

## Supporting Information

# Automated Multistep Synthesis of 2-Pyrazolines in Continuous Flow

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## General experimental details

**General methods:** Reagents were obtained from commercial sources and used without purification. Solvents used in this work were HPLC grade and used without purification. Triethylamine was dried with calcium hydride, distilled, and stored under argon with molecular sieves (4 Å). The removal of solvent under reduced pressure was carried out on a standard rotary evaporator. All the olefins are commercially available and were used as without further purification. Activated MnO<sub>2</sub> was purchased from Sigma-Aldrich (prod. number 217646). Hydrazones were synthesized following methods previously reported.<sup>[1]</sup> 2-furoyl hydrazone readily dimerized. To avoid dimers formation the hydrazone was prepared and its solvent evaporated with the rotatory evaporator's bath at room temperature. Excess solvent and hydrazine were removed on a high vacuum pump for 6 hours. The crude was stored overnight in a freezer and diluted with THF:MeOH 90:10 to 0.5 mol L<sup>-1</sup> before being used. Unless otherwise stated, all the flow reactions were performed using a Vapourtec R-series platform. In-line IR spectroscopy was performed using the Mettler Toledo FlowIR® device.

**Chromatography:** Analytical thin-layer chromatography (TLC) was carried out on pre-coated glass plates (silica gel 60 F<sub>254</sub>) from Merck. Compound spots were visualised under ultraviolet (UV) light (254 nm) and using ninhydrin or KMnO<sub>4</sub> stain solutions. Purification of the products were performed on SiliCycle SiliaSep<sup>TM</sup> 40–63mm 60 Å flash cartridges using an automated Biotage<sup>TM</sup> flash chromatography coupled with UV detector at 254 nm.

**NMR spectroscopy:** <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra were recorded on a Bruker DPX-250 spectrometer or Bruker Avance III 500 MHz spectrometer with the residual solvent peak as the internal reference (CDCl<sub>3</sub> = 7.26 ppm, *d*<sub>6</sub>-DMSO = 2.50 ppm, CD<sub>3</sub>OD = 3.31 ppm). <sup>1</sup>H resonances are reported to the nearest 0.01 ppm. <sup>13</sup>C-NMR spectra were recorded on a Bruker DPX-250 spectrometer or Bruker Avance III 500 MHz spectrometer with the central resonance of the solvent peak as the internal reference (CDCl<sub>3</sub> = 77.16 ppm, *d*<sub>6</sub>-DMSO = 39.52 ppm, CD<sub>3</sub>OD = 49.00 ppm). All <sup>13</sup>C resonances are reported to the nearest 0.1 ppm. The multiplicity of <sup>1</sup>H signals are indicated as: s = singlet, d = doublet, dd = doublet of doublet, ddd = doublet of doublet of doublet, t = triplet, q = quadruplet, quint = quintet, sext = sextet, m = multiplet, br = broad, or combinations of thereof. Coupling constants (*J*) are quoted in Hz and reported to the nearest 0.1 Hz. Where appropriate, averages of the signals from peaks displaying multiplicity were used to calculate the value of the coupling constant. It was noted that some of the compounds were not completely stable when in CDCl<sub>3</sub> solution. That was observed when storing at 4°C overnight even after prior base washing of the CDCl<sub>3</sub>.

**High-resolution mass spectrometry (HRMS)** High-resolution mass spectra were recorded on a Synapt mass spectrometer. The injection was performed by direct infusion using 15-30 V sampling cone, 3.5 kV capillarity, 4.0 V extraction cone, 120 °C source temperature, 175 °C desolvation temperature, with 1.0 min acquisition time with scans of 1.0 second and interscan time of 0.02 s.

Unless otherwise stated, reported mass correspond to the parent molecular ion associated with a proton  $[M+H]^+$  or a sodium cation  $[M+Na]^+$  ( $^{23}\text{Na}$  isotope). All  $m/z$  values are reported to four decimal places.

## General procedure for the activation of the $\text{MnO}_2$ column

A glass column (Omnifit<sup>®</sup> column, 6.6 mm i.d.  $\times$  100 mm length) was packed with 3 g of activated  $\text{MnO}_2$  (Sigma-Aldrich 217646) between pads of celite ( $\sim 10$  mm each). A solution of THF/ $\text{Et}_3\text{N}$ /MeOH 70:20:10 (v/v/v), 32 mL was passed through the column at a flow rate of 0.5 mL/min followed by a mixture of THF/MeOH 9:1 (3 mL).

## Pumping a solution of hydrazine in THF

An initial problem was identified when the hydrazine solution was pumped continuously through the double reciprocating piston pumps (Vapourtec R-series). After a short period of operation, the performance of the pump was compromised owing to hydrazine reacting with the polymeric secondary seals of the pump, which produces particulates that sequentially caused the check valves to malfunction. Figure S1 shows secondary seals that have been exposed overnight to a hydrazine solution in THF (2 mol/L). Removal of these secondary seals from the hydrazine pump and inclusion of 10% methanol in the stock solution however prevented such blockages. This modification did not affect the operation of the pumps. Furthermore, in order to keep the primary seal clean, the back of the pump was flushed with solvent periodically using the flushing ports.



**Figure S1:** Left: Pump secondary seal exposed overnight to a solution of hydrazine in THF (1 mol/L). Right: Pump secondary seal exposed overnight to THF.

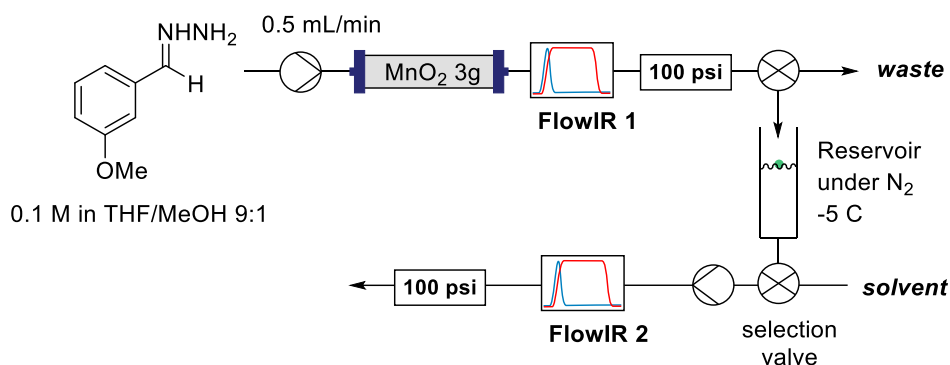
## General procedure for the synthesis of 2-pyrazolines

A 1 mL sample loop was loaded with a solution containing a 0.5 M solution of the hydrazone in THF/MeOH 90:10 (v/v). The reaction mixture was pumped with THF/MeOH 90:10 (v/v) at a flow rate of 0.5 mL/min, through an activated column containing MnO<sub>2</sub> (see above) at room temperature. Formation of the diazo was observed as the appearance of an intense reddish solution. The diazo was collected in a vial under inert atmosphere and at 0 °C. To this diazo mixture was added a solution containing the olefin (1.2 eq., 0.6 mmol) and the reaction was stirred for 2 hours at room temperature. Up to 5 injections were done using the same column.

The crude reaction mixture was concentrated under reduced pressure, and it was purified by column chromatography (hexanes/EtOAc 5-100%).

## Reservoir implementation

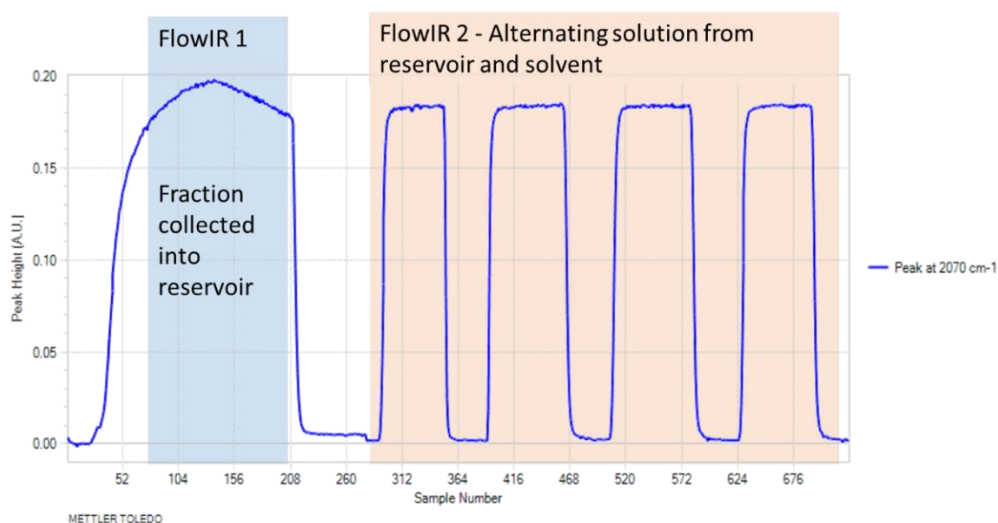
The reservoir utilised in this work was custom built in double jacketed glass, with connections compatible with the Vapourtec R-series module, where the temperature could be controlled to -5 °C and kept under nitrogen atmosphere (Figure S2).



**Figure S2:** Left) MnO<sub>2</sub> column and custom-built reservoir. Right) Equipment setup for the evaluation of the reservoir storing the diazo compound solution.



To evaluate the stability of the solution containing the diazo compound a system was setup with 2 FlowIR units (Figure S2). In this setup a solution of a hydrazone was pumped through a column containing MnO<sub>2</sub> to generate the respective diazo compounds, which was directed to the reservoir at -5°C and under nitrogen atmosphere. Once a reasonable volume of the solution was collected, it was pumped from the reservoir through a second FlowIR unit, while switching between solvent and solution. This aimed to evaluate if the solution could be split between different coupling partners. Both FlowIR units were following the IR signal at 2070 cm<sup>-1</sup>, which corresponds to the diazo functional group (Figure S3).



**Figure S3:** IR signal at 2070 cm<sup>-1</sup> during production of diazo solution and pumping it from the reservoir.

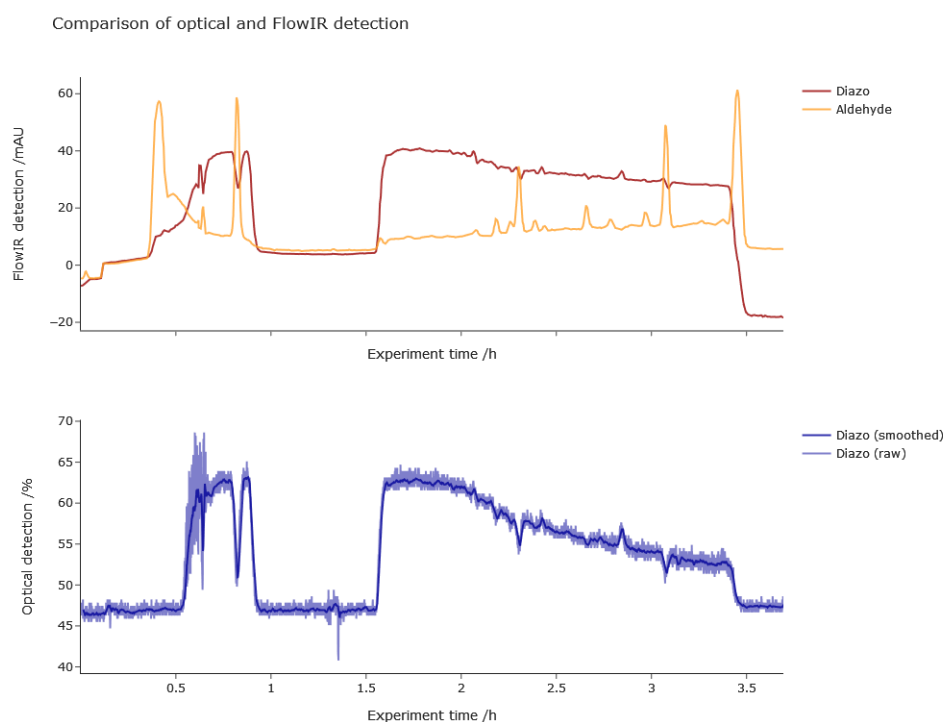
## Webcam colorimeter

A colorimeter was therefore constructed by wrapping a clear PFA tubing around a white cylinder. A 3D printed part led to a small improvement, having walls to allow two layers of tubing in the coil which intensified the colour. A USB webcam was focused on the coiled tubing, and image processing functionality within the Octopus software was used to generate a measurement of the amount of diazo present. Two separate tubing-colorimeters were monitored using a single camera image. The tubing coils were clamped in position so that their positions within the image did not change during the process. To generate a numerical value, the following computation was carried out: First the area of interest for a particular coil was selected from the image using a crop function. Then the Euclidian colour distance of each pixel from yellow was calculated to give a grayscale image. After passing through an invert function, this produces a grayscale image which is brighter when more diazo species is present.

A numerical value was then generated by taking the mean intensity of each pixel in this processed image. This calculated data had a reasonably high degree of noise from the camera; after some optimisation it was found that taking the average value of a 45 second rolling window, the signal was

smoothed to a satisfactory level. Finally, the rate at which the 'diazo intensity' variable was updated was limited to once every 5 seconds. This was useful to reduce the frequency at which the flow rate was changed for any pumps that were controlled based on this signal.

When compared to an IR measurement taken at the same point in the system, we were pleased to find that although the colorimeter had a much lower sensitivity than the IR spectrometer, it produced the same signal profile, so we were confident that this could be used to trigger actions within the control protocol.



## Iterative development of the two-step automated reaction system

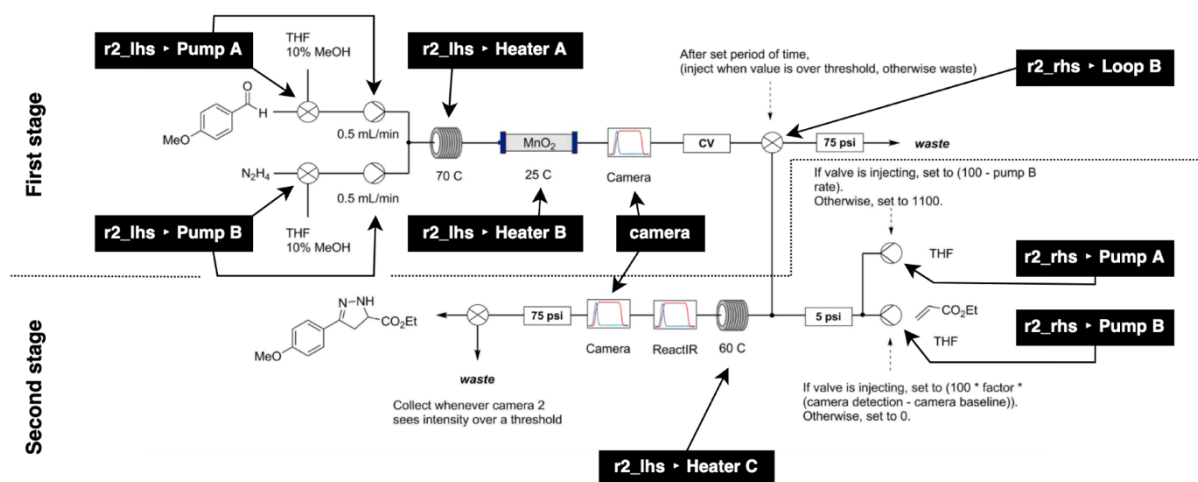
In this section, three iterations of process development for the automated two-step reaction are described, to illustrate the application of the Blocktopus GUI for implementing the control system.

### Control of a two-stage experiment

#### Instruments

- First Vapourtec R2+/R4 system (**r2\_lhs**) providing:
  - Pump for aldehyde, with low-pressure reagent/solvent selector valve (**r2\_lhs ▶ Pump A**)
  - Pump for hydrazine, with low-pressure reagent/solvent selector valve (**r2\_lhs ▶ Pump B**)
  - Reactor coil for aldehyde/hydrazine incubation (**r2\_lhs ▶ Heater A**)
  - Reactor column with  $\text{MnO}_2$  (**r2\_lhs ▶ Heater B**)
  - Reactor coil for diazo/olefin incubation (**r2\_lhs ▶ Heater C**)

- High-pressure valve for hydrazine direction to waste/second stage (**r2\_lhs ▸ Loop B**)
- Low-pressure collection/waste valve (**r2\_lhs ▸ Output**)
- Second Vapourtec R2+/R4 system (**r2\_rhs**) providing:
  - Pump for solvent (**r2\_rhs ▸ Pump A**)
  - Pump for olefin (**r2\_rhs ▸ Pump B**)
- Microsoft LifeCam Cinema webcam (**camera**) observing both colorimetry coils.
- Mettler-Toledo FlowIR detector (**ir**), initially not used in the control protocol.



