Supporting Information

Lewis acids modulation in phosphorus phenol nickel catalyzed ethylene polymerization and copolymerization

Wenbing Wang,^a Nan Nie,^a Menghe Xu,^{a,*} Chen Zou^{a,*}

^a CAS Key Laboratory of Soft Matter Chemistry, Hefei National Laboratory for Physical Sciences at the Microscale, Department of Polymer Science and Engineering, University of Science and Technology of China, Hefei, 230026, China

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1. Supplementary Figures and Tables

Ent.	Cat.	Lewis Acid	Yield/g	Act. ^b	T _m (°C) ^c	$M_{\rm n}(10^3)^{\rm d}$	PDI ^d
1	-	Ni(OAc)2	-	-	-	-	-
2	-	Zn(OAc) ₂	-	-	-	-	-
3	-	Zn(TMEDA)(OAc) ₂	-	-	-	-	-

Supplementary Table S1 Ethylene homopolymerization.^a

^{*a*}Conditions: Lewis Acid 1 µmol, ethylene 8 atm, toluene 20 mL, t=15 min. ^{*b*} Activity is in unit of 10⁶ g mol⁻¹ h⁻¹. ^{*c*} Determined by differential scanning calorimetry (DSC; second heating). ^{*d*} Determined by GPC in trichlorobenzene at 160 °C with polystyrene standards. M_n is in unit of kg mol⁻¹.

Supplementary Table S2 Ethylene homopolymerization.^a

Ent.	Cat.	Lewis Acid	Yield/g	Act. ^b	T _m (°C) ^c	$M_{\rm n}(10^3)^{ m d}$	PDI ^d
1	Ni1	Ni(COD)2	1.2	4.8	119.1	6.8	2.4
2	Ni2	Ni(COD)2	1.8	7.2	129.1	12.4	3.7
3	Ni3	Ni(COD)2	3.3	13.2	130.5	28.6	5.3

^{*a*}Conditions: **Ni** catalyst 1 µmol, ethylene 8 atm, toluene 20 mL, Ni(COD)₂ 1eq, t=15 min. ^{*b*} Activity is in unit of 10⁶ g mol⁻¹ h⁻¹. ^{*c*} Determined by differential scanning calorimetry (DSC; second heating). ^{*d*} Determined by GPC in trichlorobenzene at 160 °C with polystyrene standards. M_n is in unit of kg mol⁻¹.



Figure S1 Stress-strain curves for every copolymer tested three times in Figure 4a.



Figure S2 The values and figures of copolymers' WCAs in Figure 4b.



Figure S5 DSC of the polymer from Table S2, Entry 3.



22496 45903 249239 37693 3.68728 1 12449 134279 **Processed Peaks** Peak No Pk Height % Height Name Start RT Max RT End RT Area % Area (mins) (mV) (mV.secs) (mins) (mins) -6.88011 1 12.25 14.67 16.72 0 848.833 100

Figure S7 GPC of the polymer from Table S1, Entry 2.



2. Experimental

2.1 General consideration

All manipulations of air- and water-sensitive compounds were carried out using standard Schlenk, high-vacuum, and glovebox techniques. Deuterated solvents used for NMR were dried and distilled prior to use. ¹H NMR spectra was recorded by a Bruker Ascend Tm 400 spectrometer at ambient temperature unless otherwise stated. The chemical shifts of the ¹H NMR spectra were referenced to the residual solvent; Coupling constants are in Hz. Molecular weight and molecular weight distribution of the polymer were determined by gel permeation chromatography (GPC) with a PL-220 equipped with two Agilent PLgel Olexis columns at 160 °C using trichlorobenzene as a solvent, and the calibration was made using polystyrene standard and are corrected for linear polyethylene by universal calibration using the Mark-Houwink parameters of Rudin: $K = 1.75 \times 10^{-2}$ cm³/g and R = 0.67 for polystyrene and $K = 5.90 \times 10^{-2}$ cm³/g and R = 0.69 for polyethylene. DSC measurements were performed on a TA Instruments DSC250. Samples (ca. 5 mg) were annealed by heating to 150 °C at 10 °C /min, cooled to 40 °C at 10 °C /min, and

then analyzed while being heated to 150 °C at 10 °C /min.

