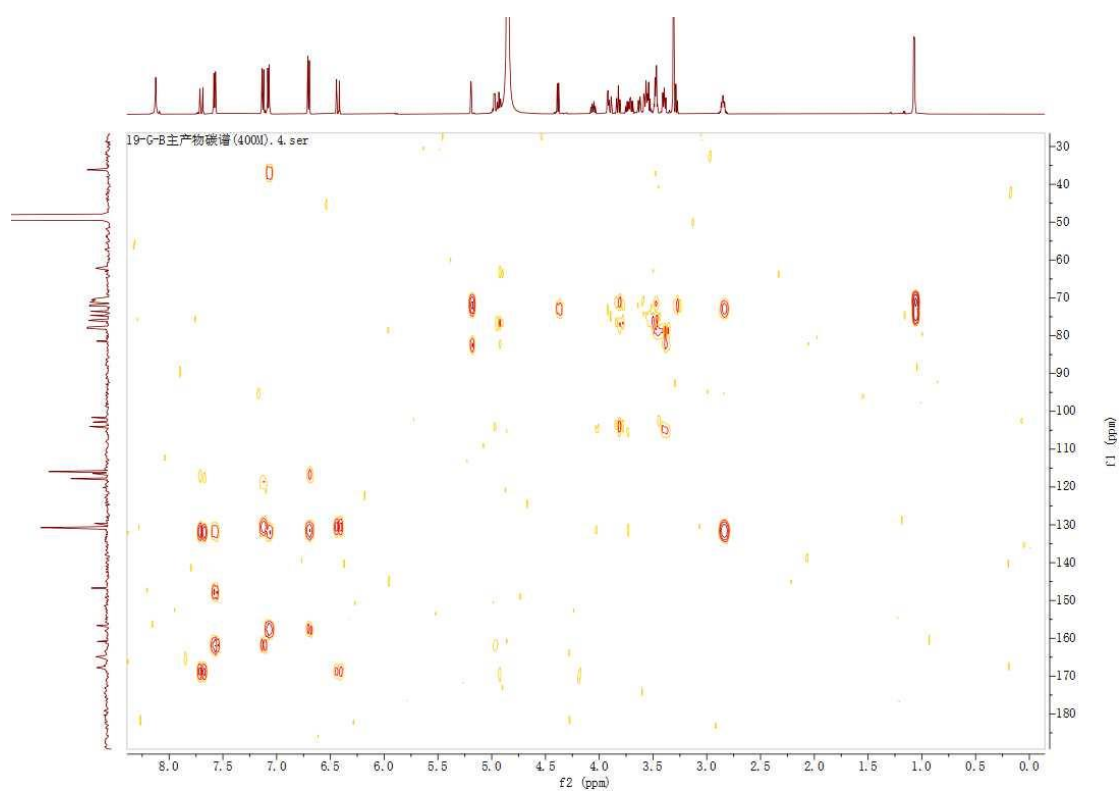


Figure S40. HMBC spectrum of glycosylated product **1a** (CD₃OD, 400 MHz).

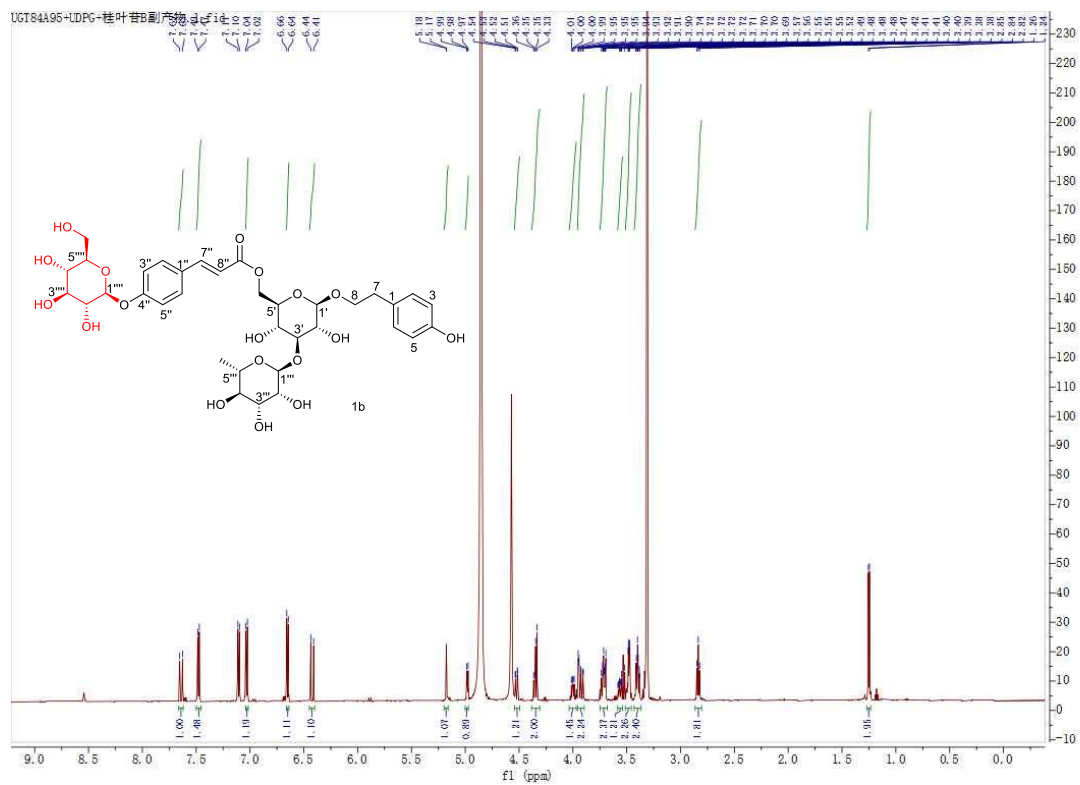


Figure S41. ^1H NMR spectrum of glycosylated product **1b** (CD_3OD , 600 MHz).

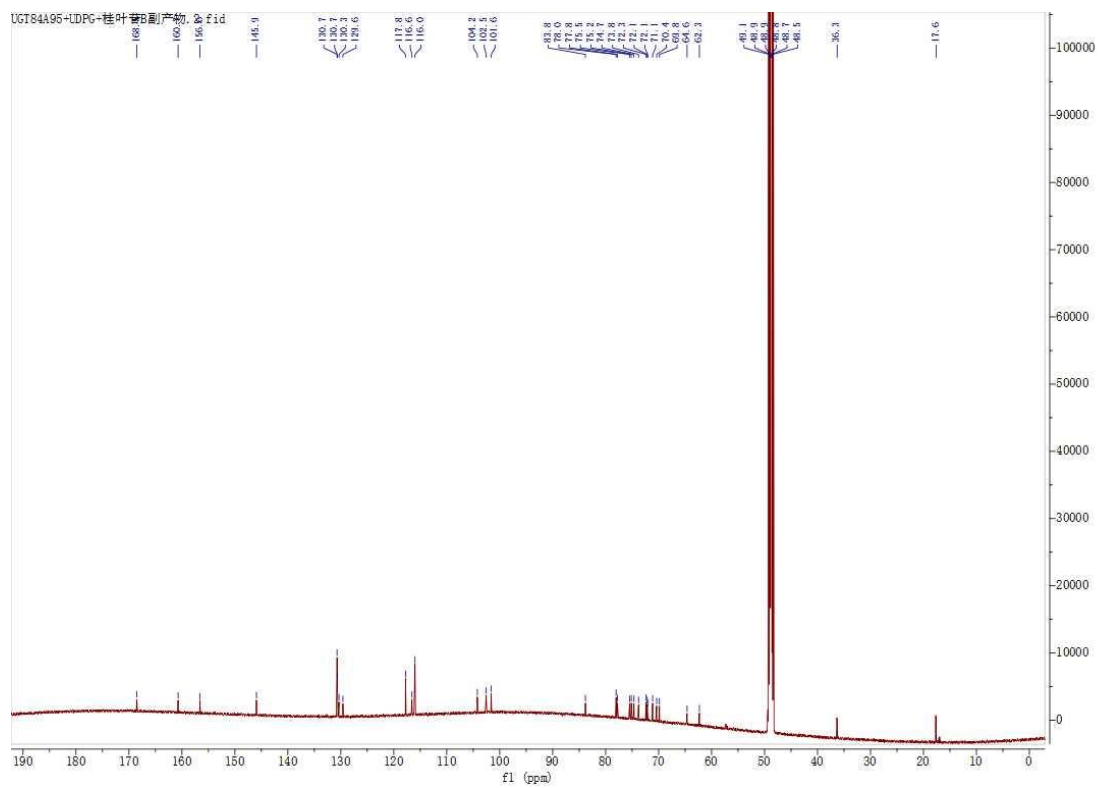


Figure S42. ^{13}C NMR spectrum of glycosylated product **1b** (CD_3OD , 600 MHz).

