Selective synthesis of an elusive *C*-functionalized bis-cyclam and study of its inhibition properties of CXCR4 chemokine receptor

Marie M. Le Roy,^a Sandra Claes,^b Nathalie Saffon-Merceron, ^c Dominique Schols,^{*c} Thibault Troadec,^{*a} and Raphaël Tripier^a

Addresses :

^a Univ. Brest, UMR CNRS 6521 CEMCA, 6 Avenue Victor le Gorgeu, 29200 Brest, France.

^b Rega Institute for Medical Research, KU Leuven, B-3000 Leuven, Belgium

^c Institut de Chimie de Toulouse (FR 2599), 118 route de Narbonne, 31062 Toulouse Cedex 9, France

email : thibault.troadec@univ-brest.fr

SUPPORTING INFORMATION

Table of content

EXPERIMENTAL DETAILS	3
Synthesis	3
Nuclear Magnetic Resonance Spectroscopy	3
Mass Spectrometry	3
Liquid Chromatography (HPLC)	3
SYNTHETIC PROTOCOLS AND CHARACTERIZATION DATA FOR COMPOU	NDS
1-10	4
SPECTRAL DATA	7
COMPOUND 1:	7
Figure S1: 1H (400 MHz, CDCl3, 298K, TMS) and 13C Jmod (75 MHz, CDCl3, 298K, TMS) N	MR 7
Figure S2: HRMS spectrum (ESI)	8
COMPOUND 2:	9
Figure S3: ¹ H (300 MHz, CDCl ₃ , 298K, TMS) and ¹³ C Jmod (75 MHz, CDCl ₃ , 298K, TMS) NM	ЛR9
Figure S4: HRMS spectrum (ESI)	10