The water contact angles on polymer films were measured with a Contact Angle Meter SL200B (Solon Tech. Co., Ltd.) using the dynamic sessile drop method. Samples for water contact angle measurements were prepared via evaporation of 3 to 5 % (w/w) solutions in toluene onto glass slides under ambient conditions. The solvent was evaporated on top of a glass slide for 10 minutes, and a second layer of the polymer solution was then applied to increase thickness. The water contact angles of the polymer thin films were measured using a contact angle goniometer at 25 °C with an accuracy of $\pm 3^{\circ}$. The reported values are the average of at least six measurements made at different positions of the film.

A standard test method, ASTM 638, was followed to measure the tensile properties of the polyethylene samples. Polymers were melt-pressed at 30 to 35°C above their melting point to obtain the dog-bone-shaped tensile-test specimens. The test specimens showed 25-mm gauge length, 2-mm width, and thickness of 0.4 mm. Stress/strain experiments were performed at 10 m/min using a Universal Test Machine (UTM2502) at room temperature. At least three specimens of each copolymer were tested.

2.2 Ethylene polymerization procedure.

For the polymerization reaction of ethylene at 8 atm pressures, a 350 mL glass thick-walled pressure vessel was charged with toluene, a desired amount of comonomer, and a magnetic stir bar in the glovebox. The pressure vessel was connected to a high-pressure line, and the solution was degassed. The vessel was warmed to the desired temperature using an oil bath and allowed to equilibrate for 5 min. The metal catalyst Ni1-Ni3 in toluene was injected into the polymerization system via syringe. With rapid stirring, the reactor was pressurized, maintained at a desired ethylene pressure, and stirred continuously for a desired period. The pressure vessel was vented, the polymerization was quenched via the addition of MeOH (5 mL), and the polymer was precipitated using excess MeOH. After filtration, the polymer sample was obtained and dried at 80 °C for 24 h under vacuum. The polymer

monomer incorporation (mol%) was calculated from NMR analysis. In particular, 30 atm ethylene polymerization takes place in a closed mechanical stirring kettle.



2.3 Ligand preparation procedures.

Ni 1: Under nitrogen, 2-(2-(tert-butyl)phenoxy)tetrahydro-2H-pyran (2.34g,10 mmol) was dissolved in 50 mL of dried THF. n-BuLi(4.6mL,2.4 M in hexane,11 mmol,1.1 equiv) was added dropwise and the reaction mixture was stirred for 2 h at 0 °C. The flask was transferred to a -78 °C bath, and the THF solution of PhPArCl (10 mmol,1.0 equiv) was added dropwise. The mixture was stirred for 1 hour, and warmed to room temperature to react for 12 hours. After quenching, the suspension was transferred to a round bottom flask and the THF was evaporated on a rotary evaporator. The crude material was then extracted with DCM (3 x 200 mL), washed with H₂O (3 x 100 mL), and then collect the organic phase and dried with Na₂SO₄. After filtration and concentration, the crude product of the protected ligand can be obtained, which can be directly used for the next reaction without further purification. Then, the protected ligand was dissolved in methanol under nitrogen, 3.0 eq of p-toluenesulfonic acid was added to react for 6 hours. Then, the reaction solution was evaporated with a rotary evaporator to evaporate methanol, the crude material was then extracted with DCM (3 x 200 mL), washed with H₂O (3 x 100 mL), and then collect the organic phase and dried with Na₂SO₄. After filtration and concentration, the pure ligand 1 was obtained by column chromatography as a white solid (1.80 g, 48%). Under nitrogen, a toluene solution of 1 mmol (377 mg) of the ligand and 1.2 eq Py₂NiMe₂ was stirred for 1 hour at room temperature. After concentrating, adding nhexane for recrystallization resulted in the formation of Ni 1 (428 mg, 81%) as a yellow solid.

Ligand 1: ¹H NMR (400 MHz, CDCl₃) ¹H NMR (400 MHz, CDCl₃) δ 7.68 (s, 1H), 7.43 – 7.28 (m, 6H), 7.23 – 7.15 (m, 1H), 7.11 (s, 2H), 6.91 (s, 1H), 6.76 (t, *J* = 7.5 Hz, 1H), 2.54 (s, 6H), 1.42 (s, 9H). ³¹P NMR (162 MHz, CDCl₃) δ -41.48. ¹³C NMR (101 MHz, CDCl₃) δ 157.61, 157.40, 132.15, 131.98, 128.97, 127.29, 127.22, 127.18, 127.04, 121.19, 120.40, 120.37, 118.90, 118.89, 76.30, 75.98, 75.67, 44.34, 44.31. ESI-MS (m/z): [M+H]⁺ Calcd for C₂₄H₂₉ONP, 378.19813; Found: 378.19809.

