

## Supplementary Information

Extraction of phytochemicals from the pomegranate (*Punica granatum L.*, *Punicaceae*) by reverse iontophoresis

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**Supplementary Table S1:**  
Chromatographic separation schedules.

Time (min)	% B (C18)	%B (HILIC)
0.0	5	100
1.0	20	-
4.0	45	-
4.5	95	-
6.5	95	-
7.0	5	-
8.0	95	-
10.0	95	-
10.5	5	-
11.5	-	30
12.0	-	100
12.5	5	-
15.0	-	100

**Supplementary Table S2:** LC-MS operational conditions for the two analysis methods employed.

Parameter	C18 method	HILIC method
Injection volume ( $\mu\text{L}$ )	5	5
Mobile phase flow rate (mL/min)	0.4	0.5
Gas temperature ( $^{\circ}\text{C}$ )	250	300
Drying gas flow rate (L/min)	12	13
Nebulising gas pressure (bar)	3.1	2.1
Sheath gas temperature ( $^{\circ}\text{C}$ )	350	350
Sheath gas flow rate (L/min)	12	12
VCap voltage (V)	3500	1500
Fragmentor voltage (V)	125	100
Skimmer voltage (V)	45	40
MS/MS scan segment collision energies (eV)	0, 20, 40	0, 20, 40

**Supplementary Table S3:** Summary of mass spectrometric data for bioactive compounds generated from *ex vivo* and *in vivo* RI experiments with their relevant physicochemical parameters. Putative hits were qualified based on precursor ppm mass error (within 5 ppm) with at least two curated fragment ions within 10 ppm mass error.

RT (min)	HPLC column	Putative phytochemical name (CAS number)	pK <sub>a</sub> (s)	Charge at pH 7.4	Metabolite Class	Molecular Formula	ES(-/+) theor. m/z	ES(-/+) found m/z	Adduct	m/z error (ppm)	MS/MS ES(-) fragment ions
0.87*	C18	Citric acid <sup>†</sup> (320-77-4)	3.1 4.8 6.4 <sup>a</sup>	-3	Organic acid	C <sub>6</sub> H <sub>8</sub> O <sub>7</sub>	191.0192	191.0199	[M-H] <sup>-</sup>	3.7	57.0346 67.0189 85.0295 87.0088 111.0088 154.9986 173.0092
6.97	C18	Luteolin 7-β-rutinoside <sup>†§</sup> (20633-84-5)	8.3 <sup>b</sup>	0	Flavonoid glycoside	C <sub>27</sub> H <sub>30</sub> O <sub>15</sub>	593.1506	593.1513	[M-H] <sup>-</sup>	1.2	227.0383 255.0302 284.0302 285.04 286.0427 327.0510
4.98	C18	(-)Epicatechin <sup>†</sup> (490-46-0)	8.3 (1)	0	Flavonoid	C <sub>15</sub> H <sub>14</sub> O <sub>6</sub>	289.0712	289.0718	[M-H] <sup>-</sup>	2.1	109.0295 179.0350 203.0714 205.0506 245.0819
6.60	C18	Rutin <sup>†</sup> (115888-40-9)	6.4 <sup>b</sup>	-1	Flavonoid glycoside	C <sub>27</sub> H <sub>30</sub> O <sub>16</sub>	609.1456	609.1457	[M-H] <sup>-</sup>	0.2	178.9986 300.0334 301.0412
6.70*	C18	Ellagic acid <sup>†‡§  </sup> (476-66-4)	5.4 6.8 (2)	-2	Tannin	C <sub>14</sub> H <sub>6</sub> O <sub>8</sub>	300.9984	300.9987	[M-H] <sup>-</sup>	1.0	257.0086 284.9994
9.49*	HILIC	Histidine <sup>§  </sup> (71-00-1)	1.8 6.0 9.2 <sup>a</sup>	0	Amino acid	C <sub>6</sub> H <sub>9</sub> N <sub>3</sub> O <sub>2</sub>	156.0773	156.0772	[M+H] <sup>+</sup>	-0.6	81.0448 82.0527 93.045 83.0601 110.0714
10.42*	HILIC	Lysine <sup>§</sup>	2.2	+1	Amino acid	C <sub>6</sub> H <sub>14</sub> N <sub>2</sub> O <sub>2</sub>	147.1133	147.1129	[M+H] <sup>+</sup>	-2.7	56.0495

