Supplementary Information

The synthesis of pharmacologically important oxindoles via the asymmetric aldol reaction of isatin, and the investigation of the organocatalytic activity of new alicyclic β -amino acid derivatives

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Contents

Contents

1. I	Experimental procedures and analytical data of compounds 3–25	S2
2. I	FT-IR spectra of the synthesized compounds 3a–25	S12
3. (Copies of ¹ H- and ¹³ C NMR spectra of 3a–25	S25
4. (Copies of HRMS-ESI Spectra of 3a–25	S50
5. I	HPLC Chromatograms of 3a-h	S67

Experimental procedures and analytical data General methods

All reactions were conducted using magnetic stirring in oven-dried glassware. To purify the products using column chromatography, Merck KGaA Darmstadt, Germany's Kieselgel 60 (0.063–0.200 mm 70–230 mesh ASTM) was employed. Thin layer chromatography was carried out on silica-coated aluminium backing plates 60A F254. Spots were visualized using an UV lamp (254 nm). Sulphuric acid and vanillin were utilized for detection.

In a microwave (CEM, Discover, SP) chamber, sealed reaction vials (10 mL) were used to conduct microwave-promoted reactions (CEM Corporation, Matthwes, NC, USA).

All high-speed ball milling measurements were carried out with Retsch 400 Mixer Mill with 10 mL agate jars and 5 mm agate balls (Retsch Gmbh Germany).

Ethyl acetate (EtOAc), *n*-hexane, chloroform, acetone, ethanol and diethyl ether (Et₂O) were used as solvents of the highest analytical grade that were purchased from Molar Chemicals Kft (Halásztelek, Hungary).

HPLC-grade *i*-PrOH, *n*-hexane and tetrahydrofuran (THF) were purchased from VWR Chemicals (Fontenary-sous-Bois, France). *N*,*N*'-Diisopropylcarbodiimide (DIC) was bought from Alfa Aesar (Thermo Fischer, Kandel, Germany), Boc₂O was from Reanal Zrt. (Budapest, Hungary). 1-Hydroxybenzotriazole hydrate (HOBt), (+)-*O*,*O*'dibenzoiltartaric acid (DBTA), *O*,*O*'-di-*p*-toluoyltartaric acid (DPTTA), (*S*)-(+)-1-(2-naphthyl)ethylamine and (*S*)-(-)-1-(1-naphthyl)ethylamine were obtained from Sigma Aldrich (St. Louis, USA). 2-Methyl-2-butanol (98%), (*R*)-(+)-1-(2-naphthyl)ethylamine and (*R*)-(+)-1-(1-naphthyl)ethylamine were bought from TCI (Tokyo Chemical Industry Co., Zwijndrecht, Belgium). (*S*)-(-)- α -Methylbenzyl isocyanate was purchased from Acros Organics (Geel, Belgium), isatin and LiOH were bought from Fluka Chemie Gmbh (Sigma Aldrich Chemie Gmbh Steinheim Germany). Anhydrous sodium sulfate (Na₂SO₄) used as drying agent, was purchased from Sigma Aldrich (Merck KGaA Darmstadt, Germany).

The samples were prepared for HPLC analysis by removing the solvent under vacuum, adding 1 mL of the HPLC eluent (*n*-hexane/*i*-PrOH 80:20), and sonicating the mixture for 5 minutes in an ultrasonic bath (Elma Elmasonic S 30/H ultrasonic device, Elma Schmidbauen GmbH Gottlieb-Daimler Straße 17, 78224 Singen).

After homogenisation, the samples were filtered with PTFE 0.45 μ m filter and injected to the HPLC. HPLC measurements were carried out with JASCO HPLC (PU-4086 Binary Semi Preparative Pump, MD-4015 Photo Diode Array Detector, JASCO 2967-5 Ishikawa-machi Hachioji-shi Tokyo Japan 192-8537). The *ee* values for **3a–h** were determined by HPLC using Chiralpak IA column (Particle size: 5 μ m, dimensions: 4.6 mm × 15 mm) (eluent: a mixture of *n*-hexane and IPA (80:20), flow rate: 0.5 mL/min.

FT-IR spectra of the compounds were obtained by Fourier-transform infrared spectroscopy (Thermo Nicolet AVATAR 330, USA) equipped with the GRAMS/AI ver. 7 program. With 150 mg dry KBr, samples were crushed and made into pastilles. The pastilles were scanned 128 times in the wavenumber range 4000–400 cm⁻¹ at a resolution of 4 cm⁻¹.

¹H NMR spectra were recorded at 500.20 MHz, while the ¹³C NMR spectra were measured at 125.62 MHz in CDCl₃ or in DMSO-d₆ at ambient temperature, with a Bruker AV NEO Ascend 500 spectrometer (Bruker Biospin, Karlsruhe, Germany) with Double Resonance Broad Band Probe (BBO). Chemical shifts are given, relative to tetramethylsilane (Me₄Si) as internal standard, in δ (ppm).

The HRMS flow injection analysis was performed with Thermo Scientific Q Exactive Plus hybrid quadrupole-Orbitrap (Thermo Fisher Scientific, Waltham, MA, USA) mass spectrometer coupled to a Waters Acquity I-Class UPLC[™] (Waters, Manchester, UK).

Optical rotations were measured with JASCO P-2000 Polarimeter. Melting points were determined with a Hinotex-X4 micro melting point apparatus (Hinotek, Ningbo, China) and are uncorrected.

General procedure for the aldol reaction in solvent using organocatalyst 6-21, synthesis of 3a-h

Organocatalysts **6–21** (corresponding catalyst loading 20 mol %) were directly added to a stirred solution of isatin **1a–h** (0.28 mmol, 1 equiv), acetone (5 mL, 325 equiv) and additive (LiOH 1 mmol) at 25 °C and stirred for the corresponding time. The resulting mixture was stirred at that temperature for specified time. After completion of the reaction (monitored by TLC) the crude mixture was purified by column chromatography on silica gel (using EtOAc). Further purification was carried out if required by crystallizing the product in Et₂O to obtain the aldol adducts **3a–h**.

In case of solvent screening: The organocatalysts **6–21** (corresponding catalyst loading 20 mol %) were directly added to a stirred solution of isatin **1a–h** (0.28 mmol, 1 equiv) acetone (2.5 mL, 162.5 equiv) and different tested solvents (2.5 mL) and additive (LiOH 1 mmol) at 25 °C and stirred for the corresponding time. The resulting mixture was stirred at that temperature for specified time. After completion of the reaction (monitored by TLC) the crude mixture was purified by column chromatography on silica gel (using EtOAc). Further purification was carried out if required by crystallizing the product in Et₂O to obtain the aldol adducts **3a–h**.

(S)-3-Hydroxy-3-(2-oxopropyl)indolin-2-one (3a)



Yield (40 mg, 70%), white solid, mp 178–183 °C, mp (lit., 1 166–168°C, $[\alpha]_D^{25}$ -23.3 (c 0.5 in MeOH, 57 % *ee* % **Table 4** entry 1), (lit. 1 $[\alpha]_D^{25}$ -18.4 (c 0.82 in MeOH, 67% *ee*%)). The *ee* % was determined by HPLC method using Chiralpak IA column *n*-hexane/*i*-PrOH 80:20, flow rate 0.5mL/min; tR = 14.0 min (*S*)-enantiomer); tR = 17.0 min (*R*)-enantiomer). ¹H-NMR δ H (500.20 MHz, DMSO-d₆, 30 °C, Me₄Si) 2.00 (3H, s, CH₃), 2.99 (1 H, d, *J* = 16.6 Hz, *CH*¹CO), 3.25 (1 H, d, *J* = 16.6 Hz, *CH*²CO), 5.95 (1 H, s, OH), 6.77 (1 H, d, *J* = 7.7 Hz, Ar), 6.90 (1H, t, *J* = 7.4 Hz, Ar), 7.16 (1 H, t, *J* = 7.5 Hz, Ar), 7.23 (1H, d, *J* = 7.3 Hz, Ar) 10.19 (1 H, brs, NH), ¹³C-NMR δ C (125.62 MHz, DMSO-d₆, 30 °C, Me₄Si) 31.04,50.74, 73.13, 109.87, 121.67, 124.15, 129.42, 131.99, 142.98, 178.59, 205.58. HRMS (ESI) [M+H]+ m/z calcd for C₁₁H₁₁NO₃: 206.07389, found: 206.08101

(S)-3-Hydroxy-5-iodo-3-(2-oxopropyl)indolin-2-one (3b)



