

Supplementary Information

Comprehensive Analysis of Cationic Dye Removal from Synthetic and Industrial Wastewater Using Semi-Natural Curcumin Grafted Biochar/Poly Acrylic Acid Composite Hydrogel

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Fig. S1. Image of the hydrogel formation, before (left) and after (right) formation of the gel.

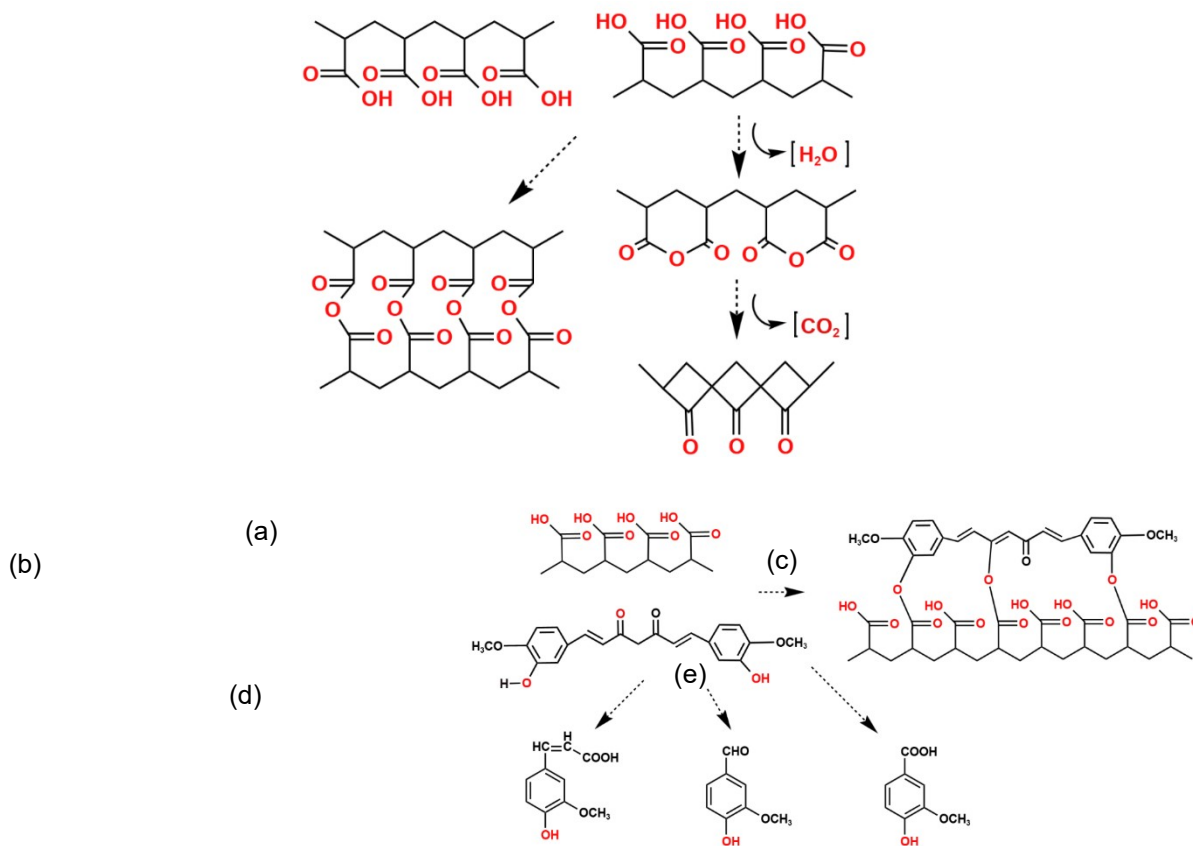


Fig. S₂ Possible degradation of synthesized composite hydrogel¹.

