

Supporting Information

**Transition-metal free oxidative carbo-carboxylation of alkenes with
formate in air**

Xu,^{†,a} Hui Xu,^{†,a} Sai Wang,^a Tian-Zi Hao,^a Si-Yi Yan,^a Dong Guo,^{*,a} Xu Zhu^{*,a,b}

^a Jiangsu Key Laboratory of New Drug Research and Clinical Pharmacy, School of Pharmacy, Xuzhou Medical University, 209 Tongshan Road, Xuzhou 221004, China.

^b Key Laboratory of Organic Synthesis of Jiangsu Province, College of Chemistry, Chemical Engineering and Materials Science, Soochow University, Suzhou, 215123, China.

Email: guo@xzhmu.edu.cn; xuzhu@xzhmu.edu.cn

Table of Contents

1. General Information.....	2
2. Optimization of Reaction Conditions.....	2
3. Synthesis of Substrates	4
4. Succinic Acid Derivatives	14
5. The Application of the Reaction	24
6. Control Experiments	26
 6.1 TEMPO Trapping Experiment.....	26
 6.2 D-Labeling Experiment.....	27
 6.3 ¹³C Isotope Labeling Experiment and Detection of H₂O₂.....	27
 6.4 Stern-Volmer Fluorescence Quenching Analysis	29
7. References.....	30
8. NMR Spectra.....	31

1. General Information

¹H NMR (400 MHz) spectra, ¹³C NMR (100 MHz) spectra, and ¹⁹F NMR (376 MHz) spectra were recorded on a JEOL ECZ400 (400 MHz) spectrometers in CDCl₃, CD₃OD or DMSO-*d*₆. The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublet, td = triplet of doublet, ddd = doublet of doublet of doublet, sep = septet, m = multiplet, br = broad. Flash column chromatography was performed using Nuo Tai silica gel (Size: 200-300) with distilled solvents. High Resolution Mass Spectra were obtained from the Xuzhou Medical University Mass Spectral facility.

All reactions were set up on the bench top and conducted under air or nitrogen atmosphere while subject to irradiation from blue LEDs (25 W, $\lambda_{\text{max}} = 450$ nm). Reagents and solvents and photocatalysts were purchased from various vendors and used as received, unless stated otherwise. Thin-layer chromatography (TLC) was performed on 0.2-0.3 mm SiliCycle silica gel F-254 plates.

2. Optimization of Reaction Conditions

General Procedure:

An oven-dried Schlenk tube (10 mL) containing a stirring bar was charged with the substrate (0.2 mmol), HCOONa (60.8 mg, 1.0 mmol, 5.0 equiv), DABCO (11.2 mg, 0.1 mmol, 50 mol%), K₃PO₄ (42.5 mg, 0.2 mmol, 1.0 equiv) and 4CzIPN (3.2 mg, 0.004 mmol, 2 mol%). Then anhydrous DMSO (2 mL) was added. Finally, the Schlenk tube was placed in an oil bath at the distance of 3 cm from a 25 W blue LED (wavelength: 450 nm) and stirred at 60 °C for 12 - 48 hours. After completion of the reaction, the reaction mixture was quenched with 2 mL of HCl (2 N), diluted with water (20 mL) and extracted with EtOAc (3 x 20 mL). The combined organic layers were washed with water and brine before dried over sodium sulfate and concentrated under reduced pressure. The residue was purified by silica gel flash chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2) to give the pure desired product.

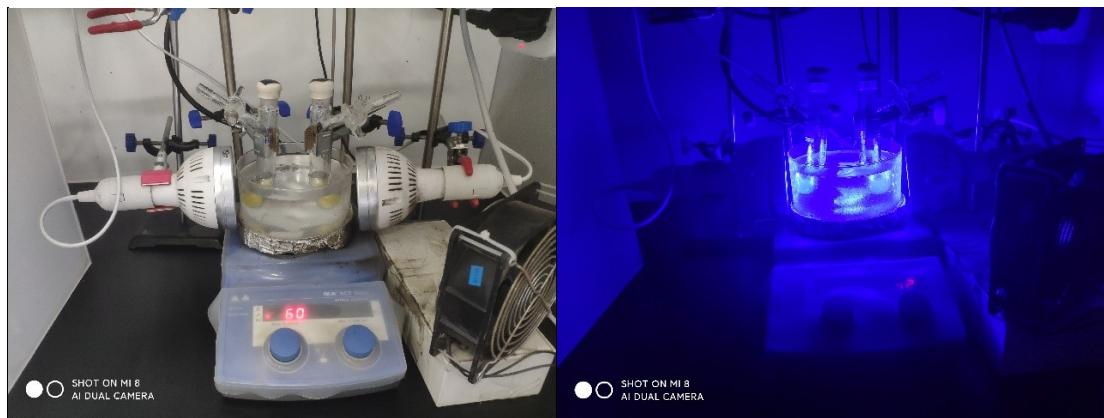
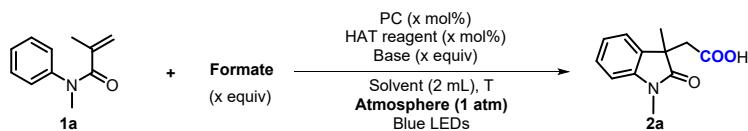


Figure S1. Standard setup for reactions

Table S1. Screening table of the reaction of *N*-methyl-*N*-phenylmethacrylamide with formate.



Entry	PC (x mol%)	Formate (x equiv)	Base (x equiv)	HAT reagent (x mol%)	Solvent (2 mL)	Atmosphere	T (°C)	Yield of 2a ^a (%)	RSM
1	4CzIPN (2)	HCOONa (5)	Cs ₂ CO ₃ (3)	DABCO(50)	DMSO	Air	rt	20	0
2	4CzIPN (2)	HCOONa (5)	Cs ₂ CO ₃ (3)	DABCO(50)	DMSO	Air	50	43	0
3	4CzIPN (2)	HCOONa (5)	Cs ₂ CO ₃ (3)	DABCO(50)	DMSO	N ₂	50	10(0) ^b	0
4	4CzIPN (2)	HCOONa (5)	Cs ₂ CO ₃ (3)	DABCO(50)	DMSO	O ₂	50	ND	25
5	4CzIPN (2)	HCOONa (5)	KH ₂ PO ₄ (3)	DABCO(50)	DMSO	Air	50	35	0
6	4CzIPN (2)	HCOONa (5)	Na ₃ PO ₄ (3)	DABCO(50)	DMSO	Air	50	42	0
7	4CzIPN (2)	HCOONa (5)	Na ₂ HPO ₄ (3)	DABCO(50)	DMSO	Air	50	40	0
8	4CzIPN (2)	HCOONa (5)	Na ₂ CO ₃ (3)	DABCO(50)	DMSO	Air	50	36	0
9	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (3)	DABCO(50)	DMSO	Air	50	47	0
10	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (0.5)	DABCO(50)	DMSO	Air	50	36	0
11	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	50	51	0
12	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (2)	DABCO(50)	DMSO	Air	50	49	0
13	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (4)	DABCO(50)	DMSO	Air	50	51	0
14	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (3)	—	DMSO	Air	50	12	68
15	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (3)	DIPEA(50)	DMSO	Air	50	17	41
16	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (3)	Quinuclidine(50)	DMSO	Air	50	23	8
17	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(20)	DMSO	Air	50	26	23
18	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(80)	DMSO	Air	50	46	13
19	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	25	39	0
20	4CzIPN (2)	HCOONa (5)	K₃PO₄ (1)	DABCO(50)	DMSO	Air	60	62^c	0
21	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	70	56	0
22	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	80	52	0
23	4CzIPN (2)	HCOOLi (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	46	0
24	4CzIPN (2)	HCOOK (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	52	0
25	4CzIPN (2)	HCOOCs (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	60	0
26	4CzIPN (2)	HCOONH₄ (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	29	0
27	4CzIPN (2)	HCOONA (1)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	20	5
28	4CzIPN (2)	HCOONA (3)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	55	0
29	4CzIPN (2)	HCOONa (7)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	42	0
30	—	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	0	100
31	4DPAIPN (2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	42	0
32	fac-Ir(ppy)₃(2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	55	0
33	Ir[(dtpy)(ppy)₂]PF₆(2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	55	0
34	Eosin B (2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	55	14
35	Eosin Y(2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	43	0
36	4CzIPN (1)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	47	0
37	4CzIPN (3)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMSO	Air	60	48	0
38	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMF	Air	60	42	0
39	4CzIPN (2)	HCOONa (5)	K ₃ PO ₄ (1)	DABCO(50)	DMA	Air	60	10	0

^aYields were determined by ¹H NMR analysis with 1,2-dichloroethane as an internal standard.

^bSolvent was degassed by three vacuum-purge/nitrogen-flush cycles or freeze-pump-thaw (in parentheses).

^cIsolated yields.

Large Scale Reaction Procedure:

An oven-dried Schlenk tube (50 mL) containing a stirring bar was charged with the *N*-(4-(*tert*-butyl)phenyl)-*N*-methylmethacrylamide (0.4627 g, 2.0 mmol), HCOONa (0.6801 g, 10.0 mmol, 5.0 equiv), DABCO (0.1122 g, 0.1 mmol, 50 mol%), K₃PO₄ (0.4245 g, 0.2 mmol, 1.0 equiv) and 4CzIPN (31.6 mg, 0.004 mmol, 2 mol%). The anhydrous DMSO (20 mL) were added. Finally, the Schlenk tube was placed in an oil bath at the distance of 3 cm from a 45 W blue LED (wavelength: 450 nm) and stirred at 60 °C for 12 hours. After completion of the reaction, the reaction mixture was quenched with 10 mL of HCl (2 N), diluted with water (60 mL) and extracted with EtOAc (3 x 60 mL). The combined organic layers were washed with water and brine before dried over sodium sulfate and concentrated under reduced pressure. The residue was purified by silica gel flash chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2) to give the pure desired product.

3. Synthesis of Substrates

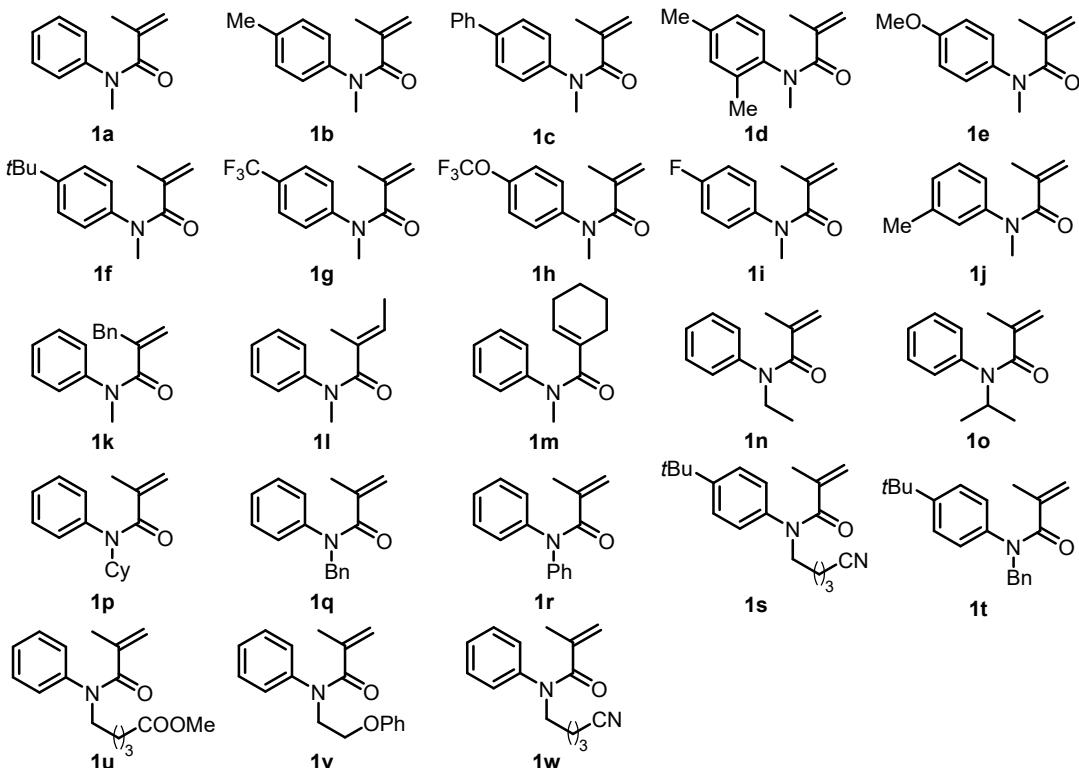
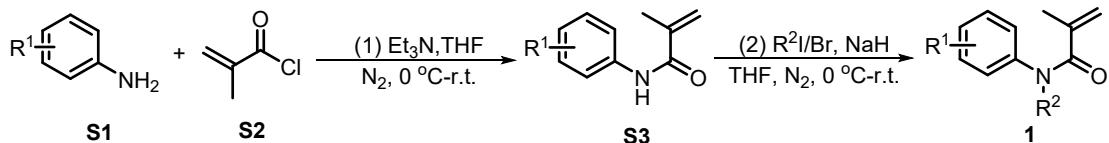


Figure S3. The scope of various alkenes.

General Procedure A: substrates **1a-1j**, **1n-1o**, **1q**, **1s-1w** were synthesized following the reported procedures in the literature.^[1]

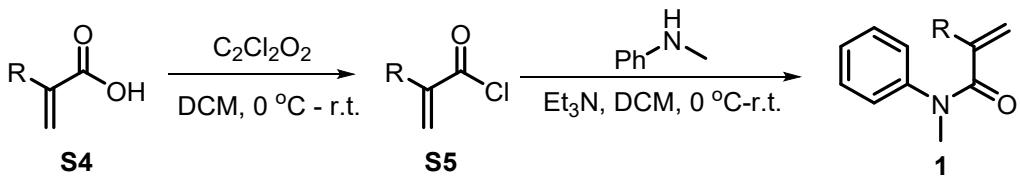
General Procedure A for Preparation of Substrates



To a 50 mL round-bottom flask was added the solution of corresponding aniline **S1** (10.0 mmol) in THF (15 mL) and triethylamine (2.1 mL, 15.0 mmol, 1.5 equiv). The mixture was stirred at 0 °C, and methacryloyl chloride **S2** (1.2 mL, 12 mmol, 1.2 equiv) was added slowly under nitrogen atmosphere. The resulting solution was stirred at room temperature for 6 hours, quenched with H₂O (15 mL), and extracted with EtOAc (20 mL x 3). The combined organic layers were washed with brine, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by flash column chromatography on a silica gel using petroleum ether and EtOAc (5:1, v/v) as the eluent to give corresponding intermediates **S3**.

To a 50 mL round-bottom flask was added the solution of corresponding intermediates **S3** (2.0 mmol) in THF (10 mL). The mixture was stirred at 0 °C, and NaH (0.07 g, 3.0 mmol, 1.5 equiv) was added slowly. Then the reaction mixture was stirred at 0 °C for 15 min followed by addition of halohydrocarbon (3.0 mmol, 1.5 equiv). The resulting solution was stirred at room temperature for 8 hours, quenched with H₂O (15 mL), and extracted with EtOAc (3 x 20 mL). The combined organic layers were washed with brine, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by flash chromatography on a silica gel using petroleum ether and EtOAc (5:1, v/v) as the eluent to give corresponding compounds **1a-1j**, **1n-1o**, **1q**, **1s-1w**.

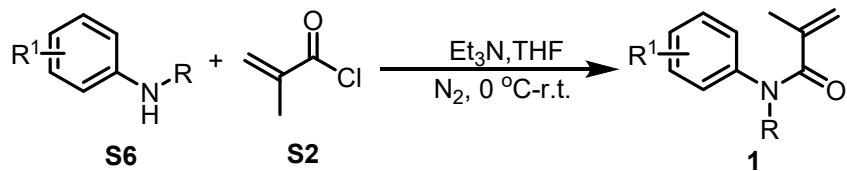
General Procedure B: substrates **1k-1m** were synthesized following the reported procedures in the literature.^[2]



The acid **S4** (2.0 mmol) and DCM (10 mL) were added to a 50 mL round-bottom flask. The mixture was stirred at 0 °C, and oxalyl chloride (3.0 mmol, 2 equiv) was added slowly. The resulting solution was then stirred at room temperature for 8 hours. The solvent was removed under reduced pressure to get product **S5**, which was used directly to the next step.

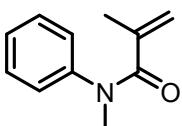
To a 50 mL round-bottom flask was added the solution of *N*-methylaniline (2.0 mmol) in DCM (15 mL) and triethylamine (4.0 mmol, 2.0 equiv). The mixture was stirred at 0 °C, and added slowly the solution of **S5** (0.31 g, 3.0 mmol, 1.5 equiv) in DCM (5 mL) under argon atmosphere. The resulting solution was stirred at room temperature for 8 hours, followed by the addition of H₂O (10 mL) to quench excess acyl chloride, and extracted with DCM (3 x 20 mL). The combined organic layer was washed with brine, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by flash chromatography on a silica gel using petroleum ether and EtOAc (5:1, v/v) as the eluent to give product **1k-1m**.

General Procedure C: substrates **1p, 1r** were synthesized following the reported procedures in the literature.^[1]



To a 50 mL round-bottom flask was added the solution of corresponding aniline **S1** (10.0 mmol) in THF (15 mL) and triethylamine (2.1 mL, 15.0 mmol, 1.5 equiv). The mixture was stirred at 0 °C, and added slowly methacryloyl chloride **S2** (1.2 mL, 12

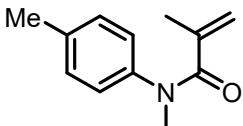
mmol, 1.2 equiv) under nitrogen atmosphere. The resulting solution was stirred at room temperature for 6 hours, quenched with H₂O (15 mL), and extracted with EtOAc (20 mL x 3). The combined organic layer was washed with brine, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by flash column chromatography on a silica gel using petroleum ether and EtOAc (5:1, v/v) as the eluent to give product **1p, 1r**.



N-methyl-N-phenylmethacrylamide (1a): Prepared according to general procedure A using aniline and iodomethane. The spectra matched with reported literature.^[1]

¹H NMR (400 MHz, CDCl₃) δ 7.35 (td, *J* = 6.8, 1.6 Hz, 2H), 7.27 (dd, *J* = 2.8, 1.6 Hz, 1H), 7.16 – 7.12 (m, 2H), 5.05 – 5.02 (m, 1H), 5.00 – 4.98 (m, 1H), 3.35 (s, 3H), 1.76 (s, 3H).

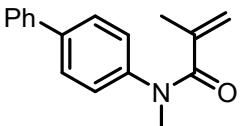
¹³C NMR (100 MHz, CDCl₃) δ 172.0, 144.7, 140.8, 129.3, 127.0, 126.6, 119.4, 37.7, 20.4.



N-methyl-N-(*p*-tolyl)methacrylamide (1b): Prepared according to general procedure A using *p*-toluidine and iodomethane. The spectra matched with reported literature.^[1]

¹H NMR (400 MHz, CDCl₃) δ 7.06 (d, *J* = 7.6 Hz, 2H), 6.94 (d, *J* = 7.6 Hz, 2H), 5.01 – 4.86 (m, 2H), 3.24 (s, 3H), 2.26 (s, 3H), 1.68 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 171.9, 141.9, 140.7, 136.7, 129.7, 126.2, 119.0, 37.6, 20.9, 20.3.

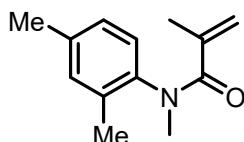


N-([1,1'-biphenyl]-4-yl)-N-methylmethacrylamide (1c): Prepared according to general procedure A [1,1'-biphenyl]-4-amine and iodomethane. The spectra matched with reported literature.^[3]

¹H NMR (400 MHz, CDCl₃) δ 7.57 (t, *J* = 7.2 Hz, 4H), 7.44 (t, *J* = 7.2 Hz, 2H), 7.35 (t, *J* = 8.0 Hz, 1H), 7.20 (d, *J* = 8.0 Hz, 2H), 5.06 (d, *J* = 10.8 Hz, 2H), 3.38 (s, 3H), 1.81 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 172.0, 143.8, 140.7, 140.0, 139.8, 128.9, 127.9, 127.6,

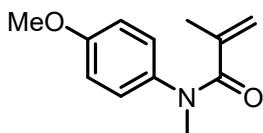
127.0, 126.8, 119.6, 37.7, 20.4.



N-(2,4-dimethylphenyl)-N-methylmethacrylamide (1d): Prepared according to general procedure A using 3,5-dimethylaniline and iodomethane.^[4]

¹H NMR (400 MHz, CDCl₃) δ 7.04 (s, 1H), 6.97 (d, *J* = 9.2 Hz, 1H), 6.92 (d, *J* = 8.0 Hz, 1H), 4.95 (d, *J* = 1.2 Hz, 2H), 3.20 (s, 3H), 2.32 (s, 3H), 2.21 (s, 3H), 1.73 (s, 3H).

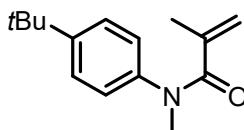
¹³C NMR (100 MHz, CDCl₃) δ 172.0, 140.6, 140.5, 137.6, 134.4, 131.8, 127.8, 127.5, 118.1, 36.6, 20.9, 20.2, 17.5.



N-(4-methoxyphenyl)-N-methylmethacrylamide (1e): Prepared according to general procedure A using 4-methoxyaniline and iodomethane. The spectra matched with reported literature.^[1]

¹H NMR (400 MHz, CDCl₃) δ 7.08 – 7.02 (m, 2H), 6.89 – 6.81 (m, 2H), 5.01 (d, *J* = 13.6 Hz, 2H), 3.81 (s, 3H), 3.31 (s, 3H), 1.74 (s, 3H).

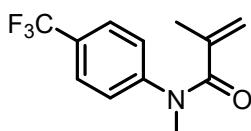
¹³C NMR (100 MHz, CDCl₃) δ 171.5, 157.9, 140.5, 136.9, 127.3, 118.4, 114.0, 54.9, 37.3, 20.0.



N-(4-(tert-butyl)phenyl)-N-methylmethacrylamide (1f): Prepared according to general procedure A using 4-(*tert*-butyl)aniline and iodomethane. The spectra matched with reported literature.^[5]

¹H NMR (400 MHz, CDCl₃) δ 7.34 (d, *J* = 8.4 Hz, 2H), 7.05 (d, *J* = 8.4 Hz, 2H), 5.02 (d, *J* = 8.0 Hz, 2H), 3.33 (s, 3H), 1.75 (s, 3H), 1.32 (s, 9H).

¹³C NMR (100 MHz, CDCl₃) δ 171.9, 149.8, 141.8, 140.7, 126.0, 125.9, 119.2, 37.6, 34.5, 31.3, 20.3.

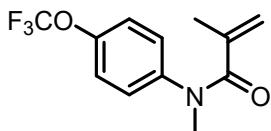


N-methyl-N-(4-(trifluoromethyl)phenyl)methacrylamide (1g): Prepared according to general procedure A using 4-(trifluoromethyl)aniline and iodomethane. The spectra matched with reported literature.^[1]

¹H NMR (400 MHz, CDCl₃) δ 7.54 (d, *J* = 8.0 Hz, 2H), 7.21 (d, *J* = 8.0 Hz, 2H), 4.97 (d, *J* = 44.6 Hz, 2H), 3.31 (s, 3H), 1.74 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 171.8, 147.8, 140.2, 128.6 (d, *J_{FC}* = 32.6 Hz), 126.4, 126.3 (d, *J_{FC}* = 3.7 Hz), 126.2(6), 123.8 (q, *J_{FC}* = 270.6 Hz), 37.4, 20.0.

¹⁹F NMR (376 MHz, CDCl₃) δ -62.4.

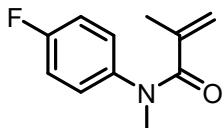


N-methyl-N-(4-(trifluoromethoxy)phenyl)methacrylamide (1h): Prepared according to general procedure A using 4-(trifluoromethoxy)aniline and iodomethane. The spectra matched with reported literature.^[5]

¹H NMR (400 MHz, CDCl₃) δ 7.21 – 7.15 (m, 4H), 5.09 (dt, *J* = 2.8, 1.2 Hz, 1H), 5.00 (p, *J* = 1.0 Hz, 1H), 3.35 (s, 3H), 1.79 (dd, *J* = 1.6, 1.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 171.9, 147.5 (d, *J_{FC}* = 1.9 Hz), 143.1, 140.4, 127.8, 120.4 (q, *J_{FC}* = 260.2 Hz), 119.7, 37.7, 20.2.

¹⁹F NMR (376 MHz, CDCl₃) δ -58.1.

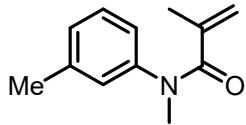


N-(4-fluorophenyl)-N-methylmethacrylamide (1i): Prepared according to general procedure A using 4-fluoroaniline and iodomethane. The spectra matched with reported literature.^[1]

¹H NMR (400 MHz, CDCl₃) δ 7.12 (dd, *J* = 8.8, 4.8 Hz, 2H), 7.04 (t, *J* = 8.4 Hz, 2H), 5.02 (d, *J* = 31.2 Hz, 2H), 3.33 (s, 3H), 1.77 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 172.1, 161.2 (d, *J_{FC}* = 245.8 Hz), 140.6, 128.4, 128.3, 119.5, 116.2 (d, *J_{FC}* = 22.6 Hz), 37.9, 20.4.

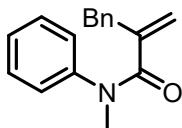
¹⁹F NMR (376 MHz, CDCl₃) δ -114.5.



N-methyl-N-(m-tolyl)methacrylamide (1j): Prepared according to general procedure A using *m*-toluidine and iodomethane. The spectra matched with reported literature.^[1]

¹H NMR (400 MHz, CDCl₃) δ 7.25 – 7.14 (m, 3H), 7.04 (d, *J* = 7.2 Hz, 1H), 4.95 (d, *J* = 5.6 Hz, 2H), 3.23 (s, 3H), 2.26 (s, 3H), 1.74 (s, 3H).

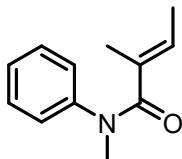
¹³C NMR (100 MHz, CDCl₃) δ 172.1, 144.7, 140.9, 139.3, 129.1, 127.8, 127.2, 123.7, 119.3, 37.8, 21.4, 20.5.



2-benzyl-N-methyl-N-phenylacrylamide (1k): Prepared according to general procedure B using 2-benzylacrylic acid. The spectra matched with reported literature.^[1]

¹H NMR (400 MHz, CDCl₃) δ 7.28 – 7.22 (m, 6H), 7.07 (d, *J* = 6.8 Hz, 2H), 6.82 (d, *J* = 6.8 Hz, 2H), 4.96 (d, *J* = 15.6 Hz, 2H), 3.46 (s, 2H), 3.29 (s, 3H).

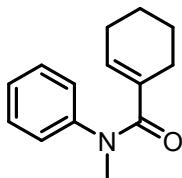
¹³C NMR (100 MHz, CDCl₃) δ 171.2, 144.5, 144.4, 137.8, 129.6, 129.2, 128.5, 127.0, 126.9, 126.6, 119.6, 40.4, 38.0.



(E)-N,2-dimethyl-N-phenylbut-2-enamide (1l): Prepared according to general procedure B using (*E*)-2-methylbut-2-enoic acid. The spectra matched with reported literature.^[2]

¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.30 (m, 2H), 7.24 – 7.19 (m, 1H), 7.10 (dt, *J* = 8.8, 2.0 Hz, 2H), 5.74 (qq, *J* = 6.8, 1.2 Hz, 1H), 3.34 (s, 3H), 1.58 – 1.52 (m, 3H), 1.48 (dd, *J* = 6.8, 1.2 Hz, 3H).

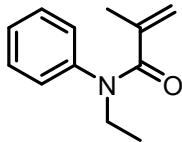
¹³C NMR (100 MHz, CDCl₃) δ 173.5, 145.1, 132.7, 130.6, 129.2, 126.5, 126.5, 37.9, 14.2, 13.4.



N-methyl-N-phenylcyclohex-1-ene-1-carboxamide (1m): Prepared according to general procedure B using cyclohex-1-ene-1-carboxylic acid. The spectra matched with reported literature.^[2]

¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.29 (m, 2H), 7.25 – 7.18 (m, 1H), 7.14 – 7.09 (m, 2H), 5.83 (dt, *J* = 3.6, 2.0 Hz, 1H), 3.34 (s, 3H), 1.99 – 1.81 (m, 4H), 1.42 (m, 4H).

¹³C NMR (100 MHz, CDCl₃) δ 172.6, 144.8, 134.4, 132.5, 128.9, 126.4, 126.3, 37.6, 25.8, 24.8, 21.9, 21.3.

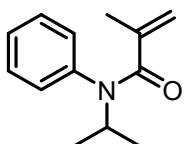


N-ethyl-N-phenylmethacrylamide (1n): Prepared according to general procedure A using aniline and bromoethane. The spectra matched with reported literature.^[6]

¹H NMR (400 MHz, CDCl₃) δ 7.35 (dd, *J* = 8.8, 6.8 Hz, 2H), 7.28 (d, *J* = 7.6 Hz, 1H),

7.11 (dd, $J = 7.6$, 2.0 Hz, 2H), 5.00 (d, $J = 12.4$ Hz, 2H), 3.84 (q, $J = 7.2$ Hz, 2H), 1.75 (s, 3H), 1.15 (t, $J = 7.2$ Hz, 3H).

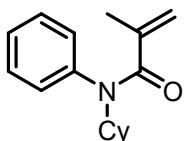
^{13}C NMR (100 MHz, CDCl_3) δ 171.5, 142.7, 140.8, 129.1, 127.5, 127.1, 119.1, 44.6, 20.3, 12.8.



N-isopropyl-N-phenylmethacrylamide (1o): Prepared according to general procedure A using aniline and 2-bromopropane. The spectra matched with reported literature.^[7]

^1H NMR (400 MHz, CDCl_3) δ 7.38 – 7.27 (m, 3H), 7.09 – 7.02 (m, 2H), 4.97 (dt, $J = 13.2$, 6.4 Hz, 1H), 4.90 (s, 1H), 4.85 (s, 1H), 1.73 (s, 3H), 1.12 (d, $J = 6.8$ Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3) δ 172.0, 141.7, 139.2, 130.4, 128.8, 127.8, 117.8, 46.8, 21.1, 20.8.

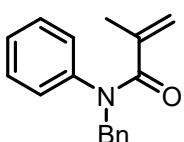


N-cyclohexyl-N-phenylmethacrylamide (1p): Prepared according to general procedure C using *N*-cyclohexylaniline.

^1H NMR (400 MHz, CDCl_3) δ 7.35 – 7.30 (m, 3H), 7.09 – 7.03 (m, 2H), 4.86 (d, $J = 24.4$ Hz, 2H), 4.56 (t, $J = 11.6$ Hz, 1H), 1.92 – 1.82 (m, 2H), 1.73 (s, 3H), 1.64 – 1.54 (m, 3H), 1.41 (qt, $J = 13.2$, 3.6 Hz, 2H), 1.13 (qd, $J = 12.4$, 3.6 Hz, 2H), 0.93 (qt, $J = 13.2$, 3.6 Hz, 1H).

^{13}C NMR (100 MHz, CDCl_3) δ 172.0, 141.7, 139.7, 130.6, 128.7, 127.8, 117.6, 54.8, 31.7, 25.9, 25.5, 20.9.

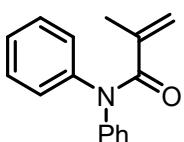
ESIHRMS: m/z Calcd. For $\text{C}_{16}\text{H}_{21}\text{NO}$: ($\text{M}+\text{H}$)⁺ 244.1696. Found: 244.1696.



N-benzyl-N-phenylmethacrylamide (1q): Prepared according to general procedure A using aniline and benzyl bromide. The spectra matched with reported literature.^[1]

^1H NMR (400 MHz, CDCl_3) δ 7.22 – 7.08 (m, 8H), 6.93 (d, $J = 7.6$ Hz, 2H), 4.96 (d, $J = 6.8$ Hz, 2H), 4.92 (s, 2H), 1.73 (s, 3H).

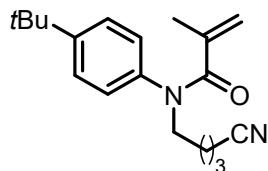
^{13}C NMR (100 MHz, CDCl_3) δ 171.4, 142.8, 140.4, 137.2, 128.8, 128.1(3), 128.0(9), 127.1, 127.0, 126.8, 119.2, 52.8, 20.1.



N,N-diphenylmethacrylamide (1r): Prepared according to general procedure C using diphenylamine. The spectra matched with reported literature.^[1]

¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.31 (m, 4H), 7.25 – 7.20 (m, 2H), 7.19 – 7.15 (m, 4H), 5.25 – 5.23 (p, *J* = 1.2 Hz, 1H), 5.17 (p, *J* = 1.2 Hz, 1H), 1.84 (t, *J* = 1.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 172.0, 143.6, 141.3, 129.2, 127.3, 126.6, 121.1, 20.1.

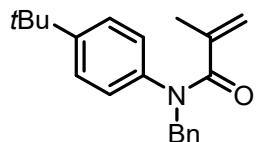


N-(4-(*tert*-butyl)phenyl)-N-(4-cyanobutyl)methacrylamide (1s): Prepared according to general procedure A using 4-(*tert*-butyl)aniline and 5-bromopentanenitrile.

¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.32 (m, 2H), 7.03 – 6.98 (m, 2H), 5.03 – 4.96 (m, 2H), 3.82 – 3.76 (m, 2H), 2.44 – 2.36 (m, 2H), 1.71 – 1.70 (m, 4H), 1.69 (s, 3H), 1.32 (s, 9H).

¹³C NMR (100 MHz, CDCl₃) δ 172.1, 150.5, 140.9, 139.9, 126.9, 126.3, 119.6, 119.4, 48.3, 34.7, 31.4, 26.8, 22.8, 20.4, 16.9.

ESIHRMS: *m/z* Calcd. For C₁₉H₂₆N₂O: (M+Na)⁺ 321.1937. Found: 321.1930.

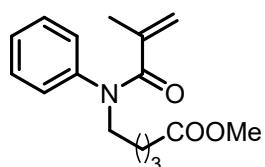


N-benzyl-N-(4-(*tert*-butyl)phenyl)methacrylamide (1t): Prepared according to general procedure A using 4-(*tert*-butyl)aniline and benzyl bromide.

¹H NMR (400 MHz, CDCl₃) δ 7.31 – 7.27 (m, 1H), 7.27 – 7.21 (m, 6H), 6.92 – 6.87 (m, 2H), 5.04 (dq, *J* = 2.8, 1.2 Hz, 2H), 4.95 (s, 2H), 1.78 – 1.73 (m, 3H), 1.28 (s, 9H).

¹³C NMR (100 MHz, CDCl₃) δ 172.0, 150.1, 140.8, 140.6, 137.8, 128.5, 128.3, 127.3, 126.8, 126.0, 119.5, 53.3, 34.6, 31.4, 20.5.

ESIHRMS: *m/z* Calcd. For C₂₁H₂₅NO: (M+H)⁺ 308.2009. Found: 308.2014.



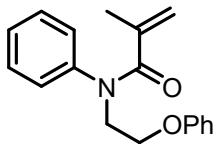
methyl 5-(N-phenylmethacrylamido)pentanoate (1u): Prepared according to general procedure A using aniline and 5-bromopentanoic acid.

¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.32 (m, 2H), 7.28 (d, *J* = 6.4 Hz, 1H), 7.11 (d, *J* = 7.6 Hz, 2H), 4.98 (d, *J* = 18.4 Hz, 2H), 3.79 (t, *J* = 6.8 Hz, 2H), 3.64 (s, 3H), 2.32 (t, *J* = 6.8 Hz, 2H), 1.74 (s, 3H), 1.63 (dq, *J* = 13.6, 7.6 Hz, 4H).

¹³C NMR (100 MHz, CDCl₃) δ 173.9, 171.9, 143.1, 141.1, 129.3, 127.6, 127.2, 119.2,

51.6, 49.2, 33.8, 27.2, 22.3, 20.5.

ESIHRMS: m/z Calcd. For $C_{16}H_{21}NO_3$: ($M+Na$)⁺ 298.1414. Found: 298.1407.

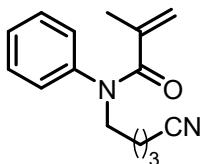


N-(2-phenoxyethyl)-N-phenylmethacrylamide (1v): Prepared according to general procedure A using aniline and (2-bromoethoxy)benzene.

1H NMR (400 MHz, CDCl₃) δ 7.34 (ddd, $J = 7.6, 6.4, 1.2$ Hz, 2H), 7.28 – 7.24 (m, 3H), 7.23 – 7.20 (m, 2H), 6.94 (tt, $J = 7.6, 1.2$ Hz, 1H), 6.87 – 6.82 (m, 2H), 5.06 – 5.03 (m, 1H), 5.01 (s, 1H), 4.22 (t, $J = 5.2$ Hz, 2H), 4.15 (td, $J = 5.2, 1.2$ Hz, 2H), 1.75 (s, 3H).

^{13}C NMR (100 MHz, CDCl₃) δ 172.2, 158.6, 143.9, 140.7, 129.5, 129.2, 127.5, 127.2, 121.0, 119.9, 114.5, 65.1, 49.8, 20.3.

ESIHRMS: m/z Calcd. For $C_{18}H_{19}NO_2$: ($M+H$)⁺ 282.1489. Found: 282.1482.



N-(4-cyanobutyl)-N-phenylmethacrylamide (1w): Prepared according to general procedure A using aniline and 5-bromopentanenitrile.

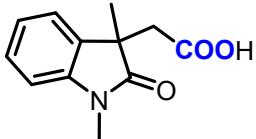
1H NMR (400 MHz, CDCl₃) δ 7.37 (ddd, $J = 7.6, 6.4, 1.2$ Hz, 2H), 7.32 – 7.27 (m, 1H), 7.13 – 7.08 (m, 2H), 5.03 – 5.00 (m, 1H), 4.99 – 4.93 (m, 1H), 3.87 – 3.77 (m, 2H), 2.45 – 2.36 (m, 2H), 1.73 (t, $J = 2.4$ Hz, 3H), 1.70 (p, $J = 3.6$ Hz, 4H).

