

# Supporting information

## Electrocatalytic O-S Bonding Reaction Targeting Biological Macromolecules

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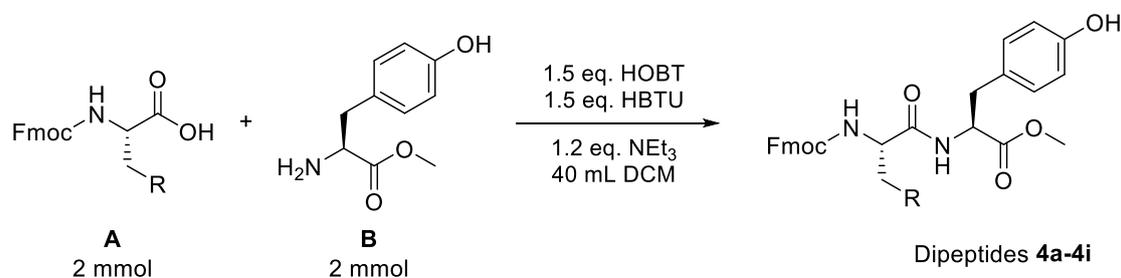
## 1. General Information

All glassware was oven dried at 110°C for hours and cooled down under vacuum. Unless otherwise noted, materials were obtained from commercial suppliers and used without further purification. The instrument for electrolysis was dual display potentiostat (DJS-292B) (made in China). The anodic electrode was graphite rod ( $\phi$  6 mm) and cathodic electrode was platinum plate (15 mm $\times$ 15 mm $\times$ 0.3 mm). Thin layer chromatography (TLC) employed glass 0.25 mm silica gel plates. Flash chromatography columns were packed with 200-300 mesh silica gel. Gradient flash chromatography was conducted eluting with a continuous gradient from dichloromethane to the methanol. High resolution mass spectra (HRMS) for dipeptides were measured with a Waters Micromass GCT instrument and accurate masses were reported for the molecular ion + Sodium (M+Na). High resolution mass spectra (HRMS) for polypeptides were measured with an ABI 5800 instrument and accurate masses were reported for the molecular ion + Hydrogen (M+H) or molecular ion + Sodium (M+Na). The  $^1\text{H}$ ,  $^{13}\text{C}$  and  $^{19}\text{F}$  NMR spectra were recorded on a Bruker 400 MHz NMR spectrometer. For  $^1\text{H}$  NMR, chemical shifts ( $\delta$ ) were given in ppm relatives to internal standard (TMS at 0 ppm, DMSO- $d_6$  at 2.50 ppm, MeOH- $d_4$  at 3.31 ppm, Acetone- $d_6$  at 2.05 ppm). For  $^{13}\text{C}$ -NMR, chemical shifts ( $\delta$ ) were reported in ppm using solvent as internal standard (CDCl $_3$  at 77.00 ppm, DMSO- $d_6$  at 39.50 ppm, MeOH- $d_4$  at 49.00 ppm, Acetone- $d_6$  at 29.84 ppm). HPLC analyses were performed on an Agilent 1260 Infinity LC system using a 100 mm Agilent Zorbax 300SB-C18 5  $\mu\text{m}$  analytical column. All of the MALDI-TOF-MS and MALDI-TOF-MS/MS spectra were acquired using 5800 MALDI-MS (AB SCIEX, Concord, Canada) equipped with a 355 nm Nd: YAG laser in the reflector positive mode. Samples of 0.6  $\mu\text{L}$  mixed with 0.6  $\mu\text{L}$  freshly prepared CHCA matrix were directly loaded onto the stainless steel MALDI plate and allowed to dry in a gentle stream of warm air. Samples were ablated with a power of 3500 while the laser rastered over the target surface. A total of 2000 laser shots were employed in each sample spot. The MS and MS/MS data processing was further performed by DataExplorer 4.0 (AB SCIEX, Concord, Canada). UV-vis absorption

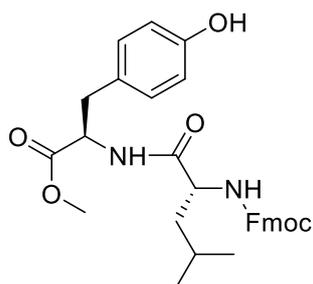
spectra were performed on a Shimadzu UV-2700 spectrophotometer or Agilent Technologies Cary 8454. Fluorescence spectra were collected on a Hitachi F-4600 fluorescence spectrophotometer. The circular dichroism spectra were collected on Chirascan<sup>TM</sup> CD spectroscopy (Applied Photophysics, Leatherhead, United Kingdom). CD spectra were collected from 180 nm to 280 nm and with a scanning speed of 200 nm/min. The bandwidth was 5 nm, and the response time was 2s. All spectra were taken at ambient temperature.

## 2. Synthesis of Starting Materials

### Synthesis of starting materials dipeptides **4a-4i**<sup>{1}{2}</sup>

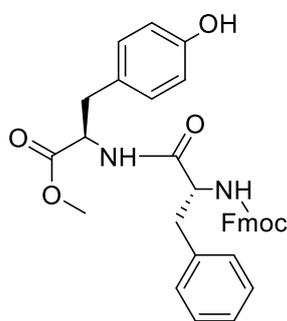


In a round bottomed flask, equipped with a stir bar, peptide **A** (2.0 mmol), HOBT (1-hydroxybenzotriazole) (3.0 mmol), HBTU (O-benzotriazole-*N, N, N', N'*-tetramethyluronium-hexafluorophosphate) (3.0 mmol), dichloromethane (40 mL) and triethylamine (2.4 mmol) were combined and added. The mixture was stirred for 30 min at room temperature, and then, peptide **B** (2.0 mmol) was added to the solution. The reaction was stirred overnight. After regular workup, the reaction mixture was washed by saturated NaHCO<sub>3</sub> solution (40 mL x 3), 2M hydrochloric acid solution (40 mL x 3) and H<sub>2</sub>O (40 mL x 3). The organic layers were combined, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated. The resulting crude product was purified by flash chromatography (DCM/ MeOH) to afford corresponding dipeptides **4a-4i**.



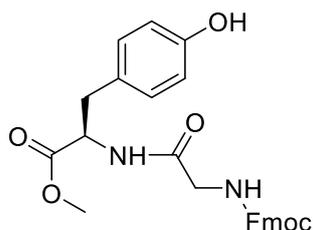
**4a**

Dipeptide **4a Fmoc-Leu-Tyr-OMe**, white solid.  $^1\text{H}$  NMR (400 MHz, Acetone- $d_6$ )  $\delta$  8.37 (s, 1H), 7.84 (d,  $J=7.6$  Hz, 2H), 7.72 – 7.68 (m, 2H), 7.63 – 7.58 (m, 1H), 7.39 (t,  $J=7.6$  Hz, 2H), 7.30 (td,  $J=7.6, 1.2$  Hz, 2H), 7.02 (d,  $J=8.4$  Hz, 2H), 6.84 – 6.80 (m, 1H), 6.75 (d,  $J=8.2$  Hz, 2H), 4.73 – 4.68 (m, 1H), 4.39 – 4.29 (m, 3H), 4.24 – 4.20 (m, 1H), 3.63 (s, 3H), 3.06 – 2.93 (m, 2H), 1.78 – 1.69 (m, 1H), 1.63 – 1.57 (m, 2H), 0.94 – 0.89 (m, 6H).  $^{13}\text{C}$  NMR (101 MHz, Acetone- $d_6$ )  $\delta$  173.18, 172.51, 157.06, 156.97, 145.02, 144.74, 141.95, 131.09, 128.43, 127.87, 126.08, 120.70, 115.96, 67.13, 54.64, 52.21, 47.89, 41.91, 37.34, 25.25, 23.41, 21.91.



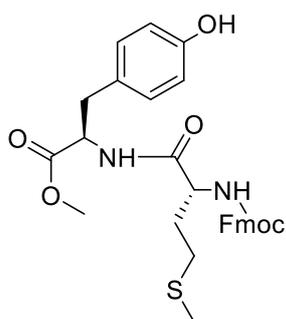
**4b**

Dipeptide **4b Fmoc-Phe-Tyr-OMe**, white solid.  $^1\text{H}$  NMR (400 MHz, Acetone- $d_6$ )  $\delta$  8.39 (s, 1H), 7.83 (d,  $J=7.6$  Hz, 2H), 7.64 – 7.62 (m, 3H), 7.39 (td,  $J=7.6, 1.2$  Hz, 2H), 7.31 – 7.15 (m, 7H), 7.04 – 7.00 (m, 2H), 6.81 (d,  $J=8.8$  Hz, 1H), 6.76 – 6.73 (m, 2H), 4.73 – 4.68 (m, 1H), 4.58 – 4.53 (m, 1H), 4.29 – 4.24 (m, 1H), 4.19 – 4.12 (m, 2H), 3.64 (s, 3H), 3.20 (dd,  $J=14.0, 4.8$  Hz, 1H), 3.07 – 2.90 (m, 3H).  $^{13}\text{C}$  NMR (101 MHz, Acetone- $d_6$ )  $\delta$  172.09, 171.71, 156.77, 156.41, 144.55, 144.47, 141.58, 138.21, 130.79, 129.85, 128.65, 128.09, 127.53, 126.85, 125.73, 120.35, 115.65, 66.87, 56.64, 54.42, 51.92, 47.44, 38.30, 37.07.



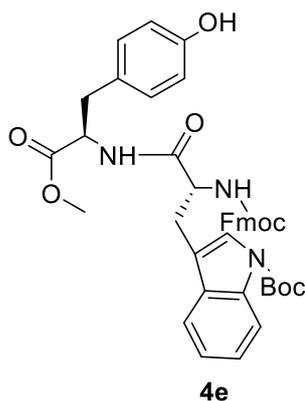
**4c**

Dipeptide **4c Fmoc-Gly-Tyr-OMe**, white solid.  $^1\text{H}$  NMR (400 MHz, Acetone- $d_6$ )  $\delta$  8.32 (s, 1H), 7.85 (d,  $J = 7.6$  Hz, 2H), 7.72 (d,  $J = 7.6$  Hz, 2H), 7.43 – 7.30 (m, 3H), 7.32 (t,  $J = 7.2$  Hz, 2H), 7.03 (d,  $J = 8.4$  Hz, 2H), 6.82 (t,  $J = 6.0$  Hz, 1H), 6.75 (d,  $J = 8.0$  Hz, 2H), 4.71 – 4.66 (m, 1H), 4.37 – 4.22 (m, 3H), 3.91 – 3.81 (m, 2H), 3.64 (s, 3H), 3.04 – 2.99 (m, 1H), 2.96 – 2.91 (m, 1H).  $^{13}\text{C}$  NMR (101 MHz, Acetone- $d_6$ )  $\delta$  172.53, 169.79, 157.45, 157.14, 144.97, 142.01, 131.14, 128.49, 128.05, 127.92, 126.13, 120.75, 116.03, 67.36, 54.60, 52.26, 47.88, 44.66, 37.47.

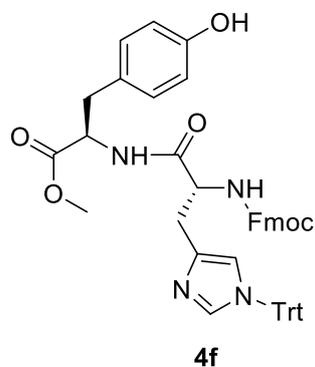


**4d**

Dipeptide **4d Fmoc-Met-Tyr-OMe**, white solid.  $^1\text{H}$  NMR (400 MHz, Acetone- $d_6$ )  $\delta$  8.32 (s, 1H), 7.85 (d,  $J = 7.6$ , 2H), 7.71 (t,  $J = 7.2$  Hz, 2H), 7.52 (d,  $J = 7.6$  Hz, 1H), 7.41 (t,  $J = 7.6$  Hz, 2H), 7.34 – 7.30 (m, 2H), 7.05 – 6.02 (m, 2H), 6.80 (d,  $J = 8.4$  Hz, 1H), 6.77 – 6.673 (m, 2H), 4.69 – 4.64 (m, 1H), 4.39 – 4.29 (m, 3H), 4.25 – 4.20 (m, 1H), 3.65 (s, 3H), 3.06 – 3.11 (m, 1H), 2.98 – 2.93 (m, 1H), 2.60 – 2.48 (m, 2H), 2.13 – 2.03 (m, 1H), 2.05 (s, 3H), 1.98 – 1.88 (m, 1H).  $^{13}\text{C}$  NMR (101 MHz, Acetone- $d_6$ )  $\delta$  172.55, 172.08, 157.13, 156.93, 145.07, 144.85, 142.03, 131.13, 128.50, 127.92, 126.14, 120.77, 116.02, 67.19, 54.82, 54.70, 52.28, 47.94, 37.26, 32.89, 30.61, 15.13.

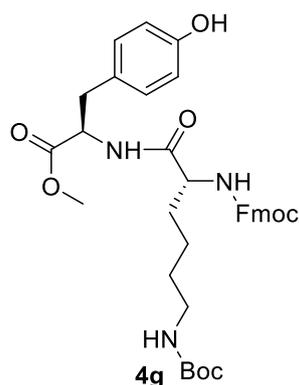


Dipeptide **4e Fmoc-Trp(Boc)-Tyr-OMe**, white solid.  $^1\text{H}$  NMR (400 MHz, DMSO-  $d_6$ )  $\delta$  9.27 (s, 1H), 8.47 (d,  $J = 7.4$  Hz, 1H), 8.02 (s, 1H), 7.85 (d,  $J = 8.0$  Hz, 2H), 7.76 (d,  $J = 7.6$  Hz, 1H), 7.68 (d,  $J = 8.8$  Hz, 1H), 7.59 (d,  $J = 13.5$  Hz, 3H), 7.41 – 7.30 (m, 4H), 7.27 – 7.17 (m, 3H), 7.01 (d,  $J = 8.5$  Hz, 2H), 4.50 – 4.39 (m, 2H), 4.22 – 4.08 (m, 3H), 3.57 (s, 3H), 3.08 – 3.01 (m, 1H), 2.97 – 2.84 (m, 3H), 1.55 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz, DMSO-  $d_6$ )  $\delta$  172.47, 172.16, 157.10, 156.79, 145.00, 144.95, 142.00, 137.55, 131.19, 128.46, 128.13, 127.94, 126.15, 124.55, 122.11, 120.73, 119.57, 119.34, 116.00, 112.15, 111.31, 67.22, 56.48, 54.75, 52.24, 47.92, 37.47, 28.77.28.09.