COMPOUND 4:	11
Figure S5: ¹ H (300 MHz, CDCl ₃ , 298K, TMS) and ¹³ C Jmod (75 MHz, CDCl ₃ , 298K, TMS) NMR	11
Figure S6: HRMS spectrum (ESI)	12
Table S1: X-ray data of compound 4	13
	4 4
	14
Figure S7: ¹ H (400 MHz, CDCl ₃ , 298K, TMS) and ¹³ C Jmod (125 MHz, CDCl ₃ , 298K, TMS) NMF	14
Figure S8: HRMS spectrum (ESI)	15
COMPOUND 6:	16
Figure S9: ¹ H (400 MHz, MeOD, 298K, TMS) and ¹³ C Jmod (125 MHz, MeOD, 298K, TMS) NM	२
	16
Figure S10: HRMS spectrum (ESI) of compound 6	17
	40
	10
Figure S11: ¹ H (400 MHz, CDCl ₃ , 298K, TMS) and ¹³ C Jmod (125 MHz, CDCl ₃ , 298K, TMS) NM	R
	18
COMPOUND 9 (MIXTURE OF DIASTEREOMERS):	19
COMPOUND 9 (MIXTURE OF DIASTEREOMERS): Figure S12: ¹ H (500 MHz, CDCl ₃ , 298K, TMS) and ¹³ C Jmod (125 MHz, CDCl ₃ , 298K, TMS) NM	19 R
COMPOUND 9 (MIXTURE OF DIASTEREOMERS): Figure S12: ¹ H (500 MHz, CDCl ₃ , 298K, TMS) and ¹³ C Jmod (125 MHz, CDCl ₃ , 298K, TMS) NM	19 R 19
COMPOUND 9 (MIXTURE OF DIASTEREOMERS): Figure S12: ¹ H (500 MHz, CDCl ₃ , 298K, TMS) and ¹³ C Jmod (125 MHz, CDCl ₃ , 298K, TMS) NM Figure S13: HRMS spectrum (ESI)	19 R 19 20
COMPOUND 9 (MIXTURE OF DIASTEREOMERS): Figure S12: ¹ H (500 MHz, CDCI ₃ , 298K, TMS) and ¹³ C Jmod (125 MHz, CDCI ₃ , 298K, TMS) NM Figure S13: HRMS spectrum (ESI)	19 R 19 20 21
COMPOUND 9 (MIXTURE OF DIASTEREOMERS): Figure S12: ¹ H (500 MHz, CDCI ₃ , 298K, TMS) and ¹³ C Jmod (125 MHz, CDCI ₃ , 298K, TMS) NM Figure S13: HRMS spectrum (ESI). COMPOUND 10 (C,C'-(M-XYLYLENE)BIS-CYCLAM). Figure S14: ¹ H (400 MHz, DrO, 208K, TMS) and ¹³ C, Imod (125 MHz, DrO, 208K, TMS) NMP	19 R 19 20 21
 COMPOUND 9 (MIXTURE OF DIASTEREOMERS): Figure S12: ¹H (500 MHz, CDCl₃, 298K, TMS) and ¹³C Jmod (125 MHz, CDCl₃, 298K, TMS) NM Figure S13: HRMS spectrum (ESI) COMPOUND 10 (<i>C</i>,<i>C'</i>-(<i>M</i>-XYLYLENE)BIS-CYCLAM) Figure S14: ¹H (400 MHz, D₂O, 298K, TMS) and ¹³C Jmod (125 MHz, D₂O, 298K, TMS) NMR Figure S15: HPMS spectrum (ESI) 	19 R 19 20 21 21
 COMPOUND 9 (MIXTURE OF DIASTEREOMERS): Figure S12: ¹H (500 MHz, CDCl₃, 298K, TMS) and ¹³C Jmod (125 MHz, CDCl₃, 298K, TMS) NM Figure S13: HRMS spectrum (ESI) COMPOUND 10 (C,C'-(M-XYLYLENE)BIS-CYCLAM) Figure S14: ¹H (400 MHz, D₂O, 298K, TMS) and ¹³C Jmod (125 MHz, D₂O, 298K, TMS) NMR Figure S15: HRMS spectrum (ESI) 	19 R 19 20 21 21 22
 COMPOUND 9 (MIXTURE OF DIASTEREOMERS): Figure S12: ¹H (500 MHz, CDCl₃, 298K, TMS) and ¹³C Jmod (125 MHz, CDCl₃, 298K, TMS) NM Figure S13: HRMS spectrum (ESI). COMPOUND 10 (C,C'-(M-XYLYLENE)BIS-CYCLAM). Figure S14: ¹H (400 MHz, D₂O, 298K, TMS) and ¹³C Jmod (125 MHz, D₂O, 298K, TMS) NMR Figure S15: HRMS spectrum (ESI) Figure S16: HPLC-ELSD chromatogram. Mobile phase H₂O + 0.1% formic acid / Acetonitrile (95: for 16 mins, gradient to 10:90 in 6 mins + 2 mins at 10:90, gradient to 95:5 in 6 mins) 	19 R 19 20 21 21 22 5 5
COMPOUND 9 (MIXTURE OF DIASTEREOMERS): Figure S12: ¹ H (500 MHz, CDCl ₃ , 298K, TMS) and ¹³ C Jmod (125 MHz, CDCl ₃ , 298K, TMS) NM Figure S13: HRMS spectrum (ESI) COMPOUND 10 (<i>C</i> , <i>C'</i> -(<i>M</i> -XYLYLENE)BIS-CYCLAM). Figure S14: ¹ H (400 MHz, D ₂ O, 298K, TMS) and ¹³ C Jmod (125 MHz, D ₂ O, 298K, TMS) NMR Figure S15: HRMS spectrum (ESI) Figure S16: HPLC-ELSD chromatogram. Mobile phase H ₂ O + 0.1% formic acid / Acetonitrile (95: for 16 mins, gradient to 10:90 in 6 mins + 2 mins at 10:90, gradient to 95:5 in 6 mins)	19 R 19 20 21 21 22 5 23
 COMPOUND 9 (MIXTURE OF DIASTEREOMERS):	 19 R 19 20 21 21 22 5 23 24
 COMPOUND 9 (MIXTURE OF DIASTEREOMERS):	 19 R 19 20 21 22 21 22 23 24
 COMPOUND 9 (MIXTURE OF DIASTEREOMERS): Figure S12: ¹H (500 MHz, CDCl₃, 298K, TMS) and ¹³C Jmod (125 MHz, CDCl₃, 298K, TMS) NM Figure S13: HRMS spectrum (ESI). COMPOUND 10 (<i>C</i>, <i>C'</i>-(<i>M</i>-XYLYLENE)BIS-CYCLAM). Figure S14: ¹H (400 MHz, D₂O, 298K, TMS) and ¹³C Jmod (125 MHz, D₂O, 298K, TMS) NMR Figure S15: HRMS spectrum (ESI). Figure S16: HPLC-ELSD chromatogram. Mobile phase H₂O + 0.1% formic acid / Acetonitrile (95: for 16 mins, gradient to 10:90 in 6 mins + 2 mins at 10:90, gradient to 95:5 in 6 mins) BIOLOGICAL ASSAYS Table S2: Calcium Signalling Assays (3 replicates, U87.CD4.CXCR4 cells). Figure S17: Calcium Signalling Assays (U87.CD4.CCR5 cells) 	 19 R 19 20 21 21 22 5 23 24 25
 COMPOUND 9 (MIXTURE OF DIASTEREOMERS):	 19 R 20 21 22 23 24 24 25 26
COMPOUND 9 (MIXTURE OF DIASTEREOMERS): Figure S12: ¹ H (500 MHz, CDCl ₃ , 298K, TMS) and ¹³ C Jmod (125 MHz, CDCl ₃ , 298K, TMS) NM Figure S13: HRMS spectrum (ESI) COMPOUND 10 (<i>C</i> , <i>C'</i> -(<i>M</i> -XYLYLENE)BIS-CYCLAM) Figure S14: ¹ H (400 MHz, D ₂ O, 298K, TMS) and ¹³ C Jmod (125 MHz, D ₂ O, 298K, TMS) NMR Figure S15: HRMS spectrum (ESI) Figure S16: HPLC-ELSD chromatogram. Mobile phase H ₂ O + 0.1% formic acid / Acetonitrile (95: for 16 mins, gradient to 10:90 in 6 mins + 2 mins at 10:90, gradient to 95:5 in 6 mins) BIOLOGICAL ASSAYS Table S2: Calcium Signalling Assays (3 replicates, U87.CD4.CXCR4 cells) Figure S18: CXCL12 inhibition Assay (Jurkat cells) Table S3: Monoclonal Antibodies (mAbs) binding assays (IC ₅₀ values, Jurkat cells)	 19 R 20 21 21 22 5 23 24 25 26 26
 COMPOUND 9 (MIXTURE OF DIASTEREOMERS): Figure S12: ¹H (500 MHz, CDCl₃, 298K, TMS) and ¹³C Jmod (125 MHz, CDCl₃, 298K, TMS) NM Figure S13: HRMS spectrum (ESI). COMPOUND 10 (<i>C</i>, <i>C'</i>-(<i>M</i>-XYLYLENE)BIS-CYCLAM). Figure S14: ¹H (400 MHz, D₂O, 298K, TMS) and ¹³C Jmod (125 MHz, D₂O, 298K, TMS) NMR Figure S15: HRMS spectrum (ESI). Figure S16: HPLC-ELSD chromatogram. Mobile phase H₂O + 0.1% formic acid / Acetonitrile (95: for 16 mins, gradient to 10:90 in 6 mins + 2 mins at 10:90, gradient to 95:5 in 6 mins) BIOLOGICAL ASSAYS Table S2: Calcium Signalling Assays (3 replicates, U87.CD4.CXCR4 cells). Figure S18: CXCL12 inhibition Assay (Jurkat cells). Table S3: Monoclonal Antibodies (mAbs) binding assays. 	 19 R 19 20 21 21 22 5 23 24 25 26 27