^{13}C NMR (100 MHz, CDCl₃) δ 171.8, 142.6, 140.6, 129.3, 127.3, 127.2, 119.4, 119.3, 48.0, 26.6, 22.6, 20.2, 16.7.

ESIHRMS: m/z Calcd. For $C_{15}H_{18}N_2O$: ($M+Na$)⁺ 265.1311. Found: 265.1311.

4. Succinic Acid Derivatives

All reactions were conducted according to the general procedure with 0.2 mmol alkenes unless otherwise stated.

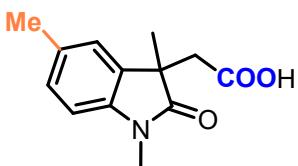


2-(1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2a): Prepared according to general procedure using *N*-methyl-*N*-phenylmethacrylamide (0.2 mmol, 35.0 mg, 1.0 equiv). After 12 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (27.2 mg, 62% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.29 (d, *J* = 7.6 Hz, 1H), 7.19 (d, *J* = 7.2 Hz, 1H), 7.06 (t, *J* = 7.6 Hz, 1H), 6.85 (d, *J* = 8.0 Hz, 1H), 3.21 (s, 3H), 2.98 (d, *J* = 16.8 Hz, 1H), 2.80 (d, *J* = 16.8 Hz, 1H), 1.38 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 180.5, 174.1, 143.3, 132.8, 128.5, 123.0, 122.5, 108.6, 45.4, 41.5, 26.6, 24.1.

ESIHRMS: *m/z* Calcd. For C₁₂H₁₃NO₃: (M+Na)⁺ 242.0788. Found: 242.0780.

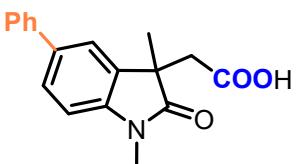


2-(1,3,5-trimethyl-2-oxoindolin-3-yl)acetic acid (2b): Prepared according to general procedure using *N*-methyl-*N*-(*p*-tolyl)methacrylamide (0.2 mmol, 37.9 mg, 1.0 equiv). After 12 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (28.0 mg, 60% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.08 (d, *J* = 8.0 Hz, 1H), 7.01 (s, 1H), 6.74 (d, *J* = 8.0 Hz, 1H), 3.20 (s, 3H), 2.97 (d, *J* = 16.4 Hz, 1H), 2.78 (d, *J* = 16.4 Hz, 1H), 2.33 (s, 3H), 1.37 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 180.6, 173.8, 140.8, 132.9, 132.6, 128.7, 123.3, 108.3, 45.5, 41.5, 26.7, 24.1, 21.3.

ESIHRMS: *m/z* Calcd. For C₁₃H₁₅NO₃: (M+Na)⁺ 256.0944. Found: 256.0934.



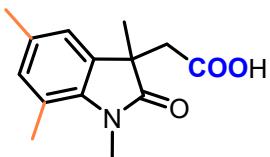
2-(1,3-dimethyl-2-oxo-5-phenylindolin-3-yl)acetic acid (2c): Prepared according to

general procedure using *N*-([1,1'-biphenyl]-4-yl)-*N*-methylmethacrylamide (0.2 mmol, 50.3 mg, 1.0 equiv). After 12 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (40.8 mg, 69% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.55 – 7.50 (m, 2H), 7.48 (dd, *J* = 8.0, 2.0 Hz, 1H), 7.45 – 7.39 (m, 3H), 7.35 – 7.29 (m, 1H), 6.89 (d, *J* = 8.0 Hz, 1H), 3.22 (s, 3H), 3.02 (d, *J* = 16.8 Hz, 1H), 2.85 (d, *J* = 16.8 Hz, 1H), 1.40 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 180.5, 174.1, 142.7, 140.9, 136.3, 133.5, 128.9, 127.3, 127.2, 127.0, 121.4, 108.8, 45.6, 41.4, 26.7, 24.3.

ESIHRMS: *m/z* Calcd. For C₁₈H₁₇NO₃: (M+H)⁺ 296.1281. Found: 296.1268.

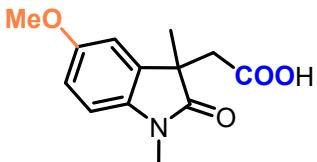


2-(1,3,5,7-tetramethyl-2-oxoindolin-3-yl)acetic acid (2d): Prepared according to general procedure using *N*-(2,4-dimethylphenyl)-*N*-methylmethacrylamide (0.2 mmol, 40.7 mg, 1.0 equiv). After 12 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as yellow solid (24.2 mg, 49% yield).

¹H NMR (400 MHz, DMSO-d₆) δ 6.93 (s, 1H), 6.74 (s, 1H), 3.31 (s, 3H), 2.80 (d, *J* = 16.4 Hz, 1H), 2.71 (d, *J* = 16.4 Hz, 1H), 2.45 (s, 3H), 2.17 (s, 3H), 1.13 (s, 3H).

¹³C NMR (100 MHz, DMSO-d₆) δ 180.4, 171.4, 139.3, 134.6, 132.1, 131.0, 121.2, 119.4, 44.8, 41.8, 29.6, 25.4, 20.9, 18.9.

ESIHRMS: *m/z* Calcd. For C₁₄H₁₇NO₃: (M+Na)⁺ 270.1101. Found: 270.1099.

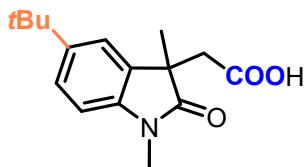


2-(5-methoxy-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2e): Prepared according to general procedure using *N*-(4-methoxyphenyl)-*N*-methylmethacrylamide (0.2 mmol, 41.1 mg, 1.0 equiv). After 12 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as brown solid (20.9 mg, 42% yield).

¹H NMR (400 MHz, CDCl₃) δ 6.83 – 6.78 (m, 2H), 6.76 (dd, *J* = 8.4, 0.8 Hz, 1H), 3.79 (s, 3H), 3.20 (s, 3H), 2.97 (d, *J* = 16.4 Hz, 1H), 2.78 (d, *J* = 16.4 Hz, 1H), 1.38 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 180.3, 173.7, 156.3, 136.7, 134.3, 112.4, 110.2, 108.9, 55.9, 45.8, 41.4, 26.7, 24.1.

ESIHRMS: *m/z* Calcd. For C₁₃H₁₅NO₄: (M+Na)⁺ 272.0893. Found: 272.0900.

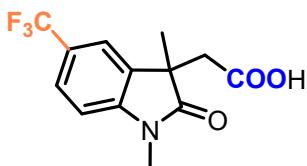


2-(5-(*tert*-butyl)-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2f): Prepared according to general procedure using *N*-(4-(*tert*-butyl)phenyl)-*N*-methylmethacrylamide (0.2 mmol, 46.3 mg, 1.0 equiv). After 19 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (43.5 mg, 79% yield).

¹H NMR (400 MHz, CDCl₃) δ 8.85 (s, 1H), 7.30 (dd, *J* = 8.0, 2.0 Hz, 1H), 7.25 (d, *J* = 2.0 Hz, 1H), 6.78 (d, *J* = 8.0 Hz, 1H), 3.20 (s, 3H), 2.98 (d, *J* = 16.4 Hz, 1H), 2.83 (d, *J* = 16.4 Hz, 1H), 2.05 (s, 2H), 1.39 (s, 3H), 1.30 (s, 9H).

¹³C NMR (100 MHz, CDCl₃) δ 180.8, 173.9, 146.3, 140.7, 132.5, 125.0, 119.7, 108.0, 45.7, 41.6, 34.7, 31.7, 26.6, 24.0.

ESIHRMS: *m/z* Calcd. For C₁₆H₂₁NO₃: (M+Na)⁺ 298.1414. Found: 298.1414.



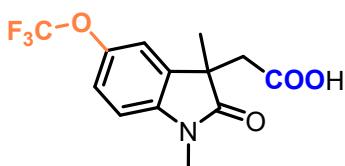
2-(1,3-dimethyl-2-oxo-5-(trifluoromethyl)indolin-3-yl)acetic acid (2g): Prepared according to general procedure using *N*-methyl-*N*-(4-(trifluoromethyl)phenyl)methacrylamide (0.2 mmol, 48.6 mg, 1.0 equiv). After 12 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (23.0 mg, 40% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.56 (dd, *J* = 8.4, 0.8 Hz, 1H), 7.43 – 7.39 (d, *J* = 1.2 Hz, 1H), 6.91 (d, *J* = 8.0 Hz, 1H), 3.24 (s, 3H), 3.04 (d, *J* = 17.2 Hz, 1H), 2.87 (d, *J* = 17.2 Hz, 1H), 1.38 (s, 3H).

¹³C NMR (100 MHz, CD₃OD) δ 182.6, 173.2, 148.3, 135.8, 126.1 (q, *J* = 269.1 Hz), 126.9 (q, *J* = 3.7 Hz), 125.7 (q, *J* = 32.2 Hz), 120.3 (q, *J* = 3.3 Hz), 109.6, 46.9, 41.9, 26.9, 24.5.

¹⁹F NMR (376 MHz, CDCl₃) δ -61.2.

ESIHRMS: *m/z* Calcd. For C₁₃H₁₂F₃NO₃: (M+Na)⁺ 310.0661. Found: 310.0658.



2-(1,3-dimethyl-2-oxo-5-(trifluoromethoxy)indolin-3-yl)acetic acid (2h): Prepared according to general procedure using *N*-methyl-*N*-(4-

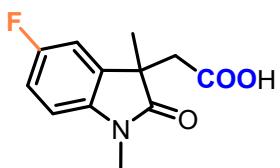
(trifluoromethoxy)phenyl)methacrylamide (0.2 mmol, 51.8 mg, 1.0 equiv). After 12 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (32.1 mg, 53% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.15 (ddd, *J* = 8.4, 2.4, 0.8 Hz, 1H), 7.10 (d, *J* = 1.2 Hz, 1H), 6.82 (d, *J* = 8.4 Hz, 1H), 3.21 (s, 3H), 3.00 (d, *J* = 16.8 Hz, 1H), 2.82 (d, *J* = 16.8 Hz, 1H), 1.37 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 180.0, 174.2, 144.9, 142.1, 134.4, 120.6 (q, *J* = 255.2 Hz), 121.4, 116.6, 108.8, 45.8, 41.1, 26.7, 24.0.

¹⁹F NMR (376 MHz, CDCl₃) δ -58.2.

ESIHRMS: *m/z* Calcd. For C₁₃H₁₂F₃NO₄: (M+Na)⁺ 326.0611. Found: 326.0605.



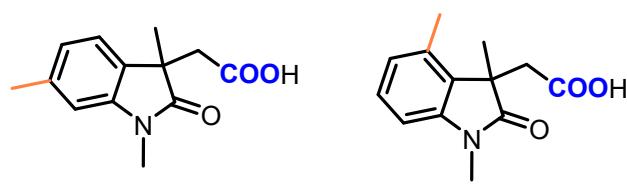
2-(5-fluoro-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2i): Prepared according to general procedure using *N*-(4-fluorophenyl)-*N*-methylmethacrylamide (0.2 mmol, 51.8 mg, 1.0 equiv). After 12 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as deep brown oil (28.0 mg, 59% yield).

¹H NMR (400 MHz, CDCl₃) δ 6.98 (t, *J* = 9.6 Hz, 2H), 6.79 – 6.76 (m, 1H), 3.21 (s, 3H), 2.99 (d, *J* = 16.8 Hz, 1H), 2.80 (d, *J* = 16.8 Hz, 1H), 1.37 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 180.1, 173.8, 159.5 (d, *J* = 239.8 Hz), 139.3, 134.5 (d, *J* = 7.8 Hz), 114.6 (d, *J* = 23.4 Hz), 110.8 (d, *J* = 24.8 Hz), 109.0 (d, *J* = 8.1 Hz), 45.9, 41.2, 26.8, 24.1.

¹⁹F NMR (376 MHz, CDCl₃) δ -120.0.

ESIHRMS: *m/z* Calcd. For C₁₂H₁₂FNO₃: (M+Na)⁺ 260.0693. Found: 260.0693.



0.6:1

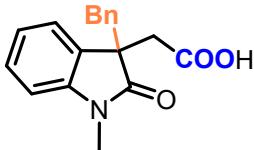
2-(1,3,6-trimethyl-2-oxoindolin-3-yl)acetic acid (2j) and 2-(1,3,4-trimethyl-2-oxoindolin-3-yl)acetic acid (2j') (0.6:1): Prepared according to general procedure using *N*-methyl-*N*-(*m*-tolyl)methacrylamide (0.2 mmol, 37.9 mg, 1.0 equiv). After 12 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (28.0 mg, 60% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.20 – 7.04 (m, 1H), 6.84 (m, 1H), 6.70 – 6.66 (m, 1H), 3.19 (d, *J* = 3.2 Hz, 3H), 3.14 – 2.73 (m, 2H), 2.37 (d, *J* = 10.4 Hz, 3H), 1.38 (d, *J* =

22.4 Hz, 3H).

^{13}C NMR (100 MHz, CDCl_3) δ 180.8, 180.3, 174.4, 174.2, 143.7, 143.3, 138.6, 133.9, 129.9, 129.4, 128.2, 125.4, 123.5, 122.2, 109.6, 106.3, 46.4, 45.2, 41.6, 40.4, 26.7, 26.6, 24.1, 22.2, 21.9, 18.2.

ESIHRMS: m/z Calcd. For $\text{C}_{13}\text{H}_{15}\text{NO}_3$: ($\text{M}+\text{Na}$) $^+$ 256.0944. Found: 256.0939.

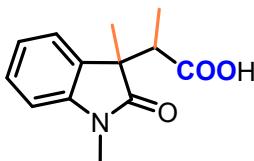


2-(3-benzyl-1-methyl-2-oxoindolin-3-yl)acetic acid (2k): Prepared according to general procedure using 2-benzyl-*N*-methyl-*N*-phenylacrylamide (0.2 mmol, 50.3 mg, 1.0 equiv). After 18 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (35.4 mg, 60% yield).

^1H NMR (400 MHz, CDCl_3) δ 7.19 (td, $J = 7.6, 1.6$ Hz, 1H), 7.11 – 7.05 (m, 2H), 7.05 – 6.99 (m, 3H), 6.77 – 6.72 (m, 2H), 6.57 (d, $J = 7.6$ Hz, 1H), 3.13 (d, $J = 16.8$ Hz, 1H), 3.07 – 3.02 (m, 2H), 2.95 (s, 3H), 2.89 (d, $J = 16.8$ Hz, 1H).

^{13}C NMR (100 MHz, CDCl_3) δ 179.1, 173.7, 143.8, 143.4, 134.6, 130.0, 128.6, 127.7, 126.9, 123.3, 122.5, 108.2, 51.2, 43.8, 40.3, 26.2.

ESIHRMS: m/z Calcd. For $\text{C}_{18}\text{H}_{17}\text{NO}_3$: ($\text{M}+\text{Na}$) $^+$ 318.1101. Found: 318.1097.



2-(1,3-dimethyl-2-oxoindolin-3-yl)propanoic acid (2l): Prepared according to general procedure using (*E*)-*N*,*N*-dimethyl-*N*-phenylbut-2-enamide (0.2 mmol, 37.9 mg, 1.0 equiv). After 48 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (15.5 + 9.4 = 24.9 mg, 53% yield).

^1H NMR (400 MHz, CDCl_3) δ 7.44 – 7.40 (m, 1H), 7.30 (td, $J = 7.6, 1.2$ Hz, 1H), 7.08 (td, $J = 7.6, 1.2$ Hz, 1H), 6.86 (d, $J = 7.6$ Hz, 1H), 3.23 (s, 3H), 3.10 (q, $J = 7.2$ Hz, 1H), 1.47 (s, 3H), 1.01 (d, $J = 7.2$ Hz, 3H).

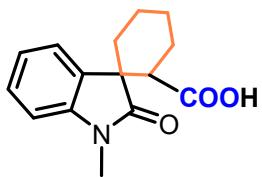
^{13}C NMR (100 MHz, CDCl_3) δ 179.6, 179.5, 143.5, 131.4, 128.4, 124.7, 122.8, 108.2, 49.5, 45.8, 26.5, 22.3, 13.1.

ESIHRMS: m/z Calcd. For $\text{C}_{13}\text{H}_{15}\text{NO}_3$: ($\text{M}+\text{Na}$) $^+$ 256.0944. Found: 256.0945.

^1H NMR (400 MHz, CDCl_3) δ 7.36 (ddd, $J = 8.0, 6.8, 2.0$ Hz, 1H), 7.23 – 7.16 (m, 2H), 6.95 (d, $J = 8.0$ Hz, 1H), 3.32 (s, 3H), 3.12 (q, $J = 7.2$ Hz, 1H), 1.55 (s, 3H), 0.97 (d, $J = 7.2$ Hz, 3H).

^{13}C NMR (100 MHz, CDCl_3) δ 182.0, 174.7, 142.6, 132.4, 128.9, 124.4, 122.4, 109.2, 49.8, 48.6, 26.8, 22.8, 13.6.

ESIHRMS: m/z Calcd. For $\text{C}_{13}\text{H}_{15}\text{NO}_3$: ($\text{M}+\text{Na}$) $^+$ 234.1125. Found: 234.1126.



1'-methyl-2'-oxospiro[cyclohexane-1,3'-indoline]-2-carboxylic acid (2m): Prepared according to general procedure using *N*-methyl-*N*-phenylcyclohex-1-ene-1-carboxamide (0.2 mmol, 43.1 mg, 1.0 equiv). After 48 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as pale yellow solid (19.4 + 12.0 = 31.4 mg, 60% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.42 (dd, *J* = 7.6, 0.8 Hz, 1H), 7.27 (td, *J* = 7.6, 1.2 Hz, 1H), 6.98 (td, *J* = 7.6, 1.2 Hz, 1H), 6.82 (dd, *J* = 7.6, 0.8 Hz, 1H), 3.15 (s, 3H), 3.00 (dd, *J* = 13.2, 4.4 Hz, 1H), 2.15 – 2.10 (m, 1H), 2.03 – 1.89 (m, 2H), 1.88 – 1.78 (m, 1H), 1.74 – 1.59 (m, 2H), 1.53 – 1.40 (m, 2H).

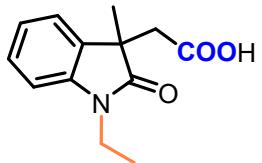
¹³C NMR (100 MHz, CDCl₃) δ 180.3, 177.2, 143.7, 131.7, 128.0, 124.7, 121.9, 108.4, 49.7, 46.3, 35.0, 26.6, 24.6, 24.0, 20.7.

ESIHRMS: *m/z* Calcd. For C₁₅H₁₇NO₃: (M+Na)⁺ 282.1101. Found: 282.1098.

¹H NMR (400 MHz, CDCl₃) δ 7.22 (td, *J* = 7.6, 1.4 Hz, 1H), 7.07 – 6.97 (m, 2H), 6.76 (d, *J* = 7.6 Hz, 1H), 3.13 (s, 3H), 2.84 (dd, *J* = 12.8, 3.6 Hz, 1H), 2.48 (qd, *J* = 13.6, 13.2, 4.2 Hz, 1H), 2.23 – 2.12 (m, 1H), 2.02 – 1.91 (m, 2H), 1.70 – 1.60 (m, 2H), 1.55 (dt, *J* = 13.2, 3.2 Hz, 1H), 1.43 (qt, *J* = 14.4, 4.2 Hz, 1H).

¹³C NMR (100 MHz, CDCl₃) δ 178.9, 178.3, 143.5, 133.8, 128.0, 122.3, 121.4, 108.0, 50.0, 46.9, 35.7, 26.2, 25.6, 24.9, 19.9.

ESIHRMS: *m/z* Calcd. For C₁₅H₁₇NO₃: (M+H)⁺ 260.1281. Found: 260.1288.

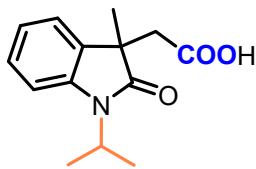


2-(1-ethyl-3-methyl-2-oxoindolin-3-yl)acetic acid (2n): Prepared according to general procedure using *N*-ethyl-*N*-phenylmethacrylamide (0.2 mmol, 37.9 mg, 1.0 equiv). After 18 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (23.3 mg, 50% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.28 – 7.24 (m, 1H), 7.20 – 7.16 (m, 1H), 7.07 – 7.01 (m, 1H), 6.87 – 6.83 (m, 1H), 3.84 (dq, *J* = 14.4, 7.2 Hz, 1H), 3.67 (dq, *J* = 14.4, 7.2 Hz, 1H), 2.99 (d, *J* = 16.4 Hz, 1H), 2.79 (d, *J* = 16.4 Hz, 1H), 1.35 (s, 3H), 1.21 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 180.0, 174.4, 142.4, 133.1, 128.3, 122.6, 122.5, 108.7, 45.3, 41.4, 34.9, 24.2, 12.2.

ESIHRMS: *m/z* Calcd. For C₁₃H₁₅NO₃: (M+Na)⁺ 256.0944. Found: 256.0934.

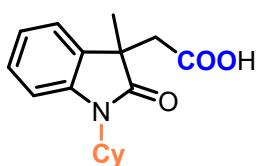


2-(1-isopropyl-3-methyl-2-oxoindolin-3-yl)acetic acid (2o): Prepared according to general procedure using *N*-isopropyl-*N*-phenylmethacrylamide (0.2 mmol, 40.7 mg, 1.0 equiv). After 12 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (27.7 mg, 56% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.23 (t, *J* = 8.0 Hz, 1H), 7.16 (d, *J* = 7.2 Hz, 1H), 7.05 – 6.98 (m, 2H), 4.61 (p, *J* = 6.8 Hz, 1H), 2.99 (d, *J* = 16.4 Hz, 1H), 2.77 (d, *J* = 16.4 Hz, 1H), 1.44 (dd, *J* = 10.8, 7.2 Hz, 6H), 1.33 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 180.1, 174.5, 142.0, 133.4, 128.1, 122.5, 122.3, 110.3, 45.1, 44.0, 41.6, 24.4, 19.4, 19.0.

ESIHRMS: *m/z* Calcd. For C₁₄H₁₇NO₃: (M+Na)⁺ 270.1101. Found: 270.1096.

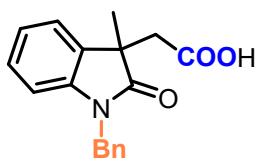


2-(1-cyclohexyl-3-methyl-2-oxoindolin-3-yl)acetic acid (2p): Prepared according to general procedure using *N*-cyclohexyl-*N*-phenylmethacrylamide (0.2 mmol, 48.7 mg, 1.0 equiv). After 24 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as brown solid (35.1 mg, 61% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.22 (td, *J* = 7.6, 1.2 Hz, 1H), 7.17 – 7.13 (m, 1H), 7.07 – 6.98 (m, 2H), 4.14 (t, *J* = 12.4 Hz, 1H), 2.98 (d, *J* = 16.8 Hz, 1H), 2.76 (d, *J* = 16.8 Hz, 1H), 2.18 – 2.01 (m, 2H), 1.88 (d, *J* = 13.2 Hz, 2H), 1.72 (d, *J* = 10.4 Hz, 2H), 1.45 – 1.37 (m, 2H), 1.33 (s, 3H), 1.30 – 1.20 (m, 2H).

¹³C NMR (100 MHz, CDCl₃) δ 180.3, 174.6, 142.4, 133.3, 128.0, 122.5, 122.2, 110.4, 52.4, 45.0, 41.6, 29.1, 28.7, 26.3, 26.1, 25.5, 24.5.

ESIHRMS: *m/z* Calcd. For C₁₇H₁₅NO₃: (M+Na)⁺ 310.1414. Found: 310.1409.

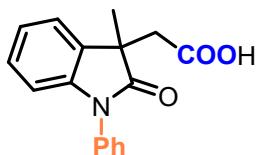


2-(1-benzyl-3-methyl-2-oxoindolin-3-yl)acetic acid (2q): Prepared according to general procedure using *N*-benzyl-*N*-phenylmethacrylamide (0.2 mmol, 50.3 mg, 1.0 equiv). After 18 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (25.4 mg, 43% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.32 – 7.27 (m, 4H), 7.26 – 7.23 (m, 1H), 7.21 – 7.18 (m, 1H), 7.14 (td, *J* = 7.6, 1.2 Hz, 1H), 7.01 (td, *J* = 7.6, 1.2 Hz, 1H), 6.69 (d, *J* = 7.6 Hz, 1H), 5.00 (d, *J* = 15.6 Hz, 1H), 4.86 (d, *J* = 15.6 Hz, 1H), 3.11 (d, *J* = 16.8 Hz, 1H), 2.90 (d, *J* = 16.8 Hz, 1H), 1.44 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 180.4, 174.6, 142.4, 135.8, 132.8, 128.9, 128.3, 127.7, 127.3, 122.9, 122.3, 109.7, 45.5, 44.1, 41.2, 25.0.

ESIHRMS: *m/z* Calcd. For C₁₈H₁₇NO₃: (M+Na)⁺ 318.1101. Found: 318.1102.

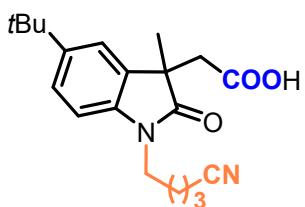


2-(3-methyl-2-oxo-1-phenylindolin-3-yl)acetic acid (2r): Prepared according to general procedure using *N,N*-diphenylmethacrylamide (0.2 mmol, 47.5 mg, 1.0 equiv). After 48 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (29.3 mg, 52% yield).

¹H NMR (400 MHz, CD₃OD) δ 7.56 (t, *J* = 8.0 Hz, 2H), 7.46 (t, *J* = 8.0 Hz, 3H), 7.36 (d, *J* = 7.6 Hz, 1H), 7.20 (t, *J* = 7.6 Hz, 1H), 7.09 (t, *J* = 7.6 Hz, 1H), 6.71 (d, *J* = 7.6 Hz, 1H), 3.06 (d, *J* = 3.2 Hz, 2H), 1.44 (s, 3H).

¹³C NMR (100 MHz, CD₃OD) δ 182.4, 173.3, 145.2, 136.3, 134.5, 130.7, 129.4, 129.1, 128.2, 124.2, 123.6, 110.5, 47.0, 42.6, 25.1.

ESIHRMS: *m/z* Calcd. For C₁₇H₁₅NO₃: (M+Na)⁺ 304.0944. Found: 304.0940.

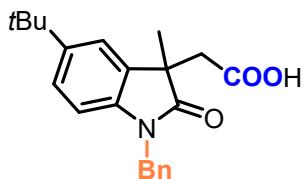


2-(5-(tert-butyl)-1-(4-cyanobutyl)-3-methyl-2-oxoindolin-3-yl)acetic acid (2s): Prepared according to general procedure using *N*-(4-(*tert*-butyl)phenyl)-*N*-(4-cyanobutyl)methacrylamide (0.2 mmol, 59.7 mg, 1.0 equiv). After 48 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (47.9 mg, 70% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.28 (d, *J* = 7.6 Hz, 1H), 7.21 (s, 1H), 6.77 (d, *J* = 8.0 Hz, 1H), 3.83 (dt, *J* = 13.6, 6.8 Hz, 1H), 3.65 (dt, *J* = 13.6, 6.4 Hz, 1H), 3.00 (d, *J* = 16.8 Hz, 1H), 2.84 (d, *J* = 16.8 Hz, 1H), 2.38 (q, *J* = 7.2 Hz, 2H), 1.84 (p, *J* = 6.8 Hz, 2H), 1.69 (p, *J* = 6.8 Hz, 2H), 1.34 (s, 3H), 1.30 (s, 9H).

¹³C NMR (100 MHz, CDCl₃) δ 180.6, 174.6, 146.2, 139.8, 132.6, 125.0, 119.7, 119.6, 107.9, 45.5, 41.2, 38.9, 34.7, 31.7, 26.4, 24.8, 22.5, 16.7.

ESIHRMS: *m/z* Calcd. For C₂₀H₂₆N₂O₃: (M+Na)⁺ 365.1836. Found: 365.1826.

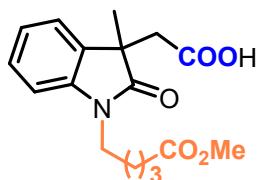


2-(1-benzyl-5-(*tert*-butyl)-3-methyl-2-oxoindolin-3-yl)acetic acid (2t): Prepared according to general procedure using *N*-benzyl-*N*-(4-(*tert*-butyl)phenyl)methacrylamide (0.2 mmol, 61.5 mg, 1.0 equiv). After 12 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (44.8 mg, 64% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.31 (d, *J* = 4.7 Hz, 4H), 7.27 – 7.25 (m, 1H), 7.23 (d, *J* = 2.0 Hz, 1H), 7.18 (dd, *J* = 8.2, 2.0 Hz, 1H), 6.66 (d, *J* = 8.0 Hz, 1H), 4.92 (q, *J* = 15.6 Hz, 2H), 3.07 (d, *J* = 16.4 Hz, 1H), 2.87 (d, *J* = 16.4 Hz, 1H), 1.49 (s, 3H), 1.26 (s, 9H).

¹³C NMR (100 MHz, CDCl₃) δ 181.1, 173.2, 146.5, 139.6, 135.8, 132.5, 128.9, 127.8, 127.5, 125.1, 119.6, 109.2, 45.6, 44.2, 41.6, 34.7, 31.7, 24.5.

ESIHRMS: *m/z* Calcd. For C₂₂H₂₅NO₃: (M+Na)⁺ 374.1727. Found: 374.1736.

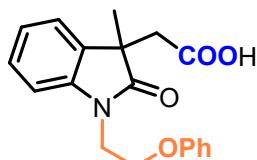


2-(1-(5-methoxy-5-oxopentyl)-3-methyl-2-oxoindolin-3-yl)acetic acid (2u): Prepared according to general procedure using methyl 5-(*N*-phenylmethacrylamido)pentanoate (0.2 mmol, 55.1 mg, 1.0 equiv). After 48 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (29.4 mg, 46% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.26 (td, *J* = 7.6, 0.8 Hz, 1H), 7.18 (d, *J* = 7.2 Hz, 1H), 7.04 (t, *J* = 7.6 Hz, 1H), 6.85 (d, *J* = 7.6 Hz, 1H), 3.83 – 3.75 (m, 1H), 3.70 – 3.67 (m, 1H), 3.65 (s, 3H), 2.99 (d, *J* = 16.4 Hz, 1H), 2.80 (d, *J* = 16.4 Hz, 1H), 2.34 (t, *J* = 6.8 Hz, 2H), 1.68 (t, *J* = 3.2 Hz, 4H), 1.36 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 180.3, 174.1, 173.9, 142.6, 133.0, 128.4, 122.7, 122.6, 108.7, 51.8, 45.4, 41.4, 39.7, 33.6, 26.7, 24.4, 22.2.

ESIHRMS: *m/z* Calcd. For C₁₇H₂₁NO₅: (M+Na)⁺ 342.1312. Found: 342.1303.



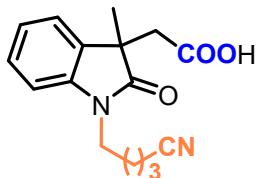
2-(3-methyl-2-oxo-1-(2-phenoxybutyl)indolin-3-yl)acetic acid (2v): Prepared according to general procedure using methyl *N*-(2-phenoxyethyl)-*N*-phenylmethacrylamide (0.2 mmol, 56.3 mg, 1.0 equiv). After 12 hours, the product was

purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as orange oil (31.8 mg, 49% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.27 – 7.22 (m, 3H), 7.15 (d, *J* = 7.2 Hz, 1H), 7.03 (dt, *J* = 7.2, 3.6 Hz, 2H), 6.93 (t, *J* = 7.2 Hz, 1H), 6.83 (d, *J* = 8.4 Hz, 2H), 4.19 – 4.05 (m, 4H), 2.98 (d, *J* = 16.8 Hz, 1H), 2.78 (d, *J* = 16.8 Hz, 1H), 1.34 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 180.7, 174.2, 158.4, 142.8, 132.7, 129.6, 128.4, 122.9, 122.4, 121.2, 114.6, 109.4, 64.9, 45.2, 41.5, 39.8, 24.2.

ESIHRMS: *m/z* Calcd. For C₁₉H₁₉NO₄: (M+Na)⁺ 348.1206. Found: 348.1192.



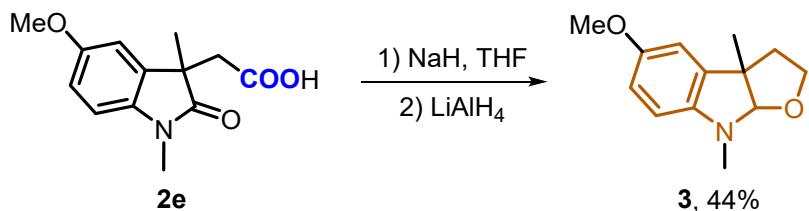
2-(1-(4-cyanobutyl)-3-methyl-2-oxoindolin-3-yl)acetic acid (2w): Prepared according to general procedure using methyl *N*-(4-cyanobutyl)-*N*-phenylmethacrylamide (0.2 mmol, 48.5 mg, 1.0 equiv). After 18 hours, the product was purified by flash column chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2 as the eluent) and obtained as white solid (35.5 mg, 62% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.27 (t, *J* = 7.6 Hz, 1H), 7.17 (d, *J* = 7.2 Hz, 1H), 7.05 (t, *J* = 7.6 Hz, 1H), 6.85 (d, *J* = 8.0 Hz, 1H), 3.86 (dt, *J* = 14.0, 7.2 Hz, 1H), 3.65 (dt, *J* = 14.0, 7.2 Hz, 1H), 3.02 (d, *J* = 16.8 Hz, 1H), 2.83 (d, *J* = 16.8 Hz, 1H), 2.44 – 2.30 (m, 2H), 1.83 (dt, *J* = 14.0, 6.8 Hz, 2H), 1.68 (dt, *J* = 14.0, 6.8 Hz, 2H), 1.33 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 180.3, 174.5, 142.4, 132.9, 128.4, 122.9, 122.5, 119.6, 108.6, 45.3, 41.1, 38.9, 26.3, 24.8, 22.5, 16.8.

ESIHRMS: *m/z* Calcd. For C₁₆H₁₈N₂O₃: (M+Na)⁺ 309.1210. Found: 309.1209.

5. The Application of the Reaction

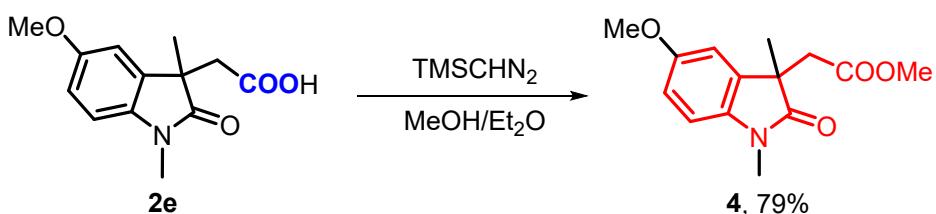


5-methoxy-3a,8-dimethyl-3,3a,8,8a-tetrahydro-2H-furo[2,3-*b*]indole (3): To a solution of **2e** (49.9 mg, 0.2 mmol) in THF (5 mL) was added NaH (20.0 mg, 0.5 mmol) at 0 °C under N₂ atmosphere. The reaction mixture was stirred at room temperature for 30 min, then LiAlH₄ (30.4 mg, 0.8 mmol) was added under N₂ atmosphere. The mixture was stirred at room temperature for 12 hours. After full conversion, the reaction was cooled to 0 °C and quenched with EtOAc. After filtration, the filtrate was concentrated in vacuo. The residue was purified by silica gel flash column chromatography (PE/EA = 10:1) to afford the product **3** as brown oil (19.1 mg, 44% yield).^[8]

¹H NMR (400 MHz, CDCl₃) δ 6.69 (d, *J* = 2.4 Hz, 1H), 6.66 (dd, *J* = 8.4, 2.4 Hz, 1H), 6.29 (d, *J* = 8.4 Hz, 1H), 5.03 (s, 1H), 3.94 (td, *J* = 8.0, 7.2, 1.6 Hz, 1H), 3.75 (s, 3H), 3.47 (ddd, *J* = 10.8, 8.8, 5.2 Hz, 1H), 2.88 (s, 3H), 2.13 (ddd, *J* = 12.0, 5.2, 1.6 Hz, 1H), 2.04 (td, *J* = 11.2, 7.2 Hz, 1H), 1.45 (s, 3H).

^{13}C NMR (100 MHz, CDCl_3) δ 152.8, 145.1, 136.1, 112.2, 110.6, 105.7, 105.4, 67.5, 56.2, 52.6, 41.6, 31.8, 24.6.

ESIHRMS: m/z Calcd. For C₁₃H₁₇NO₂: (M+H)⁺ 220.1332. Found: 220.1332.

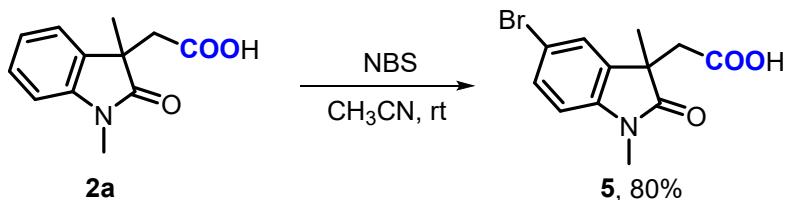


methyl 2-(5-methoxy-1,3-dimethyl-2-oxoindolin-3-yl)acetate (4): To a solution of **2e** (49.9 mg, 0.2 mmol,) in 10 mL MeOH/Et₂O (1:1, v:v) was added TMSCHN₂ solution in hexane (0.4 mL, 0.8 mmol, 2M hexane solution) at 0 °C. The reaction mixture was stirred for 4 hours at room temperature. The solvent was removed in vacuo and the residue was purified by flash column chromatography (PE/EA=10:1) to afford the ester product as a colorous oil (41.7 mg, 79% yield).^[8]

¹H NMR (400 MHz, CDCl₃) δ 6.83 (dd, *J* = 2.4, 0.8 Hz, 1H), 6.81 – 6.75 (m, 2H), 3.79 (s, 3H), 3.48 (s, 3H), 3.24 (s, 3H), 3.00 (d, *J* = 16.4 Hz, 1H), 2.83 (d, *J* = 16.4 Hz, 1H), 1.37 (s, 3H).

^{13}C NMR (100 MHz, CDCl_3) δ 179.6, 170.4, 155.9, 137.2, 134.4, 111.9, 110.2, 108.4, 55.8, 51.7, 45.9, 41.3, 26.6, 24.4.