## Protocol

### Setup

1. Configure the instruments (the FlowIR is not used in the control protocol for this experiment).
2. Split the image from the single camera into two separate variables:
  - a. Separately crop the image into two regions of interest;
  - b. Extract the “yellowness” of each;
  - c. Calculate their mean intensities (the image is 8-bit, so this is a value from 0-255);
  - d. Take the mean value over a 45 second rolling window, so smooth out noise;
  - e. Store the result in numeric variables (colour intensity lhs, colour intensity rhs).

A new image is read from the camera every 1 second, and the variables are updated with each new image.

### First stage

3. Set the reactor flow rates and heater temperatures.
4. Wait at least 5 minutes, and then until the heater at 70C (Heater A) is at the desired temperature.
5. Inject the reagents for 64 min;
6. Then flush the first half of the system with solvent for 20 min;
7. Then turn off heater A and heater B.

### Second stage

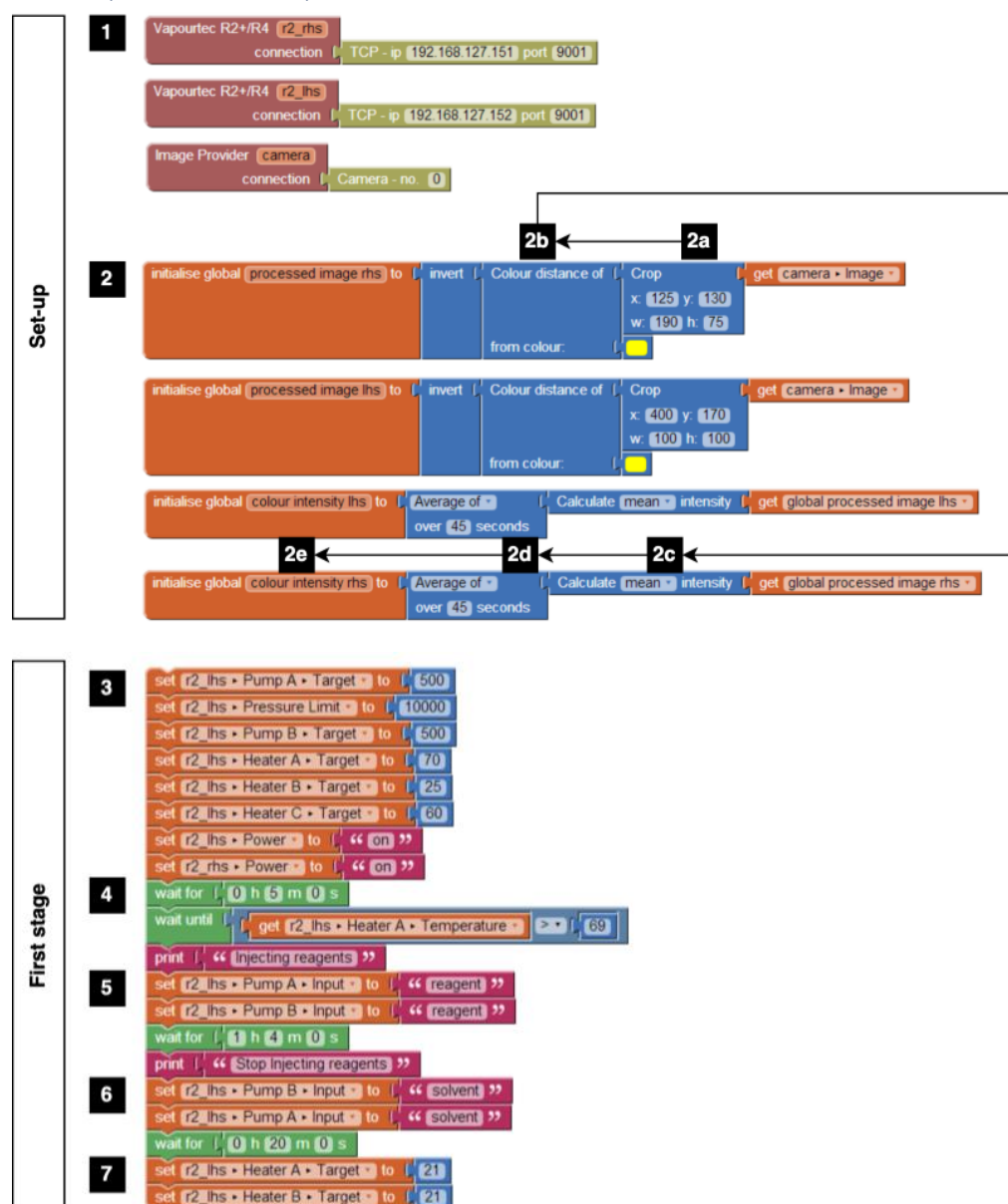
8. Allows 15 min for the reagents to pass through the heater and the MnO<sub>2</sub> column.
9. Wait for the detection of the first colorimeter to reach a set threshold (95 units; empirically determined),
10. Direct the intermediate through to the second half of the system whenever the first colorimeter reading is above a threshold of 90 units (empirically determined)
11. Until this reading drops below another threshold (88 units; empirically determined) consistently for 10 min,

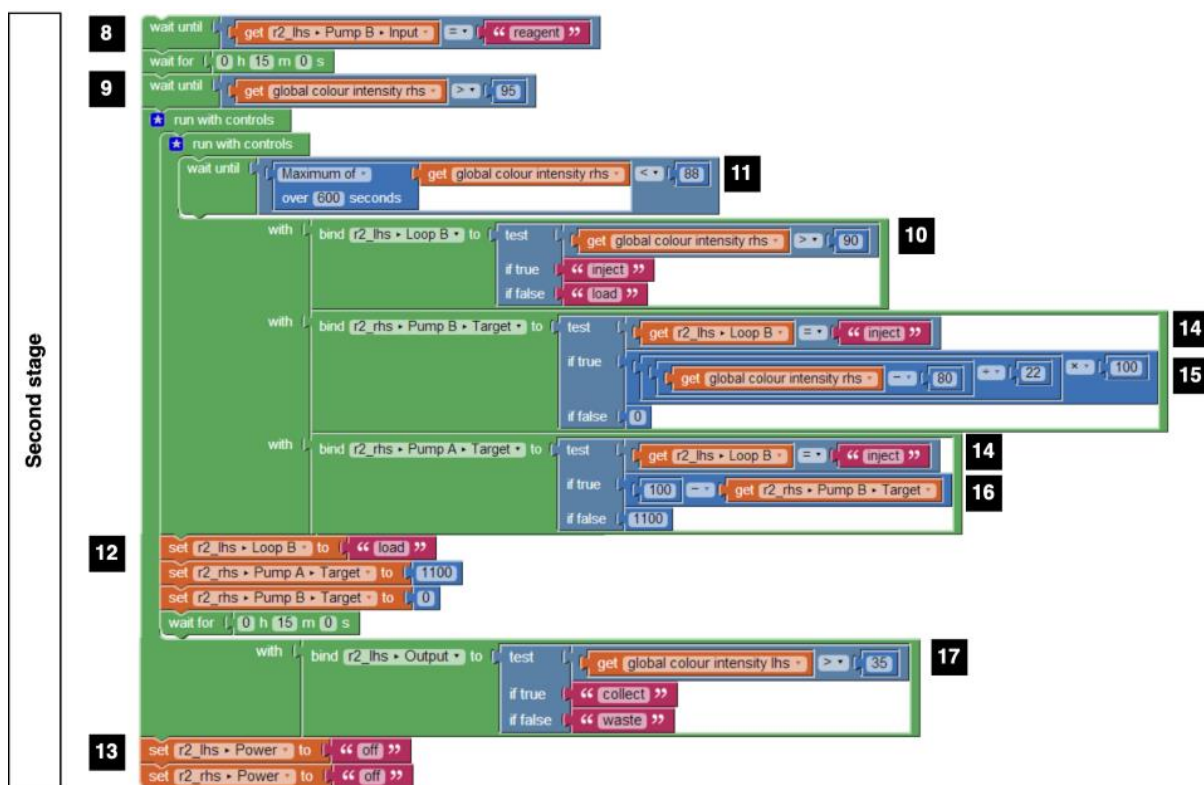
12. at which point the second half of the system is flushed with solvent ( $r2\_rhs > \text{Pump A}$ ) for 15 min,
13. and then the reactors are turned off.

### During the second stage

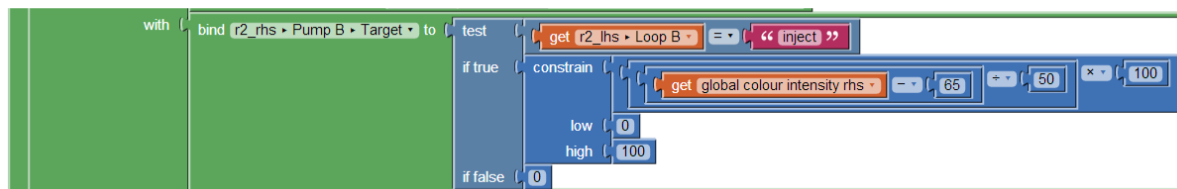
14. The second set of reagents are injected whenever the intermediate is being collected.
15. The flow rate of the pump injecting the second reagent is varied to match the amount of intermediate present (by an empirically determined factor)
16. The flow rate of make-up solvent is adjusted as well, to keep the total flow rate constant.
17. The output is directed to collection as long as the second colorimeter is detecting over a threshold value of 35 units (empirically determined).

### Blocktopus control script



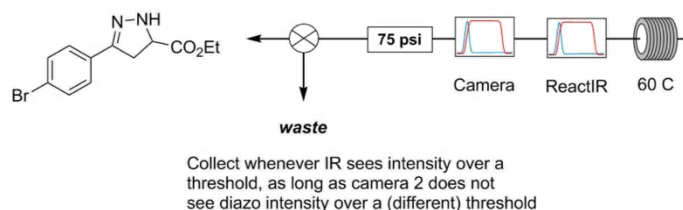


After testing, this protocol was adjusted to constrain **r2\_rhs = Pump B = Target** to have values only between 0 and 100 uL/min.

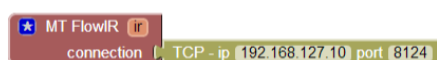


## Inclusion of FlowIR data

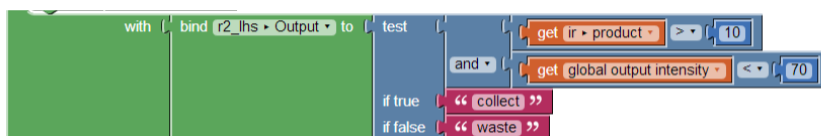
With this observation of the FlowIR data under the conditions in hand, the protocol was adjusted to include the IR data.



Add a connection to the FlowIR (**ir**) in the setup section (**1**). This required a separate script to be running on the PC alongside the Mettler Toledo software, providing access to data from a CSV file exported by the MT software at this IP address and port.



The product valve logic (17) was updated with the aim of increasing the purity of the product collected. The pyrazoline was collected whenever a certain threshold was met in the IR detection (indicating the presence of the pyrazoline C=O) but where the colorimetry detection was under another threshold (indicating a lack of unreacted diazo intermediate)



The colorimetry processing logic (2) was also modified. Since the intensity value was being used to set the flow rate of the pumps **r2\_rhs > Pump A** and **r2\_rhs > Pump B**, the fact the value of the variable was changing every second meant that there were frequent pump speed changes, which reduced the performance of the pump. By limiting the frequency at which the variable was updated, this was avoided.

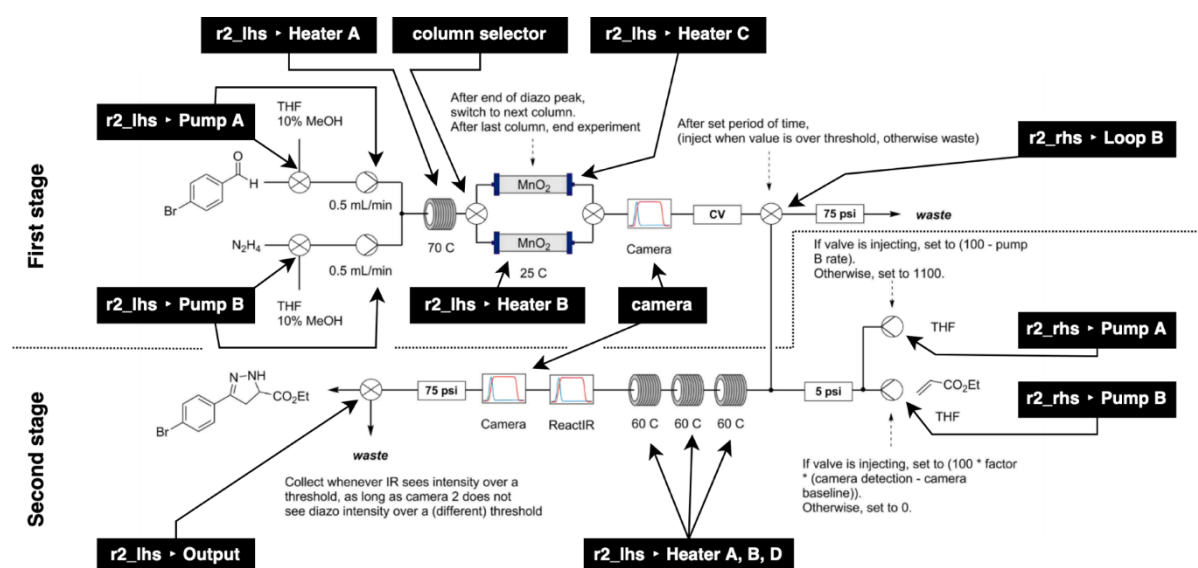


Disappointingly the use of IR data did not lead to improved reliability of the reagent collection, so control was reverted to the colorimetry data in the next iteration.

### Addition of a column switcher

In a further modification with the intention of prolonging the duration of the experiment, a Vici Valco column switching valve (**column selector**) was added, such that multiple columns of the MnO<sub>2</sub> reagent could be used in sequence.

Residence in the second stage was extended by the addition of two extra reactor coils, and these were moved to the second R2+/R4 so that heater slots on the first R2+/R4 instrument were available for extra column reactors.



When the colorimeter detected a fall in the concentration of diazo intermediate, it triggered a column switch so that fresh MnO<sub>2</sub> could be used. The remainder of the control protocol could remain the same,

since the controlled diazo selection valve (**r2\_lhs ▶ Loop B**) ensured that the intermediate was only directed towards the second stage of the reactor whenever its concentration was sufficient for the detection to be over the threshold.

## Protocol

### Setup

1. Configure the instruments (the FlowIR is not used in the control protocol for this experiment).
2. Split the image from the single camera into two separate variables:
  - a. Separately crop the image into two regions of interest;
  - b. Extract the “yellowness” of each;
  - c. Calculate their mean intensities (the image is 8-bit, so this is a value from 0-255);
  - d. Take the mean value over a 45 second rolling window, so smooth out noise;
  - e. Set the maximum refresh frequency to once per 5 seconds.
  - f. Store the result in numeric variables (**colour intensity lhs**, **colour intensity rhs**). A new image is read from the camera every 1 second, and the variables are updated with each new image.

### First stage

3. Set the reactor flow rates and heater temperatures.
4. Wait at least 10 minutes, and then until the heater at 70C (Heater A) is at the desired temperature.
5. Inject the reagents. Reagent injection is now stopped by the column-switching logic.

### Second set-up

6. Create a variable (**diazo trigger intensity**) to contain the threshold intensity (empirically determined), which is used to determine if the diazo intermediate is collected. The variable is never modified, but this allows the same value to be used in many formulae without having to enter it each time.
7. Create a variable to hold the current column number.
8. Only start the second stage once diazo begins to be injected.
9. Set the required temperatures and flow rates for **r2\_rhs**.

### Column switching

10. Allows 15 min for the reagents to pass through the heaters and the MnO<sub>2</sub> column.
11. Wait for the detection of the first colorimeter to reach a set threshold (20 units below the defined threshold frequency);
12. For each of the two columns:
  - a. Select the column using the valve.
  - b. Wait until the detection level rises above the threshold.
  - c. Wait at least 40 minutes, and then until the detection level falls below the threshold level, consistently for 10 minutes.

