#### **Supporting Information**

### Portable Self-powered Photoelectrochemical Immunosensor Based on Cu<sub>3</sub>SnS<sub>4</sub>

### Nanoflower for Ultra-sensitive and Real-time Detection of Human Cytochrome c

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**Figure S1**. (**A**) SEM image of CuSn(OH)<sub>6</sub> (500 nm) and (**B**) SEM image of CuSn(OH)<sub>6</sub> (100 nm).

X-ray diffraction (XRD) was employed to analyze the purity and crystal structure of the products. As indicated in **Figure S2**, CuSn(OH)<sub>6</sub> is a tetragonal sulfide with five strong peaks located at 20 positions of 12.5°, 21.8°, 25.2°, 29.7°, 32.4°, 33.5°, 39.3°, 44.4°, 45.4°, 49.1°, 51.7°, 54.3°, 60.8°, 67.7° and 78.3°, corresponding to diffractions from the (100), (110), (200), (201), (102), (210), (202), (220), (212), (103), (400), (203), (410), (204) and (332) planes of the CuSn(OH)<sub>6</sub> crystal, which is consistent with the description in the literature<sup>1, 2</sup>.



Figure S2. (A) XRD patterns of CuSn(OH)<sub>6</sub>; (B) XPS spectra of Cu<sub>3</sub>SnS<sub>4</sub>.



Figure S3. (A) Plots of  $(\alpha hv)^2$  vs. (hv) used to evaluate the optical bandgaps of Cu<sub>3</sub>SnS<sub>4</sub>; (B)

Mott-Schottky curves of Cu<sub>3</sub>SnS<sub>4</sub>.



Figure S4. Photocurrent response of electrodes of different materials.



Figure S5. Effects of (A) the concentration of  $Cu_3SnS_4$ ; (B) the pH value of the PBS; (C) the concentration of AA; (D) the incubation time of Cyt *c*.



Figure S6. (A) Reproducibility (Cyt c: 100 pM) and (B) Storage stability of the PEC immunosensor.



Figure S7. Real-time detection of Cyt *c* system.

Electrode	$R_{\rm s}\left(\Omega ight)$	$R_{\rm et}\left(\Omega\right)$	Ι
$Cu_3SnS_4$	22.86	22.13	49.8μΑ
Cu <sub>3</sub> SnS <sub>4</sub> /CS	23.53	42.08	35.6µА
Cu <sub>3</sub> SnS <sub>4</sub> /CS/anti-Cyt c	23.97	68.29	28.8µA
Cu <sub>3</sub> SnS <sub>4</sub> /CS/anti-Cyt c/BSA	23.70	79.75	19.6µA
Cu <sub>3</sub> SnS <sub>4</sub> /CS/anti-Cyt <i>c</i> /BSA/Cyt <i>c</i>	23.30	97.00	12.6µA

# Table S1 Simulation parameters of the equivalent circuit components

# Table S2 Different Cyt c detection methods were compared

Method	Nanomaterials	Linear range	Detection limit	Referenc es
Fluorescence	CdTe	0.5 μΜ-2.5 μΜ	0.5 μΜ	3
Fluorescence	VS <sub>2</sub> -Nanosheet	0.75 nM-50 nM	0.5 nM	4

Fluorescence	DNA-AgNCs@tween 80	0.8 nM-20000 nM	0.8 nM	5
Colorimetric, chemiluminescence	β-Co(OH) <sub>2</sub> CMk	1 pM-5 μM, 50 μM-1 mM	2 fM	6
EIS	Cyt-c/Polypyrrole/SPE	10 pM-1 nM	5 pM	7
EIS	CdS/graphene	1 nM-100 nM	0.3 nM	8
PEC	CdS/CuInS <sub>2</sub> /Au/TiO <sub>2</sub>	5 pM-100 nM	5 pM	9
PEC	Cu <sub>3</sub> SnS <sub>4</sub>	1 fM-1000 nM	0.35 fM	This work

Table S3. Analytical results of Cyt c in human serum samples

Sample	Added (nM)	Detection content (nM, n=3)	Average content (nM)	RSD (%)	Recovery (%)
Serum #789	0.1	0.085、0.095、0.108	0.096	1.55	96.17
Serum #757	0.1	0.085, 0.116, 0.92	0.098	1.89	97.63
Serum #730	0.5	0.465、0.554、0.486	0.502	3.64	100.4
Serum #729	1	1.065、0.851、0.956	0.958	1.34	95.76

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