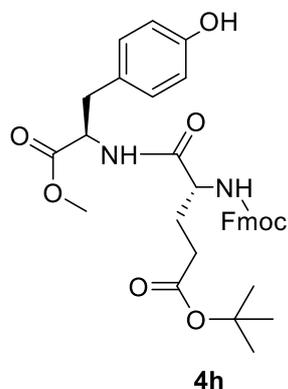


Dipeptide **4f Fmoc-His(Trt)-Tyr-OMe**, white solid.  $^1\text{H}$  NMR (400 MHz, Chloroform-  $d$ )  $\delta$  7.73 (d,  $J = 7.6$  Hz, 2H), 7.59 (dd,  $J = 7.6, 3.9$  Hz, 2H), 7.44 (dd,  $J = 6.4, 4.7$  Hz, 1H), 7.40 (d,  $J = 1.5$  Hz, 1H), 7.37 – 7.33 (m, 2H), 7.27 – 7.24 (m, 9H), 7.06 – 7.03 (m, 6H), 6.89 (d,  $J = 8.5$  Hz, 2H), 6.66 (s, 1H), 6.57 (d,  $J = 8.5$  Hz, 2H), 6.32 (d,  $J = 7.7$  Hz, 1H), 4.85 – 4.79 (m, 1H), 4.52 (s, 1H), 4.29 – 4.24 (m, 2H), 4.14 (d,  $J = 7.5$  Hz, 1H), 3.55 (s, 3H), 3.11 – 2.90 (m, 4H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-  $d$ )  $\delta$  171.63, 171.03, 156.40, 156.10, 143.94, 142.07, 141.22, 138.14, 136.55, 130.49, 129.73, 128.17, 128.12, 127.68, 127.15, 127.11, 126.61, 125.38, 125.33, 119.91, 119.73, 115.82, 67.30,

55.62, 53.48, 52.21, 47.10, 37.36, 31.38.

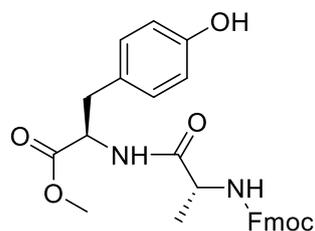


Dipeptide **4g Fmoc-Lys(Boc)-Tyr-OMe**, white solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  9.23 (s, 1H), 8.23 (d,  $J = 7.4$  Hz, 1H), 7.89 (d,  $J = 7.5$  Hz, 2H), 7.75 – 7.71 (m, 2H), 7.45 – 7.40 (m, 3H), 7.33 (t,  $J = 7.4$  Hz, 2H), 6.98 (d,  $J = 8.4$  Hz, 2H), 6.77 (t,  $J = 5.7$  Hz, 1H), 6.65 (d,  $J = 8.4$  Hz, 2H), 4.37 (q,  $J = 7.3$  Hz, 1H), 4.31 – 4.14 (m, 4H), 4.03 – 3.95 (m, 1H), 3.56 (s, 3H), 2.93 – 2.82 (m, 4H), 1.63 – 1.42 (m, 3H), 1.37 (s, 9H), 1.28 – 1.16 (m, 3H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  172.63, 172.19, 156.31, 156.05, 149.55, 149.48, 144.39, 144.22, 141.18, 134.30, 130.97, 128.09, 127.51, 125.77, 120.54, 120.06, 120.01, 77.81, 66.10, 54.87, 53.88, 52.26, 47.16, 36.25, 32.09, 29.69, 28.73, 23.20 .



Dipeptide **4h Fmoc-Glu(tBu)-Tyr-OMe**, white solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  9.25 (s, 1H), 8.26 (d,  $J = 7.4$  Hz, 1H), 7.90 (d,  $J = 7.5$  Hz, 2H), 7.74 (t,  $J = 7.4$  Hz, 2H), 7.51 (d,  $J = 8.4$  Hz, 1H), 7.42 (t,  $J = 6.9$  Hz, 2H), 7.33 (t,  $J = 7.5$  Hz, 2H), 7.00 (d,  $J = 8.4$  Hz, 2H), 6.66 (d,  $J = 8.5$  Hz, 2H), 4.39 (dt,  $J = 13.6, 6.9$  Hz, 1H), 4.26 (dd,  $J = 17.2, 8.0$  Hz, 3H), 4.06 (td,  $J = 8.5, 5.3$  Hz, 1H), 3.58 (s, 3H), 2.92 – 2.83 (m, 2H), 2.23 (t,  $J = 8.0$  Hz, 2H), 1.92 – 1.80 (m, 1H), 1.73 (dq,  $J = 16.5, 8.1$  Hz, 1H), 1.40 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  172.12, 172.00, 156.27, 149.55, 149.49, 144.38, 144.19,

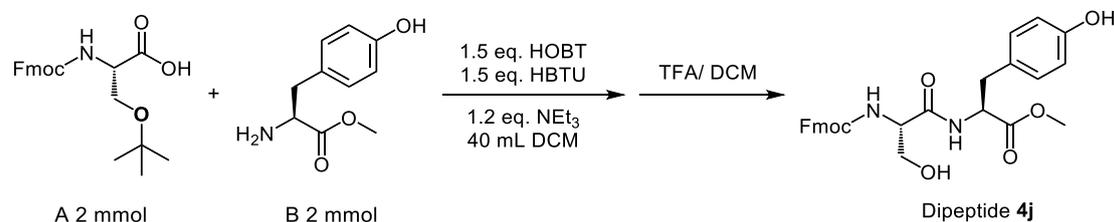
141.19, 134.30, 130.97, 128.10, 127.51, 125.77, 120.56, 120.07, 120.03, 80.15, 66.14, 54.05, 53.93, 52.30, 47.14, 36.14, 31.69, 28.22, 27.80.



**4i**

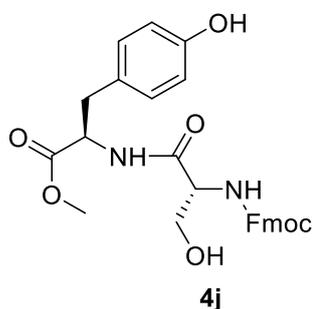
Dipeptide **4i** **Fmoc-Ala-Tyr-OMe**, white solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  8.25 (d,  $J = 8.1$  Hz, 1H), 8.14 (d,  $J = 8.0$  Hz, 1H), 7.87 (dd,  $J = 17.3, 7.5$  Hz, 4H), 7.74 (d,  $J = 4.9$  Hz, 1H), 7.42 (t,  $J = 7.4$  Hz, 2H), 7.34 (q,  $J = 7.2$  Hz, 2H), 6.98 (dd,  $J = 8.3, 1.7$  Hz, 2H), 6.68 – 6.61 (m, 2H), 4.42 (tt,  $J = 8.9, 5.0$  Hz, 1H), 4.27 – 4.17 (m,  $J = 12.3, 6.2$  Hz, 3H), 4.04 (dt,  $J = 29.4, 7.4$  Hz, 1H), 3.62 (d,  $J = 2.0$  Hz, 3H), 2.97 – 2.73 (m, 2H), 1.08 – 1.02 (m, 3H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  173.03, 172.31, 156.02, 149.51, 149.45, 144.34, 144.28, 141.17, 134.37, 131.10, 128.10, 127.53, 125.81, 120.56, 120.02, 119.97, 66.13, 53.61, 52.42, 50.23, 47.07, 36.52, 18.78.

#### Synthesis of starting materials dipeptides **4j**<sup>{1}{2}</sup>



In a round bottomed flask, equipped with a stir bar, peptide **A** (2.0 mmol), HOBT (1-hydroxybenzotriazole) (3.0 mmol), HBTU (O-benzotriazole-*N, N, N', N'*-tetramethyluronium-hexafluorophosphate) (3.0 mmol), dichloromethane (40 mL) and triethylamine (2.4 mmol) were combined and added. The mixture was stirred for 30 min at room temperature, and then, peptide **B** (2.0 mmol) was added to the solution. The reaction was stirred overnight. After regular workup, the reaction mixture washed by saturated  $\text{NaHCO}_3$  solution (40 mL x 3), 2M hydrochloric acid solution (40 mL x 3) and  $\text{H}_2\text{O}$  (40 mL x 3). The organic layers were combined, dried over  $\text{Na}_2\text{SO}_4$ , and concentrated. Without further purification, the 95% TFA / DCM solution (8 mL) was added dropwise. The mixture was stirred for 2 h at room temperature. The resulting

crude product was purified by flash chromatography (DCM / MeOH) to afford corresponding dipeptides **4j**.



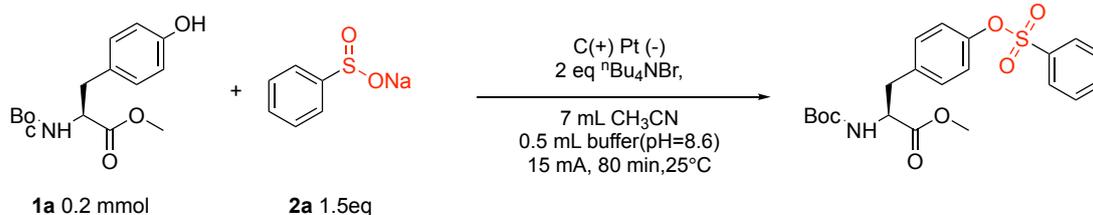
Dipeptide **4j** **Fmoc-Ser-Tyr-OMe**, white solid.  $^1\text{H}$  NMR (400 MHz, Acetone- $d_6$ )  $\delta$  8.32 (s, 1H), 7.88 (d,  $J = 7.6$  Hz, 2H), 7.76 – 7.73 (m, 2H), 7.60 (d,  $J = 7.6$  Hz, 1H), 7.43 (t,  $J = 7.6$  Hz, 2H), 7.34 (t,  $J = 7.6$  Hz, 2H), 7.07 (d,  $J = 8.0$  Hz, 2H), 6.77 (d,  $J = 8.0$  Hz, 2H), 6.69 (d,  $J = 8.0$  Hz, 1H), 4.74 – 4.69 (m, 1H), 4.36 – 4.31 (m, 3H), 4.28 – 4.22 (m, 2H), 3.86 – 3.74 (m, 2H), 3.65 (s, 3H), 3.07 – 3.02 (m, 1H), 3.02 – 2.97 (m, 1H).  $^{13}\text{C}$  NMR (101 MHz, Acetone- $d_6$ )  $\delta$  172.54, 171.02, 157.15, 157.05, 145.02, 144.91, 142.01, 131.20, 128.50, 127.95, 126.15, 120.76, 116.03, 67.42, 63.25, 57.53, 54.77, 52.34, 47.89, 37.35.

### 3. General Procedure for Bioconjugation of Tyrosine and Sodium benzenesulfinate

#### 3.1 Reaction Optimization

In an oven-dried undivided three-necked bottle (25 mL) equipped with a stir bar, protected tyrosine (0.20 mmol), Sodium benzenesulfinate (0.3 mmol),  $^n\text{Bu}_4\text{NBr}$  (0.40 mmol) and MeCN / buffer(pH=8.6) (7.0 mL / 0.5 mL) were combined and added. The bottle was equipped graphite rod ( $\phi$  6 mm, about 15 mm immersion depth in solution) as the anode and platinum plate (15 mm $\times$ 15 mm $\times$ 0.3 mm) as the cathode. The reaction mixture was stirred and electrolyzed at constant current under room temperature. When the reaction finished, The pure product was obtained by flash column chromatography on silica gel. A summary of optimization results is presented in **Table S1** below.

**Table S1. Effects of reaction parameters**



Entry	Variation from the standard conditions	Isolated yields
1	none	87%
2	8 mA instead of 15 mA, 160 min	64%
3	20 mA instead of 15 mA, 60 min	52%
4	H <sub>2</sub> O instead of buffer	42%
5	add to 1 eq HCl	trace
6	1.5 ml buffer	64%
7	without buffer	27%
8	CH <sub>3</sub> OH instead of CH <sub>3</sub> CN	trace
9	CH <sub>2</sub> Cl <sub>2</sub> instead of CH <sub>3</sub> CN	75%
10	C(+) C (-) instead of C(+) Pt (-)	56%
11	C(+) Ni (-) instead of C(+) Pt (-)	52%
12	Pt(+) Pt (-) instead of C(+) Pt (-)	75%
13	<sup>n</sup> Bu <sub>4</sub> NBF <sub>4</sub> instead of <sup>n</sup> Bu <sub>4</sub> NBr	30%
14	<sup>n</sup> Bu <sub>4</sub> NI instead of <sup>n</sup> Bu <sub>4</sub> NBr	24%
15	<sup>n</sup> Bu <sub>4</sub> NClO <sub>4</sub> instead of <sup>n</sup> Bu <sub>4</sub> NBr	trace
16	KBr instead of <sup>n</sup> Bu <sub>4</sub> NBr	74%
17	under N <sub>2</sub>	83%
18	no electric current	n.r
19	Serine	n.d
20	Threonine	n.d
21	Hydroxyproline	n.d

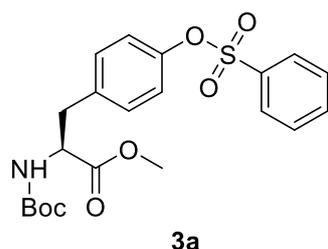
[a] Reaction conditions: graphite rod anode, platinum plate cathode, constant current = 15 mA, **1a** (1.0 equiv., 0.20 mmol), **2a** (1.5 equiv, 0.3 mmol), <sup>n</sup>Bu<sub>4</sub>NBr (2 equiv, 0.40 mmol), 7.0 mL MeCN, 0.5 mL buffer (pH = 8.6), 25°C, 80min. Yields of isolated products are shown. n r = no reaction. n d = no detected.

### 3.2 Gram-Scale Experiments

General procedure for Gram-Scale Experiments: In an oven-dried undivided three-necked bottle (250 mL) equipped with a stir bar, tyrosine (5.0 mmol), Sodium benzenesulfinate (7.5 mmol), and <sup>n</sup>Bu<sub>4</sub>NBr (10.0 mmol), buffer (pH = 8.6, 12 mL) were

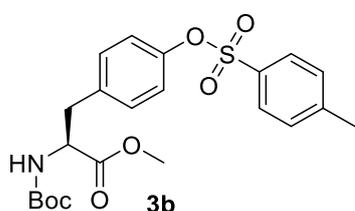
combined and added. Then, CH<sub>3</sub>CN (160 mL) were injected into the tubes via syringes. The bottle was equipped with carbon rod (ϕ 6 mm) as the anode and platinum plate (15 mm×15 mm×0.3 mm) as the cathode. The reaction mixture was stirred and electrolysis at constant current of 15 mA under 25°C overnight. The solvent was removed under vacuum. The crude product was purified by flash column chromatography on silica gel to afford pure product.

