Supplementary Material (ESI) for Organic and Biomolecular Chemistry

Synthesis of Optically Active SARS-CoV-2 MPro inhibitor drug Nirmatrelvir (Paxlovid): an Approved Treatment of COVID-19

Arun K. Ghosh*,^{a,b} and Monika Yadav^a

^aDepartment of Chemistry and ^bDepartment of Medicinal Chemistry and Molecular

Pharmacology, Purdue University, 560 Oval Drive, West Lafayette, IN 47907.

Table of Contents

1.	General Experimental Procedures	S1- S2
2.	Copies of ¹ H and ¹³ C spectra of compounds	S3-S11
3.	HPLC Data	S12-13
4.	X-ray crystallographic data	S14-16
5.	References	S16

Experimental Procedures

General Methods. All chemicals and reagents were purchased from commercial suppliers and used without further purification unless otherwise noted. Tetrahydrofuran was distilled from Na/Benzophenone prior to use. All reactions were performed in oven-dried round-bottom flasks followed by flame-drying under vacuum in the case of moisture-sensitive reactions. The flasks were fitted with rubber septa and kept under a positive pressure of argon. Cannula were used in the transfer of moisture-sensitive liquids. Heated reactions were ran using an oil bath on a hot plate equipped with a temperature probe. TLC analysis was conducted using glass-backed thinlayer silica gel chromatography plates (60 Å, 250 μ m thickness, F-254 indicator). Flash

chromatography was done using a 230–400 mesh, a 60 Å pore diameter silica gel. Organic solutions were concentrated at 30–35 °C on rotary evaporators capable of achieving a minimum pressure of \sim 25 Torr and further concentrated on a Hi-vacuum pump capable of achieving a minimum pressure of ~4 Torr. ¹H NMR spectra were recorded on 400, 500, and 800 MHz spectrometers. ¹³C NMR spectra were recorded at 100, 125, and 200 MHz on the respective NMRs. Chemical shifts are reported in parts per million and referenced to the deuterated residual solvent peak (CDCl₃, 7.26 ppm for ¹H and 77.16 ppm for ¹³C). NMR data are reported as δ value (chemical shift), J-value (Hz), and integration, where s = singlet, bs = broad singlet, d = doublet, t = triplet, q = quartet, p = quintet, m = multiplet, dd = doublet doublets, and so on. Optical rotations were measured on a Rudolph's AUTOPOL-III automatic digital polarimeter with a sodium lamp and are reported as follows: $[\alpha]\lambda T ^{\circ}C$ (c = g/100 mL, solvent). Low resolution mass spectra (LRMS) spectra were recorded using a quadrupole LCMS under positive electrospray ionization (ESI+). High-resolution mass spectrometry (HRMS) spectra were recorded at the Purdue University Department of Chemistry Mass Spectrometry Center. These experiments were performed under ESI+ and positive atmospheric pressure chemical ionization (APCI+) conditions using an Orbitrap XL Instrument. HPLC analysis was done an on Agilent 1260 series instrument using a YMC Pack ODS-A (4.6x150mm) C-18 column and eluent: 60% MeOH/ H₂O.



Figure S1. ¹H NMR (400 MHz, DMSO- d_6) of amide **10**.





Figure S3. ¹H NMR (400 MHz, CDCl₃) of trifluoroacetamide epimer-8.



Figure S4. ¹³C NMR (100 MHz, CDCl₃) of trifluoroacetamide epimer-8.



Figure S5. ¹H NMR (400 MHz, CDCl₃) of trifluoroacetamide 8a.











Figure S9. ¹H NMR (400 MHz, CDCl₃) of benzamide **8b**.







Figure S11. ¹H NMR (400 MHz, CDCl₃) of *tert*-butyl amide **8c**.





Figure S13. ¹H NMR (400 MHz, CDCl₃) of trichloroacetamide 8d.







Figure S15. ¹H NMR (400 MHz, DMSO-*d*₆) of nirmatrelvir **1**.







Reverse Phase HPLC analysis of epimer 8

Sample Info : YMC PAK ODS-A 60% MeOH 5 uL inj. 0.5 mL/min 254 nm 215 nm 1 mg/mL



Signal 2: DAD1 B, Sig=215,4 Ref=off

Peak	RetTime	Туре	Width	Area	Height	Area	
#	[min]		[min]	[mAU*s]	[mAU]	%	
1	42.162	BB	1.5991	4164.03906	38.95371	55.1795	
2	48.018	BB	2.1042	3382.31860	22.93942	44.8205	

Note: Sample has no chromophore and hence it's detected only in 215nM wavelength. Retention time at 6.274 min

corresponds to solvent impurities.

Reverse Phase HPLC analysis of 8a

Sample Info : 60% MeOH/H20 0.5 mL/min 215 nm 5 uL inj. 1 mg/mL YMCPAK ODS-A





Signal 1: VWD1 A, Wavelength=215 nm

Peak #	RetTime [min]	Туре	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.755	BV R	0.1573	73.88456	6.93667	2.6129
2	10.727	BB	0.5976	131.00914	3.21387	4.6331
3	42.457	MM	1.0583	2533.55444	39.89836	89.5983
4	49.823	MM	0.9024	89.23389	1.64804	3.1557

Note: Sample has no chromophore and hence it's detected only in 215nM wavelength. Retention times at 5.7 min and 10.72 correspond to solvent impurities.