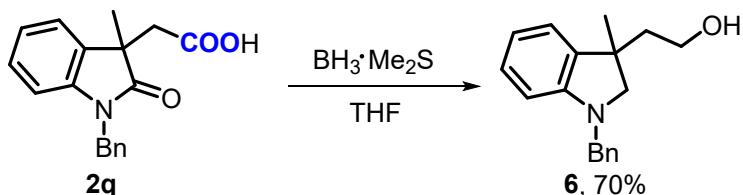
ESIHRMS: m/z Calcd. For $C_{14}H_{17}NO_4$: ($M+H$)⁺ 264.1230. Found: 264.1236.



2-(5-bromo-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (5): To a solution of **2a** (43.8 mg, 0.2 mmol) in 4 mL CH₃CN was added NBS (0.26 mmol, 46.3 mg). The reaction was stirred at room temperature for 22 hours. The solvent was removed in vacuo and the residue was purified by flash column chromatography using petroleum/ethyl acetate (5:1) to afford the **5** as a white solid (47.7 mg, 80% yield).^[8]

¹H NMR (400 MHz, CDCl₃) δ 7.40 (dd, *J* = 8.4, 2.0 Hz, 1H), 7.30 (d, *J* = 2.0 Hz, 1H), 6.72 (d, *J* = 8.4 Hz, 1H), 3.18 (s, 3H), 3.00 (d, *J* = 16.8 Hz, 1H), 2.80 (d, *J* = 16.8 Hz, 1H), 1.34 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 179.7, 174.3, 142.6, 135.0, 131.2, 125.7, 115.4, 109.9, 45.6, 41.2, 26.7, 24.2.



2-(1-benzyl-3-methylindolin-3-yl)ethan-1-ol (6): To a solution of **2q** (59.1 mg, 0.2 mmol) in 2 mL anhydrous THF was added $\text{BH}_3\text{C}_2\text{Me}_2\text{S}$ solution in THF (1 mL, 2 mmol, 2M THF solution) under nitrogen atmosphere at 0 °C. The reaction was stirred at room temperature for 24 hours and quenched with 3 mL MeOH. The solvent was removed in vacuo and the residue was purified by flash column chromatography using petroleum/EtOAc (5:1, v:v) to afford indoline **6** as a colorous oil (39.4 mg, 70% yield).^[8]

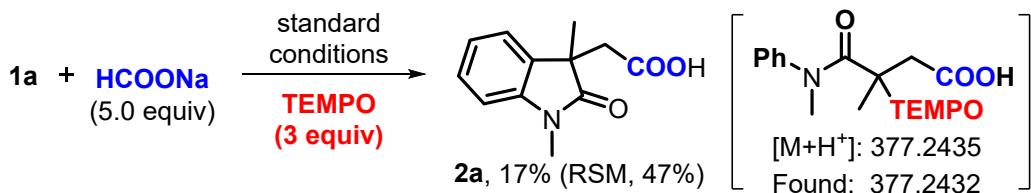
¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.31 (m, 4H), 7.28 (ddd, *J* = 8.4, 3.2, 2.0 Hz, 1H), 7.11 (td, *J* = 7.4, 1.2 Hz, 1H), 7.01 (dd, *J* = 7.4, 1.2 Hz, 1H), 6.78 (td, *J* = 7.4, 1.2 Hz, 1H), 6.63 (d, *J* = 8.0 Hz, 1H), 4.43 (d, *J* = 14.4 Hz, 1H), 4.00 (d, *J* = 14.4 Hz, 1H), 3.53 (dt, *J* = 11.2, 5.6 Hz, 1H), 3.37 (ddd, *J* = 11.2, 8.4, 5.6 Hz, 1H), 3.30 (d, *J* = 8.8 Hz, 1H), 2.94 (d, *J* = 8.8 Hz, 1H), 1.92 (ddd, *J* = 14.4, 8.4, 6.0 Hz, 1H), 1.78 – 1.68 (m, 1H), 1.35 (s, 3H).

^{13}C NMR (100 MHz, CDCl_3) δ 151.8, 137.9, 137.1, 128.8, 128.3, 128.0, 127.6, 122.6, 119.3, 108.6, 66.6, 60.5, 54.3, 44.6, 42.9, 26.9.

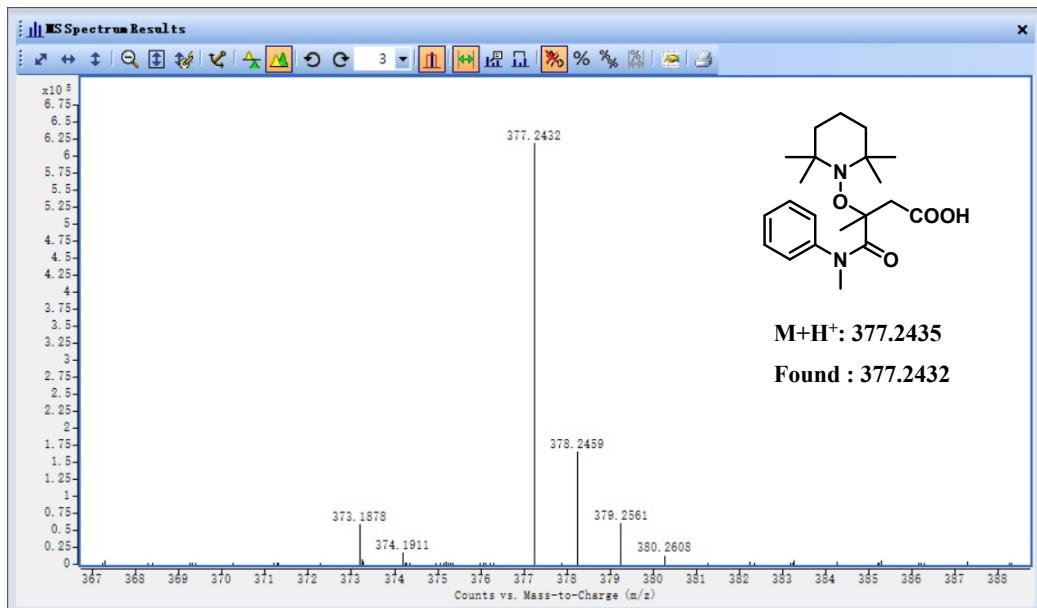
ESIHRMS: m/z Calcd. For $C_{18}H_{21}NO$: ($M+H$)⁺ 268.1696. Found: 268.1697.

6. Control Experiments

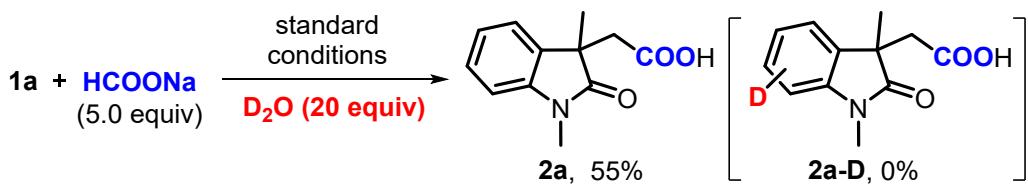
6.1 TEMPO Trapping Experiment



An oven-dried Schlenk tube (10 mL) containing a stirring bar was charged with *N*-methyl-*N*-phenylmethacrylamide (35.0 mg, 0.2 mmol, 1.0 equiv), HCOONa (60.8 mg, 1.0 mmol, 5.0 equiv), DABCO (11.2 mg, 0.1 mmol, 50 mol%), K₃PO₄ (42.5 mg, 0.2 mmol, 1.0 equiv) TEMPO (31.3 mg, 0.2 mmol, 1 equiv) and 4CzIPN (3.2 mg, 0.004 mmol, 2 mol%). Then anhydrous DMSO (2 mL) were added. Finally, the Schlenk tube was placed in an oil bath at the distance of 3 cm from a 45 W blue LED (wavelength: 450 nm) and stirred at 60 °C for 12 hours. After completion of the reaction, the reaction mixture was quenched with 2 mL of HCl (2 N), diluted with water (20 mL) and extracted with EtOAc (3 x 20 mL). The combined organic layers were washed with water and brine before dried over sodium sulfate and concentrated under reduced pressure. The residue was purified by silica gel flash chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2) to give the desired product 2a (7.5 mg, 17% yield) and 47% of starting material was recovered. **HRMS showed trapping of the reactive radical intermediate by TEMPO.**

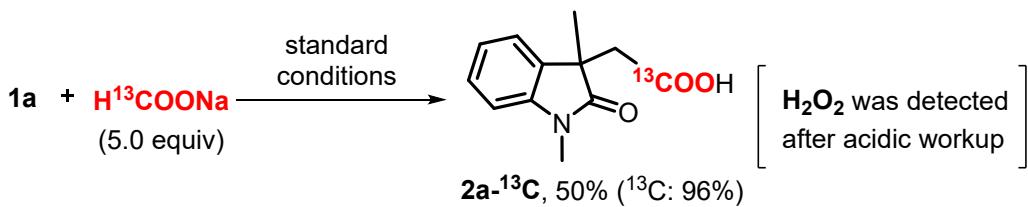


6.2 D-Labeling Experiment

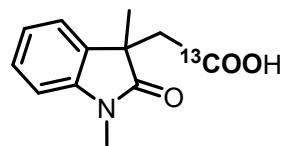
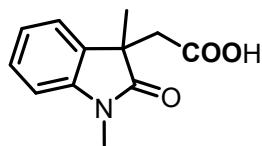


An oven-dried Schlenk tube (10 mL) containing a stirring bar was charged with *N*-methyl-*N*-phenylmethacrylamide (35.0 mg, 0.2 mmol, 1.0 equiv), HCOONa (60.8 mg, 1.0 mmol, 5.0 equiv), DABCO (11.2 mg, 0.1 mmol, 50 mol%), K₃PO₄ (42.5 mg, 0.2 mmol, 1.0 equiv) and 4CzIPN (3.2 mg, 0.004 mmol, 2 mol%). Then D₂O (4.0 mol, 20 equiv) and anhydrous DMSO (2 mL) were added. Finally, the Schlenk tube was placed in an oil bath at the distance of 3 cm from a 25 W blue LED (λ_{max} : 450 nm) and stirred at 60 °C for 12 hours. After completion of the reaction, the reaction mixture was quenched with 2 mL of HCl (2 N), diluted with water (30 mL) and extracted with EtOAc (3 x 30 mL). The combined organic layers were washed with water and brine before dried over sodium sulfate and concentrated under reduced pressure. The residue was purified by silica gel flash chromatography (5% AcOH in petroleum ether/ EtOAc 1/5 ~ 1/2) to give the desired product **2a** (24.1 mg, 55% yield).

6.3 ¹³C Isotope Labeling Experiment and Detection of H₂O₂



The experiment was conducted following the standard reaction condition but using H¹³COONa instead of HCOONa. The products obtained in the reaction were detected by HRMS. The result showed **2a-¹³C** in a ratio of 96%.



Chemical Formula: C₁₂H₁₃NO₃

calculated m/z:

219.0895 (100.0%),

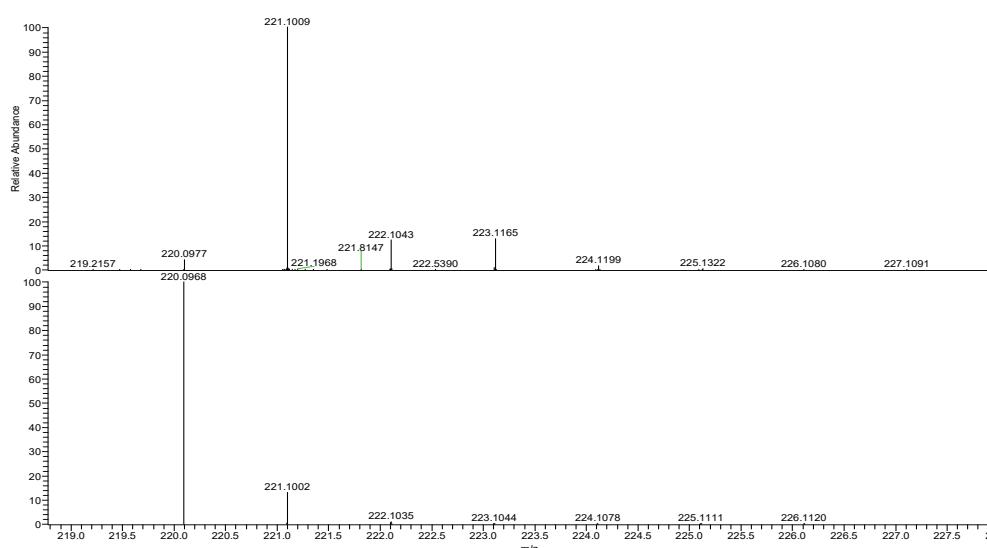
220.0929 (13.0%)

Chemical Formula: C₁₁¹³CH₁₃NO₃

calculated m/z:

220.0929 (100.0%),

221.0963 (11.9%)

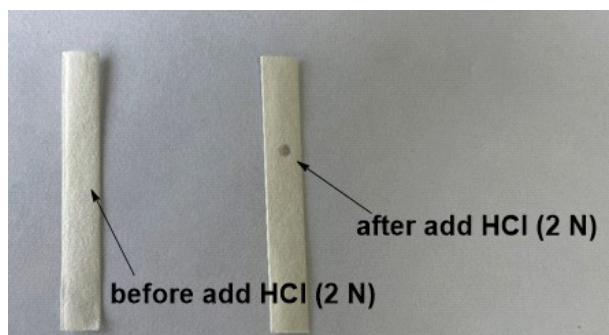


NL:
2.16E5
X:H282#11 RT:
0.29 AV: 1 T:
FTMS + c ESI
Full ms
[150.00-2000.00]

NL:
8.68E5
C₁₂H₁₄NO₃:
C₁₂H₁₄N₁O₃
p_a Chrg 1

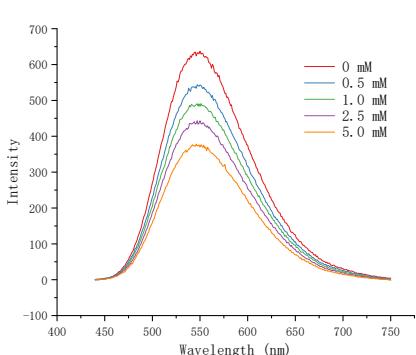
up: experimental data

When the reaction was completed, dipping the solution of the reaction mixture on the repotassium iodide-starch test paper showed no color change. After addition of HCl (2 N), dipping the solution of this reaction on the repotassium iodide-starch test paper showed the color changed to blue.

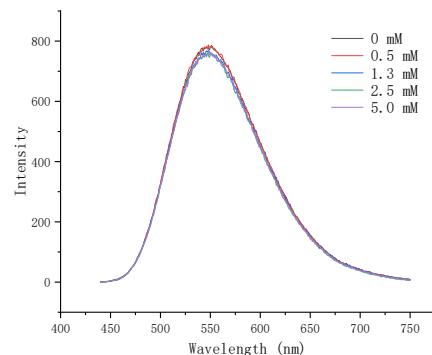


6.4 Stern-Volmer Fluorescence Quenching Analysis

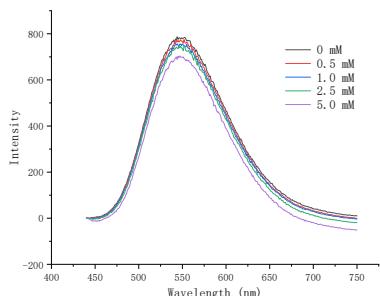
Stern-Volmer fluorescence quenching experiments were run with freshly prepared solution of 5×10^{-5} M 4CzIPN in degassed anhydrous DMSO at room temperature. The solution was irradiated at 440 nm and the fluorescence was measured from 440 nm to 750 nm. Data was collected on an Agilent Technologies F-4600 spectrofluorometer at 25 °C. Parameters: data interval = 1 nm, scan rate = 645 nm/min, Averaging time = 0.1 sec. The samples were measured in Jingke ES quartz cuvettes (chamber volume = 2.5 mL, H × W × D = 56 nm × 12.5 nm × 12.5 nm).



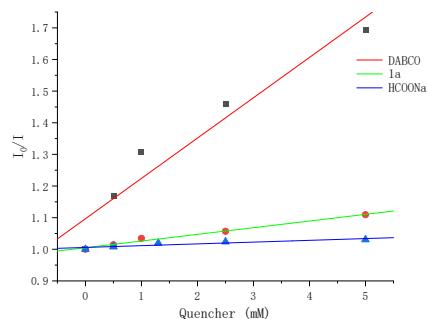
DABCO



HCOONa



Substrate 1a



Stern-Volmer Quenching

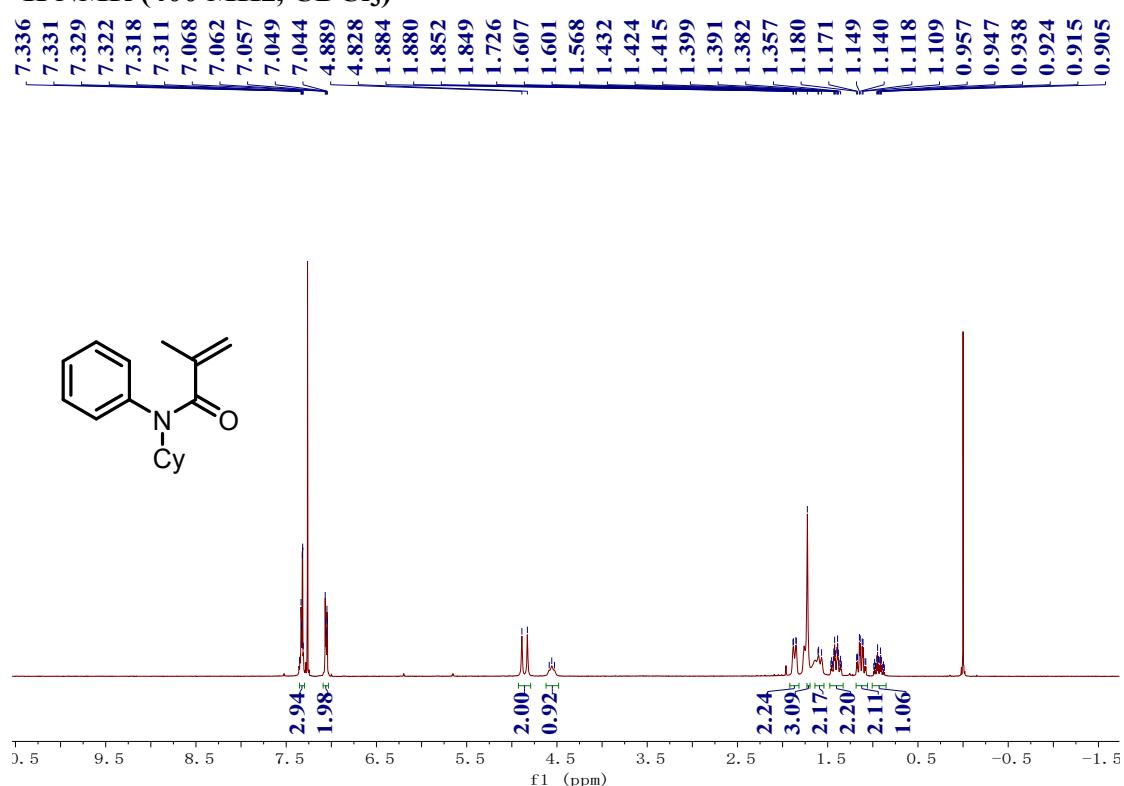
7. References

- [1] L. Zheng, H. Huang, C. Yang and W. Xia, UV Light-Mediated Difunctionalization of Alkenes through Aroyl Radical Addition/1,4-/1,2-Aryl Shift Cascade Reactions, *Org. Lett.*, 2015, **17**, 1034-1037.
- [2] A. J.-L. Ayitou and J. Sivaguru, Light-Induced Transfer of Molecular Chirality in Solution: Enantiospecific Photocyclization of Molecularly Chiral Acrylanilides, *J. Am. Chem. Soc.*, 2009, **131**, 5036-5037.
- [3] X.-Y. Duan, X.-L. Yang, P.-P. Jia, M. Zhang and B. Han, *Org. Lett.*, 2015, **17**, 6022-6025.
- [4] K. Lu, X.-W. Han, W.-W. Yao, Y.-X. Luan, Y.-X. Wang, H. Chen, X.-T. Xu, K. Zhang and M. Ye, Hydrazonyl Radical-Participated Tandem Reaction: A Strategy for the Synthesis of Pyrazoline-Functionalized Oxindoles, *ACS Catal.*, 2018, **8**, 3913-3917.
- [5] X. Sun, J.-P. Zhu, Q.-C. Qiu, Y.-L. He, D.-R. Hu, X.-L. Li, G.-P. Lu, Y.-H. Yuan, X.-F. Zhang, X. Xu, M. Yu and B. Wu, Metal-Free Visible-Light-Driven Cascade Cyclization Reaction to Synthesize 2-Oxindoles via Benzoyl and Phenylsulfinyl Radicals with Acrylamide Derivatives. *Org. Biomol. Chem.*, 2022, **20**, 8042-8048.
- [6] T. Nishio, H. Asai and T. Miyazaki, Photochemical Reactions of *N*-(2-Halogenoalkanoyl) Derivatives of Anilines, *Helv. Chim. Acta.*, 2000, **83**, 1475-1483.
- [7] S. Ding, H. Ren, M. Zhu, Q. Ma, Z. Miao and P. Li, Silver(I)-Mediated Oxidation/Cyclization of Acrylamides with Alkyl Trifluoroborates, *Synthetic Commun.*, 2021, **51**, 593-600.
- [8] X.-W. Chen, J.-P. Yue, K. Wang, Y.-Y. Gui, Y.-N. Niu, J. Liu, C.-K. Ran, W. Kong, W.-J. Zhou and D.-G. Yu, Nickel-Catalyzed Asymmetric Reductive Carbo-Carboxylation of Alkenes with CO₂, *Angew. Chem., Int. Ed.*, 2021, **60**, 14068-14075; *Angew. Chem.*, 2021, **133**, 14187-14194.

8. NMR Spectra

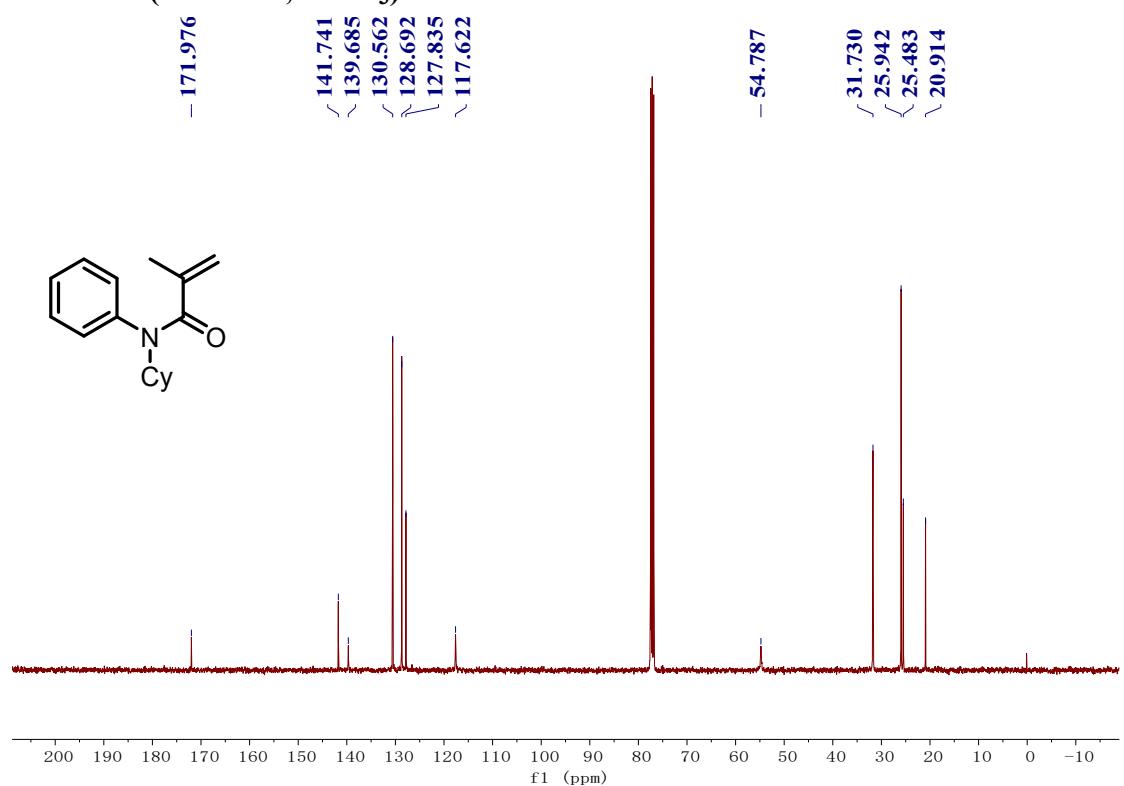
N-cyclohexyl-*N*-phenylmethacrylamide (1p)

¹H NMR (400 MHz, CDCl₃)



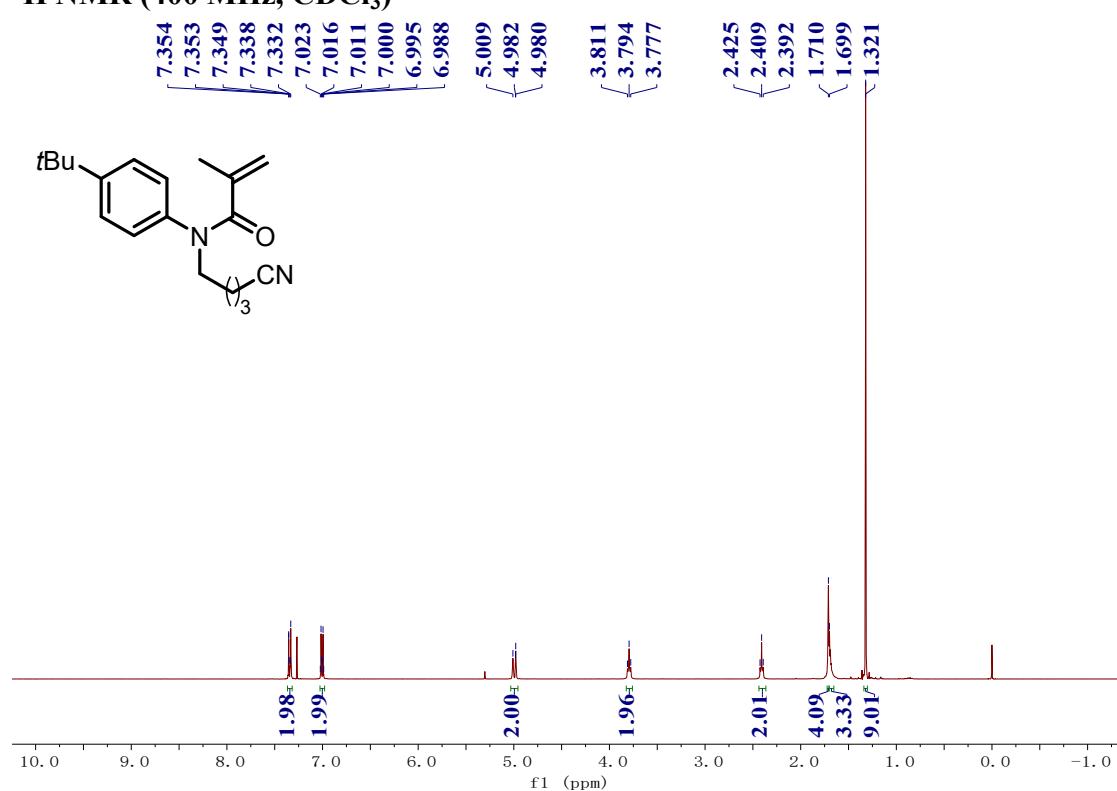
N-cyclohexyl-*N*-phenylmethacrylamide (1p)

¹³C NMR (100 MHz, CDCl₃)



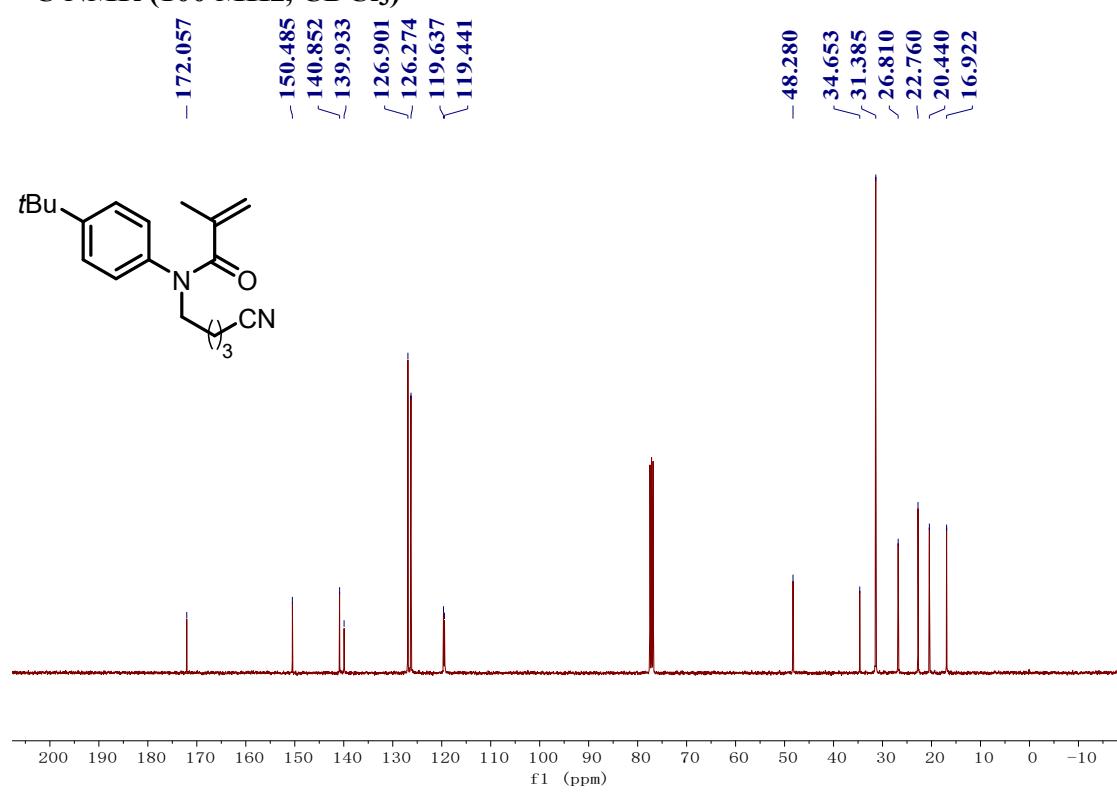
N-(4-(*tert*-butyl)phenyl)-*N*-(4-cyanobutyl)methacrylamide (**1s**)

¹H NMR (400 MHz, CDCl₃)



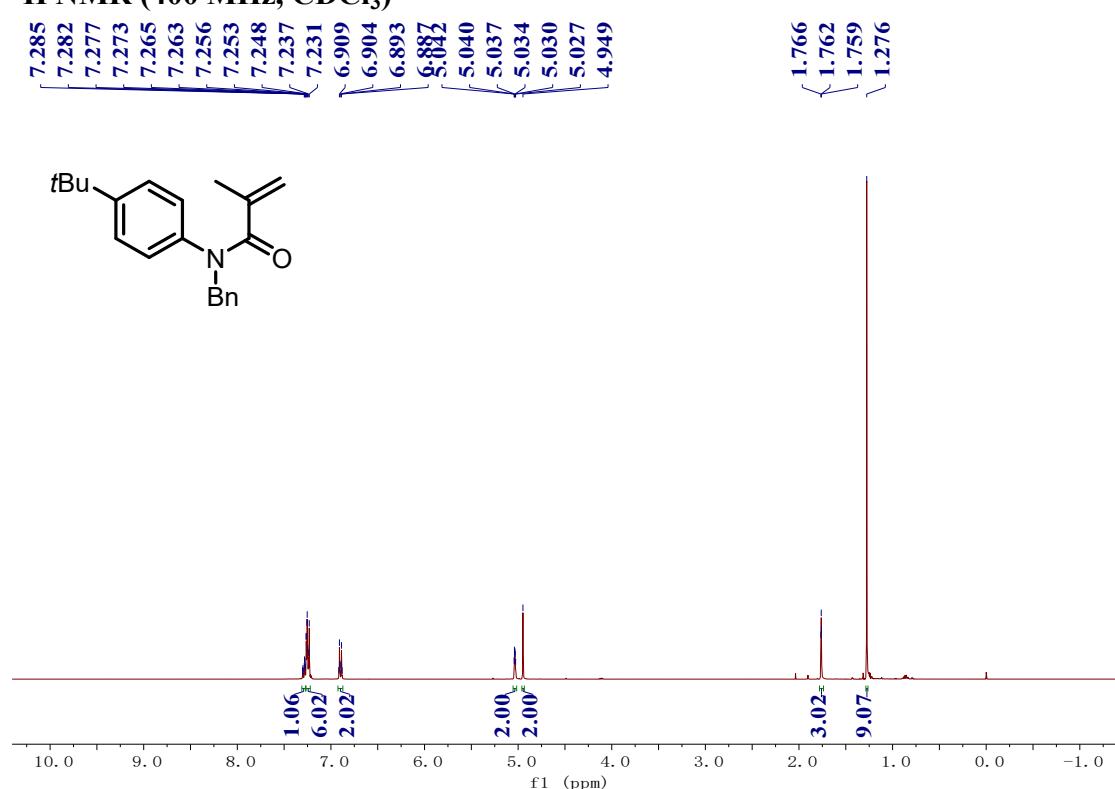
N-(4-(*tert*-butyl)phenyl)-*N*-(4-cyanobutyl)methacrylamide (**1s**)

¹³C NMR (100 MHz, CDCl₃)



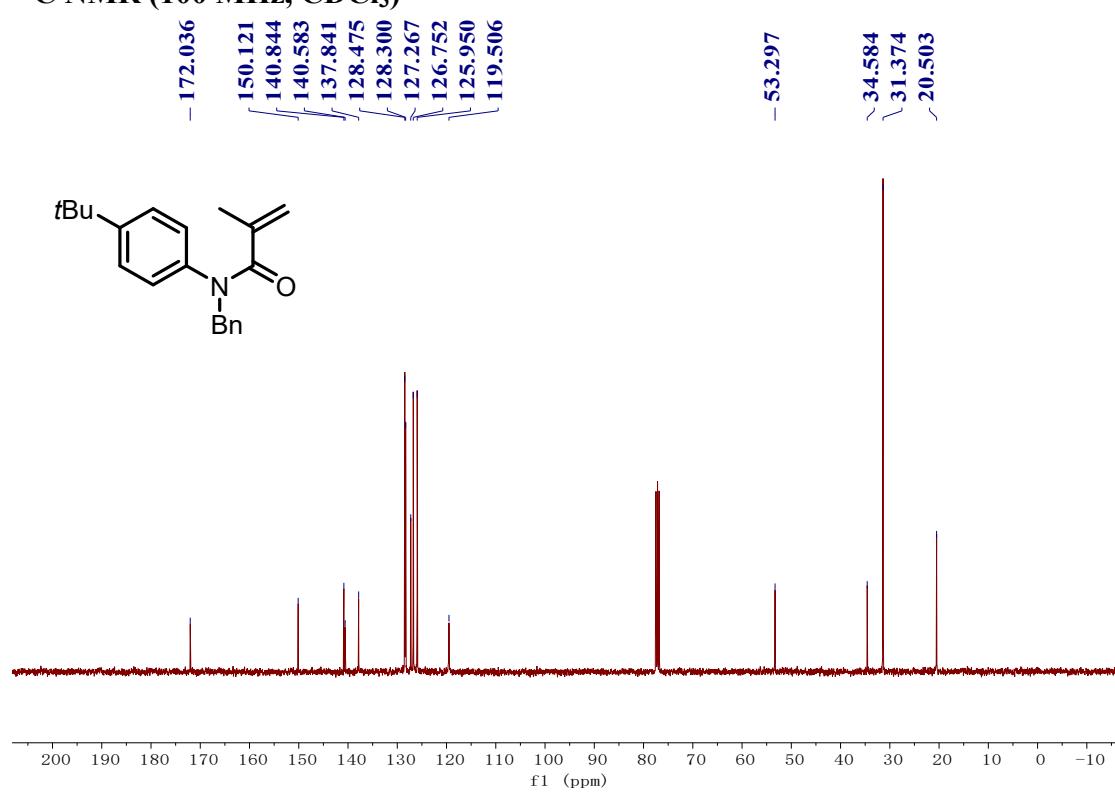
N-benzyl-*N*-(4-(*tert*-butyl)phenyl)methacrylamide (**1t**)

¹H NMR (400 MHz, CDCl₃)



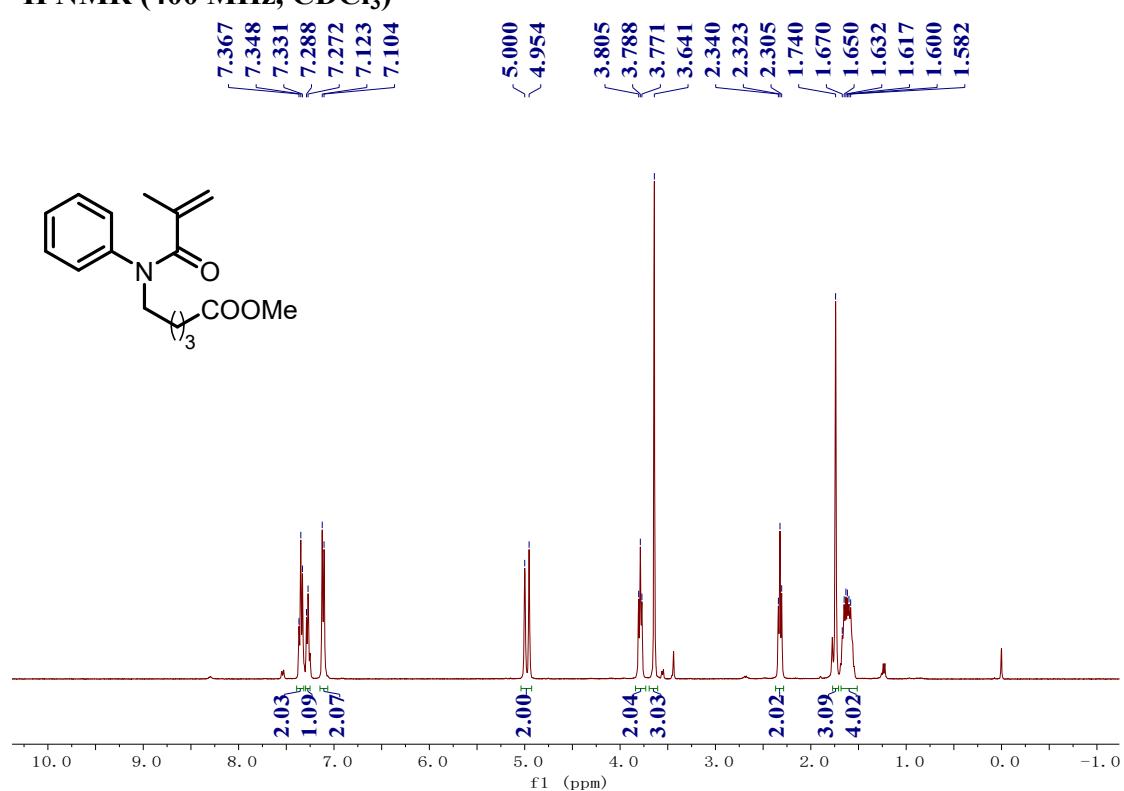
N-benzyl-*N*-(4-(*tert*-butyl)phenyl)methacrylamide (**1t**)