Yield (40 mg, 40%), white solid, mp 222–225 °C, $[α]_D^{25}$ -5.8 (c 0.5 in MeOH, 65 % *ee* % **Table 7** entry 8) The *ee* % was determined by HPLC method using Chiralpak IA column *n*-hexane/*i*-PrOH 80:20, flow rate 0.5mL/min; tR = 13.5 min (*S*)-enantiomer); tR = 19.5 min (*R*)-enantiomer). ¹H-NMR δH (500.20 MHz, DMSO-d₆, 30 °C, Me₄Si) 2.00 (3H, s, CH₃), 3.04 (1 H, d, *J* = 17.3 Hz, *CH*¹CO), 3.34 (1 H, d, *J* = 17.3 Hz, *CH*²CO), 6.06 (1 H, s, OH), 6.63 (1 H, t, *J* = 8.1 Hz, Ar), 7.50 (1H, d, *J* = 8.1 Hz Ar), 7.55 (1H, d, *J* = 1.7 Hz, Ph), 10.17 (1 H, s, NH). ¹³C-NMR δC (125.62 MHz, DMSO-d₆, 30 °C, Me₄Si) 30.8, 50.5, 72.9, 84.4, 112.4, 132.5, 134.9, 137.8, 143.0, 178.0, 205,8. HRMS (ESI) [M+Na]+ m/z calcd for C₁₁H₁₀INO₃Na: 353.96031, found: 353.95972

(S)-7-Chloro-3-hydroxy-3-(2-oxopropyl)indolin-2-one (3c)



Yield (31 mg, 42 %), white solid, mp 188–189 °C, mp (lit.2 175–177 °C), $[\alpha]_D^{25}$ -33.4 (c 0.4 in MeOH, 98% *ee*% **Table 7** entry 12), (lit 2. $[\alpha]_D^{20}$ -14.5 (c 0.4 in MeOH, 66% *ee*%)). The *ee* was determined by HPLC method using Chiralpak IA column *n*-hexane/*i*-PrOH 80:20, flow rate 0.5mL/min; tR = 12.6 min (*S*)-enantiomer); tR = 17.2 min (*R*)-enantiomer). ¹H-NMR δ H (500.20 MHz, DMSO-d₆, 30 °C, Me₄Si) 2.00 (3H, s, CH₃), 3.08 (1 H, d, *J* = 17.2 Hz, *CH*¹CO), 3.34 (1 H, d, *J* = 17.3 Hz, *CH*²CO), 6.11 (1 H, s, OH), 6.93 (1 H, t, *J* = 7.6 Hz, Ar), 7.21 (1H, d, *J* = 7.5 Hz Ar), 7.23 (1H, d, *J* = 7.9 Hz, Ph), 10.62 (1 H, s, NH).¹³C-NMR δ C (125.62 MHz, DMSO-d₆, 30 °C, Me₄Si) 30.8, 50.6, 73.6, 114.1, 122.7, 123.1, 129.8, 134,1, 140,8, 178.6, 205.7. HRMS (ESI) [M+Na]+ m/z calcd for C₁₁H₁₀CINO₃Na: 262.02469, found: 262.02413

(S)-5-fluoro-3-hydroxy-3-(2-oxopropyl)indolin-2-one (3d)



Yield: (50 mg, 50%), white solid, mp 158–161 °C, mp (lit. 6 183–185 °C); $[\alpha]_{D}^{25}$ -2.9 (c 0.1 in MeOH, 37% *ee*% **Table 7** entry 15), (lit. 6 $[\alpha]_{D}^{25}$ -17.0 (c 1.0 in MeOH, 77% *ee*%)). The *ee* was determined by HPLC method using Chiralpak IA column *n*-hexane/*i*-PrOH 80:20, flow rate 0.5mL/min; tR = 14.1 min (*S*)-enantiomer); tR = 19.4 min (*R*)-enantiomer). ¹H-NMR δ H (500.20 MHz, DMSO-d₆, 30 °C, Me₄Si) 2.01 (3H, s, CH₃), 3.04 (1 H, d, *J* = 17.2 Hz, *CH*¹CO), 3.34 (1 H, s, *CH*²CO), 6.07–6.78 (1 H, m, Ar), 6.96–7.02 (1 H, m, Ar), 7.12–7.17 (1 H, m, Ar), 10.22 (1H, s, NH). ¹³C-NMR δ C (125.62 MHz, DMSO-d₆, 30 °C, Me₄Si) 30.4, 49.0, 74.5, 113.2, 123.3, 128.5, 128.8, 129.6, 129.8, 130.9, 143.0, 177.8, 205.8. HRMS (ESI) [M+H]+ m/z calcd for C₁₁H₁₁O₃NF: 224.07137, found: 224.07175

(S)-3-hydroxy-5-nitro-3-(2-oxopropyl)indolin-2-one (3e)



Yield: (30 mg, 30%), white solid, mp 226–230 °C, mp (lit. 9 222–224 °C); $[\alpha]_{D}^{25}$ -4.0 (c 0.2 in MeOH, 41% *ee*% **Table 7** entry 15), (lit. 9 $[\alpha]_{D}^{20}$ -17.3 (c 1.82 in MeOH, 45% *ee*%)). The *ee* was determined by HPLC method using Chiralpak IA column *n*-hexane/*i*-PrOH 80:20, flow rate 0.5mL/min; tR = 15.0 min (*S*)-enantiomer); tR = 20.0 min (*R*)-enantiomer). ¹H-NMR δ H (500.20 MHz, DMSO-d₆, 30 °C, Me₄Si) 2.02 (3H, s, CH₃), 3.17 (1 H, d, *J* = 17.85 Hz, *CH*¹CO), 3.60 (1 H, d, *J* = 17.84 Hz, *CH*²CO), 6.27 (1 H, s, OH), 6.96–7.00 (1 H, m, Ar), 8.15–8.18 (2 H, m, Ar), 10.94 (1H, s, NH).¹³C-NMR δ C (125.62 MHz, DMSO-d₆, 30 °C, Me₄Si) 0.58, 30.5, 50.3, 72.6, 110.0, 120.0, 126.9, 133.2, 149.9, 179.0, 206.0. HRMS (ESI) [M+H]+ m/z calcd for C₁₁H₁₁O₅N: 251.06579, found: 251.06625

(S)-3-hydroxy-5-methyl-3-(2-oxopropyl)indolin-2-one (3f)



Yield: (24 mg, 27%), white solid, mp 170–173 °C, mp (lit.6 158–160 °C); $[\alpha]_D^{25}$ -4.5 (c 0.1 in MeOH, 39% *ee*% **Table 7** entry 18), (lit. 6 $[\alpha]_D^{25}$ -13.0 (c 1.0 in MeOH, 66% *ee*%)). The *ee* was determined by HPLC method using Chiralpak IA column *n*-hexane/*i*-PrOH 80:20, flow rate 0.5mL/min; tR = 12.9 min (*S*)-enantiomer); tR = 16.9 min (*R*)-enantiomer). ¹H-NMR δ H (500.20 MHz, DMSO-d₆, 30 °C, Me4Si) 2.00 (3H, s, CH₃), 2.22 (3 H, s, CH₃Ar), 2.97 (1 H, d, *J* = 16.51 Hz, *CH*¹CO), 3.22 (1 H, d, *J* = 16.47 Hz, *C1*²CO), 5.90 (1 H, s, OH), 6.65 (1 H, d, *J* = 7.78 Hz Ar), 6.96 (1 H, d, *J* = 7.8 Hz, Ar), 7.1 (1 H, s, Ar), 10.09 (1H, s, NH).¹³C-NMR δ C (125.62 MHz, DMSO-d₆, 30 °C, Me4Si) 21.1, 29.5, 31.0, 50.8, 73.2, 109.6, 124.8, 130.4, 132.1, 140.5, 178.6, 205.6. HRMS (ESI) [M+Na]+ m/z calcd for C₁₂H₁₃O₅NNa: 242.07825, found: 242.07876

(S)-4,7-dichloro-3-hydroxy-3-(2-oxopropyl)indolin-2-one (3g)



Yield: (99 mg, 96%), white solid, mp 204–208 °C, mp (lit. 10 235–237 °C); $[\alpha]_D^{25}$ -21.1 (c 0.4 in MeOH, *97*% **Table 7** entry 21). The *ee* was determined by HPLC method using Chiralpak IA column *n*-hexane/*i*-PrOH 80:20, flow rate 0.5mL/min; tR = 13.0 min (*S*)-enantiomer); tR = 26.6 min (*R*)-enantiomer). ¹H-NMR δ H (500.20 MHz, DMSO-d₆, 30 °C, Me₄Si) 2.03 (3H, s, CH₃), 3.21 (1 H, d, *J* = 17.72 Hz, *CH*¹CO), 3.69 (1 H, d, *J* = 17.72 Hz, *CH*²CO), 6.30 (1 H, s, OH), 6.92 (1 H, d, *J* = 8.71 Hz Ar), 7.29 (1 H, d, *J* = 8.71 Hz, Ar), 10.16 (1H, s, NH).¹³C-NMR δ C (125.62 MHz, DMSO-d₆, 30 °C, Me₄Si) 30.4, 49.0,74.5, 113.3, 123.7, 128.5, 128.8, 129.6, 129.8, 130.9, 143.0, 177.8, 205.8. HRMS (ESI) [M+Na]+ m/z calcd for C₁₁H₉O₃NCl₂Na: 295.98459, found: 295.98517