### 3.3 Sodium arenesulfonates scope and characterization



#### **methyl (S)-2-((tert-butoxycarbonyl)amino)-3-(4-((phenylsulfonyl)oxy)phenyl)propanoate(3a);**

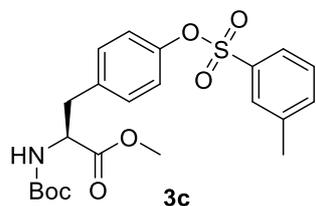
75.7 mg (yield: 87%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.86 – 7.81 (m, 2H), 7.81 – 7.77 (m, 1H), 7.65 (t, *J* = 7.9 Hz, 2H), 7.31 (d, *J* = 8.2 Hz, 1H), 7.23 (d, *J* = 8.7 Hz, 2H), 6.93 (d, *J* = 8.7 Hz, 2H), 4.21 – 4.11 (m, 1H), 3.57 (s, 3H), 2.97 (dd, *J* = 13.8, 5.3 Hz, 1H), 2.83 (dd, *J* = 13.8, 10.2 Hz, 1H), 1.30 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.84, 155.81, 148.10, 137.51, 135.39, 134.89, 131.08, 130.20, 128.61, 122.17, 78.77, 55.30, 52.24, 36.20, 28.55. HRMS (ESI) calcd. for (M+Na)<sup>+</sup> C<sub>21</sub>H<sub>25</sub>NO<sub>7</sub>S: 458.1243, found, 458.1248.



#### **methyl (S)-2-((tert-butoxycarbonyl)amino)-3-(4-(tosyloxy)phenyl)propanoate(3b);**

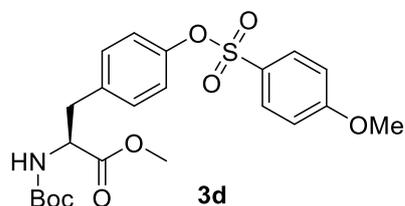
62.1 mg (yield: 69%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.71 (d, *J* = 8.4 Hz, 2H), 7.45 (d, *J* = 7.9 Hz, 2H), 7.30 (d, *J* = 8.2 Hz, 1H), 7.22 (d, *J* = 8.7 Hz, 2H), 6.92 (d, *J* = 8.7 Hz, 2H), 4.15 (dd, *J* = 18.4, 5.3 Hz, 1H), 3.57 (s, 3H), 2.96 (dd, *J* = 13.8, 5.3 Hz, 1H), 2.82 (dd, *J* = 13.9, 10.2 Hz, 1H), 2.41 (s, 3H), 1.31 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.85, 155.81, 148.15, 146.13, 137.40, 132.01,

131.05, 130.62, 128.64, 122.17, 78.77, 55.30, 52.24, 36.19, 28.55, 21.64. HRMS (ESI) cald. for (M+Na)<sup>+</sup> C<sub>22</sub>H<sub>27</sub>NO<sub>7</sub>S: 472.1400, found, 472.1423.



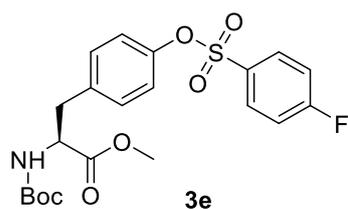
**methyl (S)-2-((tert-butoxycarbonyl)amino)-3-(4-((m-tolylsulfonyl)oxy)phenyl)propanoate(3c);**

54.8 mg (yield: 61%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO- *d*<sub>6</sub>) δ 7.70 (s, 1H), 7.63 (d, *J* = 7.0 Hz, 2H), 7.54 (t, *J* = 7.7 Hz, 1H), 7.33 (d, *J* = 8.3 Hz, 1H), 7.25 (d, *J* = 8.7 Hz, 2H), 6.95 (d, *J* = 8.5 Hz, 2H), 4.22 – 4.14 (m, 1H), 3.60 (s, 3H), 2.99 (dd, *J* = 13.8, 5.1 Hz, 1H), 2.85 (dd, *J* = 13.8, 10.2 Hz, 1H), 2.41 (s, 3H), 1.33 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO- *d*<sub>6</sub>) δ 172.85, 155.82, 148.12, 140.23, 137.46, 136.00, 134.87, 131.06, 129.96, 128.60, 125.78, 122.19, 78.77, 55.33, 52.23, 36.19, 28.55, 21.11. HRMS (ESI) cald. for (M+Na)<sup>+</sup> C<sub>22</sub>H<sub>27</sub>NO<sub>7</sub>S: 472.1400, found, 472.1425.



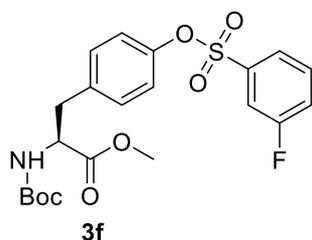
**methyl (S)-2-((tert-butoxycarbonyl)amino)-3-(4-(((4-methoxyphenyl)sulfonyl)oxy)phenyl)propanoate(3d);**

54.0 mg (yield: 58%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO- *d*<sub>6</sub>) δ 7.77 (d, *J* = 9.0 Hz, 2H), 7.33 (d, *J* = 8.2 Hz, 1H), 7.25 (d, *J* = 8.7 Hz, 2H), 7.16 (d, *J* = 9.0 Hz, 2H), 6.94 (d, *J* = 8.7 Hz, 2H), 4.23 – 4.14 (m, 1H), 3.87 (s, 3H), 3.60 (s, 3H), 2.99 (dd, *J* = 13.8, 5.3 Hz, 1H), 2.85 (dd, *J* = 13.9, 10.2 Hz, 1H), 1.33 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO- *d*<sub>6</sub>) δ 172.86, 164.41, 155.83, 148.21, 137.33, 131.02, 126.11, 122.23, 115.32, 78.78, 56.34, 55.33, 52.23, 36.20, 28.54. HRMS (ESI) cald. for (M+Na)<sup>+</sup> C<sub>22</sub>H<sub>27</sub>NO<sub>8</sub>S: 488.1349, found, 488.1343.



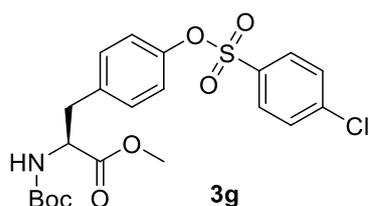
**methyl (S)-2-((tert-butoxycarbonyl)amino)-3-(4-((4-fluorophenyl)sulfonyl)oxy)phenyl)propanoate(3e);**

63.5 mg (yield: 70%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.91 (dd, *J* = 8.9, 5.0 Hz, 2H), 7.48 (t, *J* = 8.8 Hz, 2H), 7.31 (d, *J* = 8.3 Hz, 1H), 7.24 (d, *J* = 8.8 Hz, 2H), 6.94 (d, *J* = 8.7 Hz, 2H), 4.16 (dd, *J* = 13.4, 10.2 Hz, 1H), 3.58 (s, 3H), 2.97 (dd, *J* = 13.9, 5.1 Hz, 1H), 2.83 (dd, *J* = 13.8, 10.2 Hz, 1H), 1.30 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.83, 167.27, 164.73, 155.81, 148.02, 137.63, 132.06, 131.96, 131.14, 131.11, 131.08, 122.21, 117.69, 117.46, 78.77, 55.29, 52.24, 36.18, 28.54. <sup>19</sup>F NMR (377 MHz, DMSO-*d*<sub>6</sub>) δ -102.57. HRMS (ESI) calcd. for (M+Na)<sup>+</sup> C<sub>21</sub>H<sub>24</sub>FNO<sub>7</sub> S: 476.1149 ,found, 476.1138.



**methyl (S)-2-((tert-butoxycarbonyl)amino)-3-(4-((3-fluorophenyl)sulfonyl)oxy)phenyl)propanoate(3f);**

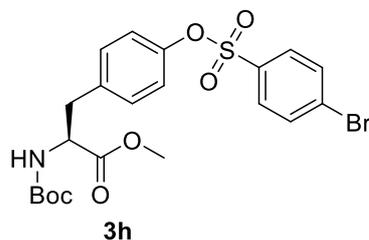
54.4 mg (yield: 60%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.75 – 7.69 (m, 4H), 7.28 (t, *J* = 9.3 Hz, 3H), 7.00 (d, *J* = 8.6 Hz, 2H), 4.24 – 4.14 (m, 1H), 3.59 (s, 3H), 3.00 (dd, *J* = 13.8, 5.2 Hz, 1H), 2.85 (dd, *J* = 13.8, 10.1 Hz, 1H), 1.34 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.80, 163.44, 160.96, 155.79, 148.00, 137.71, 136.74, 136.67, 132.70, 132.62, 131.18, 125.07, 125.04, 122.85, 122.64, 122.12, 115.84, 115.59, 78.76, 55.25, 52.21, 36.22, 28.53. <sup>19</sup>F NMR (377 MHz, DMSO-*d*<sub>6</sub>) δ -57.04. HRMS (ESI) calcd. for (M+Na)<sup>+</sup> C<sub>21</sub>H<sub>24</sub>FNO<sub>7</sub> S: 476.1149, found, 476.1141.



**methyl (S)-2-(((tert-butoxycarbonyl)amino)-3-(4-(((4-chlorophenyl)sulfonyl)oxy)phenyl)propanoate(3g);**

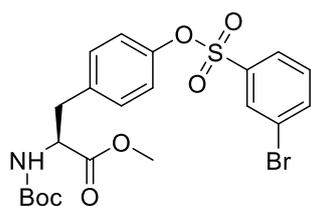
56.4 mg (yield: 60%, 0.2 mmol scale), white solid.<sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.86 (d, *J* = 8.7 Hz, 2H), 7.73 (d, *J* = 8.7 Hz, 2H), 7.28 (dd, *J* = 13.5, 8.4 Hz, 3H), 6.98 (d, *J* = 8.6 Hz, 2H), 4.24 – 4.12 (m, 1H), 3.59 (s, 3H), 2.99 (dd, *J* = 13.8, 5.1 Hz, 1H), 2.85 (dd, *J* = 13.9, 10.1 Hz, 1H), 1.32 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.79, 155.80, 148.01, 140.48, 137.69, 133.68, 131.18, 130.57, 130.41, 122.16, 78.77, 55.27, 52.22, 36.21, 28.55.

HRMS (ESI) cald. for (M+Na)<sup>+</sup> C<sub>21</sub>H<sub>24</sub>ClNO<sub>7</sub>S: 492.0854, found, 492.0857



**methyl (S)-3-(4-(((4-bromophenyl)sulfonyl)oxy)phenyl)-2-(((tert-butoxycarbonyl)amino)propanoate(3h);**

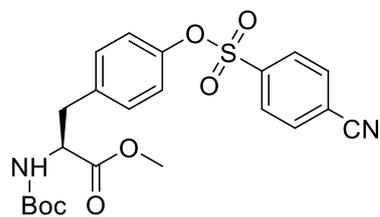
77.2 mg (yield: 75%, 0.2 mmol scale), white solid.<sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.86 (d, *J* = 8.7 Hz, 2H), 7.76 (d, *J* = 8.8 Hz, 2H), 7.32 (d, *J* = 8.2 Hz, 1H), 7.25 (d, *J* = 8.7 Hz, 2H), 6.97 (d, *J* = 8.7 Hz, 2H), 4.20 – 4.09 (m, 1H), 3.58 (s, 3H), 2.98 (dd, *J* = 13.8, 5.2 Hz, 1H), 2.83 (dd, *J* = 13.8, 10.2 Hz, 1H), 1.30 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.83, 155.81, 147.98, 137.70, 134.05, 133.36, 131.20, 130.56, 129.65, 122.18, 78.76, 55.28, 52.25, 36.17, 28.54. HRMS (ESI) cald. for (M+Na)<sup>+</sup> C<sub>21</sub>H<sub>24</sub>BrNO<sub>7</sub>S: 536.0349, found, 536.0346.



3i

**methyl (S)-3-(4-(((3-bromophenyl)sulfonyl)oxy)phenyl)-2-((tert-butoxycarbonyl)amino)propanoate(3i);**

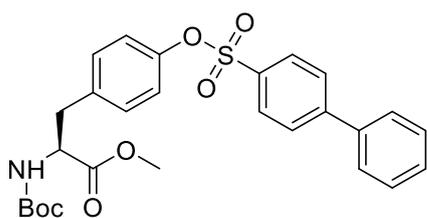
80.2 mg (yield: 78%, 0.2 mmol scale), white solid.  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  8.04 (ddd,  $J = 8.0, 2.0, 1.0$  Hz, 1H), 8.00 (t,  $J = 1.9$  Hz, 1H), 7.87 (d,  $J = 9.7$  Hz, 1H), 7.63 (t,  $J = 8.0$  Hz, 1H), 7.32 – 7.25 (m, 3H), 7.01 (d,  $J = 8.7$  Hz, 2H), 4.25 – 4.16 (m, 1H), 3.60 (s, 3H), 3.00 (dd,  $J = 13.8, 5.3$  Hz, 1H), 2.86 (dd,  $J = 13.8, 10.1$  Hz, 1H), 1.32 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  172.80, 155.79, 147.95, 138.33, 137.75, 136.80, 132.38, 131.20, 130.70, 127.73, 122.98, 122.14, 78.77, 55.26, 52.22, 36.23, 28.55. HRMS (ESI) cald. for  $(\text{M}+\text{Na})^+$   $\text{C}_{21}\text{H}_{24}\text{BrNO}_7\text{S}$  : 536.0349, found, 536.0342.



3j

**methyl (S)-2-((tert-butoxycarbonyl)amino)-3-(4-(((4-cyanophenyl)sulfonyl)oxy)phenyl)propanoate(3j);**

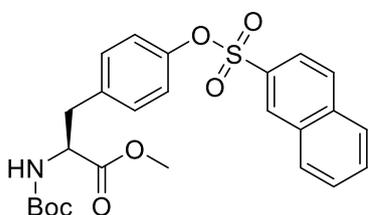
45.1 mg (yield: 49%, 0.2 mmol scale), white solid.  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  8.14 (d,  $J = 8.6$  Hz, 2H), 8.04 (d,  $J = 8.7$  Hz, 2H), 7.27 (dd,  $J = 11.6, 8.4$  Hz, 3H), 6.99 (d,  $J = 8.6$  Hz, 2H), 4.18 (dd,  $J = 18.4, 5.1$  Hz, 1H), 3.59 (s, 3H), 2.99 (dd,  $J = 13.9, 5.2$  Hz, 1H), 2.85 (dd,  $J = 13.9, 10.2$  Hz, 1H), 1.31 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  172.78, 155.80, 147.88, 138.85, 137.88, 134.30, 131.27, 129.42, 122.13, 117.74, 117.73, 78.78, 55.24, 52.24, 36.19, 28.54. HRMS (ESI) cald. for  $(\text{M}+\text{Na})^+$   $\text{C}_{22}\text{H}_{24}\text{N}_2\text{O}_7\text{S}$  : 483.1196, found, 483.1188.