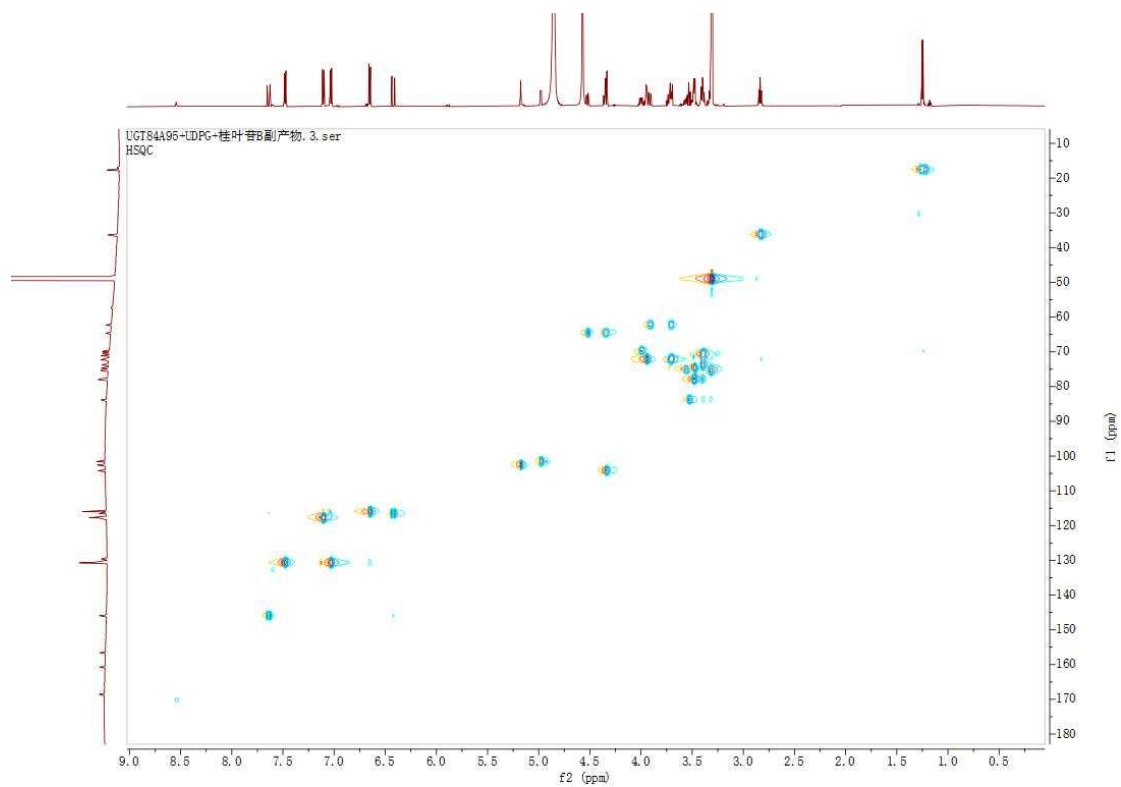


Figure S43. HSQC spectrum of glycosylated product **1b** (CD₃OD, 600 MHz).

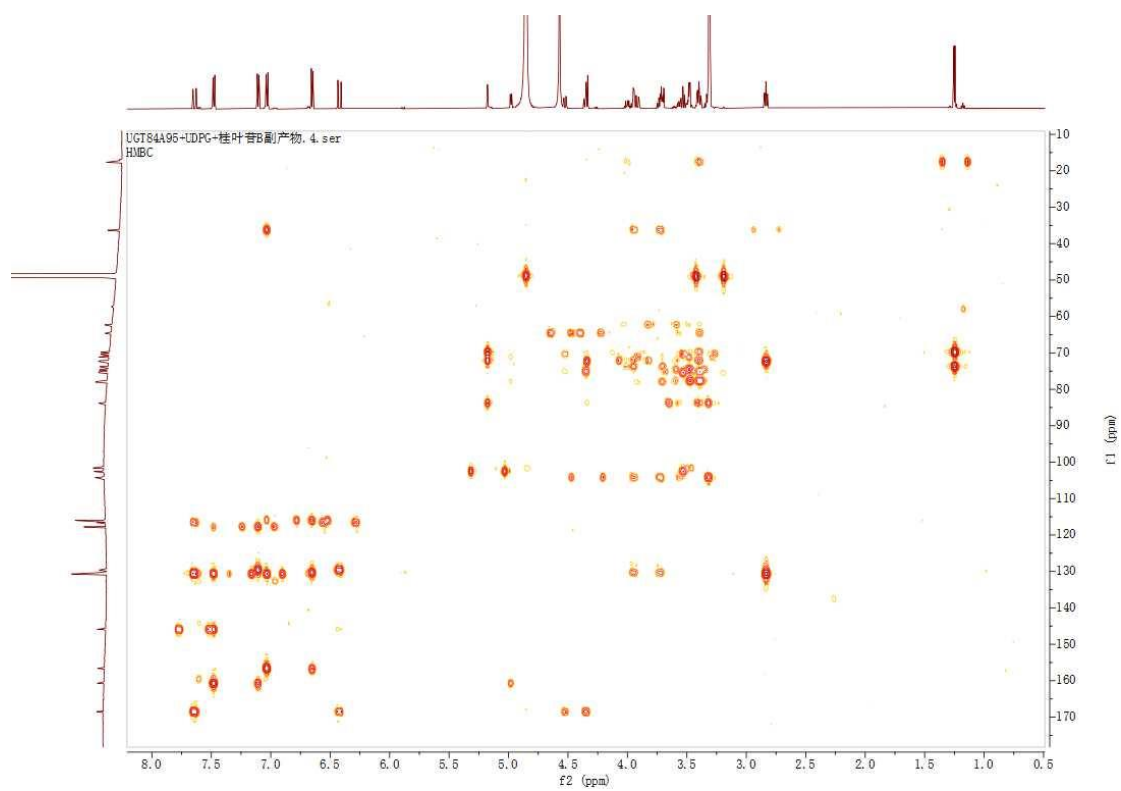


Figure S44. HMBC spectrum of glycosylated product **1b** (CD₃OD, 600 MHz).

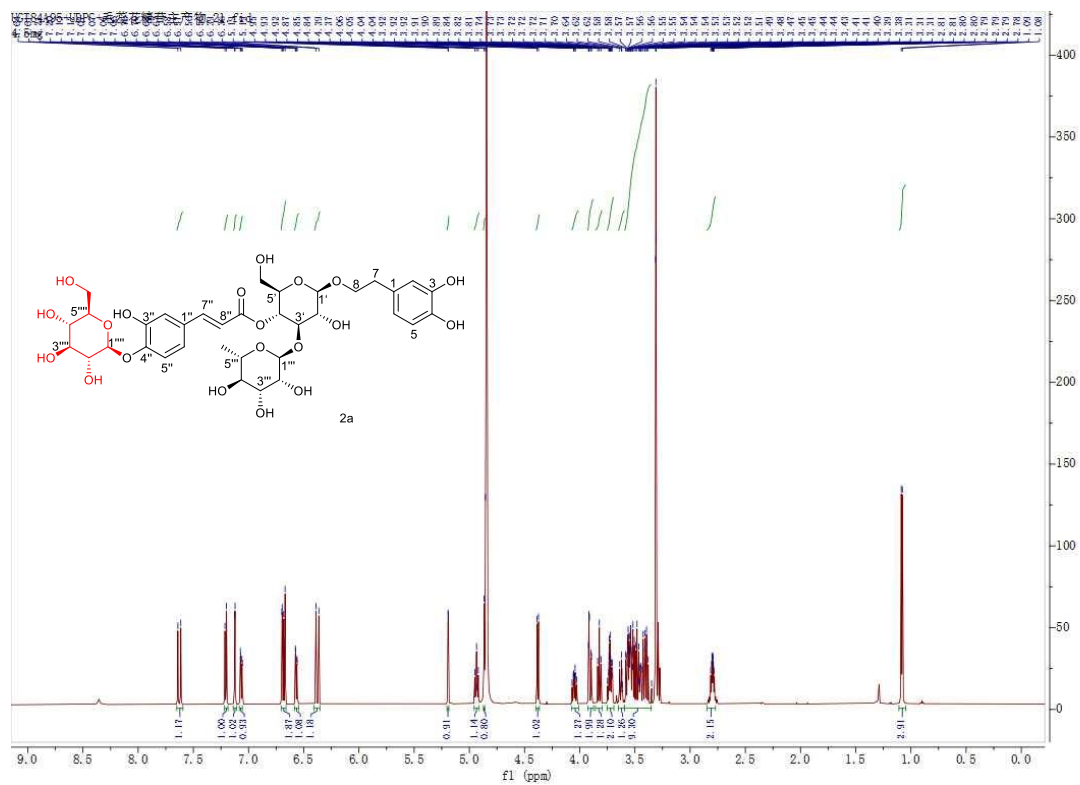


Figure S45. ^1H NMR spectrum of glycosylated product **2a** (CD_3OD , 600 MHz).