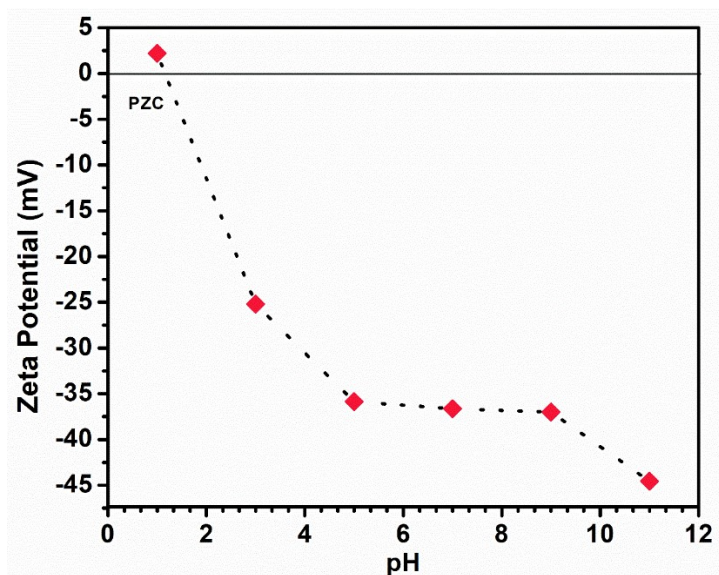


Fig. S₃ Zeta potential results of adsorbent versus pH.

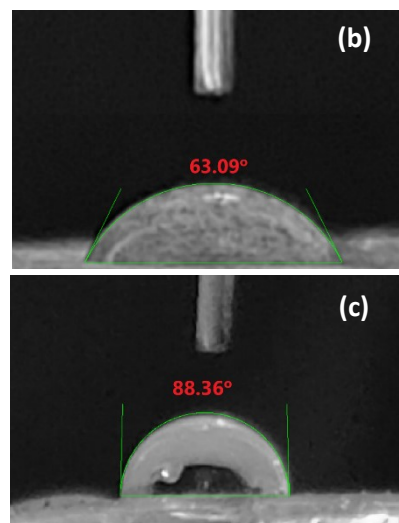
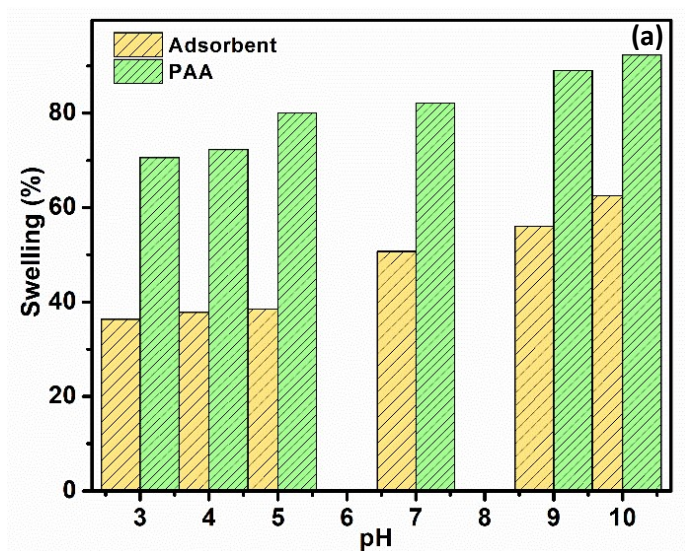


Fig.S₄ Swelling of the adsorbent and pure PAA hydrogel (a), contact angle of PAA (b) and adsorbent (c).

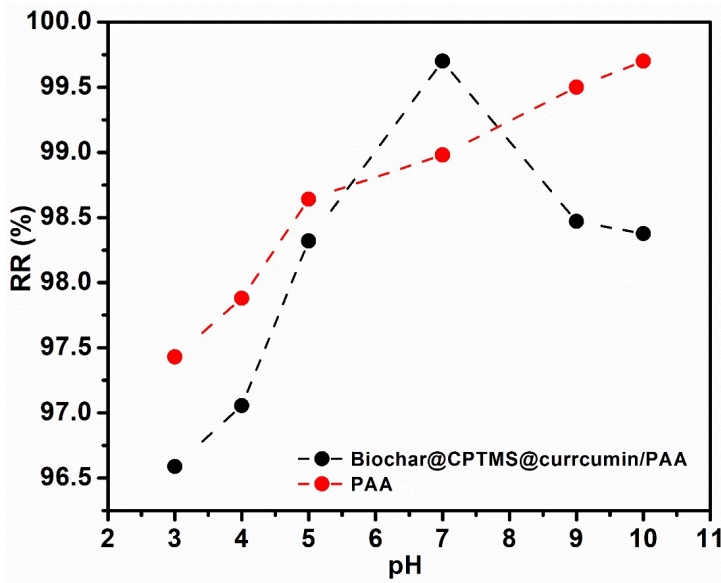


Fig. S₅ stability analysis data with pH variation after 10 days.

