

Supporting Information

Porous Organic Polymers Incorporating BODIPY Moieties for Efficient Removal of Organic Dyes from Aqueous Solutions

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Characterization. Fourier transform infrared (FT-IR) spectra were measured within a 4000 to 400 cm^{-1} region on a Bruker TENSOR-27 infrared spectrophotometer (KBr pellet). ^1H NMR, ^{13}C NMR, ^{19}F NMR, and ^{11}B NMR spectra were measured on a Bruker AVANCE-300 or 400 NMR spectrometer. Solid-state ^{13}C and ^{29}Si cross-polarization/magic-angle-spinning (CP/MAS) NMR spectra were recorded on a Bruker AVANCE-500 NMR spectrometer operating at a magnetic field strength of 9.4 T. The resonance frequencies at this field strength were 125 and 99 MHz for ^{13}C and ^{29}Si NMR, respectively. A chemagnetics 5 mm triple-resonance MAS probe was used to acquire ^{13}C and ^{29}Si NMR spectra. ^{29}Si MAS NMR spectra with high power proton decoupling were recorded by using a $p/2$ pulse length of 5 ms, a recycle delay of 120 s, and a spinning rate of 5 kHz. Elemental analyses were conducted using an Elementar vario EL III elemental analyzer.

Thermogravimetric analysis (TGA) was conducted under N_2 using a TA SDTQ600 at a temperature range of room temperature to 800°C with a heating rate of $10^\circ\text{C min}^{-1}$. Field-emission scanning electron microscopy (FE-SEM) experiments were determined by using HITACHI S4800 Spectrometer. Powder X-ray diffraction (PXRD) were carried out on a Rigaku D/MAX 2550 diffractometer with $\text{Cu-K}\alpha$ radiation, 40 kV, 20 mA with the 2θ range of $10^\circ\sim 90^\circ$ (scanning rate of $10^\circ \text{ min}^{-1}$) at room temperature. Nitrogen sorption isotherm measurements were performed using a Micro meristics surface area and a pore size analyzer. Before measurement, the samples were degassed at 100°C for at least 12 h. A sample of ca. 100 mg and a UHP-grade nitrogen (99.999%) gas source were used for the nitrogen sorption measurements conducted at 77K and

collected on a Quantachrome Quadrasorb apparatus. Brunauer-Emmett-Teller (BET) surface areas were determined over a P/P_0 range from 0.01 to 0.20. Nonlocal density functional theory (NLDFT) pore size distributions were determined using the carbon/slit cylindrical pore mode of the Quadrawin software. X-ray photoelectron spectroscopy (XPS) was conducted on a Thermo Fischer ESCALAB 250Xi using a monochromatic Al K α (1486.8 eV) X-ray source with a spot size of 500 μm . The anode was operated at 12.5 kV and 16 mA. Ultraviolet absorption (UV) spectra were performed with TU-1901 double UV-Vis spectrophotometer.