### Second stage (runs until the last column is exhausted)

13. Direct the intermediate through to the second half of the system whenever the first colorimeter reading is above the threshold.
14. The second set of reagents are injected whenever the intermediate is being collected.
15. The flow rate of the pump injecting the second reagent is varied to match the amount of intermediate present (by an empirically determined factor)



16. The flow rate of make-up solvent is adjusted as well, to keep the total flow rate constant.

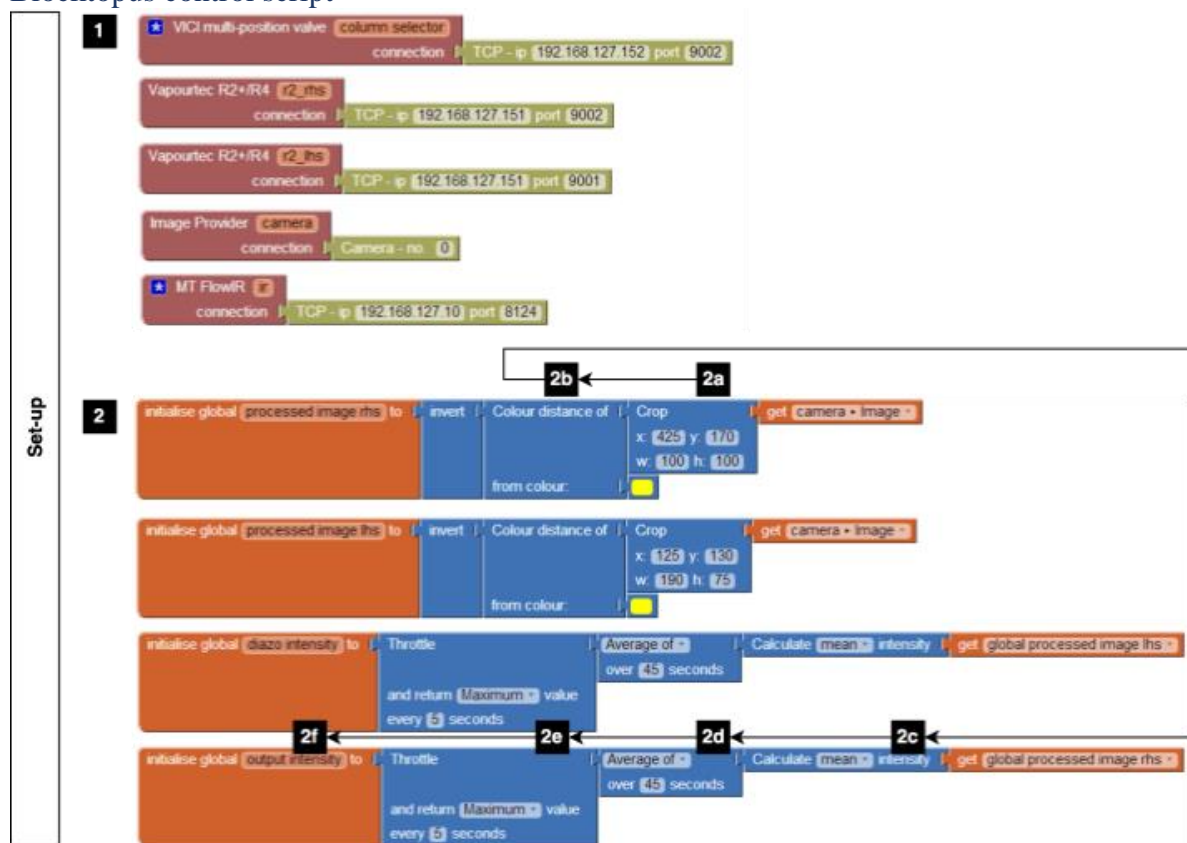
### Flushing

17. Once the last column is exhausted, stop collecting the intermediate and switch the second-stage pumps to inject solvent only.
18. Flush the second-stage part of the system for 35 min.
19. At the same time, stop injecting the aldehyde and hydrazine reagents.
20. Flush the first-stage part of the system for 20 min,
21. Turn off the heaters in the first-stage part of the system.
22. Once the flushing stages are complete, turn off the reactors.

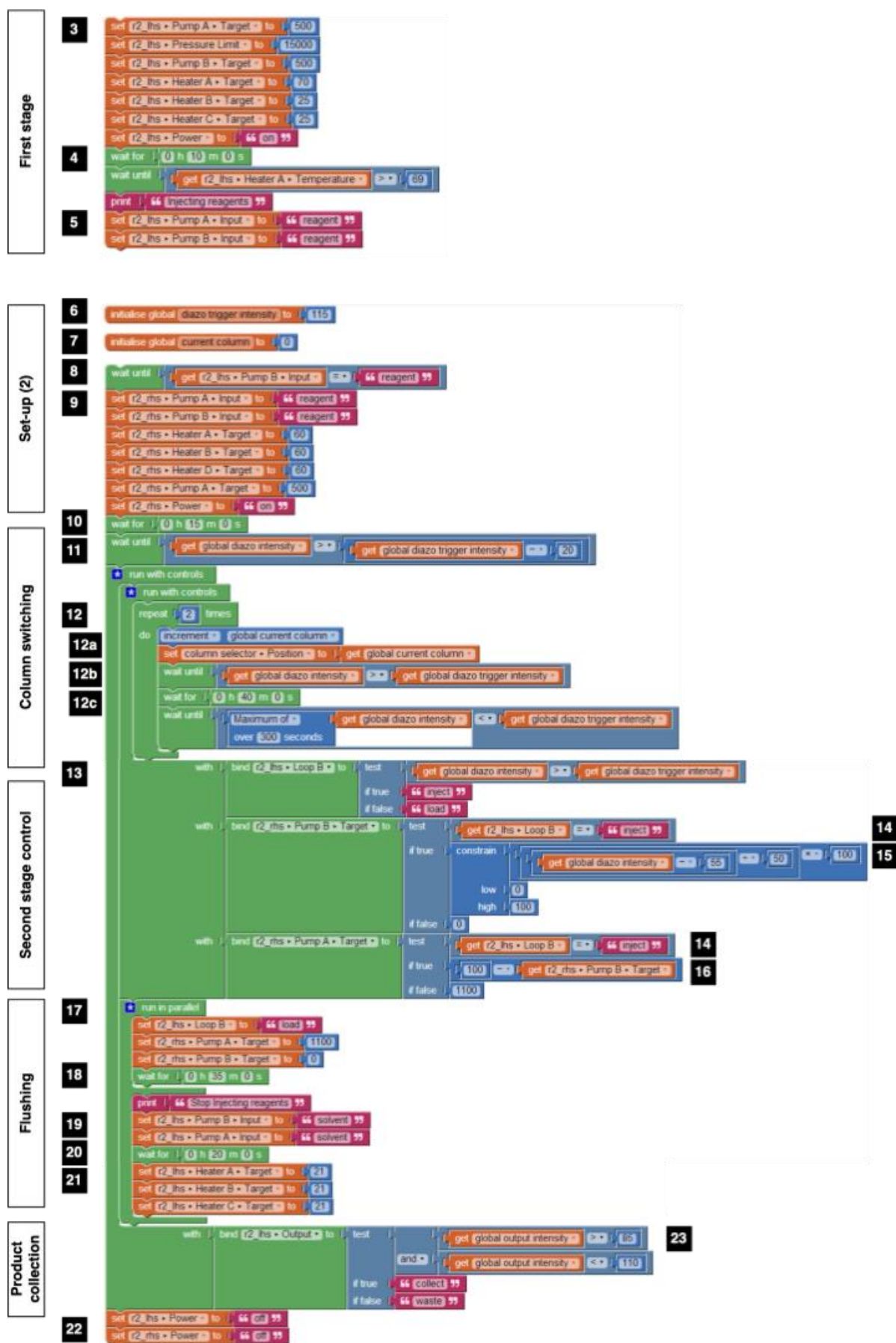
### Product collection

23. The output is directed to collection as long as the second colorimeter is detecting between 85 and 110 units (empirically determined). This had proved to be more effective than using the reading from the inline IR spectrometer.

### Blocktopus control script







## Characterization of the isolated compounds

### Ethyl 5-(4-bromophenyl)-4,5-dihydro-1H-pyrazole-3-carboxylate (1)

Isolated as a pale-yellow oil in 81% yield.

**<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, ppm)** δ 7.47 (d, *J* = 7.8 Hz, 2H), 7.18 (d, *J* = 8.2 Hz, 2H), 4.97 (t, *J* = 10.9 Hz, 1H), 4.30 (q, *J* = 7.2 Hz, 2H), 3.37 (dd, *J* = 17.2, 11.1 Hz, 1H), 2.86 (dd, *J* = 17.3, 10.1 Hz, 1H), 1.34 (t, *J* = 7.1 Hz, 3H)

**<sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, ppm)** δ 162.5 (C<sub>0</sub>), 141.6 (C<sub>0</sub>), 140.5 (C<sub>0</sub>), 132.1 (CH), 128.0 (CH), 122.0 (C<sub>0</sub>), 65.0 (CH), 61.3 (CH<sub>2</sub>), 40.0 (CH<sub>2</sub>), 14.3 (CH<sub>3</sub>)

**HRMS** expected: 319.0058 [M+Na]<sup>+</sup> found: 319.0027 [M+Na]<sup>+</sup>

### 5-(4-bromophenyl)-4,5-dihydro-1H-pyrazole-3-carbonitrile (6)

Isolated as a pale-yellow oil in 73% yield.

**<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, ppm)** δ 7.52 (d, *J* = 8.4 Hz, 2H), 7.18 (d, *J* = 8.5 Hz, 2H), 6.71 (br s, 1H), 5.02 (t, *J* = 11.1 Hz, 1H), 3.35 (dd, *J* = 16.9, 12.0 Hz, 1H), 2.84 (dd, *J* = 16.8, 10.0 Hz, 1H)

**<sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, ppm)** δ 139.1 (C<sub>0</sub>), 132.3 (CH), 128.0 (CH), 122.5 (C<sub>0</sub>), 122.3 (C<sub>0</sub>), 114.5 (C<sub>0</sub>), 64.5 (CH), 41.8 (CH<sub>2</sub>)

**HRMS** expected: 249.9980 [M+H]<sup>+</sup> found: 249.9991 [M+H]<sup>+</sup>

### Ethyl 5-(4-bromophenyl)-4-(trifluoromethyl)-4,5-dihydro-1H-pyrazole-3-carboxylate (11)

Isolated as a pale-yellow oil in 89% yield.

**<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, ppm)** δ 7.51 (d, *J* = 8.5 Hz, 2H), 7.09 (d, *J* = 8.4 Hz, 2H), 5.14 (d, *J* = 5.2 Hz, 1H), 4.32-4.24 (m, 2H), 3.88 (qd, *J* = 8.5, 5.4 Hz, 1H), 1.33 (t, *J* = 7.1 Hz, 3H)

**<sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, ppm)** δ 161.5 (C<sub>0</sub>), 138.3 (C<sub>0</sub>), 132.6 (CH), 132.3 (C<sub>0</sub>), 127.4 (CH), 124.7 (C<sub>0</sub>, q, *J* = 280.3 Hz), 123.0 (C<sub>0</sub>), 65.9 (CH), 61.6 (CH<sub>2</sub>), 56.6 (CH, q, *J* = 29.5 Hz), 14.1 (CH<sub>3</sub>)

**<sup>19</sup>F NMR (235 MHz, CDCl<sub>3</sub>, ppm)** δ -69.45 (d, *J* = 8.6 Hz, major isomer, 90%) -76.73, (d, *J* = 7.1 Hz, minor isomer, 10%)

**HRMS** expected: 365.0113 [M+H]<sup>+</sup> found: 365.0096 [M+H]<sup>+</sup>

### 5-(4-bromophenyl)-3-(phenylsulfonyl)-4,5-dihydro-1H-pyrazole (16)

Isolated as a pale-orange solid in 58% yield.

**<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, ppm)** δ 7.99-7.97 (m, 2H), 7.70 (t, *J* = 7.5 Hz, 1H), 7.60 (t, *J* = 7.6 Hz, 2H), 7.46 (d, *J* = 8.5 Hz, 2H), 7.12 (d, *J* = 8.4 Hz, 2H), 5.03 (t, *J* = 10.9 Hz, 1H), 3.42 (dd, *J* = 16.9, 11.6 Hz, 1H), 2.91 (dd, *J* = 16.8, 10.3 Hz, 1H)

**<sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, ppm)** δ 149.8 (C<sub>0</sub>), 139.1 (C<sub>0</sub>), 138.5 (C<sub>0</sub>), 134.0 (CH), 132.1 (CH), 129.3 (CH), 128.3 (CH), 127.9 (CH), 122.4 (C<sub>0</sub>), 66.0 (CH), 39.1 (CH)

**HRMS** expected: 364.9959 [M+H]<sup>+</sup> found: 364.9952 [M+H]<sup>+</sup>

### Ethyl 5-(3-nitrophenyl)-4,5-dihydro-1H-pyrazole-3-carboxylate (2)

Isolated as a white solid in 30% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 8.21-8.15 (m, 2H), 7.70 (d, *J* = 7.8 Hz, 1H), 7.57 (t, *J* = 7.6 Hz, 1H), 6.63 (br s, 1H), 5.16 (t, *J* = 11.1 Hz, 1H), 4.33 (q, *J* = 7.1 Hz, 2H), 3.49 (dd, *J* = 17.4, 11.9 Hz, 1H), 2.91 (dd, *J* = 17.3, 10.4 Hz, 1H), 1.37 (t, *J* = 7.2 Hz, 3H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 162.3 (C<sub>0</sub>), 148.6 (C<sub>0</sub>), 143.5 (C<sub>0</sub>), 141.9 (C<sub>0</sub>), 132.5 (CH), 130.1 (CH), 123.1 (CH), 121.5 (CH), 64.7 (CH), 61.4 (CH<sub>2</sub>), 40.3 (CH<sub>2</sub>), 14.3 (CH<sub>3</sub>)

**HRMS** expected: 264.0984 [M+H]<sup>+</sup> found: 264.0985 [M+H]<sup>+</sup>

### 5-(3-nitrophenyl)-4,5-dihydro-1H-pyrazole-3-carbonitrile (7)

Isolated as a pale-yellow oil in 77% yield

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 8.25-8.12 (m, 2H), 7.72-7.50 (m, 2H), 6.70 (br s, 1H), 5.18 (td, *J* = 11.2, 2.3 Hz, 1H), 3.44 (dd, *J* = 16.9, 12.0 Hz, 1H), 2.88 (dd, *J* = 16.8, 10.7 Hz, 1H)

**<sup>13</sup>C NMR (63 MHz, DMSO-*d*<sub>6</sub>, ppm)** δ 148.4 (C<sub>0</sub>), 143.9 (C<sub>0</sub>), 133.8 (CH), 130.8 (CH), 123.2 (CH), 121.8 (CH), 119.8 (C<sub>0</sub>), 116.2 (C<sub>0</sub>), 63.6 (CH), 40.8 (CH<sub>2</sub>)

**HRMS** expected: 217.0926 [M+H]<sup>+</sup> found: 217.0716 [M+H]<sup>+</sup>

### Ethyl 5-(3-nitrophenyl)-4-(trifluoromethyl)-4,5-dihydro-1H-pyrazole-3-carboxylate (12)

Isolated as a white solid in 45% yield.