**3k**

**methyl (S)-3-(4-(((1,1'-biphenyl)-4-ylsulfonyl)oxy)phenyl)-2-((tert-butoxycarbonyl)amino)propanoate(3k);**

46.1 mg (yield: 45%, 0.2 mmol scale), white solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  7.94 (q,  $J = 8.7$  Hz, 4H), 7.77 (d,  $J = 8.0$  Hz, 2H), 7.54 (t,  $J = 7.3$  Hz, 2H), 7.48 (t,  $J = 7.2$  Hz, 1H), 7.27 (dd,  $J = 13.9, 8.4$  Hz, 3H), 7.00 (d,  $J = 8.6$  Hz, 2H), 4.22 – 4.12 (m, 1H), 3.58 (s, 3H), 2.99 (dd,  $J = 13.9, 5.2$  Hz, 1H), 2.84 (dd,  $J = 13.9, 10.2$  Hz, 1H), 1.30 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  172.82, 155.81, 148.14, 146.61, 138.31, 137.52, 133.61, 131.13, 129.69, 129.50, 129.30, 128.24, 127.69, 122.18, 78.77, 55.29, 52.22, 36.20, 28.54. HRMS (ESI) calcd. for  $(\text{M}+\text{Na})^+$   $\text{C}_{27}\text{H}_{29}\text{NO}_7\text{S}$  : 534.1556, found, 534.1558.

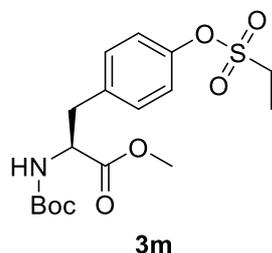


**3l**

**methyl (S)-2-((tert-butoxycarbonyl)amino)-3-(4-((naphthalen-2-ylsulfonyl)oxy)phenyl)propanoate(3l);**

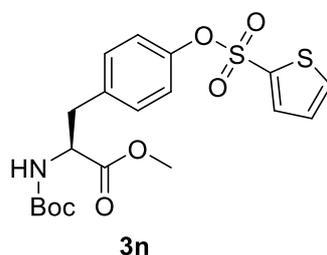
72.0 mg (yield: 74%, 0.2 mmol scale), white solid.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  8.56 (s, 1H), 8.22 – 8.18 (m, 2H), 8.09 (d,  $J = 8.2$  Hz, 1H), 7.88 (dd,  $J = 8.7, 2.1$  Hz, 1H), 7.76 (t,  $J = 7.6$  Hz, 1H), 7.69 (t,  $J = 7.5$  Hz, 1H), 7.27 (d,  $J = 8.2$  Hz, 1H), 7.22 (d,  $J = 8.7$  Hz, 2H), 6.98 (d,  $J = 8.6$  Hz, 2H), 4.24 – 4.13 (m, 1H), 3.55 (s, 3H), 2.97 (dd,  $J = 13.9, 5.2$  Hz, 1H), 2.83 (dd,  $J = 13.9, 10.1$  Hz, 1H), 1.28 (s, 9H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  172.80, 155.80, 148.20, 137.47, 135.52, 132.01, 131.93, 131.09, 130.64,

130.41, 130.37, 130.05, 128.50, 128.45, 122.96, 122.17, 78.74, 55.25, 52.16, 36.22, 28.51. HRMS (ESI) cald. for (M+Na)<sup>+</sup> C<sub>25</sub>H<sub>27</sub>NO<sub>7</sub>S : 508.1400, found, 508.1408.



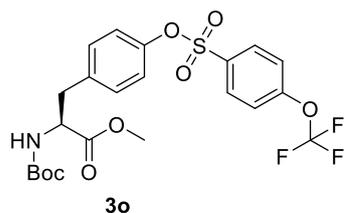
**methyl (S)-2-((tert-butoxycarbonyl)amino)-3-(4-((ethylsulfonyl)oxy)phenyl)propanoate(3m);**

47.3 mg (yield: 61%, 0.2 mmol scale), white solid.<sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.39 – 7.33 (m, 3H), 7.26 (d, *J* = 8.7 Hz, 2H), 4.27 – 4.18 (m, 1H), 3.64 (s, 3H), 3.48 (q, *J* = 7.3 Hz, 2H), 3.05 (dd, *J* = 13.8, 5.0 Hz, 1H), 2.89 (dd, *J* = 13.8, 10.3 Hz, 1H), 1.38 (t, *J* = 7.3 Hz, 3H), 1.34 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.89, 155.87, 148.07, 137.28, 131.22, 122.26, 78.80, 55.41, 52.28, 44.90, 36.20, 28.55, 8.51. HRMS (ESI) cald. for (M+Na)<sup>+</sup> C<sub>17</sub>H<sub>25</sub>NO<sub>7</sub>S : 410.1243, found, 410.1241.



**methyl (S)-2-((tert-butoxycarbonyl)amino)-3-(4-((thiophen-2-ylsulfonyl)oxy)phenyl)propanoate(3n);**

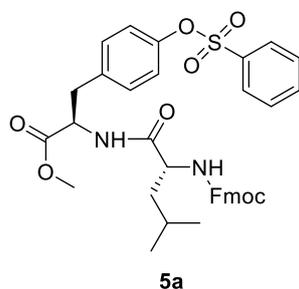
61.8 mg (yield: 70%, 0.2 mmol scale), white solid.<sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.19 (dd, *J* = 5.0, 1.4 Hz, 1H), 7.75 (dd, *J* = 3.9, 1.5 Hz, 1H), 7.33 (d, *J* = 8.2 Hz, 1H), 7.28 – 7.24 (m, 3H), 6.97 (d, *J* = 8.6 Hz, 2H), 4.17 (dd, *J* = 16.8, 6.7 Hz, 1H), 3.58 (s, 3H), 2.98 (dd, *J* = 13.8, 5.2 Hz, 1H), 2.84 (dd, *J* = 13.8, 10.2 Hz, 1H), 1.31 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.86, 155.81, 148.17, 137.76, 137.51, 136.81, 133.54, 131.14, 128.87, 122.06, 78.79, 55.32, 52.26, 36.20, 28.56. HRMS (ESI) cald. for (M+Na)<sup>+</sup> C<sub>19</sub>H<sub>23</sub>NO<sub>7</sub>S<sub>2</sub> : 464.0808, found, 464.0802.



**methyl (S)-2-((tert-butoxycarbonyl)amino)-3-(4-(((4-(trifluoromethoxy)phenyl)sulfonyl)oxy)phenyl)propanoate(3o);**

70.6 mg (yield: 68%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.02 (d, *J* = 9.0 Hz, 2H), 7.65 (d, *J* = 10.0 Hz, 2H), 7.29 (t, *J* = 9.4 Hz, 3H), 7.00 (d, *J* = 8.7 Hz, 2H), 4.27 – 4.14 (m, 1H), 3.60 (s, 3H), 3.01 (dd, *J* = 13.8, 5.2 Hz, 1H), 2.87 (dd, *J* = 13.9, 10.1 Hz, 1H), 1.32 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.78, 155.80, 152.96, 152.94, 147.99, 137.73, 133.61, 131.49, 131.17, 122.12, 122.04, 121.52, 118.95, 78.74, 55.27, 52.17, 36.20, 28.48. <sup>19</sup>F NMR (377 MHz, DMSO) δ -109.15 (t, *J* = 17.8 Hz). HRMS (ESI) calcd. for (M+Na)<sup>+</sup> C<sub>22</sub>H<sub>24</sub>F<sub>3</sub>NO<sub>8</sub>S : 542.1066, found, 542.1054.

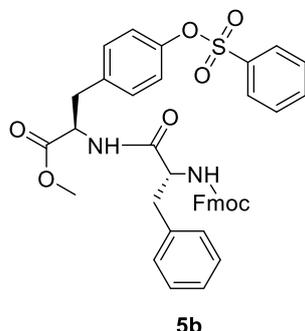
**3.4 Dipeptide scope and characterization**



**methyl (R)-2-(((R)-2-(((9H-fluoren-9-yl)methoxy)carbonyl)amino)-4-methylpentanamido)-3-(4-((phenylsulfonyl)oxy)phenyl)propanoate(5a);**

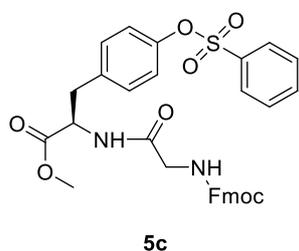
83.2 mg (yield: 62%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.37 (d, *J* = 7.5 Hz, 1H), 7.91 (d, *J* = 8.5 Hz, 2H), 7.83 – 7.80 (m, 2H), 7.78 (d, *J* = 7.4 Hz, 1H), 7.73 (dd, *J* = 7.5, 4.0 Hz, 2H), 7.66 – 7.61 (m, 2H), 7.47 (d, *J* = 8.5 Hz, 1H), 7.43 (t, *J* = 7.5 Hz, 2H), 7.33 (t, *J* = 8.6 Hz, 2H), 7.23 (d, *J* = 8.8 Hz, 2H), 6.90 (d, *J* = 8.7 Hz, 2H), 4.49 – 4.42 (m, 1H), 4.34 – 4.28 (m, 1H), 4.27 – 4.19 (m, 2H), 4.10 – 4.04 (m, 1H), 3.55 (s, 3H), 3.04 – 2.91 (m, 2H), 1.59 (dd, *J* = 17.1, 10.4 Hz, 1H), 1.48 – 1.40 (m, 1H), 1.39 – 1.32 (m, 1H), 0.89 (d, *J* = 6.7 Hz, 3H), 0.85 (d, *J* = 6.5 Hz, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.99, 172.15, 156.26, 148.13, 144.42, 144.17, 141.21,

141.19, 137.08, 135.40, 134.76, 131.12, 130.20, 128.62, 128.12, 127.52, 125.76, 122.18, 120.60, 120.58, 65.99, 53.67, 53.23, 52.27, 47.16, 41.10, 36.17, 24.55, 23.45, 21.94. HRMS (ESI) cald. for (M+Na)<sup>+</sup> C<sub>37</sub>H<sub>38</sub>N<sub>2</sub>O<sub>8</sub>S : 693.2241, found, 693.2244.



**methyl (R)-2-((R)-2-(((9H-fluoren-9-yl)methoxy)carbonyl)amino)-3-phenylpropyl anamido)-3-(4-((phenylsulfonyl)oxy)phenyl)propanoate(5b);**

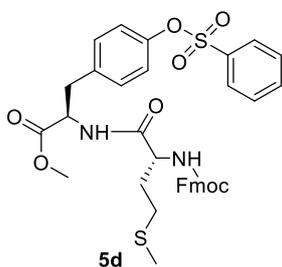
74.7 mg (yield: 53%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.50 (d, *J* = 7.6 Hz, 1H), 7.88 (d, *J* = 7.5 Hz, 2H), 7.81 (d, *J* = 10.6 Hz, 2H), 7.74 (d, *J* = 8.8 Hz, 1H), 7.64 (d, *J* = 10.2 Hz, 2H), 7.62 – 7.57 (m, 3H), 7.41 (t, *J* = 7.5 Hz, 2H), 7.32 (d, *J* = 8.7 Hz, 3H), 7.31 – 7.28 (m, 2H), 7.25 (t, *J* = 8.5 Hz, 4H), 7.19 (t, *J* = 7.2 Hz, 1H), 6.95 – 6.90 (m, 2H), 4.59 – 4.49 (m, 1H), 4.36 – 4.28 (m, 1H), 4.19 (d, *J* = 12.4 Hz, 1H), 4.14 (t, *J* = 6.2 Hz, 2H), 3.57 (s, 3H), 3.10 – 3.02 (m, 1H), 3.01 – 2.94 (m, 2H), 2.77 (dd, *J* = 13.8, 10.6 Hz, 1H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.27, 172.08, 156.19, 148.19, 144.25, 144.15, 141.13, 138.51, 137.01, 135.36, 134.77, 131.14, 130.17, 129.71, 128.62, 128.51, 128.09, 127.52, 126.74, 125.72, 122.26, 120.55, 64.80, 56.34, 53.83, 52.35, 47.02, 37.37, 34.20. HRMS (ESI) cald. for (M+Na)<sup>+</sup> C<sub>40</sub>H<sub>36</sub>N<sub>2</sub>O<sub>8</sub>S : 727.2084, found, 727.2081.



**methyl (R)-2-(2-(((9H-fluoren-9-yl)methoxy)carbonyl)amino)acetamido)-3-(4-((phenylsulfonyl)oxy)phenyl)propanoate(5c);**

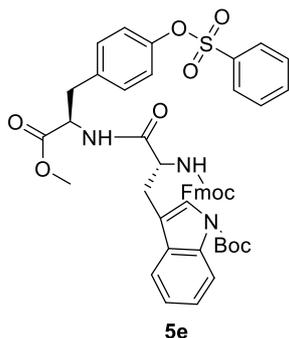
78.6 mg (yield: 64%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.32 (d, *J* = 7.8 Hz, 1H), 7.90 (s, 1H), 7.88 (s, 1H), 7.85 – 7.81 (m, 2H), 7.80 – 7.77 (m,

1H), 7.72 (d,  $J = 7.5$  Hz, 2H), 7.65 (t,  $J = 8.0$  Hz, 2H), 7.49 (t,  $J = 6.2$  Hz, 1H), 7.43 (d,  $J = 7.5$  Hz, 2H), 7.33 (t,  $J = 6.8$  Hz, 2H), 7.21 (d,  $J = 8.6$  Hz, 2H), 6.92 (d,  $J = 8.6$  Hz, 2H), 4.51 – 4.44 (m, 1H), 4.29 (d,  $J = 8.0$  Hz, 2H), 4.23 (d,  $J = 6.2$  Hz, 1H), 3.62 (t,  $J = 5.1$  Hz, 2H), 3.57 (s, 3H), 3.01 (dd,  $J = 13.8, 5.9$  Hz, 1H), 2.91 (dd,  $J = 13.8, 8.9$  Hz, 1H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  172.17, 169.66, 156.90, 148.17, 144.30, 141.19, 137.02, 135.43, 134.73, 131.15, 130.22, 128.65, 128.11, 127.55, 125.72, 122.26, 120.59, 66.20, 53.78, 52.35, 47.07, 43.49, 36.42. HRMS (ESI) cald. for  $(\text{M}+\text{Na})^+$   $\text{C}_{33}\text{H}_{30}\text{N}_2\text{O}_8\text{S}$  : 637.1615, found, 637.1619.