Experimental details

Synthesis. Reagent used for synthesis were purchased from SIGMA-ALDRICH[®], TCI chemicals[®], ACROS ORGANICS[®] and Ambeed[®] and used without further purification. Solvents for synthesis were obtained from a MBraun MB-SPS 800 purification system. Ultrapure water was freshly obtained from a Milli-Q dispenser.

Compounds 3 and 8 were synthesized according to previously reported procedures. 38,32

Nuclear Magnetic Resonance Spectroscopy. NMR data were recorded at the "Service Général des Plateformes (SGPLAT)" of the Université de Bretagne Occidentale. ¹H, ¹³C and 2D NMR spectra were recorded on a Bruker Avance III HD 500 (500.25 MHz for ¹H and 125.79 MHz for ¹³C), Bruker Avance 400 (400.13 MHz for ¹H and 100.62 MHz for ¹³C) or Bruker AMX-3 300 (300.13 MHz for ¹H and 75.47 MHz for ¹³C) spectrometers. Deuterated solvents from Eurisotop[®] are used to reference spectra, ¹H and ¹³C shifts are reported in ppm and the δ scales are relative to TMS.

The signals are indicated as follows: chemical shift, multiplicity (s for singlet; br for broad singlet, d for doublet; t for triplet; q for quadruplet; m for multiplet), coupling constants J in Hertz (Hz), assignment: $CH_2\alpha N$, $CH_2\beta N$ or $CH_2\gamma N$ correspond to CH_2 located in alpha, beta or gamma position of considered nitrogen atom, type of nuclei is indicated in italic. Ar is a generic term used in subscript for all H or C aromatic atoms.

Mass Spectrometry. High-Resolution Mass Spectrometry (HRMS) analyses were performed at the Institute of Organic and Analytic Chemistry (ICOA Orléans) on a HRMS Q-Tof MaXis, sources ESI, APCI, APPI and nano-ESI. MALDI TOF were recorded at the "service commun" of the Université de Bretagne Occidentale using MALDI TOF-TOF AuTOFLEX III from Bruker Daltonics.

Liquid Chromatography (HPLC). HPLC analysis was carried out on Shimadzu LD20 system equipped with an Agilent Zorbax SB-C18 column (4.6 x 250 mm, 5-micron) and a Shimadzu ELSD LTII detector (80° C, N₂ as nebulizing gas, 350kPa). Elution with H₂O (0.1% formic acid)/Acetonitrile was performed with 30 minutes program set up as: 95:5 for 16 mins, gradient to 10:90 in 6 mins + 2 mins at 10:90, gradient to 95:5 in 6 mins). Analysis and data were processed with LabSolutions software.

Synthetic protocols and characterization data for compounds 1-10

Synthesis of 1: Isophtaldehyde (1.000 g, 7.46 mmol), Meldrum Acid (2.146 g, 14.9 mmol) and Hantzch ester (3.777 g, 14.9 mmol) were solubilized in MeOH (26 mL). Then, *L*-proline (0.172 g, 1.49 mmol, 20 %) was added and the reaction mixture was stirred at 25 °C for 12 h. The precipitate was filtered and washed with diethyl ether. The crude product was purified by column chromatography on silica gel (Hexane/Ethyl acetate/Methanol 50:50:0 to 0:90:10) to afford compound **1** as a white powder (2.460 g, 85 %).

¹*H NMR* (CDCl₃, 400 MHz, 298 K) δ (ppm) 7.24-7.17 (m, 4H, *CH*_{Ar}), 3.77 (t, 2H, *CH*, J = 5.21 Hz), 3.44 (d, 4H, *CH*₂, J = 5.21 Hz), 1.52 (s, 6H, *CH*₃), 1.55 (s, 6H, *CH*₃). ¹³*C Jmod NMR* (CDCl₃, 75 MHz, 298 K) δ (ppm) 165.3 (*C*0), 137.8 (*C*_{ipso}), [130.7, 129.0, 128.6 (x2)] (*CH*_{Ar}), 105.4 (*C*_{sp}), 48.2 (*C*H), 31.9 (*C*H₂), [28.6, 27.2] (*C*H₃). *HRMS* (ESI, positive, H₂O) m/z calcd. for [C₂₀H₂₆NO₈]⁺ 408.1653 found [M+NH₄]⁺ 408.1651, calcd. for [C₂₀H₂₂NaO₈]⁺ 413.1207 found [M+Na]⁺ 413.1207. *Mp*: 180 °C

Synthesis of 2: Compound **1** (2.460 g, 6.30 mmol) and Eschenmoser's salt (5.831 g, 31.5 mmol) were solubilized in MeOH (25 mL), and the reaction was stirred at 65 °C for 12 h. After cooling to room temperature, the solvent was removed under vacuum. The crude product was dissolved in CHCl₃ (100 mL) and washed with NaHCO_{3(sat)} (50 mL), KHSO_{4(10 % weight)} (50 mL) and NaCl_(sat) (50 mL). The organic layer was dried over MgSO₄, filtered, and concentrated *in vacuo* to afford compound **2** as an orange oil (1.674 g, 97 %).