¹³C NMR (100 MHz, CDCl₃)



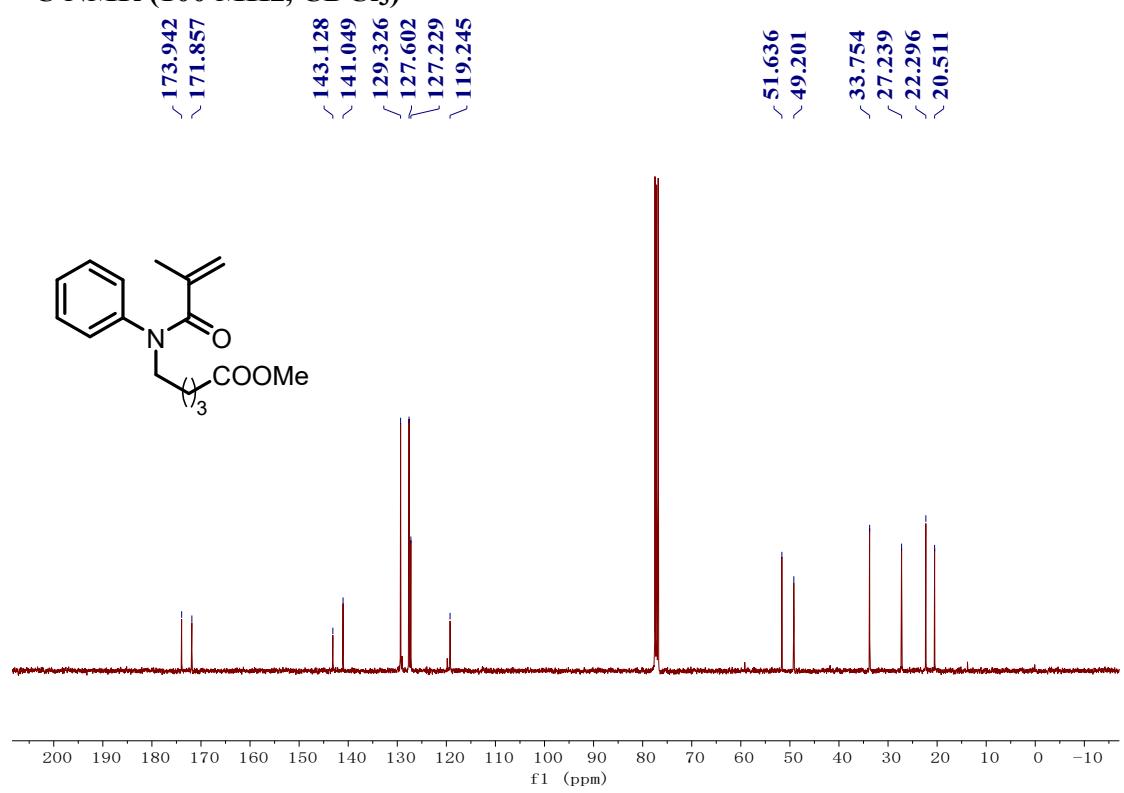
methyl 5-(N-phenylmethacrylamido)pentanoate (1u)

¹H NMR (400 MHz, CDCl₃)



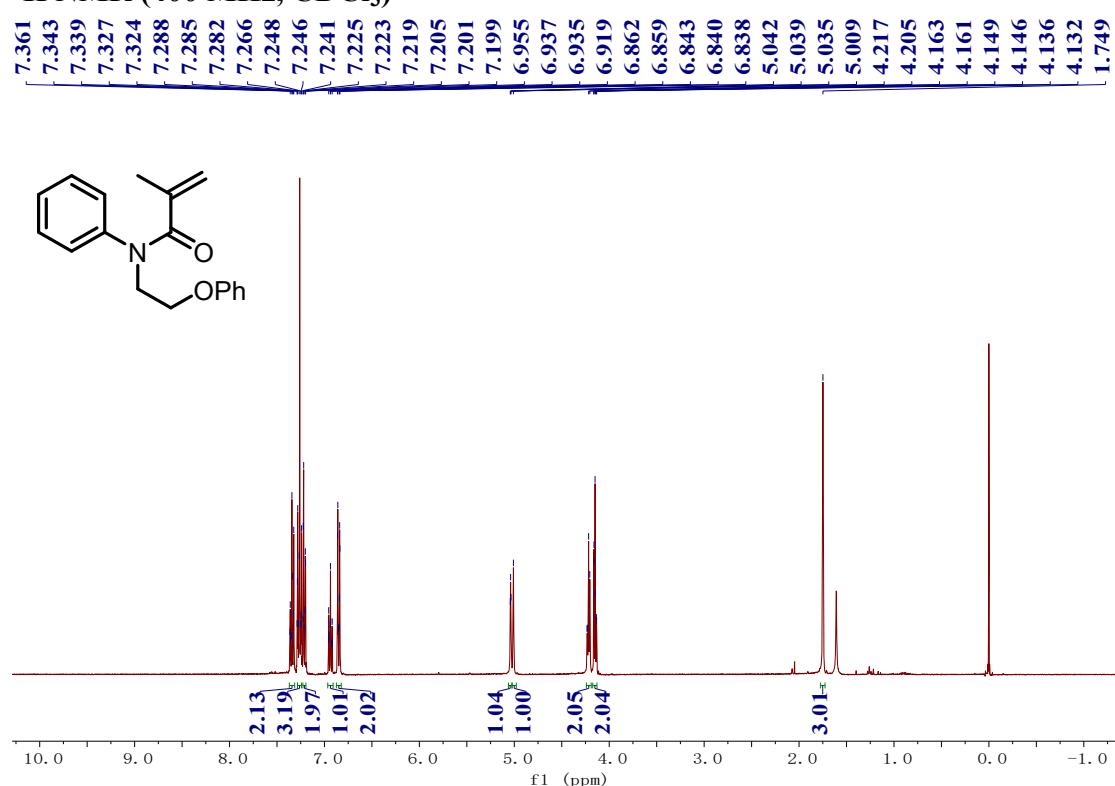
methyl 5-(N-phenylmethacrylamido)pentanoate (1u)

¹³C NMR (100 MHz, CDCl₃)



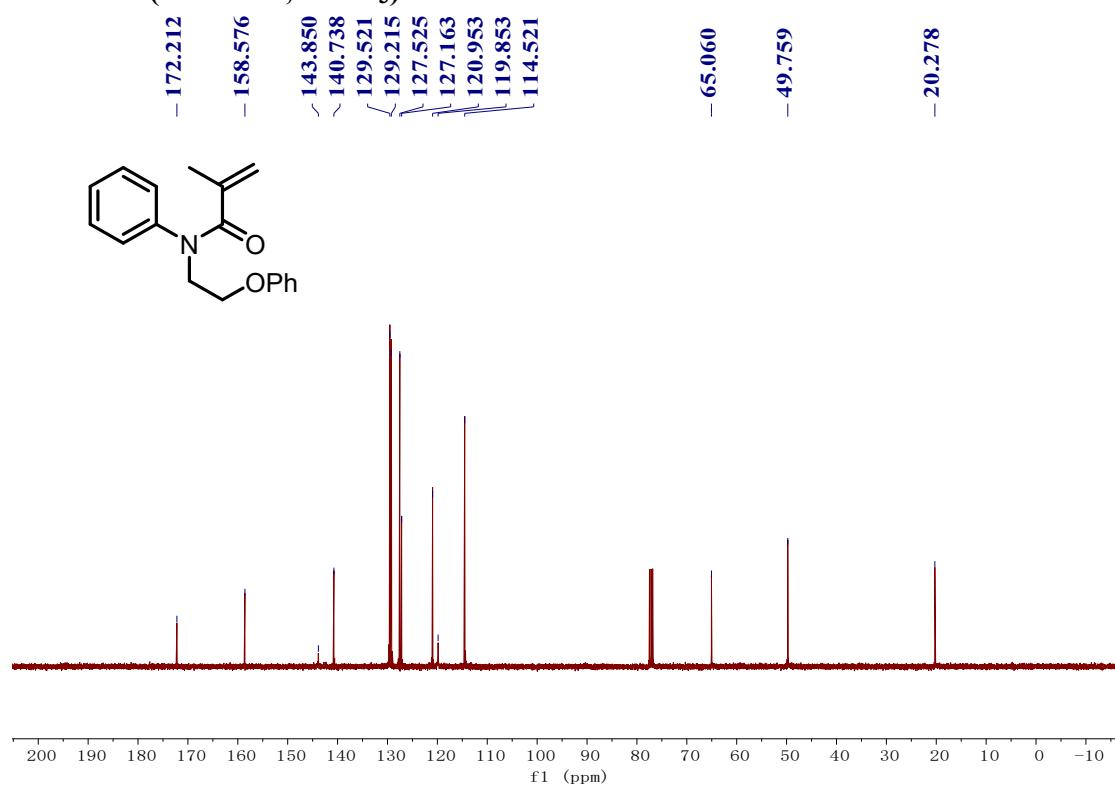
***N*-(2-phenoxyethyl)-*N*-phenylmethacrylamide (**1v**)**

¹H NMR (400 MHz, CDCl₃)



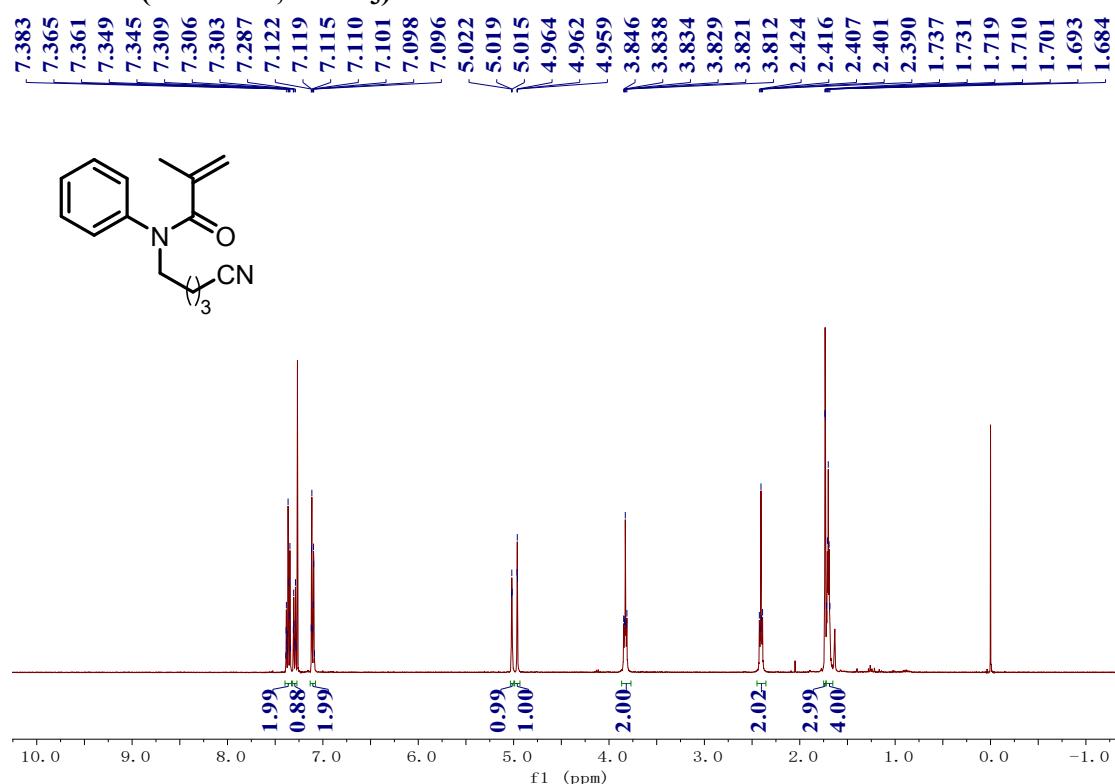
***N*-(2-phenoxyethyl)-*N*-phenylmethacrylamide (**1v**)**

¹³C NMR (100 MHz, CDCl₃)



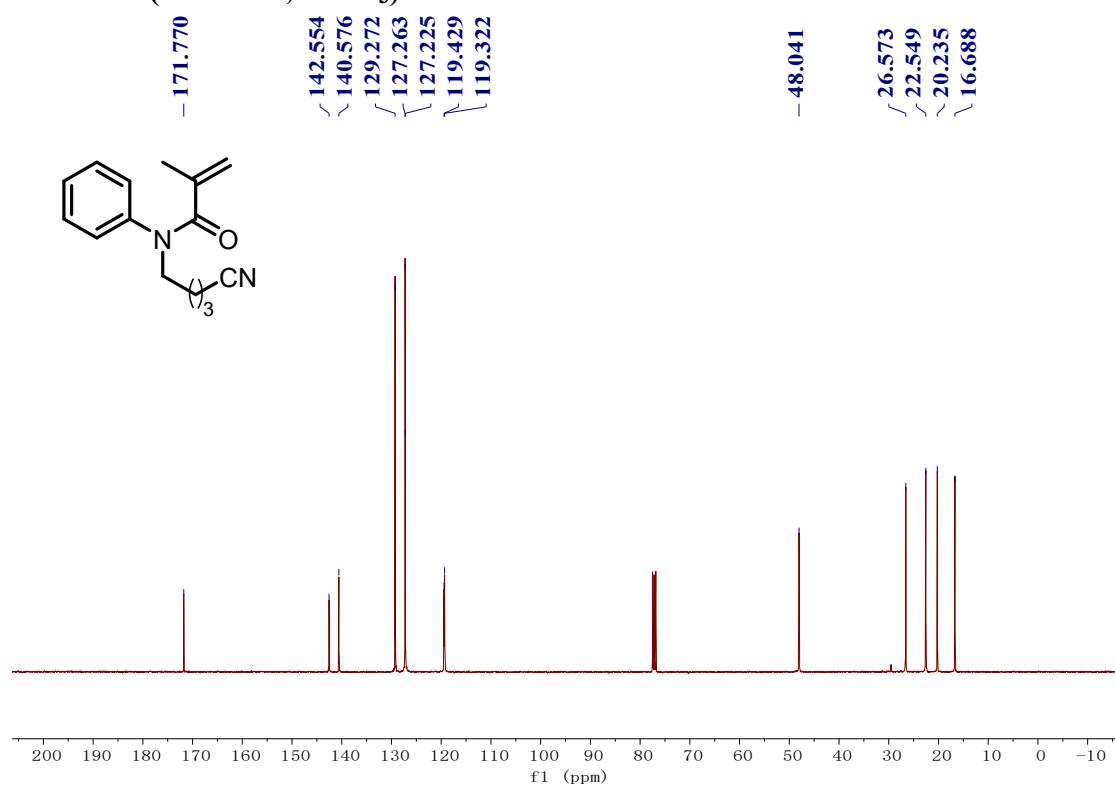
N-(4-cyanobutyl)-*N*-phenylmethacrylamide (**1w**)

¹H NMR (400 MHz, CDCl₃)



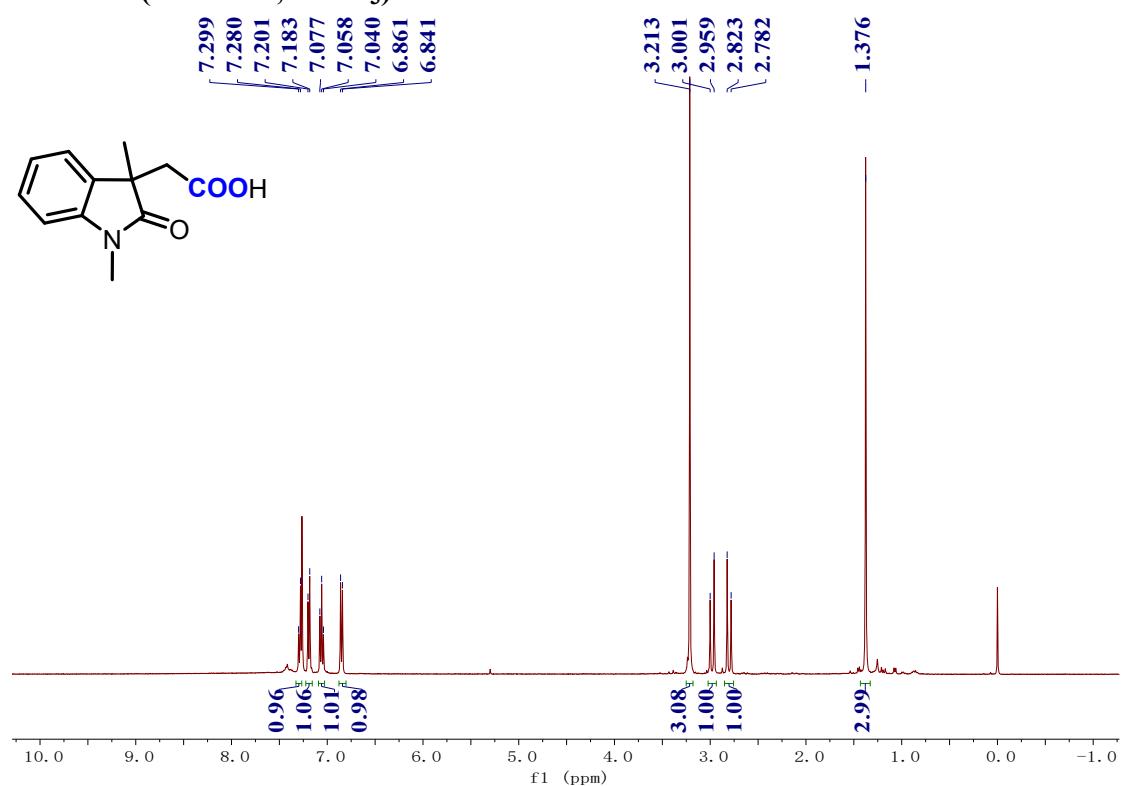
N-(4-cyanobutyl)-*N*-phenylmethacrylamide (**1w**)

¹³C NMR (100 MHz, CDCl₃)



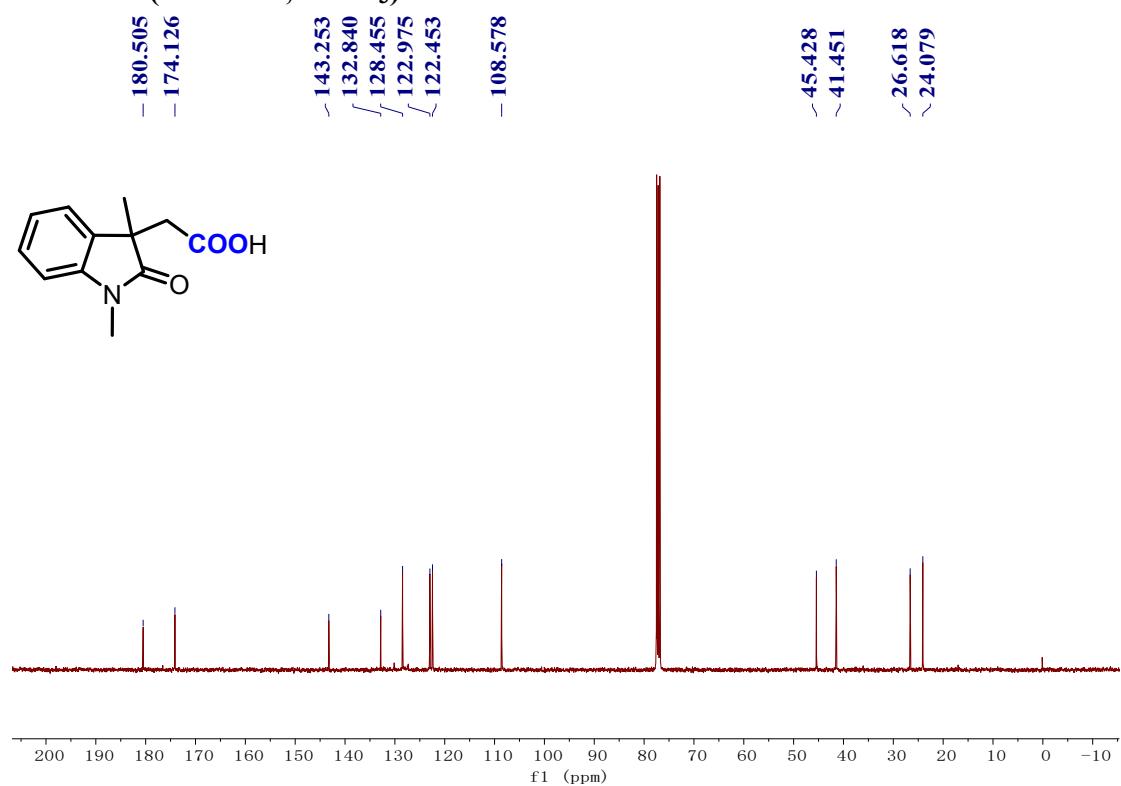
2-(1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2a)

¹H NMR (400 MHz, CDCl₃)



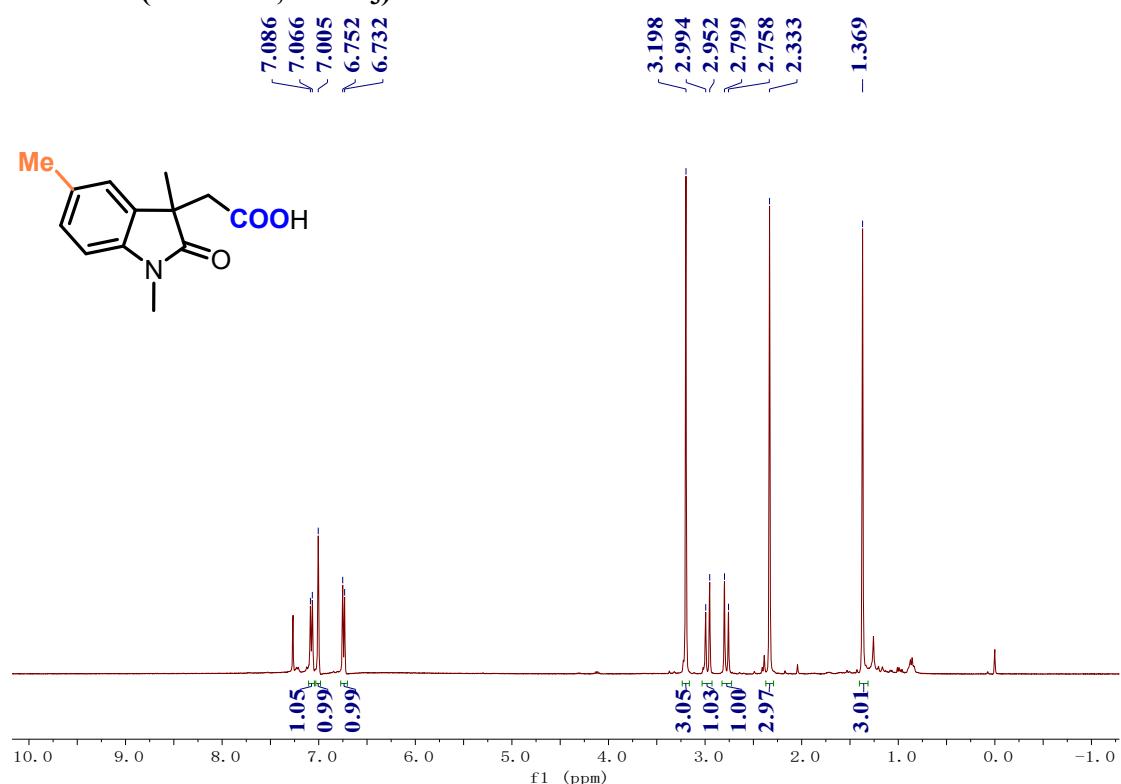
2-(1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2a)

¹³C NMR (100 MHz, CDCl₃)



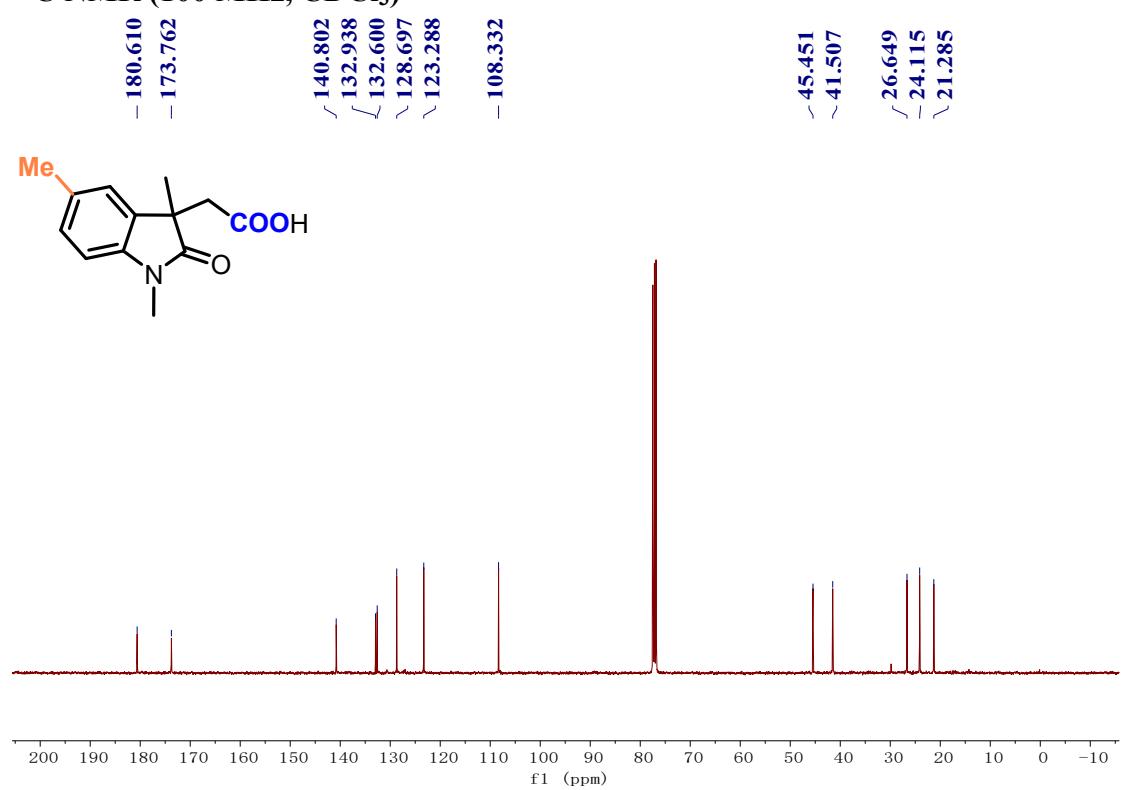
2-(1,3,5-trimethyl-2-oxoindolin-3-yl)acetic acid (2b)

¹H NMR (400 MHz, CDCl₃)



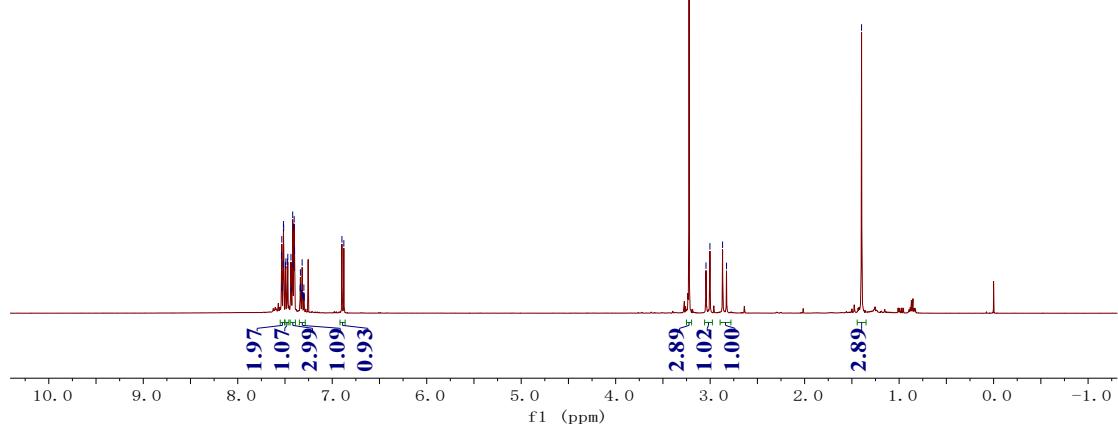
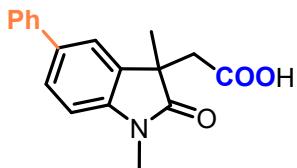
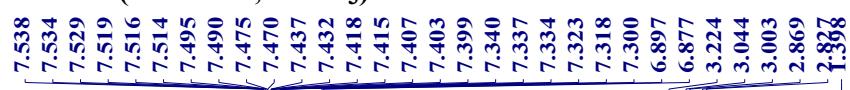
2-(1,3,5-trimethyl-2-oxoindolin-3-yl)acetic acid (2b)

¹³C NMR (100 MHz, CDCl₃)



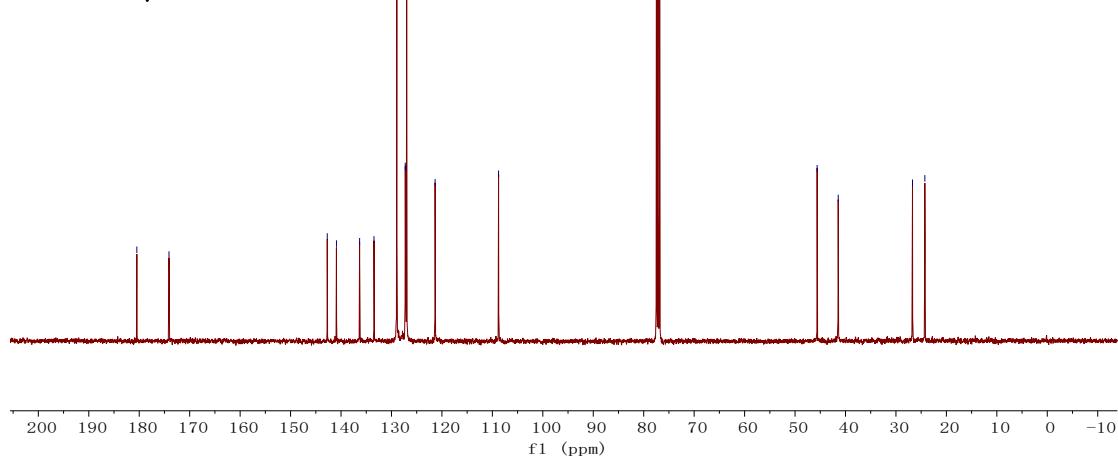
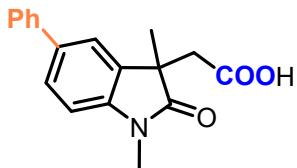
2-(1,3-dimethyl-2-oxo-5-phenylindolin-3-yl)acetic acid (2c)

¹H NMR (400 MHz, CDCl₃)



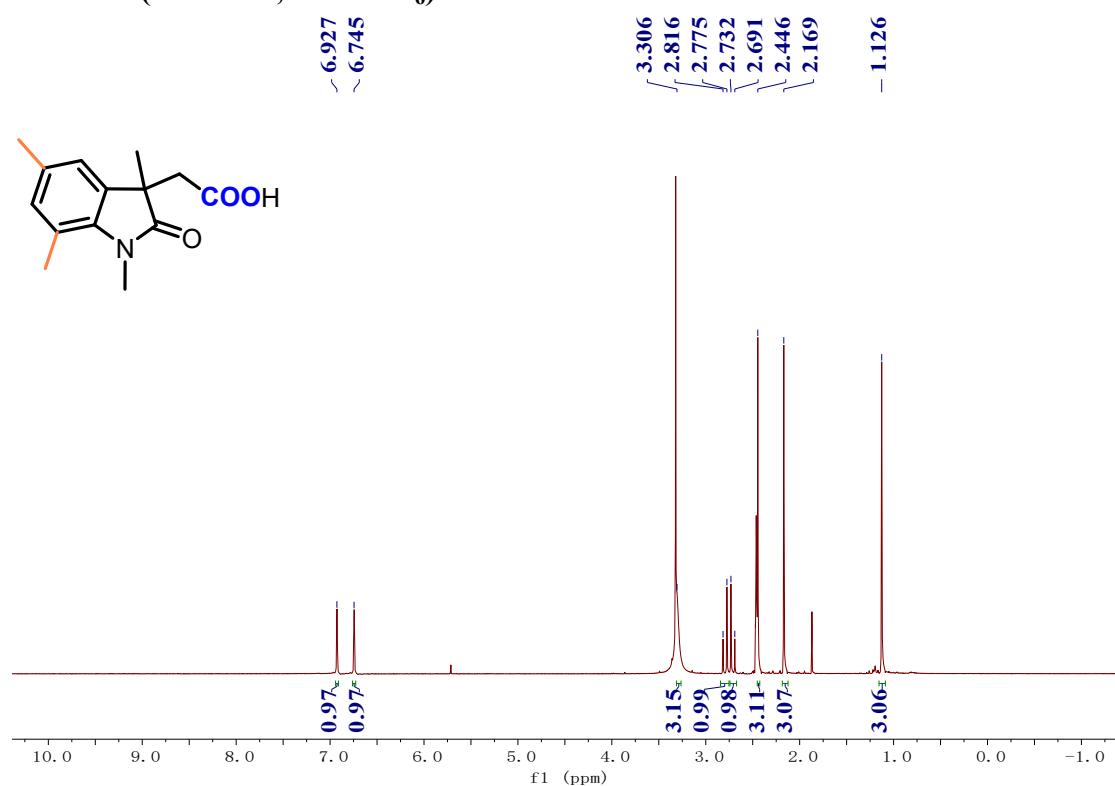
2-(1,3-dimethyl-2-oxo-5-phenylindolin-3-yl)acetic acid (2c)

¹³C NMR (100 MHz, CDCl₃)



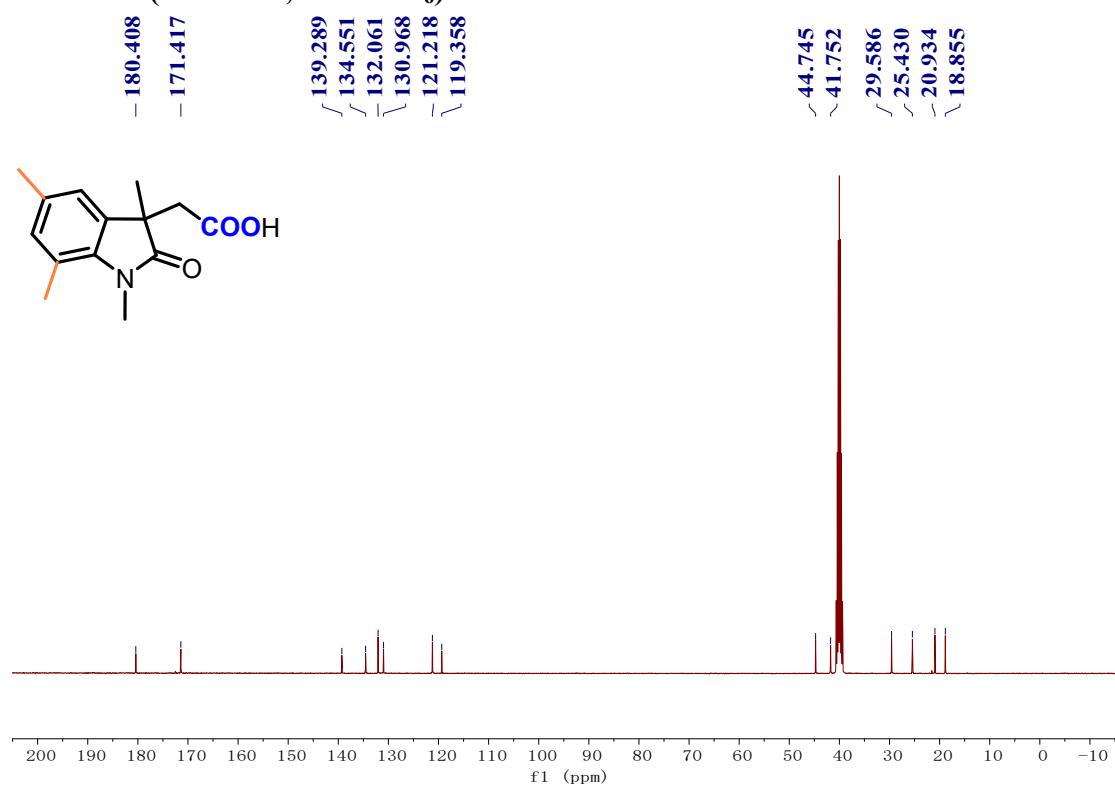
2-(1,3,5,7-tetramethyl-2-oxoindolin-3-yl)acetic acid (2d)

¹H NMR (400 MHz, DMSO-d₆)



