Supporting Information

Pillar[5]arene-based supramolecular pseudorotaxane polymer material for ultra-sensitive detection of Fe³⁺ and F⁻

Wen-Bo Zhu *a, Tai-Bao wei b*, Hao-Bin Hu a, Zhi-Jun Li a, Yu-Quan Zhang a,

Yan-Chun Li^a, Liang Zhang^a, Xiao-Wei Zhang^a

^a Key Gansu Key Laboratory of Protection and Utilization for Biological Resources and Ecological Restoration, College of Chemistry and Chemical Engineering, Longdong University, Qingyang, Gansu, 745000, P. R. China. E-mail: zwb-00@163.com

^b Key Laboratory of Eco-Environment-Related Polymer Materials, Ministry of Education of China; Key Laboratory of Polymer Materials of Gansu Province; College of Chemistry and Chemical Engineering, Northwest Normal University, Lanzhou, Gansu, 730070, P. R. China. E-mail: weitaibao@126.com

1. Materials and methods

All reagents and solvents were commercially available at analytical grade and were used without further purification. All cations were purchased from Alfa Aesar and used as received. Fresh double distilled water was used throughout the experiment. ¹H NMR (600 MHz) and ¹³C NMR spectra (151 MHz) were carried out with a Mercury-600 BB spectrometer. Chemical shifts are reported in ppm downfield from tetramethylsilane (TMS, δ scale with solvent resonances as internal standards). High-resolution mass spectra were recorded with a Bruker Esquire 3000 Plus spectrometer. Fluorescence spectra were recorded on a Shimadzu RF-5301PC spectrofluorophotometer. The infrared spectra were performed on a Digilab FTS-3000 FT-IR spectrophotometer. The X-ray diffraction (XRD) pattern was generated using a Rigaku RINT2000 diffract meter equipped (copper target; $\lambda = 0.154073$ nm). Scanning electron microscopy (SEM) images of the xerogels were investigated using JSM-6701F instrument. Melting points were measured on an X-4 digital melting-point apparatus (uncorrected).



2. Synthesis of functionalized pillar[5]arene P5C

Scheme S1 Synthesis of functionalized pillar[5]arene P5C.

Synthesis of compound M1

Compound **M1** was synthesized as previously reported^[S1]. The ethyl 2mercaptoacetate functionalized pillar[5]arene (0.62 g, 0.68 mmol), hydrazine hydrate (3 mL, 93.6 mmol) and ethanol (30.0 mL) were mixed in a 100 mL round-bottom flask and stirred at 72 °C for 8h. After the solution cooled to room temperature, the crude product was filtered and then recrystallized with ethanol to get the solid compound **M1** as a white solid (0.46 g, yield 76%). Mp: 107-109 °C. ¹H NMR (CDCl₃, 600 MHz), δ /ppm: 6.78 - 6.72 (m, 10H), 4.02 - 3.98 (q, *J* = 7.1 Hz, 2H), 3.84 - 3.83 (t, *J* = 6.0 Hz, 2H), 3.77 - 3.75 (dd, *J* = 8.8, 5.8 Hz, 10H), 3.65 - 3.61 (m, 27H), 3.2 (s, 2H), 2.72 -2.69 (t, *J* = 7.0 Hz, 2H), 1.84 - 1.83 (d, *J* = 5.3 Hz, 2H), 1.08 - 1.05 (t, *J* = 7.1 Hz, 2H).

$\begin{array}{c} 6.79\\ 6.77\\ 6.77\\ 6.77\\ 6.77\\ 6.77\\ 6.74\\$



Fig. S1 ¹H NMR spectra (600 MHz, CDCl₃) of compound M1.

Synthesis of functionalized pillar[5]arene P5C.