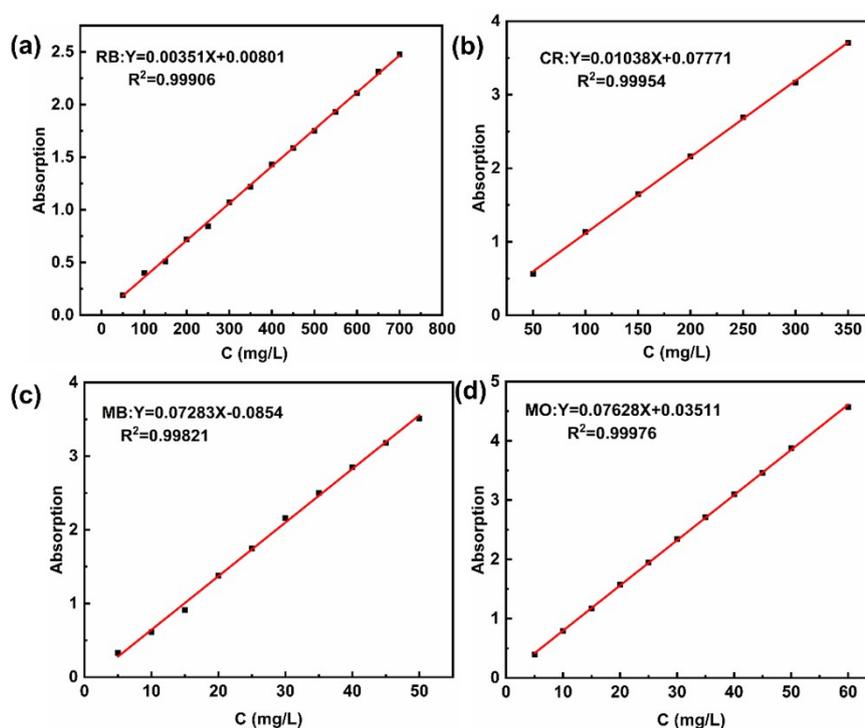


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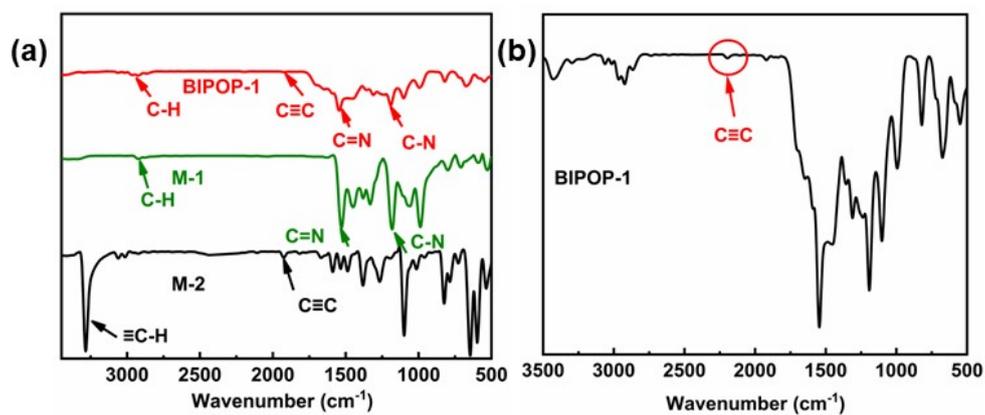


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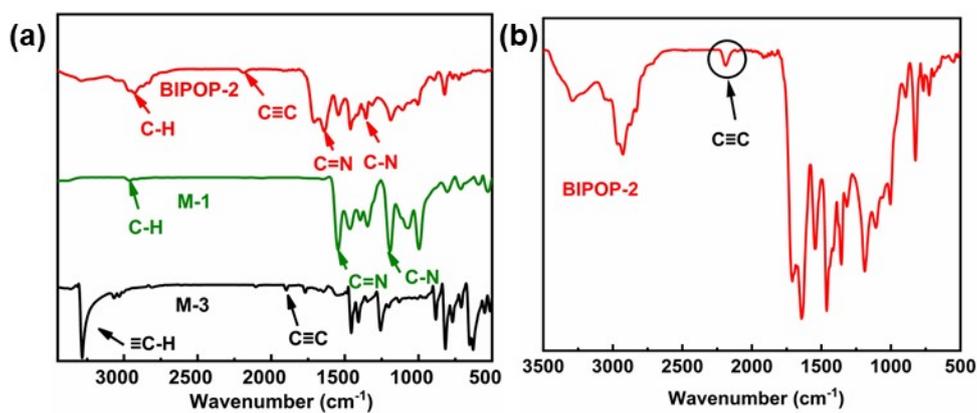


Fig. S3. (a) FT-IR spectroscopy of BIPOP-2, M-1 and M-3, (b) FT-IR spectroscopy of BIPOP-2 indicating the presence of C≡C