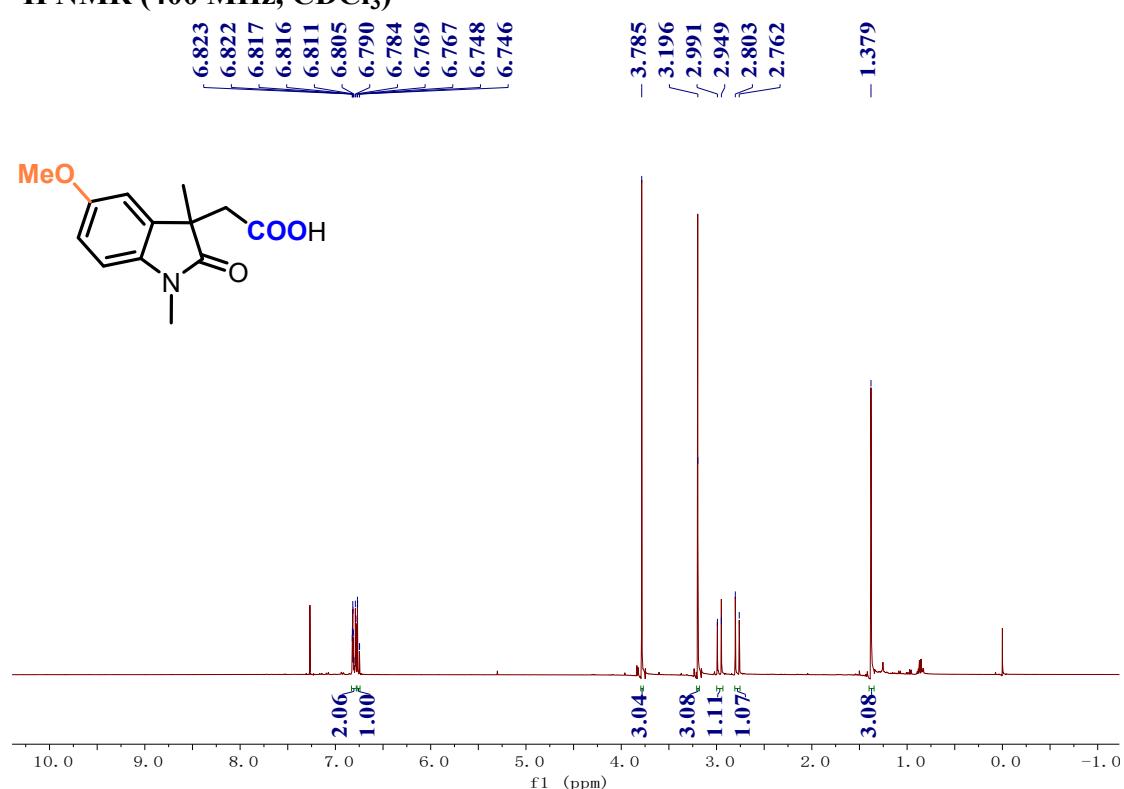
2-(1,3,5,7-tetramethyl-2-oxoindolin-3-yl)acetic acid (2d)

¹³C NMR (100 MHz, DMSO-d₆)



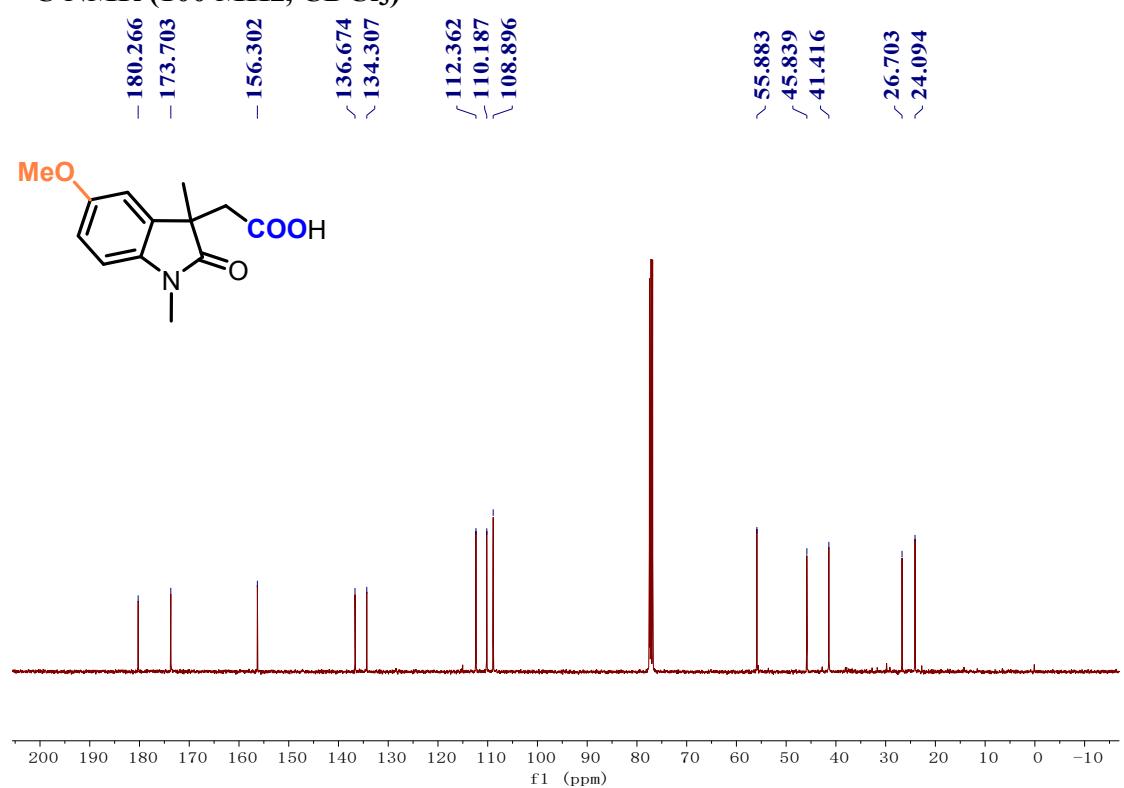
2-(5-methoxy-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2e)

¹H NMR (400 MHz, CDCl₃)



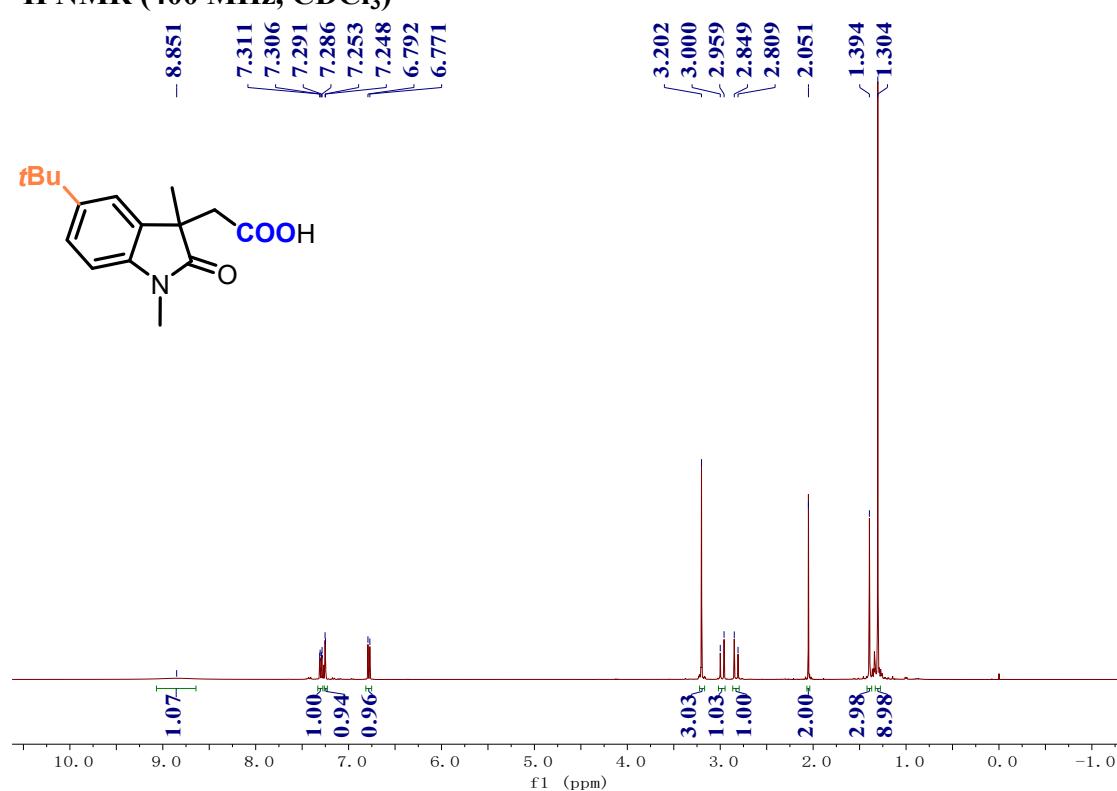
2-(5-methoxy-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2e)

¹³C NMR (100 MHz, CDCl₃)



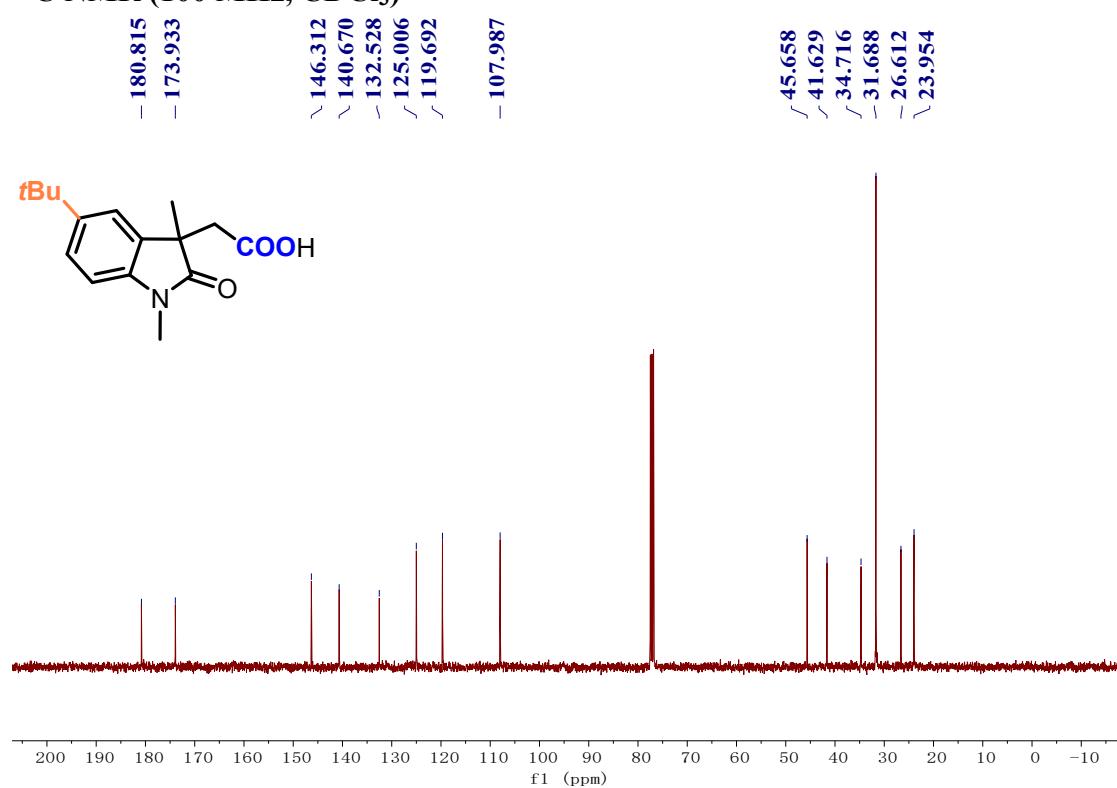
2-(5-(*tert*-butyl)-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2f)

¹H NMR (400 MHz, CDCl₃)



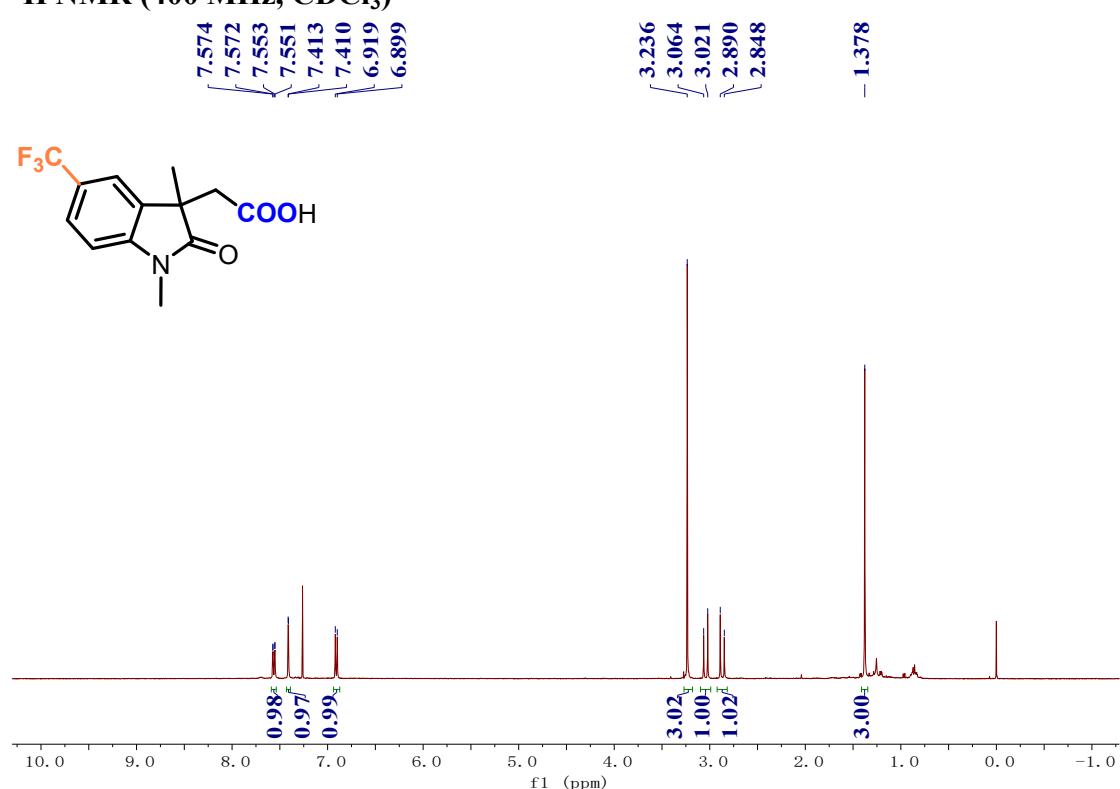
2-(5-(*tert*-butyl)-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2f)

¹³C NMR (100 MHz, CDCl₃)



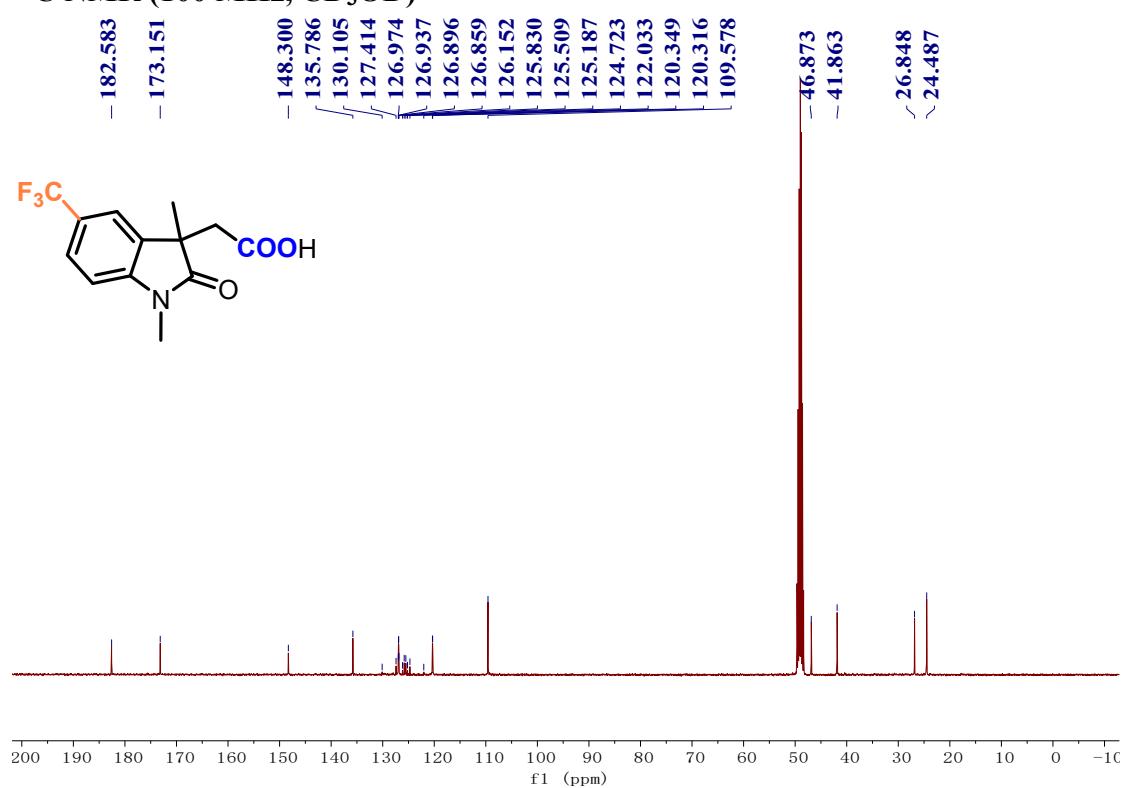
2-(1,3-dimethyl-2-oxo-5-(trifluoromethyl)indolin-3-yl)acetic acid (2g)

¹H NMR (400 MHz, CDCl₃)



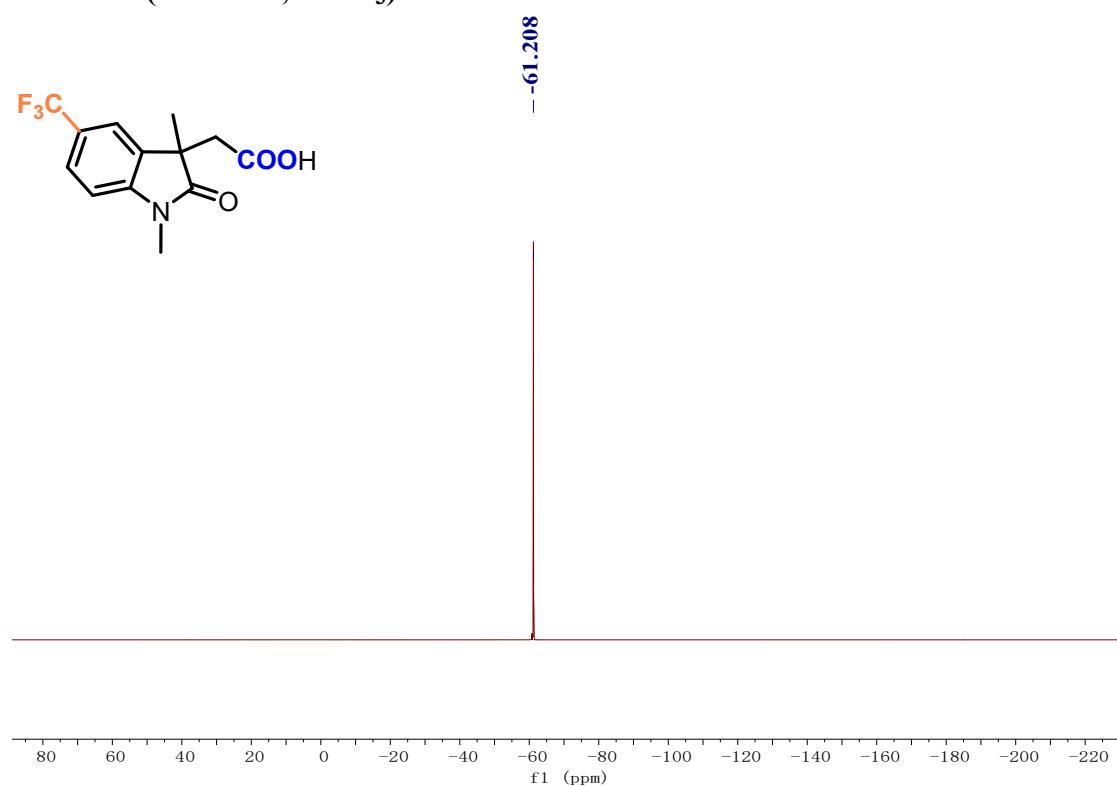
2-(1,3-dimethyl-2-oxo-5-(trifluoromethyl)indolin-3-yl)acetic acid (2g)

¹³C NMR (100 MHz, CD₃OD)



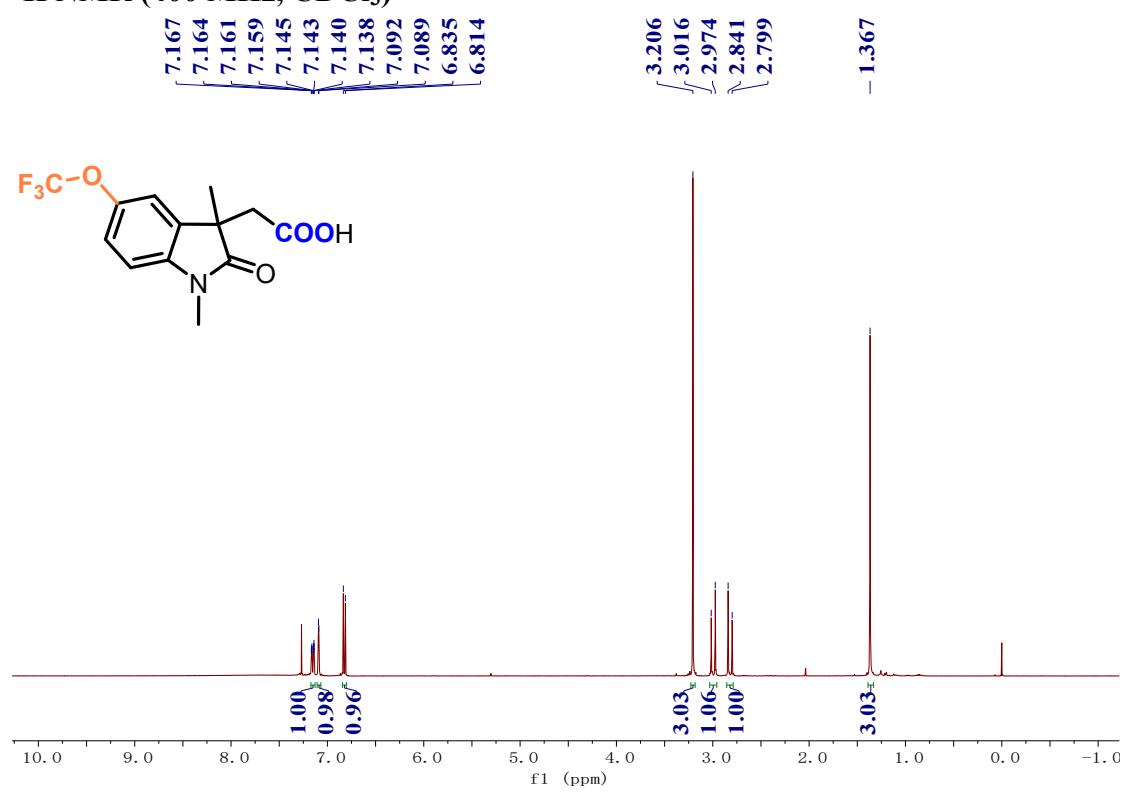
2-(1,3-dimethyl-2-oxo-5-(trifluoromethyl)indolin-3-yl)acetic acid (2g)

¹⁹F NMR (376 MHz, CDCl₃)



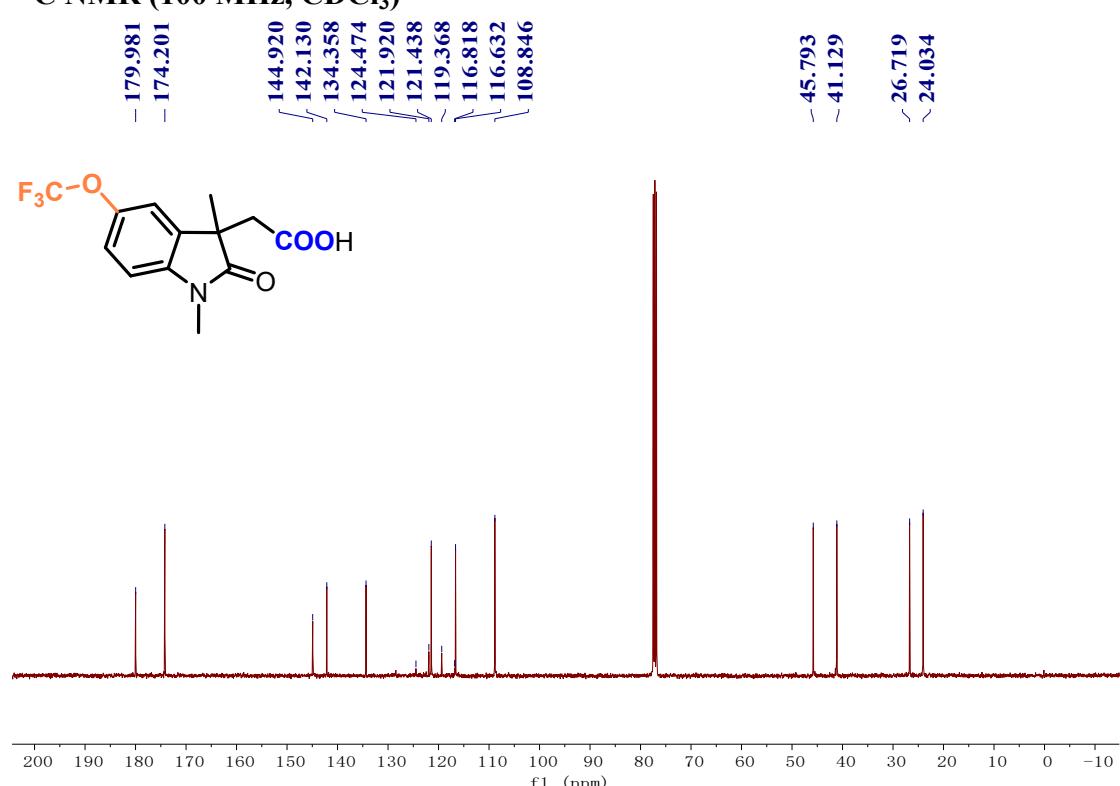
2-(1,3-dimethyl-2-oxo-5-(trifluoromethoxy)indolin-3-yl)acetic acid (2h)

¹H NMR (400 MHz, CDCl₃)



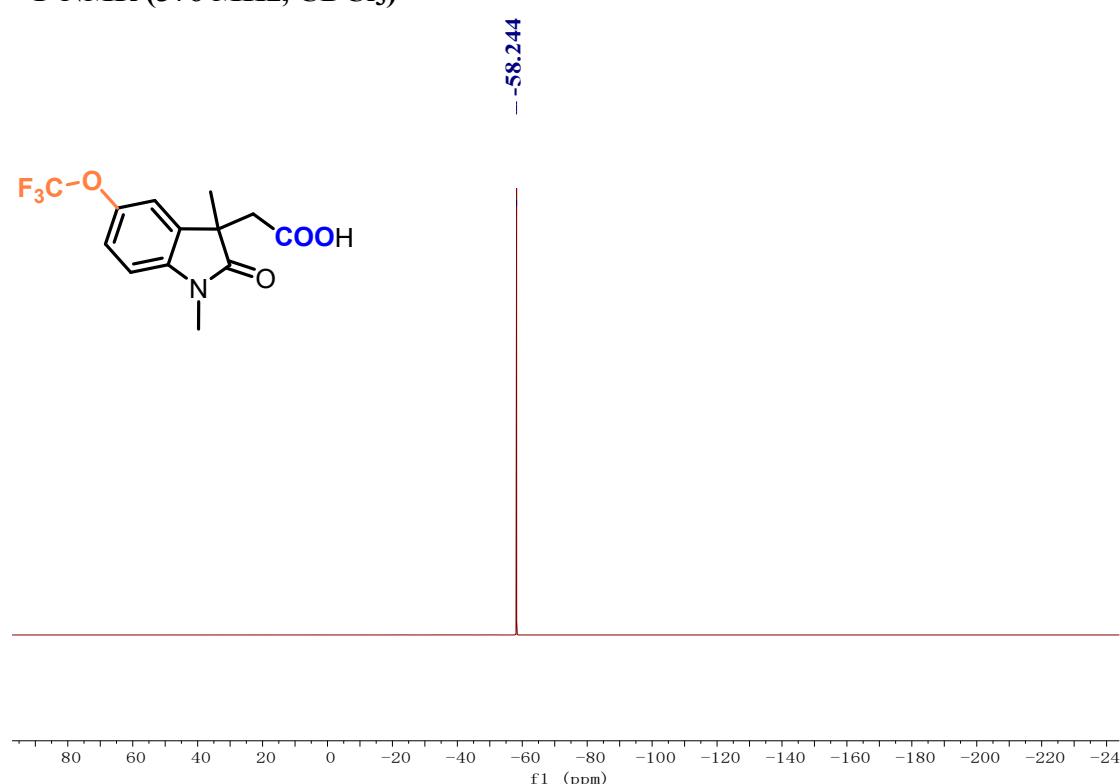
2-(1,3-dimethyl-2-oxo-5-(trifluoromethoxy)indolin-3-yl)acetic acid (2h)

¹³C NMR (100 MHz, CDCl₃)



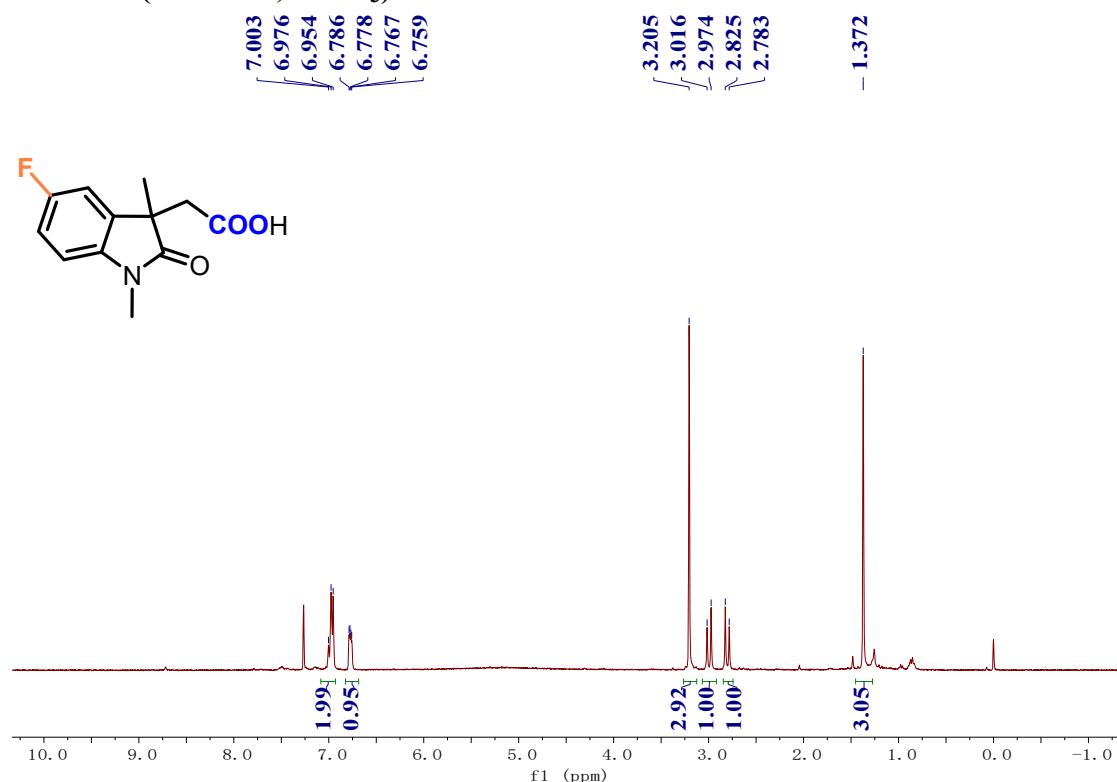
2-(1,3-dimethyl-2-oxo-5-(trifluoromethoxy)indolin-3-yl)acetic acid (2h)

¹⁹F NMR (376 MHz, CDCl₃)



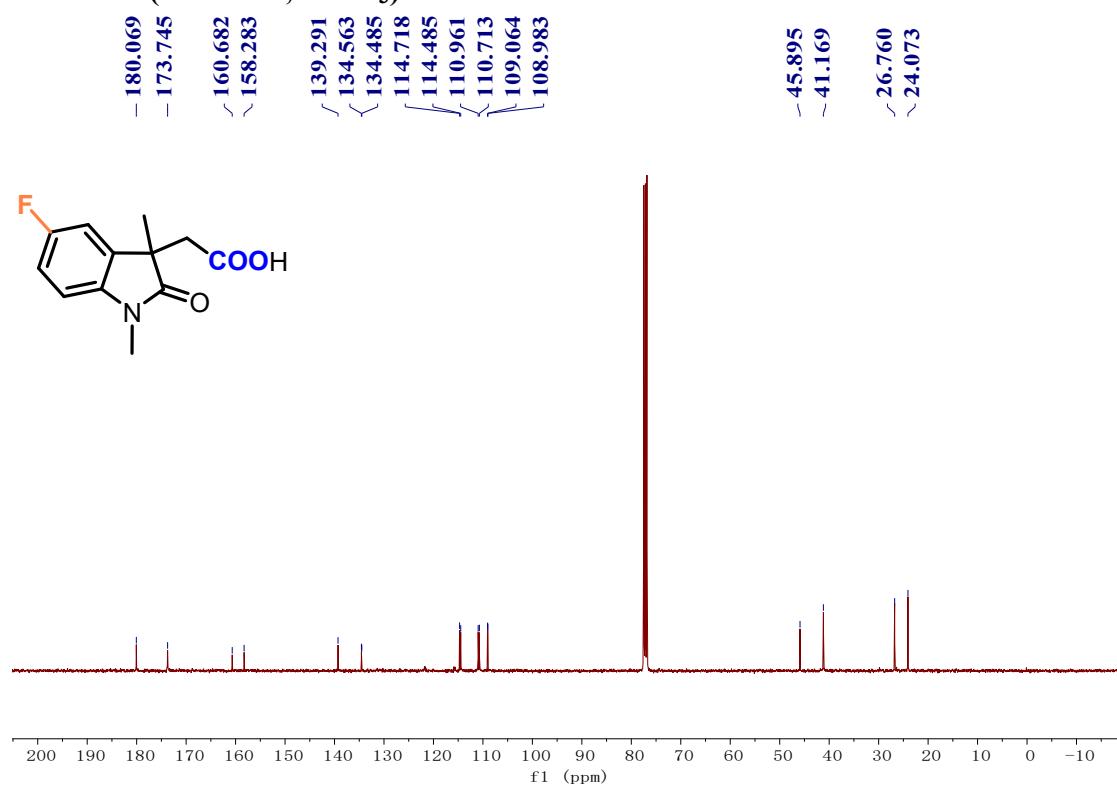
2-(5-fluoro-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2i)

¹H NMR (400 MHz, CDCl₃)



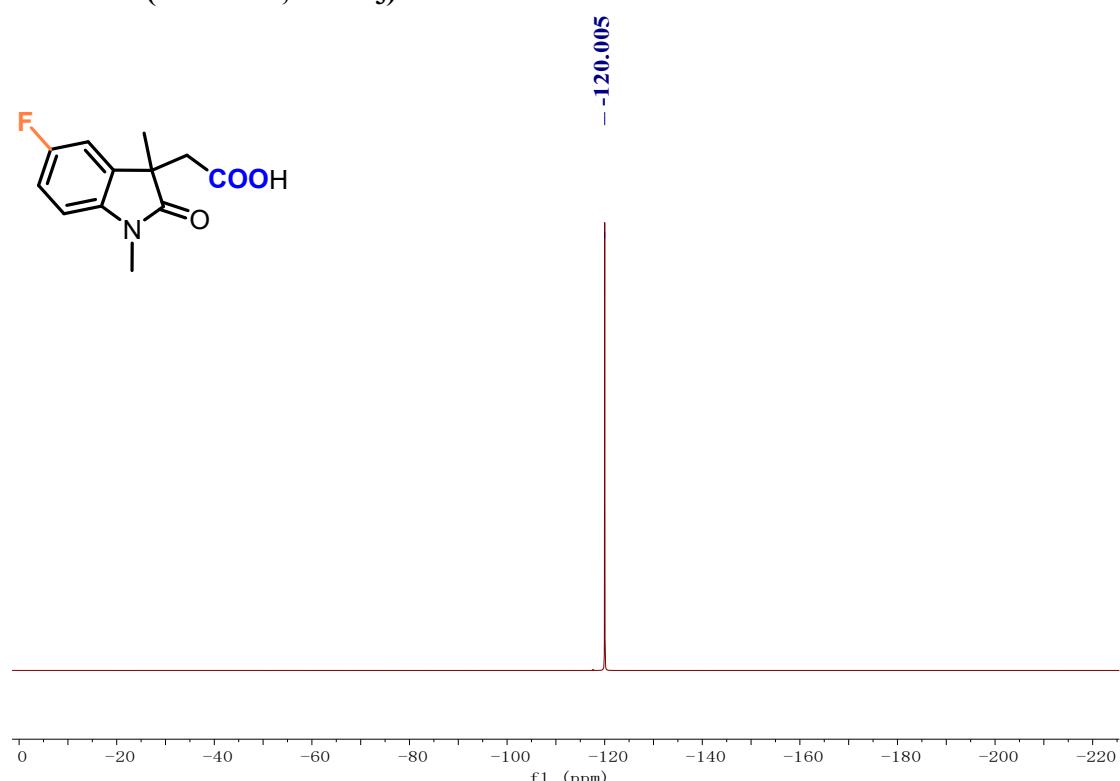
2-(5-fluoro-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2i)

¹³C NMR (100 MHz, CDCl₃)



