Synthesis of α -amino acid derived (1,2,3-triazol-4-yl)picolinamide (tzpa) ligands and their corresponding luminescent Tb(III) complexes

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Electronic Supplementary Information (ESI)





Figure S1. 13 C NMR spectrum (150 MHz, DMSO) of 4.



Figure S2. ¹³ C NMR spectrum (150 MHz, DMSO) of 5.



Figure S3. 1H NMR spectrum (600 MHz, DMSO) of 6.



Figure S4. ¹³C NMR spectrum (150 MHz, DMSO) of 6.



Figure S5. ¹*H* NMR spectra of (top) **4** and (bottom) [*Tb*(**4**)₃]³⁺ (400 MHz, *CD*₃*CN*).



Figure S6. ¹H NMR spectrum (400 MHz, CD₃CN) of Tb.5₃.



Figure S7. ¹H NMR spectrum (400 MHz, CD₃CN) of Tb.6₃.



Figure S8. The calculated and experimental isotopic distribution patterns for Tb.4₂ showing the 1:2 metal:ligand stoichiometric pattern for a molecular species of the formula $[Tb(4)_2](CF_3SO_3)_{2^+}$



Figure S9. The calculated and experimental isotopic distribution patterns for $Tb.5_2$ showing the 1:2 metal:ligand stoichiometric pattern for a molecular species of the formula $[Tb(5)_2](CF_3SO_3)_{2^+}$



Figure S10. (Left) The UV-Vis absorbance and excitation ($\lambda_{em} = 545 \text{ nm}$) spectra of $[Tb(4)_3]^{3+}$ recorded in CH₃CN (2.7 × 10⁻⁵M), and (Right) of $[Tb(5)_3]^{3+}$ recorded in CH₃CN (2.5 × 10⁻⁵M).



Figure S11. Delayed luminescence spectra $[Tb(5)_3]^{3+}$ (2.5 × 10⁻⁵M) recorded in CH₃CN showing characteristic Tb(III) transitions ${}^5D_4 \rightarrow {}^7F_{6,5,4,3,2}$.



Figure S12. The overall changes in the (left) UV-visible absorption spectra and (right) fluorescence emission spectra (excitation wavelength $\lambda = 230$ nm) upon titrating **5** (1×10⁻⁵ M) against Tb(CF₃SO₃)₃ (0→3 equiv.) in CH₃CN at RT. **Inset**: corresponding experimental binding isotherms of absorbance at $\lambda = 220$, 250 and 280 nm.



Figure S13. (left) The overall changes to the Tb(III)-centred phosphorescence spectra upon titrating **5** (1×10^{-5} M) against Tb(CF₃SO₃)₃ (0 \rightarrow 3 equiv.) in CH₃CN at RT. (right) corresponding experimental binding isotherms of phosphorescence at I = 492, 545, 583 and 620 nm.



Figure S14. The speciation distribution diagram obtained from (Left) the fit of the UV-visible absorption titration data of ligand **5** against $Tb(CF_3SO_3)_3$ in CH_3CN and (Right) the fit of the experimental binding isotherms using non-linear regression analysis software ReactLab.

Tuble 51 Crystal data and s	
Identification code	4
Empirical formula	$C_{27}H_{25}N_5O_5$
Formula weight	499.52
Temperature/K	100(2)
Crystal system	monoclinic
Space group	P21
a/Å	9.9204(3)
b/Å	9.6670(3)
c/Å	13.2315(4)
α/°	90
β/°	106.3690(10)
γ/°	90
Volume/Å ³	1217.47(6)
Z	2
$\rho_{calc}g/cm^3$	1.363
μ/mm^{-1}	0.793
F(000)	524.0
Crystal size/mm ³	$0.14 \times 0.13 \times 0.1$
Radiation	$CuK\alpha \ (\lambda = 1.54178)$
20 range for data collection/° 9.29 to 136.496	
Index ranges	$-11 \le h \le 11, -11 \le k \le 11, -15 \le l \le 15$
Reflections collected	12684
Independent reflections	4398 [$R_{int} = 0.0369, R_{sigma} = 0.0383$]
Data/restraints/parameters	4398/1/336
Goodness-of-fit on F ²	1.021
Final R indexes [I>= 2σ (I)]	$R_1 = 0.0275, wR_2 = 0.0716$
Final R indexes [all data]	$R_1 = 0.0281, wR_2 = 0.0722$
Largest diff. peak/hole / e Å ⁻³	0.13/-0.14
Flack parameter	0.04(7)
CCDC No.	2239920

Table S1 Crystal data and structure refinement for 4.

Experimental

Materials and Methods

Solvents and reagents were purchased from purchased from Sigma-Aldrich, Alfa Aesar or TCI Ltd. unless otherwise stated and used without further purification.. NMR spectra were recorded using a Bruker AV-600 instrument operating at 600.1 MHz for ¹H NMR and 150.9 MHz for ¹³C NMR. Chemical shifts are reported in commercially available deuterated solvents; δ in ppm relative to SiMe₄ (= 0 ppm) referenced relative to the internal solvent signals. Electrospray mass spectra were determined on a Micromass LCT spectrometer and high resolution mass spectra were determined relative to a standard of leucine enkephaline. Maldi-Q-TOF mass spectra were carried out on a MALDI-Q-TOF-Premier (Waters Corporation, Micromass MS technologies, Manchester, UK) and high-resolution mass spectrometry was performed using Glu-Fib with an internal reference peak of m/z1570.6774. Infrared spectra were recorded on a Perkin Elmer Spectrum One FT-IR spectrometer equipped with universal ATR sampling accessory. UV-Visible absorption spectra were measured in 1-cm quartz cuvettes on a Varian Cary 50 spectrophotometer, and emission (fluorescence, phosphorescence and excitation) spectra and lifetimes were recorded on a Varian Cary Eclipse Fluorimeter, with samples dissolved in spectrophotometric grade solvents.

X-ray Crystallography

X-ray data were collected on a Bruker APEX-II DUO diffractometer using microfocus Cu K α (λ = 1.5405 Å) radiation. All data collections were carried out using standard ω and φ scans at 100 K with temperature control provided by a Cobra cryostream. The data were reduced and multi-scan absorption corrections applied using SADABS[1] within the Bruker APEX3 software suite.[2] Datasets were solved using the intrinsic phasing routine within SHELXT [3] and refined on F² using least squares techniques with SHELXL[4] operating within the OLEX-2 GUI.[5] Non-hydrogen atoms were located from their residuals within the Fourier difference map, while hydrogen atoms were either placed in calculated positions with U_{iso} dependencies derived from their carrier atoms, or (where appropriate for hydrogen bonding species) were manually located from the Fourier map and restrained with distance restraints and U_{iso} dependencies manually. CCDC 2239920.

References

- 1. Bruker APEX-3, Bruker-AXS Inc., Madison, WI, 2016.
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- 3. G. M. Sheldrick, Acta Crystallogr., Sect. A: Found. Adv., 2015, 71, 3–8.
- 4. G. M. Sheldrick, Acta Crystallogr., Sect. C: Struct. Chem., 2015, 71, 3-8.
- 5. O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard and H. Puschmann, J. Appl. Crystallogr., 2009, 42, 339–341.