The mixture of coumarin-3-carboxylic acid (0.5g, 2.5 mmol), sulfoxide chloride (20 mL) and DMF (20 uL) as catalyst was refluxed for 6 h. Then the solvent was evaporated under vacuum to give a white solid, and the product was put into the next reaction without purification. The product and compound M1 (0.4g, 0.45 mmol) were dissolved in CH₂Cl₂, with triethylamine (60 uL) as a catalyst for 30 min at 0-10 $^{\circ}$ C and then returned to room temperature. After the reaction completed, the mixture solution was washed with water (100 mL), and the organic phase was dried with anhydrous sodium sulfate, evaporated, and recrystallized with EtOH/H₂O to give a yellow solid. Mp: 107-109 °C. ¹H NMR (600 MHz, CDCl₃) δ/ppm: 10.93 (d, *J* = 4.6 Hz, 1H), 9.24 (s, 1H), 8.83 (s, 1H), 7.71 – 7.67 (m, 2H), 7.44 – 7.37 (m, 2H), 6.76 – 6.71 (m, 10H), 3.86 - 3.84 (t, J = 5.9 Hz, 2H), 3.77 - 3.72 (m, 10H), 3.65 - 3.59 (m, 27H), 3.33 (s, 2H), 2.70 - 2.68 (t, J = 7.0 Hz, 2H), 1.88 - 1.82 (m, 4H). ¹³C NMR (151 MHz, CDCl₃) δ/ppm: 165.54, 160.61, 158.32, 154.54, 150.84, 150.81, 150.79, 150.76, 149.93, 148.99, 134.64, 129.95, 128.29, 128.22, 128.15, 125.46, 118.33, 116.84, 115.04, 114.22, 114.12, 114.00, 67.79, 55.88, 55.86, 55.82, 55.80, 55.76, 55.72, 40.94, 29.71, 29.62, 28.80, 25.92. ESI-MS m/z: (M+Na)⁺ Calcd for C₆₀H₆₄N₂NaO₁₄S 1091.3976;



Fig. S2 ¹H NMR spectra (600 MHz, CDCl₃) of compound P5C.



Fig. S3 ¹³C NMR spectra (151 MHz, CDCl₃) of compound P5C.



Fig. S4 Mass data of compound P5C.

Entry	Solvent	State ^a	CGC ^b (%)	T _{gel} ^c (°C,wt%)
1	Water	Р	\	١
2	Acetone	S	\	١
3	Methanol	S	١	١
4	Ethanol	S	١	١
5	Isopropanol	Р	١	١
6	Isoamylol	Р	١	١
7	Acetonitrile	S	١	١
8	THF	S	\	١
9	DMF	S	\	١
10	DMSO	S	١	١
11	n-hexane	Р	١	١
12	CH_2Cl_2	S	١	١
13	CHCl ₃	S	١	١
14	Ethyl acetate	S	\	٨
15	Cyclohexanol	G	10	58-60°C

 Table S1 Gelation Properties of Organogel PCDP-G in Organic Solvents.

State: $n_{P5C}/n_{DP5} = 2/1$

^aG, P and S denote gelation, precipitation and solution, respectively.

^bThe critical gelation concentration (wt%, 10mg/ml = 1.0%).

^cThe gelation temperature (°C)



Fig. S5 Temperature-dependent fluorescence spectra of PCDP-G (in cyclohexanol, 10% (w/v)) during the gelation process ($\lambda_{ex} = 365$ nm).



Fig. S6 Cycle index of the sol-gel transition process.



Fig. S7 Partial concentrations-dependent ¹HNMR spectra of (a) free P5C with different concentrations of DP5 in DMSO- d_6 , from bottom to top: 3.7×10^{-3} M, 9.3×10^{-3} M, 1.9×10^{-2} M, 2.8×10^{-2} M, 3.7×10^{-2} M.





Fig. S9 SEM images of (a) xerogel PCDP-G; (b) PCDP-GFe and

(c) **PCDP-GFe** + F⁻.



Fig. S10 The fluorescence spectra of PCDP-G in cyclohexanol solution with increasing concentrations of Fe³⁺ (0.1 M, water solution), $\lambda_{ex} = 365$ nm.



Fig. S11 The photograph of the linear range for Fe³⁺.

The result of the analysis as follows:

Linear Equation: $Y = -30947.67 \times X + 394.74$ $R^2 = 0.98287$

$$S = 3.0947 \times 10^{10}$$
$$\delta = \sqrt{\frac{\sum_{i=1}^{N} (Fi - \overline{F})}{N - 1}} = 4.29$$
(N = 20) K = 3



Fig. S12 The fluorescence spectra of PCDP-GFe in cyclohexanol solution with

increasing concentration of F⁻ (0.1 M, water solution), $\lambda_{ex} = 365$ nm.



Fig. S13 The photograph of the linear range for F⁻.