(S)-5-bromo-3-hydroxy-3-(2-oxopropyl)indolin-2-one (3h)



Yield: (30 mg, 34%), white solid, mp 211–214 °C, mp (lit.1 145–147 °C); $[\alpha]_{D}^{25}$ -6.2(c 0.1 in MeOH, 49% **Table 7** entry 24), (lit.1 $[\alpha]_{D}^{25}$ -17.6 (c 0.7 in MeOH, 75% *ee*%)). The *ee* was determined by HPLC method using Chiralpak IA column *n*-hexane/*i*-PrOH 80:20, flow rate 0.5mL/min; tR = 13.9 min (*S*)-enantiomer); tR = 20.0 min (*R*)-enantiomer). ¹H-NMR δ H (500.20 MHz, DMSO-d₆, 30 °C, Me₄Si) 2.01 (3H, s, CH₃), 3.06 (1 H, d, *J* = 17.25 Hz, *CH*²CO), 3.38 (1 H, d, *J* = 17.25 Hz, *CH*²CO), 6.07 (1 H, s, OH), 6.74 (1 H, d, *J* = 8.21 Hz Ar), 7.33–7.36 (1 H, m, Ar), 7.42 (1 H, d, *J* = 2.00 Hz), 10.33 (1H, s, NH).¹³C-NMR δ C (125.62 MHz, DMSO-d₆, 30 °C, Me₄Si) 30.8, 50.4, 73.1, 111.8, 113.4, 127.1, 132.0, 134.6, 142.5, 178.2, 205.7. HRMS (ESI) [M+Na]+ m/z calcd for C₁₁H₁₀O₃NBrNa: 305.97325, found: 305.97363

General procedure for the aldol reaction in High Speed Ball Milling (HSBM)

All High Speed Ball Milling measurements were carried out with Retsch 400 Mixer Mill with 10 mL agate jars and 5 mm agate balls (Retsch Gmbh Germany). In case of examining the effect of ball size under already optimized catalyst loading conditions: the measurements were done with 25 mL stainless steel jars and 10 mm balls. By all studies regarding the sampling, 10 minutes long pauses were taken. The applied frequencies and the reaction times were changed by every attempts. Then the gained samples from jars were solved in EtOAc, were sonicated completely, and purificated by column chromatography and after that were samples measured by HPLC.

General procedure for synthesis of urea type of organocatalysts (-)-6 and (-)-8

The enantiomeric *diendo* ethyl 3-aminobicyclo[2.2.1]hept-5-ene-2-carboxylate (–)-4 and (+)-4 were prepared from racemic *diendo* norbornene β -amino ester (±)-4 by diastereomeric salt formation with (+)-DBTA (0.5 equiv) or (–)-DBTA (0.5 equiv) according to the literature.³ Amino ester base (–)-4 or (+)-4 (100 mg, 0.55 mmol) was dissolved in anhydrous Et₂O (20 mL) and a 10 % excess of (S)-(–)- α -methylbenzyl isocyanate (100 mg, 0.55 mmol) was added The reaction mixture was allowed to stand for 24 h at room temperature. After evaporation, the resulting white crystalline urea adduct was recrystallized from *i*Pr₂O.

(1R,2S,3R,4S)-Ethyl 3-(3-((S)-1-phenylethyl)ureido)bicyclo[2.2.1]hept-5-ene-2-carboxylate (-)-6

Yield (139 mg, 77 %), white solid, mp 108–111 °C, $[\alpha]_D^{25}$ –12.3 (c 0.4 in EtOH). ¹H-NMR δ H (500.20 MHz, CDCl₃, 30 °C, Me₄Si) 1.19 (3 H, t, *J* = 7.1 Hz, CH₂*CH*₃), 1.29 (1 H, d, *J* = 8.9 Hz, 7-H), 1.36–1.42 (4 H, m, CH*CH*₃, 7-H), 2.94 (1 H, s, 2-H), 3.03 (1 H, s, 4-H), 3.11 (1 H, dd, *J*₁ = 9.2 Hz, *J* = 3.3 Hz, 1-H), 3.94–4.08 (2 H, m, O*CH*₂CH₃), 4.53–4.63 (2 H, m, 3-H, *NH*CO), 4.65-4.71 (1 H, m, NH*C*HCH₃), 4.97 (1 H, d, *J* = 9.2 Hz, *NH*), 5.63 (1 H, s, 5-H), 6.16–6.21 (1 H, m, 6-H), 7.21–7.35 (5 H, m, Ar), ¹³C-NMR δ C (125.62 MHz, CDCl₃, 30 °C, Me₄Si) 14.2, 23.7, 46.9, 47.5, 48.0, 50.6, 53.1, 60.3, 63.4 65.2, 125.9, 127.3, 128.6, 133.5, 137.3, 144.1, 156.7. HRMS (ESI) [M+H]+ m/z calcd for C₁₉H₂₄N₂O₃: 329.17869, found: 329.18597

(15,2R,35,4R)-Ethyl 3-(3-((S)-1-phenylethyl)ureido)bicyclo[2.2.1]hept-5-ene-2-carboxylate (-)-8



Yield (120 mg, 66 %), white solid crystals, (lit. 3 mp^{lit} 187–189 °C), mp 179–182 °C, (lit 3 $[\alpha]_{D}^{20}$ –14.5 (c 0.5 in EtOH), $[\alpha]_{D}^{25}$ –19.6 (c 0.4 in EtOH).¹H-NMR δ H (500.20 MHz, CDCl₃, 30 °C, Me₄Si) 1.21 (3 H, t, *J* = 7.2 Hz, OCH₃CH₂), 1.35 (1 H, d, *J* = 8.9 Hz, 7-H), 1.40–1.47 (4 H, m, *J* = 6.8 Hz, CHCH₃, 7-H), 3.07 (2 H, m, 2-H, 4-H), 3.11–3.18 (1 H, m, 1-H), 3.91–4.06 (2 H, m, CH₃CH₂), 4.41 (1 H, d, *J* = 7.14 Hz, NH), 4.65-4.71 (1 H, m, NHCHCH₃), 4.73–4.81 (1 H, m, 3-H), 5.05 (1 H, d, *J* = 8.8 Hz, NH), 6.10–6.16 (1 H, m, 6-H), 6.26–6.32 (1 H, m, 5-H), 7.21–7.24 (1 H, m, Ar), 7.28–7.35 (4 H, m, Ar). ¹³C-NMR δ C (125.62 MHz, CDCl₃, 30 °C, Me₄Si) 14.2, 23.2, 47.0, 47.6, 47.60, 47.8, 50.1, 53.4, 60.3, 118.3, 126.0, 127.2, 128.6, 133.8, 137.5, 144.1, 156.8, 173.4. HRMS (ESI) [M+H]+ m/z calcd for C₁₉H₂₄N₂O₃: 329.17869, found: 329.18540

General procedure for synthesis of amide type of organocatalysts (-)-7 and 9-16

The synthesis of enantiomeric Boc-protected amino acid (+)-**5** and (-)-**5** were performed according to the literature.⁴ They were then dissolved (140 mg, 0.55 mmol, 1 equiv) in THF using the corresponding amine (0.55 mmol, 1 equiv) in the presence of DIC (0.72 mmol, 1.3 equiv) and HOBt (0.66 mmol, 1.2 equiv) stirring for a day at room temperature. Different chiral amine, were used for the synthesis of organocatalysts: (*S*)-(-)-1-(2-naphthyl)-ethylamine (for (-)-**9**), (*S*)-(-)-1-(1-naphthyl)-ethylamine (for (-)-**10**), (*R*)-(+)-1-(2-naphthyl)-ethylamine (for (+)-**11** or (+)-**12**), (*R*)-(+)-1-(1-naphthyl)-ethylamine (for (+)-**13** or (+)-**14**) and propargylamine (for (-)-**15**). After completion of the reaction (checked by TLC), the solvent was evaporated. In order to remove the HOBt and DIC, the residue was dissolved in EtOAc/H₂O mixture (15 mL), the aqueous layer extracted with EtOAc (3 × 5 mL). The combined extracts were dried, filtered and evaporated. The product was purified by column chromatography using EtOAc or *n*-hexane/EtOAc 1:1 eluent. The appropriate Boc-protected amides (+)-**11** and (+)-**13** (0.20 mmol) was deprotected by a 20 % ethanolic HCl solution (10 mL) at r.t. for 6 h. After evaporation HCl salts of (+)-**12** and (+)-**14** were used without purification.

tert-Butyl ((15,2R,35,4R)-3-(((S)-1-phenylethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (-)-7