**<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, ppm)** δ 8.25-8.19 (m, 1H), 8.12 (s, 1H), 7.63-7.60 (m, 2H), 7.31 (br s, 1H), 5.33 (d, *J* = 5.8 Hz, 1H), 4.40-4.21 (m, 2H), 3.94 (qd, *J* = 8.3, 5.9 Hz, 1H), 1.34 (t, *J* = 7.1 Hz, 3H)

**<sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, ppm)** δ 161.2 (C<sub>0</sub>), 148.7 (C<sub>0</sub>), 141.3 (C<sub>0</sub>), 132.5 (C<sub>0</sub>), 131.9 (CH), 124.4 (C<sub>0</sub>, q, *J* = 280 Hz), 123.6 (CH), 121.0 (CH), 65.2 (CH<sub>2</sub>, q, *J* = 2.4 Hz), 61.6 (CH), 56.9 (CH<sub>2</sub>, q, *J* = 30 Hz), 14.0 (CH<sub>3</sub>)

**<sup>19</sup>F NMR (235 MHz, CDCl<sub>3</sub>, ppm)** δ -69.22 (d, *J* = 4.7 Hz)

**HRMS** expected: 354.0677 [M+Na]<sup>+</sup> found: 354.0648 [M+Na]<sup>+</sup>

### **5-(3-nitrophenyl)-3-(phenylsulfonyl)-4,5-dihydro-1H-pyrazole (17)**

Isolated as a colourless oil in 79% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 8.17-8.12 (m, 2H), 7.99-7.96 (m, 2H), 7.72-7.50 (m, 5H), 5.20 (t, *J* = 11.0 Hz, 1H), 3.52 (dd, *J* = 16.7, 11.2 Hz, 1H), 2.91 (dd, *J* = 16.8, 10.6 Hz, 1H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 149.5 (C<sub>0</sub>), 148.3 (C<sub>0</sub>), 142.2 (C<sub>0</sub>), 138.2 (C<sub>0</sub>), 134.1 (CH), 132.5 (CH), 130.0 (CH), 129.4 (CH), 128.1 (CH), 123.2 (CH), 121.3 (CH), 65.6 (CH), 39.3 (CH<sub>2</sub>)

**HRMS** expected: 354.0525 [M+Na]<sup>+</sup> found: 354.0520 [M+Na]<sup>+</sup>

### **Ethyl 5-(pyridin-3-yl)-4,5-dihydro-1H-pyrazole-3-carboxylate (3)**

Isolated as a pale-yellow solid in 70% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 8.48-8.45 (m, 2H), 7.63 (dt, *J* = 7.9, 1.8 Hz, 1H), 7.23 (dd, *J* = 7.9, 4.9 Hz, 1H), 7.01 (br s, 1H), 4.99 (t, *J* = 11.0 Hz, 1H), 4.24 (q, *J* = 7.2 Hz, 2H), 3.36 (dd, *J* = 17.3, 11.9 Hz, 1H), 2.82 (dd, *J* = 17.2, 10.2 Hz, 1H), 1.28 (t, *J* = 7.1 Hz, 3H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 162.2 (C<sub>0</sub>), 149.1 (CH), 147.8 (CH), 141.1 (C<sub>0</sub>), 136.8 (C<sub>0</sub>), 133.8 (CH), 123.6 (CH), 62.8 (CH), 61.0 (CH<sub>2</sub>), 39.6 (CH<sub>2</sub>), 14.0 (CH<sub>3</sub>)

**HRMS** expected: 220.1086 [M+H]<sup>+</sup> found: 220.1074 [M+H]<sup>+</sup>

### **5-(pyridin-3-yl)-4,5-dihydro-1H-pyrazole-3-carbonitrile (8)**

Isolated as a pale-yellow solid in 41% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 8.60 (dd, *J* = 5.0, 1.4 Hz, 1H), 8.54 (d, *J* = 1.8 Hz, 1H), 7.68 (dt, *J* = 8.0, 1.9 Hz, 1H), 7.35 (dd, *J* = 7.8, 4.8 Hz, 1H), 6.88 (br s, 1H), 5.10 (dd, *J* = 11.2, 1.8 Hz, 1H), 3.41 (dd, *J* = 16.8, 12.0 Hz, 1H), 2.89 (dd, *J* = 16.9, 10.3 Hz, 1H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 150.1 (CH), 148.1 (CH), 135.6 (C<sub>0</sub>), 133.9 (CH), 124.1 (CH), 122.5 (C<sub>0</sub>), 114.2 (C<sub>0</sub>), 62.8 (CH), 41.8 (CH<sub>2</sub>)

**HRMS** expected: 173.0827 [M+H]<sup>+</sup> found: 173.0810 [M+H]<sup>+</sup>

**Ethyl 5-(pyridin-3-yl)-4-(trifluoromethyl)-4,5-dihydro-1H-pyrazole-3-carboxylate (13)**

Isolated as a pale-yellow oil in 56% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 8.53-8.51 (m, 1H), 8.43 (s, 1H), 7.54 (d, *J* = 8.0, 1H), 7.33-7.27 (m, 1H), 5.19 (d, *J* = 5.6 Hz, 1H), 4.34-4.20 (m, 2H), 3.95-3.82 (m, 1H), 1.31 (t, *J* = 7.1 Hz, 3H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 161.3 (C<sub>0</sub>), 150.1 (CH), 147.3 (CH), 135.0 (C<sub>0</sub>), 133.6 (CH), 131.8 (C<sub>0</sub>), 124.5 (C<sub>0</sub>, q, *J* = 280.0 Hz), 124.1 (CH), 64.0 (q, *J* = 2.4 Hz), 61.4 (CH<sub>2</sub>), 56.4 (CH, q, *J* = 29.7 Hz), 14.0 (CH<sub>3</sub>)

**<sup>19</sup>F NMR (235 MHz, CDCl<sub>3</sub>, ppm)** δ -69.45 (d, *J* = 8.4 Hz)

**HRMS** expected: 288.0960 [M+H]<sup>+</sup> found: 288.0973 [M+H]<sup>+</sup>

**Ethyl 5-(4-fluorophenyl)-5-methyl-4,5-dihydro-1H-pyrazole-3-carboxylate (4)**

Isolated as a pale-yellow oil in 74% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 7.39-7.33 (m, 2H), 7.06-6.99 (m, 2H), 4.30 (q, *J* = 7.1 Hz, 2H), 3.12 (d, *J* = 17.1 Hz, 1H), 3.02 (d, *J* = 17.1 Hz, 1H), 1.60 (s, 3H), 1.35 (t, *J* = 7.1 Hz, 3H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 162.7 (C<sub>0</sub>), 161.9 (C<sub>0</sub>, d, *J* = 246.1 Hz), 141.2 (C<sub>0</sub>, d, *J* = 3.2 Hz), 140.7 (C<sub>0</sub>), 127.0 (CH, d, *J* = 8.1 Hz), 115.4 (CH, d, *J* = 21.4 Hz), 69.4 (C<sub>0</sub>), 61.1 (CH<sub>2</sub>), 46.5 (CH<sub>2</sub>), 27.4 (CH<sub>3</sub>), 14.3 (CH<sub>3</sub>)

**<sup>19</sup>F NMR (235 MHz, CDCl<sub>3</sub>, ppm)** δ -115.53 (tt, *J* = 8.8, 5.2 Hz)

**HRMS** expected: 273.1015 [M+Na]<sup>+</sup> found: 273.0987 [M+Na]<sup>+</sup>

**5-(4-fluorophenyl)-5-methyl-4,5-dihydro-1H-pyrazole-3-carbonitrile (9)**

Isolated as a pale-yellow oil in 82% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 7.39-7.31 (m, 2H), 7.11-7.02 (m, 2H), 3.03 (s, 2H), 1.64 (s, 3H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 162.0 (C<sub>0</sub>, d, *J* = 246.9 Hz), 139.9 (C<sub>0</sub>, d, *J* = 3.2 Hz), 126.8 (CH, d, *J* = 7.8 Hz), 121.3 (C<sub>0</sub>), 115.7 (CH, d, *J* = 21.4 Hz), 114.8 (C<sub>0</sub>), 69.5 (C<sub>0</sub>), 48.4 (CH<sub>2</sub>), 27.0 (CH<sub>3</sub>)

**<sup>19</sup>F NMR (235 MHz, CDCl<sub>3</sub>, ppm)** δ -114.64 (tt, *J* = 8.6, 5.1 Hz)

**HRMS** expected: 204.0937 [M+H]<sup>+</sup> found: 204.0904 [M+H]<sup>+</sup>

### **5-(4-fluorophenyl)-5-methyl-3-(phenylsulfonyl)-4,5-dihydro-1H-pyrazole (19)**

Isolated as a pale-yellow oil in 38% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 8.07-7.94 (m, 2H), 7.70-7.54 (m, 3H), 7.32-7.26 (m, 2H), 7.04-6.97 (m, 2H), 3.16 (d, *J* = 16.5 Hz, 1H), 3.02 (d, *J* = 16.5 Hz, 1H), 1.58 (s, 3H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 162.0 (C<sub>0</sub>, d, *J* = 246.8 Hz), 148.6 (C<sub>0</sub>), 140.1 (C<sub>0</sub>, d, *J* = 3.3 Hz), 138.8 (C<sub>0</sub>), 134.0 (CH), 129.4 (CH), 128.2 (CH), 127.0 (CH, d, *J* = 8.1 Hz), 115.5 (CH, d, *J* = 21.4 Hz), 71.1 (C<sub>0</sub>), 45.5 (CH<sub>2</sub>), 27.0 (CH<sub>3</sub>)

**<sup>19</sup>F NMR (235 MHz, CDCl<sub>3</sub>, ppm)** δ -114.88 (tt, *J* = 8.7, 5.2 Hz)

**HRMS** expected: 319.0916 [M+H]<sup>+</sup> found: 319.0905 [M+H]<sup>+</sup>

### **Ethyl 5-methyl-5-phenyl-4,5-dihydro-1H-pyrazole-3-carboxylate (14)**

Isolated as a pale-yellow oil in 86% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 7.42-7.25 (m, 5H), 4.29 (q, *J* = 7.2 Hz, 2H), 3.09 (d, *J* = 2.3 Hz, 2H), 1.60 (s, 3H), 1.34 (t, *J* = 7.2 Hz, 3H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 162.8 (C<sub>0</sub>), 145.4 (C<sub>0</sub>), 140.8 (C<sub>0</sub>), 128.7 (CH), 127.3 (CH), 125.1 (CH), 69.9 (C<sub>0</sub>), 61.1 (CH<sub>2</sub>), 46.3 (CH<sub>2</sub>), 27.5 (CH<sub>3</sub>), 14.3 (CH<sub>3</sub>)

**HRMS** expected: 255.1109 [M+Na]<sup>+</sup> found: 255.1095 [M+Na]<sup>+</sup>

### **5-methyl-5-phenyl-4,5-dihydro-1H-pyrazole-3-carbonitrile (18)**

Isolated as a pale-yellow oil in 68% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 7.42-7.32 (m, 5H), 3.05 (s, 2H), 1.65 (s, 3H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 144.0 (C<sub>0</sub>), 128.9 (CH), 127.8 (CH), 125.0 (CH), 121.5 (C<sub>0</sub>), 114.8 (C<sub>0</sub>), 69.9 (C<sub>0</sub>), 48.4 (CH<sub>2</sub>), 27.0 (CH<sub>3</sub>)

**HRMS** expected: 186.1031 [M+H]<sup>+</sup> found: 186.1026 [M+H]<sup>+</sup>

### **Ethyl 5-(furan-2-yl)-4,5-dihydro-1H-pyrazole-3-carboxylate (5)**

Isolated as a pale-yellow oil in 61% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 7.39-7.37 (m, 1H), 6.33 (dd, *J* = 3.2, 1.9 Hz, 1H), 6.24 (d, *J* = 3.6 Hz, 1H), 5.04 (dd, *J* = 11.0, 8.4 Hz, 1H), 4.33 (q, *J* = 7.2 Hz, 2H), 3.26 (dd, *J* = 17.0, 11.1 Hz, 1H), 3.15 (dd, *J* = 17.2, 8.4 Hz, 1H), 1.37 (t, *J* = 7.1 Hz, 3H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 162.4 (C<sub>0</sub>), 153.3 (C<sub>0</sub>), 142.7 (CH), 142.6 (C<sub>0</sub>), 110.4 (CH), 106.5 (CH), 61.3 (CH<sub>2</sub>), 58.3 (CH), 36.1 (CH<sub>2</sub>), 14.3 (CH<sub>3</sub>)

**HRMS** expected: 209.0926 [M+H]<sup>+</sup> found: 209.0918 [M+H]<sup>+</sup>

### **5-(furan-2-yl)-4,5-dihydro-1H-pyrazole-3-carbonitrile (16)**

Isolated as a pale-yellow oil in 65% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 7.40 (d, *J* = 1.1 Hz, 1H), 6.36 (dd, *J* = 3.2, 1.8 Hz, 1H), 6.28 (d, *J* = 3.3 Hz, 1H), 5.07 (dd, *J* = 11.5, 8.2 Hz, 1H), 3.23 (dd, *J* = 16.8, 11.5 Hz, 1H), 3.09 (dd, *J* = 16.7, 8.2 Hz, 1H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 151.9 (C<sub>0</sub>), 143.0 (CH), 123.1 (C<sub>0</sub>), 114.3 (C<sub>0</sub>), 110.5 (CH), 107.3 (CH), 57.9 (CH), 38.0 (CH<sub>2</sub>)

**HRMS** expected: 162.0656 [M+H]<sup>+</sup> found: 162.0667 [M+H]<sup>+</sup>

### **Ethyl 5-(furan-2-yl)-4-(trifluoromethyl)-4,5-dihydro-1H-pyrazole-3-carboxylate (15)**

Isolated as a pale-yellow oil in 51% yield.

**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 7.41-7.40 (m, 1H), 6.65 (br s, 1H), 6.38-6.35 (m, 1H), 6.33-6.31 (m, 1H), 5.22 (d, *J* = 4.5 Hz, 1H), 4.41-4.29 (m, 3H), 1.39 (t, *J* = 7.1 Hz, 3H)

**<sup>13</sup>C NMR of the major isomer (63 MHz, CDCl<sub>3</sub>, ppm)** δ 161.3 (C<sub>0</sub>), 150.8 (C<sub>0</sub>), 146.0 (C<sub>0</sub>), 143.6 (CH), 124.6 (C<sub>0</sub>, q, *J* = 280.5 Hz), 110.7 (CH), 107.7 (CH), 61.6 (CH<sub>2</sub>), 60.1 (CH, q, *J* = 2.7 Hz), 53.3 (CH, q, *J* = 30.0 Hz), 14.1 (CH<sub>3</sub>)

**<sup>19</sup>F NMR (235 MHz, CDCl<sub>3</sub>, ppm)** δ -69.41 (d, *J* = 8.7 Hz, 77%), -76.27 (d, *J* = 7.0 Hz, 23%)

**HRMS** expected: 299.0620 [M+Na]<sup>+</sup> found: 299.0634 [M+Na]<sup>+</sup>

### **5-(furan-2-yl)-3-(phenylsulfonyl)-4,5-dihydro-1H-pyrazole (20)**

Isolated as a pale-yellow oil in 75% yield.