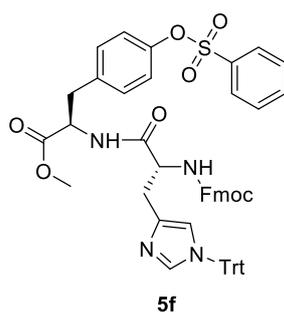
**methyl (R)-2-((R)-2-(((9H-fluoren-9-yl)methoxy)carbonyl)amino)-4-(methylthio)butanamido)-3-(4-((phenylsulfonyl)oxy)phenyl)propanoate(5d);**

64.7 mg (yield: 48%, 0.2 mmol scale), white solid.  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  8.46 (d,  $J = 8.2$  Hz, 1H), 7.91 (d,  $J = 7.4$  Hz, 2H), 7.86 – 7.83 (m, 2H), 7.80 (d,  $J = 6.1$  Hz, 1H), 7.75 (d,  $J = 7.5$  Hz, 2H), 7.68 – 7.64 (m, 2H), 7.54 (d,  $J = 8.5$  Hz, 1H), 7.43 (t,  $J = 7.5$  Hz, 2H), 7.34 (t,  $J = 7.4$  Hz, 2H), 7.23 (d,  $J = 8.7$  Hz, 2H), 6.89 (d,  $J = 8.7$  Hz, 2H), 4.53 – 4.46 (m, 1H), 4.30 (d,  $J = 12.7$  Hz, 1H), 4.25 – 4.21 (m, 2H), 4.12 (td,  $J = 8.6, 5.3$  Hz, 1H), 3.62 (s, 3H), 3.06 (dd,  $J = 13.7, 5.0$  Hz, 1H), 2.89 (dd,  $J = 13.7, 10.1$  Hz, 1H), 2.36 – 2.30 (m, 2H), 2.01 (s, 3H), 1.70 – 1.60 (m, 2H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  144.34, 141.18, 137.14, 135.42, 134.82, 131.22, 130.23, 128.61, 128.13, 127.53, 120.60, 66.16, 53.99, 53.58, 52.44, 47.81, 37.13, 33.77, 31.09, 14.96. HRMS (ESI) cald. for  $(\text{M}+\text{Na})^+$   $\text{C}_{36}\text{H}_{36}\text{N}_2\text{O}_8\text{S}_2$  : 711.1805, found, 711.1811.



**tert-butyl 3-((R)-2-(((9H-fluoren-9-yl)methoxy)carbonyl)amino)-3-(((R)-1-methoxy-1-oxo-3-(4-((phenylsulfonyl)oxy)phenyl)propan-2-yl)amino)-3-oxopropyl)-1H-indole-1-carboxylate(5e);**

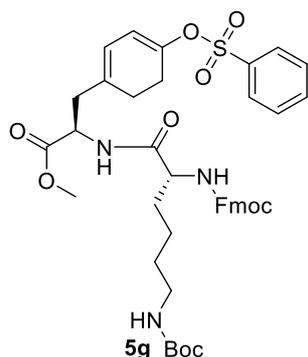
99.5 mg (yield: 59%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.64 (d, *J* = 7.7 Hz, 1H), 8.07 (d, *J* = 8.3 Hz, 1H), 7.88 (d, *J* = 7.5 Hz, 2H), 7.82 (t, *J* = 7.4 Hz, 2H), 7.75 (t, *J* = 9.1 Hz, 3H), 7.62 (t, *J* = 7.8 Hz, 5H), 7.43 – 7.32 (m, 4H), 7.30 – 7.21 (m, 5H), 6.93 (d, *J* = 8.7 Hz, 2H), 4.60 – 4.51 (m, 1H), 4.49 – 4.41 (m, 1H), 4.23 – 4.10 (m, 3H), 3.58 (s, 3H), 3.12 – 2.93 (m, 4H), 1.57 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.13, 172.04, 148.19, 144.21, 144.12, 141.15, 137.04, 135.34, 134.78, 131.15, 130.74, 130.17, 128.62, 128.08, 127.47, 125.76, 125.68, 124.78, 124.61, 122.89, 122.26, 120.57, 119.97, 117.14, 115.16, 85.07, 63.72, 55.50, 53.86, 52.34, 46.02, 35.13, 28.08, 26.35. HRMS (ESI) calcd. for (M+Na)<sup>+</sup> C<sub>47</sub>H<sub>45</sub>N<sub>3</sub>O<sub>10</sub>S :866.2717, found, 866.2711.



**methyl (R)-2-((R)-2-(((9H-fluoren-9-yl)methoxy)carbonyl)amino)-3-(1-trityl-1H-imidazol-4-yl)propanamido)-3-(4-((phenylsulfonyl)oxy)phenyl)propanoate(5f);**

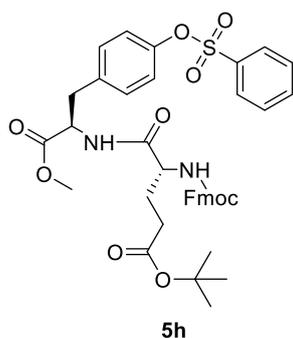
95.6 mg (yield: 51%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.40 (d, *J* = 7.6 Hz, 1H), 7.90 (d, *J* = 7.6 Hz, 2H), 7.79 (d, *J* = 7.2 Hz, 2H), 7.74 (d, *J* = 7.5 Hz, 1H), 7.66 – 7.58 (m, 4H), 7.46 – 7.38 (m, 4H), 7.33 (s, 8H), 7.27 (d, *J* = 7.9 Hz, 2H), 7.20 (d, *J* = 8.7 Hz, 3H), 7.02 (dd, *J* = 6.8, 3.0 Hz, 6H), 6.88 (d, *J* = 8.6 Hz, 2H),

6.74 (s, 1H), 4.50 – 4.43 (m, 1H), 4.35 – 4.27 (m, 1H), 4.16 (d,  $J = 6.1$  Hz, 2H), 4.11 (d,  $J = 5.9$  Hz, 1H), 3.48 (s, 3H), 3.03 – 2.88 (m, 2H), 2.88 – 2.81 (m, 1H), 2.76 – 2.67 (m, 1H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  172.06, 171.94, 156.15, 148.14, 144.20, 144.17, 142.54, 141.16, 141.13, 138.14, 137.35, 137.01, 135.37, 134.72, 131.12, 130.18, 129.70, 129.64, 128.78, 128.62, 128.59, 128.47, 128.12, 127.54, 125.76, 122.23, 120.58, 119.68, 75.11, 66.26, 54.79, 53.74, 52.30, 47.05, 36.26, 31.22. HRMS (ESI) cald. for  $(\text{M}+\text{Na})^+$   $\text{C}_{56}\text{H}_{48}\text{N}_4\text{O}_8\text{S}$  : 959.3085, found, 959.3092.



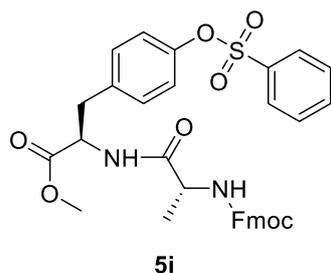
**methyl (R)-2-((R)-2-(((9H-fluoren-9-yl)methoxy)carbonyl)amino)-5-((tert-butoxycarbonyl)amino)pentanamido)-3-(4-((phenylsulfonyl)oxy)cyclohexa-1,3-dien-1-yl)propanoate(5g);**

91.4 mg (yield: 58%, 0.2 mmol scale), white solid.  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  8.31 (d,  $J = 7.6$  Hz, 1H), 7.89 (d,  $J = 6.8$  Hz, 2H), 7.81 (d,  $J = 8.6$  Hz, 2H), 7.76 (d,  $J = 7.5$  Hz, 1H), 7.74 – 7.69 (m, 2H), 7.65 – 7.60 (m, 2H), 7.44 – 7.37 (m, 3H), 7.33 (t,  $J = 7.4$  Hz, 2H), 7.22 (d,  $J = 8.7$  Hz, 2H), 6.90 (d,  $J = 8.7$  Hz, 2H), 6.78 – 6.71 (m, 1H), 4.51 – 4.42 (m, 1H), 4.31 – 4.18 (m, 3H), 4.02 – 3.95 (m, 1H), 3.55 (s, 3H), 3.01 (dd,  $J = 14.0, 5.9$  Hz, 1H), 2.93 (dd,  $J = 14.0, 8.6$  Hz, 3H), 1.58 – 1.48 (m, 2H), 1.38 (s, 9H), 1.35 – 1.18 (m, 4H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  172.62, 172.10, 156.31, 156.05, 148.16, 144.39, 144.21, 141.19, 137.02, 135.35, 134.83, 131.10, 130.17, 128.59, 128.10, 127.52, 125.75, 122.17, 120.56, 77.82, 66.08, 54.83, 53.69, 52.26, 47.15, 36.26, 32.05, 29.68, 28.74, 23.20. HRMS (ESI) cald. for  $(\text{M}+\text{Na})^+$   $\text{C}_{41}\text{H}_{45}\text{N}_3\text{O}_{10}\text{S}$  : 794.2717, found, 794.2712.



**tert-butyl (R)-4-(((9H-fluoren-9-yl)methoxy)carbonyl)amino)-5-(((R)-1-methoxy-1-oxo-3-(4-((phenylsulfonyl)oxy)phenyl)propan-2-yl)amino)-5-oxopentanoate(5h);**

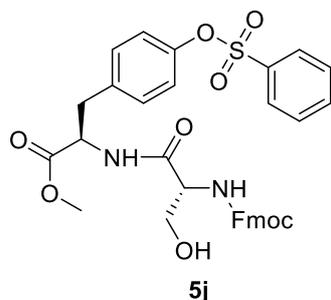
90.6 mg (yield: 61%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.39 (d, *J* = 7.5 Hz, 1H), 7.91 (d, *J* = 7.7 Hz, 2H), 7.84 – 7.72 (m, 5H), 7.64 (t, *J* = 8.0 Hz, 2H), 7.52 (d, *J* = 8.4 Hz, 1H), 7.43 (t, *J* = 7.5 Hz, 2H), 7.34 (t, *J* = 7.5 Hz, 2H), 7.23 (d, *J* = 8.7 Hz, 2H), 6.90 (d, *J* = 8.7 Hz, 2H), 4.50 – 4.44 (m, 1H), 4.33 – 4.20 (m, 3H), 4.07 – 4.01 (m, 1H), 3.57 (s, 3H), 3.06 – 2.91 (m, 2H), 2.23 (t, *J* = 8.0 Hz, 2H), 1.88 – 1.69 (m, 2H), 1.41 (s, 9H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 172.13, 172.08, 172.03, 156.28, 148.14, 144.38, 144.17, 141.20, 137.04, 135.40, 134.74, 131.13, 130.20, 128.63, 128.13, 127.53, 125.77, 122.22, 120.60, 80.17, 66.13, 54.02, 53.75, 52.32, 47.11, 36.12, 31.67, 28.21, 27.77. HRMS (ESI) calcd. for (M+Na)<sup>+</sup> C<sub>40</sub>H<sub>42</sub>N<sub>2</sub>O<sub>10</sub>S : 765.2452, found, 765.2439.



**methyl (R)-2-((R)-2-(((9H-fluoren-9-yl)methoxy)carbonyl)amino)propanamido)-3-(4-((phenylsulfonyl)oxy)phenyl)propanoate(5i);**

88.0 mg (yield: 70%, 0.2 mmol scale), white solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 8.36 (d, *J* = 8.3 Hz, 1H), 7.89 (d, *J* = 8.2 Hz, 2H), 7.84 – 7.81 (m, 2H), 7.78 (d, *J* = 7.5 Hz, 1H), 7.74 (dd, *J* = 7.4, 3.1 Hz, 2H), 7.67 – 7.62 (m, 2H), 7.47 (d, *J* = 7.9 Hz, 1H), 7.42 (t, *J* = 7.5 Hz, 2H), 7.33 (t, *J* = 7.5 Hz, 2H), 7.21 (d, *J* = 8.7 Hz, 2H), 6.89 (d, *J* = 8.6 Hz, 2H), 4.55 – 4.48 (m, 1H), 4.27 – 4.18 (m, 3H), 4.04 (q, *J* = 7.3 Hz, 1H), 3.61

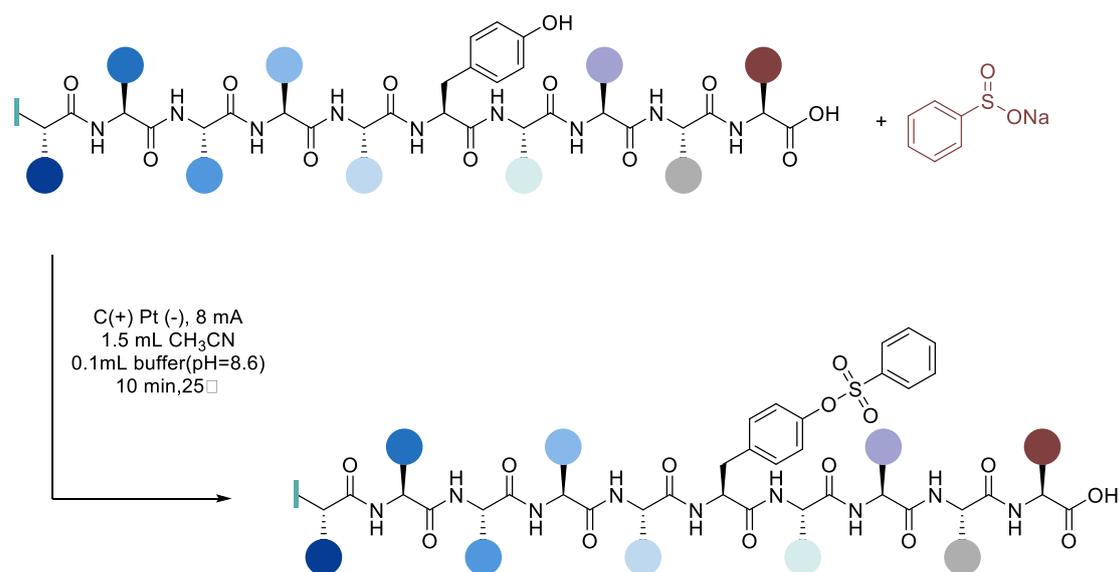
(s, 3H), 3.06 (dd,  $J = 13.8, 4.9$  Hz, 1H), 2.86 (dd,  $J = 13.8, 10.2$  Hz, 1H), 0.99 (d,  $J = 7.2$  Hz, 3H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  173.03, 172.23, 144.28, 143.05, 137.07, 134.77, 131.23, 130.21, 129.39, 128.61, 128.11, 127.75, 127.53, 125.79, 121.84, 120.56, 120.49, 110.19, 66.50, 55.37, 52.39, 50.23, 47.09, 35.19, 18.85. HRMS (ESI) cald. for  $(\text{M}+\text{Na})^+$   $\text{C}_{34}\text{H}_{32}\text{N}_2\text{O}_8\text{S}$  : 651.1771, found, 651.1778.