Yield (104 mg, 53 %), white solid, mp 140–145 °C, $[α]_{D}^{25}$ –7.6 (c 0.5 in EtOH). ¹H-NMR δH (500.20 MHz, CDCl₃, 30 °C, Me₄Si) 1.17 (1H, d J= 6.4Hz, 7-H), 1.35 (9 H, brs, Boc), 1.44–1.52 (3 H, t, *J* = 6.9 Hz, CH₃), 1.63 (1H, s, 7-H), 2.99 (1H, dd, *J* = 2.3 Hz, *J* = 9.6 Hz, 2-H), 3.07 (2 H, s, 4-H, 1-H), 4.52-4.58 (1 H, m, NHCHCH₃), 5.03-5.08 (1 H, m, 3-H), 5.13 (1 H, d, *J* = 9.1 Hz, *NH*), 5.89 (1 H, d, *J* = 5.4 Hz, *NH*), 6.11–6.21 (1 H, m, 5-H), 6.51 (1 H, s, 6-H), 7.22-7.36 (5 H, m, Ar). ¹³C-NMR δC (125.62 MHz, CDCl₃, 30 °C, Me₄Si) 21.9, 28.3, 47.1, 47.5, 47.6, 49.1, 50.5, 54.1, 79.2, 126.1, 127.2, 128.6, 132.1, 138.8, 143.0, 156.0, 171.0. HRMS (ESI) [M+H]+ m/z calcd for C₂₁H₂₈N₂O₃: 357.21000, found: 357.21727.

tert-Butyl ((1*R*,2*S*,3*R*,4*S*)-3-(((*S*)-1-(naphthalen-2-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (–)-9



Yield (156 mg, 70 %), light yellow oil, $[\alpha]_D^{25}$ -67.1 (c 0.2 in EtOH). ¹H-NMR δ H (500.20 MHz, CDCl₃, 30 °C, Me₄Si) 1.31 (2 H, d, *J* = 8.6 Hz, 7-H), 1.43 (9 H, s, Boc), 1.51 (3 H, d, *J* = 6.9 Hz, CH₃), 2.96-3.05 (3H, m, 2-H, 4-H, 1-H), 4.59 (1 H, m, NH*CH*CH₃), 4.97 (1 H, d, *J* = 9.7 Hz, NH), 5.13–5.21 (1 H, m, 3-H), 5.95 (1 H, d, *J* = 7.0 Hz, *NH*), 6.08–6.13 (1 H, m, 5-H), 6.51-6.57 (1 H, m, 6-H), 6.98 (1 H, s, Ar), 7.38–7.50 (3 H, m, Ar), 7.73 (1 H, s, Ar), 7.79–7.84 (3 H, m, Ar). ¹³C-NMR δ C (125.62 MHz, CDCl₃, 30 °C, Me₄Si) 21.2, 22.0, 28.5, 30.3, 34.2, 46.6, 47.5, 47.6, 49.1, 51.4, 54.2, 79.4, 124.5, 124.6, 125.5, 125.8, 126.2, 127.6, 127.9, 128.5, 131.2, 132.8, 133.4, 139.9, 170.9. HRMS (ESI) [M+H]+ m/z calcd for C₂₅H₃₀N₂O₃: 407.22564, found: 407.23291

tert-Butyl ((1*R*,2*S*,3*R*,4*S*)-3-(((*S*)-1-(naphthalen-1-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (–)-10



Yield (159 mg, 71 %), light oil, $[\alpha]_D^{25}$ -25.3 (c 0.4 in EtOH).¹H-NMR δ H (500.20 MHz, CDCl₃, 30 °C, Me₄Si) 1.25-1.27 (2 H, m, 7-H), 1.43 (9 H, s, Boc), 1.57–1.63 (4 H, m, CH₃, 7-H), 2.91 (1H, d, 2-H), 2.96 (1H, s, 4-H), 3.03 (1 H, s, 1-H), 3.43–3.52 (1 H, m, 3-H), 4.51–4.60 (1 H, m, CH₃*CH*NH), 5.11 (1 H, d, *J* = 9.5 Hz, *NH*), 5.76-5.80 (1 H, brs, *NH*), 6.11 (1 H, m, 5-H), 6.53-6.60 (1 H, m, 6-H), 7.41–7.48 (5 H, m, Ar), 7.78 (1 H, d, *J* = 7.9 Hz, Ar), 7.86 (1 H, d, *J* = 8.1 Hz, Ar), 8.06 (1 H, d, *J* = 8.4 Hz, Ar).¹³C-NMR δ C (125.62 MHz, CDCl₃, 30 °C, Me₄Si) 20.8, 28.5, 30.3, 44.6, 46.8, 47.6, 51.0, 54.1, 79.3, 122.5, 123.3, 125.3, 125.5, 125.8, 126.5, 128.3, 128.8, 131.0, 131.5, 134.0, 138.4, 139.6, 156.0, 170.6. HRMS (ESI) [M+Na]+ m/z calcd for C₂₅H₃₀N₂O₃Na: 429.21541, found: 429.21539

tert-Butyl ((1*R*,2*S*,3*R*,4*S*)-3-(((*R*)-1-(naphthalen-2-yl) ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-11



Yield (179 mg, 80 %), light yellow solid crystals, mp 137–141 °C, $[\alpha]_{D}^{25}$ +93.4 (c 0.3 in EtOH).¹H-NMR δ H (500.20 MHz, CDCl₃, 30 °C, Me₄Si) 1.22 (9 H, m, Boc), 1.30-1.37 (2 H, s, 7-H), 1.56 (3 H, d, *J* = 6.87 Hz, CH₃), 2.98 (1 H, dd, *J* = 2.7 Hz, *J* = 9.4 Hz, 2-H), 3.06 (2 H, d, *J* = 12.9 Hz, 4-H, 1-H), 4.45-4.55 (1 H, m, 3-H), 5.11 (1 H, d, *J* = 7.9 Hz, *NH*), 5.14–5.21 (1 H, m, CH₃*CH*NH), 5.92 (1 H, brs, *NH*), 6.09–6.16 (1 H, m, 5-H), 6.46–6.51 (1 H, m, 6-H), 7.33–7.50 (3 H, m, Ar), 7.70 (1H, s, Ar), 7.74–7.86 (3 H, m, Ar).¹³C-NMR δ C (125.62 MHz, CDCl₃, 30 °C, Me₄Si) 21.8, 28.2, 47.1, 47.6, 49.2, 50.5, 54.2, 79.3, 111.0, 117.5, 124.6, 124.7, 125.8, 126.1, 127.5, 128.0, 128.5, 132.1, 132.7, 133.3, 138.9, 140.3, 156.1, 171.2. HRMS (ESI) [M+H]+ m/z calcd for C₂₅H₃₀N₂O₃: 407.22564, found: 407.23292

(1*S*,2*R*,3*S*,4*R*)-3-Amino-N-((*R*)-1-(naphthalen-2-yl)ethyl)bicyclo[2.2.1]hept-5-ene-2-carboxamide hydrochloride (+)-12



Yield (118 mg, 70 %, light yellow oil, $[α]_{0}^{25}$ +18.9 (c 0.1 in EtOH).¹H-NMR δH (500.20 MHz, DMSO-d6, 30 °C, Me4Si) 1.09 (1 H, t, *J*= 7.0 Hz, 7-H), 1.21 (3 H, t, *J*= 7.1 Hz,CH₃), 1.41 (2 H, d, *J*= 8.9 Hz, 2-H, 4-H), 1.44 (1 H, s, 7-H), 2.36 (1 H, s, 1-H), 2.64 (1 H, s, 3-H), 5.00-5.05 (1 H, m, CH₃*CH*NH), 6.15-6.20 (1 H, m,NH₂), 6.29-6.32 (1 H, m, 5-H), 6.33-6.36 (1 H, m, 6-H), 7.46–7.53 (3 H, m, Ar), 7.81 (1 H, s, Ar), 7.86-7.90 (3 H, m, Ar), 8.58 (1 H, d, *J*= 8.1 Hz, NH) ¹³C-NMR δC (125.62 MHz, DMSO-d6, 30 °C, Me4Si) 22.6, 43.9, 47.3, 47.4, 48.5, 48.6, 52.9, 66.8, 124.7, 125.6, 126.1, 126.6, 127.9, 128.1, 128.3, 132.7, 133.3, 138.6, 142.0, 171.6. HRMS (ESI) [M+H]+ m/z calcd for C₂₀H₂₃ClN₂O: 307.17321, found: 307.18049

tert-Butyl ((1*R*,2*S*,3*R*,4*S*)-3-(((*R*)-1-(naphthalen-1-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-13