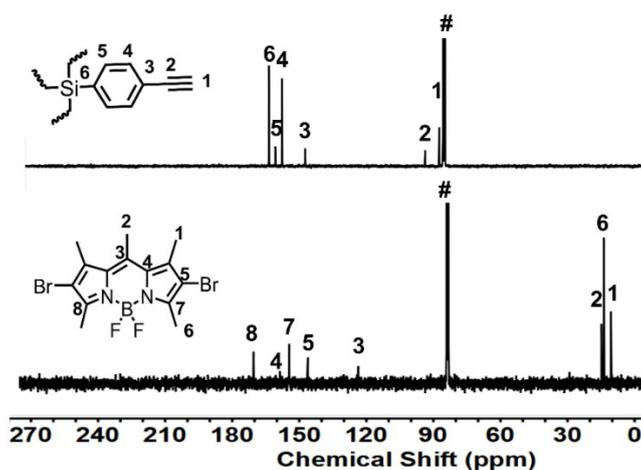


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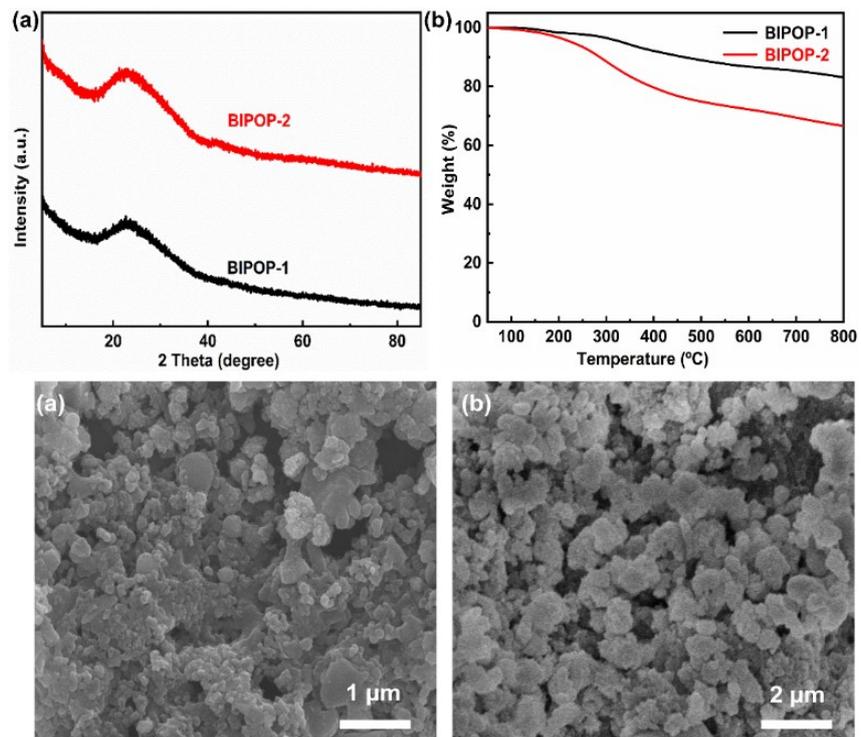


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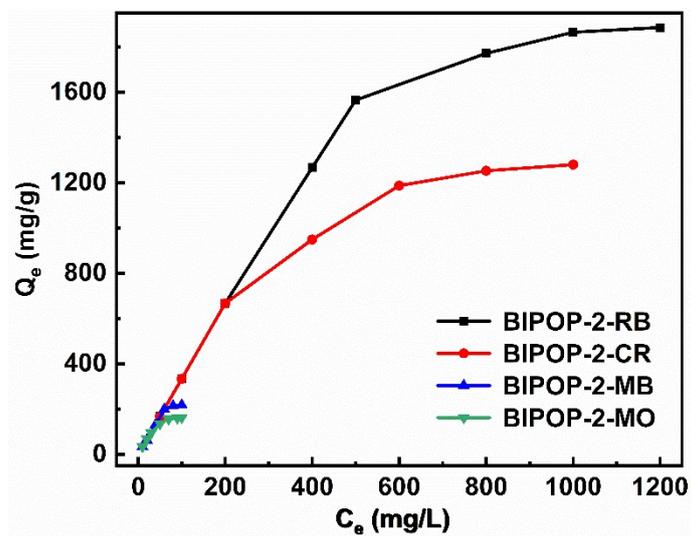


Fig. S6. Equilibrium adsorption isotherms of BIPOP-2 towards various dyes

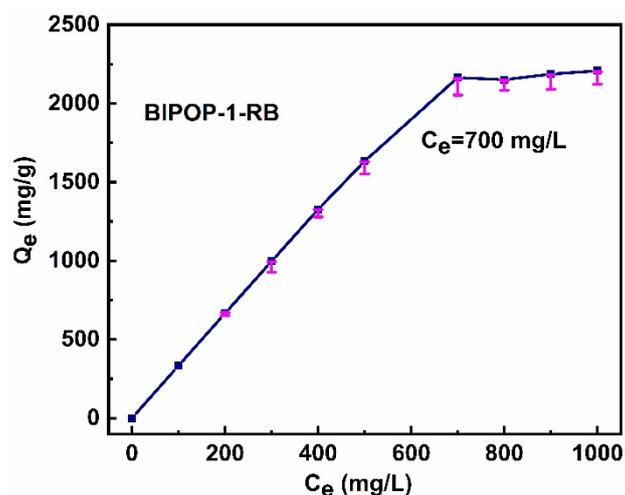


Fig. S7. Equilibrium adsorption isotherms of BIPOP-1 towards RB with error bars based on triplicate measurement