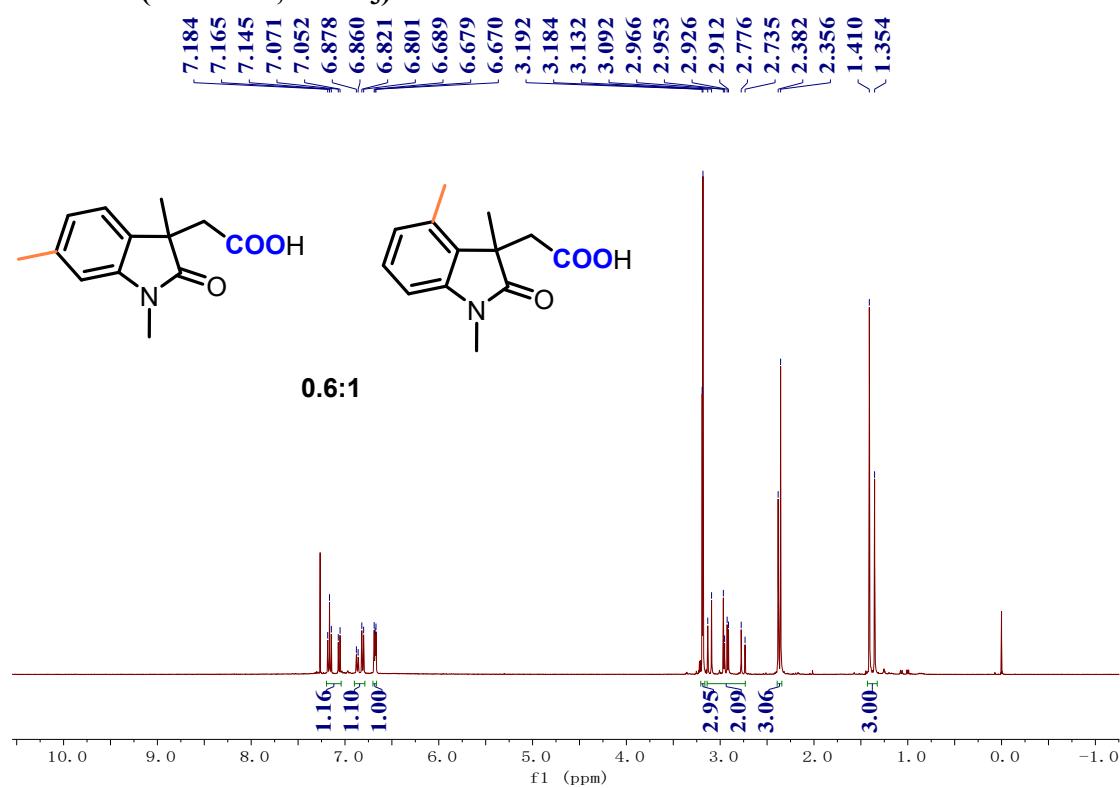
2-(5-fluoro-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (2i)

^{19}F NMR (376 MHz, CDCl_3)



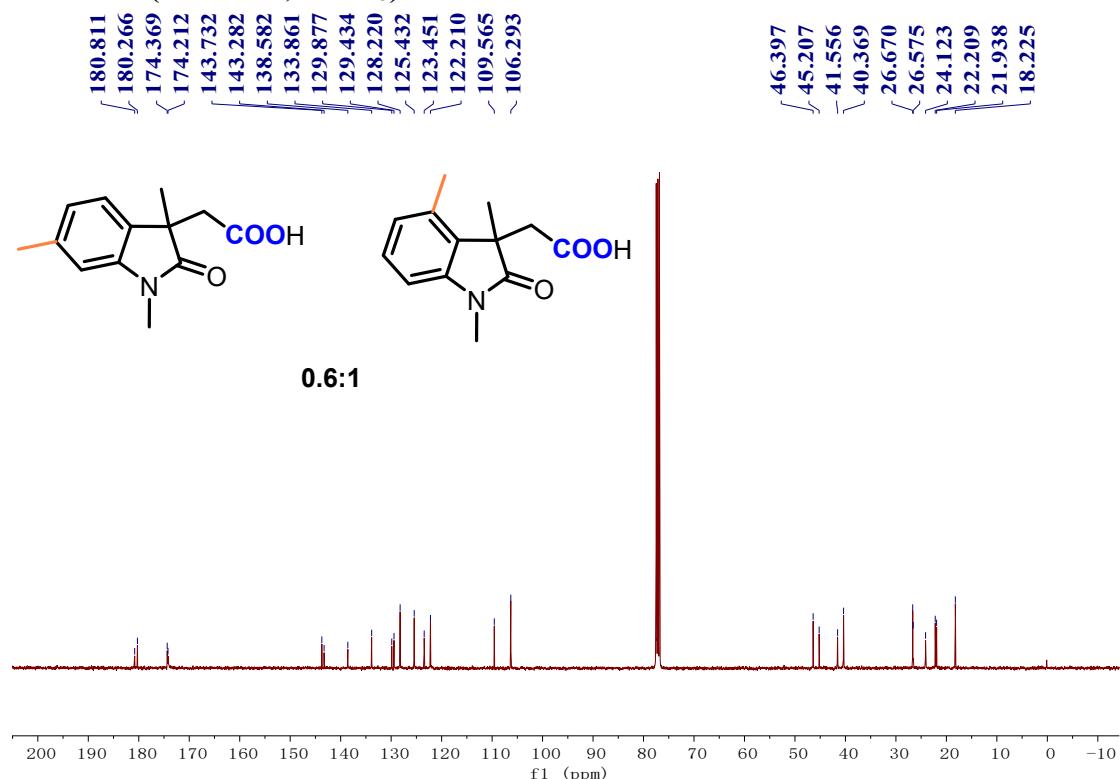
2-(1,3,6-trimethyl-2-oxoindolin-3-yl)acetic acid (2j) and 2-(1,3,4-trimethyl-2-oxoindolin-3-yl)acetic acid (2j') (0.6:1)

^1H NMR (400 MHz, CDCl_3)



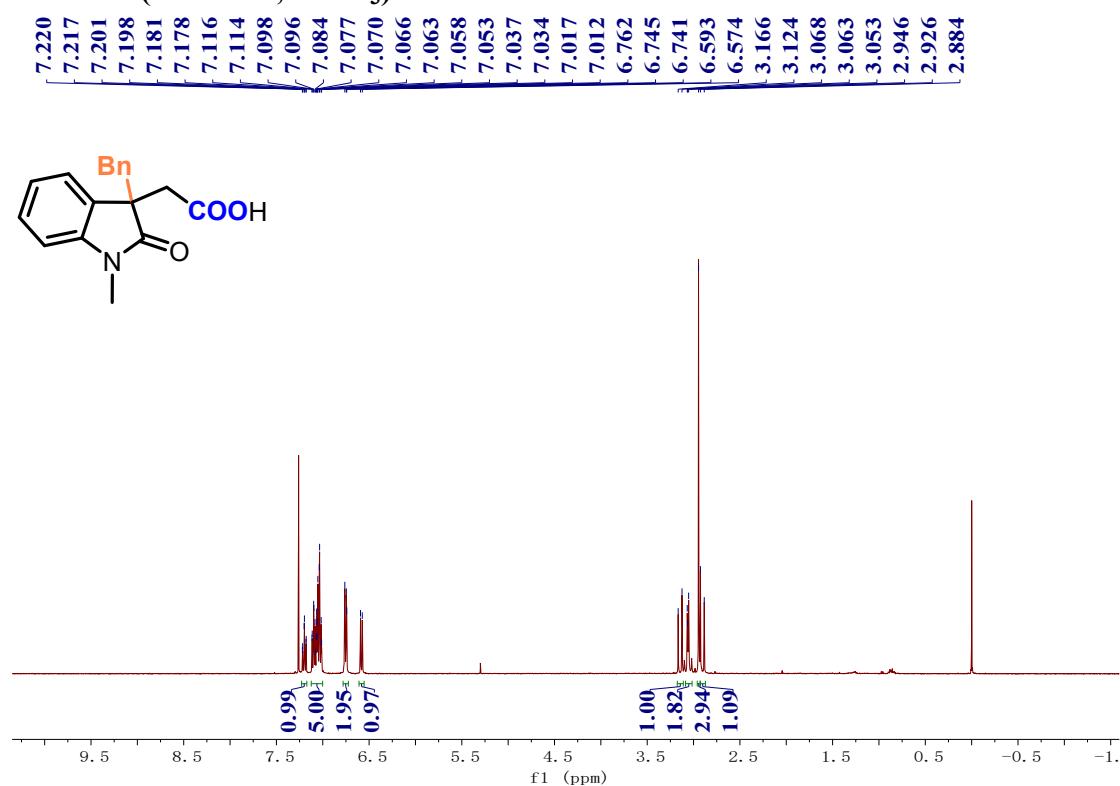
2-(1,3,6-trimethyl-2-oxoindolin-3-yl)acetic acid (2j) and 2-(1,3,4-trimethyl-2-oxoindolin-3-yl)acetic acid (2j') (0.6:1)

¹³C NMR (100 MHz, CDCl₃)



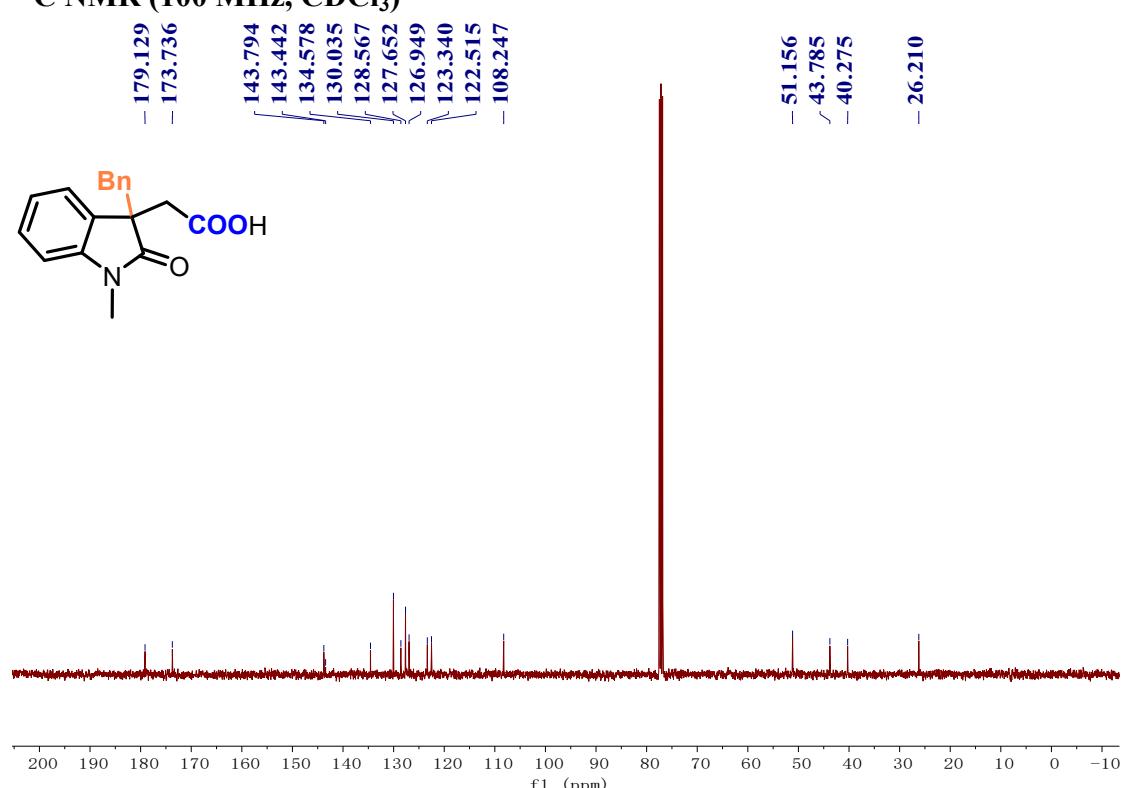
2-(3-benzyl-1-methyl-2-oxoindolin-3-yl)acetic acid (2k)

¹H NMR (400 MHz, CDCl₃)



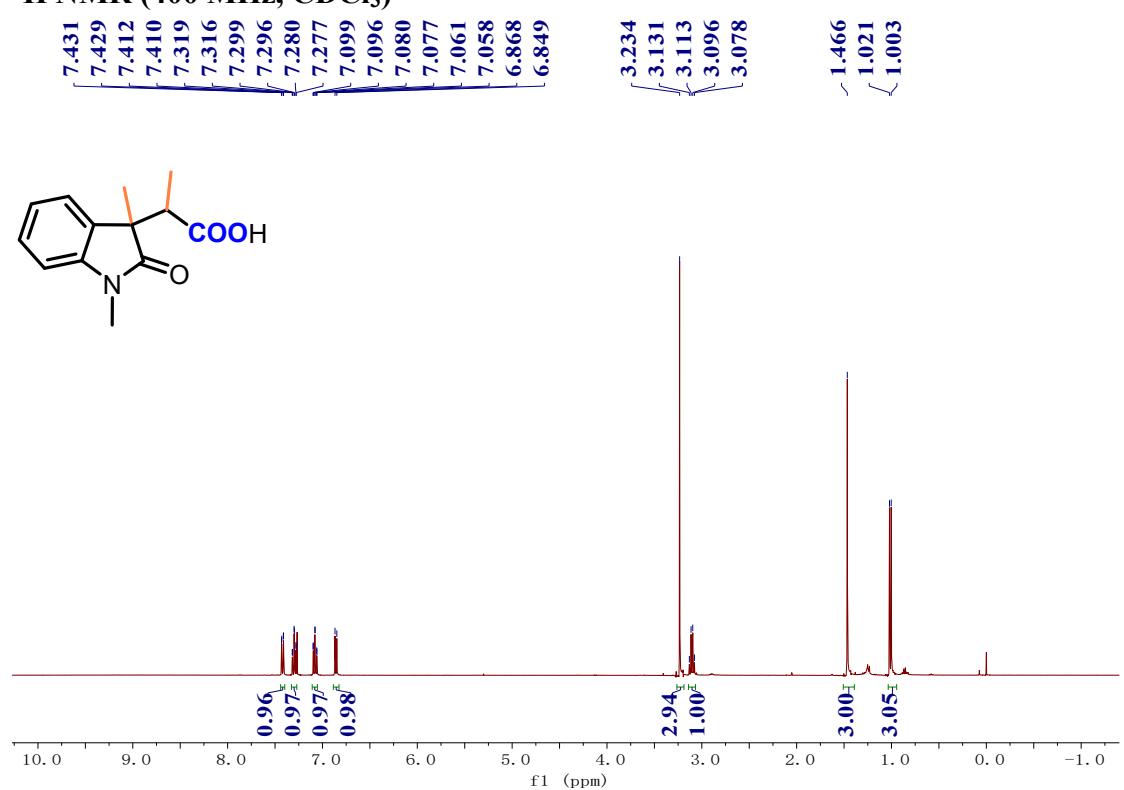
2-(3-benzyl-1-methyl-2-oxoindolin-3-yl)acetic acid (2k)

¹³C NMR (100 MHz, CDCl₃)



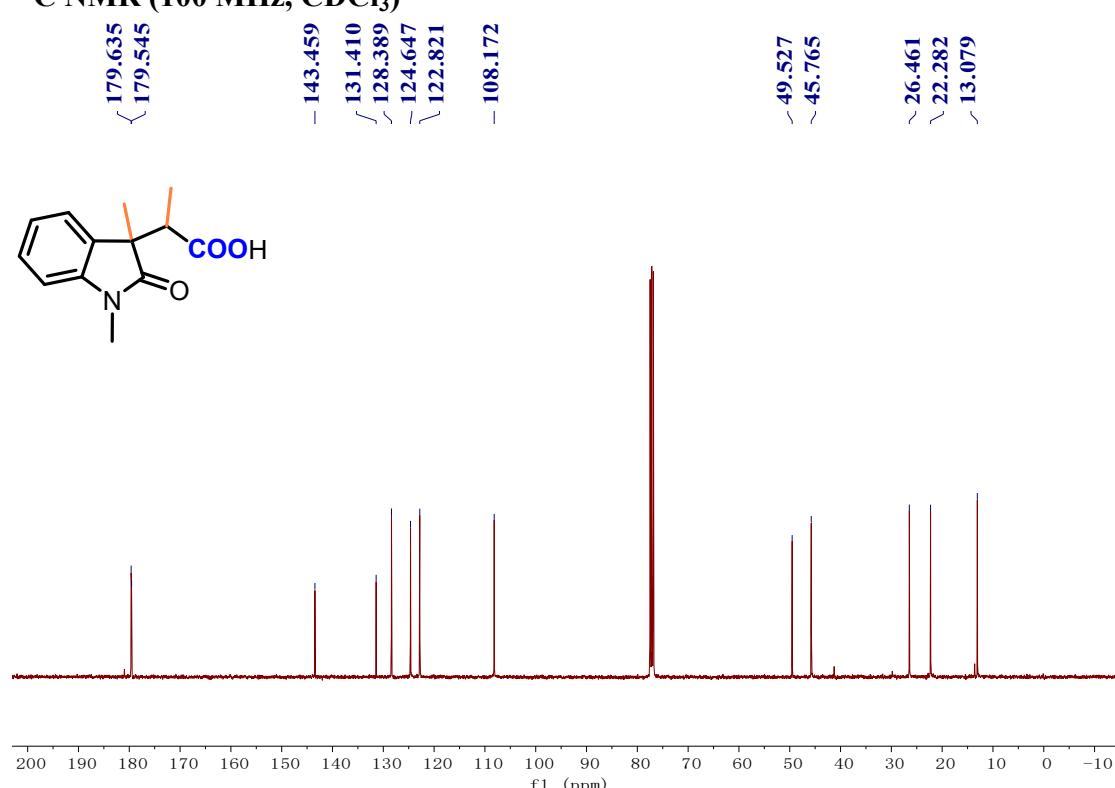
2-(1,3-dimethyl-2-oxoindolin-3-yl)propanoic acid (2l)

¹H NMR (400 MHz, CDCl₃)



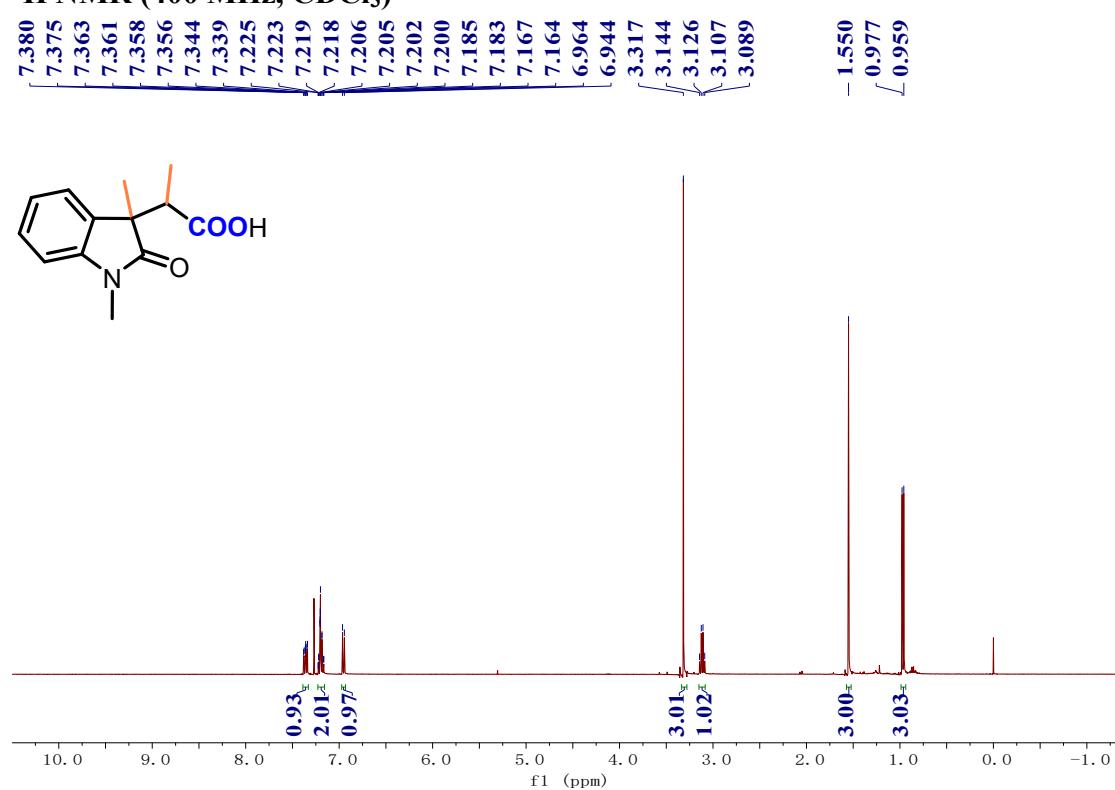
2-(1,3-dimethyl-2-oxoindolin-3-yl)propanoic acid (2l)

¹³C NMR (100 MHz, CDCl₃)



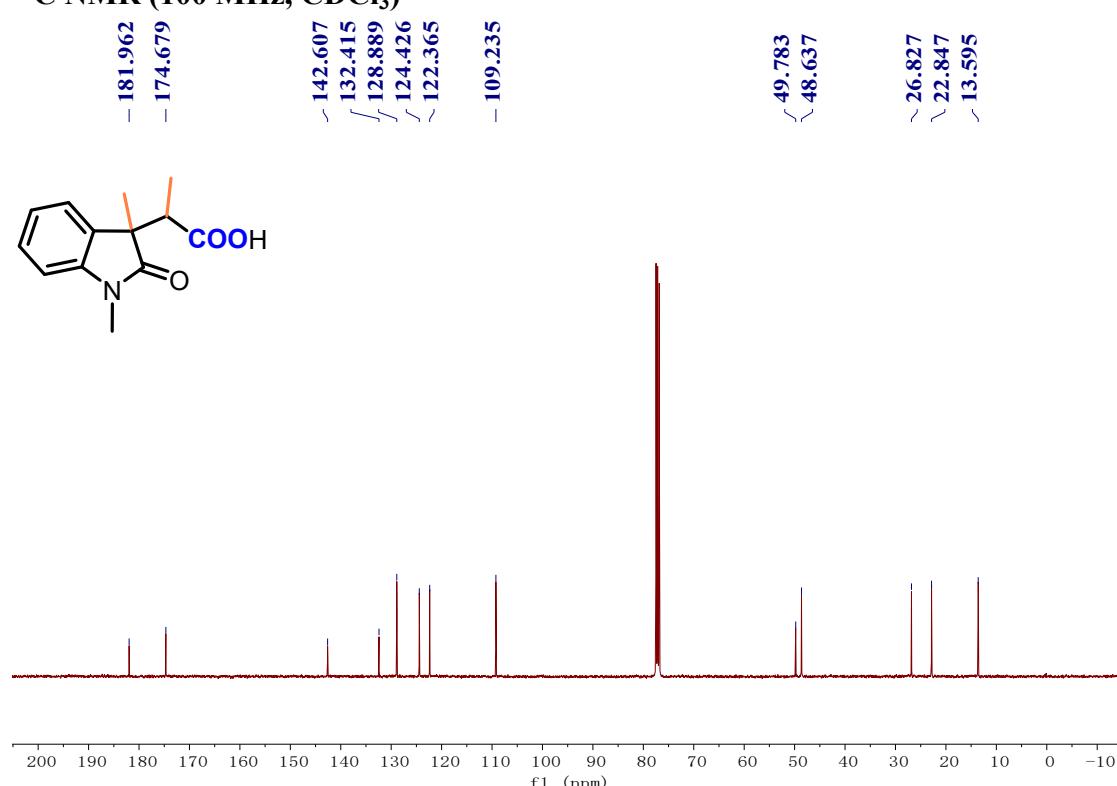
2-(1,3-dimethyl-2-oxoindolin-3-yl)propanoic acid (2l')

¹H NMR (400 MHz, CDCl₃)



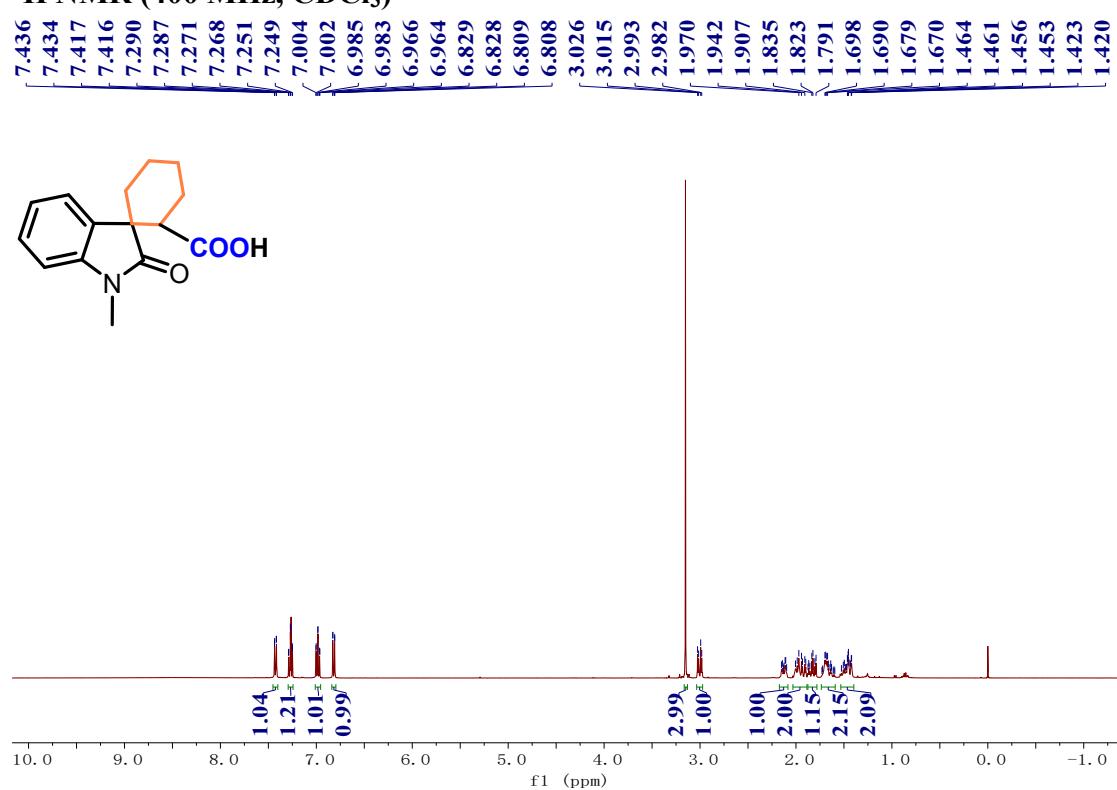
2-(1,3-dimethyl-2-oxoindolin-3-yl)propanoic acid (2l')

¹³C NMR (100 MHz, CDCl₃)



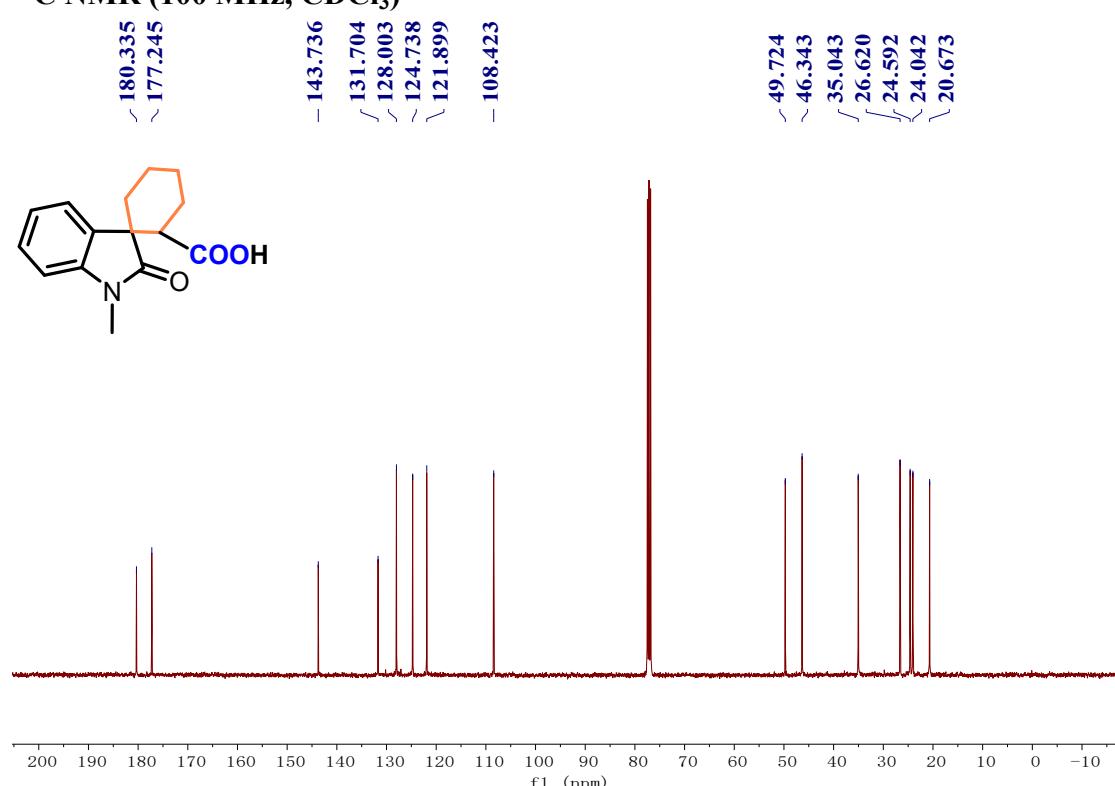
1'-methyl-2'-oxospiro[cyclohexane-1,3'-indoline]-2-carboxylic acid (2m)

¹H NMR (400 MHz, CDCl₃)



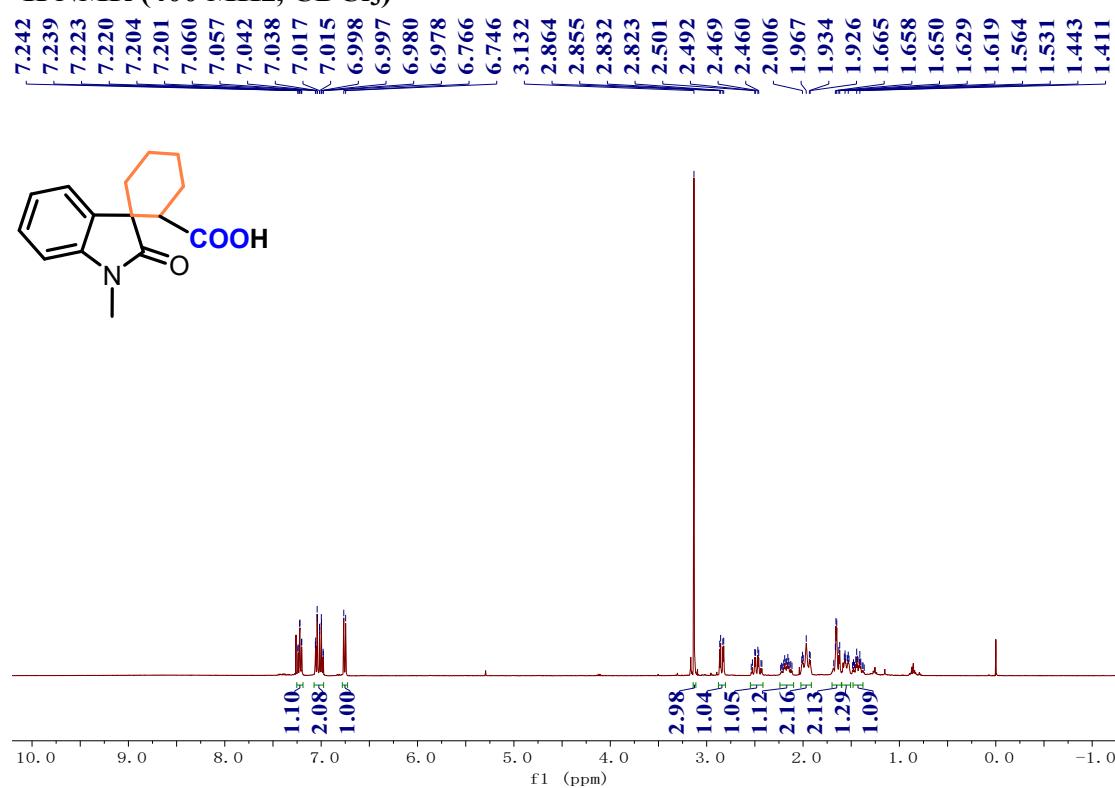
1'-methyl-2'-oxospiro[cyclohexane-1,3'-indoline]-2-carboxylic acid (2m)

^{13}C NMR (100 MHz, CDCl_3)

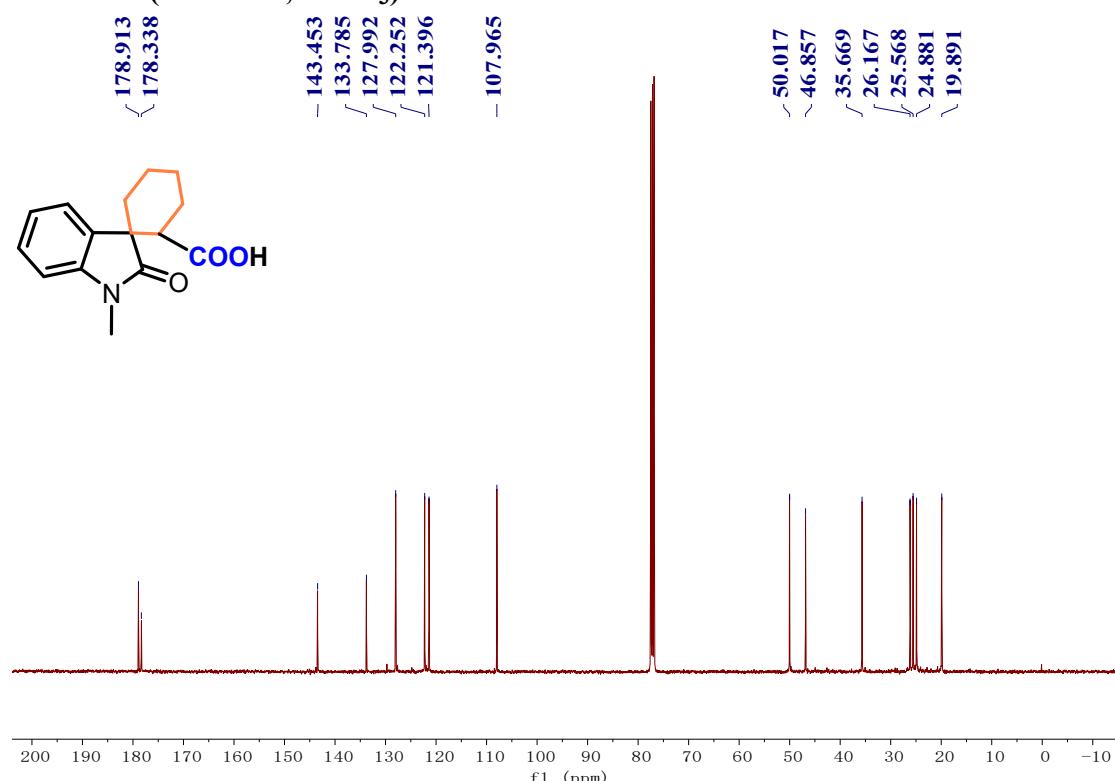


1'-methyl-2'-oxospiro[cyclohexane-1,3'-indoline]-2-carboxylic acid (2m')

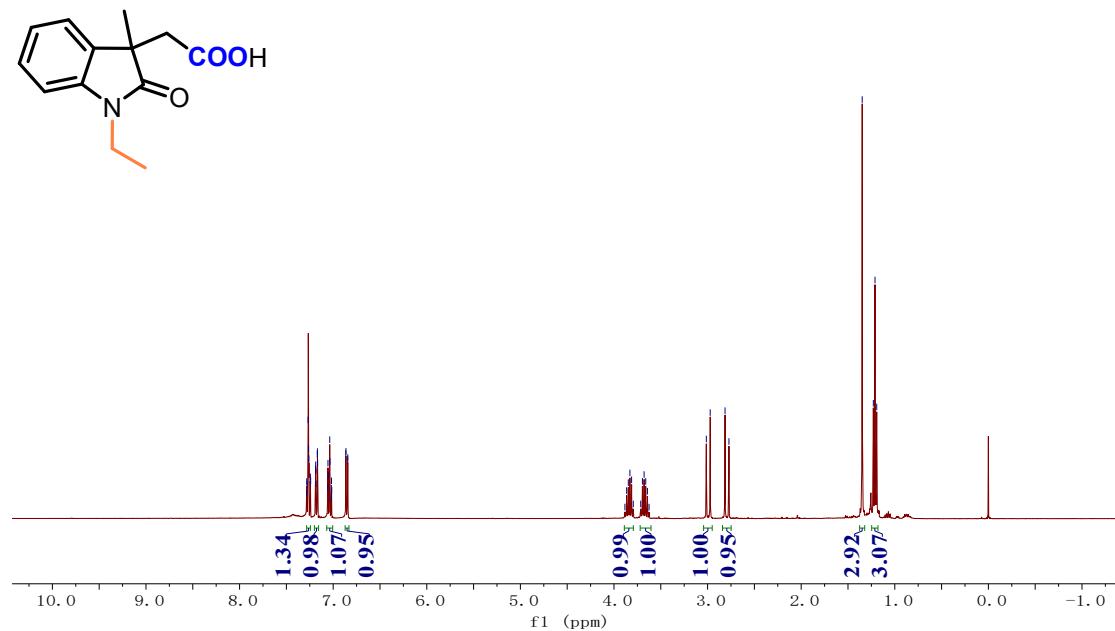
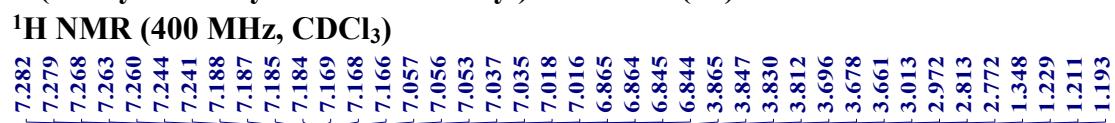
^1H NMR (400 MHz, CDCl_3)



1'-methyl-2'-oxospiro[cyclohexane-1,3'-indoline]-2-carboxylic acid (2m')
¹³C NMR (100 MHz, CDCl₃)

