Supplementary data file

Selective fluorescence turn-on detection of combination cisplatin-etoposide

chemotherapy based on N-CDs/GSH-CuNCs nanoprobe

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Fluorescence quantum yields (QYs) of N-CDs/GSH-CuNCs

The QYs of N-CDs and CuNCs were calculated using quinine sulfate (0.1 M H₂SO₄ as solvent) as a standard reference, i.e., at the excitation wavelength of 350 nm, its QY was 56% in 0.1 M H₂SO₄ solution. The fluorescent spectra of quinine sulfate, N-CDs and CuNCs were measured at excitation wavelengths of 350 nm and the absorbance was kept under 0.05. The QYs were determined by plotting the integrated fluorescence intensities against absorbance for the synthesized nanoparticles and quinine sulfate reference standards. The slopes of the linear fits for quinine sulfate, N-CDs, and CuNCs were 99.83, 51.7, and 60.72, respectively. These slope values were used along with the refractive indices and literature QY for quinine sulfate (56%) in the quantitative QY calculations according to the standard equation. The excellent linearity ($R^2 > 0.99$) of the intensity versus absorbance plots for all three samples provided robust slope values for input into the QY formula to determine accurate fluorescence efficiencies of 29.0 % for N-CDs and 34.0 % for CuNCs. The QYs were calculated according to the following equation:

$$\phi_{Sm} = \phi_{Qs} \times \frac{F_{Sm}}{F_{Qs}} \times \frac{A_{Qs}}{A_{Sm}} \times \frac{\eta_{Sm}}{\eta_{Qs}}$$

where Q denotes the QY of N-CDs or CuNCs; F and A are the integral area of fluorescence emission peak and UV–Vis absorbance intensity at excitation wavelength, respectively; η is the refractive index of the solvent; and the Qs and Sm represent quinine sulfate and the test samples, respectively.



Fig. S1. (A) Raw XPS spectrum of the N-CDs; High-resolution XPS spectra of C 1s (B), N 1s (C), and O 1s (D).













Fig. S2. Stability of the fluorescence response of N-CDs and GSH-CuNCs under various conditions: Effect of pH (2-11) on fluorescence intensity of (A) N-CDs, (B) GSH-CuNCs. Effect of NaCl concentration (0.01-2.0 mol L⁻¹) on fluorescence intensity of (C) N-CDs, (D) GSH-CuNCs. Effect of storage time (0-60 days) on fluorescence intensity of (E) N-CDs, (F) GSH-CuNCs. Effect of UV irradiation time (0-3 hrs) on fluorescence intensity of (G) N-CDs, GSH-CuNCs. Effect of temperature (20-60°C) on fluorescence intensity of (I) N-CDs, (J) GSH-CuNCs.



Fig. S3. Influence of (A) pH (3–11) and (B) incubation time (0-500 s) on fluorescence intensity of N-CDs/GSH-CuNCs upon reaction with CIS/ETP.



Fig. S4. The influence of different concentrations of (A) CIS $(20 - 160 \text{ ng mL}^{-1})$ and (B) ETP $(25 - 200 \text{ ng mL}^{-1})$ on the fluorescence emission of N-CDs/GSH-CuNCs.

Table S1: Comparison of this work with other reported fluorescent probes for the detection of

 CIS

Probe	LOD	Linear range	Matrix	Ref.
	(ng mL ⁻¹)	(µg mL ⁻¹)		
Rhodamine	24	0-3.0	Mitochondria	[1]
dithiocarbamate				
Rhodamine 640	68	0.6 -15	Cells and zebrafish	[2]
Fluorescent Sensor	Not given	0.15 - 1.5	Human Plasma	[3]
Array				
G-quadruplex/	Not given	0.003 - 0.15	Urine samples	[4]
Thioflavin T				
G-quadruplex	216	0.3 - 3.0	Urine samples	[5]
DNA-based				
Coumarin-466	32.7	0 - 48	Intracellular	[6]
/rhodamine-6G				
N-CDs/GSH-	5.2	0.02 - 0.16	Rabbit plasma	This
CuNCs				work



Fig. S5. TEM images of (A) aggregated CuNCs, (B) aggregated N-CDs and (C) disaggregated

N-CDs



Fig. S6. Plot of (F/F_0) versus concentration of GNT (25 – 160 ng mL⁻¹) and ETP (30 – 200 ng mL⁻¹) in spiked rabbit plasma.

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