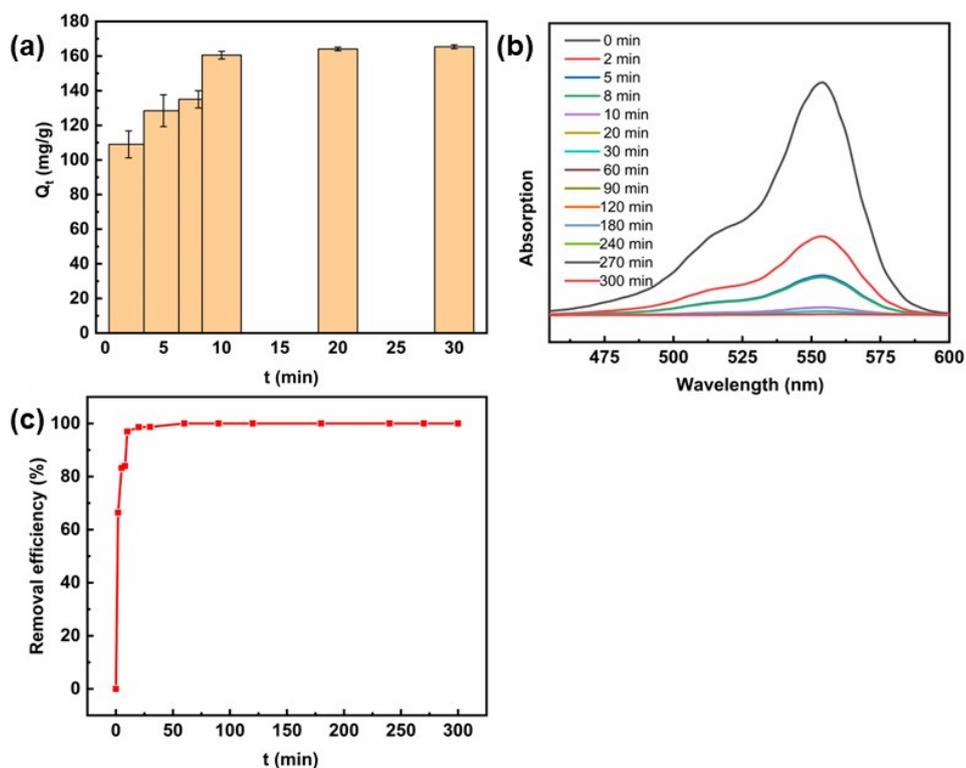


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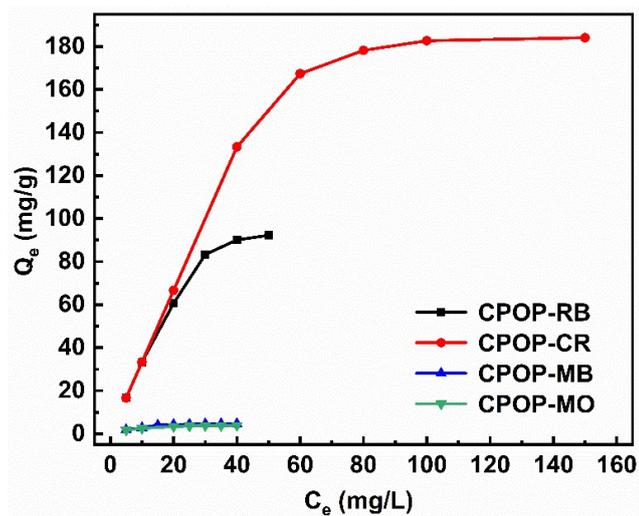


