Supplementary Materials for

Lasiodipline G and other diketopiperazine metabolites produced by

Lasiodiplodia chiangraiensis

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ABSTRACT

Lasiodiplodia fungi are known to colonize plants as both pathogens and/or endophytes; hence, they can be exploited for their beneficial roles. Many compound classes from the genus have exhibited their potential biotechnological applications. Herein, we report two new metabolites **1** and **2** together with three known cylco-(D-Ala-D-Trp) (**3**), indole-3-carboxylic acid (**4**) and a cyclic pentapeptide clavatustide B (**5**), isolated from the submerged cultures of a recently described species *L. chiangraiensis*. Chemical structures of the isolated compounds were determined by extensive NMR spectroscopic analyses together with HRESIMS. The absolute configurations of the new compounds were established based on comparing experimental and calculated time-dependent density functional theory circular dichroism (TDDFT-ECD) spectra. Compound **1** exhibited significant cytotoxic activities against an array of cell lines with IC₅₀ values of 2.9-12.6 μ M, as well as moderate antibacterial effects.

Keywords: Lasiodiplodia, new species, diketopiperazine, cytotoxic, antimicrobial.

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Pos.	¹ H- ¹ H COSY ^a	HMBC ^a	ROESY ^a
1			
2			
4			
5			
6-N <i>H</i>			
7	H-9w ^a	4, 5, 8, 9, 16	H-9, H-15
8			
9	H-7w ^a	7, 8, 11, 12w ^a , 15w ^a , 16	H-7
10-NH			
11			
12	H-13, H-14w ^a	14, 16	
13	H-12, H-14	11, 12w ^a , 15	
14	H-13, H-15	11w ^a , 12, 16	
15	H-14, H-13w ^a	8, 11, 13, 16w ^a	H-7
16			
2-Me		1, 2	3-NMe
3-NMe		2,4	2-Me

Table S1. 2D (COSY, HMBC and ROESY) NMR correlations of **1** in methanol- d_4 at 700 MHz.

^a "w" denotes weak correlation.

Table S2. 2D (COSY, HMBC and ROESY) NMR correlations of **2** in methanol- d_4 at 700 MHz.

Pos.	¹ H- ¹ H COSY	НМВС	ROESY	
1				
2	2-C <u>H</u> ₃	C-1, 2- <u>C</u> H ₃ , C-4	H-5	
3-NOH				
4				
5		C-1, C-4, C-7, C-8,	H-2	
6-N <i>H</i>				
7		C-1w ^a , C-4, C-5, C-8, C-9, C-16	H-15	
8				
9	$C\underline{H}_2$ -7 w^a	C-5w ^a , C-7, C-8, C-11, C-16		
10-NH				
11				
12	H-13, H-14w ^a	C-14, C-16		
13	H-12, H-14, H-15w ^a	C-11, C-12w ^a , C-14w ^a , C-15		
14	H-12w ^a , H-13, H-15	C-12, C-13w ^a , C-15w ^a , C-16		
15	H-13w ^a , H-14, H-15	C-8, C-11, C-13, C-16w ^a	C <u>H</u> ₂ -7	
16				
2-Me	H-2	C-1, C-2		

^a "w" denotes weak correlation.