		(923-27-3)	9.0 10.5 <sup>a</sup>								84.0808
9.88*	HILIC	Arginine <sup>§</sup> (74-79-3)	2.2 9.0 12.5 <sup>a</sup>	+1	Amino acid	C <sub>6</sub> H <sub>14</sub> N <sub>4</sub> O <sub>2</sub>	175.1195	175.1193	[M+H] <sup>+</sup>	-1.1	60.0570 70.0651 116.0706 130.0975
8.88*	HILIC	Aspartic acid <sup>§  </sup> (56-84-8)	1.9 3.7 9.6 <sup>a</sup>	-1	Amino acid	C <sub>4</sub> H <sub>7</sub> NO <sub>4</sub>	134.0450	134.0453	[M+H] <sup>+</sup>	-2.2	46.0286 70.0288 74.0238 116.0341
9.36*	HILIC	Glutamylglutamic acid <sup>§  </sup> (3929-61-1)	2.2 4.3 9.7 <sup>ad</sup>	-2	Dipeptide	C <sub>10</sub> H <sub>16</sub> N <sub>2</sub> O <sub>7</sub>	277.1035	277.1041	[M+H] <sup>+</sup>	2.2	84.0450 130.0496 148.06
0.81	C-18	Malic acid <sup>§  </sup> (6915-15-7)	3.5 5.0 <sup>c</sup>	-2	Organic acid	C <sub>4</sub> H <sub>6</sub> O <sub>5</sub>	133.0137	133.0144	[M-H] <sup>-</sup>	5.3	71.0139 72.9930 115.0033
7.13	C-18	Abscisic acid <sup>§</sup> (7773-56-0)	4.7 <sup>b</sup>	-1	Phytohormone	C <sub>15</sub> H <sub>20</sub> O <sub>4</sub>	263.1283	263.1288	[M-H] <sup>-</sup>	1.9	153.0919 203.1077 219.1386

<sup>a</sup> <https://www.rsc.org/merck-index>

<sup>b</sup> <https://hmdb.ca/metabolites>

<sup>c</sup> <https://pubchem.ncbi.nlm.nih.gov>

<sup>d</sup> pK<sub>a</sub> values based on those of glutamic acid

\*Verified with analytical reference standard

† RI extraction (isolated peel) confirmed from unknown cultivar (Israel)

‡ RI extraction *in fructo* confirmed from Hicaz (Turkey)

§ RI extraction *in fructo* confirmed from Aco (South Africa)

|| RI extraction *in fructo* confirmed from Wonderful (Peru)

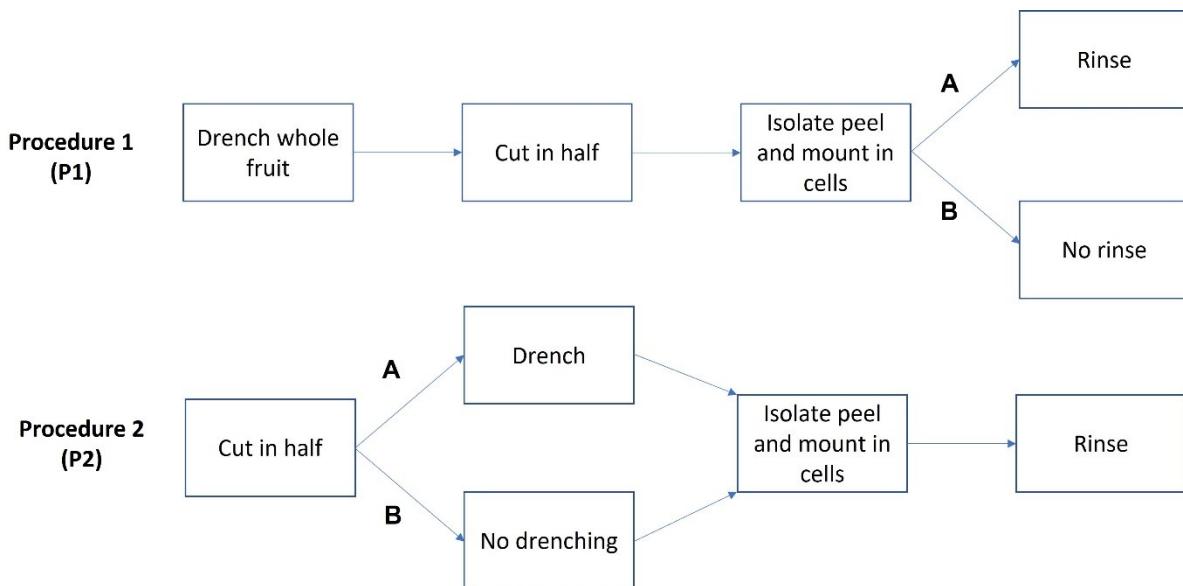
**Supplementary Table S4:** Intra-fruit variability in two Hicaz (Turkey) pomegranates