X-Ray Diffraction Data for acid 2

Single crystals of compound 2 were formed by slow evaporation of the solution of the compound 2 in 1 mL THF and 4 mL hexanes over a week. Examination and data collection were performed with Cu K α radiation ($\lambda = 1.54178$ Å) at 150 K. Data were collected, reflections were indexed and processed, and the files scaled and corrected for absorption using APEX3¹. The space groups were assigned, and the structures were solved by direct methods using XPREP within the SHELXTL suite of programs^{2,3} and refined by full matrix least squares against F2 with all reflections using Shelx12018⁴ using the graphical interface Shelxle. If not specified otherwise H atoms attached to carbon were positioned geometrically and constrained to ride on their parent atoms. C-H bond distances were constrained to 0.95 Å for aromatic and alkene C-H and CH2 and alkyne C-H moieties, and to 1.00, 0.99 and 0.98 Å for aliphatic C-H, CH2 and CH3 moieties, respectively. Methyl H atoms were allowed to rotate but not to tip to best fit the experimental electron density. $U_{iso}(H)$ values were set to a multiple of $U_{eq}(C)$ with 1.5 for CH3, and 1.2 for C-H and CH2 units, respectively. Additional data collection and refinement details can be found in the Supporting Information. Complete crystallographic data, in CIF format, have been deposited with the Cambridge Crystallographic Data Centre. CCDC 2181895 contains the supplementary crystallographic data for compound 2. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data request/cif. X-ray diffraction data were collected using an instrument funded by the NSF (CHE-1625543).

 Table 1. Crystal data and structure refinement for carboxylic acid 2.



Crystal data	
Chemical formula	C16H23F3N2O4
Mr	364.36
Crystal system, space group	Trigonal, R3:H
Temperature (K)	150
a, c (Å)	23.5990 (6), 10.4104 (4)
V (Å3)	5020.9 (3)
Ζ	9
F(000)	1728
Radiation type	Cu Ka
No. of reflections for cell measurement	9772
θ range (°) for cell measurement	3.8–72.5
μ (mm-1)	0.81
Crystal shape	Needle
Colour	Colorless
Crystal size (mm)	
Data collection	
Diffractometer	Bruker AXS D8 Quest
diffractometer with PhotonIII_C14 charge-integrating and photon counting pixel array detector	
Radiation source	I-mu-S microsource X-ray tube
Monochromator	Laterally graded multilayer (Goebel) mirror
Scan method	ω and phi scans
Absorption correction	Multi-scan
SADABS 2016/2: Krause, L., Herbst-Irmer, R., Sheldrick G.M. & Stalke D. (2015). J. Appl. Cryst. 48, 3-10.	
Tmin, Tmax	0.635, 0.754

No. of measured, independent and				
observed $[I > 2\sigma (I)]$ reflections				
Rint	0.046			
θ values (°)	$\theta_{\rm max}=72.5,\theta_{\rm min}=4.8$			
$(\sin \theta / \lambda)_{max}$ (Å-1)	0.619			
Range of h, k, l	$h = -29 \rightarrow 29, k = -29 \rightarrow 29, l = -9 \rightarrow 12$			
Refinement				
Refinement on	F2			
$R[F^2 > 2 \sigma(F^2)], wR(F^2), S$	0.026, 0.070, 1.06			
No. of reflections	3995			
No. of parameters	272			
No. of restraints	160			
H-atom treatment	H atoms treated by a mixture of independent and constrained refinement			
Weighting scheme	$w = 1/[\sigma^2(F_o^2) + (0.0429P)^2 + 0.6317P]$ where $P = (F_o^2 + 2F_c^2)/3$			
$(\Delta/\sigma)_{max}$	0.009			
$\Delta \square_{\max}, \Delta \square_{\min} (e \text{ Å}^{-3})$	0.10, -0.10			
Extinction method	SHELXL2018/3 (Sheldrick 2018), Fc*=kFc[1+0.001xFc ² λ^{3} /sin(20)] ^{-1/4}			
Extinction coefficient	0.00030 (6)			
Absolute structure	Flack x determined using 1676 quotients [(I+)-(I-)]/[(I+)+(I-)] (Parsons, Flack and Wagner, Acta Cryst. B69 (2013) 249-259).			
Absolute structure parameter	-0.09 (6)			

References

- 1. Bruker (2016). Apex3 v2016.9-0, Saint V8.34A, SAINT V8.37A, Bruker AXS Inc.: Madison (WI), USA, 2013/2014.
- 2. SHELXTL suite of programs, Version 6.14, 2000-2003, Bruker Advanced X-ray Solutions, Bruker AXS Inc., Madison, Wisconsin: USA.
- 3. Sheldrick, G. Acta Crystallogr A 64 (1), 112 (2008).
- 4. Sheldrick, G. Acta. Crystallogr. Sect. C. Struct. Chem. 71 (1), 3 (2015).