Yield (201 mg, 90 %), light yellow solid crystals, mp 139–143 °C, $[\alpha]_{D}^{25}$ +34.9 (c 0.3 in EtOH). ¹H-NMR δ H (500.20 MHz, CDCl₃, 30 °C, Me₄Si) 1.27 (9 H, brs, Boc), 1.43 (2 H, s, 7-H),1.63 (3 H, d, *J* = 6.7 Hz, CH₃), 2.94 (1 H, d, *J* = 8.9 Hz, 4-H), 3.07 (2 H, s, 2-H, 1-H), 4.38–4.50 (1 H, m, 3-H), 5.29 (1 H, d, *J* = 8.8 Hz, NH), 5.78–5.87 (2 H, m, CH₃*CH*NH, NH), 6.10–6.19 (1 H, m, H-5), 6.46 (1 H, s, 6-H), 7.40–7.54 (5 H, m, Ar), 7.76 (1 H, d, *J* = 7.9 Hz, Ar), 7.84 (1 H, d, *J* = 7.9 Hz, Ar), 8.04 (1 H, d, *J* = 8.4 Hz, Ar).¹³C-NMR δ C (125.62 MHz, CDCl₃, 30 °C, Me₄Si) 21.3, 28.3, 30.3, 45.0, 47.4, 47.5, 47.6, 49.9, 54.2, 79.1, 122.4, 123.3, 125.2, 125.5, 125.8, 126.5, 128.8, 130.9, 132.7, 134.0, 138.3, 156.0, 171.0.HRMS (ESI) [M+H]+ m/z calcd for C₂₅H₃₀N₂O₃: 407.22564, found: 407.23292

((1*S*,2*R*,3*S*,4*R*)-3-Amino-N-((*R*)-1-(naphthalen-1-yl)ethyl)bicyclo[2.2.1]hept-5-ene-2-carboxamide hydrochloride (+)-14



Yield (160 mg, 72 %), white solid, mp 255–260 °C, $[α]_{D}^{25}$ +14,5 (c 0.1 in EtOH).¹H-NMR δH (500.20 MHz, DMSO-d₆, 30 °C, Me₄Si 1.37-1.47 (2 H, m, 7-H), 1.51 (3 H, d, J = 6.76 Hz, CH₃), 3.11 (1 H, d, J = 8.1 Hz, 2-H), 3.15 (1H, s, 4-H), 3. 27 (1H, s, 1-H), 3.84 (1H, s, 3-H), 5.62–5.71 (1H, m, CH₃CHNH), 6.15-6.22 (1 H, m, 5-H), 6.26-6.350 (1 H, s, 6-H), 7.47–7.66 (4 H, m, Ar), 7.61 (1 H, d, *J* = 7.0 Hz, Ar), 7.74 (3H, brs, NH₂.HCl)7.83 (1 H, d, *J* = 7.9 Hz, Ar), 7.94 (1 H, d, *J* = 7.7 Hz, Ar), 8.11 (1 H, d, *J* = 8.2 Hz, Ar), 9.00 (1 H, d, *J* = 7.22 Hz, Ar).¹³C-NMR δC (125.62 MHz, DMSO-d₆, 30 °C, Me₄Si) 22.2, 23.8, 42.9, 44.7, 47.1, 47.4, 48.8, 52.6, 123.49, 123.5, 126.0, 126.1, 126.8, 127.8, 129.1, 130.7, 133.0, 133.8, 140.2, 171.6. HRMS (ESI) [M+H]+ m/z calcd for C₂₀H₂₃ClN₂O: 407.22564, found: 407.23292

tert-Butyl ((1R,2S,3R,4S)-3-(prop-2-yn-1-ylcarbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (-)-15



Yield (128 mg, 80 %), white solid, mp 163–164 °C, (mp lit. 4 161–163. $[\alpha]_D^{25}$ -5.1 (c 0.5 in EtOH)) (lit 4 $[\alpha]_D^{25}$ -3.7 (c 0.5 in EtOH) .¹H-NMR δ H (500.20 MHz, CDCl₃, 30 °C, Me₄Si). The ¹H NMR and ¹³C NMR spectroscopic data for the compound **15** are similar to those for the literature compounds.: ¹H-NMR δ H (500.20 MHz, CDCl₃, 30 °C, Me₄Si) = 1.29–1.50 (m, 11H, CH₃, H-7, H-7) 2.19 (t, J = 2.3 Hz, 1H, C≡CH) 2.98–3.10 (m, 3H, H-1, H-2, H-4),3.75–3.83 (m, H, H-3), 4.03–4.13 (m, 1H, NCH₂), 4.78–4.86 (m, 1H, NCH₂) 5.99 (s, 1H, NH) 6.06–6.11 (m, 1H, H-6) 6.59–6.61 (m, 1H, H-5), 13C NMR (125 MHz, CDCl₃= 28.4, 29.4, 46.3, 47.1, 47.7, 53.0, 71.7, 79.3, 79.5, 137.5, 138.8, 155.9, 172.8 ppm. HRMS (ESI) [M+H]+ m/z calcd for C₁₆H₂₃N₂O₃: 291.17032, found: 291.17053, [M+Na]+ m/z calcd for C₁₆H₂₂N₂O₃Na: 313.15226, found: 313.15261, [2M+Na]+ m/z calcd for C₃₂H₄₄N₄O₆Na: 603.31531, found: 603.31740.

General procedure for synthesis of organocatalysts 18–20

Enantiomeric *diexo* 2-aminonorbornene ester (+)-**16** was prepared from racemic *diexo* 2-aminonorbornene ester (±)-**16** by diastereomeric salt formation with *O*, *O'*-di-*p*-toluoyltartaric acid ((+)-DPTTA, 0.5 equiv) according to the literature.³ Ester (+)-**16** was transformed into amino acid via hydrolysis in microwave reactor. The enantiomeric *diexo* norbornene β -amino acid (135 mg, 0.55 mmol, 1 equiv) was dissolved in a mixture of acetone/water 2:1 (60 ml) and reacted with Fmoc-OSu (92 mg, 0.5 equiv). Crude product was evaporated, treated with aqueous Na₂CO₃ solution, than it was extracted with the mixture of CH₂Cl₂/H₂O, dried with anhydrous Na₂SO₄ and it was evaporated to get (+)-**17**.

Acid-amine coupling reactions were done with (+)-**17** in order to get **18–20** as demonstrated in **Scheme 4**. Chiral amines (0.55 mmol, 1 equiv) were added to N-Fmoc-protected *diexo* norbornene β -amino acid in presence of *N*,*N*'-diisopropylcarbodiimide (DIC) (0.72 mmol, **1**.3 equivalent) and hydroxybenzotriazole (HOBt) (0.66 mmol, **1**.2 equivalent). The reaction mixture was stirred for a day at room temperature according to the literature.⁵ The following chiral amines were used to create a catalyst: (*R*)-(+)-1-(2-naphthyl)-ethylamine (+)-**18**), (*S*)-(-)-1-phenylethylamine (-)-**19**), (*R*)-(+)-1-phenylethylamine (+)-**20**).

(9*H*-Fluoren-9-yl)methyl ((1*R*,2*R*,3*S*,4*S*)-3-(((*R*)-1-(naphthalen-2-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-18



Yield (189 mg, 65 %), white solid, mp 190–194 °C, [α]_D²⁵ +15.7 (c 0.2 in EtOH)

¹H-NMR δH (500.20 MHz, DMSO-d₆, 30 °C, Me₄Si) 1.29 (3 H, d, J = 6.8 Hz, CH₃), 1.34 (1 H, d, J = 8.5 Hz, 7-H), 2.17 (1 H, d, J = 8.5 Hz 7-H), 2.59 (1 H, s, 2-H), 2.70 (1 H, s, 4-H), 3.76 (1 H, t, J = 8.4 Hz, 1-H) 4.30-4.39 (1 H, m, CH-Fmoc) 5.00–5.09 (1 H, m, CH₃CHNH), 5.47(2 H, d, J = 7.2 Hz, CH₂-FmOc), 6.22 (2 H, d, J = 10.8 Hz, 5-H, 6-H), 6.96 (1 H, d, J = 8.8 Hz, NH), 7.32–7.96 (16 H, m, Ar), 8.43 (1 H, d, J = 7.7 Hz, NH). ¹³C-NMR δC (125.62 MHz, DMSO-d₆, 30 °C, Me₄Si) 22.7, 45.1, 45.6, 46.4, 47.2, 47.9, 48.6, 53.6, 66.2, 120.6, 124.4, 125.4, 125.6, 125.7, 126.1, 126.6, 127.6, 127.9, 128.1, 128.2, 128.4, 132.5, 133.3, 137.3, 139.8, 141.2, 142.6, 144.3, 156.1, 171.9, HRMS (ESI) [M+H]+ m/z calcd for C₃₅H₃₂N₂O₃: 529.24129, found: 529.24910.