**methyl (R)-2-((R)-2-(((9H-fluoren-9-yl)methoxy)carbonyl)amino)-3-hydroxypropanamido)-3-(4-((phenylsulfonyl)oxy)phenyl)propanoate(5j);**

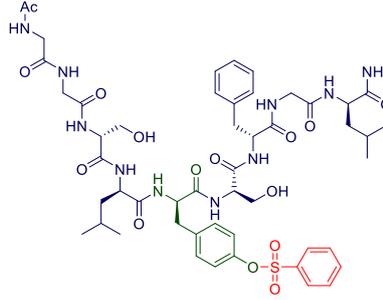
60.6 mg (yield:47%, 0.2 mmol scale), white solid.  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  8.32 (d,  $J = 7.6$  Hz, 1H), 7.90 (d,  $J = 7.5$  Hz, 2H), 7.81 (d,  $J = 6.0$  Hz, 2H), 7.77 (d,  $J = 7.4$  Hz, 1H), 7.74 (dd,  $J = 8.1, 3.8$  Hz, 2H), 7.66 – 7.62 (m, 2H), 7.42 (t,  $J = 7.0$  Hz, 2H), 7.35 – 7.30 (m, 3H), 7.21 (d,  $J = 8.7$  Hz, 2H), 4.89 (t,  $J = 5.7$  Hz, 1H), 4.50 – 4.43 (m, 1H), 4.28 (d,  $J = 5.9$  Hz, 2H), 4.22 (d,  $J = 6.1$  Hz, 1H), 4.13 – 4.07 (m, 1H), 3.58 (d,  $J = 4.7$  Hz, 1H), 3.55 (s, 3H), 3.47 (d,  $J = 11.1$  Hz, 1H), 3.04 – 2.90 (m, 2H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  171.99, 170.68, 156.38, 148.15, 144.35, 144.24, 141.19, 137.00, 135.41, 134.72, 131.19, 130.21, 128.63, 128.13, 127.56, 125.79, 122.21, 120.59, 66.21, 62.11, 57.63, 53.76, 52.34, 47.08, 36.30. HRMS (ESI) cald. for  $(\text{M}+\text{Na})^+$   $\text{C}_{34}\text{H}_{32}\text{N}_2\text{O}_9\text{S}$  : 667.1720, found, 667.1718.

### 3.5 Polypeptide scope and characterization



General Procedure for Bioconjugation of Tyrosine and Sodium benzenesulfite : In an oven-dried undivided three-necked bottle (15 mL) equipped with a stir bar, polypeptides (5 mg), Sodium benzenesulfite (10 mg), CH<sub>3</sub>CN (1.5 mL), buffer (pH =8.6, 0.1 mL ), <sup>n</sup>Bu<sub>4</sub>NBr (0.08 mmol) were combined and added. The bottle was equipped graphite rod (φ 6 mm) as the anode and platinum plate (10 mm×10 mm×0.3 mm) as the cathode and then charged. The reaction mixture was stirred and electrolyzed at constant current of 8 mA under 25°C for 10 min. After completion of the reaction, the solution was analyzed by LC-MS/MS spectroscopy. The reaction was analyzed by reversed-phase HPLC on a 250 mm long ChromCore C18 5μm column using a gradient of 5% to 50% buffer B within 30 minutes. HPLC analysis used buffers A (water + 0.1% TFA) and B (9:1 acetonitrile : water + 0.1% TFA). Conversion reported as a % conversion as determined.





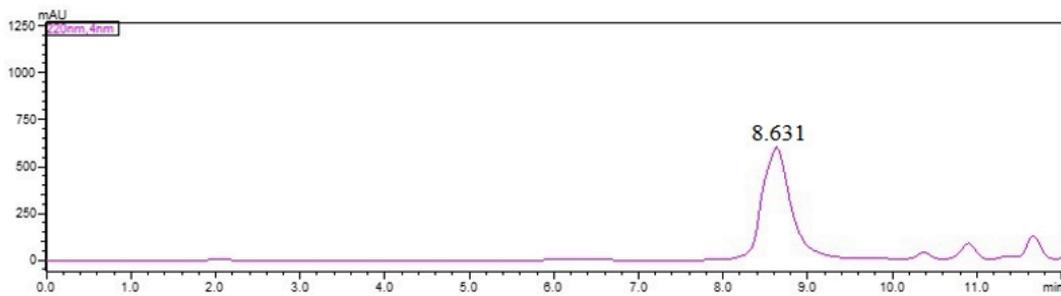
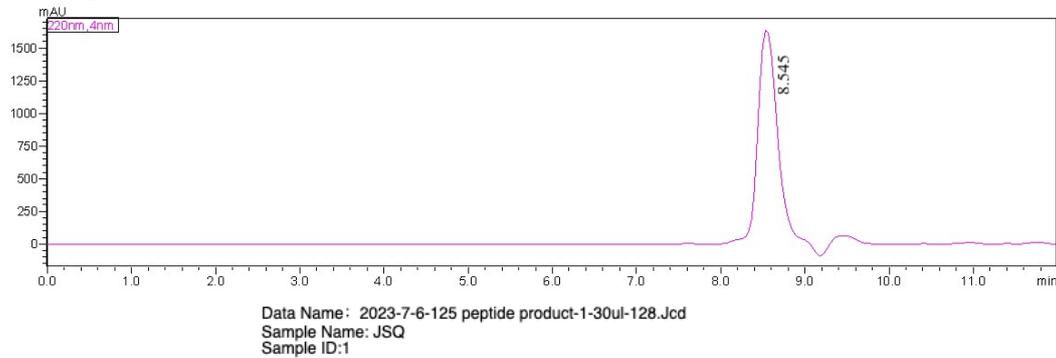
**Allatostation: GGSLYSFGL**

**HPLC: >99% conversion.**

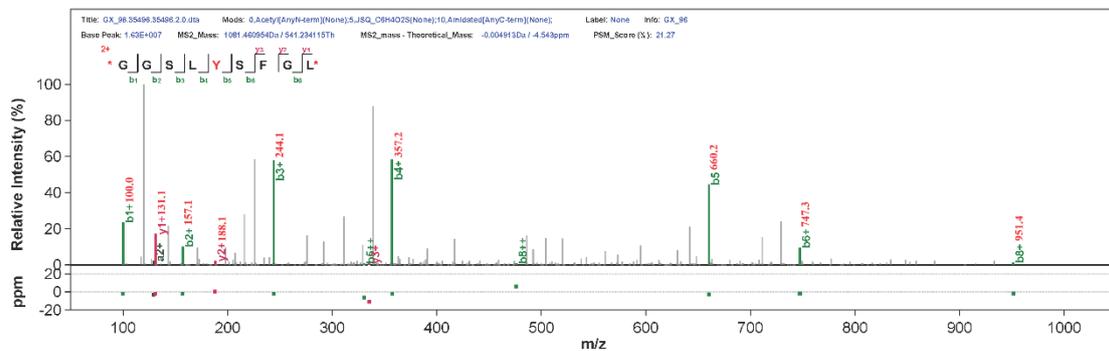
**Product 6b** is a peak that elute at 50% buffer B (9:1 acetonitrile: water + 0.1% TFA) with retention times of 8.631 min. Reactant is a peak that elutes at 50% buffer B with a retention time of 8.545 min.

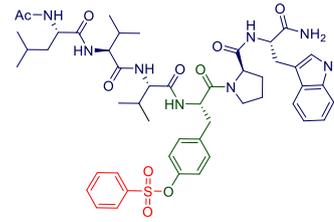
**HRMS (ESI-TOF): m/z calculated for C<sub>50</sub>H<sub>68</sub>N<sub>10</sub>O<sub>15</sub>S, [M+H]<sup>+</sup>, 1081.4737, found 1081.4736.**

**HPLC Spectra:**



**MALDI-TOF-MS/MS Spectra:**





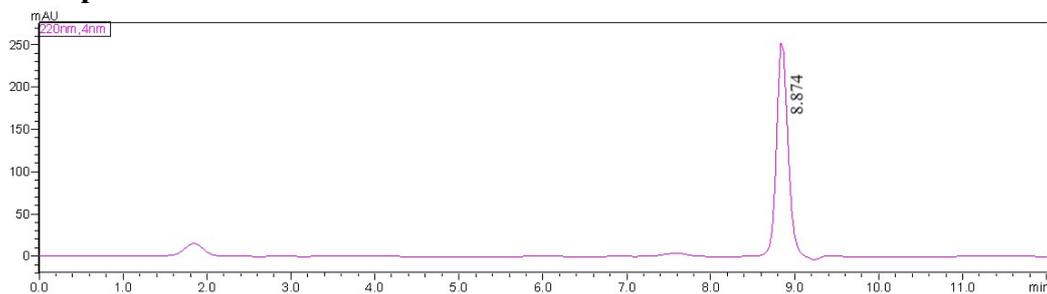
## Mylopeptide-2(MP-2):LVVYPW

HPLC: >99% conversion.

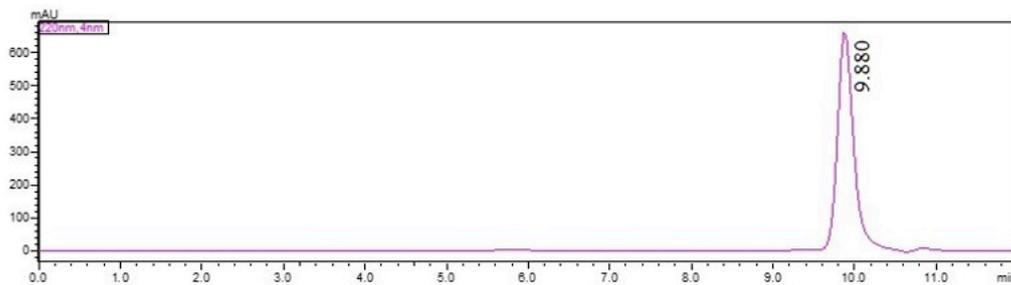
Product **6c** is a peak that elute at 50% buffer B (9:1 acetonitrile: water + 0.1% TFA) with retention times of 8.874 min. Reactant is a peak that elutes at 50% buffer B with a retention time of 9.880 min.

HRMS (ESI-TOF): m/z calculated for  $C_{49}H_{64}N_8O_{10}S$ ,  $[M+H]^+$ , 957.4539, found 957.4539.

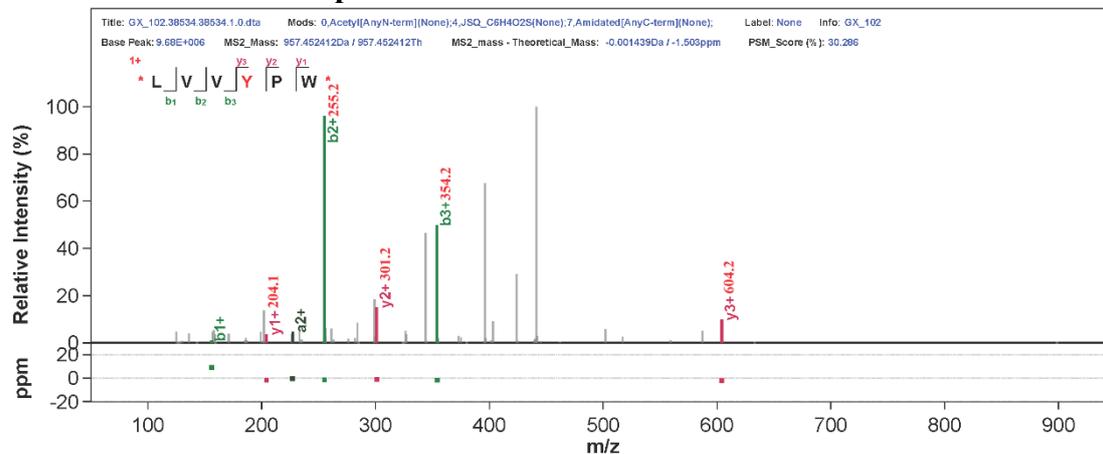
## HPLC Spectra:

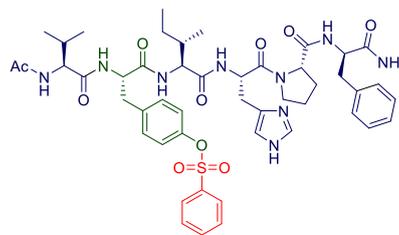


Data Name: 2023-7-11-815 peptide product-1-20ul-161.Jcd  
Sample Name: JSQ  
Sample ID:1



## MALDI-TOF-MS/MS Spectra:





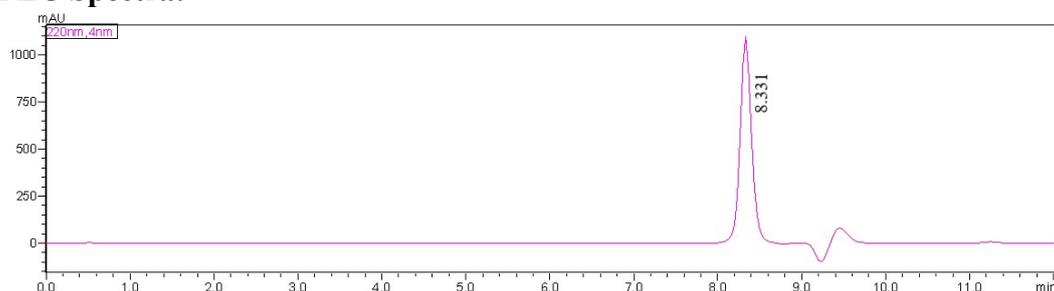
### 3-8-Angiotensin II:VYIHPF

HPLC: >99% conversion.

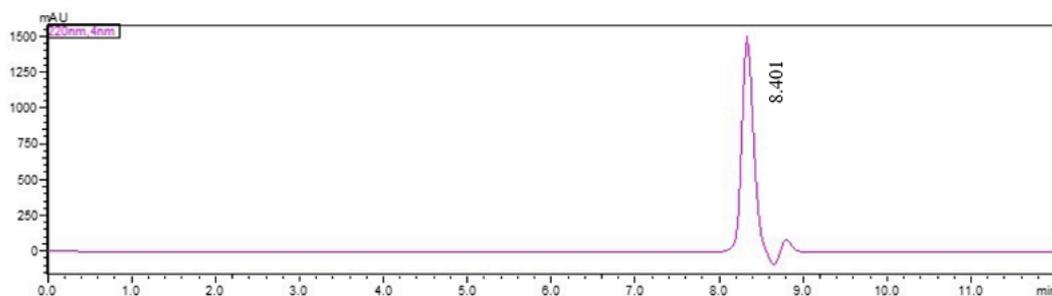
Product **6c** is a peak that elute at 50% buffer B (9:1 acetonitrile: water + 0.1% TFA) with retention times of 8.401 min. Reactant is a peak that elutes at 50% buffer B with a retention time of 8.331 min.

HRMS (ESI-TOF): m/z calculated for  $C_{48}H_{61}N_9O_{10}S$ ,  $[M+H]^+$ , 956.4334, found 956.4331.