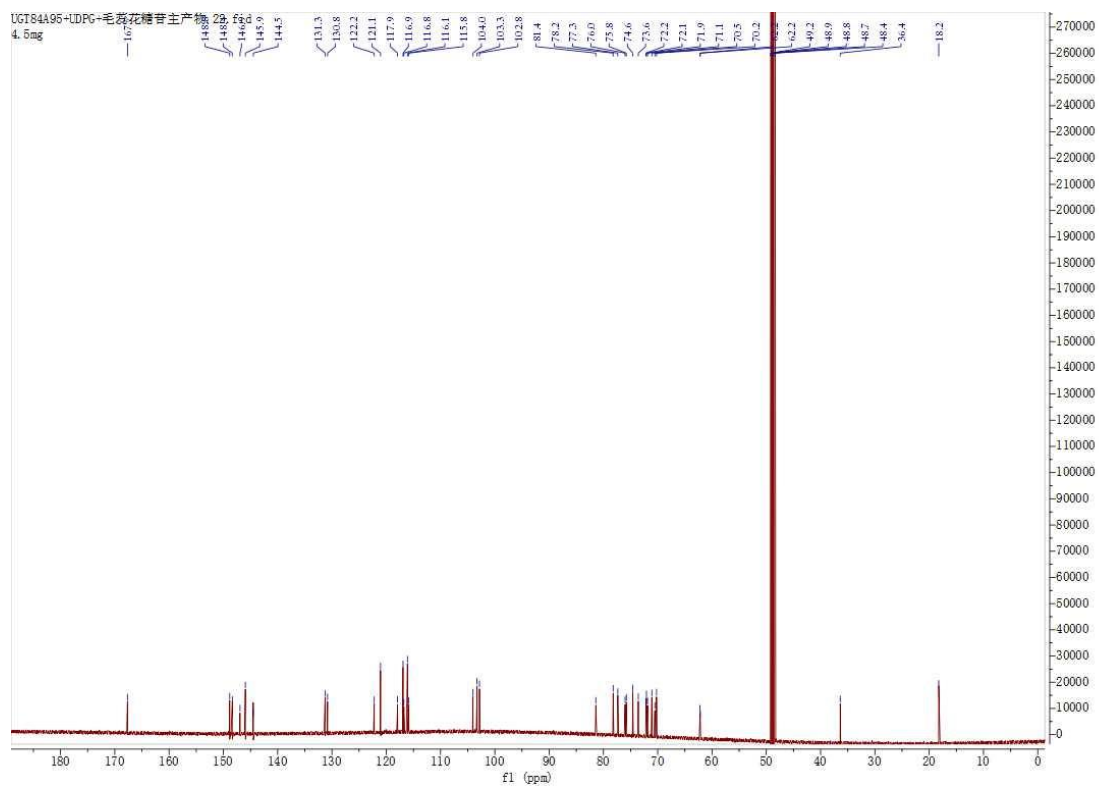


Figure S46. ^{13}C NMR spectrum of glycosylated product **2a** (CD_3OD , 600 MHz).

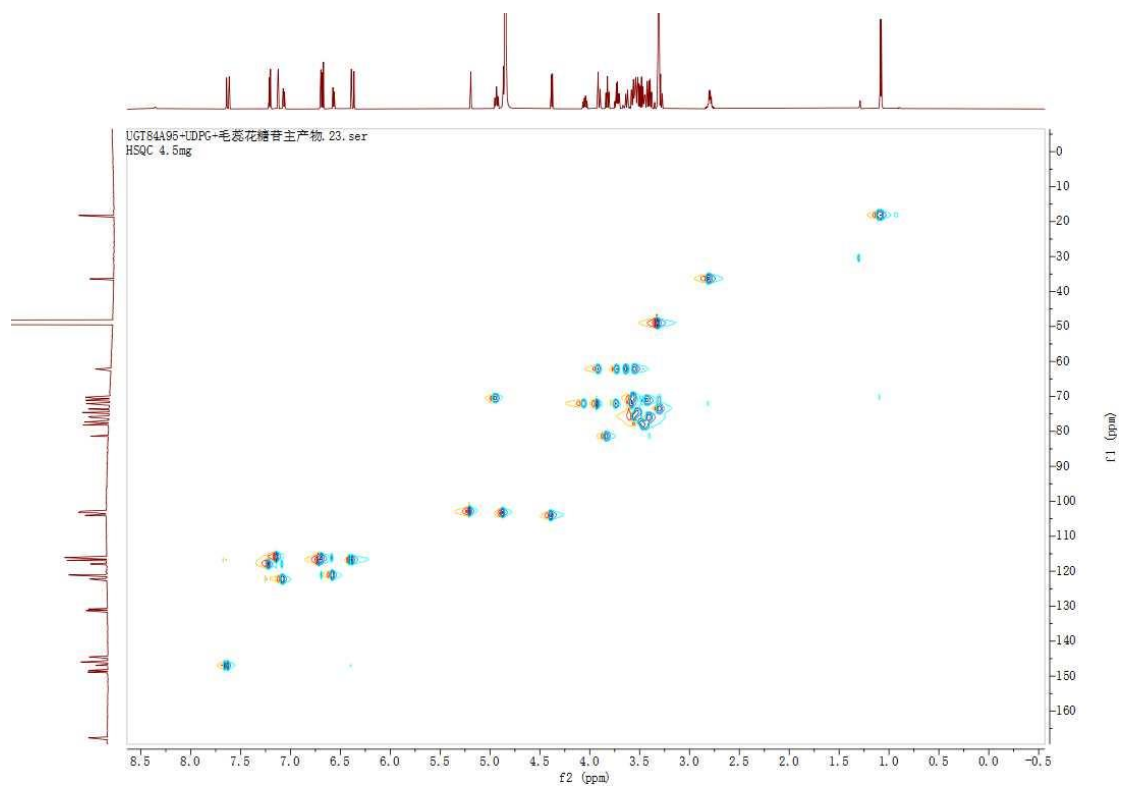


Figure S47. HSQC NMR spectrum of glycosylated product **2a** (CD₃OD, 600 MHz).