Peel section	Ellagic acid concentration in dry peel ( $\mu\text{g}/\text{mg}$ )	
	Experiment P1A & P1B	Experiment P2A & P2B
Top	2.62 ( $\pm 0.19$ )	1.53 ( $\pm 0.10$ ) <sup>b</sup>
Middle	1.96 ( $\pm 0.22$ ) <sup>a,b</sup>	1.57 ( $\pm 0.04$ ) <sup>b</sup>
Bottom	2.72 ( $\pm 0.20$ )	1.94 ( $\pm 0.06$ )

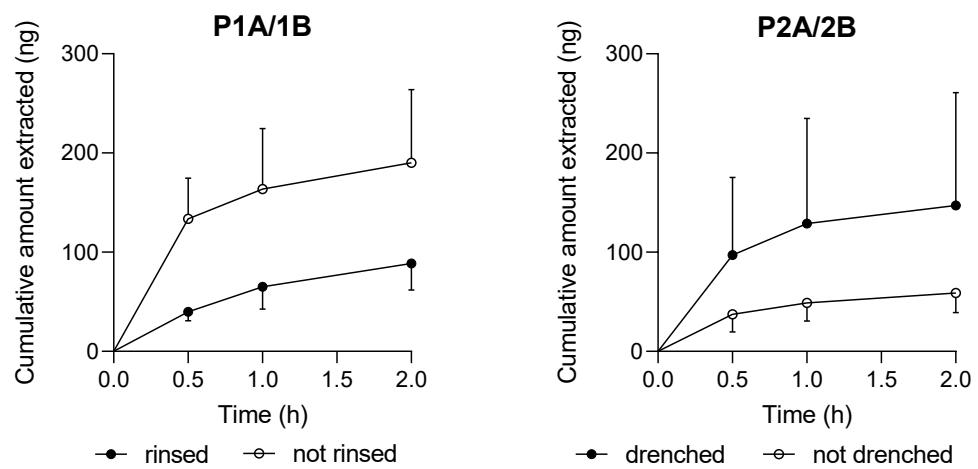
<sup>a</sup> Significantly different to Top<sup>b</sup> Significantly different to Bottom**Supplementary Table S5:** Measured parameters from the reverse iontophoresis extraction of ellagic acid (EA) on intact pomegranates

Cultivar	% loss on drying	pH of dry peel suspension	Ellagic acid in dry peel ( $\mu\text{g}/\text{mg}$ )	EA extracted in 1 <sup>st</sup> hr (ng)	EA extracted in 2 <sup>nd</sup> hr (ng)
Aco (South Africa)	69	3.7	3.78 ( $\pm 1.70$ )	2060 ( $\pm 1001$ )	1321( $\pm 1069$ )
Wonderful (Peru)	63	3.4	1.09 ( $\pm 0.04$ )	53 ( $\pm 33$ )	58 ( $\pm 32$ )

**Supplementary Figure S1:** Details of the four peel preparation procedures examined.

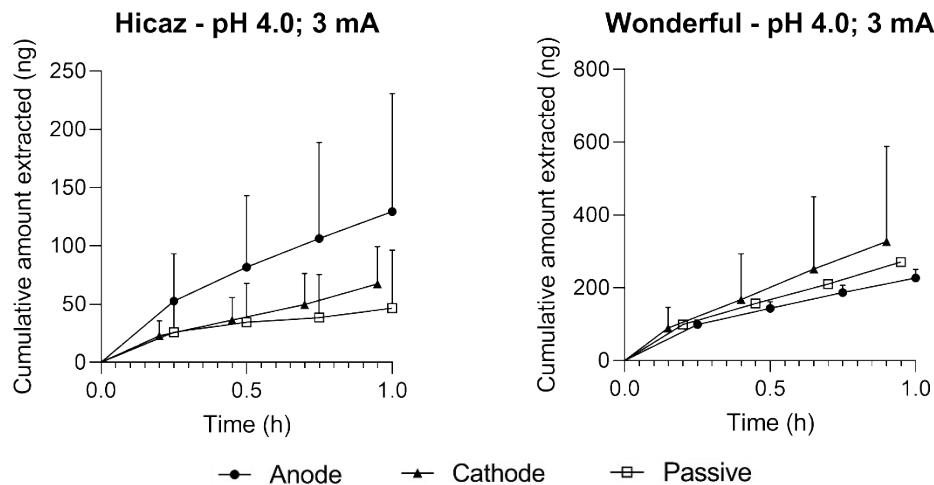


**Supplementary Figure S2:** Cumulative passive extraction of ellagic acid following [left panel] P1A and P1B, and [right panel] P2A and P2B (mean  $\pm$  SD;  $n = 5$  except for P2B for which  $n = 6$ ).