¹*H NMR* (CDCl₃, 300 MHz, 298 K) δ (ppm) 7.22-7.19 (m, 1H, CH_{Ar}), 7.06-7.03 (m, 3H, CH_{Ar}), 6.22 (s, 2H, =CH), 5.43 (s, 2H, =CH), 3.73 (s, 6H, CH₃), 3.60 (s, 4H, CH₂). ¹³*C Jmod NMR* (CDCl₃, 75 MHz, 298 K) δ (ppm) 167.3 (CO), [140.1, 138.8] (C_{ipso}), [129.8, 128.5, 127.1 (x2)] (CH_{Ar}), 126.2 (=CH₂), 51.8 (CH₃), 37.9 (CH₂Ar). *HRMS* (ESI, positive, H₂O) m/z calcd. for [C₁₆H₁₉O₄]⁺ 275.1278 found [M+H]⁺ 275.1281, calcd. for [C₁₂H₂₂NO₄]⁺ 292.1543 found [M+NH₄]⁺ 292.1546, calcd. for [C₁₆H₁₈NaO₄]⁺ 297.1097 found [M+Na]⁺ 297.1099

Synthesis of 4: A solution of **2** (0.387 g, 1.41 mmol) in ethyl acetate (2 mL) was added dropwise to a solution of **3** (0.514 g, 2.82 mmol) in ethyl acetate (5 mL). The reaction was stirred at 25 °C for 4 months. The solvent was removed under reduced pressure at room temperature and the crude product was precipitated with THF. The solid was filtered off, washed with THF and dried under reduced pressure to afford compound **4** as a white solid (0.243 g, 30 %).

¹*H NMR* (CDCl₃, 300 MHz, 298 K) δ (ppm) 7.19-7.12 (m, 1H, CH_{Ar}), 7.01-6.93 (m, 3H, CH_{Ar}), 4.50 (d, 2H, CH, J = 13.10 Hz), 4.32 (d, 2H, N-CH-N, J = 2.5 Hz), 3.54-3.41 (m, 4H, CH₂-Ar), 3.22 (d, 2H, N-CH-N, J = 2.5 Hz), 3.19-2.09 (m, 30H, CH₂ α N), 1.37-1.23 (m, 2H, CH₂ β N). ¹³*C Jmod NMR* (CDCl₃, 75 MHz, 298 K) δ (ppm) 171.2 (CO), 139.7 (C_{ipso}), [129.6, 128.7, 127.0] (CH_{Ar}), [76.0, 70.6] (CH_{cis}), 55.7 (CH₂Ar), [54.1, 53.2, 52.9, 44.3, 44.2, 40.2] (CH₂ α N), 37.0 (CH), 35.3 (CH₂ α N), 19.5 (CH₂ β N). *HRMS* (ESI, positive, H₂O)

m/z calcd. for $[C_{32}H_{47}N_8O_2]^+$ 575.3816 found $[M+H]^+$ 575.3819, calcd. for $[C_{32}H_{46}N_8NaO_2]^+$ 597.3636 found $[M+Na]^+$ 597.3640, calcd. for $[C_{32}H_{48}N_8O_2]^{2+}$ 288.1945 found $[M+2H]^{2+}$ 288.1949

Synthesis of 5: A suspension of NaH (0.5018 g, 20.91 mmol) in THF (10 mL) was cooled to 0 °C and diethyl malonate (3.257 g, 20.4 mmol) was added dropwise and stirring was continued until the solution became clear. Then *m*-dibromoxylene (1.500 g, 5.10 mmol) was added, and the solution was allowed to warm to room temperature and stirred for 1 h, then heated for 18 h at 50 °C. The reaction mixture was quenched with $NH_4Cl_{(sat)}$ and the product was extracted from the aqueous layer with ethyl acetate. The crude product was purified by flash chromatography on silica gel (Hexane/EtOAc, 100:0 to 0:100) to afford compound **5** as a colorless oil (1.1674 g, 54 %).

¹*H NMR* (CDCl₃, 400 MHz, 298 K) δ (ppm) 7.20-7.15 (m, 1H, CH_{Ar}), 7.06-7.03 (m, 3H, CH_{Ar}), 4.20-4.10 (m, 8H, O-CH₂-CH₃), 3.59 (t, 2H, CH, J = 7.8 Hz), 3.16 (d, 4H, CH₂Ar, J = 7.8 Hz), 1.20 (t, 12H,O-CH₂-CH₃, J = 7.2 Hz). ¹³*C Jmod NMR* (CDCl₃, 125 MHz, 298 K) δ (ppm) 168.9 (CO), 138.3 (C_{ipso}), [129.4, 128.7, 127.3] (CH_{Ar}), 61.5 (O-CH₂-CH₃), 53.9 (CH), 34.6 (CH₂Ar), 14.1 (O-CH₂-CH₃). *HRMS* (ESI, positive, H₂O) m/z calcd. for [C₂₂H₃₁O₈]⁺ 423.2013 found [M+H]⁺ 423.2015, calcd. for [C₂₂H₃₀NaO₈]⁺ 445.1832 found [M+Na]⁺ 445.1837, calcd. for [C₂₂H₃₀KO₈]⁺ 461.1572 found [M+K]⁺ 461.1573