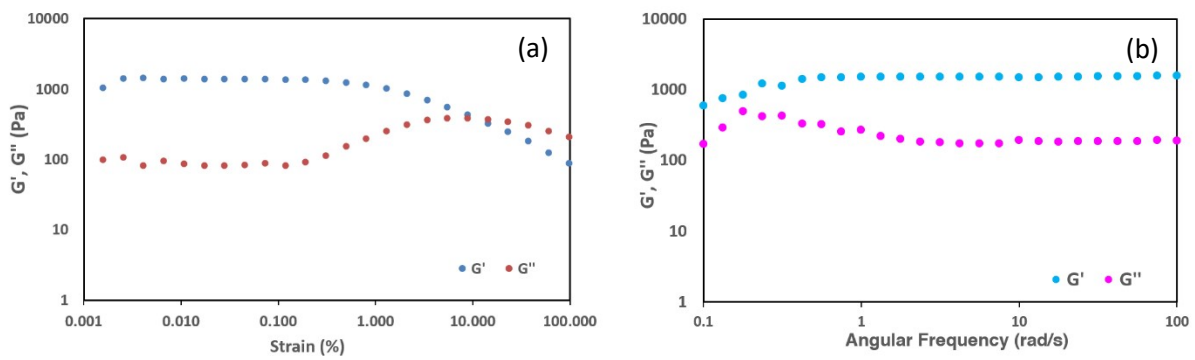


Fig. S₆. Dependence of storage (G') and loss (G'') modulus on the strain in strain sweep test (a) and on the angular frequency in frequency sweep test (b)