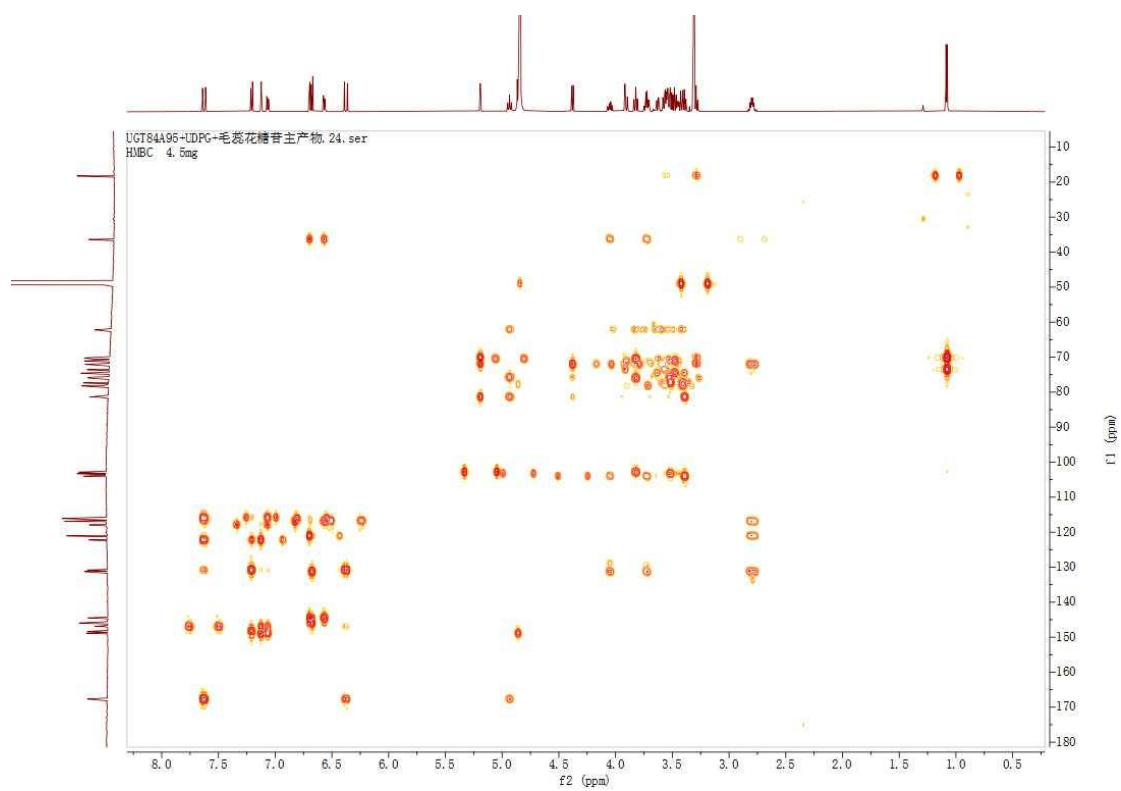
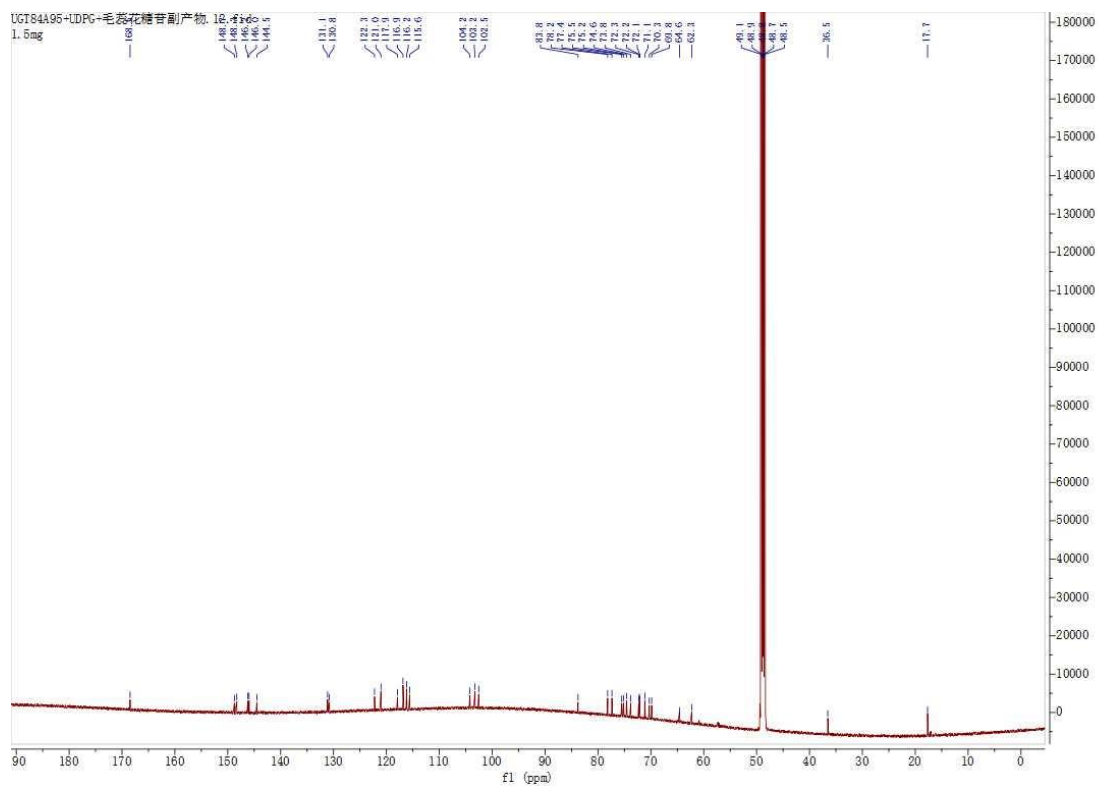
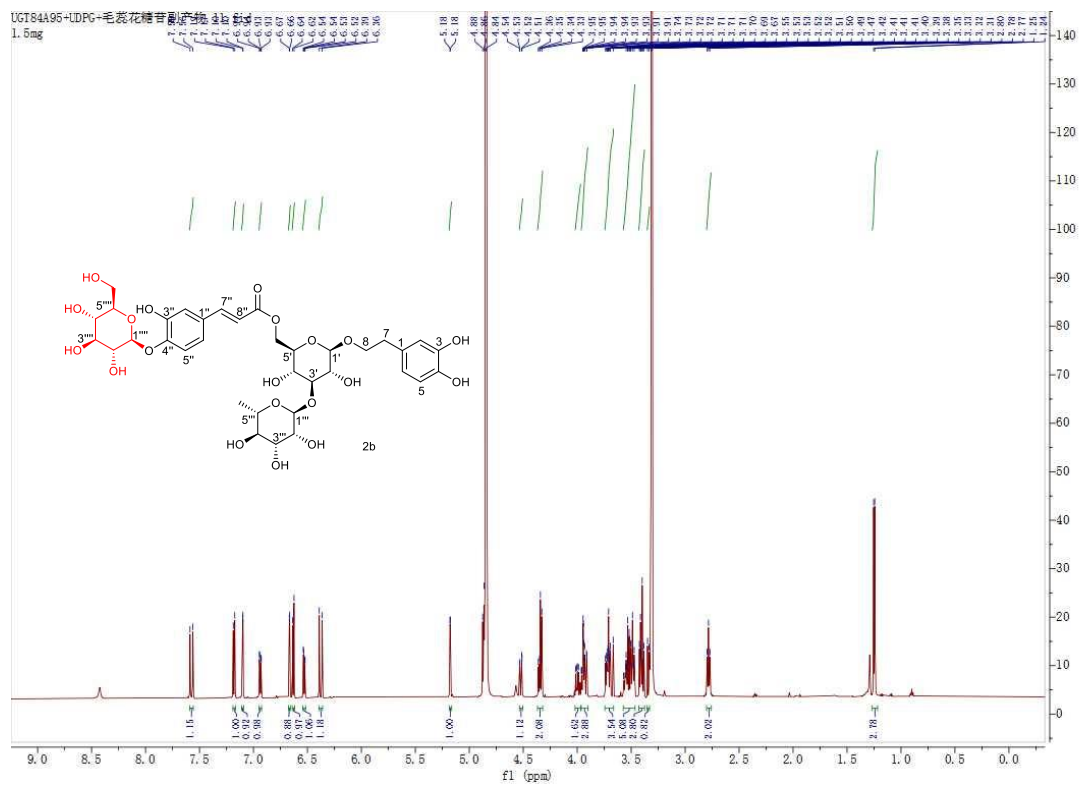


Figure S48. HMBC NMR spectrum of glycosylated product **2a** (CD₃OD, 600 MHz).



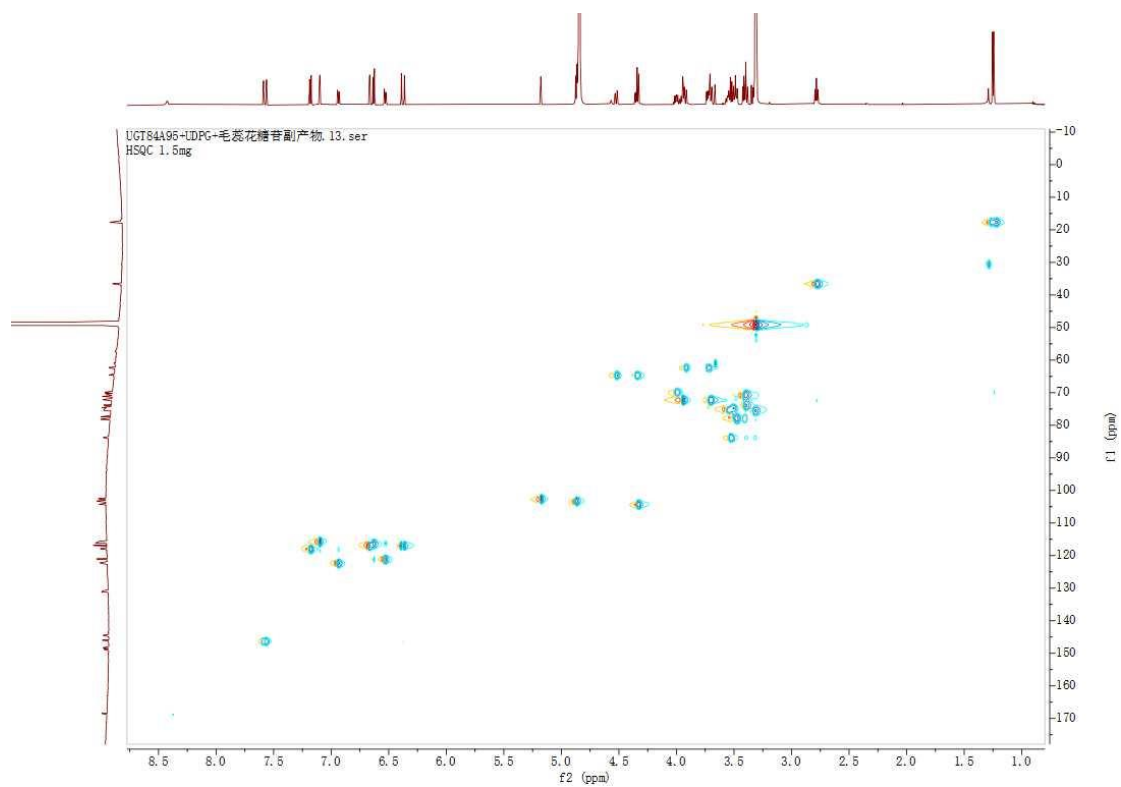


Figure S51. HSQC spectrum of glycosylated product **2b** (CD₃OD, 600 MHz).