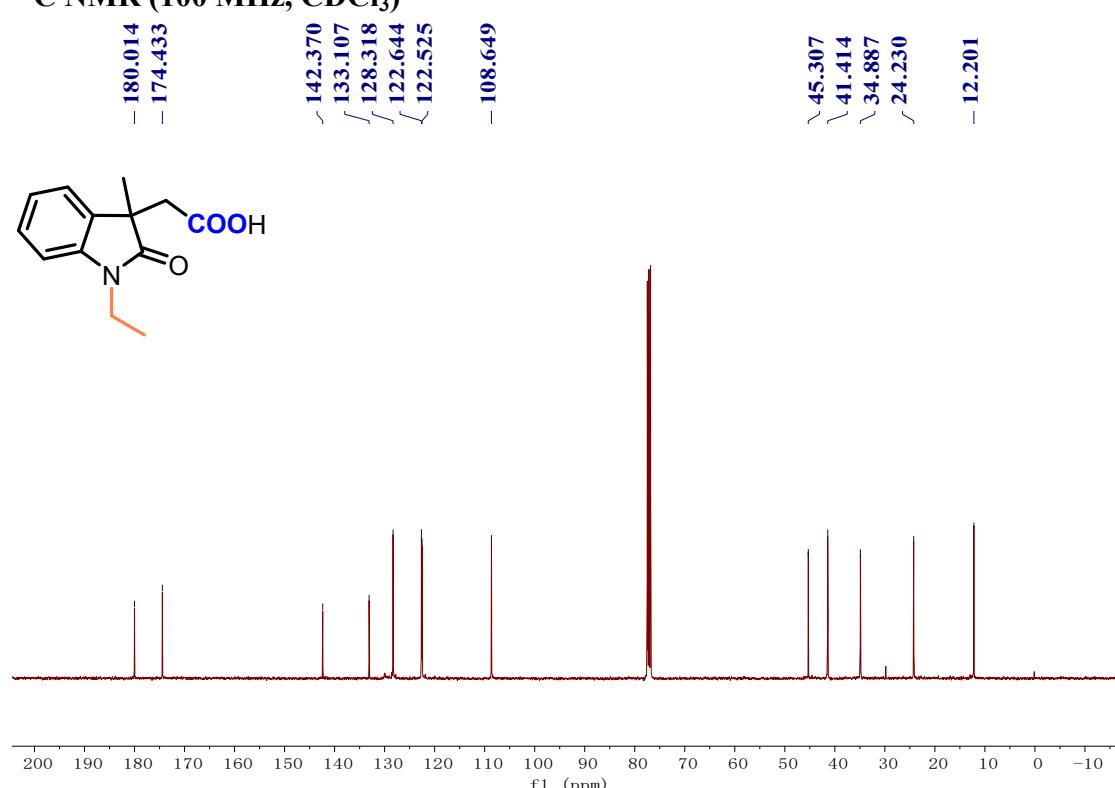


2-(1-ethyl-3-methyl-2-oxoindolin-3-yl)acetic acid (2n)



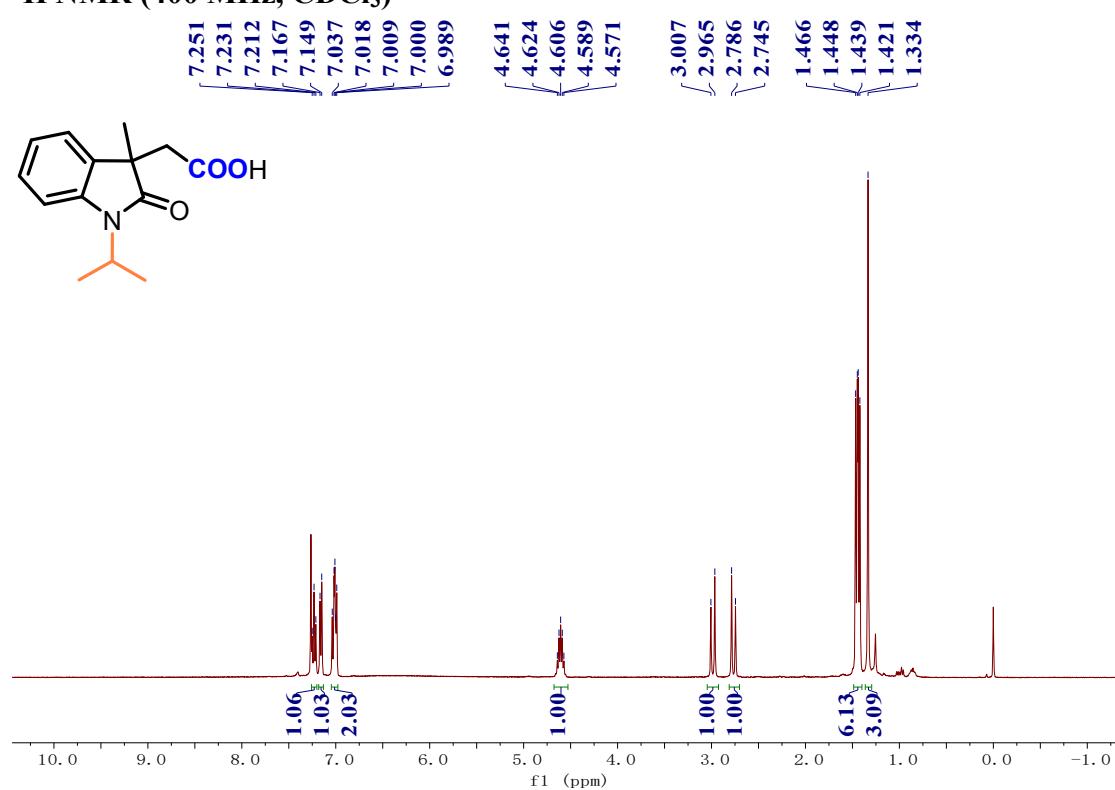
2-(1-ethyl-3-methyl-2-oxoindolin-3-yl)acetic acid (2n)

¹³C NMR (100 MHz, CDCl₃)



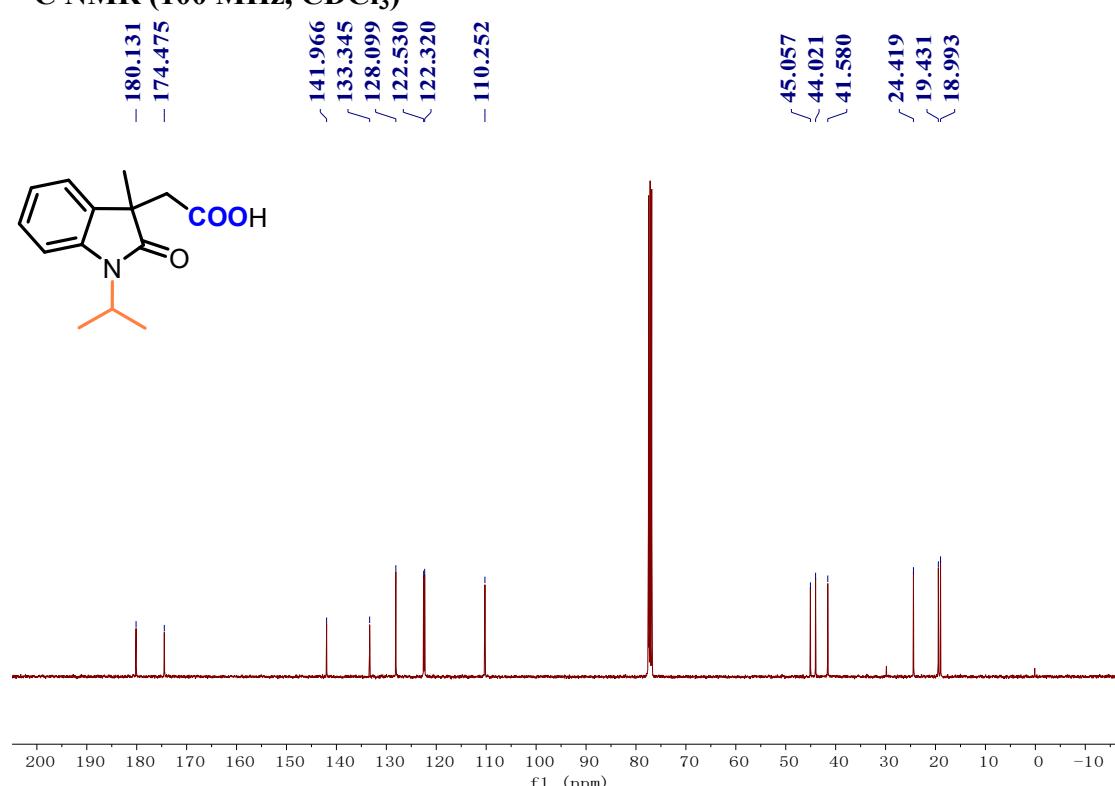
2-(1-isopropyl-3-methyl-2-oxoindolin-3-yl)acetic acid (2o)

¹H NMR (400 MHz, CDCl₃)



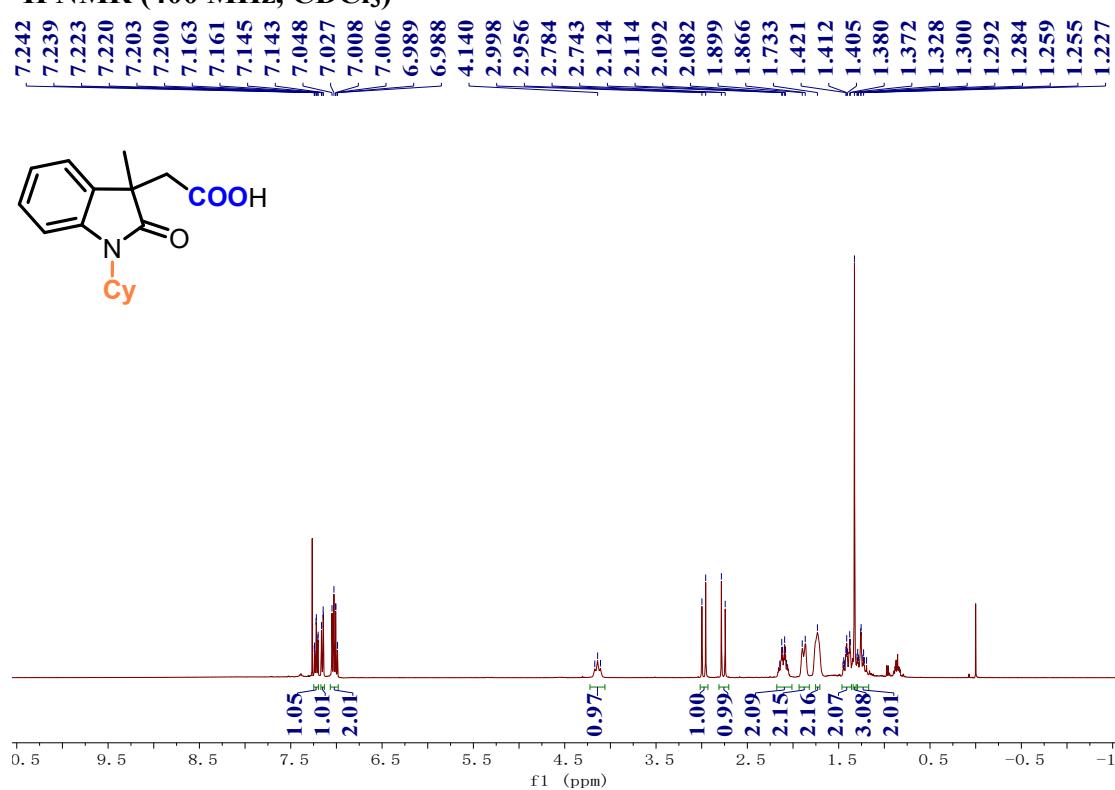
2-(1-isopropyl-3-methyl-2-oxoindolin-3-yl)acetic acid (2o)

¹³C NMR (100 MHz, CDCl₃)



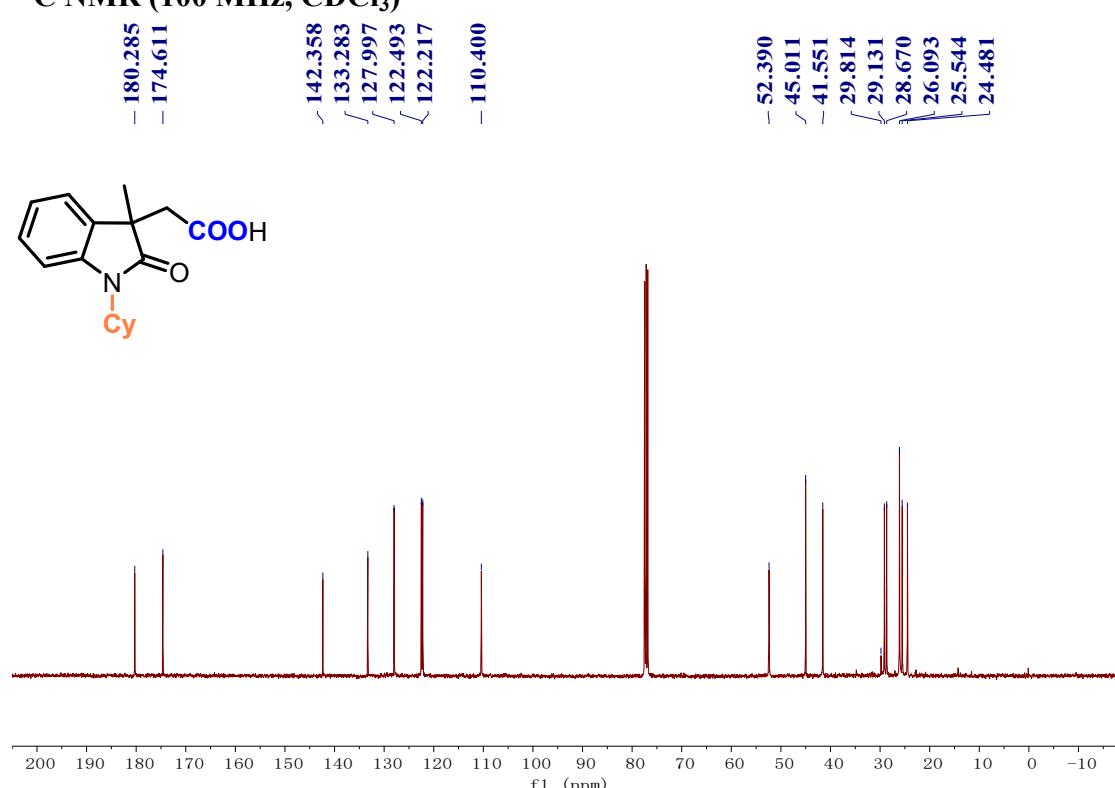
2-(1-cyclohexyl-3-methyl-2-oxoindolin-3-yl)acetic acid (2p)

¹H NMR (400 MHz, CDCl₃)



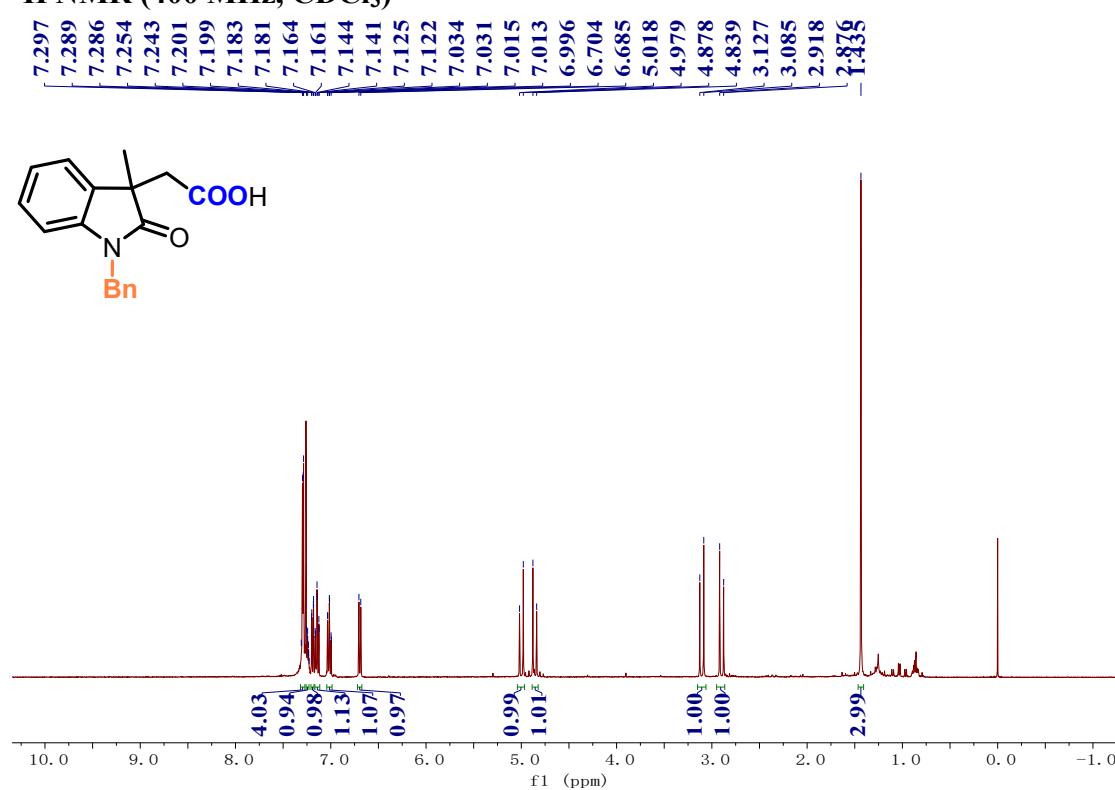
2-(1-cyclohexyl-3-methyl-2-oxoindolin-3-yl)acetic acid (2p)

¹³C NMR (100 MHz, CDCl₃)



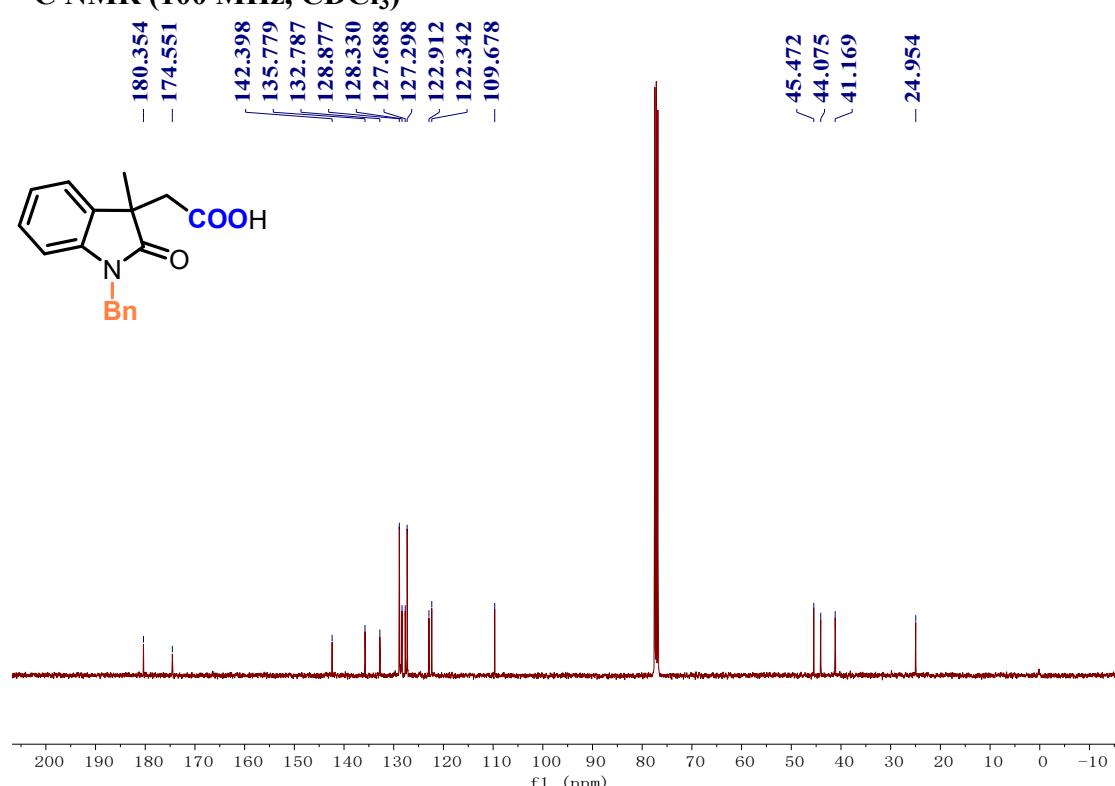
2-(1-benzyl-3-methyl-2-oxoindolin-3-yl)acetic acid (2q)

¹H NMR (400 MHz, CDCl₃)



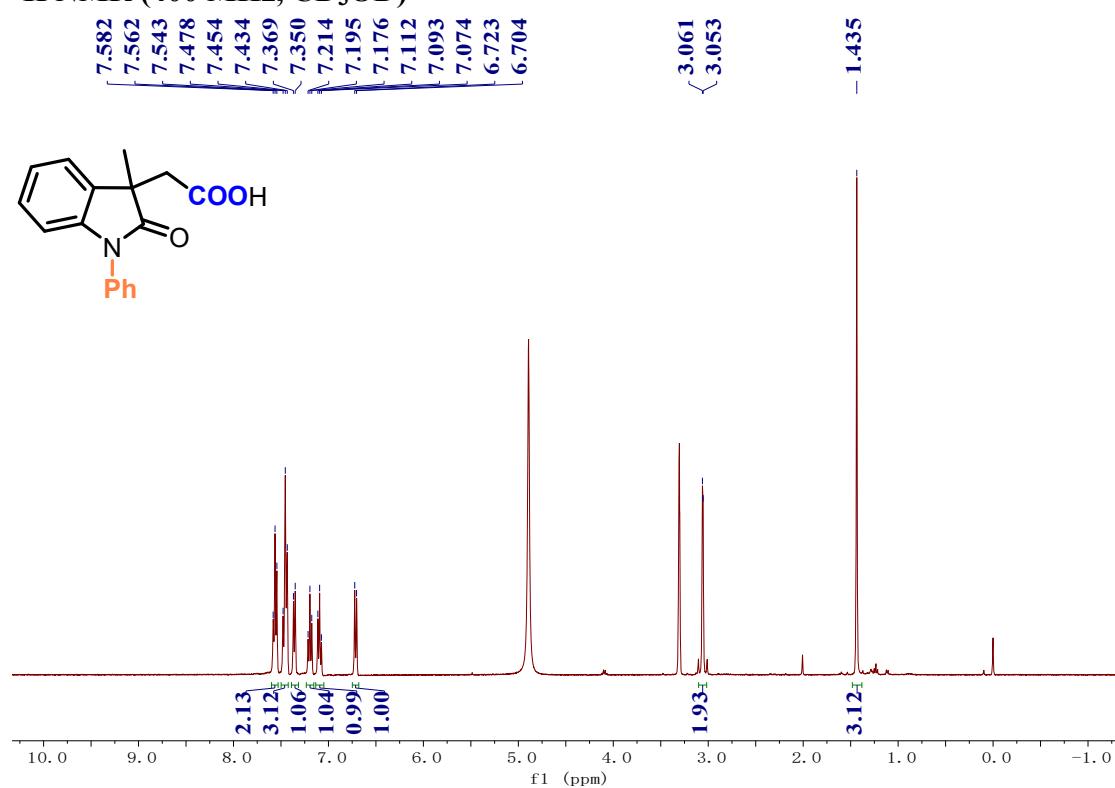
2-(1-benzyl-3-methyl-2-oxoindolin-3-yl)acetic acid (2q)

¹³C NMR (100 MHz, CDCl₃)



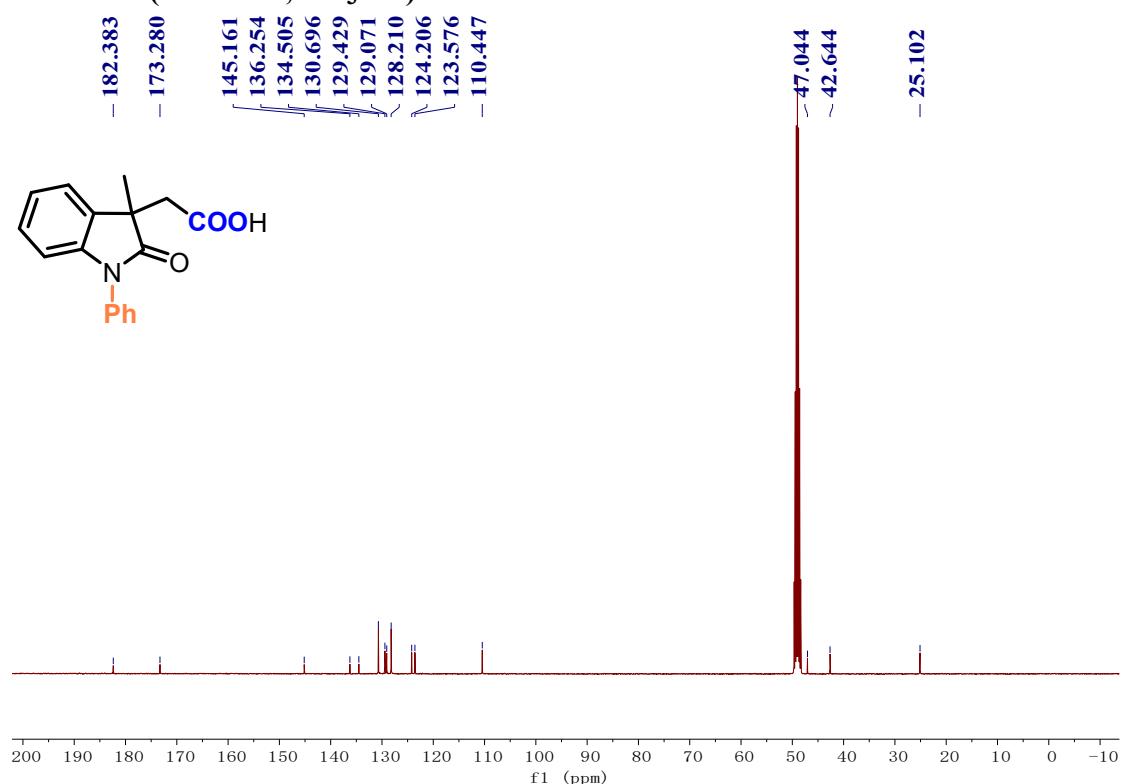
2-(3-methyl-2-oxo-1-phenylindolin-3-yl)acetic acid (2r)

¹H NMR (400 MHz, CD₃OD)



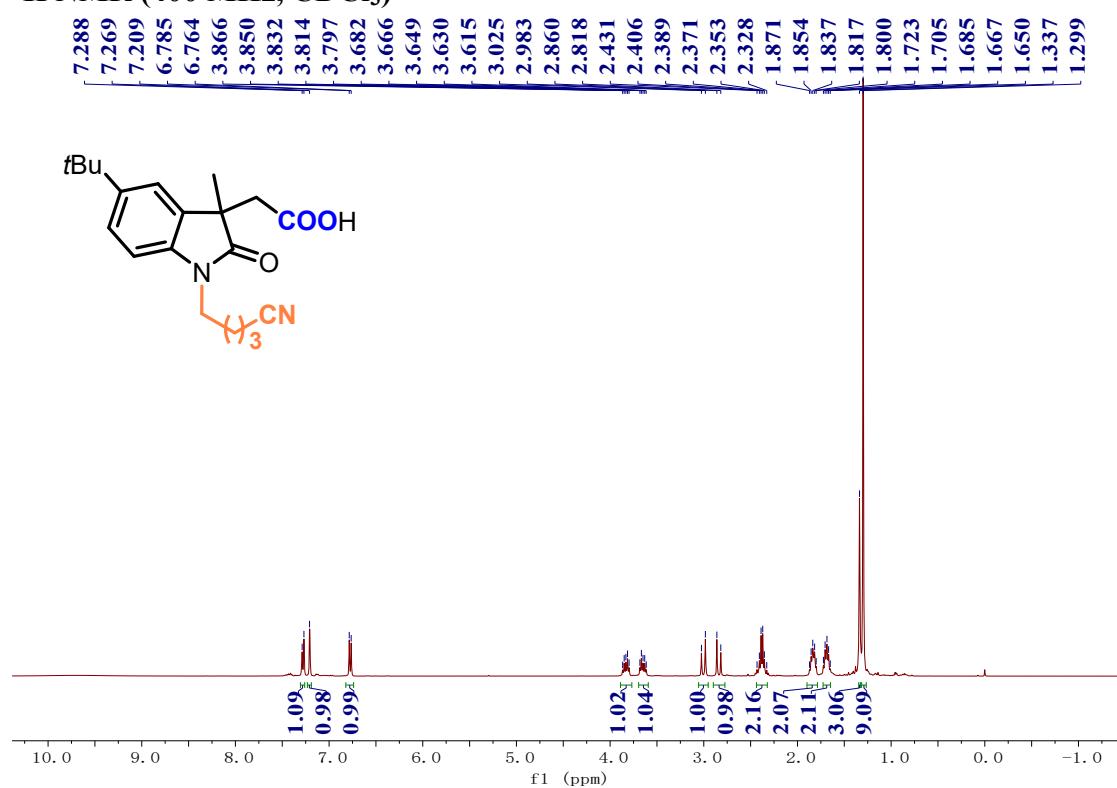
2-(3-methyl-2-oxo-1-phenylindolin-3-yl)acetic acid (2r)

¹³C NMR (100 MHz, CD₃OD)



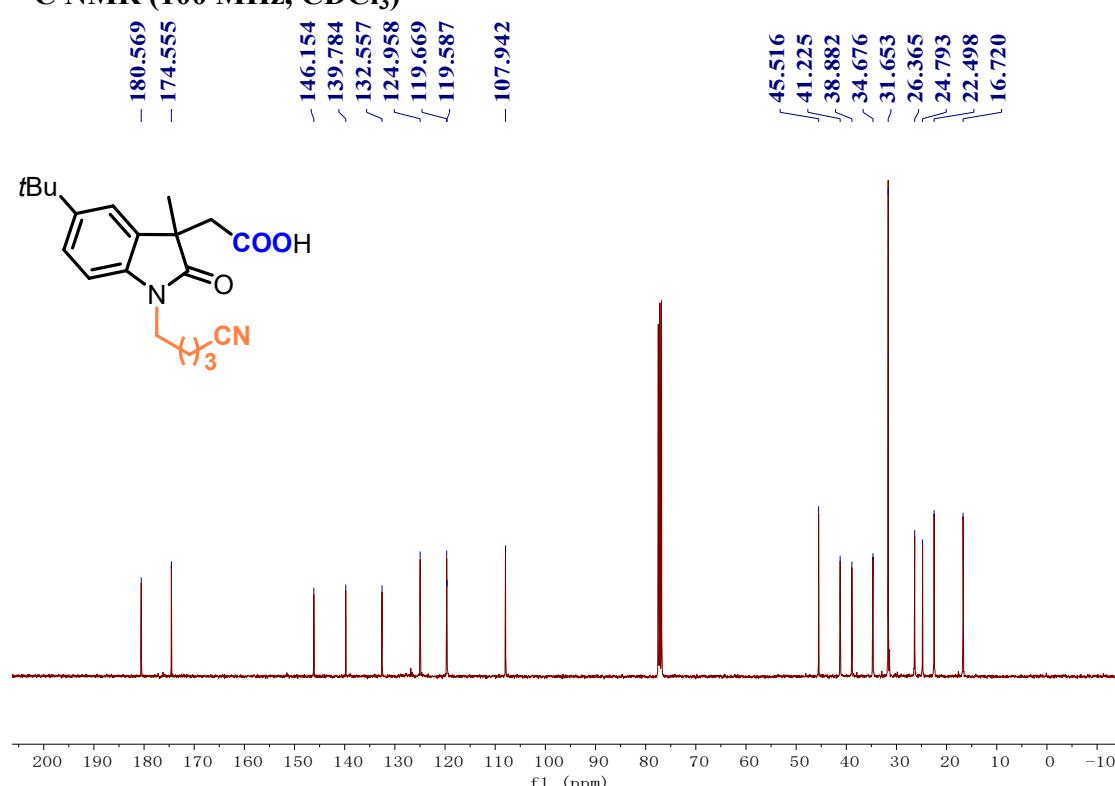
2-(5-(tert-butyl)-1-(4-cyanobutyl)-3-methyl-2-oxoindolin-3-yl)acetic acid (2s)

¹H NMR (400 MHz, CDCl₃)



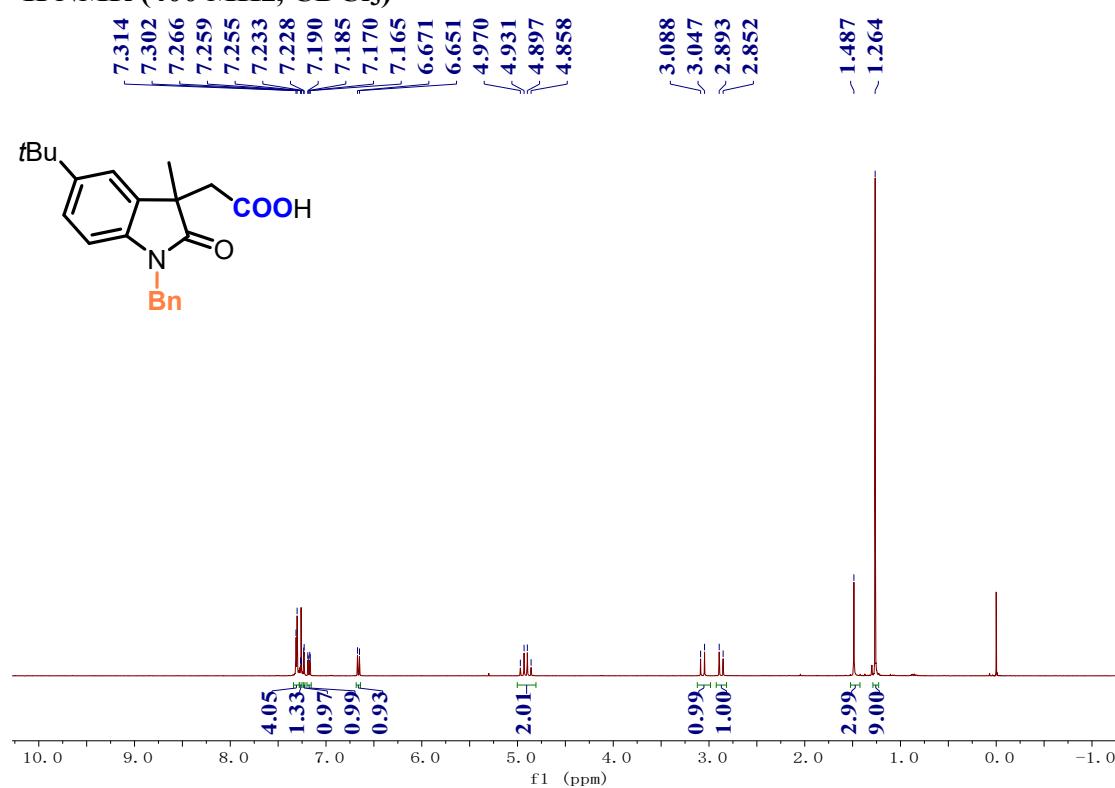
2-(5-(*tert*-butyl)-1-(4-cyanobutyl)-3-methyl-2-oxoindolin-3-yl)acetic acid (2s)

¹³C NMR (100 MHz, CDCl₃)

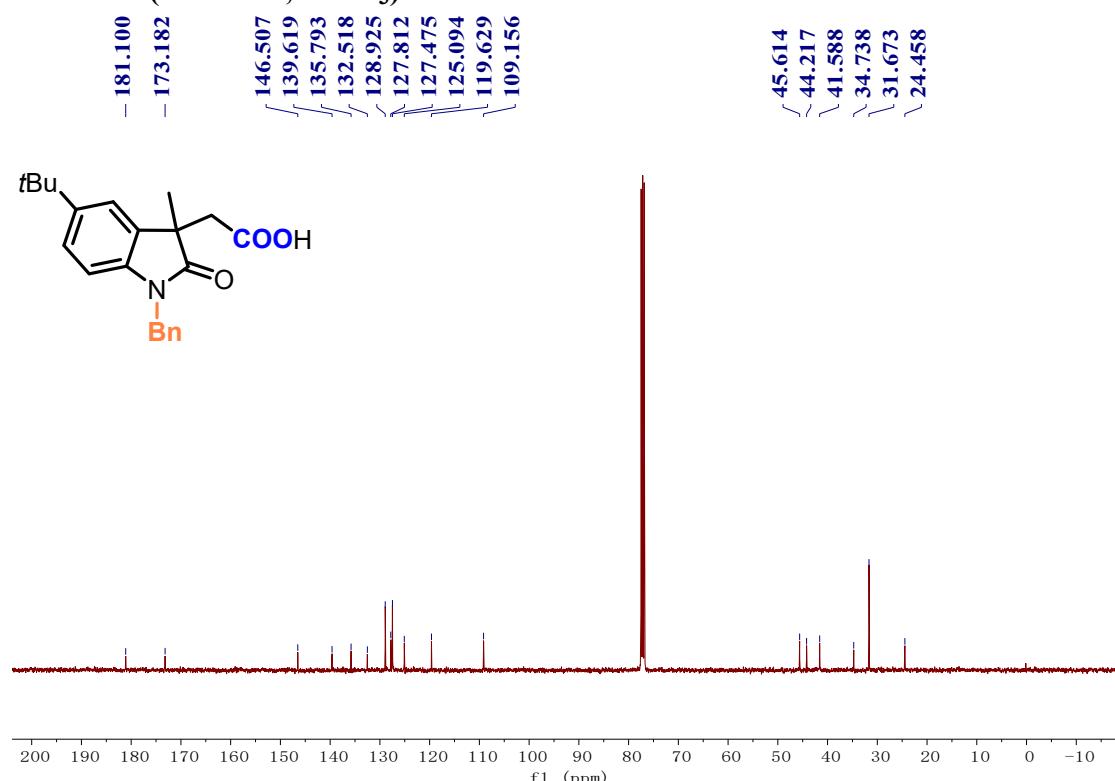


2-(1-benzyl-5-(*tert*-butyl)-3-methyl-2-oxoindolin-3-yl)acetic acid (2t)