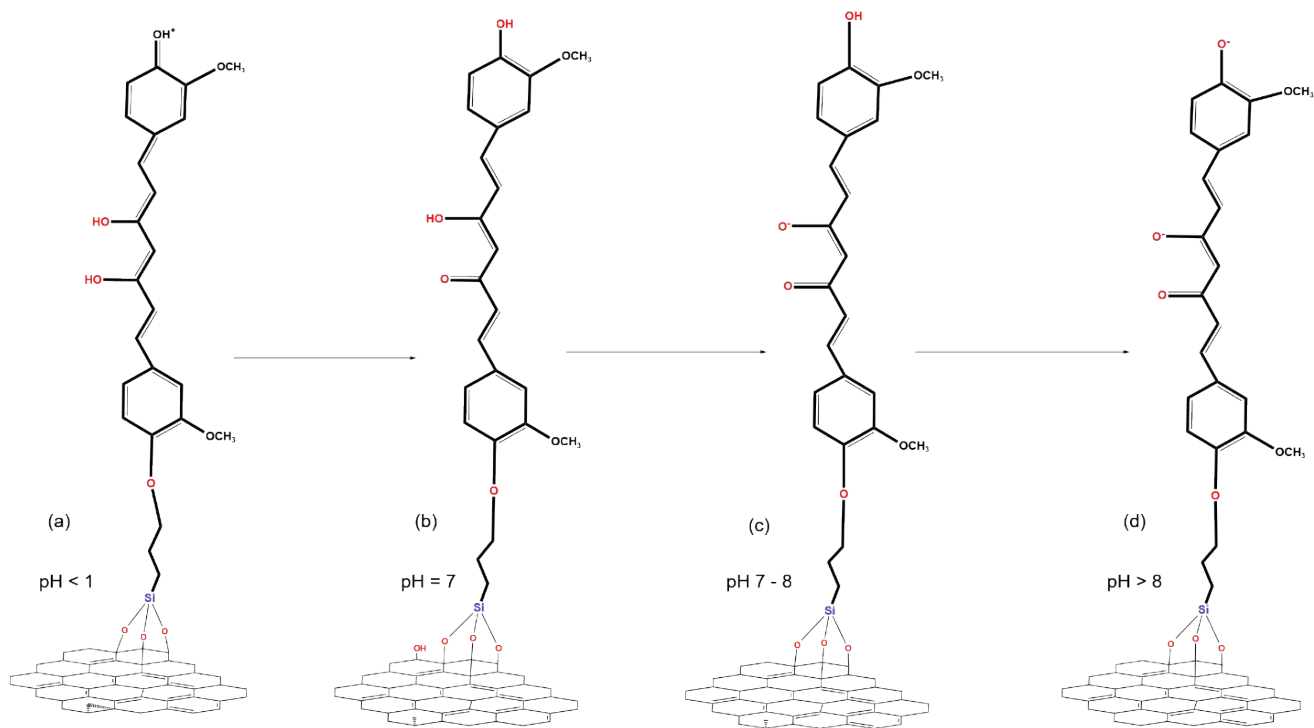


Fig. S₇ pH dependency of curcumin structure¹.

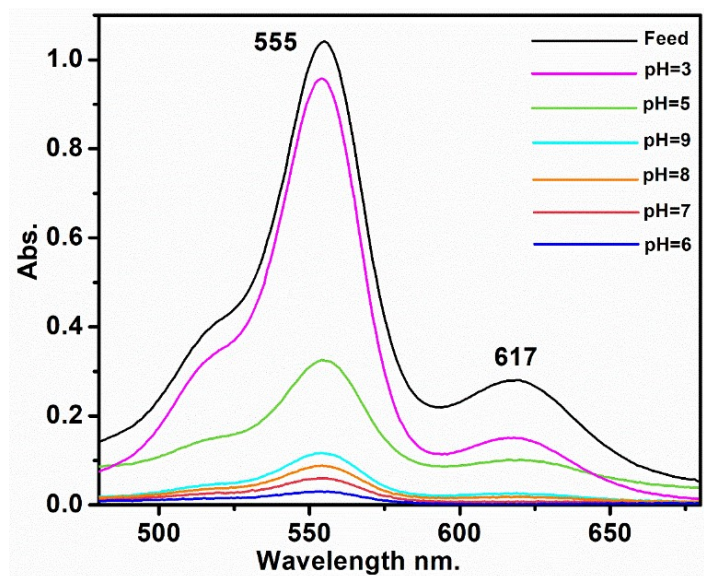


Fig. S₈ UV-Vis spectra of mixed dye solutions before and after adsorption in different pH.

Table. S₁ Linear and non-linear form of applied Isotherms and Kinetic models.