Ni 1: ¹H NMR (400 MHz, C₆D₆) δ 8.50 (s, 2H), 7.58 (t, J = 7.7 Hz, 2H), 7.19 - 7.01 (m, 2H), 6.79 (s, 5H), 6.54 (s, 2H), 6.24 (d, J = 5.8 Hz, 3H), 2.44 (s, 6H), 1.32 (s, 9H), -0.78 (d, J = 4.6 Hz, 3H). ³¹P NMR (162 MHz, C₆D₆) δ 13.36. ¹³C NMR (101 MHz, C₆D₆) δ 173.98, 173.77, 157.25, 157.14, 137.11, 137.02, 132.60, 131.84, 131.75, 130.10, 129.48, 126.88, 126.64, 126.39, 123.62, 123.54, 122.11, 122.05, 118.69, 118.17, 112.08, 112.01, 44.72, 33.87, 28.30, -17.19 (d, J = 38.5 Hz, Ni-*Me*). Anal. Calcd for C₃₀H₃₆N₂NiOP: C, 67.95; H, 6.84; N, 5.28. Found: C, 67.76; H, 6.87; N, 5.33.

Ni 2: A procedure similar to the Ni 1;



Ligand 2(10mmol,1.90g,46%):¹H NMR (400 MHz, CDCl₃) δ 7.47 - 7.34 (m, 7H), 7.31 (dd, J = 7.5, 1.8 Hz, 1H), 7.22 (m, J = 7.8, 4.7, 0.9 Hz, 1H), 7.12 (t, J = 7.5 Hz, 1H), 6.94 (m, J = 7.6, 4.7, 1.5 Hz, 1H), 6.87 - 6.75 (m, 2H), 3.59 (t, J = 4.4 Hz, 4H), 2.97 - 2.84 (m, 2H), 2.63 (d, J = 10.5 Hz, 2H), 1.46 (s, 9H). ³¹P NMR (162 MHz, CDCl₃) δ -41.83. ¹³C NMR (101 MHz, CDCl₃) δ 158.27, 158.06, 155.19, 155.03, 135.95, 133.85, 133.66, 133.06, 132.46, 130.18, 128.77,128.60, 128.53, 125.98, 125.96, 122.43, 122.41, 120.14, 120.11, 77.40, 77.08, 76.76, 67.19, 52.82, 34.98, 34.96, 29.62. ESI-MS (m/z): [M+H]⁺ Calcd for C₂₆H₃₁O₂NP, 420.20869; Found: 420.20844.

Ni 2 (1mmol,468 mg, 82%): ¹H NMR (400 MHz, C_6D_6) δ 8.49 (s, 2H), 7.66 (dd, J =

8.3, 4.8 Hz, 2H), 7.08 - 6.94 (m, 2H), 6.89 - 6.75 (m, 7H), 6.63 (s, 1H), 6.51 (s, 2H), 6.26 - 6.12 (m, 3H), 4.19 - 3.68 (m, 4H), 2.84 (s, 2H), 2.45 (s, 2H), 1.29 (s, 9H), -0.78 (d, J = 4.9 Hz, 3H). ³¹P NMR (162 MHz, C₆D₆) δ 18.72. ¹³C NMR (101 MHz, C₆D₆) δ 156.64, 156.55, 138.26, 138.17, 134.64, 134.61, 133.26, 133.17, 131.60, 131.54, 131.53, 129.92, 129.25, 129.23, 129.00, 128.49, 128.40, 127.95, 127.83, 127.71, 127.59, 127.46, 125.25, 125.17, 124.05, 123.99, 121.13, 113.15, 113.08, 67.34, 53.97, 34.89, 34.87, 29.44, -15.82 (d, J = 39.2 Hz, Ni-*Me*). Anal. Calcd for C₃₂H₃₈N₂NiO₂P: C, 67.16; H, 6.69; N, 4.89. Found: C, 67.02; H, 6.77; N, 4.92.