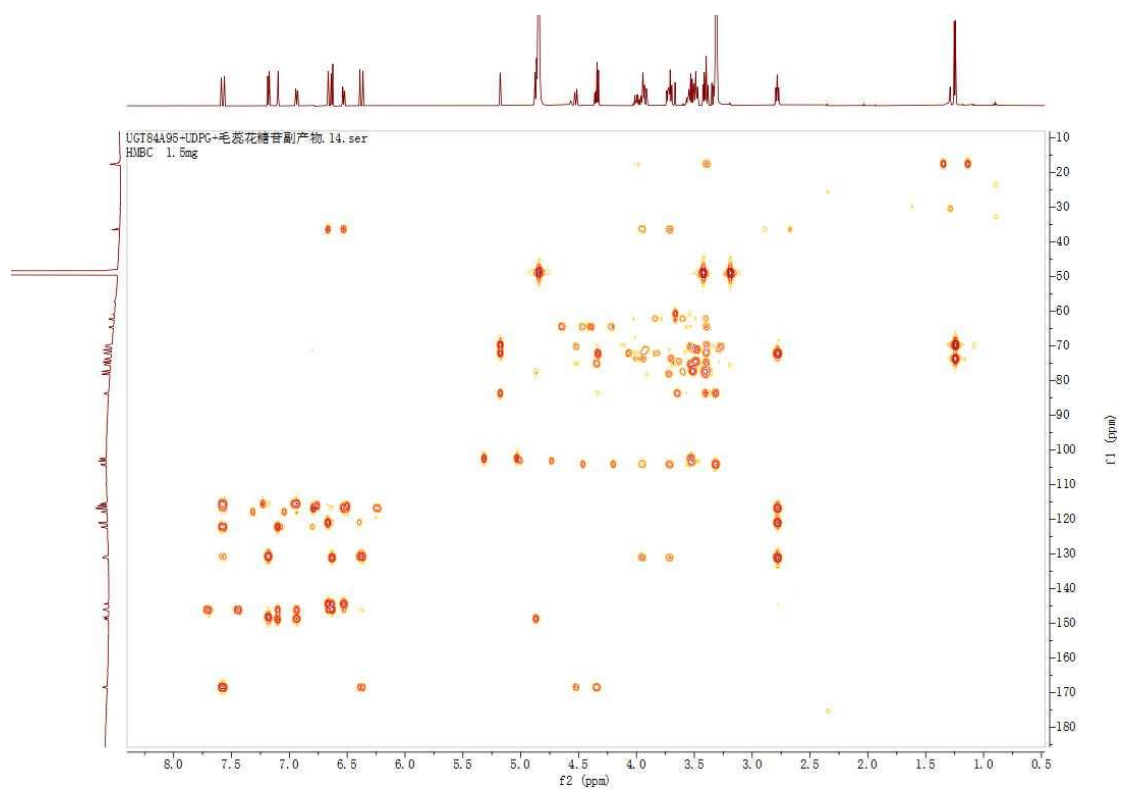


Figure S52. HMBC spectrum of glycosylated product **2b** (CD₃OD, 600 MHz).

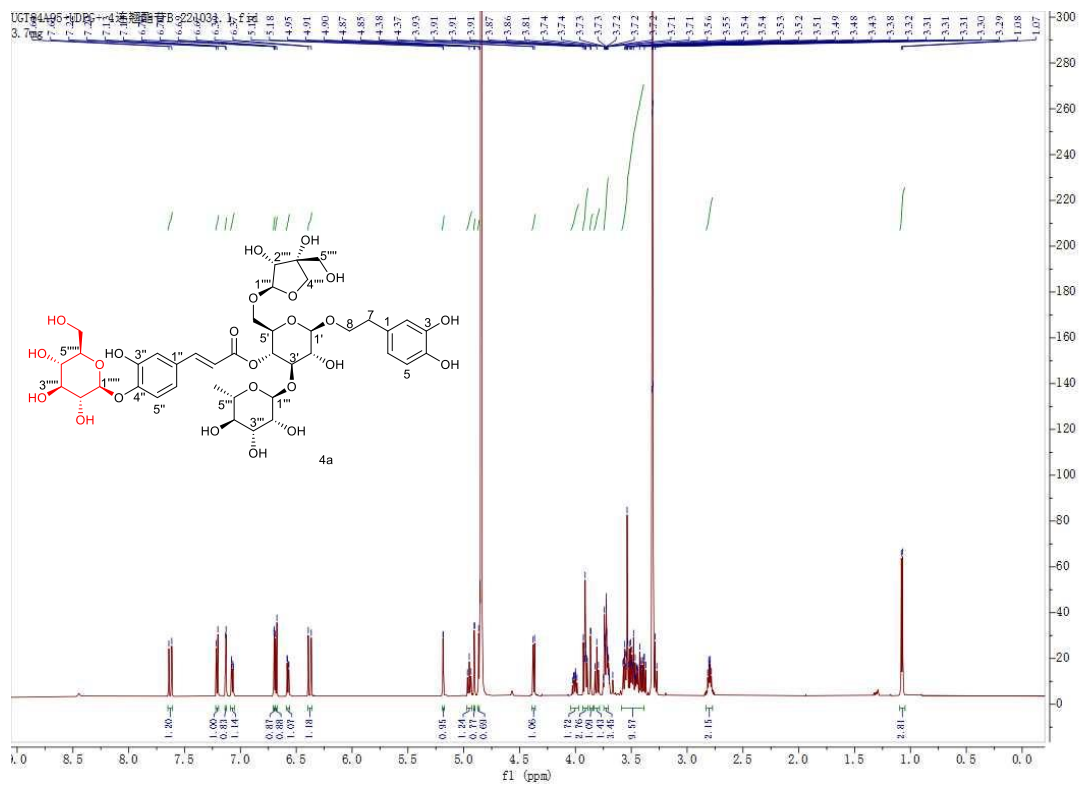


Figure S53. ¹H NMR spectrum of glycosylated product 4a (CD₃OD, 600 MHz).

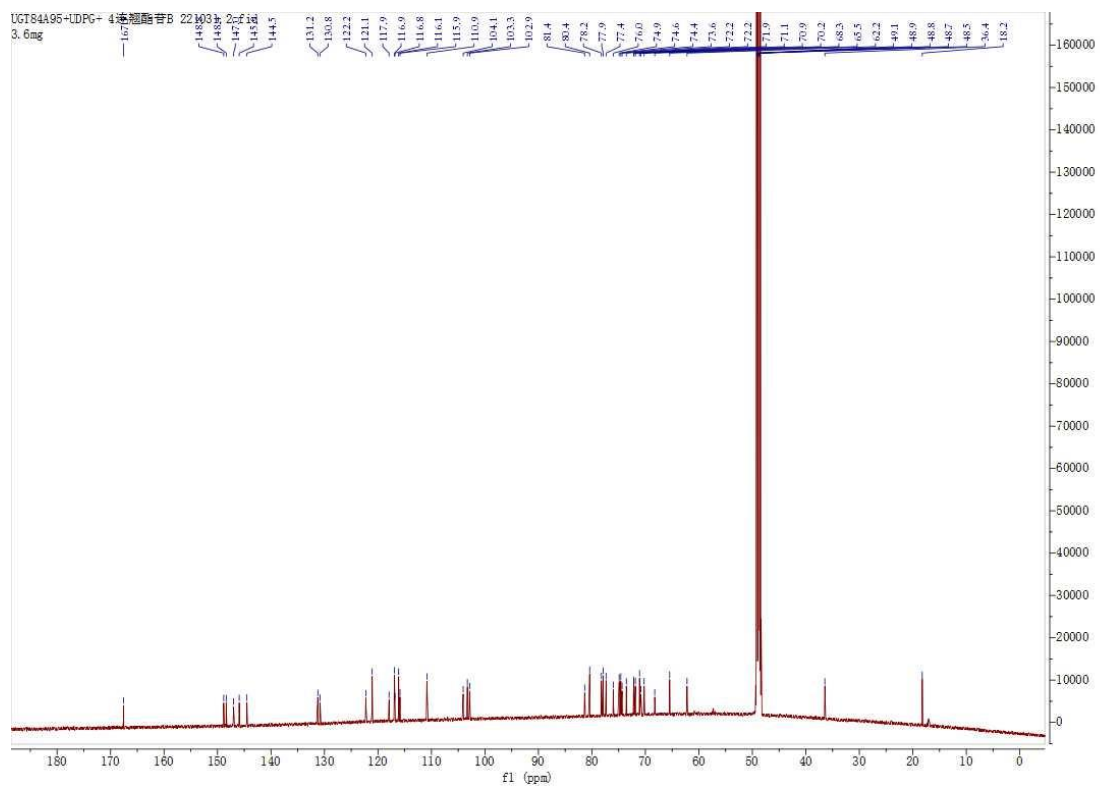


Figure S54. ¹³C NMR spectrum of glycosylated product 4a (CD₃OD, 600 MHz).