Synthesis of 6: A suspension of LiAlH₄ (0.7329 g, 19.4 mmol) in THF (25 mL) was cooled to -10 °C and a solution of **5** (1.3635 g, 3.23 mmol) in THF (7 mL) was added dropwise. The reaction mixture was warmed slowly to room temperature (~2 h) and heated to 50 °C for 18 h. Celite^{*} and Et₂O were added before the addition of H₂O (1 mL), HCl (3M, 1 mL), H₂O (1 mL) and the mixture was filtered off. The filtrate was concentrated under reduced pressure to afford compound **6** as a colorless oil (0.7443 g, 91 %).

¹*H NMR* (MeOD, 400 MHz, 298 K) δ (ppm) 7.21-7.16 (m, 1H, CH_{Ar}), 7.09-7.01 (m, 3H, CH_{Ar}), 3.54 (d, 8H, CH₂-OH, J = 5.7 Hz), 2.62 (d, 4H, CH₂-Ar, J = 7.3 Hz), 1.94-1.86 (q, 2H, CH, J = 7.3, 5.7 Hz). ¹³C Jmod NMR (MeOD, 125 MHz, 298 K) δ (ppm) 141.8 (C_{ipso}), [131.1, 129.2, 127.8] (CH_{Ar}), 63.1 (CH₂-OH), 46.6 (CH), 35.1 (CH₂-Ar). *HRMS* (ESI, positive, H₂O) m/z calcd. for [$C_{14}H_{23}O_4$]⁺ 255.1591 found [M+H]⁺ 255.1594, calcd. for [$C_{14}H_{22}NaO_4$]⁺ 277.1410 found 277.1415

Synthesis of 7: Compound **6** (0.350 g, 1.38 mmol) and triphenylphosphine (7.2075 g, 27.5 mmol) were solubilized in DMF (60 mL) and carbon tetrabromide (9.1275 g, 27.5 mmol) was added. The reaction was stirred at 30 °C for 18 h. The solvent was removed under reduced pressure. The crude product was dissolved in H₂O (40 mL) and extracted with dichloromethane (2 × 100 mL). Then, purification on silica gel was performed (Hexane/DCM 100:0 to 80:20) to afford compound **7** as a colorless oil (0.4362 g, 63 %).

¹*H NMR* (CDCl₃, 400 MHz, 298 K) δ (ppm) 7.30 (m, 1H, CH_{Ar}), 7.14-7.08 (m, 3H, CH_{Ar}), 3.59 (dd, 4H, CH₂-Br, J = 10.5, 4.6 Hz), 3.43 (dd, 4H, CH₂-Br, J = 10.5, 6.3 Hz), 2.18 (d, 4H, CH₂-Ar, J = 7.5 Hz), 2.32-2.22 (m, 2H, CH). ¹³*C Jmod NMR* (CDCl₃, 125 MHz, 298 K) δ (ppm) 138.9 (C_{ipso}), [130.0, 129.3, 127.6] (CH_{Ar}), 43.8 (CH), 37.5 (CH₂-Ar), 35.8 (CH₂-Br). *MALDI-TOF* (matrix: dithranol): m/z calcd for [C₁₄H₁₉Br₄] 506.820 found [M+H]⁺ 506.254

Synthesis of 9: To a solution of **7** (0.4362 g, 0.86 mmol) in dry acetonitrile (7.5 mL), a solution of compound **8** (0.372 g, 1.77 mmol) in dry acetonitrile (7.5 mL) was added dropwise. Then, K_2CO_3 (2.3770 g, 17.2 mmol, 20 equiv.) was added and the reaction was stirred at 60 °C for 10 days. K_2CO_3 was removed through filtration with cannula and filtrate was concentrated under reduced pressure. Crude product was purified by flash column chromatography on silica gel (DCM/Et₃N/MeOH 97:3:0 to 87:3:10) to afford compound **9** as a white foam (0.0836 g, 16 %).

¹*H NMR* (CDCl₃, 500 MHz, 298 K) δ (ppm) 7.15-7.07 (m, 1H, CH_{Ar}), 6.99-6.81 (m, 3H, CH_{Ar}), 3.98 (br, 1H), 3.82-3.58 (m,3H), 3.44-3.02 (m, 5H), 2.90-2.11 (m, 31H, CH₂ α N), 1.75 (br, 1H), 1.27 (s, 6H, CH₃), 1.22 (s, 6H, CH₃), 1.17-1.07 (m, 2H, CH₂ β N). ¹³*C Jmod NMR* (CDCl₃, 125 MHz, 298 K) δ (ppm) [142.0 (x2), 141.9, 139.6 (x2)] (*C*_{ipso}), [129.8, 129.7, 129.5, 128.3, 128.2, 126.7, 126.6, 126.4, 126.3] (CH_{Ar}), 74.1 (N-*C*(CH₃)-N), [55.6, 53.1, 53.0, 50.9, 50.9, 50.8, 50.7, 49.3, 46.7, 46.0, 45.0, 45.0, 44.9] (CH₂ α N), 39.1 (CH₂ γ N), [34.4, 28.7] (CH), [18.1, 18.0] (CH₂ β N), [11.3, 11.1, 10.9, 10.0] (CH₃). *HRMS* (ESI, positive, H₂O) m/z calcd. for [C₃₆H₅₉N₈]⁺ 603.4857 found [M+H]⁺ 603.4853, calcd. for [C₃₆H₆₀N₈]²⁺ 302.2465 found [M+2H]²⁺ 302.2466, calcd. for [C₃₆H₆₁N₈]³⁺ 201.8334, found [M+3H]³⁺ 201.8336.