Fig. S9. Equilibrium adsorption isotherms of CPOP towards various dyes

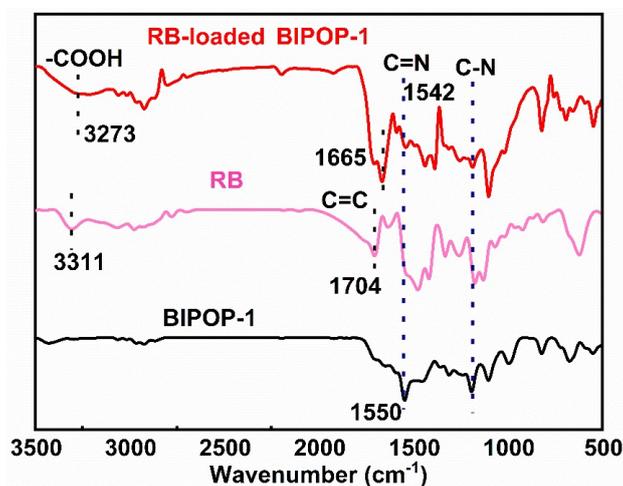


Fig. S10. FT-IR spectroscopy of BIPOP-1, RB and RB-loaded BIPOP-1

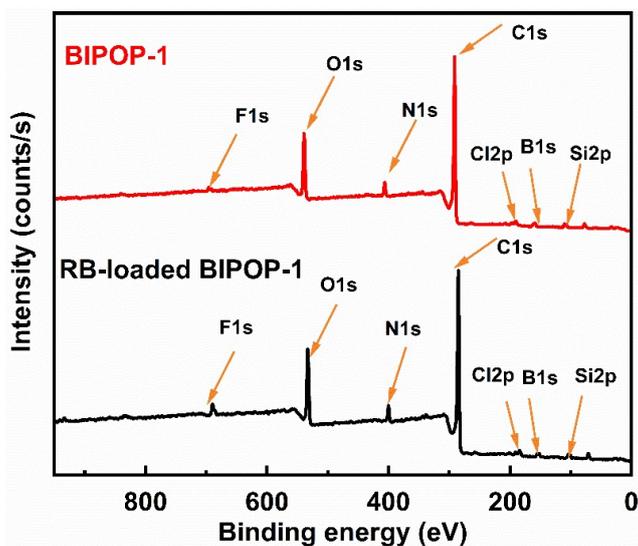


Fig. S11. Wide-scan XPS spectra of BIPOP-1 and RB-loaded BIPOP-1

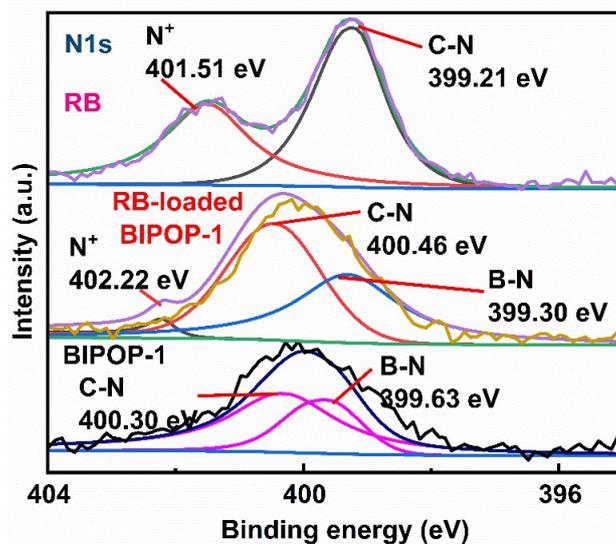


Fig. S12. High resolution XPS spectra of N1s

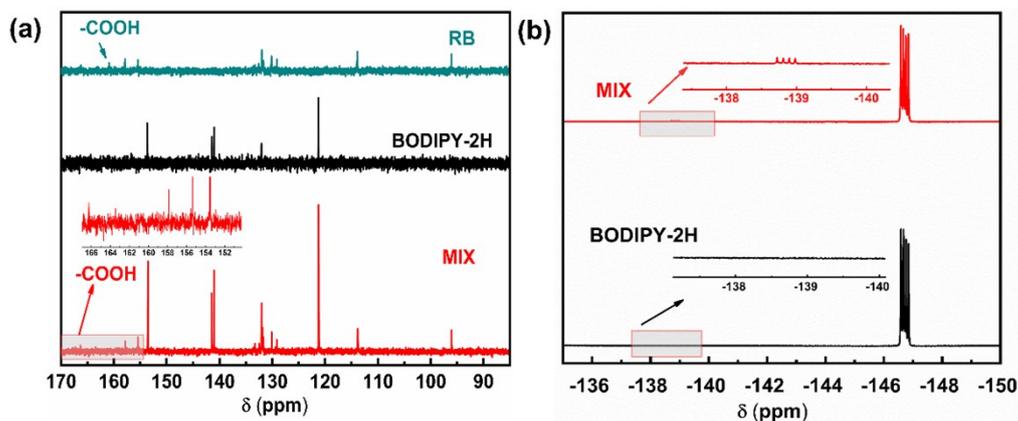


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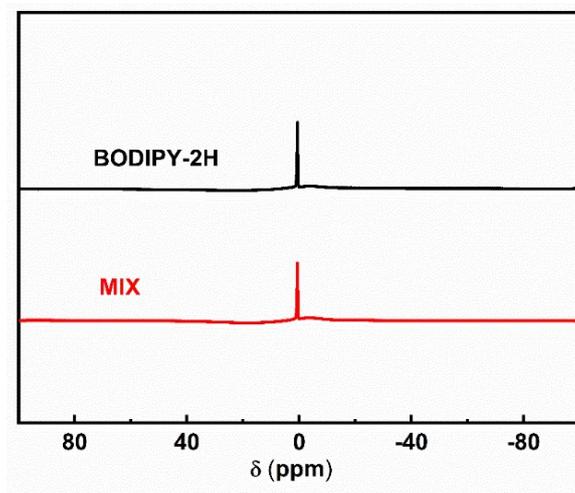