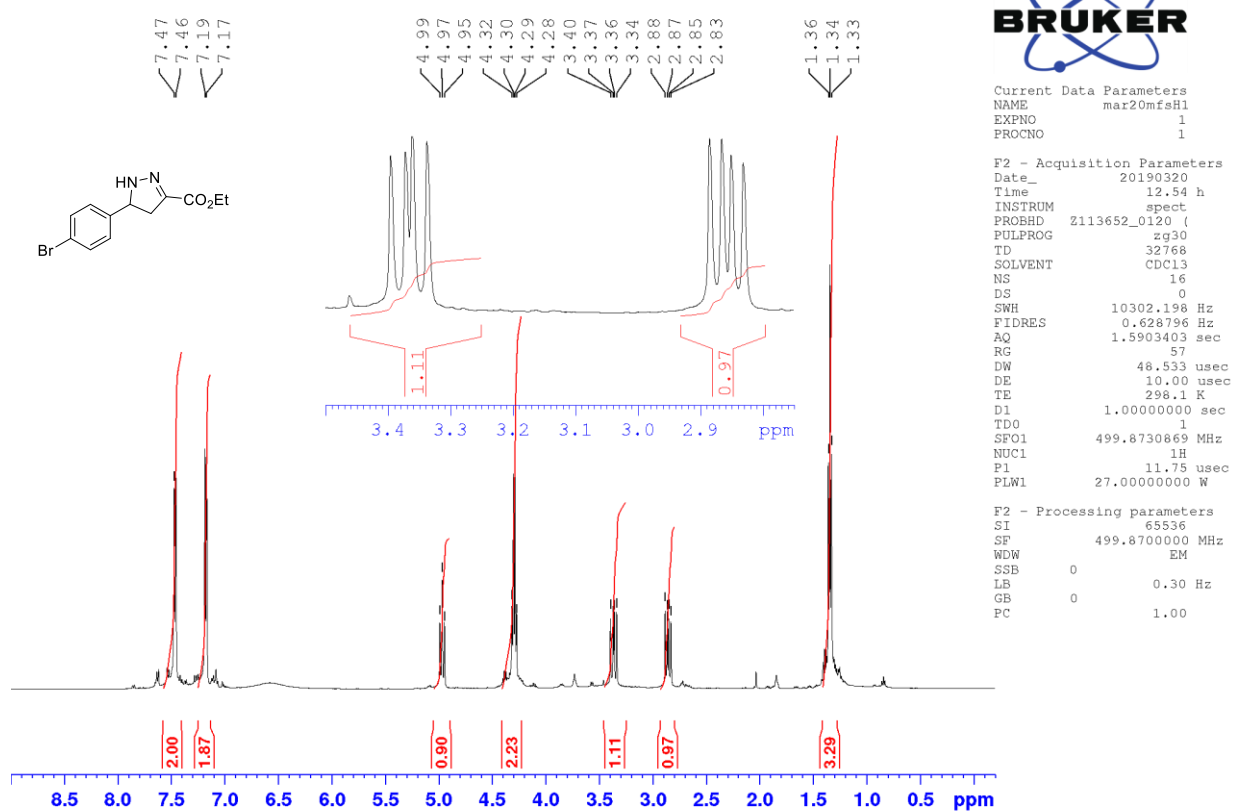
**<sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>, ppm)** δ 8.03-7.97 (m, 2H), 7.72-7.55 (m, 3H), 7.34-7.32 (m, 1H), 6.30 (dd, *J* = 3.1, 2.0 Hz, 1H), 6.18 (d, *J* = 3.3 Hz, 1H), 5.07 (dd, *J* = 11.0, 8.3 Hz, 1H), 3.30 (dd, *J* = 16.7, 11.0 Hz, 1H), 3.16 (dd, *J* = 16.7, 8.3 Hz, 1H)

**<sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>, ppm)** δ 152.1 (C<sub>0</sub>), 150.6 (C<sub>0</sub>), 143.0 (CH), 138.7 (C<sub>0</sub>), 134.1 (CH), 129.4 (CH), 128.4 (CH), 110.5 (CH), 107.3 (CH), 59.5 (CH), 35.6 (CH<sub>2</sub>)

**HRMS** expected: 277.0647 [M+H]<sup>+</sup> found: 277.0663 [M+H]<sup>+</sup>

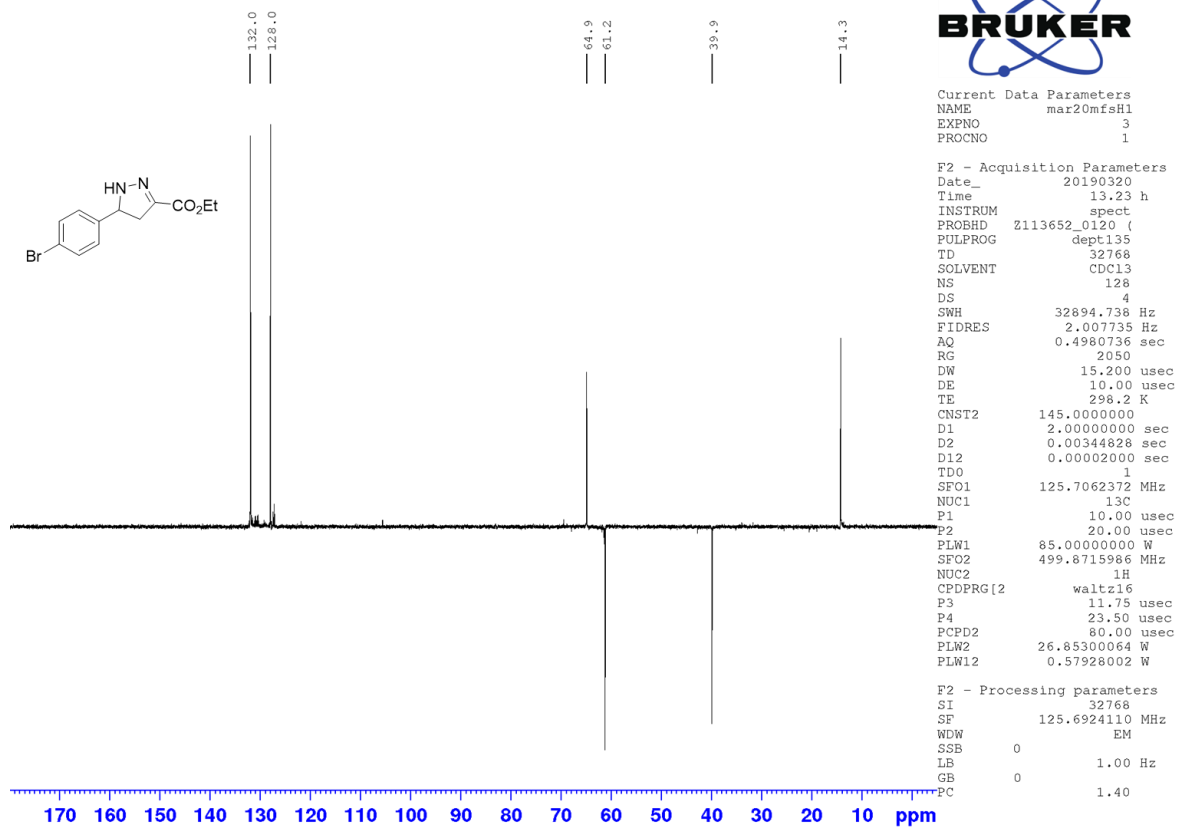
# NMR Spectra for isolated compounds

## Ethyl 5-(4-bromophenyl)-4,5-dihydro-1H-pyrazole-3-carboxylate

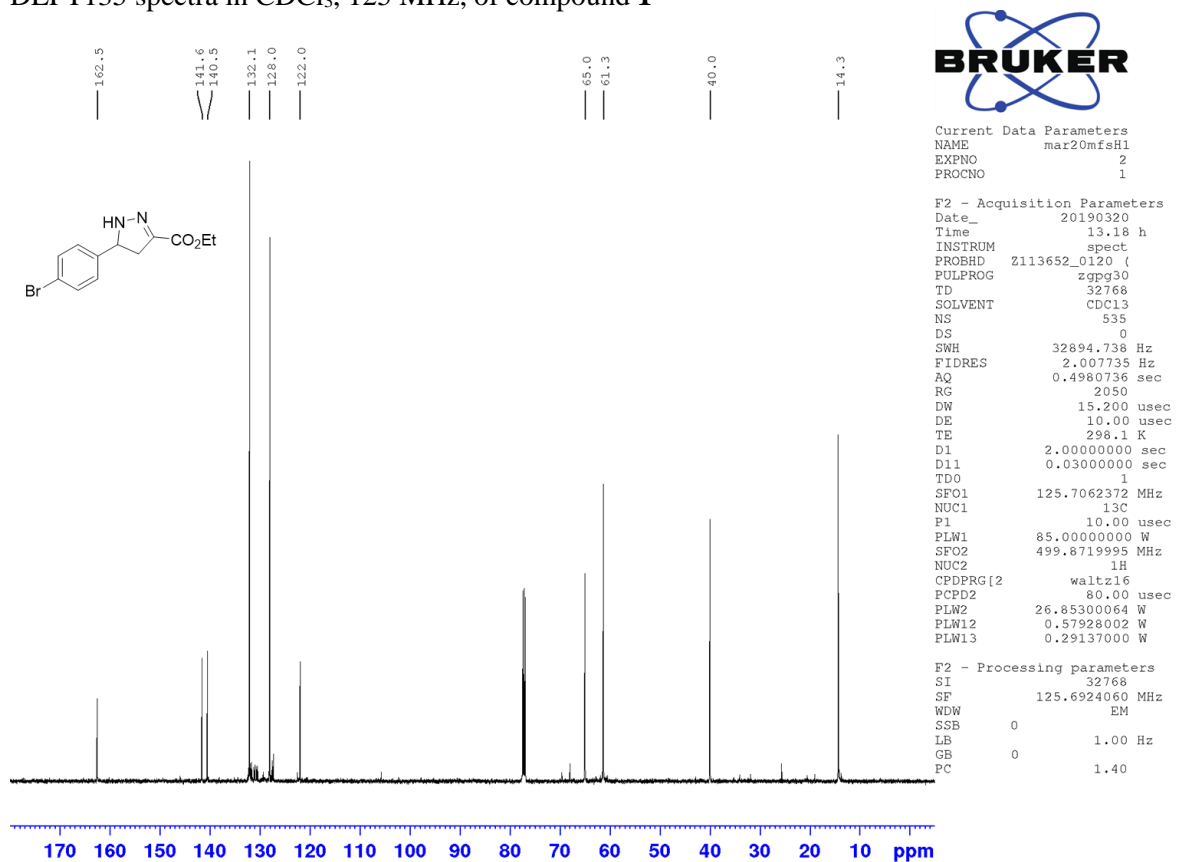


<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 500 MHz, of compound **1**