Synthesis of 10: Compound 9 (0.0836 g, 0.139 mmol) was solubilized in HCl (3 M, 14 mL) and the reaction was stirred at room temperature for 24 h. An extraction with $CHCl_3$ (3 × 20 mL) was performed to remove organic impurities. Then the aqueous layer was concentrated under reduced pressure and the product was purified by flash chromatography on reversed-phase C18 silica (H₂O(0.1 % formic acid)/CH₃CN 100:0 to 0:100) to afford compound *C,C'-(m-xylylene)bis-cyclam* as a brown film (0.071 g, 59 %).

¹*H NMR* (D₂O, 500 MHz, 298 K) δ (ppm) 8.18 (s, 0.3H, *H*COOH), 7.38 (m, 1H, *CH*_{Ar}), 7.31-7.22 (m, 3H, *CH*_{Ar}), 3.70-3.54 (m, 13H), 3.53-3.33 (m, 20H), 3.28-3.18 (m, 6H), 3.00-2.85 (m, 6H), 2.28 (br, 4H, *CH*₂ β N). ¹³*C Jmod NMR* (D₂O, 125 MHz, 298 K) δ (ppm) 140.3 (*C*_{ipso}), [132.9, 132.4, 131.0] (*C*H_{Ar}), [50.5, 45.4, 44.5, 43.5] (*C*H₂ α N), 38.4 (*C*H₂ γ N), 36.1 (*C*H), 22.6 (*C*H₂ β N). *HRMS* (ESI, positive, H₂O) m/z calcd. for [C₂₈H₅₆N₈]⁺ 252.2308 found [M+2H]²⁺ 252.2309, calcd. for [C₂₈H₅₇N₈]³⁺ 168.4897 found [M+3H]³⁺ 168.4902, calcd. for [C₂₈H₅₈N₈]⁴⁺ 126.6191 found [M+4H]⁴⁺ 126.6192.

Spectral data Compound 1:



Figure S1: 1H (400 MHz, CDCl3, 298K, TMS) and 13C Jmod (75 MHz, CDCl3, 298K, TMS) NMR

HRAM



Figure S2: HRMS spectrum (ESI)

CÕZ





Figure S3: ¹H (300 MHz, CDCl₃, 298K, TMS) and ¹³C Jmod (75 MHz, CDCl₃, 298K, TMS) NMR



Figure S4: HRMS spectrum (ESI)

Compound 4:

Formula: C₃₂H₄₆N₈O₂ 0 0 Molecular Weight: 574.77 g.mol⁻¹ H. H N´ Description: white solid Н Н Ň N Yield: 30 % ---7.16 ---7.00 2.25 2.19 2.25 2.25 2.25 2.19 333 .21 1.00 U 2.02 1.97 2 8 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.0 1.5 1.0 2.5 ppm -171.2 -139.7 9 - 0 -76.0 70.6 -19.5 129. 128. 52. 55. 54. 4 2 Ś 4 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 ppm

Figure S5: ¹H (300 MHz, CDCl₃, 298K, TMS) and ¹³C Jmod (75 MHz, CDCl₃, 298K, TMS) NMR

icoa

Fédération de Recherche Physique et Chimie du Vivant (FR2708 : CBM/ICOA) HRAM Plate-forme de Spectrométrie de Masse Haute Résolution

Acquisition Date Instrument / Ser# Method Analysis Info 06/01/2021 17:29:04 Sample Name Analysis Name MLR 095 X058842CYC.d maXis 255552.00086 Positif.m Acquisition Parameter Source Type Scan Begin Scan End ESI 50 m/z 3000 m/z Positive 4500 V 1800.0 Vpp 0.6 Bar 200 °C 7.0 **I**/min Ion Polarity Set Nebulizer Set Dry Heater Set Dry Gas Set Capillary Set Collision Cell RF Intens. x10⁵ +MS, 0.41min #24, Background Subtracted (#6-7) 2+ 288.1949 4-3-2-1+ 575.3820[+ |597.3640 1-425.2537 141.9588 0-100 200 300 400 500 6<u>0</u>0 m/z Intens. x10⁴ X058842CYC.d: +MS, 0.41min #24, Background Subtracted (#6-7) 1+ 575.3820 8 6 1+ 576.3851 4 2-1+ 577.3885 573.3657 574.3710 0. x10⁴ C32H47N8O2, 575.3816 1+ 575.3816 8 6 1+ 576.3846 4 2-1+ 577.3874 0 573 574 575 576 577 578 579 580 . m/z e⁻ Conf Meas. m/z 288.194939 Ion Formula m/z 288.194463 err [ppm] 1.7 mSigma rdb 14.0 z # 14.3 9.8 2+ 1+ C32H48N8O2 even 1 575.381999 C32H47N8O2 575.381649 -0.6 14.0 even C32H46N8NaO2 597.364017 597.363593 1+ 1 -0.7 11.1 14.0 even