Fig. S14. The ^{11}B NMR spectra of BODIPY-2H and its mixture with RB

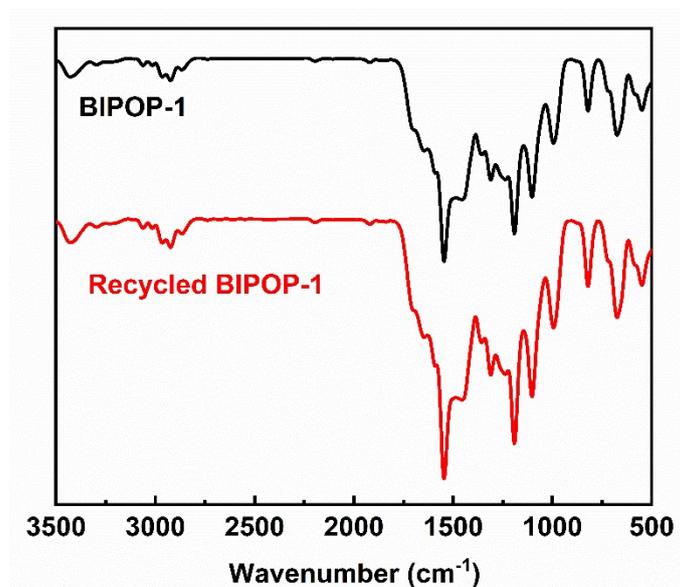


Fig. S15. The FT-IR spectroscopy of the pristine and recycled BIPOP-1

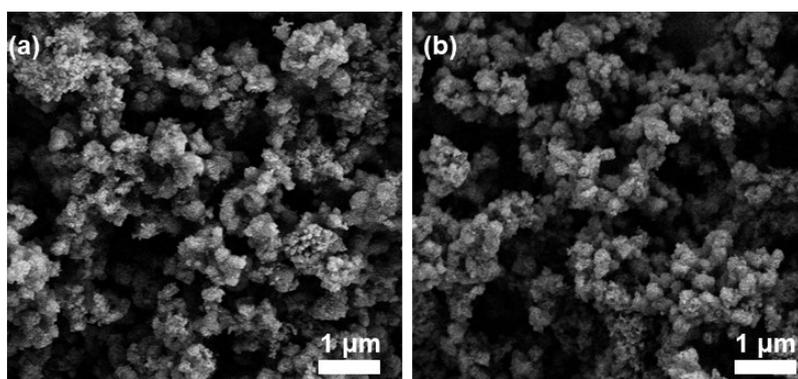


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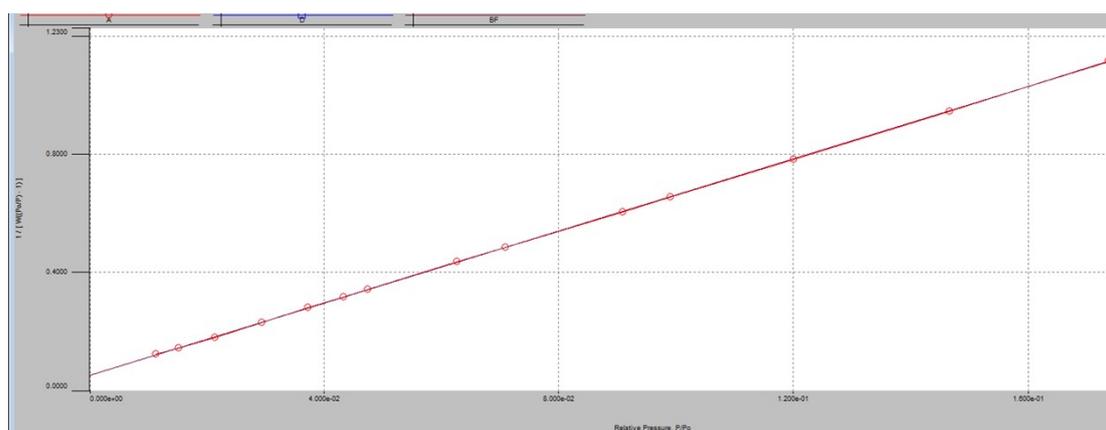