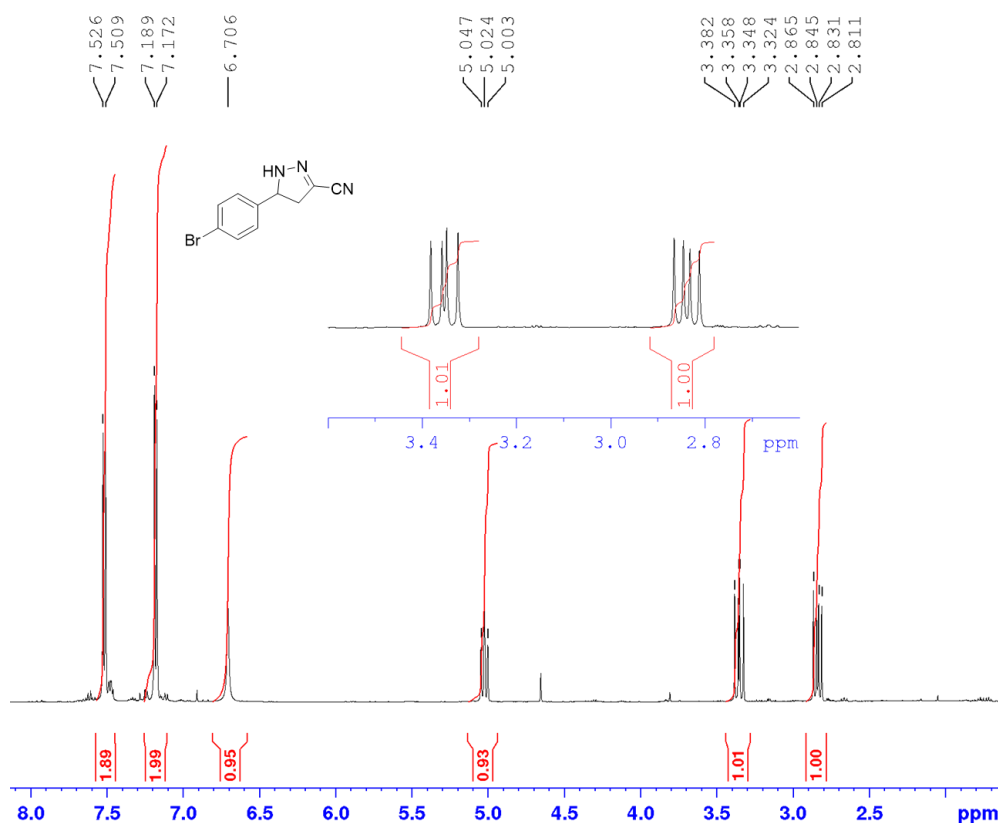


DEPT135 spectra in CDCl<sub>3</sub>, 125 MHz, of compound 1

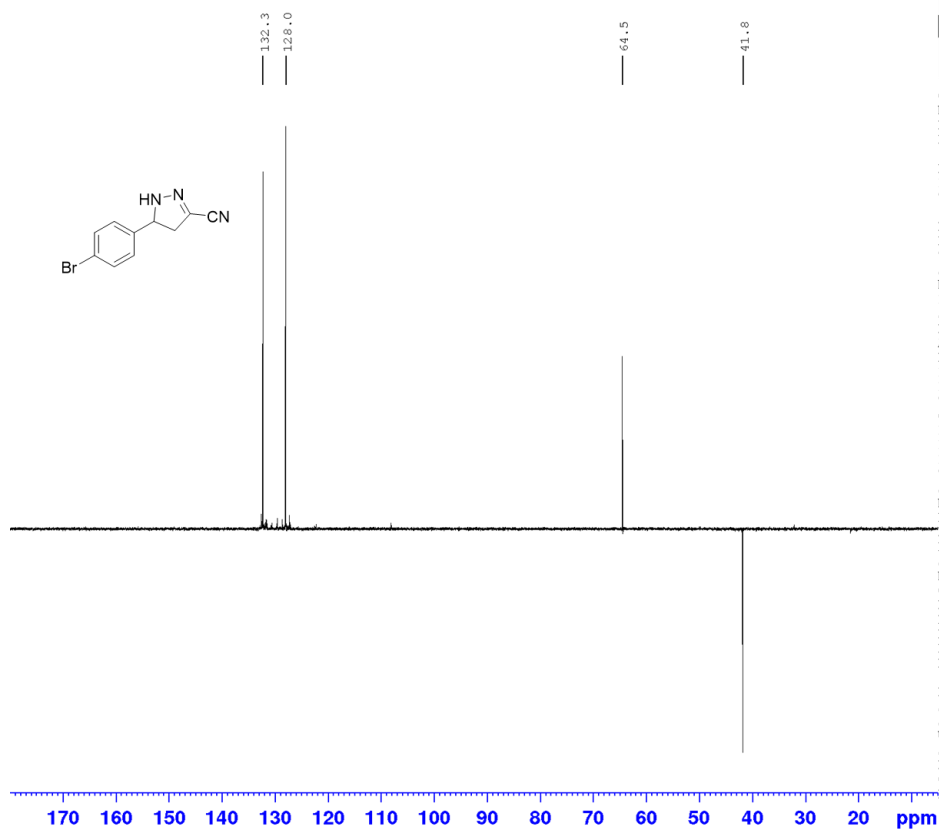


<sup>13</sup>C NMR spectra in CDCl<sub>3</sub>, 125 MHz, of compound 1

# 5-(4-bromophenyl)-4,5-dihydro-1H-pyrazole-3-carbonitrile



<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 500 MHz, of compound 6



DEPT135 NMR spectrum in CDCl<sub>3</sub>, 125 MHz, of compound 6



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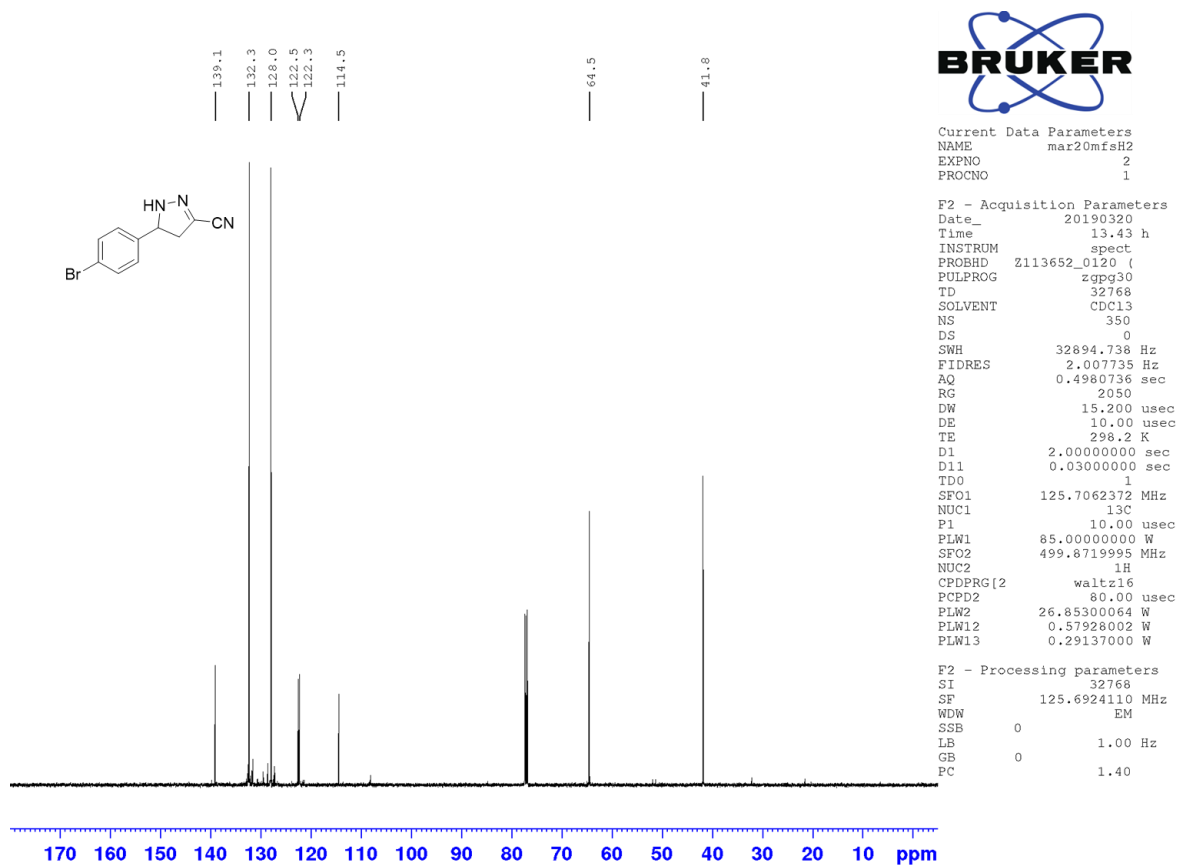
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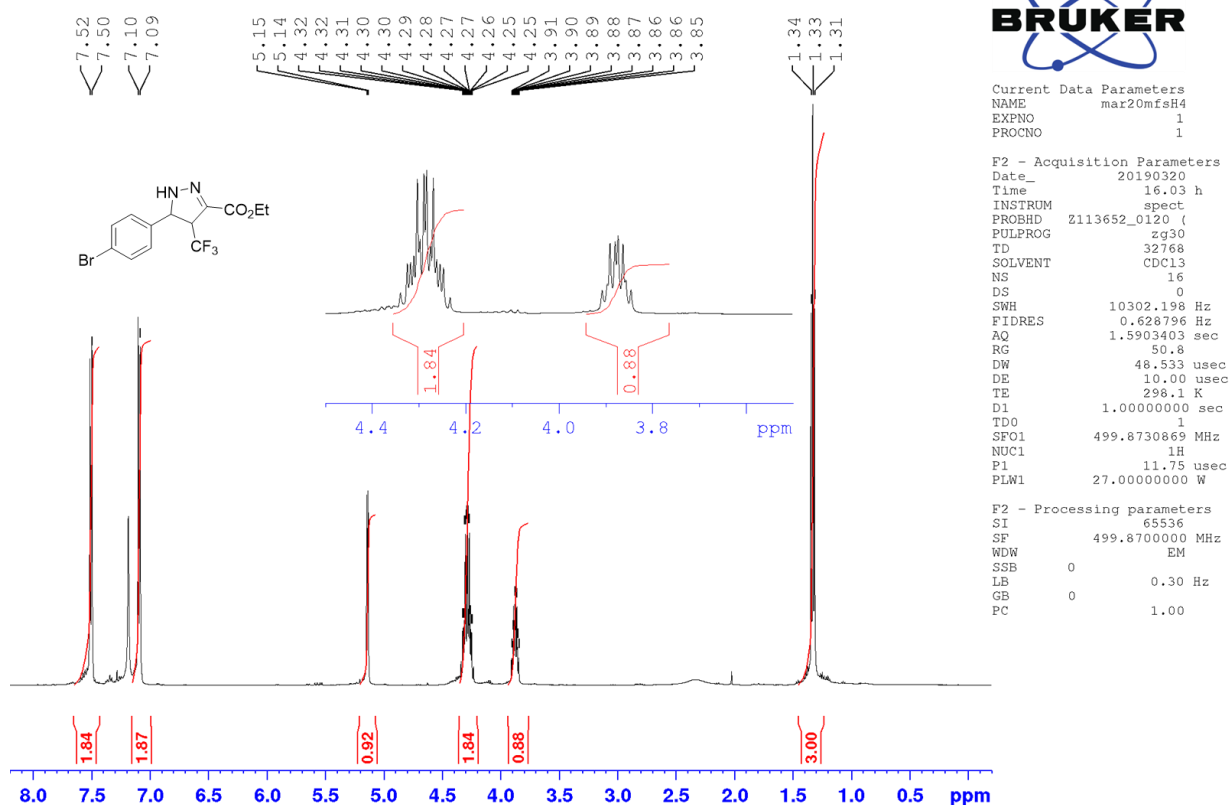
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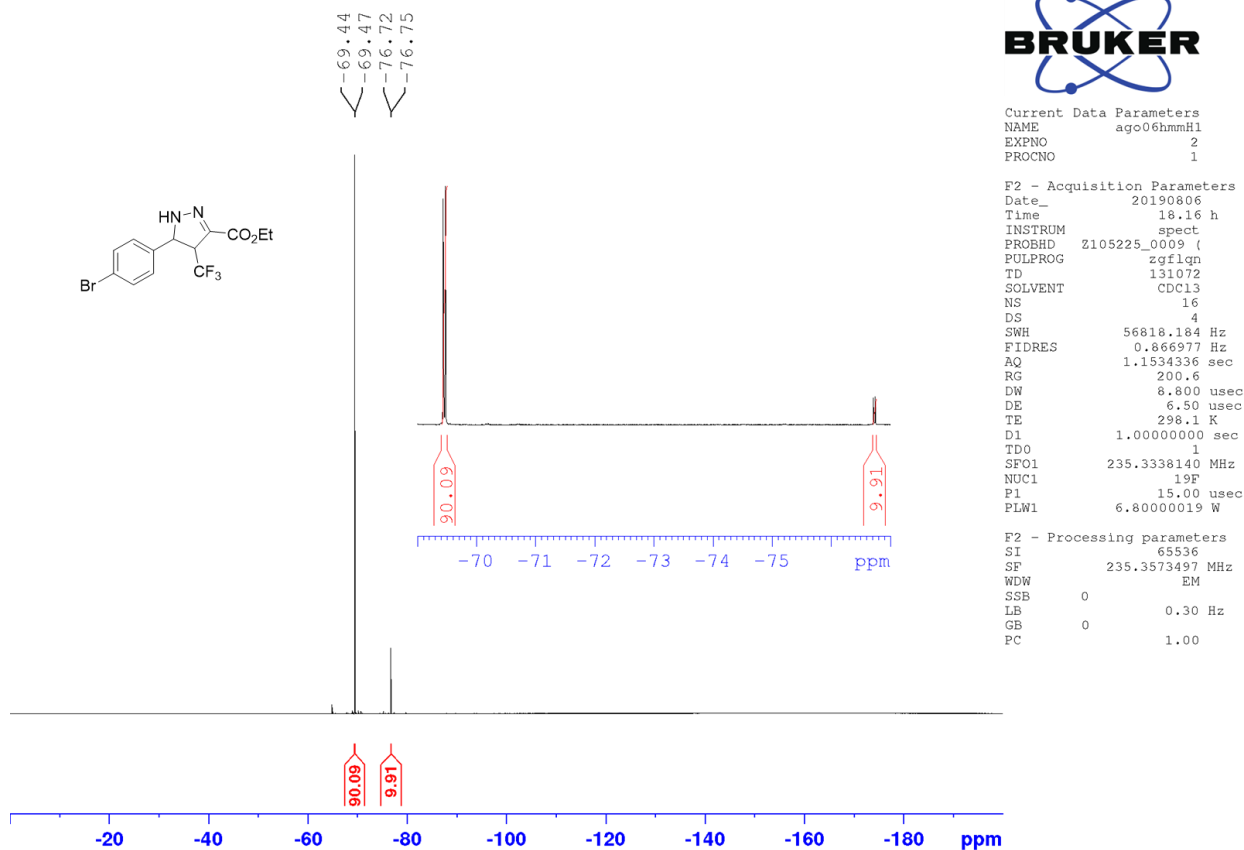


<sup>13</sup>C NMR spectrum in CDCl<sub>3</sub>, 125 MHz, of compound **6**

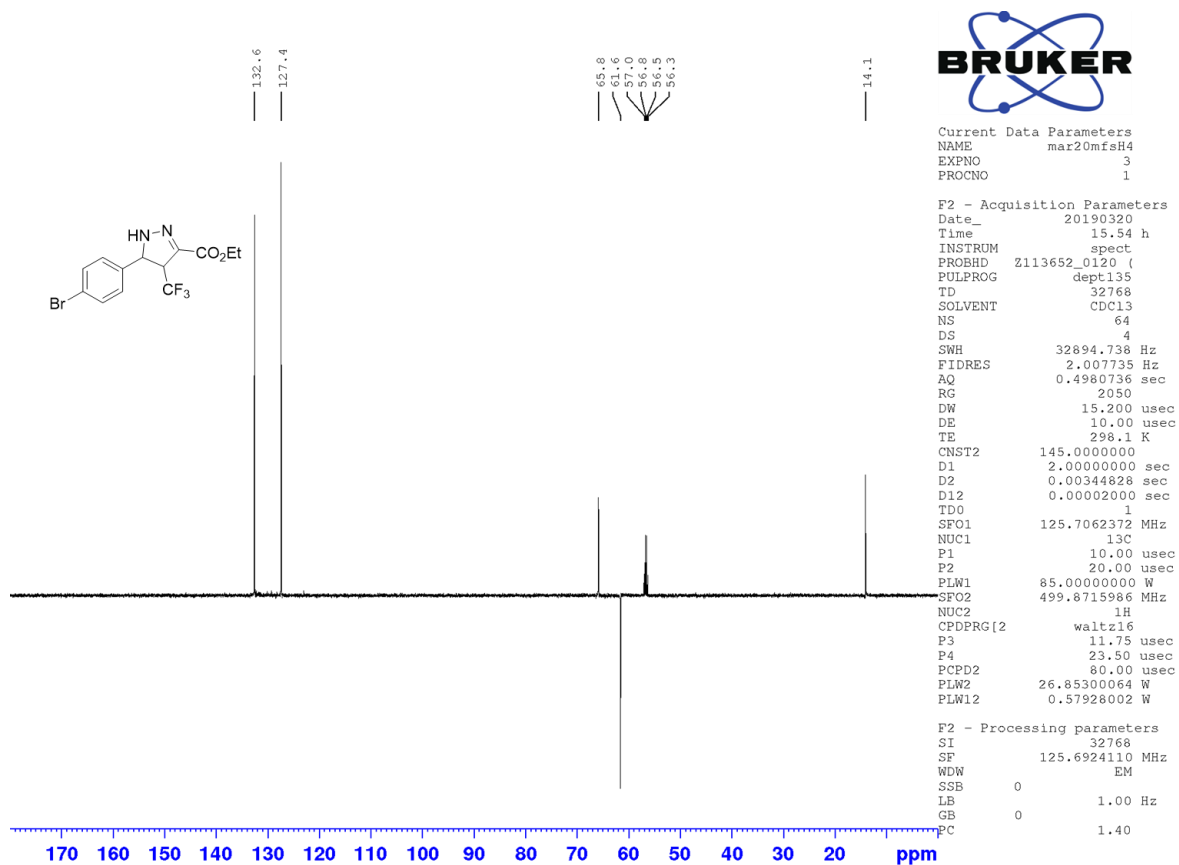
# Ethyl 5-(4-bromophenyl)-4-(trifluoromethyl)-4,5-dihydro-1H-pyrazole-3-carboxylate



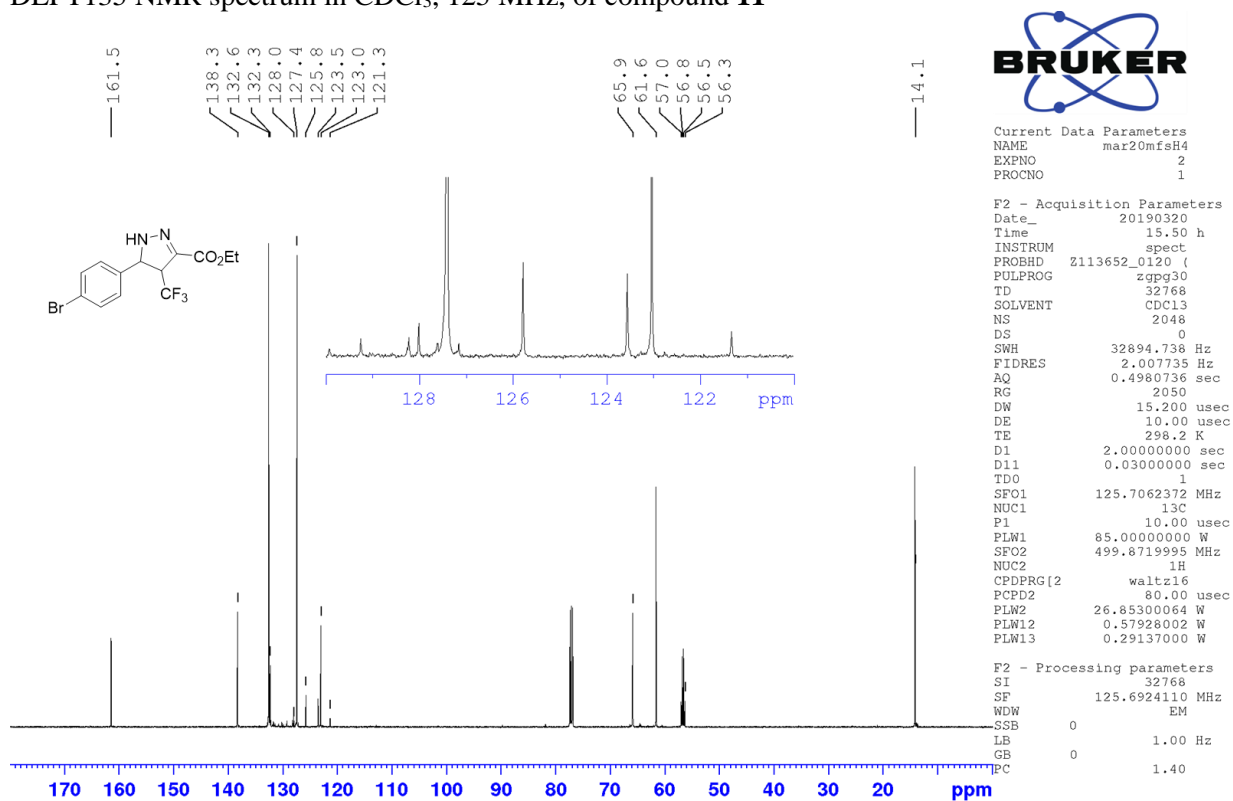
<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 500 MHz, of compound **11**