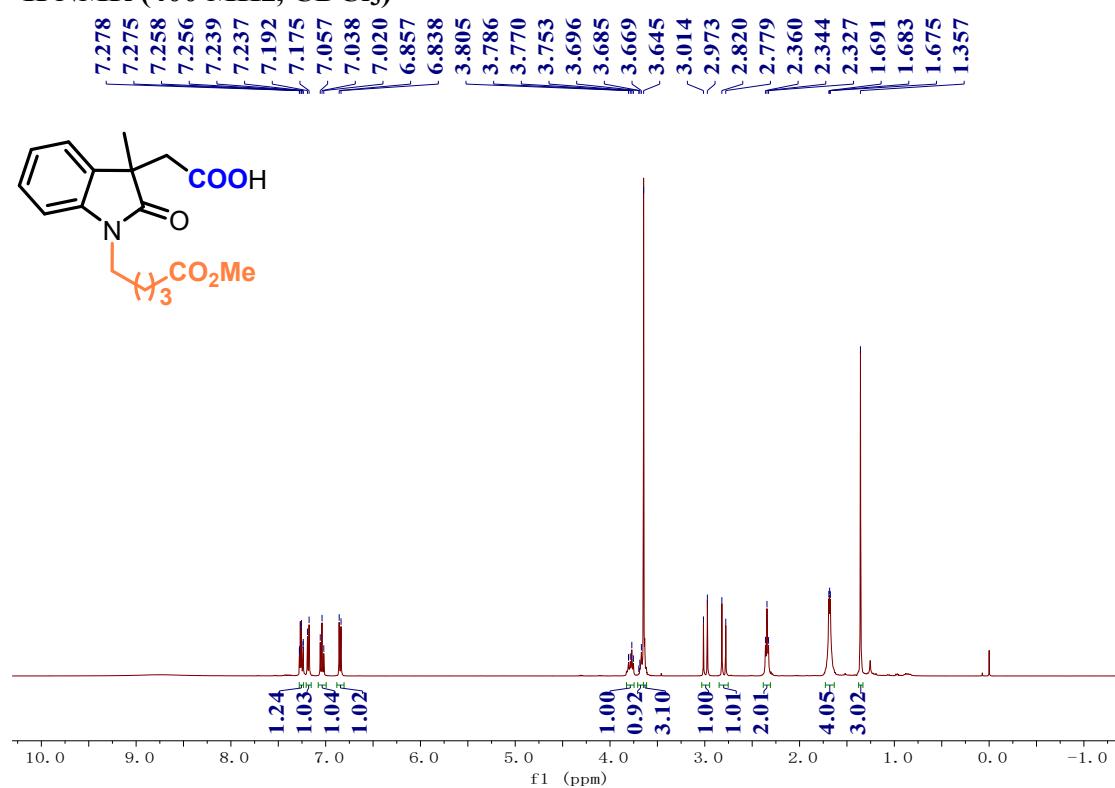
¹H NMR (400 MHz, CDCl₃)



2-(1-benzyl-5-(*tert*-butyl)-3-methyl-2-oxoindolin-3-yl)acetic acid (2t)
¹³C NMR (100 MHz, CDCl₃)

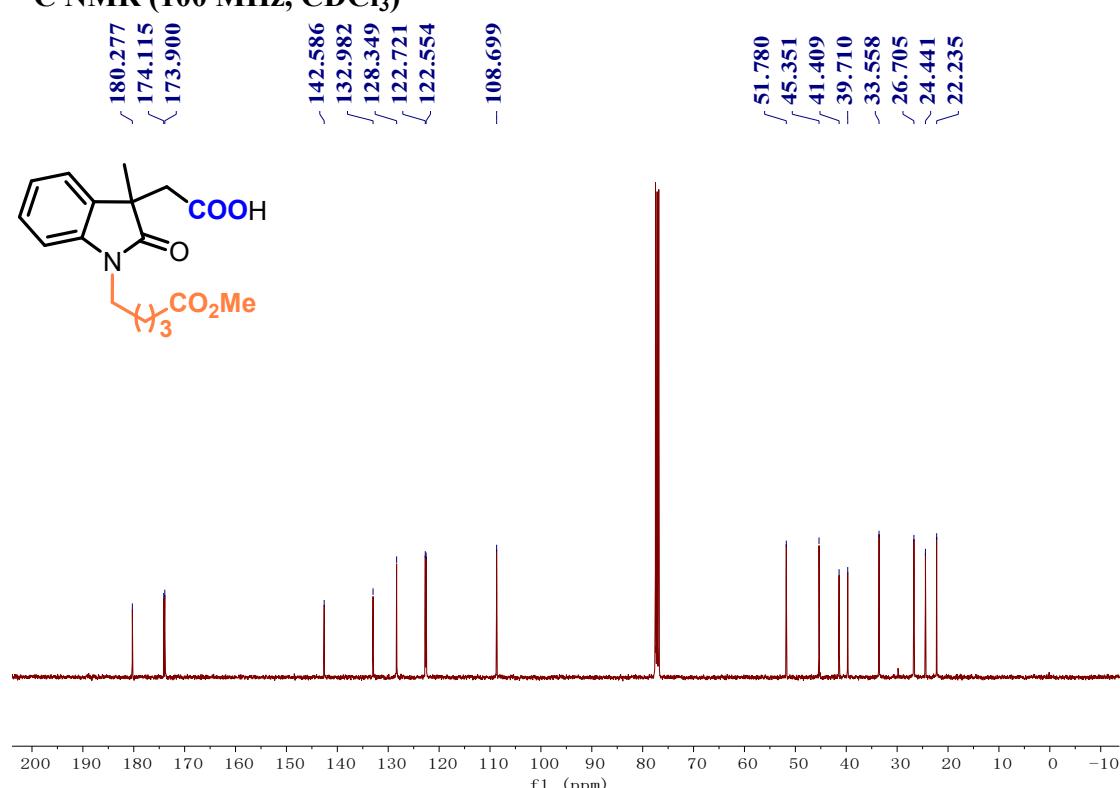


2-(1-(5-methoxy-5-oxopentyl)-3-methyl-2-oxoindolin-3-yl)acetic acid (2u)
¹H NMR (400 MHz, CDCl₃)



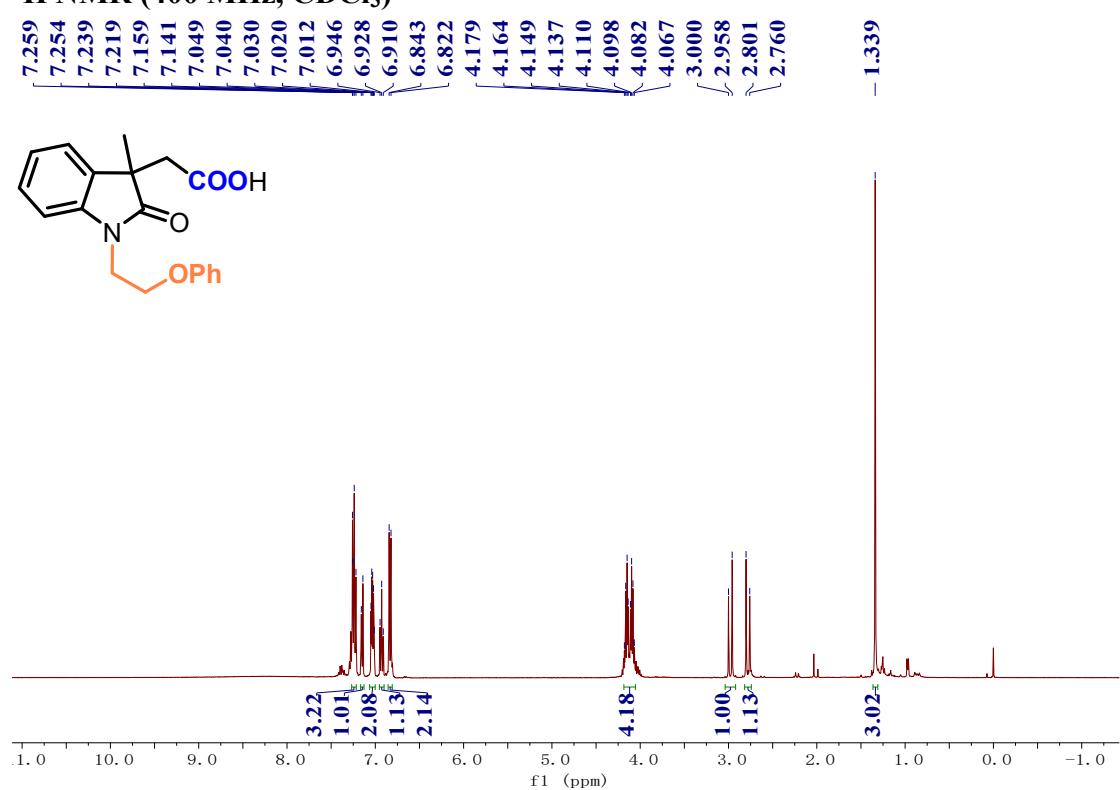
2-(1-(5-methoxy-5-oxopentyl)-3-methyl-2-oxoindolin-3-yl)acetic acid (2u)

¹³C NMR (100 MHz, CDCl₃)



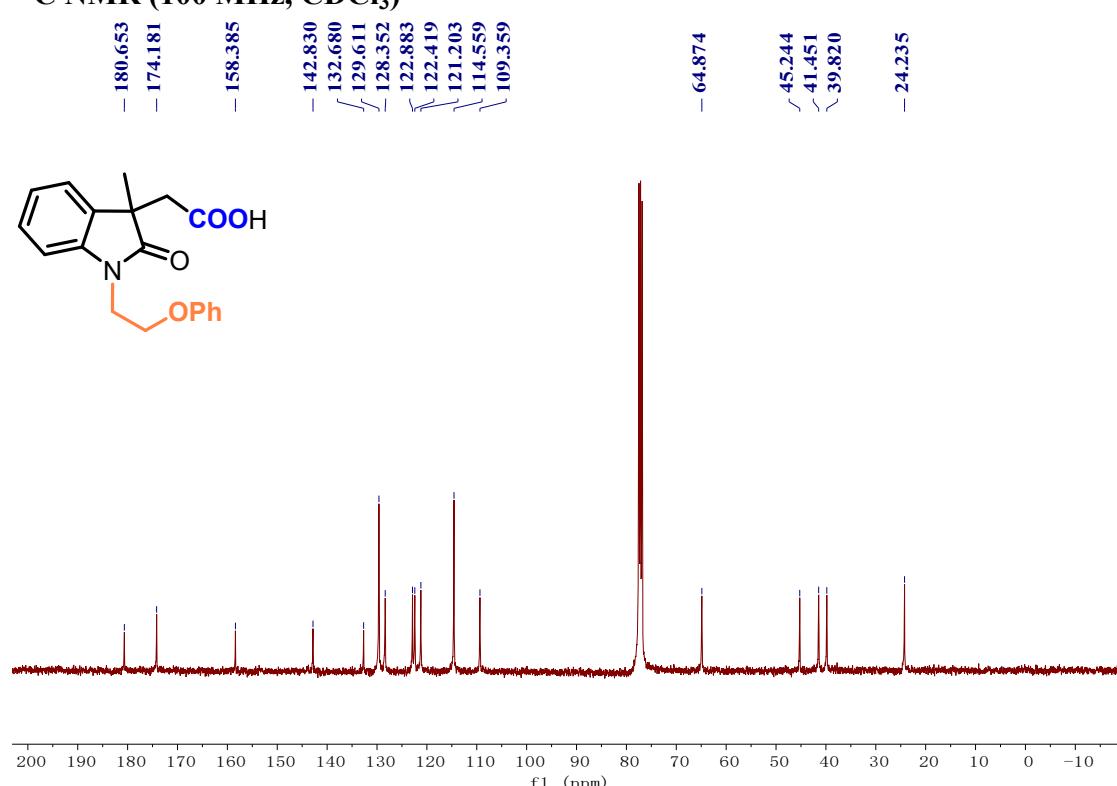
2-(3-methyl-2-oxo-1-(2-phenoxybutyl)indolin-3-yl)acetic acid (2v)

¹H NMR (400 MHz, CDCl₃)



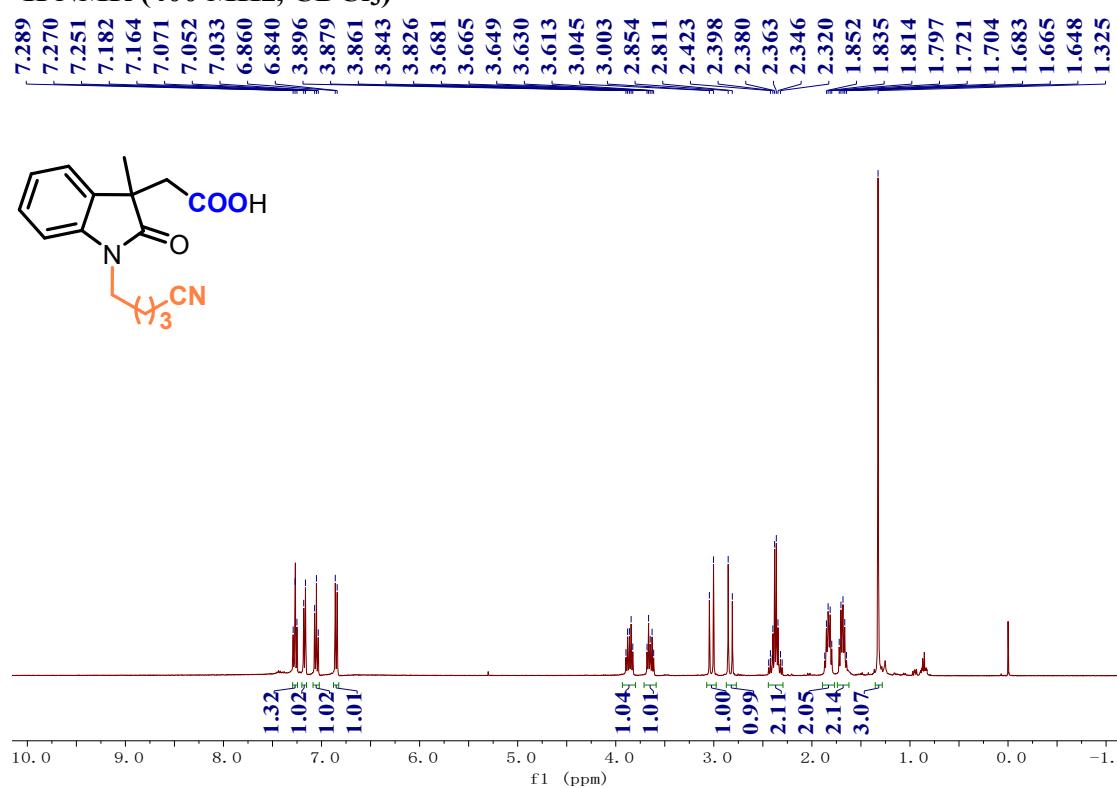
2-(3-methyl-2-oxo-1-(2-phenoxybutyl)indolin-3-yl)acetic acid (2v)

¹³C NMR (100 MHz, CDCl₃)



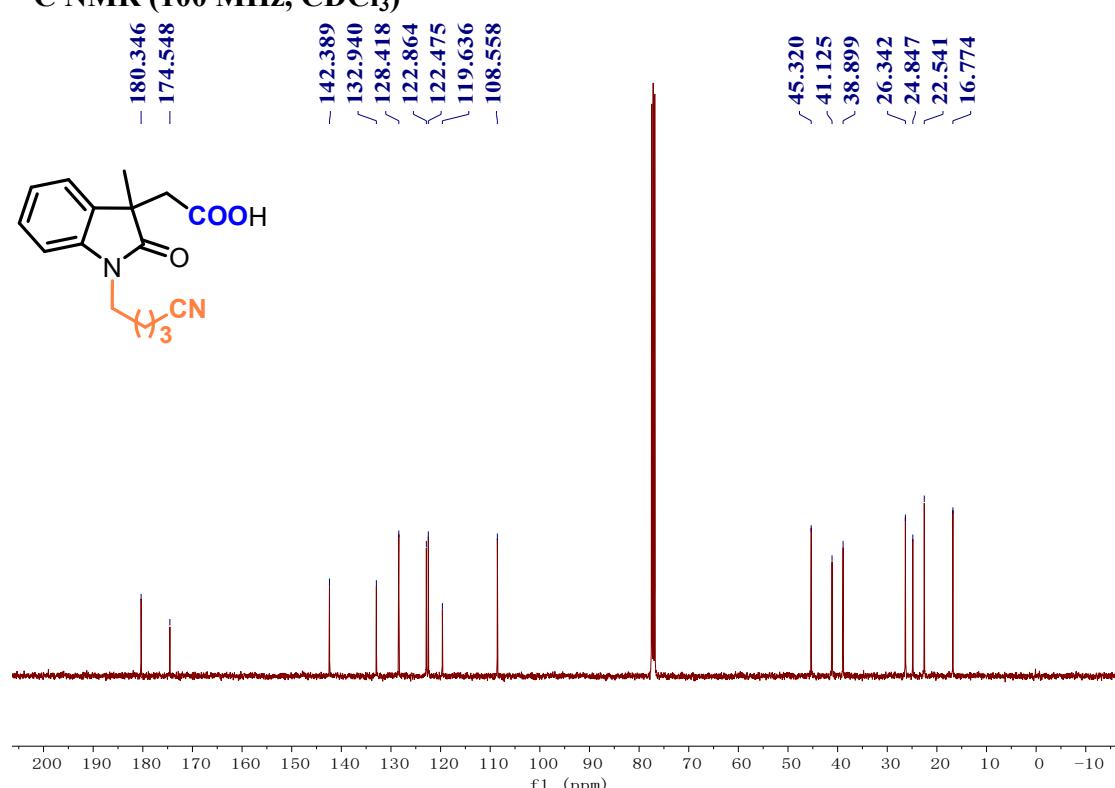
2-(1-(4-cyanobutyl)-3-methyl-2-oxoindolin-3-yl)acetic acid (2w)

¹H NMR (400 MHz, CDCl₃)



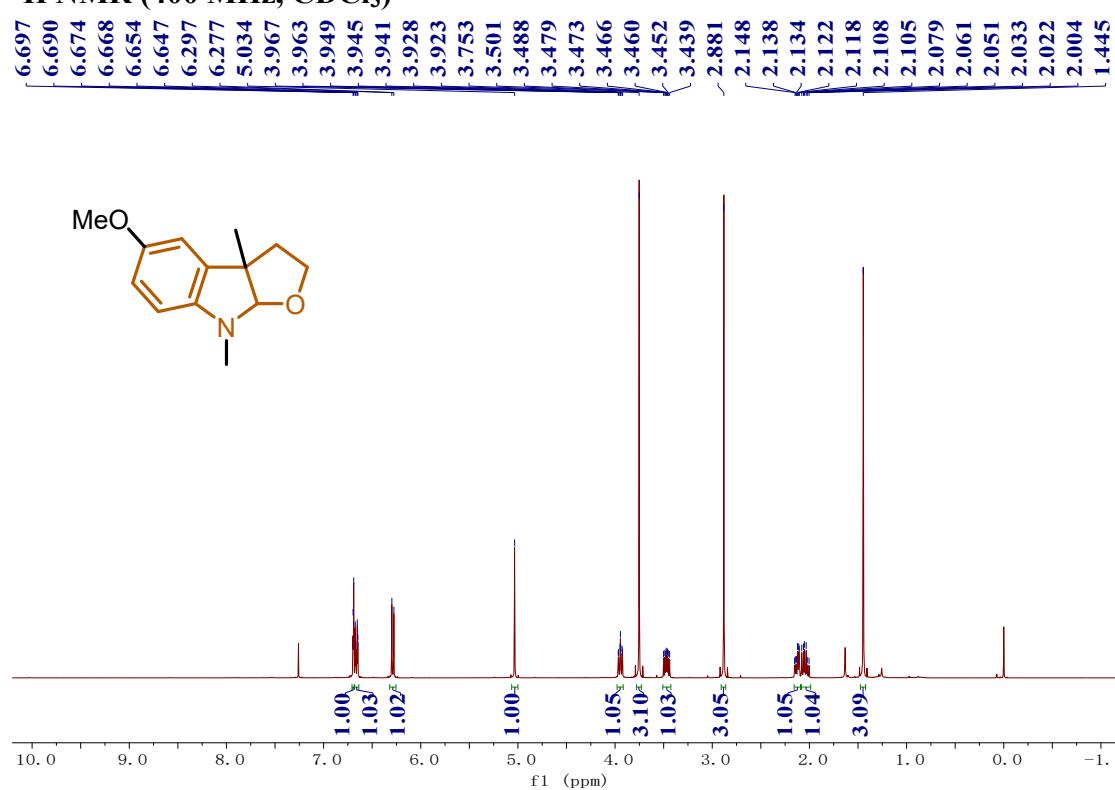
2-(1-(4-cyanobutyl)-3-methyl-2-oxoindolin-3-yl)acetic acid (2w)

¹³C NMR (100 MHz, CDCl₃)



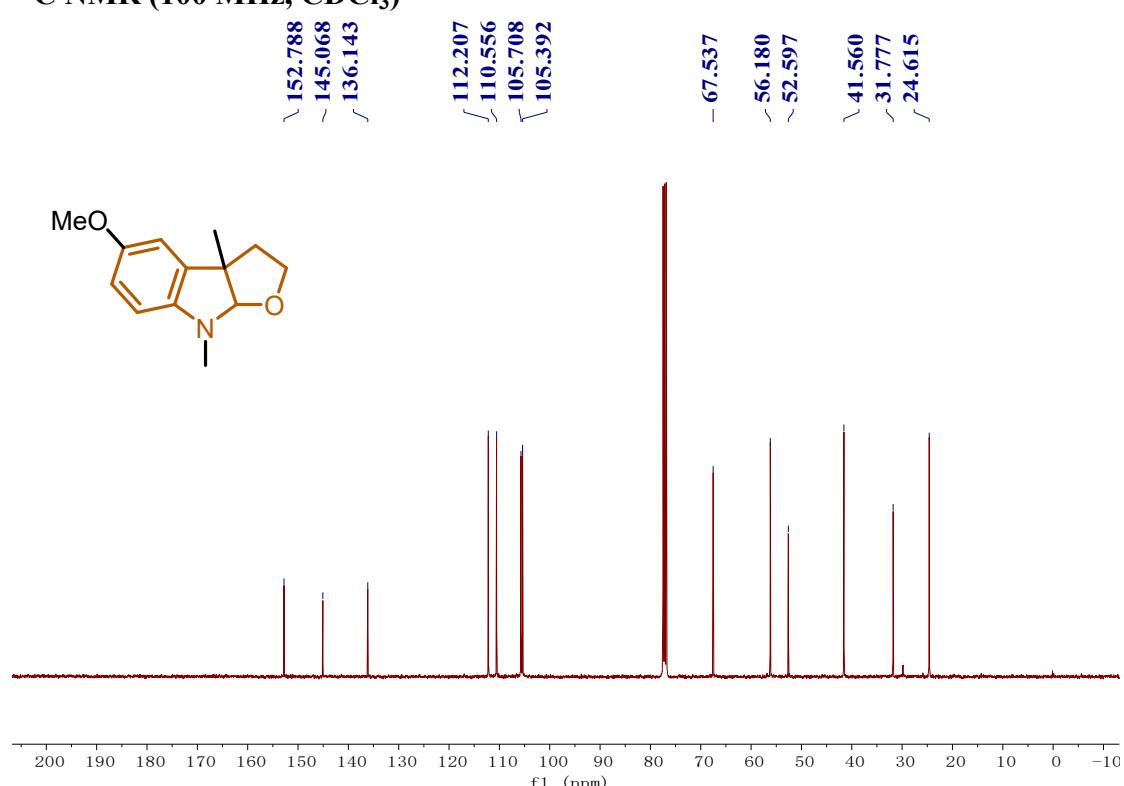
5-methoxy-3a,8-dimethyl-3,3a,8,8a-tetrahydro-2*H*-furo[2,3-*b*]indole (3)

¹H NMR (400 MHz, CDCl₃)



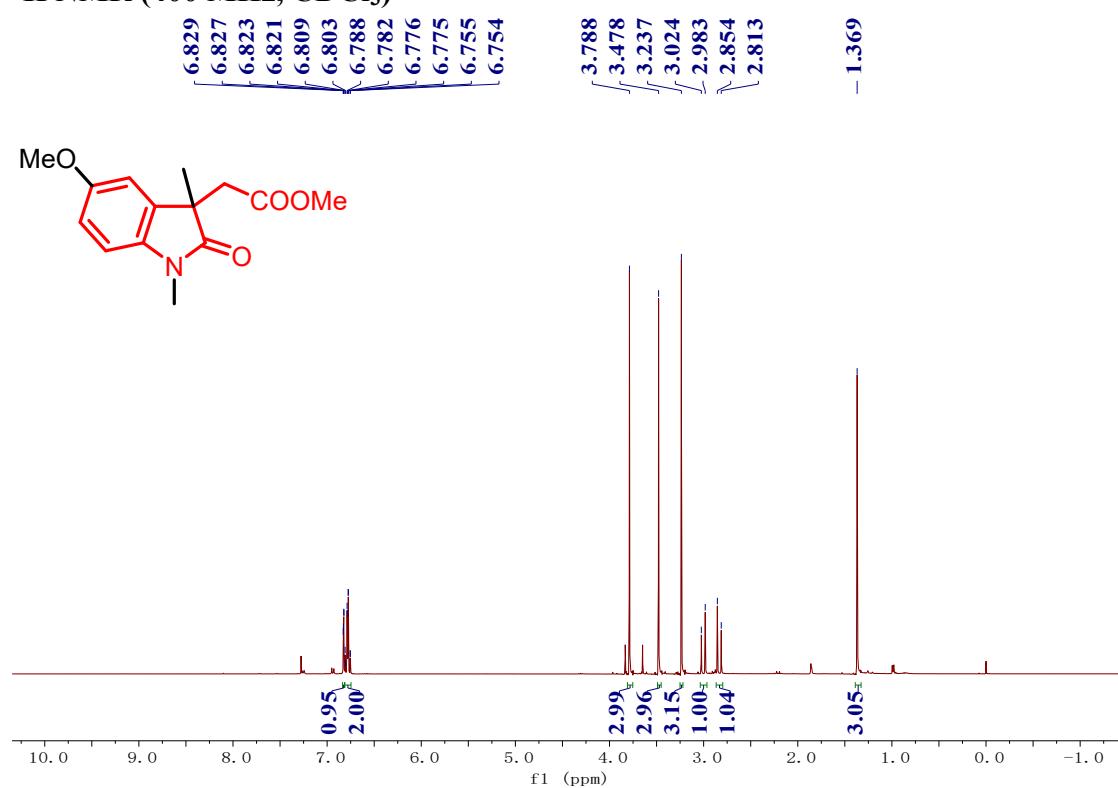
5-methoxy-3a,8-dimethyl-3,3a,8,8a-tetrahydro-2*H*-furo[2,3-*b*]indole (3)

¹³C NMR (100 MHz, CDCl₃)



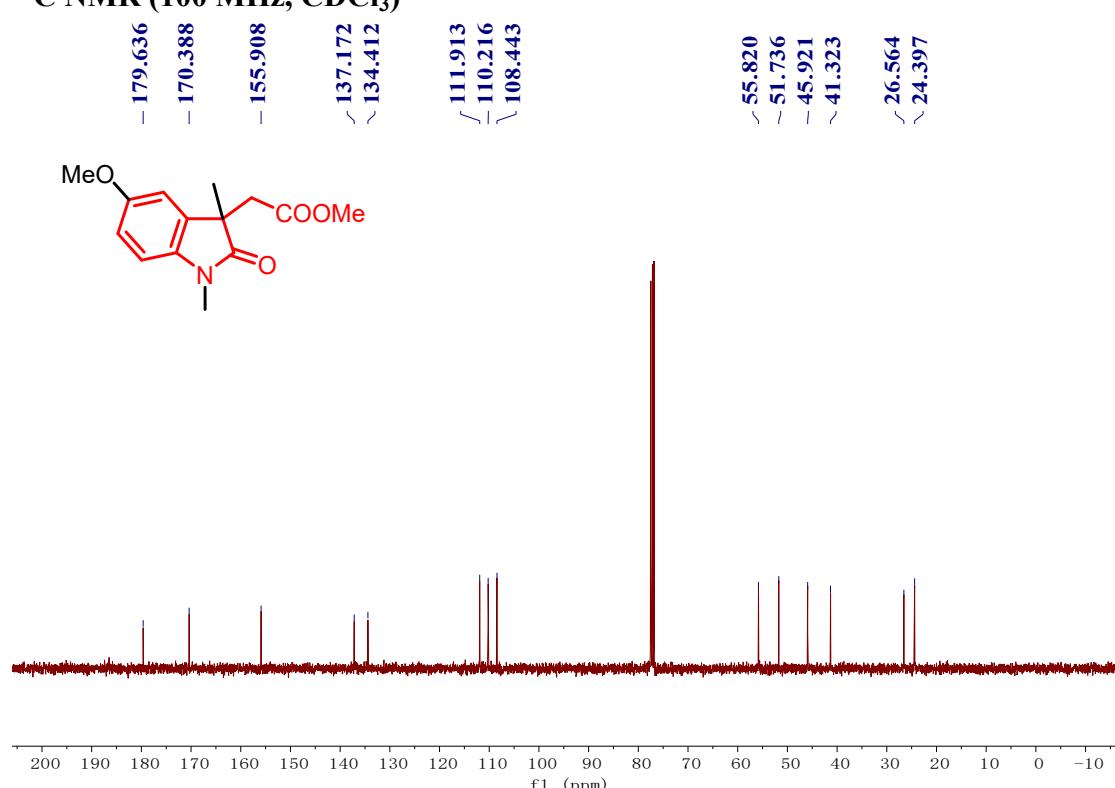
methyl 2-(5-methoxy-1,3-dimethyl-2-oxoindolin-3-yl)acetate (4)

¹H NMR (400 MHz, CDCl₃)



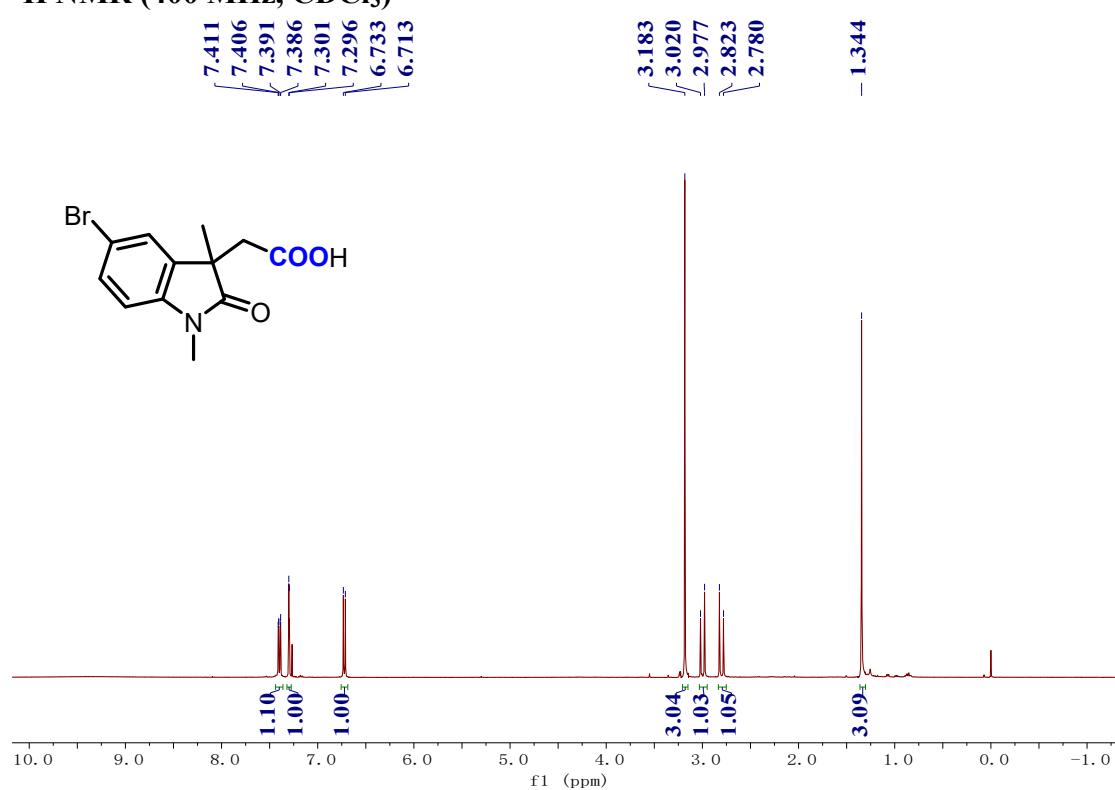
methyl 2-(5-methoxy-1,3-dimethyl-2-oxoindolin-3-yl)acetate (4)

¹³C NMR (100 MHz, CDCl₃)



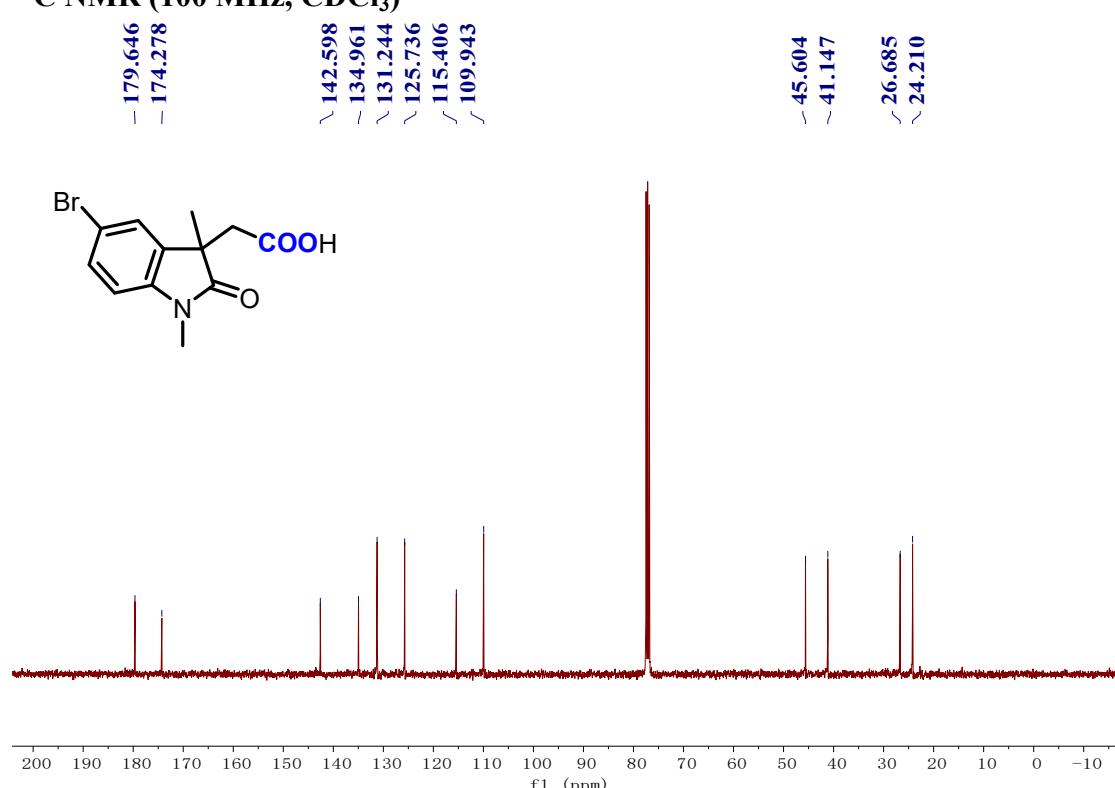
2-(5-bromo-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (5)

¹H NMR (400 MHz, CDCl₃)



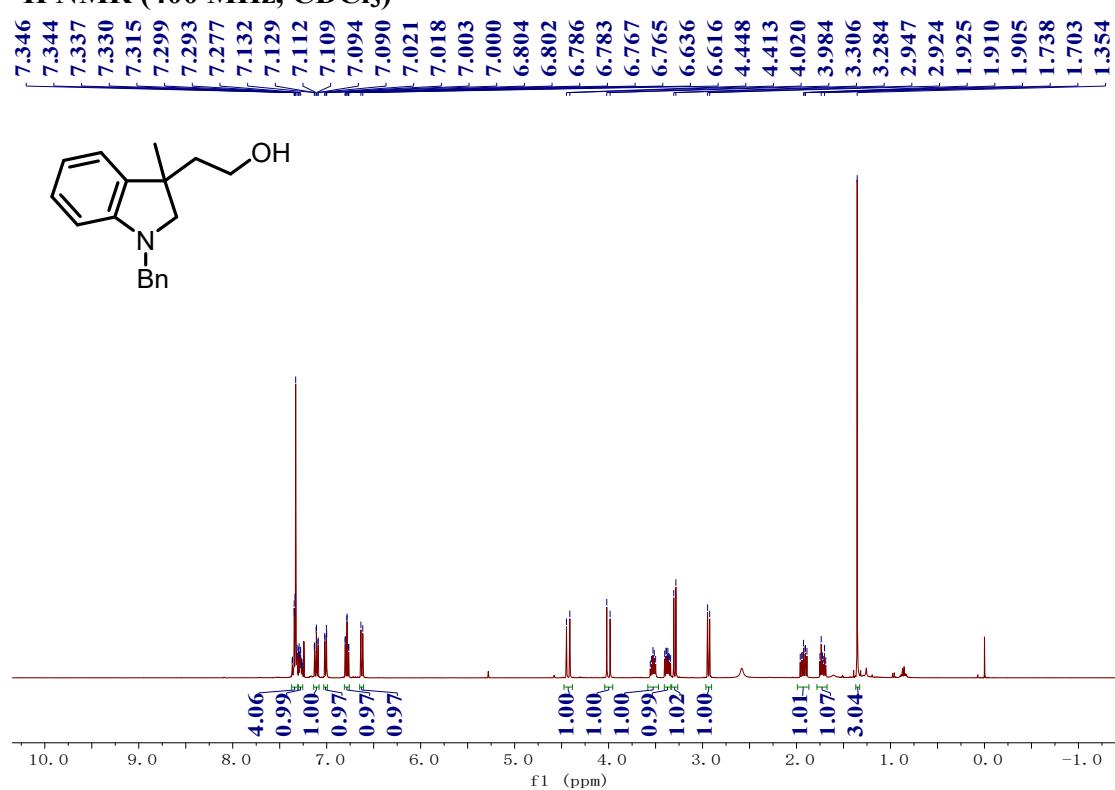
2-(5-bromo-1,3-dimethyl-2-oxoindolin-3-yl)acetic acid (5)

¹³C NMR (100 MHz, CDCl₃)



2-(1-benzyl-3-methylindolin-3-yl)ethan-1-ol (6)

¹H NMR (400 MHz, CDCl₃)



2-(1-benzyl-3-methylindolin-3-yl)ethan-1-ol (6)

¹³C NMR (100 MHz, CDCl₃)