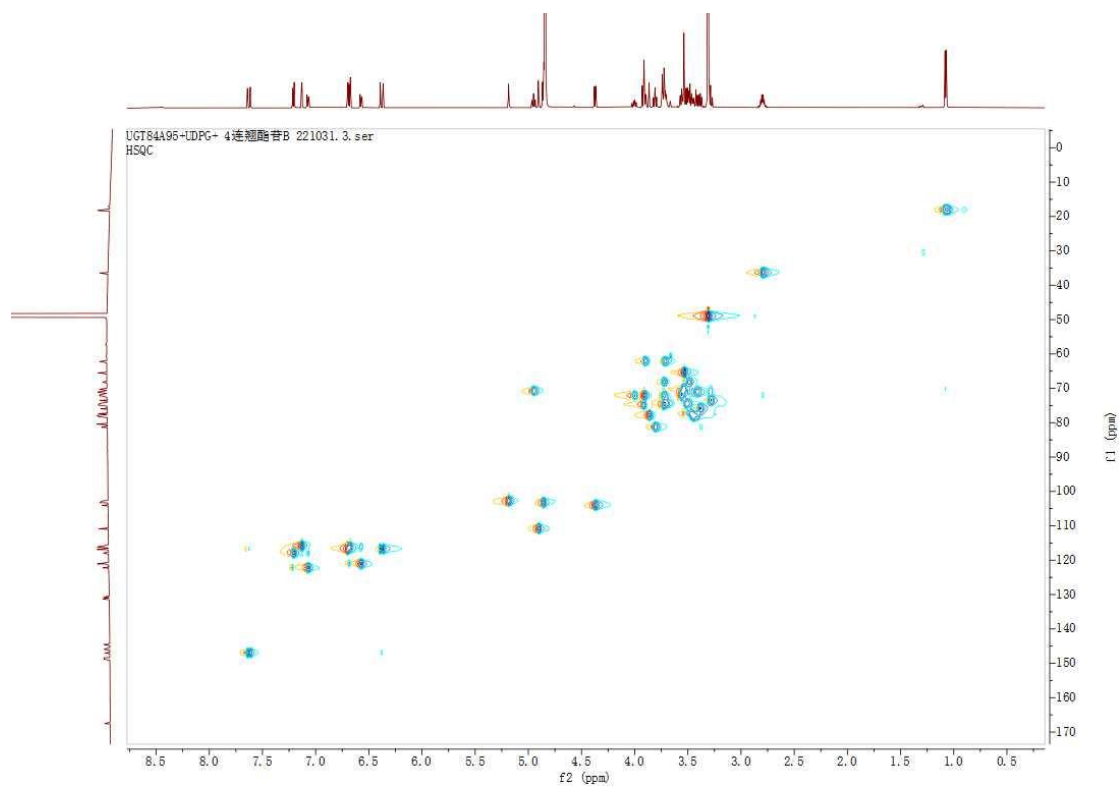


Figure S55. HSQC spectrum of glycosylated product **4a** (CD₃OD, 600 MHz).

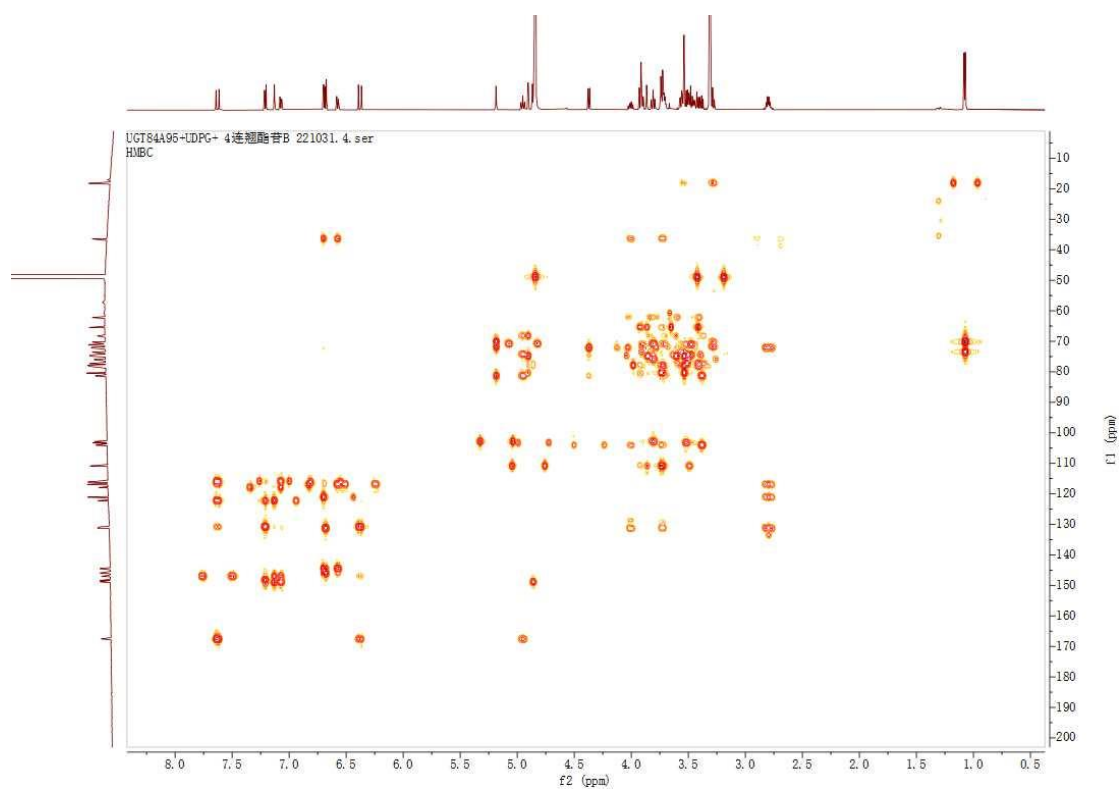
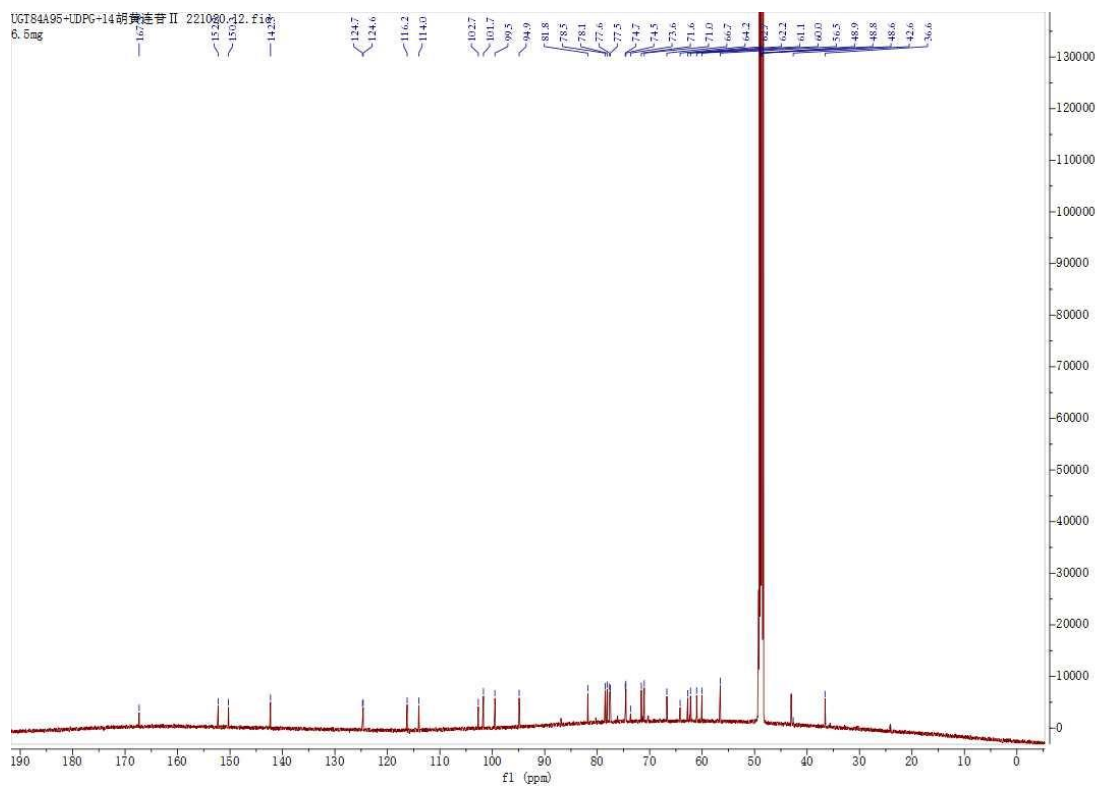
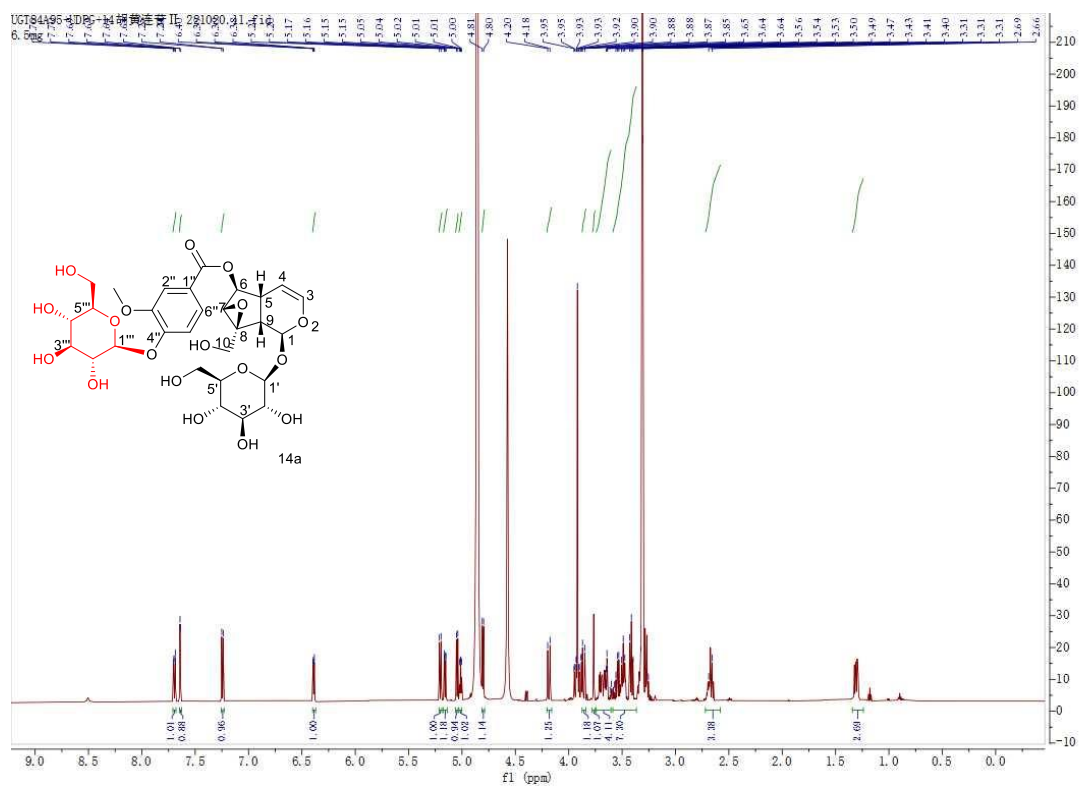


Figure S56. HMBC spectrum of glycosylated product **4a** (CD₃OD, 600 MHz).