<sup>19</sup>F NMR spectrum in CDCl<sub>3</sub>, 235 MHz, of compound **11**

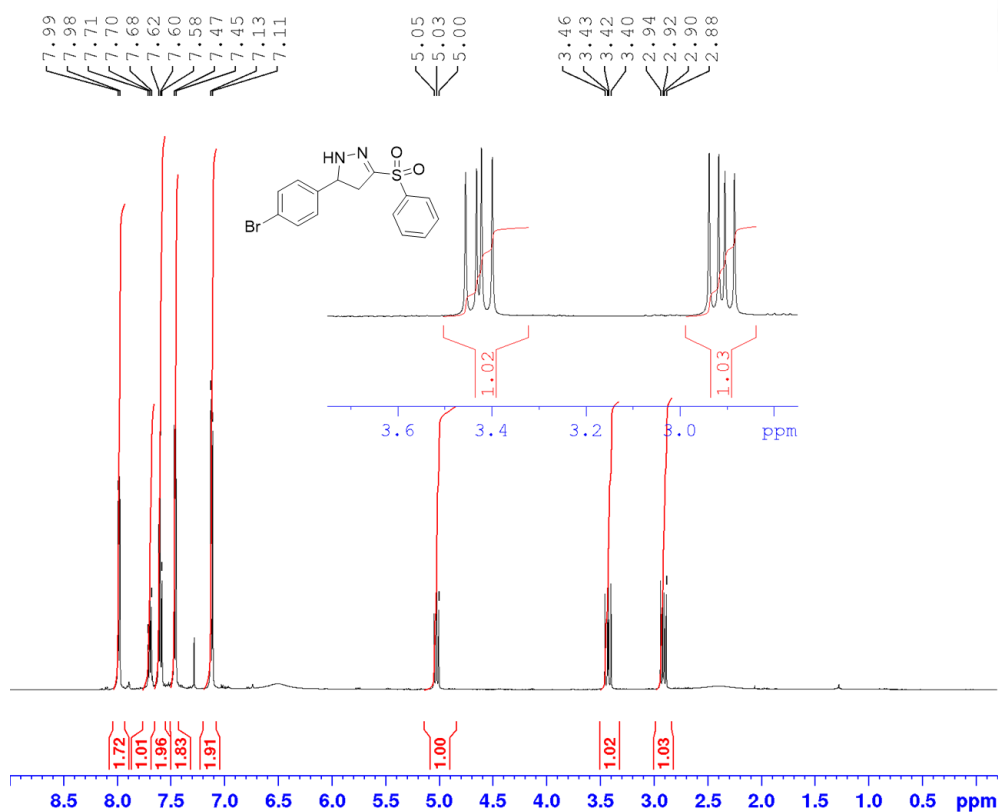


DEPT135 NMR spectrum in CDCl<sub>3</sub>, 125 MHz, of compound **11**

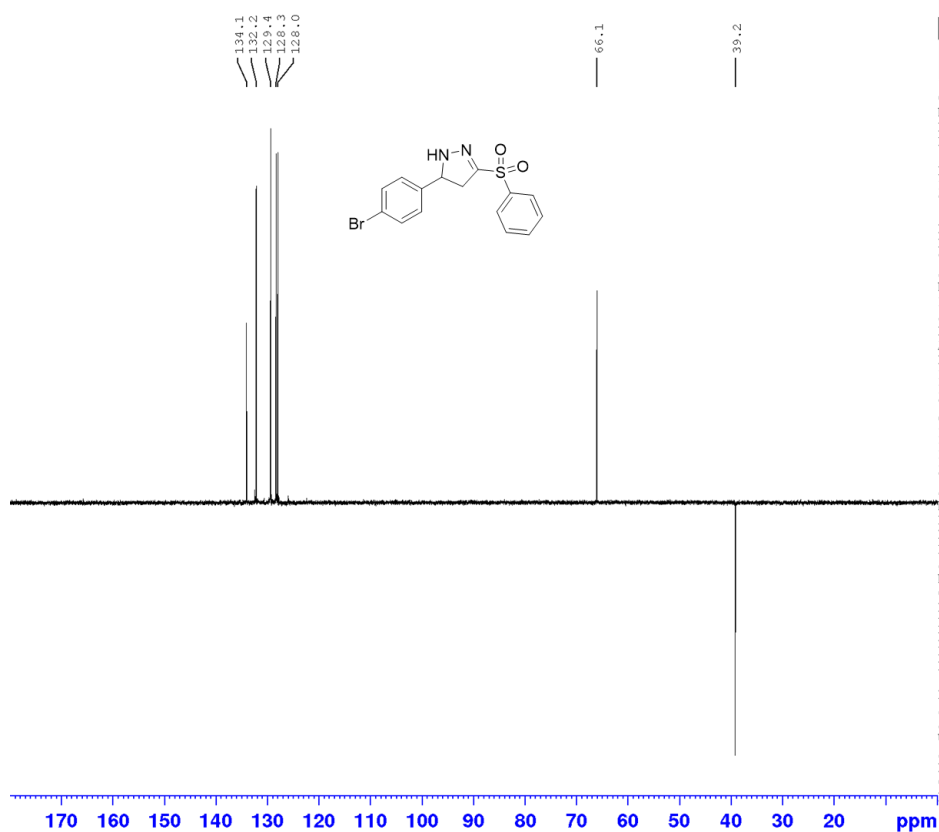


<sup>13</sup>C NMR spectrum in CDCl<sub>3</sub>, 125 MHz, of compound **11**

# 5-(4-bromophenyl)-3-(phenylsulfonyl)-4,5-dihydro-1H-pyrazole



<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 500 MHz, of compound **16**



DEPT135 NMR spectrum in CDCl<sub>3</sub>, 125 MHz, of compound **16**



Current Data Parameters  
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PROCNO 1

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DS 0  
SWH 10302.198 Hz  
FIDRES 0.628796 Hz  
AQ 1.5903403 sec  
RG 128  
DW 48.533 usec  
DE 10.00 usec  
TE 298.2 K  
D1 1.00000000 sec  
TD0 1  
SFO1 499.8730869 MHz  
NUC1 1H  
P1 11.75 usec  
PLW1 27.00000000 W

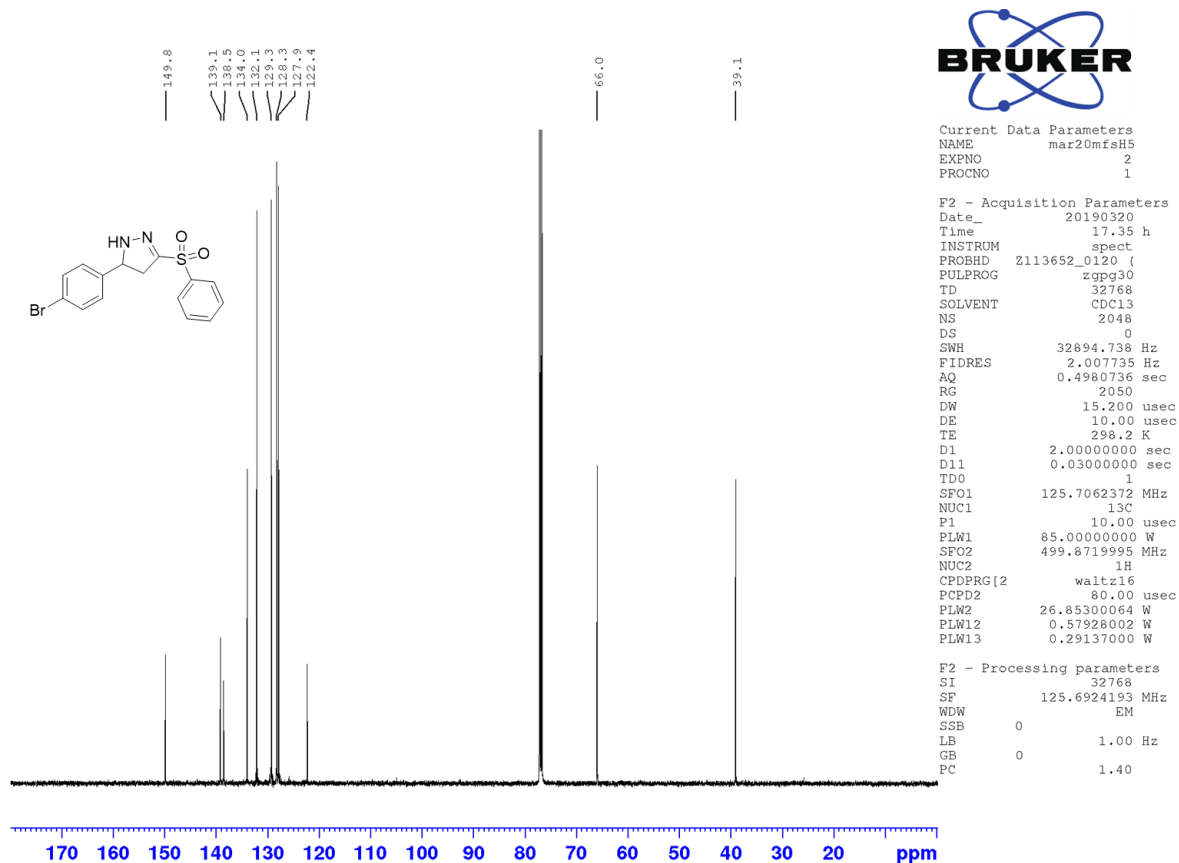
F2 - Processing parameters  
SI 65536  
SF 499.8700000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
NAME mar20mfsH5  
EXPNO 3  
PROCNO 1

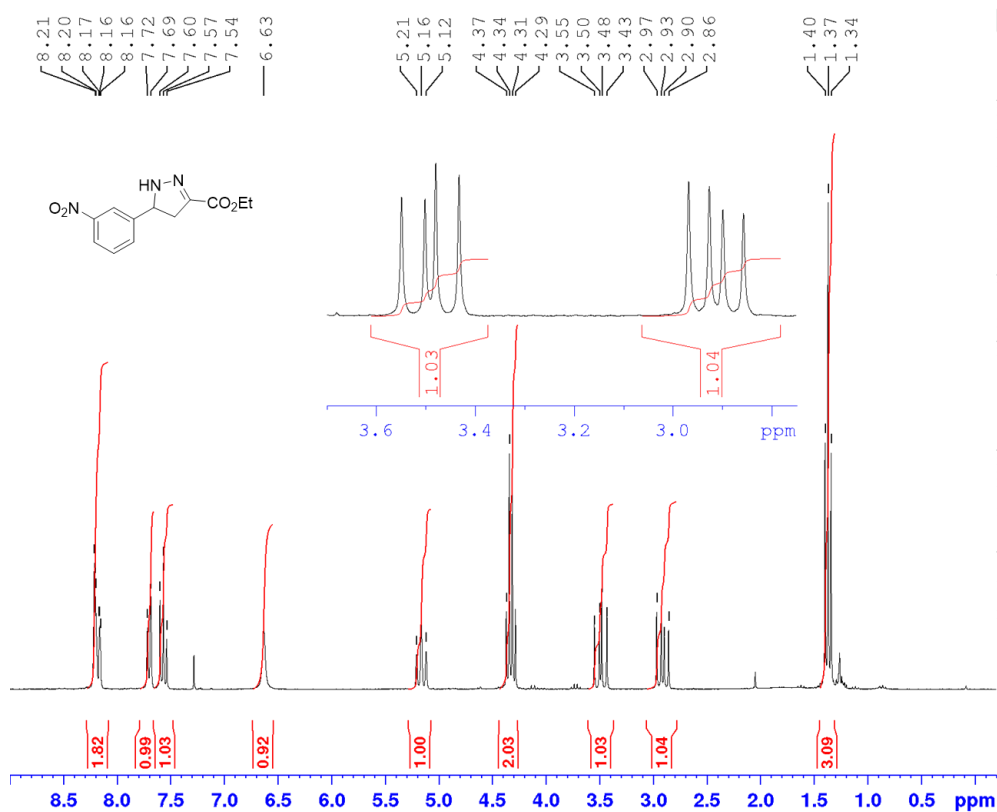
F2 - Acquisition Parameters  
Date\_ 20190320  
Time 17.57 h  
INSTRUM spect  
PROBHD Z113652\_0120 ( )  
PULPROG dept135  
TD 32768  
SOLVENT CDCl3  
NS 512  
DS 4  
SWH 32894.738 Hz  
FIDRES 2.007735 Hz  
AQ 0.4980736 sec  
RG 2050  
DW 15.200 usec  
DE 10.00 usec  
TE 298.2 K  
CNST2 145.0000000  
D1 2.00000000 sec  
D2 0.00344828 sec  
D12 0.00002000 sec  
TD0 1  
SFO1 125.7062372 MHz  
NUC1 13C  
P1 10.00 usec  
P2 20.00 usec  
PLW1 85.00000000 W  
SFO2 499.8715986 MHz  
NUC2 1H  
CPDPRG2 waltz16  
P3 11.75 usec  
P4 23.50 usec  
PCPD2 80.00 usec  
PLW2 26.85300064 W  
PLW12 0.57928002 W

F2 - Processing parameters  
SI 32768  
SF 125.6924110 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

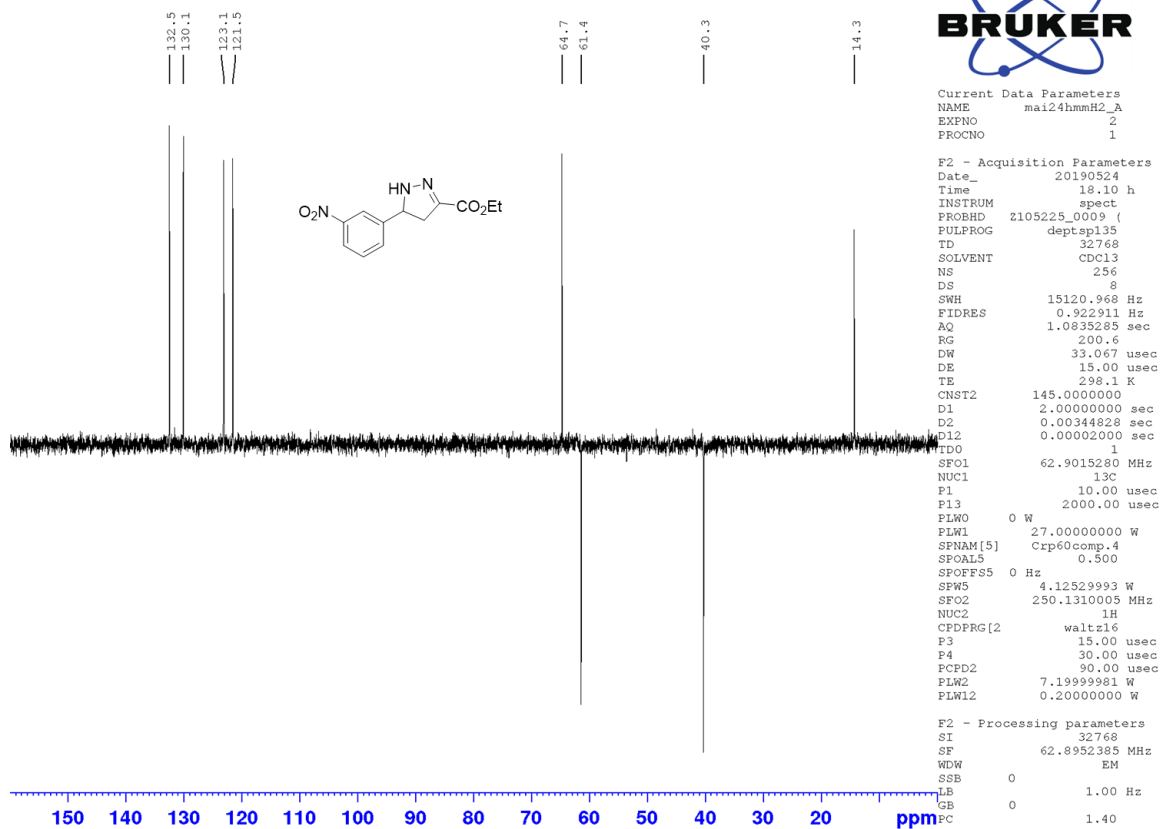


<sup>13</sup>C NMR spectrum in CDCl<sub>3</sub>, 125 MHz, of compound **16**

# Ethyl 5-(3-nitrophenyl)-4,5-dihydro-1H-pyrazole-3-carboxylate



<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound 2



DEPT135 NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound 2



Current Data Parameters  
NAME mai24hmmH2\_A  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190524  
Time 17.41 h  
INSTRUM spect  
PROBHD Z105225\_0009 ( )  
PULPROG zg30  
TD 32768  
SOLVENT CDCl3  
NS 8  
DS 0  
SWH 5000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 200.6  
DE 100.000 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 250.1315446 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 7.19999981 W

F2 - Processing parameters  
SI 55536  
SF 250.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