(9*H*-Fluoren-9-yl)methyl ((1*R*,2*R*,3*S*,4*S*)-3-(((*S*)-1-phenylethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (–)-19



Yield (113 mg, 43 %), white solid, mp 153–157 °C, $[\alpha]_{D}^{25}$ –24.3 (c 0.3 in EtOH). ¹H-NMR δ H-NMR (500.20 MHz, CDCl₃, 30 °C, Me₄Si) 1.5 (1 H, d, *J* = 6.8 Hz, CH₃), 1.60 (1 H, d, *J* = 8.9 Hz, 7-H), 2.23 (1 H, d, *J* = 8.9 Hz, 7-H), 2.34 (1 H, d, *J* = 8.0 Hz, 2-H) 2.74 (1 H, s, 4-H), 2.99 (1 H, s, 1-H), 3.94 (1 H, t, *J* = 8.8 Hz, 3-H), 3.97–4.02 (1 H, m, CH₂Fmoc), 4.10 (1 H, t, J = 7,2 Hz CH-Fmoc), 4.21-4.30 (1 H, m, CH₂Fmoc), 5.06–5.16 (1 H, m, CH₃CHNH), 5.83 (1H, d, J = 9.1 Hz, NH), 5.89 (1 H, d, J = 7.7 Hz, NH), 6.19-6.27 (2 H, m, 5-H, 6-H), 7.10-7.82 (13 H, m, Ar). ¹³C-NMR δ C (125.62 MHz, CDCl₃, 30 °C, Me₄Si) 22.0, 44.6, 46.5, 47.2, 47.7, 48.5, 49.1, 53.2, 67.0, 119.9, 125.3, 126.1, 127.1, 127.4, 127.7, 128.7, 137.6, 138.8, 141.2, 141.3, 142.7, 144.0, 144.1, 156.1, 172.3. HRMS (ESI) [M+H]+ m/z calcd for C₃₁H₃₀N₂O₃: 479.22564, found: 479.23292

(9*H*-Fluoren-9-yl)methyl ((1*R*,2*R*,3*S*,4S*R*)-3-(((*R*)-1-phenylethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-20



Yield (168 mg, 64 %), white solid, mp 167–172 °C, $[\alpha]_{D}^{25}$ +20.7 (c 0.2 in EtOH). ¹H-NMR δ H-NMR (500.20 MHz, CDCl₃, 30 °C, Me₄Si) 1.38 (1 H, d, *J* = 6.7 Hz, CH₃), 1.59 (1 H, d, *J* = 9.0 Hz, 7-H), 2.18 (1 H, d, *J* = 8.9 Hz, 7-H), 2.29-2.36 (1 H, m, 2-H) 2.75 (1 H, s, 4-H), 2.93 (1 H, s, 1-H), 3.97 (1 H, t, *J* = 9.0 Hz, 3-H), 4.21 (1 H, t, *J* = 7.2 Hz CH-Fmoc) 4.26–4.32 (1 H, m, CH₂-Fmoc), 4.39-4.45 (1 H, m, CH₂-Fmoc), 5.04–5.12 (1 H, m, CH₃*CH*NH), 5.73 (1H, d, *J* = 9.5 Hz, NH), 5.82 (1 H, d, *J* = 7.2 Hz, NH), 6.15-6.25 (2 H, m, 5-H, 6-H), 7.15-7.79 (13 H, m, Ar). ¹³C-NMR δ C (125.62 MHz, CDCl₃, 30 °C, Me₄Si) 21.7, 44.8, 45.9, 47.2, 47.9, 48.3, 49.1, 53.3, 67.1, 120.0, 120.0, 125.1, 125.3, 126.2, 127.1, 127.1, 127.4, 127.7, 127.8, 128.8, 137.4, 139.0, 141.3, 141.3, 143.0, 143.9, 144.0, 156.3, 172.0. HRMS (ESI) [M+H]+ m/z calcd for C₃₁H₃₀N₂O₃: 479.22564, found: 479.23292

(S)–Phenyalanine lithium salt (–)-21

Synthesis of **21** was done according to literature procedure.⁶ White solid, mp 213-215 °C, $[\alpha]_D^{25}$ –12.0 (c 0.5 in H₂O).

(S)-Methyl 2-(3-(1-phenylethyl)ureido)benzoate (+)-22



Anthranilic acid methyl ester base (151 mg,1 mmol) was dissolved in anhydrous Et₂O (20 mL) and a 10 % excess of (S)-(–)- α -methylbenzyl isocyanate (161 mg, 1.1 mmol) was added. The reaction mixture was allowed to stand for 24 h at room temperature. After evaporation, the resulting white crystalline urea adduct was recrystallized from iPr₂O. Yield (138 mg, 49 %), light solid, mp 122–124 °C, [α]_D²⁵ +17.8 (c 0.5 in EtOH). ¹H-NMR δ H-NMR (500.20 MHz, DMSO-d₆, 30 °C, Me₄Si) 1.39 (1 H, d, *J* = 7.0 Hz, CH₃), 3.86 (3 H, s, COO*CH*₃), 4.80–4.88 (1 H, m, CH₃*CH*NH), 6.93-7.00 (1 H, m Ar), 7.20-7.50 (5 H, m, Ar), 7.42–7.51 (1 H, m, Ar), 7.88 (1H, dd, J₁=8.2 Hz, J₂ = 1.5 Hz, Ar) 7.99 (1H, d, *J* = 7.5 Hz, Ar) 8.33 (1H, d, *J* = 8.5 Hz, NH) 9.79 (1H, brs, NH) ¹³C-NMR δ C (125.62 MHz, DMSO-d₆, 30 °C, Me₄Si) 23.3, 49.3, 52.7, 114.5, 120.0, 120.8, 126.4, 127.0, 128.7, 130.90, 134.40, 143.1, 145.8, 154.3, 168.2. HRMS (ESI) [M+H]+ m/z calcd for C₁₇H₁₈N₂O₃: 283.13488, found: 283.18059

(S)-2-Amino-N-(1-phenylethyl)benzamide (+)-23



1.1 Equiv of the (S)- α -methylbenzylamine (133 mg, 1.1 mmol) was added to a suspension of isatoic anhydride (1 equiv, 163 mg, 1 mmol) in THF (50 mL). The reaction mixture was refluxed for a day. The solution was then concentrated under reduced pressure. The crude product was purified by column chromatography (hexane-ethyl acetate, 1:1) to afford (+)-**23**, as a white solid: Yield (97 mg, 40% yield), mp 140–144 °C, (lit. 7, 136–138°C) [α] $_{D}^{25}$ +114.1 (c 0.5 in EtOH) (lit. 7 +110.9 (c 1.0 in CDCl₃)). ¹H-NMR δ H-NMR (500.20 MHz, CDCl₃, 30 °C, Me₄Si) 1.58 (3 H, d, *J* = 6.9 Hz, CH₃), 5.23–5.32 (1 H, m, *CH*₃*CH*NH), 5.50 (2H, brs, NH₂) 6.24 (1 H, brs, NH), 6.59–6.70 (2 H, m, Ar), 7.14–7.46 (7 H, m, Ar). ¹³C-NMR δ C (125.62 MHz, CDCl₃, 30 °C, Me₄Si) 22.0, 48.9, 116.0, 116.6, 117.4, 126.1, 127.1, 127.4, 128.8, 132.3, 143.4, 148.9, 168.4. HRMS (ESI) [M+H]+ m/z calcd for C₁₅H₁₆N₂O: 241.13354, found: 241.13344

tert-Butyl ((15,65)-6-(((R)-1-phenylethyl)carbamoyl)cyclohex-3-en-1-yl)carbamate (+)-24



The synthesis of (1*S*,6*S*)-6-(*tert*-butoxycarbonylamino)cyclohex-3-enecarboxylic acid were performed according to the literature.⁸ The Boc protected amino acid were then dissolved (140 mg, 0.55 mmol, 1 equiv) in THF using the (S)- α -methylbenzylamine (73 mg, 0.6 mmol, 1.1 equiv) in the presence of DIC (0.72 mmol, 1.3 equiv) and HOBt (0.66 mmol, 1.2 equiv) stirring for a day at room temperature. The general method's instructions for processing the reaction were followed. Yield (113 mg, 60 %), light beige solid, mp 155–162 °C, (α]_D²⁵ +62.9 (c 0.4 in EtOH) ¹H-NMR δ H-NMR (500.20 MHz, DMSO-d₆, 30 °C, Me₄Si) 1.34 (3 H, d, *J* = 6.9 Hz, CH₃), 1.37 (9 H, s, Boc), 1.87.–1.96 (1 H, m, CH₂), 2.06–2.15 (1 H, m, CH₂), 2.18–2.30 (2 H, m,CH₂), 2.36 (1 H, m, 1-H), 3.53-3.65 (1H, m, 6-H), 4.86–4.94 (1 H, m, CH₃*CH*NH), 5.51-5.65 (2H, m, 3-H, 4-H), 6.53 (1 H, d, *J* = 8.4 Hz, NH), 7.19–7.25 (1 H, m, Ph), 7.38–7.43 (4 H, m, Ph), 7.85 (1 H, d, *J* = 7.8 Hz, NH). ¹³C-NMR δ C (125.62 MHz, DMSO-d₆, 30 °C, Me₄Si) 22.9, 28.7, 29.0, 32.8, 45.2, 48.1, 48.3, 77.9, 125.1, 126.0, 126.4, 127.1, 128.7, 144.9, 155.1, 172.5. HRMS (ESI) [M+H]+ m/z calcd for C₂₀H₂₈N₂O₃, Exact Mass: 344,20999,Molecular Weight: 344,44792

tert-Butyl ((15,2R)-2-(((R)-1-(naphthalen-2-yl)ethyl)carbamoyl)cycloheptyl)carbamate (+)-25