Isotherm			
Title	Linear Equation	Nonlinear Equation	Parameters
Henry's	$Q_e = K_{HE} C_e$	-----	K_{HE} denotes the Henry's adsorption constant ² .
Langmuir	$\frac{1}{Q_e} = \frac{1}{Q_m} + \frac{1}{k_L Q_m C_e}$ $R_L = \frac{1}{1 + k_L C_e}$	$Q_e = \frac{Q}{1 + kC_e}$	k_L (L.mg ⁻¹) and Q_m (mg.g ⁻¹) signify Langmuir adsorption constant and maximum adsorption capacity at equilibrium and R_L shows the favorability of model ³ .
Freundlich	$\log Q_e = \log k_f + \frac{1}{n} \log C_e$	$Q_e = k_f C_e^{\frac{1}{n}}$	k_F ((mg.g ⁻¹) (L.mg) ^{1/n}) is the Freundlich constant and 1/n implies the surface heterogeneity and shows the relative distribution of energy ⁴ .
Temkin	$Q_e = \frac{RT}{b_T} \ln A_T + \left(\frac{RT}{b_T}\right) \ln C_e$	$Q_e = \frac{RT}{b_T} \ln A_T C_e$	b_T and A_T represent Temkin models constant (KJ.mol ⁻¹) and equilibrium binding constant (L.g ⁻¹) ⁵ .
Jovanovic	$\ln Q_e = \ln Q_{max} - k_J C_e$	$Q_e = Q_{max}(1 - e^{-k_J C_e})$	k_J (L.g ⁻¹) exhibits the Jovanovic models constant and Q_{max} (mg.g ⁻¹) indicates the maximum adsorption capacity at equilibrium ⁶ .
K-C	$\frac{1}{Q_e} = \frac{1}{A_{KC} C_e^n} + \frac{B_{KC}}{A}$	$Q_e = \frac{A_{KC} C_e^n}{1 + B_{KC} C_e^n}$	A_{KC} (L ⁿ .mg ¹⁻ⁿ .g ⁻¹), B_{KC} (L.mg ⁻¹) ⁿ and n are Koble–Corrigan isotherm constants ⁷ .
Hill	$\log \left(\frac{Q_e}{Q_{SH} - Q_e} \right) = n_H \log C_e - \log k_D$	$Q_e = \left(\frac{Q_{SH} C_e^{n_H}}{K_D + C_e^{n_H}} \right)$	Q_{SH} (mg.g ⁻¹) and k_D signify the Hill maximum uptake and isotherm constant while n_H is the cooperativity of binding interaction ⁸ .
R-P	$\ln \left(k_R \frac{C_e}{Q_e} - 1 \right) = g \ln(C_e) + \ln \alpha_R$	$Q_e = \frac{k_R C_e}{1 + \alpha_R C_e^g}$	k_R (L.g ⁻¹) and α_R (L.mg ⁻¹) are the R-P isotherm constant and g implies the models exponent ⁹ .
Sips	$\frac{1}{Q_e} = \frac{1}{Q_s k_s} \left(\frac{1}{C_e} \right)^{\frac{1}{n}} + \frac{1}{Q_s}$	$Q_e = \frac{k_s Q_s C_e^{\frac{1}{n}}}{1 + k_s C_e^{\frac{1}{n}}}$	k_S (mg.L ⁻¹) and Q_s (mg.g ⁻¹) reveal the Sips adsorption isotherm constant and maximum uptake at equilibrium respectively and 1/n discloses the surface heterogeneity ¹⁰ .
Kinetic			
PFO	$Q_t = Q_e (1 - \exp^{-k_1 t})$	$\ln(Q_e - Q_t) = \ln Q_e - k_1 t$	K_1 (min ⁻¹) implies the PFO kinetic constant and Q_e (mg.g ⁻¹) is the maximal uptake at equilibrium ¹¹ .
PSO	$Q = \frac{K_2 Q_e^2 t}{1 + K_2 Q_e t}$	$\frac{t}{Q_t} = \frac{1}{K_2 Q_e^2} + \frac{t}{Q_e}$	K_2 (g.mg ⁻¹ .min ⁻¹) shows the PSO kinetic constant and Q_e (mg.g ⁻¹) is the maximum adsorption capacity ¹² .
Elovich	$Q_t = \frac{1}{\beta} \ln \alpha \beta t$	$Q_t = \frac{1}{\beta} \ln(\alpha \beta) + \frac{1}{\beta} \ln t$	α_{El} (g.mg ⁻¹) and β_{El} (g.mg ⁻¹) denoting preliminary rate of adsorption and Elovich model constant ¹³ .
LFD	$Q_t = Q_{\infty} [1 - \exp^{-Rt}]$	$\ln \left(1 - \frac{Q_t}{Q_{\infty}} \right) = -Rt$	R (min ⁻¹) is fractional attainment of equilibrium and Q_{∞} (mg.g ⁻¹) is maximum adsorption capacity at equilibrium ¹⁴ .
ID	$Q_t = k_{id} \sqrt{t} + I$		K_{id} (mg/g.min ^{0.5}) indicates the intraparticle diffusion rate constant and I (mg.g ⁻¹) is a constant correlated to thickness of boundary layer ¹² .
Bangham	$Q_t = Q_m [1 - \exp(-k_b t^n)]$	$\log \log \left(\frac{C_i}{C_i - Q_t M} \right) = \log \left(\frac{K_j M}{2.303 V} \right)$	Q_m and k_b signify the maximal uptake and Bangham rate coefficient, while n is the Bangham models power ¹⁵ .

