SUPPORTING INFORMATION

Supramolecular aggregates constructed by pillar[5]arene-based host-guest interaction with aggregation-induced emission

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Experimental Section

General Methods

¹H NMR, ¹³C NMR and 2D NOESY spectra were measured on a Brüker 400 spectrometer. The ESI-TOF-MS were acquired on an AB Sciex TripleTOF[®] 6600 mass spectrometer. Fluorescent spectra were recorded on a HITACHI F-7000 spectrophotometer. SEM images were recorded on a ZEISS Sigma 300 apparatus. DLS were recorded on a Malvern Zetasizer Nano ZS90. Fluorescent quantum efficiencies were determined using a Hamamatsu C11347-13 Quantaurus-QY spectrometer.

Materials

Chemicals were used as received from Adamas. All solvents were reagent grade and were dried and distilled prior to use according to standard procedures. The molecular structures were confirmed using ¹H NMR, ¹³C NMR spectroscopies and ESI-TOF-MS.

Synthesis

The compound **A**, compound **B**, compound **C**, compound **D** and pillar[5]arene (P5) were synthesized according to previous reports ^[1-5].



Fig. S1. The synthesis routes of H1 and G

Synthesis of H1

H1 was prepared in a closed reaction flask with a [A]:[B]:[C]:[CuI]:[DIEA] ratio of 1:1:1:2.5:20. Compound A (14.561 mg, 0.0376 mmol), compound B (30.000 mg, 0.0376 mmol), compound C (5.719 mg, 0.0376 mmol), CuI (17.892 mg, 0.0939 mmol) and DIEA (97.132 mg, 0.7516 mmol) were dissolved in 10.0 ml dry THF, then the reaction mixture was reaction under 80 °C for 12 hours with argon. The mixture was dissolved in 50.0 ml dichloromethane, washed twice with pure water, and then dried with MgSO₄. After filtering, the solvent was removed by evaporation. The residue was purified by column chromatography (SiO₂, dichloromethane/Methanol=100:1) to give purified monomer H1 (yield = 35.3 %). ¹H NMR (CDCl₃, 400 MHz, 298K): δ 7.66 (s, 1H), 7.62 (s, 1H), 7.10-6.98 (m, 29H), 5.45-5.27 (m, 6H), 3.85-3.73 (m, 34H), 1.71(s, 1H), 1.31-1.13 (m, 3H), -0.70 (d, *J* = 6.4 Hz, 4H), -1.61 (t, *J* = 6.2 Hz, 4H). ¹³C NMR (CDCl₃, 100 MHz, 298K): δ 151.0, 150.7, 150.6, 150.2, 150.1, 150.0, 149.8, 146.4, 144.9, 144.4, 143.5, 143.4, 143.3, 141.8, 139.5, 132.6, 132.0, 131.3, 131.2, 130.0, 129.1, 129.0, 128.7, 128.5, 128.2, 128.0, 127.8, 127.7, 127.6, 127.3, 126.7, 126.6, 124.9, 122.8, 120.7, 115.8, 114.9, 114.1, 114.0, 113.7, 113.4, 113.3, 113.1, 113.0, 112.8, 61.9, 59.6, 57.7, 56.3, 56.1, 55.6, 55.4, 55.3, 55.2, 53.8, 53.4, 49.5, 30.6, 29.6, 29.3, 28.5, 27.4, 26.2, 23.6, 22.6, 13.6. ESI-TOF-MS(m/z): [M+H]⁺: calcd for C₈₃H₈₄N₇O₁₀,1338.6274; found,1338.6275.