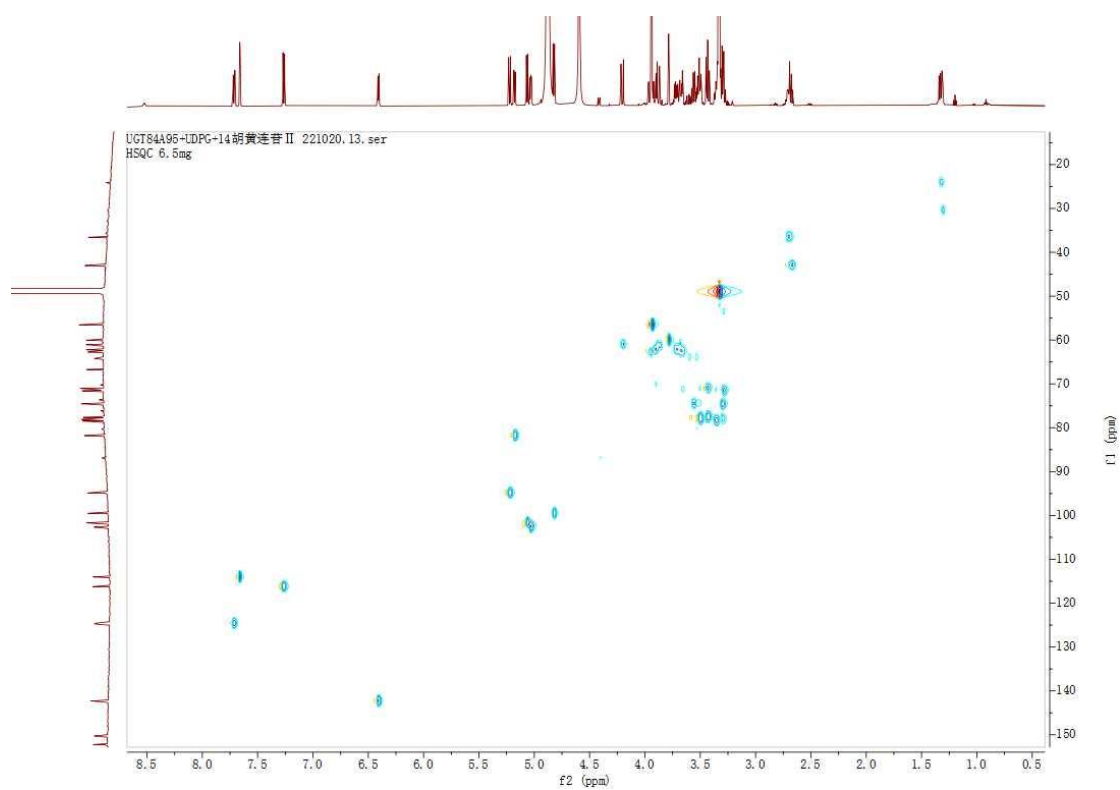


Figure S59. HSQC spectrum of glycosylated product **14a** (CD_3OD , 600 MHz).

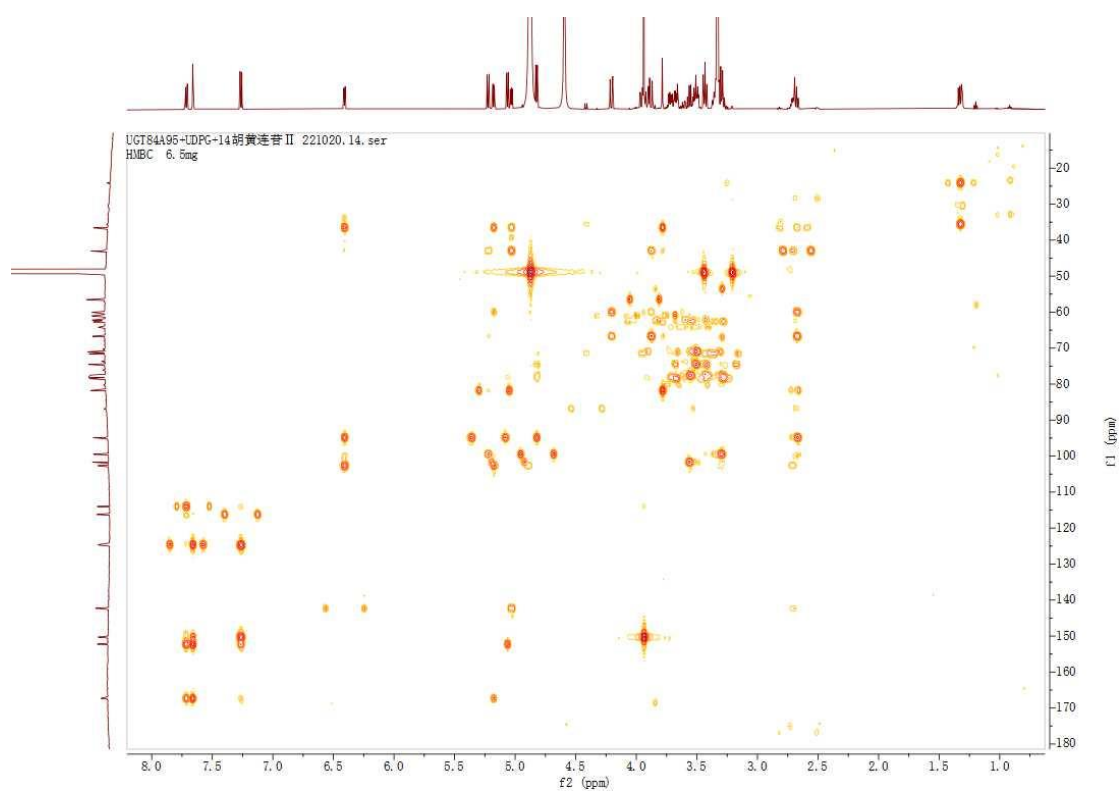
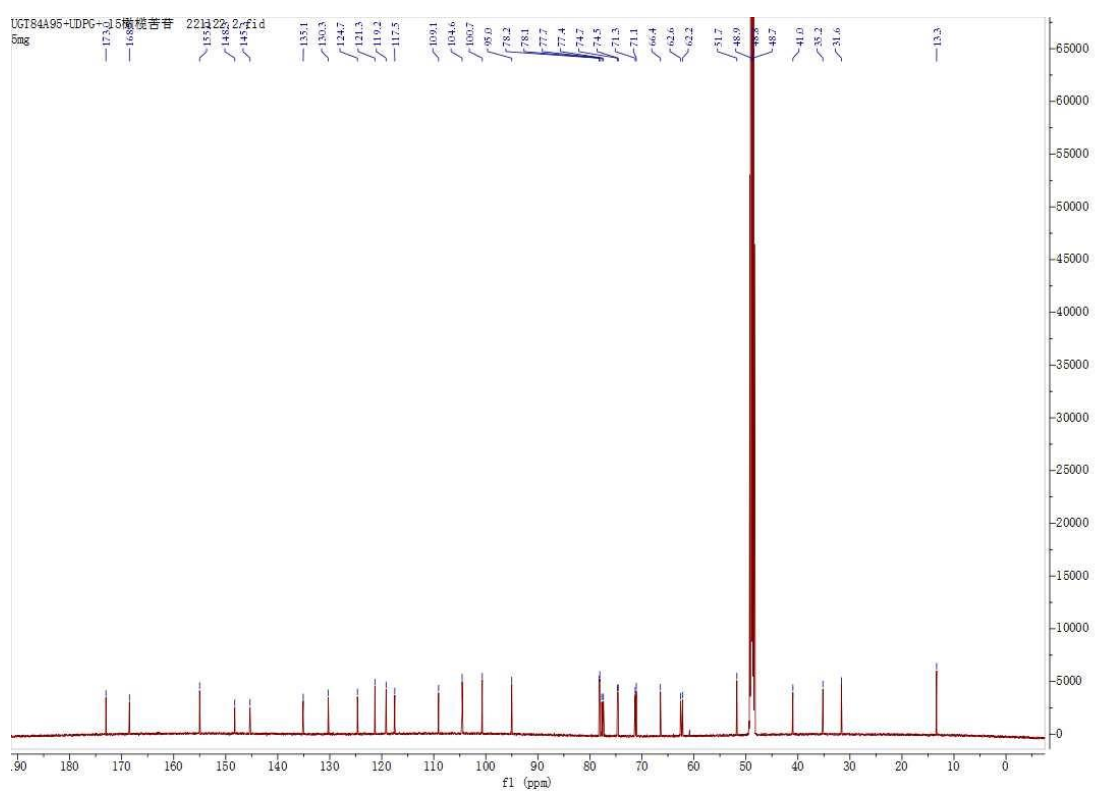
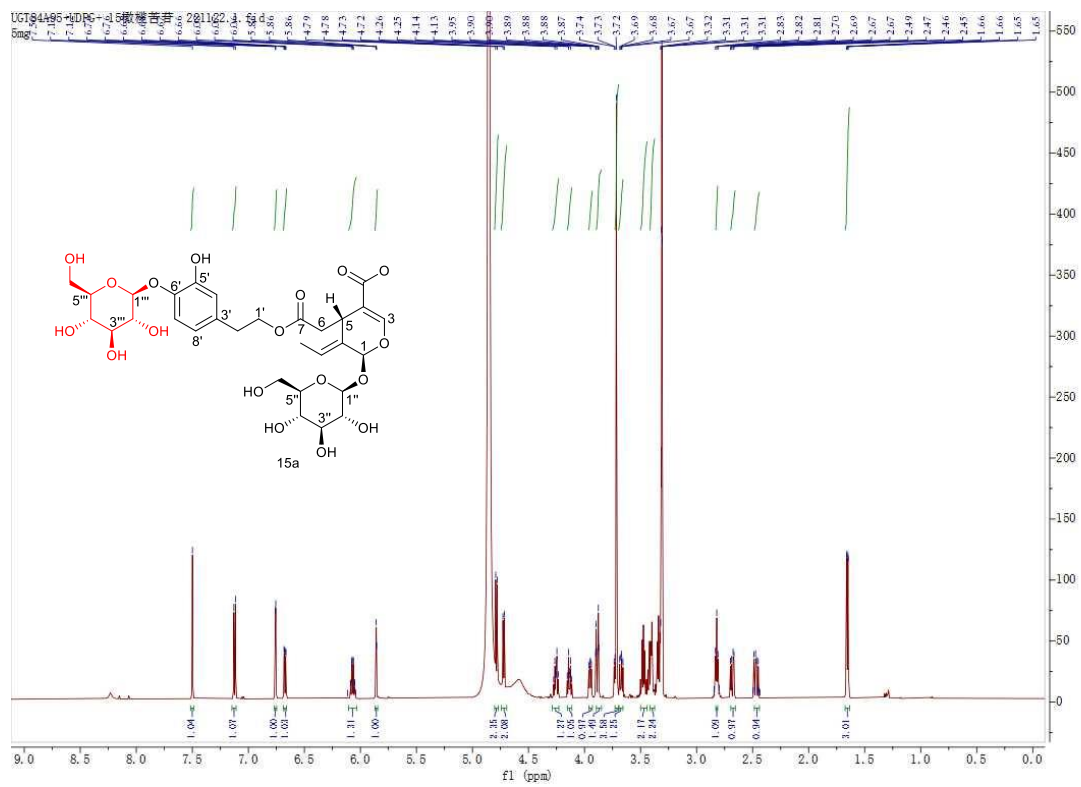