	IC50 (µM)			Positive Control	
Test Cell Line	1	2	3	4	Epothilone B (µM)
Mouse fibroblast (L929)	5.8	n.a.	n.a.	87	0.65
Human endocervival adenocarcinoma (KB3.1)	8.4	n.a.	n.a.	n.a.	0.17
Human prostate carcinoma (PC-3)	6.3	n.d.	n.d.	n.d.	0.09
Human breast adenocarcinoma (MCF-7)	3.9	n.d.	n.d.	n.d.	0.07
Human ovarian cancer (SKOV-3)	3.9	n.d.	n.d.	n.d.	0.09
Human epidermoid carcinoma (A431)	2.9	n.d.	n.a.	n.a.	0.06
Human lung carcinoma (A549)	12.6	n.d.	n.d.	n.d.	0.05
Test Microorganism		MIC	(µg/mL)	Positive Control (µg/mL)	
Staphylococcus aureus	33.3	n.i.	n.i.	n.i.	0.21 ^G
Escherichia coli	n.d.	n.i.	n.i.	n.i.	0.83 ^G
Bacillus subtilis	n.i.	n.i.	n.i.	n.i.	16.6 ⁰
Pseudomonas aeruginosa	n.d.	n.i.	n.i.	n.i.	0.22 ^G
Pichia anomala	n.d.	n.i.	66.6	n.i.	4.20 ^N
Candida albicans	n.d.	n.i.	n.i.	n.i.	2.20 ^N
Acinetobacter baumanii	n.d.	n.i.	n.i.	n.i.	0.52 ^C
Chromobacterium violaceum	n.d.	n.i.	n.i.	n.i.	1.70 ^G
Schizosaccharomyces pombe	n.d.	n.i.	n.i.	n.i.	2.10 ^N
Mucor hiemails	n.d.	n.i.	n.i.	n.i.	2.10 ^N
Rhodotorula glutinis	n.d.	n.i.	n.i.	n.i.	2.30 ^N
Mycobacterium smegmatis	n.d.	n.i.	n.i.	n.i.	1.70 ^K

Table S3. Cytotoxicity (IC_{50}) and antimicrobial activity (MIC) of 1-4.

n.a.: No activity. n.i.: No inhibition up to 67 µg/mL. n.d.: Not determined.

G: Gentamycin; O: Oxytetracycline; N: Nystatin; C: Ciprofloxacin; K: Kanymycin.



Figure S1. HPLC chromatogram of Lasiodiplodia chiangraiensis cultured on cotton seed (Q6) medium.





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Figure S4. ¹H NMR spectrum of **1** in methanol- d_4 at 700 MHz.



Figure S5. ¹³C NMR spectrum of **1** in methanol- d_4 at 175 MHz.



Figure S6. ¹H-¹H COSY spectrum of **1** in methanol- d_4 at 700 MHz.



Figure S7. HMBC spectrum of $\mathbf{1}$ in methanol- d_4 at 700 MHz.























Figure S14. ¹H-¹H COSY spectrum of 2 in methanol- d_4 at 700 MHz.



Figure S15. HMBC spectrum of 2 in methanol- d_4 at 700 MHz.



Figure S16. HSQC spectrum of 2 in methanol- d_4 at 700 MHz.



Figure S17. ¹H NMR spectrum of **2** in DMSO- d_6 at 700 MHz.



Figure S18. ¹³C NMR spectrum of **2** in DMSO- d_6 at 175 MHz.



Figure S19. ¹H-¹H COSY spectrum of **2** in DMSO- d_6 at 700 MHz.



Figure S20. ROESY spectrum of 2 in DMSO- d_6 at 700 MHz.



Figure S21. LRESIMS of 3.





Figure S23. ¹H NMR spectrum of **3** in methanol- d_4 at 700 MHz.



Figure S24. ¹H-¹H COSY spectrum of **3** in methanol- d_4 at 700 MHz.



Figure S25. ROESY spectrum of **3** in methanol- d_4 at 700 MHz.



Figure S26. ¹H NMR spectrum of **3** in DMSO- d_6 at 700 MHz.



Figure S27. ¹³C NMR spectrum of **3** in DMSO- d_6 at 175 MHz.



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Figure S28. LRESIMS of 4.



Figure S29. HRESIMS of 4.

Figure S30. ¹H NMR spectrum of **4** in methanol- d_4 at 500 MHz.

Figure S31. HMBC spectrum of **4** in methanol- d_4 at 500 MHz.

Generic Display Report

Figure S32. LRESIMS of 5.

Figure S33. HRESIMS of 5.

Figure S34. ¹H NMR spectrum of **5** in methanol- d_4 at 700 MHz.

Figure S35. ¹H-¹H COSY spectrum of **5** in methanol- d_4 at 700 MHz.

Figure S36. HMBC spectrum of **5** in methanol- d_4 at 700 MHz.

Figure S37. HSQC spectrum of $\mathbf{5}$ in methanol- d_4 at 700 MHz.