The result of the analysis as follows:

Linear Equation: $Y = 18858.89 \times X + 158.68$ $R^2 = 0.97902$

 $S = 1.8858 \times 10^{10}$

 $\delta = 4.29$ (N = 20) K = 3

 $LOD = K \times \delta/S = 0.682 \text{ nM}$



Fig. S13 Fluorescent "on-off-on" cycles of PCDP-G, controlled by the alternative addition of Fe³⁺ and F⁻ ($\lambda_{ex} = 365$ nm).

Table S2. Comparison the LODs of Different Fluorescence Sensors

Refs	Solvent	LOD
S2	aqueous solution	900 nM
S3	deionized water	450 nM
S4	CH ₃ CN-H ₂ O (1:1, v/v)	261 nM
S5	drinking water	240 nM
This work	cyclohexanol solution	0.416 nM

for Fe³⁺

Table S3. Comparison the LODs of Different Fluorescence Sensors

Refs	Solvent	LOD
S 6	DMF/H ₂ O (v:v=1:1)	9.24 nM
S7	DMSO	1.44 µM
S 8	DMSO	289 ppm
S9	DMSO - H ₂ O (1:1, v/v)	108 nM

for F-

Reference:

- S1 W.-B. Zhu, T.-B. Wei, Y.-Q. Fan, W.-J. Qu, W. Zhu, X.-Q. Ma, H. Yao, Y.-M. Zhang and Q. Lin, A pillar[5]arene-based and OH⁻ dependent dual-channel supramolecular chemosensor for recyclable CO₂ gas detection: High sensitive and selective off-onoff response, *Dyes Pigm.*, 2020, 174, 108073.
- S2 C.-X. Yang, H.-B. Ren and X.-P. Yan, Fluorescent Metal–Organic Framework MIL-53(Al) for Highly Selective and Sensitive Detection of Fe³⁺ in Aqueous Solution, *Anal. Chem.*, 2013, 85, 7441.
- S3 X. Zhu, Z. Zhang, Z. Xue, C. Huang, Y. Shan, C. Liu, X. Qin, W. Yang, X. Chen and T. Wang, Understanding the Selective Detection of Fe³⁺ Based on Graphene Quantum Dots as Fluorescent Probes: The Ksp of a Metal Hydroxide-Assisted Mechanism, *Anal. Chem.*, 2017, 89, 12054.
- S4 S. D. Padghan, A. L. Puyad, R. S. Bhosale, S. V. Bhosale and S. V. Bhosale, A pyrene based fluorescent turn-on chemosensor: aggregation-induced emission enhancement and application towards Fe³⁺ and Fe²⁺ recognition, *Photochem. Photobiol. Sci.*, 2017, 16, 1591.
- S5 M. Faraz, A. Abbasi, F. K. Naqvi, N. Khare, R. Prasad, I. Barman and R. Pandey, Polyindole/cadmium sulphide nanocomposite based turn-on, multi-ion fluorescence sensor for detection of Cr³⁺, Fe³⁺ and Sn²⁺ ions, *Sens. Actuators B*, **2018**, 269, 195.
- S6 Z. A. Consty, Y. Zhang and Y. Xu, A simple sensor based on imidazo[2,1-b]thiazole for recognition and differentiation of Al³⁺, F⁻and PPi, *J. Photochem. Photobio. A.*, 2020, 112578.
- S7 B. Kaur, A. Gupta and N. Kaur, A Simple Schiff base as a multi responsive and sequential sensor towards Al³⁺, F⁻ and Cu²⁺ ions, *J. Photochem. Photobio. A.*, 2020, 112140.
- **S8** R. Joseph, A. Asok and K. Joseph, Quinoline appended pillar[5]arene (QPA) as Fe³⁺

sensor and complex of Fe³⁺ (FeQPA) as a selective sensor for F⁻, arginine and lysine in the aqueous medium, *Spectrochim. Acta A Mol. Biomol. Spectrosc.*, **2020**, 224, 117390.

S9 G. J. Shree, S. Murugesan and A. Siva, A highly sensitive and selective Schiff-base probe as a colorimetric sensor for Co²⁺ and a fluorimetric sensor for F⁻ and its utility in bio-imaging, molecular logic gate and real sample analysis, *Spectrochim. Acta A Mol. Biomol. Spectrosc.*, **2020**, 226, 117163.