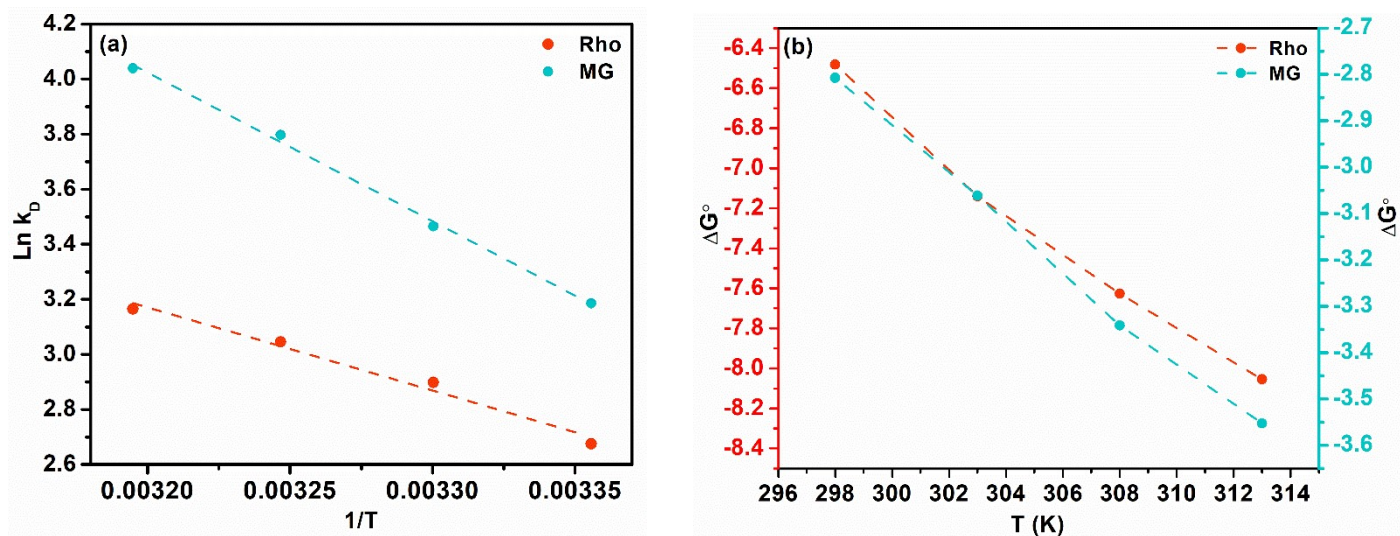


Fig. S₉ Thermodynamic plot (a), and Gibbs free energy plot (b) for the adsorption of Rho and MG onto F-Biochar.

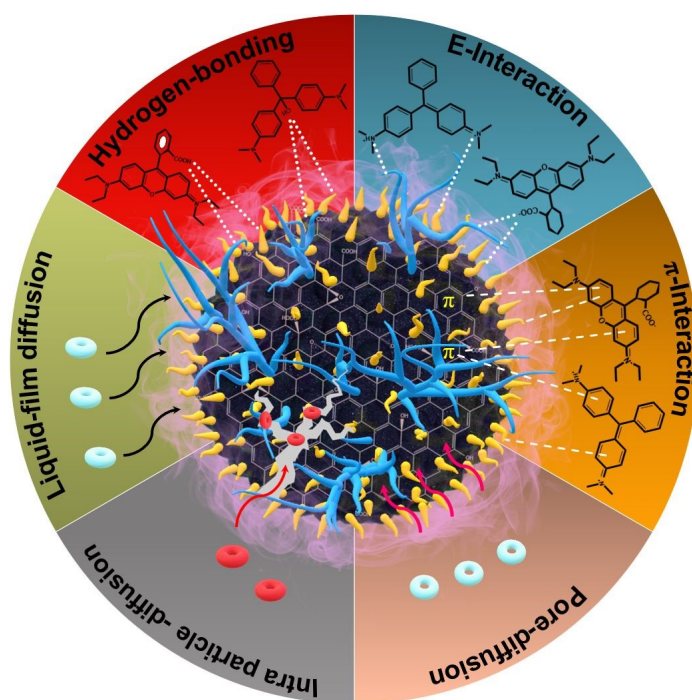


Fig. S₁₀ Graphical representation of potential adsorption phenomena during the adsorption of dye.