The synthesis of (+)-**25** were performed similar of (+)-**11**, using (1*R*,2*S*)-2-(*tert*-butoxycarbonylamino)cycloheptane carboxylic acid (141 mg, 0.55 mmol) as starting material. Yield (108 mg, 48 %), light beige solid, mp 119–124 °C, $[\alpha]_D^{25}$ +61.8 (c 0.3 in EtOH) ¹H-NMR δ H-NMR (500.20 MHz, DMSO-d₆, 30 °C, Me₄Si) 1.25 (9 H, s, Boc), 1.32–1.39 (3H, m, CH₂) 1.42 (3 H, d, *J* = 6.9 Hz CH₃), 1.62-1.92 (7 H, m, CH₂), 2.67–2.75 (1 H, m, 1–H), 3.80-3.91 (1 H, m, 2-H), 4.99–5.10 (1 H, m, CH₃*CH*NH), 5.93 (1 H, brs, NH), 7.43–7.50 (3 H, m, Ar), 7.77 (1 H, s, Ar), 7.81–7.90 (3 H, m, Ar), 8.20 (1 H, d, *J* = 7.7 Hz, NH).¹³C-NMR δ C (125.62 MHz, DMSO-d₆, 30 °C, Me₄Si) 22.5, 23.8, 24.3, 25.9, 26.7, 28.6, 28.7, 33.0, 48.1, 48.4, 52.5, 78.0, 124.4, 125.5, 126.0, 126.4, 127.8, 128.1, 128.2, 132.5, 133.3, 142.5, 155.2, 173.5. HRMS (ESI) [M+H]+ m/z calcd for C₂₅H₃₄N₂O₃, Exact Mass: 410,25694,Molecular Weight: 410,54906

1. Notes and references

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2. FT-IR spectra of the synthesized compounds 3a-25



(S)-3-Hydroxy-3-(2-oxopropyl) indolin-2-one 3a

(S)-3-Hydroxy-5-iodo-3-(2-oxopropyl) indolin-2-one 3b







(S)-5-fluoro-3-hydroxy-3-(2-oxopropyl)indolin-2-one 3d



(S)-3-hydroxy-5-nitro-3-(2-oxopropyl)indolin-2-one 3e



(S)-3-hydroxy-5-methyl-3-(2-oxopropyl)indolin-2-one 3f



(S)-4,7-dichloro-3-hydroxy-3-(2-oxopropyl)indolin-2-one 3g



(S)-5-bromo-3-hydroxy-3-(2-oxopropyl)indolin-2-one **3h**





(1R,2S,3R,4S)-Ethyl 3-(3-((S)-1-phenylethyl)ureido)bicyclo[2.2.1]hept-5-ene-2-carboxylate (-)-6



tert-Butyl ((15,2R,35,4R)-3-(((S)-1-phenylethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (-)-7









tert-Butyl ((1*R*,2*S*,3*R*,4*S*)-3-(((*S*)-1-(naphthalen-1-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate





tert-Butyl ((1*R*,2*S*,3*R*,4*S*)-3-(((*R*)-1-(naphthalen-2-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-**11**

(1*S*,2*R*,3*S*,4*R*)-3-Amino-N-((*R*)-1-(naphthalen-2-yl)ethyl)bicyclo[2.2.1]hept-5-ene-2-carboxamide hydrochloride (+)-**12**





tert-Butyl ((1*R*,2*S*,3*R*,4*S*)-3-(((*R*)-1-(naphthalen-1-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-**13**

((1*S*,2*R*,3*S*,4*R*)-3-Amino-N-((*R*)-1-(naphthalen-1-yl)ethyl)bicyclo[2.2.1]hept-5-ene-2-carboxamide hydrochloride (+)-**14**





tert-Butyl ((1R,2S,3R,4S)-3-(prop-2-yn-1-ylcarbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (-)-15

(9*H*-Fluoren-9-yl)methyl ((1*R*,2*R*,3*S*,4*S*)-3-(((*R*)-1-(naphthalen-2-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-**18**





(9*H*-Fluoren-9-yl)methyl ((1*R*,2*R*,3*S*,4*S*)-3-(((*S*)-1-phenylethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (–)-**19**

(9*H*-Fluoren-9-yl)methyl ((1*R*,2*R*,3*S*,4S*R*)-3-(((*R*)-1-phenylethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-**20**







(S)-2-Amino-N-(1-phenylethyl)benzamide (+)-23





tert-Butyl ((15,65)-6-(((R)-1-phenylethyl)carbamoyl)cyclohex-3-en-1-yl)carbamate (+)-24

tert-Butyl ((15,2R)-2-(((R)-1-(naphthalen-2-yl)ethyl)carbamoyl)cycloheptyl)carbamate (+)-25



3. ¹H and ¹³C NMR spectra of the compounds 3a-25

(S)-3-Hydroxy-3-(2-oxopropyl)indolin-2-one **3a**



(S)-3-Hydroxy-5-iodo-3-(2-oxopropyl)indolin-2-one **3b**



(S)-7-Chloro-3-hydroxy-3-(2-oxopropyl)indolin-2-one ${\bf 3c}$





(S)-5-fluoro-3-hydroxy-3-(2-oxopropyl)indolin-2-one **3d**

(S)-3-hydroxy-5-nitro-3-(2-oxopropyl)indolin-2-one **3e**





(S)-3-hydroxy-5-methyl-3-(2-oxopropyl)indolin-2-one **3f**



(S)-4,7-dichloro-3-hydroxy-3-(2-oxopropyl)indolin-2-one **3g**

(S)-5-bromo-3-hydroxy-3-(2-oxopropyl)indolin-2-one **3h**





(1R,2S,3R,4S)-Ethyl 3-(3-((S)-1-phenylethyl)ureido)bicyclo[2.2.1]hept-5-ene-2-carboxylate (-)-6

tert-Butyl ((15,2R,35,4R)-3-(((S)-1-phenylethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (-)-7





(15,2R,3S,4R)-Ethyl 3-(3-((S)-1-phenylethyl)ureido)bicyclo[2.2.1]hept-5-ene-2-carboxylate (–)-8

tert-Butyl ((1R,2S,3R,4S)-3-(((S)-1-(naphthalen-2-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate

(–)-9


tert-Butyl ((1*R*,2*S*,3*R*,4*S*)-3-(((*S*)-1-(naphthalen-1-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (–)-**10**



tert-Butyl ((1*R*,2*S*,3*R*,4*S*)-3-(((*R*)-1-(naphthalen-2-yl) ethyl) carbamoyl) bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-**11**



(1*S*,2*R*,3*S*,4*R*)-3-Amino-*N*-((*R*)-1-(naphthalen-2-yl)ethyl)bicyclo[2.2.1]hept-5-ene-2-carboxamide hydrochloride (+)-**12**



tert-Butyl ((1*R*,2*S*,3*R*,4*S*)-3-(((*R*)-1-(naphthalen-1-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-**13**



((1*S*,2*R*,3*S*,4*R*)-3-Amino-*N*-((*R*)-1-(naphthalen-1-yl)ethyl)bicyclo[2.2.1]hept-5-ene-2-carboxamide hydrochloride (+)-**14**



tert-Butyl ((1R,2S,3R,4S)-3-(prop-2-yn-1-ylcarbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (-)-15







(9*H*-Fluoren-9-yl)methyl ((1*R*,2*R*,3*S*,4*S*)-3-(((*S*)-1-phenylethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (–)-**19**



(9*H*-Fluoren-9-yl)methyl ((1*R*,2*R*,3*S*,4S*R*)-3-(((*R*)-1-phenylethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-**20**



((S)-Methyl 2-(3-(1-phenylethyl)ureido)benzoate) (+)-22



(S)-2-Amino-N-(1-phenylethyl)benzamide (+)-23









tert-Butyl ((15,2R)-2-(((R)-1-(naphthalen-2-yl)ethyl)carbamoyl)cycloheptyl)carbamate (+)-25