Ni 3: A procedure similar to the Ni 1;



Ligand 3(10mmol,2.25g,45%): ¹H NMR (400 MHz, CDCl₃) δ 8.03 (dd, J = 17.7, 7.5 Hz, 2H), 7.57 (d, J = 7.5 Hz, 1H), 7.50 (s, 1H), 7.38 (d, J = 16.1 Hz, 2H), 7.31 (s, 1H), 7.26 – 7.20 (m, 4H), 7.20 – 7.10 (m, 5H), 7.04 (d, J = 8.1 Hz, 1H), 6.88 (dd, J = 6.8, 5.7 Hz, 1H), 6.73 (t, J = 7.6 Hz, 1H), 6.64 (d, J = 8.0 Hz, 1H), 1.19 (s, 9H). ³¹P NMR (162 MHz, CDCl₃) δ -44.19. ¹³C NMR (101 MHz, CDCl₃) δ 158.54, 158.32, 133.63, 133.44, 128.50, 128.43, 125.46, 120.17, 119.95, 119.67, 119.64, 77.35, 77.24, 77.04, 76.72, 34.68, 34.66, 29.35. ESI-MS (m/z): [M+H]⁺ Calcd for C₃₄H₃₀NOP, 500.21378; Found: 500.21375.

Ni **3** (1mmol,527 mg, 81%): ¹H NMR (400 MHz, C_6D_6) δ 8.06 - 7.97 (m, 2H), 7.75 (dd, J = 23.7, 13.9 Hz, 6H), 7.43 - 7.22 (m, 4H), 7.03 (m, J = 40.8, 21.5, 5.6 Hz, 8H), 6.77 - 6.56 (m, 4H), 6.33 (s, 2H), 1.30 (d, J = 3.4 Hz, 9H), -0.83 (t, J = 4.4 Hz, 3H). ³¹P NMR (162 MHz, C_6D_6) δ -18.94. ¹³C NMR (101 MHz, C_6D_6) δ 149.09, 143.00, 142.61, 137.59, 134.83, 134.10, 133.08, 131.79, 131.70, 130.80, 128.95, 128.26, 127.56, 127.46, 127.09, 126.85, 126.61, 125.13, 124.24, 123.84, 122.95, 121.55, 119.02, 118.79, 118.43, 116.82, 112.59, 111.71, 110.76, 33.77, 28.43, -14.59 (d, J = 3.4 Hz, J = 4.4 Hz, 36.3 Hz, Ni-*Me*). Anal. Calcd for C₄₀H₃₈N₂NiOP: C, 73.64; H, 5.87; N, 4.29. Found:
C, 73.52; H, 5.78; N, 4.22.

3. ¹H NMR and ¹³C NMR of the ligands and catalyst





Figure S11 ¹³C NMR spectrum of L1. (101 MHz, CDCl₃).



Figure S12 ESI-MS spectrum of L1.



Figure S13 ¹H NMR spectrum of L2. (400 MHz, CDCl₃).



Figure S14 ³¹P NMR spectrum of L2. (162 MHz, CDCl₃).



Figure S15¹³C NMR spectrum of L2. (101 MHz, CDCl₃).



Figure S16 ESI-MS spectrum of L2.



Figure S17 ¹H NMR spectrum of L3. (400 MHz, CDCl₃).



Figure S18 ³¹P NMR spectrum of L3. (162 MHz, CDCl₃).







Figure S20 ESI-MS spectrum of L3.



Figure S21 ¹H NMR spectrum of Ni1. (400 MHz, C₆D₆). (*solution)



Figure S22 ³¹P NMR spectrum of Ni1. (162 MHz, C₆D₆).



Figure S23 13 C NMR spectrum of Ni1. (101 MHz, C₆D₆).



Figure S24 ¹H NMR spectrum of Ni2. (400 MHz, C₆D₆). (*solution)



Figure S25 ³¹P NMR spectrum of Ni2. (162 MHz, C_6D_6).



Figure S26 ¹³C NMR spectrum of Ni2. (101 MHz, C₆D₆). (*Hex)







Figure S28 ³¹P NMR spectrum of Ni3. (162 MHz, C₆D₆).