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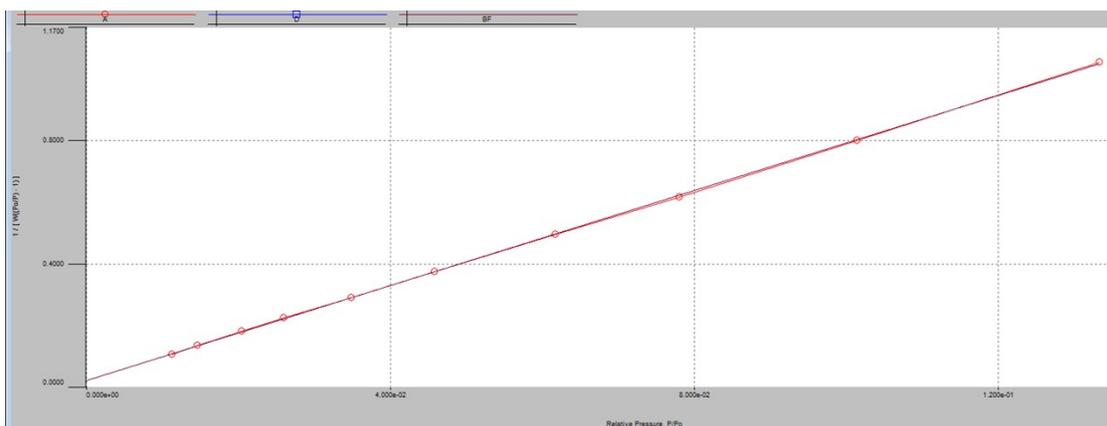


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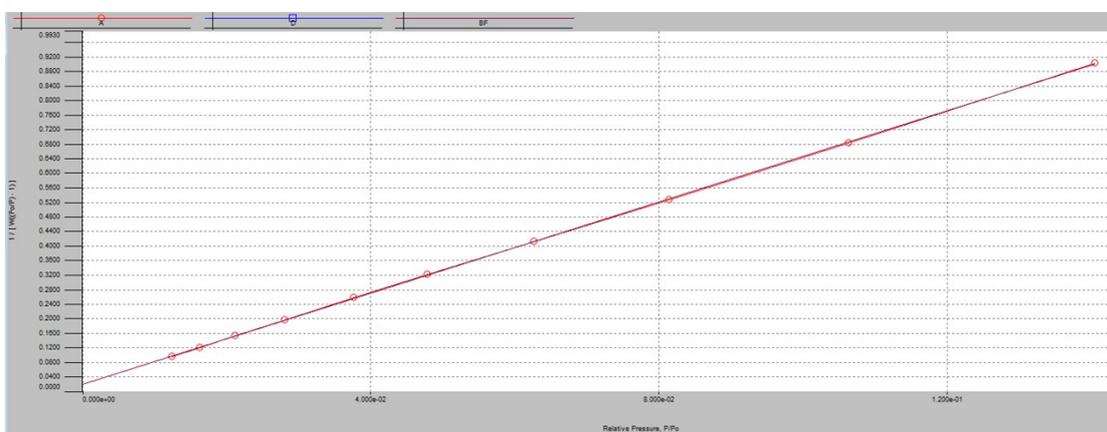


Fig. S19. BET plots of CPOP ($r = 0.999977$, $C = 325.499$)

Table S1. Comparison of the adsorption capacities of BIPOPs towards MO with other adsorbents

Adsorbents	Adsorption Capacity /mg g ⁻¹	Ref.
BIPOP-1	211.8	This work
BIPOP-2	162.0	This work
Zeolitic imidazole frameworks	1340	1
amorphous carbon nanotubes	253.26	2
UiO-66-NH ₂	242.72	3
activated carbon prepared from date pits	434	4
QPEI/SiO	105.4	5
SiO ₂ nanofibers	730.9	6
rambutan-like MnCo ₂ O ₄	185.14	7
Lignin derived ZSM-5 (PZ)	514	8

halloysite nanotubes/polypyrrole nanocomposites	214.6	9
chitosan/polyvinyl alcohol/zeolite electrospun composite nanofibrous membrane	153	10
copper-modified nanoalum Modified nickel ferrite nanocomposite/functionalized chitosan	139	11
calcined Zn-Al layered double hydroxide	551.2	12
magnetic aminated lignosulfonate/carbon	451.21	13
Sensitized Bentonite	121.28	14
sugar beet bagasse	99.3	15
$\text{Cu}_2\text{O}/\text{Bi}_2\text{O}_3$	221.5	16
Superparamagnetic nanosorbent	1533.2	17
CTA-CSM	240	18
	131.9	19