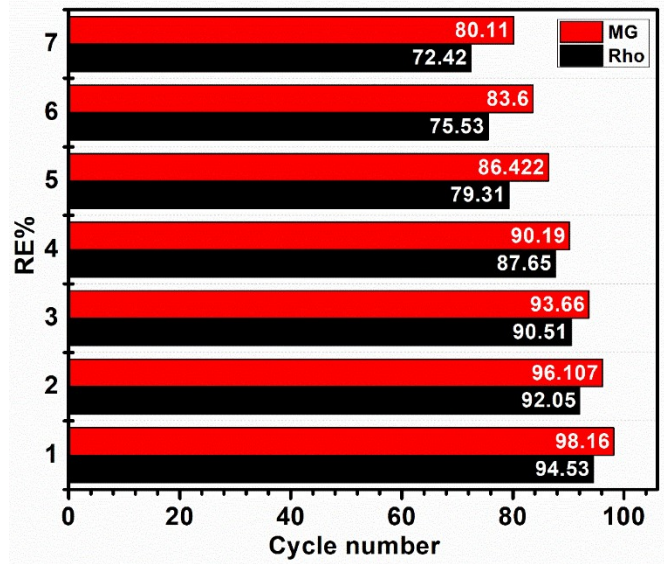


Fig. S₁₁ Reusability results.

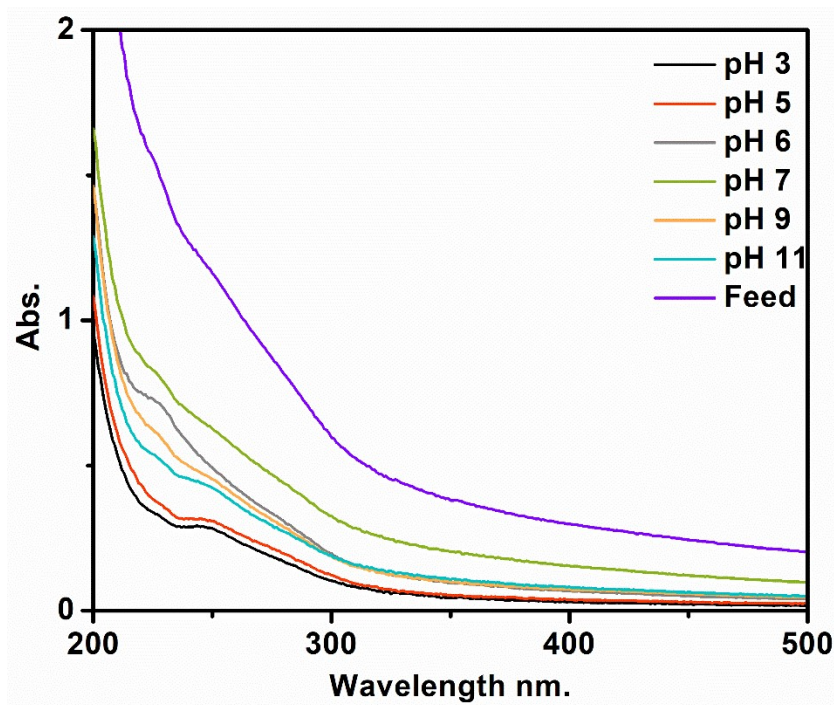


Fig. S₁₂ UV graph of industrial sample before and after adsorption in different pH.

Table.S₂ Comparison with Other Literature.

Adsorbent	Adsorbate	Q _e (mg. g ⁻¹)	Ref
CAC MercK	MG	222.22	16
Fe ₃ O ₄ @AMCA-MIL-53 (Al)	MG	262.52	17
Sawdust carbon	MG	74.5	18
Fe ₃ O ₄ @AJPL	MG	318.3	19
PB clay	MG	497.15	20
NCH based on carrageenan, AA, and AgCl	MG	270	21
poly(AAm/AACNa) hydrogel	Rho	469.48	22
chitosan hydrogel	Rho	556.9	23
Pyruvic acid (PA)-modified activated carbons	Rho	384.6	24
Gum ghatti and Fe ₃ O ₄ magnetic nanoparticles	Rho	654.87	3
Acrylic acid functionalized graphene oxide	Rho	437.1	25
PAA-AM/FA	Rho	366	26
CTS-g-P(AA-co-AMPS)/GO	MG	625.3	27
Biochar@curcumin/poly AA	MG	521.92	This Study
Biochar@curcumin/poly AA	Rho	742.53	

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