4. Copies of HRMS-ESI Spectra of 3a-25



(S)-3-Hydroxy-3-(2-oxopropyl)indolin-2-one 3a

(S)-3-Hydroxy-5-iodo-3-(2-oxopropyl)indolin-2-one 3b



(S)-7-Chloro-3-hydroxy-3-(2-oxopropyl)indolin-2-one 3c



(S)-5-fluoro-3-hydroxy-3-(2-oxopropyl)indolin-2-one 3d





(S)-3-hydroxy-5-nitro-3-(2-oxopropyl)indolin-2-one 3e

200000

245

250



262.55158

265

260

m/z

269.06228

270

275

280

256.04325

255

(S)-3-hydroxy-5-methyl-3-(2-oxopropyl)indolin-2-one 3f



(S)-4,7-dichloro-3-hydroxy-3-(2-oxopropyl)indolin-2-one 3g



(S)-5-bromo-3-hydroxy-3-(2-oxopropyl)indolin-2-one 3h





(1R,2S,3R,4S)-Ethyl 3-(3-((S)-1-phenylethyl)ureido)bicyclo[2.2.1]hept-5-ene-2-carboxylate (-)-6



m/z

332.5

334.0

334.5

335.5

335.0

336.0

329.5

330.0

330.5

331.0

331.5

332.0







tert-Butyl ((1R,2S,3R,4S)-3-(((S)-1-(naphthalen-2-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate







(-)-10



tert-Butyl ((1R,2S,3R,4S)-3-(((R)-1-(naphthalen-2-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate

(+)-11



(1*S*,2*R*,3*S*,4*R*)-3-Amino-N-((*R*)-1-(naphthalen-2-yl)ethyl)bicyclo[2.2.1]hept-5-ene-2-carboxamide hydrochloride (+)-**12**



tert-Butyl ((1R,2S,3R,4S)-3-(((R)-1-(naphthalen-1-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate

(+)-13



((1*S*,2*R*,3*S*,4*R*)-3-Amino-N-((*R*)-1-(naphthalen-1-yl)ethyl)bicyclo[2.2.1]hept-5-ene-2-carboxamide hydrochloride (+)-**14**





tert-Butyl ((1R,2S,3R,4S)-3-(prop-2-yn-1-ylcarbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (-)-15

(9*H*-Fluoren-9-yl)methyl ((1*R*,2*R*,3*S*,4*S*)-3-(((*R*)-1-(naphthalen-2-yl)ethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-**18**



(9*H*-Fluoren-9-yl)methyl ((1*R*,2*R*,3*S*,4*S*)-3-(((*S*)-1-phenylethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (–)-**19**



(9*H*-Fluoren-9-yl)methyl ((1*R*,2*R*,3*S*,4S*R*)-3-(((*R*)-1-phenylethyl)carbamoyl)bicyclo[2.2.1]hept-5-en-2-yl)carbamate (+)-**20**



(S)-Methyl 2-(3-(1-phenylethyl)ureido)benzoate (+)-22

T:FTMS + p ESI Full lock ms [150.0000-1000.0000] 0.66-0.72 AV: 13 NL: -



(S)-2-Amino-N-(1-phenylethyl)benzamide (+)-23





tert-Butyl ((15,65)-6-(((R)-1-phenylethyl)carbamoyl)cyclohex-3-en-1-yl)carbamate (+)-24





5. HPLC Chromatograms of 3a-h

Sample 1

Catalyst **7** applied in the isatin and acetone reaction using the optimum conditions (5) 2 budgeneral 2/2 even particular indexing 2 and (2a)

(S)-3-hydroxy-3-(2-oxopropyl) indolin-2-one (3a)

Method: ChiralPak[®] IA column Particle size: 5 μm, dimensions: 4.6 mm x 15 mm, mobile phase: hexane:i-PrOH = 80:20; flow rate: 0.5 mL/min, at range 200–600nm.



Chiral HPLC using catalyst **7** (5 mol %) in the aldol reaction of isatin **1a** (0.28 mmol) and acetone (5 mL) in the presence of LiOH (5 mol %), giving the (*S*)-enantiomer as a major product 3a in 57 % *ee* % and 96 % yield after 0.5 h (as in the paper **Table 4**, entry 1)

Catalyst **21** applied in the isatin/acetone reaction using the optimum conditions

(S)-3-hydroxy-5-iodo-3-(2-oxopropyl)indolin-2-one (3b)

Method: ChiralPak[®] IA column Particle size: 5 μ m, dimensions: 4.6 mm x 15 mm mobile phase: hexane:i-PrOH = 80:20; flow rate: 0.5 mL/min, at range 200–600nm.



Chiral HPLC using catalyst **21** (20 mol %) in the aldol reaction of 5-lodoisatin **1b** (0.28 mmol) and acetone (5 mL), giving the (*S*)-enantiomer as a major product **3b** in 65 % *ee*% and 78% yield after 48 h (as in the paper **Table 7**, entry 5)

Catalyst **21** applied in the isatin/acetone reaction using the optimum conditions **(S)-7-chloro-3-hydroxy-3-(2-oxopropyl)indolin-2-one (3c)**

Method: ChiralPak[®] IA column Particle size: 5 μ m, dimensions: 4.6 mm x 15 mm mobile phase: hexane:i-PrOH = 80:20; flow rate: 0.5 mL/min, at range 200–600nm.



Chiral HPLC using catalyst **21** (20 mol %) in the aldol reaction of 7-Chloroisatin **1c** (0.28 mmol) and acetone (5 mL), giving the (*S*)-enantiomer as a major product **3c** in 98 % *ee*% and 81 % yield after 48 h (as in the paper **Table 7**, entry 9)

Catalyst **21** applied in the isatin/acetone reaction using the optimum conditions **(S)-5-fluoro-3-hydroxy-3-(2-oxopropyl)indolin-2-one (3d)**

Method: ChiralPak[®] IA column Particle size: 5 μ m, dimensions: 4.6 mm x 15 mm mobile phase: hexane:i-PrOH = 80:20; flow rate: 0.5 mL/min, at range 200–600nm.



Chiral HPLC using catalyst **21** (20 mol %) in the aldol reaction of 5-Fluoroisatin **1d** (0.28 mmol) and acetone (5 mL), giving the (*S*)-enantiomer as a major product **3d** in 37 % *ee*% and 72% yield after 72 h (as in the paper **Table 7**, entry 12)

Catalyst **21** applied in the isatin/acetone reaction using the optimum conditions **(S)-3-hydroxy-5-nitro-3-(2-oxopropyl)indolin-2-one (3e)**

Method: ChiralPak[®] IA column Particle size: 5 μ m, dimensions: 4.6 mm x 15 mm mobile phase: hexane:i-PrOH = 80:20; flow rate: 0.5 mL/min, at range 200–600nm.



Chiral HPLC using catalyst **21** (20 mol %) in the aldol reaction of 5-Nitroisatin **1e** (0.28 mmol) and acetone (5 mL), giving the (*S*)-enantiomer as a major product **3e** in 41 % *ee*% and 73 % yield after 72 h (as in the paper **Table 7**, entry 15)

Catalyst **21** applied in the isatin/acetone reaction using the optimum conditions **(S)-3-hydroxy-5-methyl-3-(2-oxopropyl)indolin-2-one (3f)**

Method: ChiralPak[®] IA column Particle size: 5 μ m, dimensions: 4.6 mm x 15 mm mobile phase: hexane:i-PrOH = 80:20; flow rate: 0.5 mL/min, at range 200–600nm.



Chiral HPLC using catalyst **21** (20 mol %) in the aldol reaction of 5-Methylisatin **1f** (0.28 mmol) and acetone (5 mL), giving the (*S*)-enantiomer as a major product **3f** in 39 % *ee*% and 65 % yield after 72 h (as in the paper **Table 7**, entry 18)
Sample 7

Catalyst **7** applied in the isatin/acetone reaction using the optimum conditions (S)-4,7-dichloro-3-hydroxy-3-(2-oxopropyl)indolin-2-one (3g)

Method: ChiralPak[®] IA column Particle size: 5 μ m, dimensions: 4.6 mm x 15 mm mobile phase: hexane:i-PrOH = 80:20; flow rate: 0.5 mL/min, at range 200–600nm.



Chiral HPLC using catalyst **7** (5 mol %) in the aldol reaction of 4,7-Dichloroisatin **1g** (0.28 mmol) and acetone (5 mL), giving the (*S*)-enantiomer as a major product **3g** in 99 % *ee*% and 70 % yield after 72 h (as in the paper **Table 7**, entry 19)

Sample 8

Catalyst **7** applied in the isatin/acetone reaction using the optimum conditions **(S)-5-bromo-3-hydroxy-3-(2-oxopropyl)indolin-2-one (3h)**

Method: ChiralPak[®] IA column Particle size: 5 μ m, dimensions: 4.6 mm x 15 mm mobile phase: hexane:i-PrOH = 80:20; flow rate: 0.5 mL/min, at range 200–600nm.



Chiral HPLC using catalyst **7** (5 mol %) in the aldol reaction of 5-Bromoisatin **1h** (0.28 mmol) and acetone (5 mL), giving the (*S*)-enantiomer as a major product **3h** in 79 % *ee*% and 64 % yield after 72 h (as in the paper **Table 7**, entry 22)