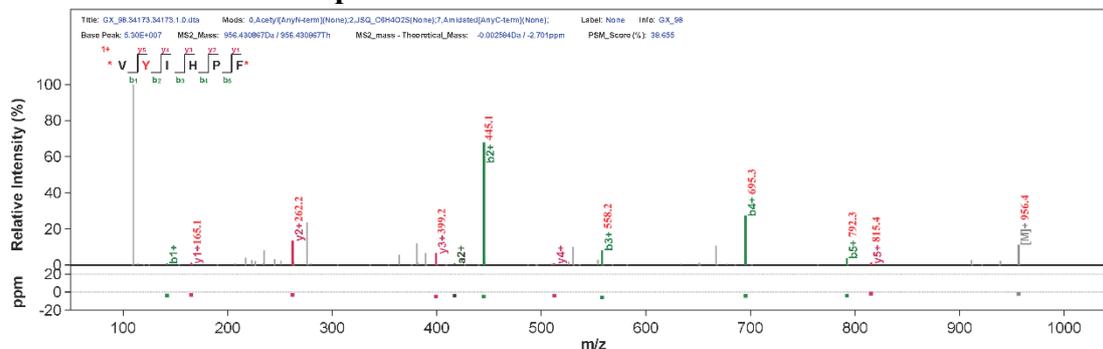
### HPLC Spectra:

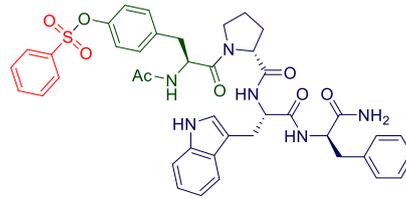


Data Name: 2023-6-29-152 peptide product-9-1-30ul-91.Jcd  
 Sample Name: JSQ  
 Sample ID:1



### MALDI-TOF-MS/MS Spectra:



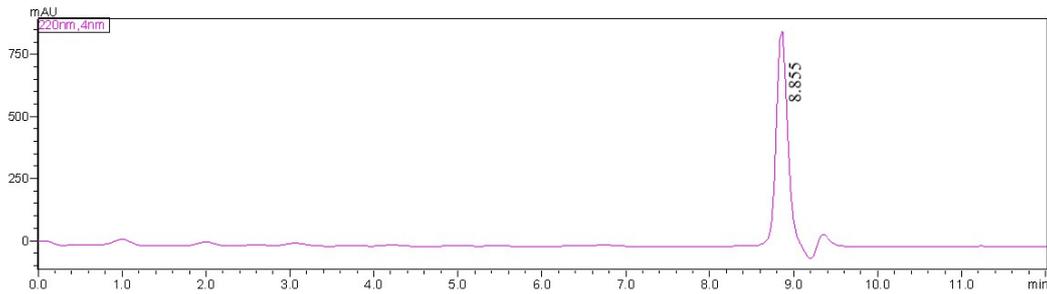


**Endomorphin 1: YPWF**  
**HPLC: >99% conversion.**

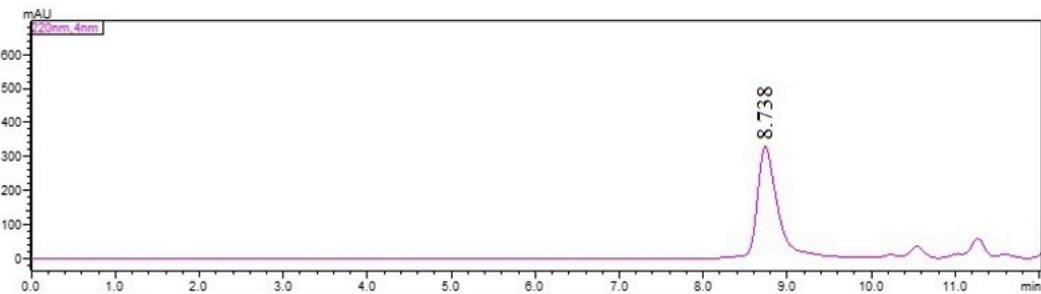
**Product 6e** is a peak that elute at 50% buffer B (9:1 acetonitrile: water + 0.1% TFA) with retention times of 8.738 min. Reactant is a peak that elutes at 50% buffer B with a retention time of 8.855 min.

**HRMS (ESI-TOF): m/z calculated for C<sub>42</sub>H<sub>44</sub>N<sub>6</sub>O<sub>8</sub>S, [M+H]<sup>+</sup>, 793.3014, found 793.3014.**

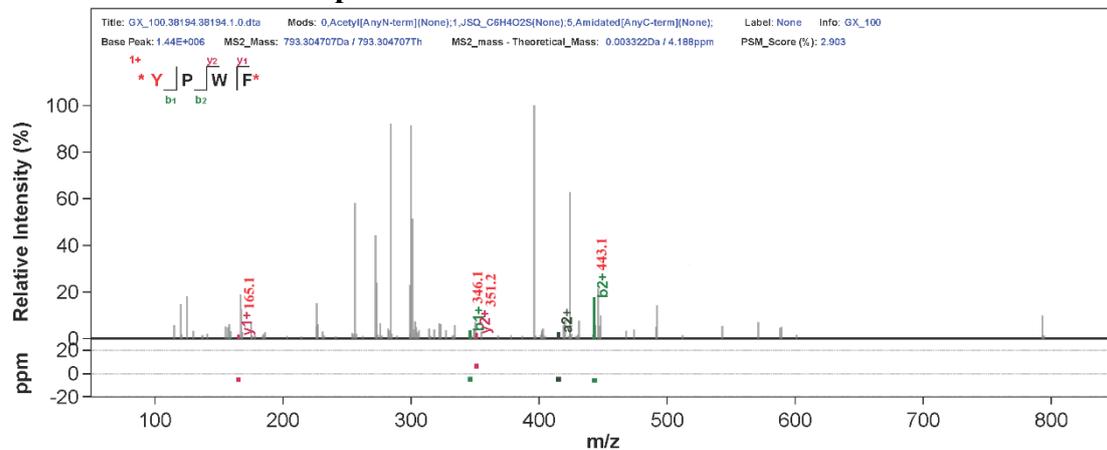
**HPLC Spectra:**



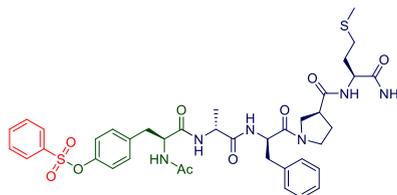
Data Name: 2023-6-29-225 peptide product-9-1-30ul-80.Jcd  
 Sample Name: JSQ  
 Sample ID:1



**MALDI-TOF-MS/MS Spectra:**



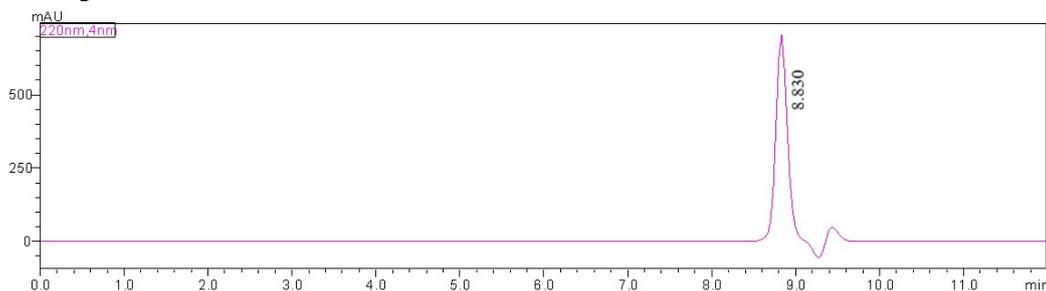
**$\beta$ -Casomorphin(1-5),amide,bovine:YAFPM**  
**HPLC: >99% conversion.**



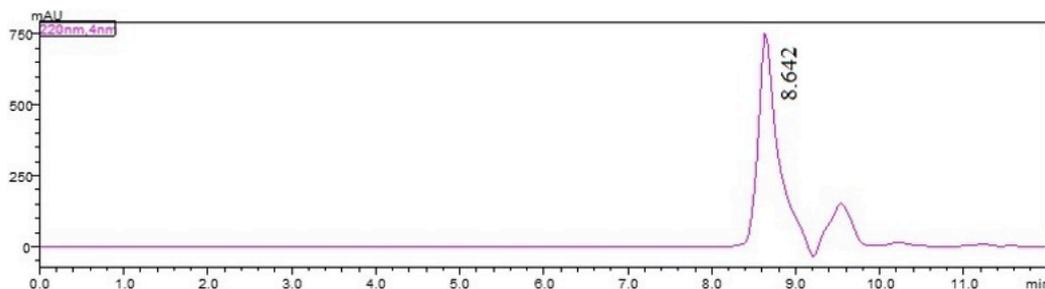
**Product 6d** is a peak that elute at 50% buffer B (9:1 acetonitrile: water + 0.1% TFA) with retention times of 8.642 min. Reactant is a peak that elutes at 50% buffer B with a retention time of 8.830 min.

**HRMS (ESI-TOF): m/z calculated for  $C_{39}H_{48}N_6O_9S_2$ ,  $[M+H]^+$ , 809.2997, found 809.2997.**

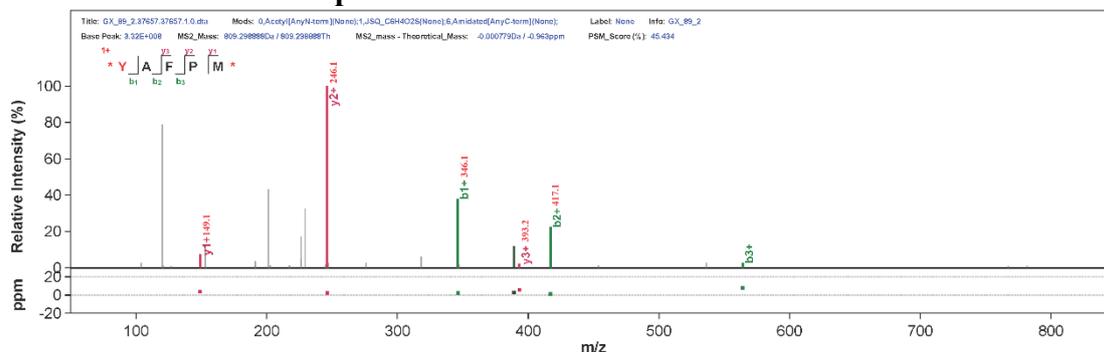
**HPLC Spectra:**

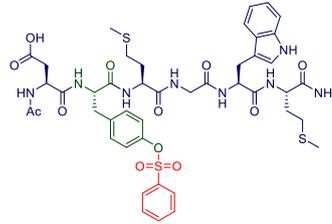


Data Name: 2023-6-20-17 peptide product-9-1-5ul-47.Jcd  
Sample Name: JSQ  
Sample ID:1



**MALDI-TOF-MS/MS Spectra:**





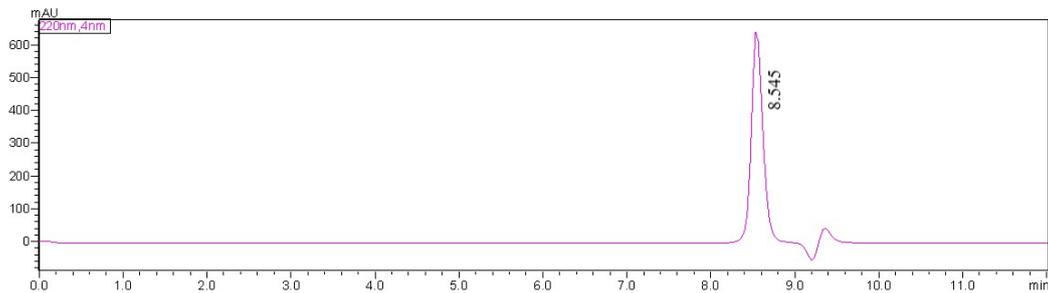
**ω-Conotoxin MVIIC: DYMGWM**

**HPLC: >99% conversion.**

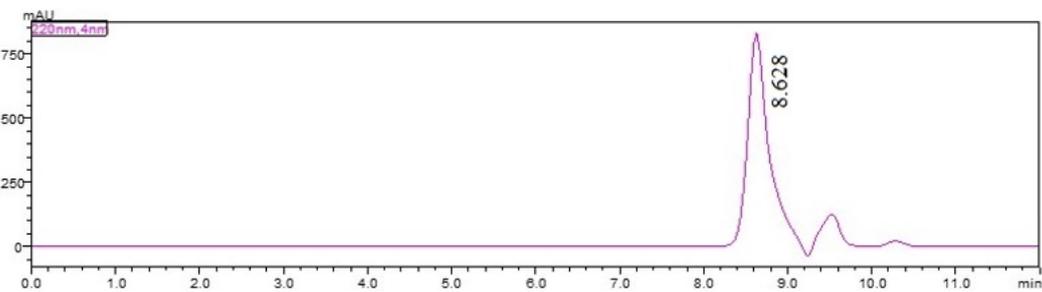
**Product 6f** is a peak that elute at 50% buffer B (9:1 acetonitrile: water + 0.1% TFA) with retention times of 8.628 min. Reactant is a peak that elutes at 50% buffer B with a retention time of 8.545 min.

**HRMS (ESI-TOF): m/z calculated for C<sub>44</sub>H<sub>54</sub>N<sub>8</sub>O<sub>12</sub>S<sub>3</sub>, [M+H]<sup>+</sup>, 983.3096, found 983.3096**

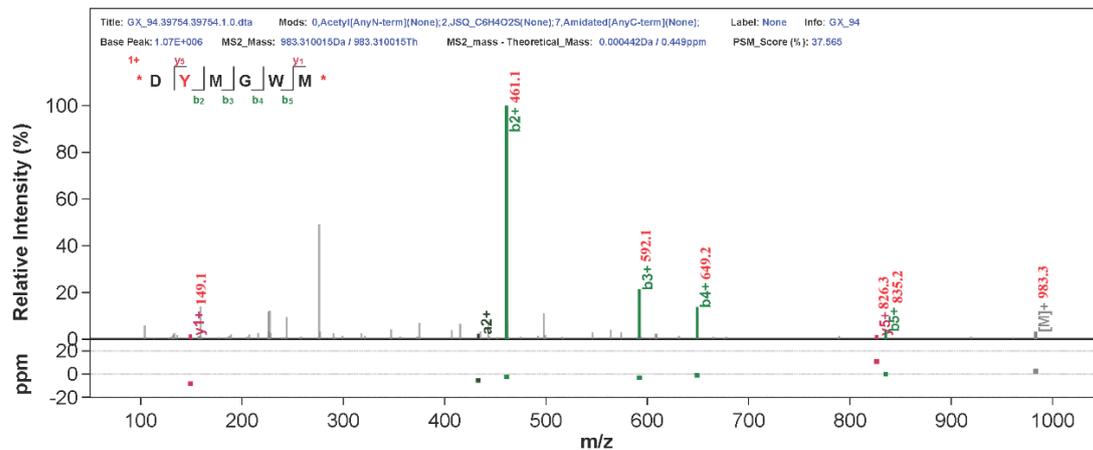
**HPLC Spectra:**

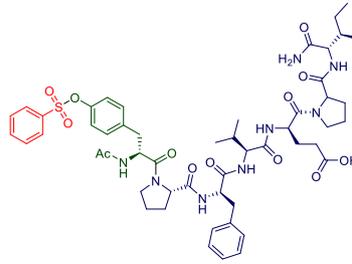


Data Name: 2023-6-14-18 peptide product-9-1-20ul-27.Jcd  
 Sample Name: JSQ  
 Sample ID:1