4. ¹H NMR of the polymers



Figure S30 ¹H NMR of the polymer from Table 3, Entry 1. (C₂D₂Cl₄, 120 °C)



Figure S31 ¹H NMR of the polymer from Table 3, Entry 2. (C₂D₂Cl₄, 120 °C)



Figure S32 ¹H NMR of the polymer from Table 3, Entry 3. ($C_2D_2Cl_4$, 120 °C)



Figure S33 ¹H NMR of the polymer from Table 3, Entry 4. (C₂D₂Cl₄, 120 °C)

Incorporation (%) =
$$\frac{\frac{I(a)/3}{I(a)}}{3} + \frac{I(CH2) - 17}{4} * 100\% = 0.8\%$$



Figure S34 ¹H NMR of the polymer from Table 3, Entry 5. (C₂D₂Cl₄, 120 °C)



Figure S35 ¹H NMR of the polymer from Table 3, Entry 7. (C₂D₂Cl₄, 120 °C)



Figure S36 ¹H NMR of the polymer from Table 3, Entry 8. (C₂D₂Cl₄, 120 °C)



Figure S38 ¹H NMR of the polymer from Table 3, Entry 10. (C₂D₂Cl₄, 120 °C)



Figure S39 ¹H NMR of the polymer from Table 3, Entry 11. (C₂D₂Cl₄, 120 °C)



Figure S40 ¹H NMR of the polymer from Table 3, Entry 12. (C₂D₂Cl₄, 120 °C)



Figure S42 ¹H NMR of the polymer from Table 3, Entry 14. (C₂D₂Cl₄, 120 °C)



Figure S43 ¹H NMR of the polymer from Table 3, Entry 15. (C₂D₂Cl₄, 120 °C)



Figure S44 ¹H NMR of the polymer from Table 3, Entry 16. (C₂D₂Cl₄, 120 °C)

5. DSC of polymers



Figure S45 DSC of the polymer from Table 1, Entry 1.



Figure S46 DSC of the polymer from Table 1, Entry 2.



Figure S48 DSC of the polymer from Table 1, Entry 4.







Figure S55 DSC of the polymer from Table 1, Entry 11.















Figure S64 DSC of the polymer from Table 2, Entry 11.



Figure S67 DSC of the polymer from Table 3, Entry 2.





Figure S70 DSC of the polymer from Table 3, Entry 5.



Figure S73 DSC of the polymer from Table 3, Entry 8.



Figure S76 DSC of the polymer from Table 3, Entry 11.



Figure S77 DSC of the polymer from Table 3, Entry 12.



Figure S78 DSC of the polymer from Table 3, Entry 13.



Figure S79 DSC of the polymer from Table 3, Entry 14.



Figure S80 DSC of the polymer from Table 3, Entry 15.



Figure S81 DSC of the polymer from Table 3, Entry 16.

6. GPC of polymers



Figure S82 GPC of the polymer from Table 1, Entry 1.



Figure S83 GPC of the polymer from Table 1, Entry 2.



Figure S84 GPC of the polymer from Table 1, Entry 3.



Figure S85 GPC of the polymer from Table 1, Entry 4.





Figure S86 GPC of the polymer from Table 1, Entry 5.

Figure S87 GPC of the polymer from Table 1, Entry 6.

S44



Figure S88 GPC of the polymer from Table 1, Entry 7.



Figure S89 GPC of the polymer from Table 1, Entry 8.



Figure S90 GPC of the polymer from Table 1, Entry 9.



Figure S91 GPC of the polymer from Table 1, Entry 10.



Figure S92 GPC of the polymer from Table 1, Entry 11.



Figure S93 GPC of the polymer from Table 1, Entry 12.



Figure S94 GPC of the polymer from Table 2, Entry 2.



Figure S95 GPC of the polymer from Table 2, Entry 3.



Figure S96 GPC of the polymer from Table 2, Entry 4.



Figure S97 GPC of the polymer from Table 2, Entry 6.







Figure S99 GPC of the polymer from Table 2, Entry 8.



Figure S100 GPC of the polymer from Table 2, Entry 10.



Figure S101 GPC of the polymer from Table 2, Entry 11.







Figure S103 GPC of the polymer from Table 3, Entry 1.







Figure S105 GPC of the polymer from Table 3, Entry 3.







Figure S107 GPC of the polymer from Table 3, Entry 5.



Figure S108 GPC of the polymer from Table 3, Entry 6.



Figure S109 GPC of the polymer from Table 3, Entry 7.



Figure S110 GPC of the polymer from Table 3, Entry 8.



Figure S111 GPC of the polymer from Table 3, Entry 9.







Figure S113 GPC of the polymer from Table 3, Entry 11.