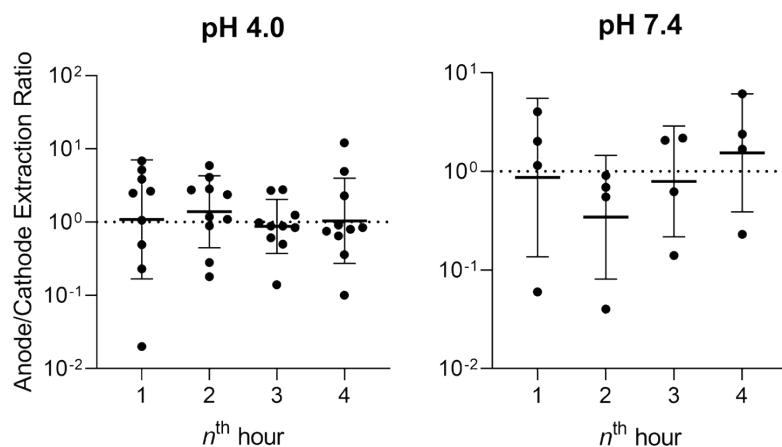




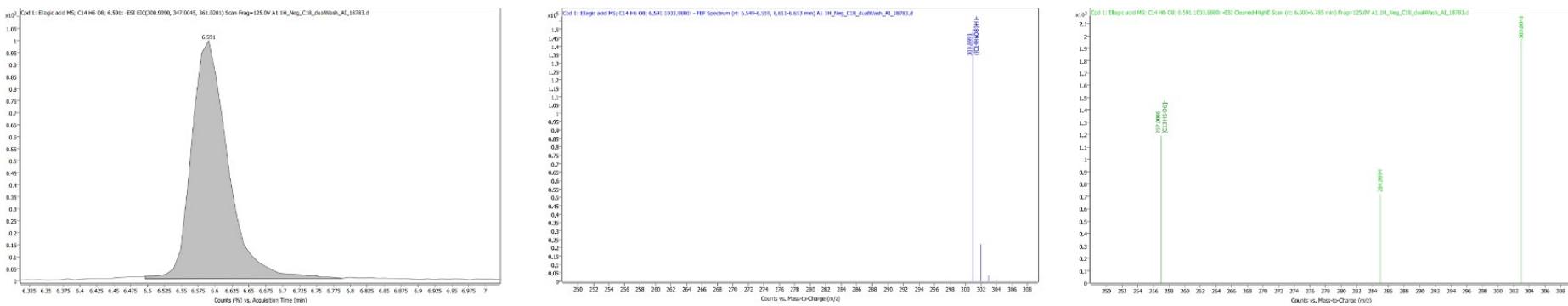
**Supplementary Figure S3:** Cumulative reverse iontophoretic extraction of ellagic acid at pH 4.0 from the isolated peels of Hicaz and Wonderful pomegranates at currents 3 mA current. Extraction to anode (closed circles,  $n = 4$  or 5), cathode (closed triangles,  $n = 5$ ) and passively (open squares,  $n = 1$  or 3) (mean  $\pm$  SD). Data points are slightly offset for clarity.



**Supplementary Figure S4:** Pooled anode-to-cathode extraction ratios of paracetamol as a function of the time of current (0.2 mA) application at pH 4.0 and pH 7.4 (geometric mean  $\pm$  SD;  $n \geq 9$  and  $n = 5$ , respectively).



**Supplementary Figure S5:** Representative ellagic acid analysis of a RI extracted anodal sample from an intact Aco pomegranate *in fructo* after 1 h of current passage. Left panel - extracted ion chromatogram (EIC). Middle panel - molecular ion [M-H]<sup>-</sup> MS/MS spectrum. Right panel - MS/MS spectrum.



**Supplementary Figure S6:** Representative histidiine analysis of a RI extracted cathodal sample from an intact Aco pomegranate *in fructo* after 1 h of current passage. Left panel - extracted ion chromatogram (EIC). Middle panel - molecular ion [M+H]<sup>+</sup> MS/MS spectrum. Right panel - MS/MS spectrum.