Institut de Chimie Organique et Analytique	cyril.colas@univ-orleans.fr	printed:	07/01/2021 16:46:45
UMR 7311 Université d'Orléans CNRS	+33 (0)2 38 49 46 61		Page 1 of 1
BP 6759 - rue de Chartres - F-45067 ORLEANS Cedex 2			

Figure S6: HRMS spectrum (ESI)

Table S1: X-ray data of compound 4

Empirical formula	$C_{32}H_{46}N_8O_{2,}C_4H_8O_2$		
Formula weight	662.87		
Temperature	193(2) К	
Wavelength	0.7107	'3 Å	
Crystal system	Triclir	nic	
Space group	P 1		
	a = 10.1829(8) Å	α = 63.913(2)°	
Unit cell dimensions	b = 13.8753(10) Å	β = 89.426(2)°	
	c = 14.2627(10) Å	γ = 73.285(2)°	
Volume	1717.5(2) Å ³	
Z	2		
Density (calculated)	1.282 Mg/m ³		
Absorption coefficient	0.086 mm ⁻¹		
F(000)	716		
Crystal size	0.200 x 0.180 x 0.100 mm ³		
Theta range for data collection	1.604 to 29.155°		
Index ranges	-13<=h<=13, -17<=k<=18, -14<=l<=19		
Reflections collected	53091		
Independent reflections	9117 [R(int)	= 0.0568]	
Completeness to theta = 25.242°	99.5	%	
Absorption correction	Semi-empirical fro	om equivalents	
Max. and min. transmission	0.7458 and	0.6737	
Refinement method	Full-matrix least-squares on F2		
Data / restraints / parameters	9117 / 0 / 435		
Goodness-of-fit on F ²	1.032		
Final R indices [I>2sigma(I)]	R1 = 0.0797, wR2 = 0.2293		
R indices (all data)	R1 = 0.1323, wR2 = 0.2686		
Extinction coefficient	n/a		
Largest diff. peak and hole	0.780 and -0.578 e.Å ⁻³		



Figure S7: ¹H (400 MHz, CDCI₃, 298K, TMS) and ¹³C Jmod (125 MHz, CDCI₃, 298K, TMS) NMR

Acquisition Date

Instrument / Ser#

HRAM

01/12/2021 18:25:31

maXis 255552.00086

Analysis Info

Sample Name MLR 194 Analysis Name X063723CYC.d



Figure S8: HRMS spectrum (ESI)

Compound 6:

HO

HO

Formula: C₁₄H₂₂O₄

Molecular Weight: 254.33 g.mol⁻¹

OH Description: colorless oil

ЮH





Figure S9: ¹H (400 MHz, MeOD, 298K, TMS) and ¹³C Jmod (125 MHz, MeOD, 298K, TMS) NMR

HRAM

Analysis Info

Acquisition Date 01/12/2021 18:24:05 Instrument / Ser# Sample Name MLR 195 f4 maXis 255552.00086 Analysis Name X063722CYC.d Method Positif.m Acquisition Parameter Source Type Scan Begin Scan End Positive 4500 V 1800.0 Vpp 0.6 Bar 200 °C 7.0 I/min ESI Ion Polarity Set Nebulizer 50 m/z 3000 m/z Set Capillary Set Collision Cell RF Set Dry Heater Set Dry Gas Intens.-x10⁵ +MS, 0.26min #15, Background Subtracted (#8-9) 277**1**415 2.0 1.5-1.0-1+ 255**.1**595 0.5 <mark>_</mark>183<mark>4</mark>168 143.0850 293.1134 0.0 100 150 200 250 300 50 m/z Intens. x10⁵ X063722CYC.d: +MS, 0.26min #15, Background Subtracted (#8-9) 1+ 255<mark>.1</mark>595 0.8 0.6 0.4 1+ 256.1631 0.2 0.0 x10⁵ C₁₄H₂₃O₄, 255.1591 1+ 255.1591 0.8 0.6 0.4 1+ 256.1625 0.2 1+ 257.1648 0.0 254.5 255.0 255.5 256.0 256.5 257.0 257.5 258.0 258.5 259.0 m/z Ion Formula e⁻ Conf Meas. m/z z # m/z err [ppm] mSigma rdb 143.085010 1+ C11H11 143.085527 3.6 35.1 7.0 even 1 7.0 8.0 171.116683 183.116782 1+ C13H15 C14H15 171.116827 183.116827 0.8 124.5 even 0.2 1 +6.4 even 1 201.127602 1+ C14H17O 201.127392 -1.0 8.5 7.0 even 219.138264 237.147600 1 +C14H19O2 C14H21O3 219.137956 -1.4 3.9 n.a. n.a. 6.0 5.0 even 237.148521 1+ even 255.159479 277.141462 C14H23O4 C14H22NaO4 1+ 255.159086 -1.5 21.6 4.0 even 277.141030 1_{+} -16 3.0 4.0 even 1 01/12/2021 18:43:04 Institut de Chimie Organique et Analytique UMR 7311 - Université d'Orléans - CNRS cyril.colas@univ-orleans.fr +33 (0)2 38 49 46 61 printed: Page 1 of 1

Figure S10: HRMS spectrum (ESI) of compound 6

BP 6759 - rue de Chartres - F-45067 ORLEANS Cedex 2



Figure S11: ¹H (400 MHz, CDCl₃, 298K, TMS) and ¹³C Jmod (125 MHz, CDCl₃, 298K, TMS) NMR



Figure S12: ¹H (500 MHz, CDCl₃, 298K, TMS) and ¹³C Jmod (125 MHz, CDCl₃, 298K, TMS) NMR

HRAM



Figure S13: HRMS spectrum (ESI)

C



Figure S14: ¹H (400 MHz, D₂O, 298K, TMS) and ¹³C Jmod (125 MHz, D₂O, 298K, TMS) NMR

HRAM

Analysis Info



Figure S15: HRMS spectrum (ESI)

Nb: Compound 10 is not UV-active at the concentrations used for HPLC identification, therefore detection was performed with ELSD.