Synthesis of G

G was synthesized in a closed reaction flask with a [**C**]:[**D**] ratio of 1:1. Compound **C** (42.600 mg, 0.308 mmol), compound **D** (50.000 mg, 0.308 mmol), CuI (88.07 mg, 0.462 mmol) and DIEA (597.64 mg, 4.62 mmol) were dissolved in 10.0 ml dry THF, then the reaction mixture was

reaction under 80 °C for 12 hours with argon. The mixture was dissolved in 50.0 ml dichloromethane, washed twice with pure water, and then dried with MgSO₄. After filtering, the solvent was removed by evaporation. The residue was purified by column chromatography (SiO₂, dichloromethane/Methanol=50:1) to give purified monomer **G** (yield = 78.8 %). ¹H NMR (CDCl₃, 400 MHz, 298K): δ 7.57 (s, 1H), 6.94-6.90 (m, 2H), 6.85-6.81 (m, 2H), 5.16 (s, 2H), 4.36(t, *J* = 7.2 Hz, 2H), 3.76 (s, 3H), 2.32 (t, *J* = 6.8 Hz, 2H), 1.97-1.89 (m, 2H), 1.67-1.60 (m, 2H), 1.52-1.45 (m, 2H), 1.38-1.31 (m, 2H). ¹³C NMR (CDCl₃, 100 MHz, 298K): δ 154.2, 152.3, 144.6, 122.4, 119.5, 115.9, 114.7, 62.8, 55.7, 50.1, 30.0, 28.0, 25.7, 25.1, 17.0. ESI-TOF-MS (m/z): [M+Na]⁺: calcd for C₁₇H₂₂N₄O₂Na, 337.1635; found, 337.1629.

Complexation between P5 and G.



Fig. S2. ¹H NMR spectra (400 MHz, CDCl₃, 298K) of **P5** at a concentration of 2.0 mM upon addition of **G** (15.0 mM): (a) 0.0 μ l, (b) 10.0 μ l, (c) 20.0 μ l, (d) 30.0 μ l, (e) 40.0 μ l, (f) 60.0 μ l, (g) 80.0 μ l, (h) 100.0 μ l, (i) 120.0 μ l, (j) 140.0 μ l, (k) 160.0 μ l, (l) 200.0 μ l.



Fig. S3. Mole ratio plot for P5 and G. The plot indicates a 1:1 stoichiometry.

Determination of the association constant.

For **G**, chemical exchange is fast on the NMR time scale (Figure S2). To determine the association constant, NMR titrations were done with solutions which had a constant concentration of **P5** and varying concentrations of **G**. Using the nonlinear curve-fitting method, the association constant was obtained for each host-guest combination from the following equation:

$$A = (A_{\infty}/[P5]_0) (0.5[G]_0 + 0.5([P5]_0 + 1/Ka) - (0.5([G]_0^2 + (2[G]_0(1/Ka - c)) + (1/Ka + [P5]_0)^2)^{0.5}))$$

Where A is the chemical shift change of aromatic proton on P5 host at $[G]_0$, A_∞ is the chemical shift change of P5 when the host is completely complexed, $[P5]_0$ is the fixed initial concentration of the P5 host, and $[G]_0$ is the initial concentration of guest.



Fig. S4. The non-linear curve-fitting (NMR titrations) for the complexation of P5 host (2.0 mM) with **G** in CDCl₃ at 298 K. The concentration of **G** was 0.375 mM, 0.75 mM, 1.12 mM, 1.15 mM, 2.25 mM, 3.00 mM, 3.75 mM, 4.50 mM, 5.25 mM, 6.00 mM, 7.50 mM.

2D NOESY spectra of H1 in CDCl3 solution

The 2D-NOESY NMR spectrum of the solution of **H1** showed the NOE peaks between protons Hb and H2, H5, H3, H4, as well as Ha/Hc and H2, H5, H3, H4, which clearly confirmed the complexation between pillar[5]arene and neutral nitrile group.



Fig. S5. 2D-NOESY analysis of H1 in CDCl₃ (400 MHz, 298 K, 32.7 mM)

Reference:

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¹H NMR, ¹³C NMR and Mass spectra

Fig. S6. ¹H NMR spectra (CDCl₃, 298 K, 400 MHz) of **H1**.



Fig. S7. ¹³C NMR spectra (CDCl₃, 298 K, 400 MHz) of H1.



Fig. S9. ¹H NMR spectra (CDCl₃, 298 K, 400 MHz) of **G**.



Fig. S10. ¹³C NMR spectra (CDCl₃, 298 K, 400 MHz) of G.



Fig. S11. ESI-TOF-MS of G.