Figure S60. HMBC spectrum of glycosylated product **14a** (CD_3OD , 600 MHz).



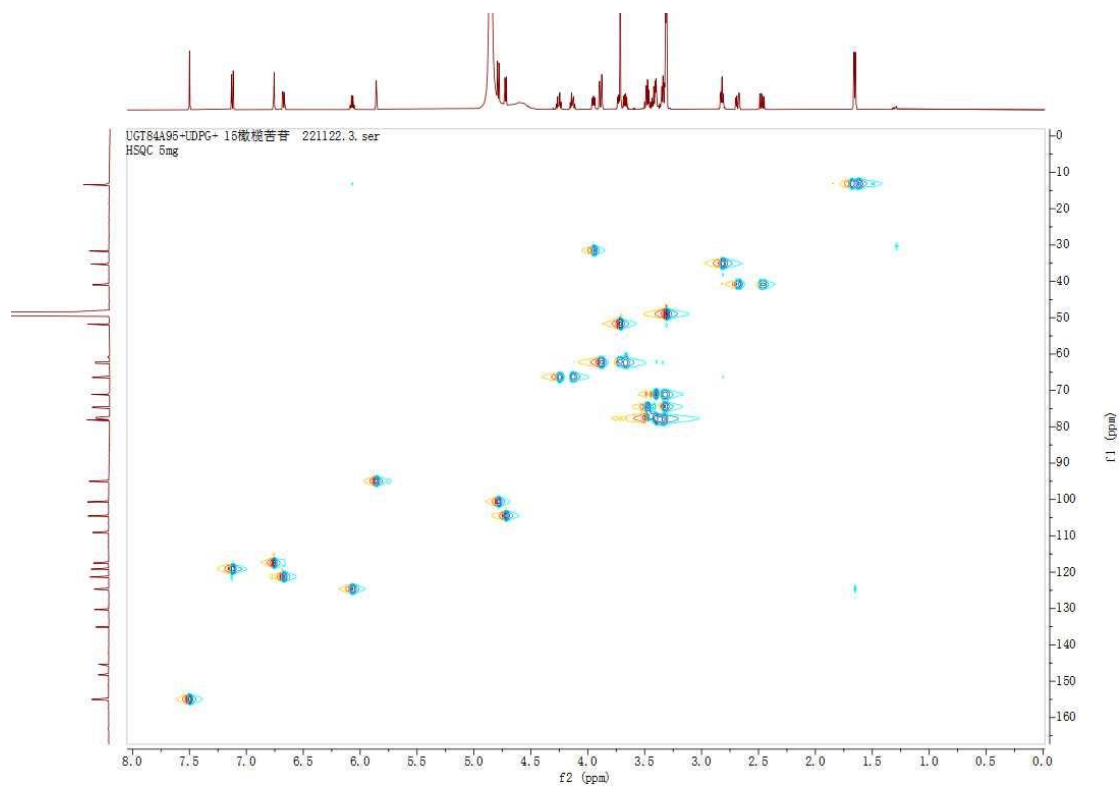


Figure S63. HSQC spectrum of glycosylated product **15a** (CD₃OD, 600 MHz).

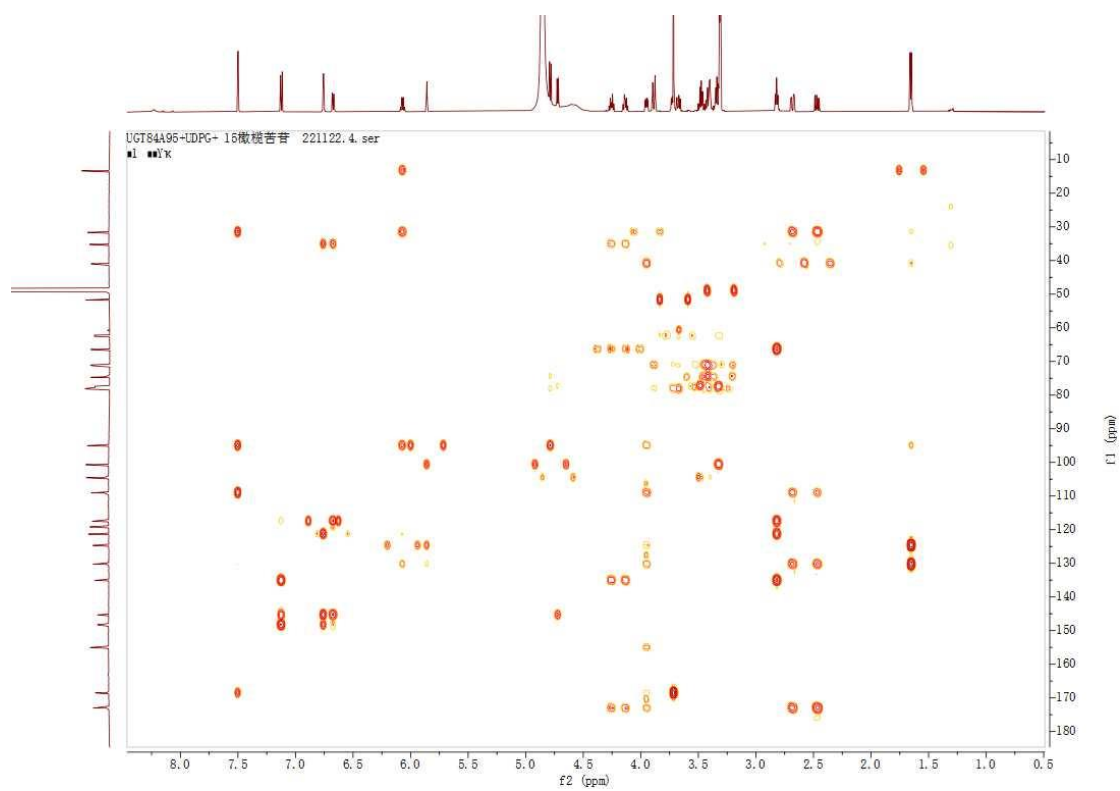


Figure S64. HMBC spectrum of glycosylated product **15a** (CD₃OD, 600 MHz).

3. Supplementary Note.

>UGT84A95

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