Figure S115 GPC of the polymer from Table 3, Entry 13.







Figure S117 GPC of the polymer from Table 3, Entry 15.



Figure S118 GPC of the polymer from Table 3, Entry 16.

7. X-ray Crystallography

Entry	Ni1
Empirical formula	C ₃₀ H ₃₅ N ₂ NiOP
Formula weight	529.28
Temperature/K	100
Crystal system	orthorhombic
Space group	P2 ₁ 2 ₁ 2 ₁
a/Å	8.6966(6)
b/Å	15.9160(12)
c/Å	19.1276(19)
α/°	90
β/°	90
$\gamma/^{\circ}$	90
Volume/Å ³	2647.5(4)
Z	4
pcalcg/cm ³	1.328
µ/mm ⁻¹	0.819
F(000)	1120.0
Crystal size/mm ³	0.19 imes 0.08 imes 0.05
Radiation	MoK α ($\lambda = 0.71073$)
20 range for data collection/°	4.97 to 52.76
Index ranges	$-10 \le h \le 10, -19 \le k \le 18, -23 \le l \le 16$
Reflections collected	11685
Independent reflections	5167 [Rint = 0.0826, Rsigma = 0.1291]
Data/restraints/parameters	5167/0/322
Goodness-of-fit on F ²	1.043

Final R indexes [I>= 2σ (I)]	$R_1 = 0.0572, wR_2 = 0.0925$
Final R indexes [all data]	$R_1 = 0.0973, wR_2 = 0.1121$
Largest diff. peak/hole / e Å ⁻³	0.43/-0.44

Summary of Data – CCDC. 2257728

Entry	Ni2
Empirical formula	$C_{32}H_{37}N_2NiO_2P$
Formula weight	571.31
Temperature/K	100
Crystal system	triclinic
Space group	P-1
a/Å	9.1318(5)
b/Å	9.5917(4)
c/Å	16.6693(9)
$\alpha/^{\circ}$	96.988(2)
β/°	92.616(2)
$\gamma/^{\circ}$	104.598(2)
Volume/Å ³	1397.99(12)
Z	2
pcalcg/cm ³	1.357
μ/mm^{-1}	0.783
F(000)	604.0
Crystal size/mm ³	0.15 imes 0.08 imes 0.05
Radiation	MoK α ($\lambda = 0.71073$)
2Θ range for data collection/°	4.43 to 52.864
Index ranges	$-11 \le h \le 11, -11 \le k \le 12, -20 \le l \le 20$
Reflections collected	16326
Independent reflections	5710 [Rint = 0.0533, Rsigma = 0.0636]
Data/restraints/parameters	5710/0/347
Goodness-of-fit on F ²	1.045
Final R indexes $[I \ge 2\sigma(I)]$	$R1 = 0.0428$, $wR_2 = 0.0823$
Final R indexes [all data]	$R1 = 0.0657, wR_2 = 0.0939$
Largest diff. peak/hole / e Å ⁻³	0.35/-0.40

Summary of Data – CCDC. 2257729

Entry	Ni3
Empirical formula	$C_{40}H_{37}N_2NiOP$
Formula weight	651.39
Temperature/K	170.0
Crystal system	triclinic
Space group	P-1
a/Å	10.4219(8)
b/Å	12.3823(10)
c/Å	12.6499(9)
α/°	88.716(2)
β/°	83.981(2)
γ/°	85.981(3)
Volume/Å ³	1619.3(2)
Z	2
pcalcg/cm ³	1.336

µ/mm ⁻¹	0.684
F(000)	684.0
Crystal size/mm ³	0.12 imes 0.07 imes 0.05
Radiation	MoKa ($\lambda = 0.71073$)
2Θ range for data collection/°	4.962 to 52.826
Index ranges	$-13 \le h \le 13, -15 \le k \le 15, -14 \le 1 \le 15$
Reflections collected	18704
Independent reflections	6567 [Rint = 0.0351, Rsigma = 0.0416]
Data/restraints/parameters	6567/0/410
Goodness-of-fit on F ²	0.990
Final R indexes $[I \ge 2\sigma(I)]$	$R1 = 0.0353$, $wR_2 = 0.0756$
Final R indexes [all data]	$R1 = 0.0448$, $wR_2 = 0.0818$
Largest diff. peak/hole / e Å ⁻³	0.50/-0.33

Summary of Data – CCDC. 2257730