**MALDI-TOF-MS/MS Spectra:**





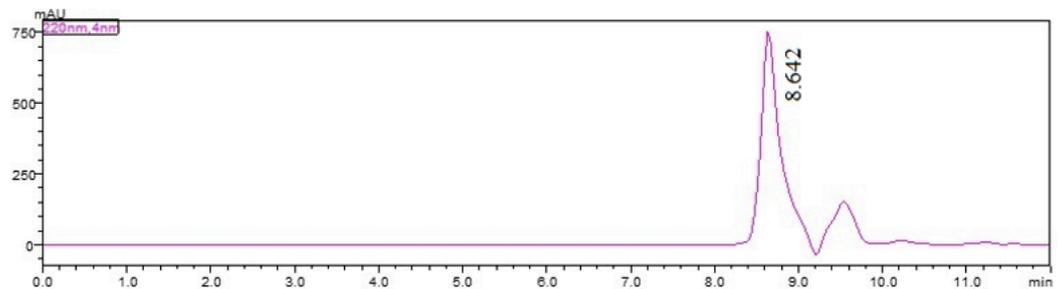
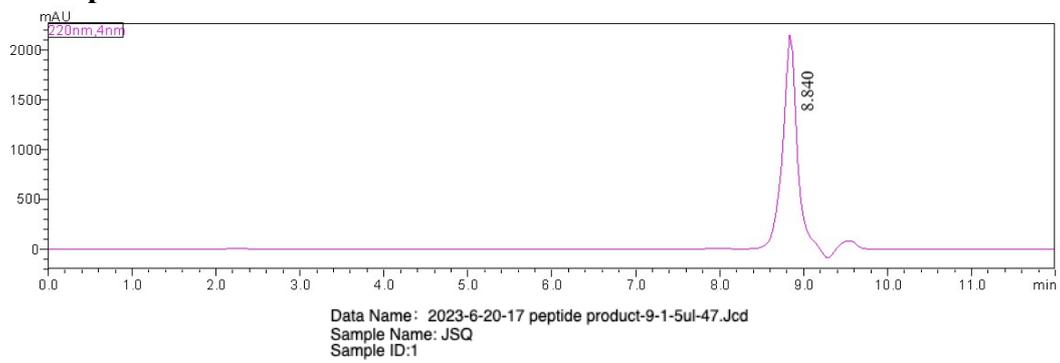
**$\beta$ -Casomorphin: YPFVEPI**

**HPLC: >99% conversion.**

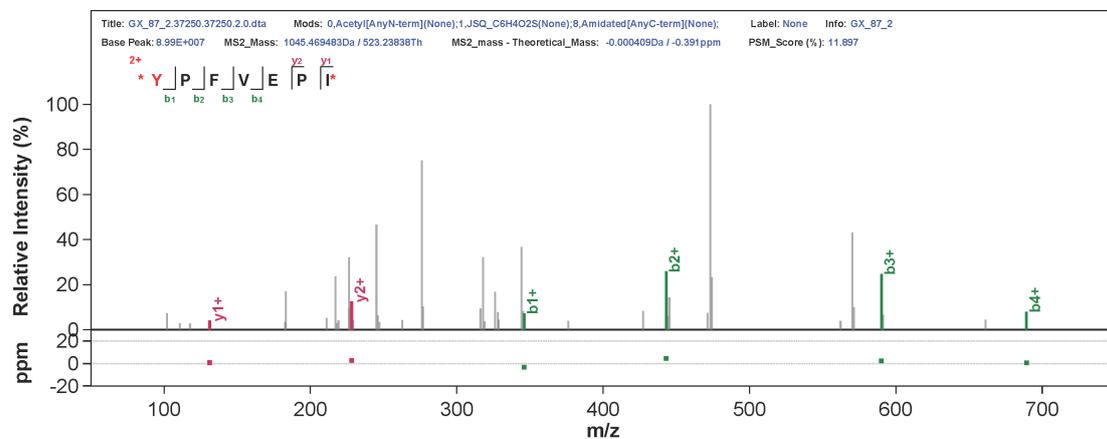
**Product 6h** is a peak that elute at 50% buffer B (9:1 acetonitrile: water + 0.1% TFA) with retention times of 8.642 min. Reactant is a peak that elutes at 50% buffer B with a retention time of 8.840 min.

**HRMS (ESI-TOF): m/z calculated for C<sub>52</sub>H<sub>68</sub>N<sub>8</sub>O<sub>13</sub>S, [M+H]<sup>+</sup>, 1045.4699, found 1045.4700.**

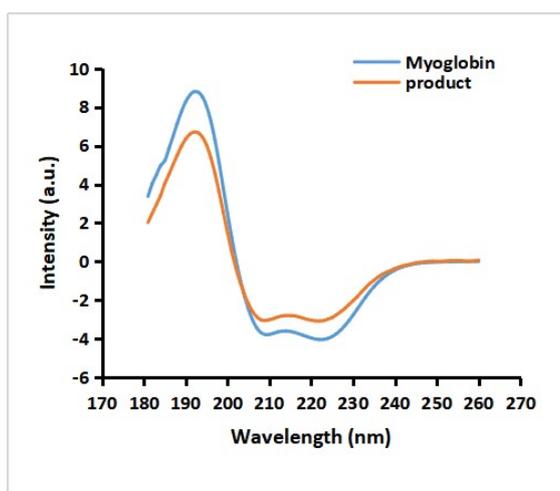
**HPLC Spectra:**



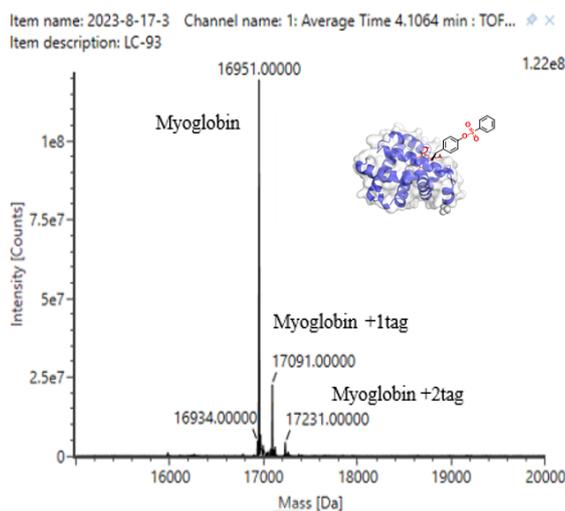
**MALDI-TOF-MS/MS Spectra:**



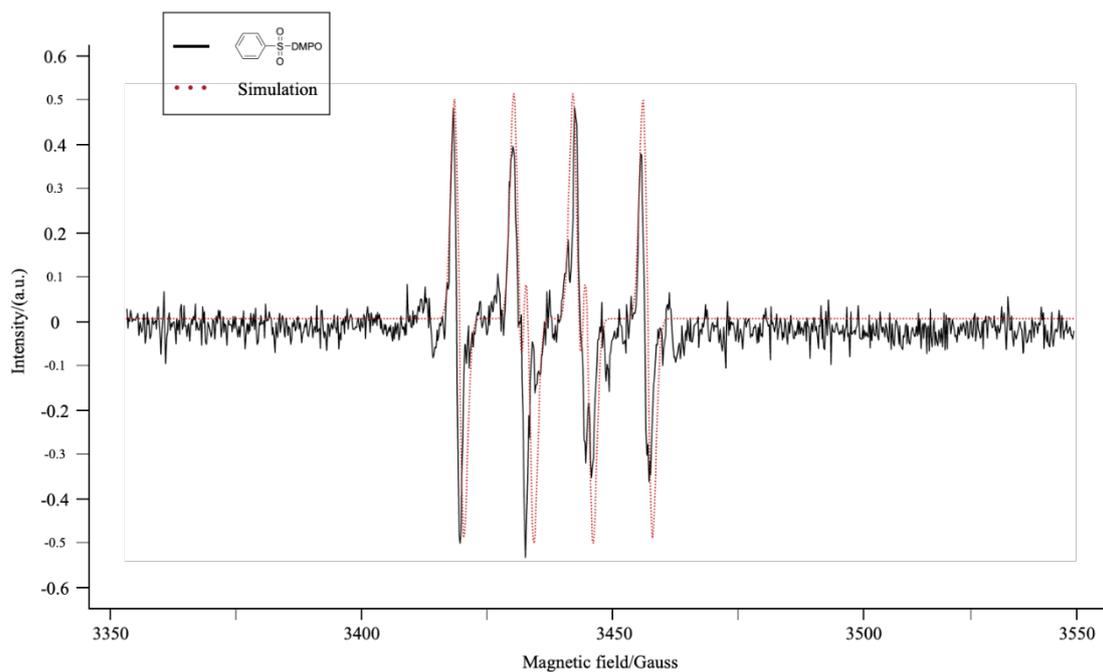
**Synthesis of 7a:** In an oven-dried undivided three-necked bottle (10 mL) equipped with a stir bar, Myoglobin (5 mg), Sodium benzenesulphonate (10 mg),  ${}^n\text{Bu}_4\text{NBr}$  (0.08mmol), MeCN / buffer(pH = 8.6) (1.5 mL / 0.1 mL) were combined and added. The bottle was equipped graphite rod ( $\phi$  6 mm) as the anode and platinum plate (10 mm $\times$ 10 mm $\times$ 0.3 mm) as the cathode and then charged. The reaction mixture was stirred and electrolyzed at constant current of 8 mA under 25 $^\circ\text{C}$  for 10 min. After completion of the reaction, the solution was analyzed by Maldi-Tof MS.



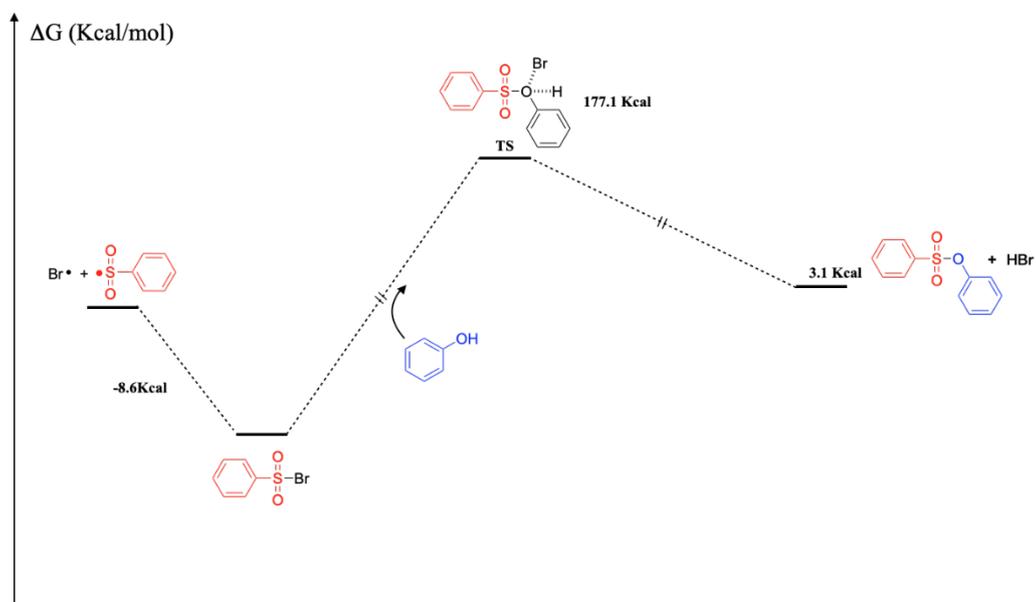
Comparison of CD spectra between Myoglobin and product (100 $\mu\text{g}/\text{mL}$  in buffer).



### 3.6 EPR and DFT

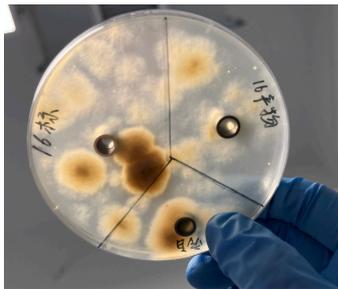


To gain additional insights into the mechanism for this reaction, we conducted DFT calculations for this reaction. The reaction diagrams were calculated at the B3LYP with 6-31G level for C and H, 6-31G+ level for S, O, and Br atoms of theory.

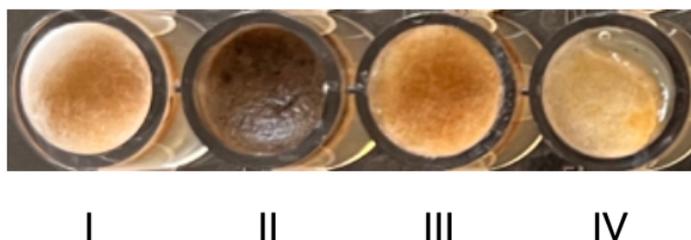


### 3.7 Anti-fungal experiment of benzenesulfonate-labeled peptide

To assess the in vivo antifungal activity of benzenesulfonate-labeled peptide, the indicator strain *Alternaria alternata* was cultivated in potato dextrose broth for 24 h, and 100  $\mu$ L culture was added to molten potato dextrose agar cooled below 55  $^{\circ}$ C. 100  $\mu$ L product 6h (3 mg/mL), substrate, solvent (water) was added to oxford cup placed onto the solidified agar respectively. Inhibition zones were recorded after 48 h at 30  $^{\circ}$ C



To assess the in vivo antifungal activity of 6h, the indicator strain *Alternaria alternata* was cultivated in potato dextrose broth for 24 h, and 100  $\mu$ L culture was added to molten potato dextrose agar (PDA) cooled below 55  $^{\circ}$ C. 100  $\mu$ L 6h (3 mg/mL), substrate, solvent (water) was added to oxford cup placed onto the solidified agar respectively. Inhibition zones were recorded after 48 h at 30  $^{\circ}$ C.



I: MeOH; II: blank; III: b-Casomorphin; IV: 6h.

**MIC Experiment:** To determine the minimal inhibitory concentration of 6h, 10  $\mu$ L dilutions ranging from 0.1-3 mg/mL were added to 200 $\mu$ L PDA in 96 well plates. At the same time, no addition, added water or substrate as positive controls. The plates were then kept at 4  $^{\circ}$ C for 4 hours to allow the diffusion of additions. The indicator strain *Alternaria alternata* was then inoculated on the surface of the agar and incubated at 28  $^{\circ}$ C for 48 h. From the results, we found that the antifungal ability of 6h increases with increasing concentration.



A

B

C

D

E

F

A: Water; B: no addition; C: b-Casomorphin; D:1mg/mL 6h; E: 2 mg/mL 6h; F: 3 mg/mL 6h.

#### 4.References

- {1} R. A. Serwa, J.-M. Swiecicki, D. Homann, C. P. R. Hackenberger, Phosphoramidate peptide synthesis by solution- and solid-phase Staudinger-phosphite reactions. *J. Pept. Sci.* 2010, 16, 563–567.
- {2} Alam J, Keller T H, Loh T P. Functionalization of peptides and proteins by Mukaiyamaaldol reaction. *J. Am. Chem. Soc.* 2010, 132, 9546-9548.