==== Shimadzu LabSolutions Data Image ====

Figure S16: HPLC-ELSD chromatogram. Mobile phase $H_20 + 0.1\%$ formic acid / Acetonitrile (95:5 for 16 mins, gradient to 10:90 in 6 mins + 2 mins at 10:90, gradient to 95:5 in 6 mins)

Biological Assays

n=1					
Group Name	Concentration (nM)	Average	Minus Neg Contrl	%inhibition	IC50
Negative Control		15,58	0		
CXCL12		91,99	76,41		
compound 10	50 µM	14,36	-1,22	101,60	
	10 µM	30,81	15,23	80,07	
	2 µM	56,32	40,74	46,68	2,34 µM
	0,4 µM	88,17	72,59	5,00	
	0,08 µM	90,95	75,37	1,36	
	0,016 µM	80,96	65,38	14,44	
	0,0032 µM	85,77	70,19	8,14	
	0,00064 µM	74,91	59,33	22,35	
AMD3100	2000 nM	16,7	1,12	98,53	
	400 nM	40,7	25,12	67,12	146,38 nM
	80 nM	61,65	46,07	39,71	
	16 nM	83,25	67,67	11,44	

Table S2: Calcium Signalling Assays (3 replicates, U87.CD4.CXCR4 ce	əlls)
---------------------------------------------------------------------	-------

n = 2					
Group Name	Concentration (nM)	Average	Minus Neg Contrl	%inhibition	IC50
Negative Control		19,24	0,00		
CXCL12		101,07	81,83		
compound 10	50 µM	14,91	-4,33	105,29	
	10 µM	29,93	10,69	86,94	
	2 µM	89,6	70,36	14,02	4,43 μM
	0,4 µM	105,57	86,33	-5,5	
	0,08 µM	102,25	83,01	-1,44	
	0,016 µM	105,73	86,49	-5,69	
	0,0032 µM	116,5	97,26	-18,86	
	0,00064 µM	122,12	102,88	-25,77	
AMD3100	2000 nM	22,69	3,45	95,78	
	400 nM	35,08	15,84	80,65	133,75 nM
	80 nM	71,92	52,68	35,62	
	16 nM	90,92	71,68	12,41	

n = 3					
Group Name	Concentration (nM)	Average	Minus Neg Contrl	%inhibition	IC50
Negative Control		19,47	0		
CXCL12		93,25	73,78		
compound 10	50 µM	18,8	-0,67	100,91	
	10 µM	34,27	14,8	79,94	
	2 µM	72,81	53,34	27,70	3,98 µM
	0,4 µM	93,5	74,03	-0,34	
	0,08 µM	87,93	68,46	7,21	
	0,016 µM	92,8	73,33	0,61	

Figure S17: Calcium Signalling Assays (U87.CD4.CCR5 cells)

Compound 10 (20 µM)+ LD78beta (50 ng/ml)



Compound 10 (20 µM- 4 µM)+ LD78beta (50 ng/ml)



Maraviroc (8 nM- 1.6 nM-0.32 nM) + LD78beta (50 ng/ml) – Positive control





 Table S3:
 Monoclonal Antibodies (mAbs) binding assays (IC₅₀ values, Jurkat cells)

<u>12G5</u>			
			AVERAGE
	IC ₅₀ (nM)	IC ₅₀ (nM)	IC₅₀ (nM)
compound 10	341,3	411,7	376,5
AMD3100	6,8	6,8	6,8

<u>44717</u>

			AVERAGE
	IC50 (nM)	IC ₅₀ (nM)	IC₅₀ (nM)
compound 10	954,1	405,6	679,9
AMD3100	110,0	16,6	63,3

<u>1D9</u>

			AVERAGE
	IC ₅₀ (nM)	IC ₅₀ (nM)	IC₅₀ (nM)
compound 10	> 12 000	> 20 000	> 20 000
AMD3100	> 12 000	> 20 000	> 20 000

<u>2B11</u>

			AVERAGE
	IC ₅₀ (nM)	IC ₅₀ (nM)	IC₅₀ (nM)
compound 10	> 12 000	> 20 000	> 20 000
AMD3100	> 12 000	> 20 000	> 20 000



Figure S19: Monoclonal Antibodies (mAbs) binding assays

Table S4: Inhibition of HIV-infection in TZM-bl, luciferase assay

	NL4.3WT	NL4.3WT	BaL	BaL	Toxicity	Toxicity
	IC₅₀ (µM)	IC₅₀ (µM)	IC₅₀ (µM)	IC₅₀ (µM)	CC₅₀ (µM)	CC₅₀ (µM)
compound 1	0,11	0,082	>20	>20	71,74	>100
AMD3100	0,00043	0,00031	-	-	>100	