Table S2. Comparison of the adsorption capacity of BIPOPs on CR with other adsorbents

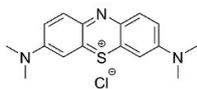
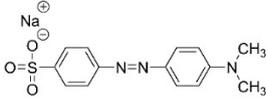
Adsorbents	Adsorption Capacity /mg g ⁻¹	Ref.
BIPOP-1	1477.4	This work
BIPOP-2	1280.1	This work
Congo red dye	1735	20
Zeolitic imidazole frameworks	3900	1
Nano-Cao	357.14	21
biochar derived from leather shavings	1916	22
Cationic Lignin Hydrogels	125.63	23
amorphous carbon nanotubes	467.97	2
titanium dioxide	152	24
$\text{Fe}_x\text{Co}_3\text{-xO}_4$ nanoparticles	128.6	25
$\text{Fe}_3\text{O}_4@n\text{SiO}_2@m\text{SiO}_2$	1428	26
fibrous xonotlite	574.71	27
Mg-Al-mixed metal oxide (MMO-E)	3470	28
Hierarchical porous Ni/Co-LDH hollow dodecahedron	909.2	29
Nano- Fe_3O_4	1395	30
Coconut husk-raw clay-Fe composite	1649.3	31

MOF-5/Cu	357.42	32
by coal-series kaolin	237.53	33
Zr-MOFs	1236.9	34

Table S3. Comparison of the adsorption capacity of BIPOPs on MB with other adsorbents

Adsorbents	Adsorption Capacity /mg g ⁻¹	Ref.
BIPOP-1	287	This work
BIPOP-2	217	This work
activated carbon prepared from date pits	455	4
carbonaceous adsorbent	140.25	35
imprinted polymer	3628.84	36
porous soy protein isolate	272.4	37
Porous Biochar	208	38
activated carbon prepared from Malawian baobab fruit shell wastes	334.45	39
Chitosan crosslinked composite	499.8	40
biochar from solid wastes	161	41
ZnCl ₂ -activated carbon	255.1	42
AC-alginate composite membrane	666	43
Mesoporous-Activated Carbon	1000	44
dimensional titanate nanosheets	3937	45
slow pyrolysis pine cone bio-char	106.4	46
Treated digested residue	285.71	47
magnetic alginate/rice husk bio-composite	274.9	48
nanofibrous membranes	3186.7	49
Stomatocyte-like hollow polydopamine nanoparticles	2896	50

Table S4. Size and molecular properties of dye molecules

Dyes	Molecular Structure	Molecular Size (nm)	Molecular Weight (g/mol)	Nature	Adsorption wavelength (nm)
MB		1.26*0.77*0.65	320	cationic	665
MO		1.31*0.55*0.18	327	anionic	464

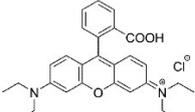
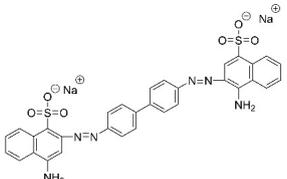
RB		1.59*1.18*0.56	479	cationic	554
CR		2.62*0.74*0.43	696	anionic	497

Table S5. The comparison of equilibrium adsorption capacities (Q_e) of RB adsorbed by BIPOP-1 in pure water and seawater from Bohai sea

	initial dye concentration (mg/L)	Q_e (mg g ⁻¹)
pure water	50	167
	500	1633
	1000	2207
seawater from Bohai sea	50	167
	500	1581
	1000	2191

Table S6. Electronic binding energy of various elements in BIPOP-1, RB, and RB-loaded BIPOP-1

		BIPOP-1	RB-loaded BIPOP-1	RB
B1s / eV	B-F	192.74	193.17	
	B-N	190.63	190.84	
F1s / eV	C-F	689.63	689.63	
	B-F	685.89	685.50	
Cl2p / eV	Cl ⁻		200.40	196.87
O1s / eV	C-O		533.45	533.48
	O-C=O		531.66	532.08
N1s / eV	C-N	400.30	400.46	399.21
	B-N	399.63	399.30	
	N ⁺		402.22	401.51

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