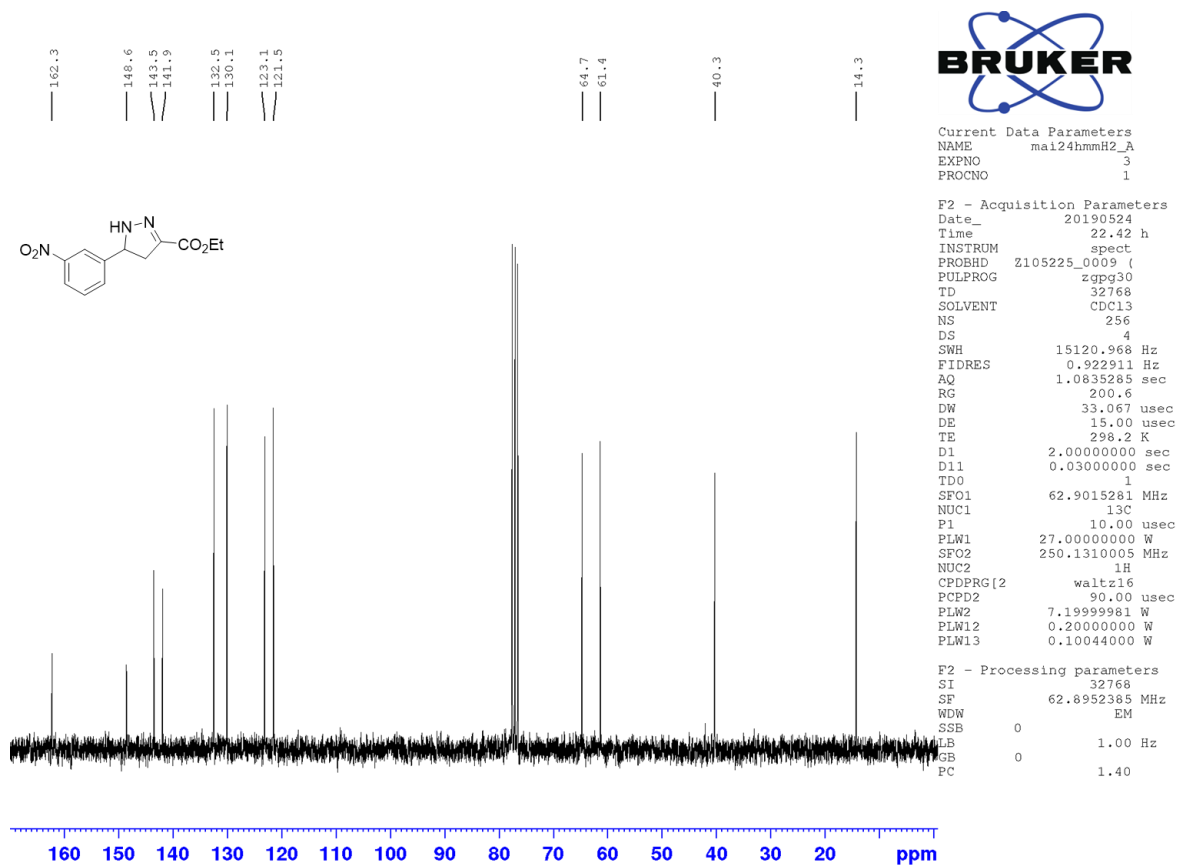


Current Data Parameters  
NAME mai24hmmH2\_A  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190524  
Time 18.10 h  
INSTRUM spect  
PROBHD Z105225\_0009 ( )  
PULPROG deptsp135  
TD 32768  
SOLVENT CDCl3  
NS 256  
DS 8  
SWH 15120.968 Hz  
FIDRES 0.922911 Hz  
AQ 1.0835285 sec  
RG 200.6  
DE 33.067 usec  
TE 298.1 K  
CNST2 145.0000000  
D1 2.00000000 sec  
D2 0.00344828 sec  
D12 0.00002000 sec  
TD0 1  
SFO1 62.9015280 MHz  
NUC1 13C  
P1 10.00 usec  
P13 2000.00 usec  
PLW0 0 W  
PLW1 27.00000000 W  
SPNAM[5] Crp60comp.4  
SFOAL5 0.500  
SPOFFS5 0 Hz  
SPW5 4.12529993 W  
SFO2 250.1310005 MHz  
NUC2 1H  
CFDPFRG2 waltz16  
P3 15.00 usec  
P4 30.00 usec  
PCPD2 90.00 usec  
PLW2 7.19999981 W  
PLW12 0.20000000 W

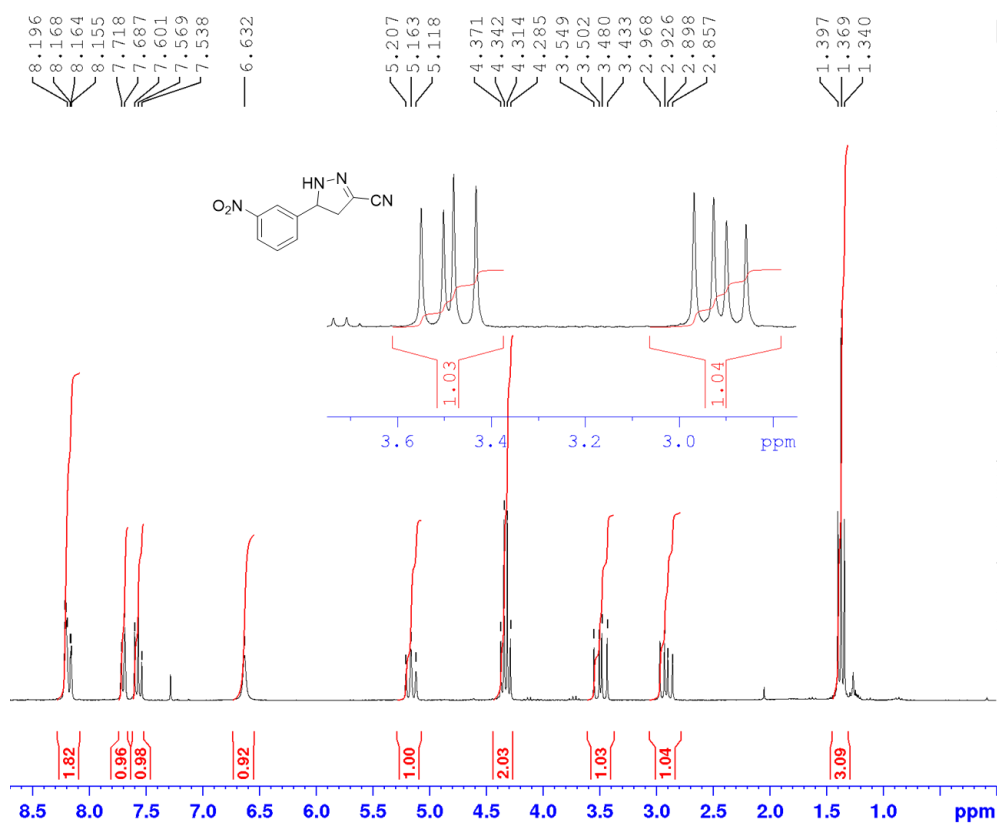
F2 - Processing parameters  
SI 32768  
SF 62.8952385 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40





$^{13}\text{C}$  NMR spectrum in  $\text{CDCl}_3$ , 63 MHz, of compound 2

# 5-(3-nitrophenyl)-4,5-dihydro-1H-pyrazole-3-carbonitrile

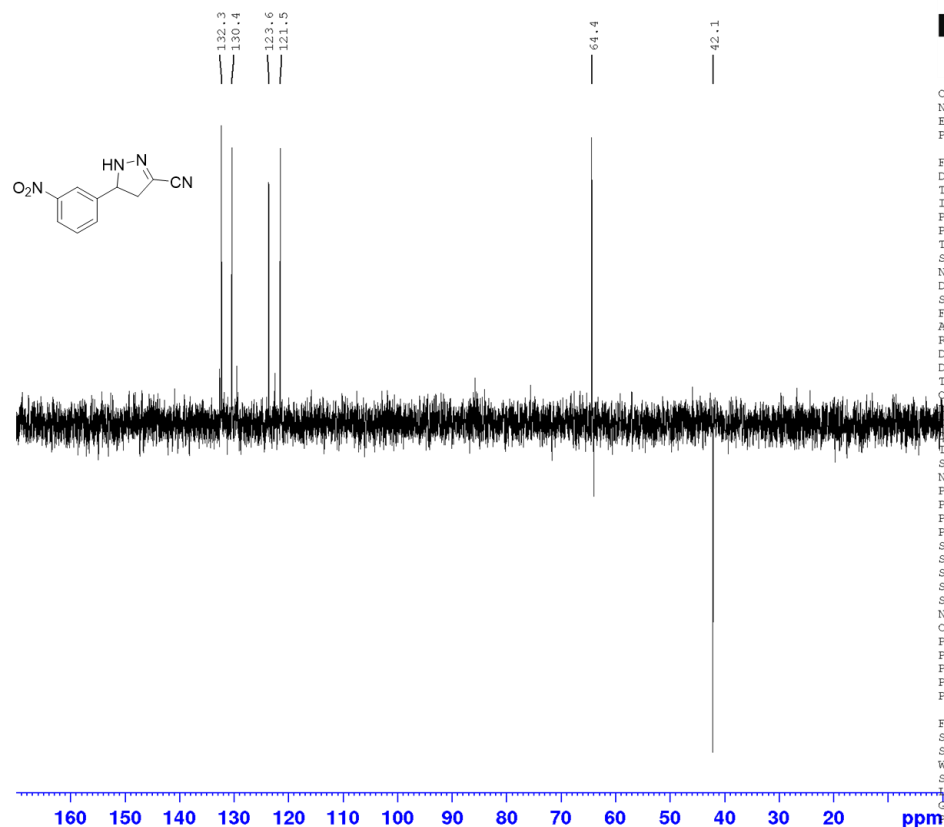


Current Data Parameters  
NAME mai24hnmh2\_A  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190524  
Time 17.41 h  
INSTRUM spect  
PROBHD Z105225\_0009 ( )  
PULPROG zg30  
TD 32768  
SOLVENT CDCl3  
NS 8  
DS 0  
SWH 5000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.276799 sec  
RG 200.6  
DW 100.000 usec  
DE 6.50 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 250.1315446 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 7.19999981 W

F2 - Processing parameters  
SI 55536  
SF 250.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound 7

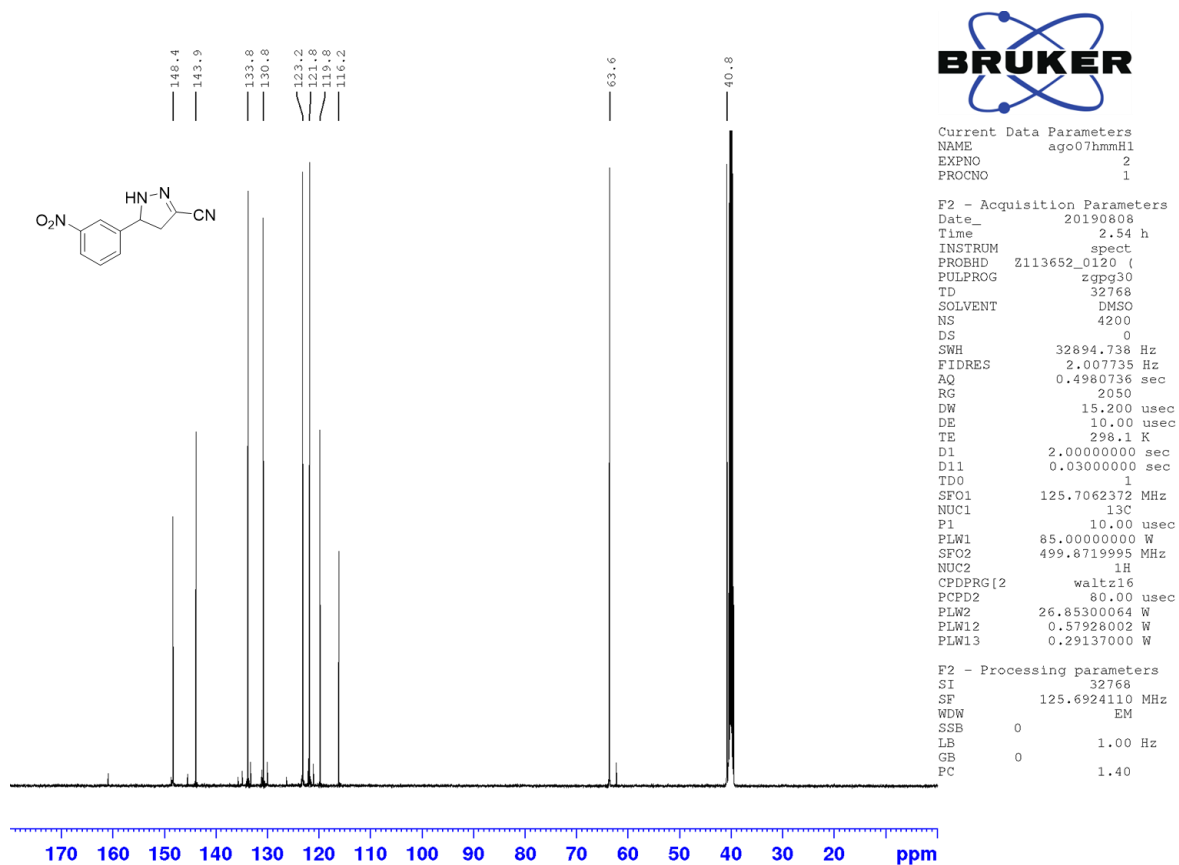


Current Data Parameters  
NAME mai24hnmh3\_B  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190524  
Time 23.31 h  
INSTRUM spect  
PROBHD Z105225\_0009 ( )  
PULPROG deptsp135  
TD 32768  
SOLVENT CDCl3  
NS 256  
DS 8  
SWH 15120.968 Hz  
FIDRES 0.922911 Hz  
AQ 1.0835285 sec  
RG 200.6  
DW 33.067 usec  
DE 15.00 usec  
TE 298.1 K  
CNST2 145.0000000  
1 2.00000000 sec  
2 0.00344828 sec  
12 0.00002000 sec  
TD0 1  
SFO1 62.9015280 MHz  
NUC1 13C  
P1 10.00 usec  
P13 2000.00 usec  
PLW0 0 W  
PLW1 27.00000000 W  
SPNAM[5] Crp60comp.4  
SFOAL5 0.500  
SPOFFS5 0 Hz  
SPW5 4.12529993 W  
SFO2 250.1310005 MHz  
NUC2 1H  
CPDPRG2 waltz16  
P3 15.00 usec  
P4 30.00 usec  
PCPD2 90.00 usec  
PLW2 7.19999981 W  
PLW12 0.20000000 W

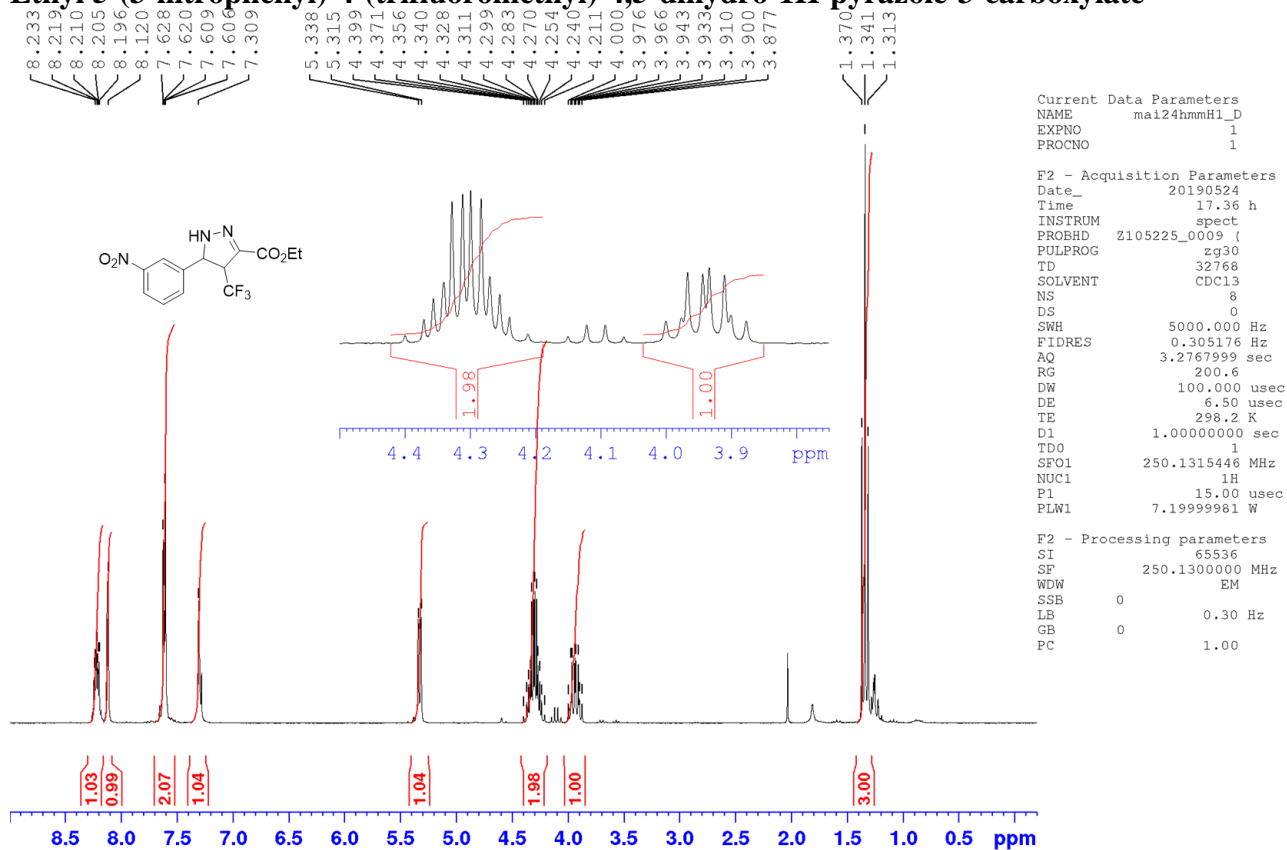
F2 - Processing parameters  
SI 32768  
SF 62.8952385 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

DEPT135 NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound 7

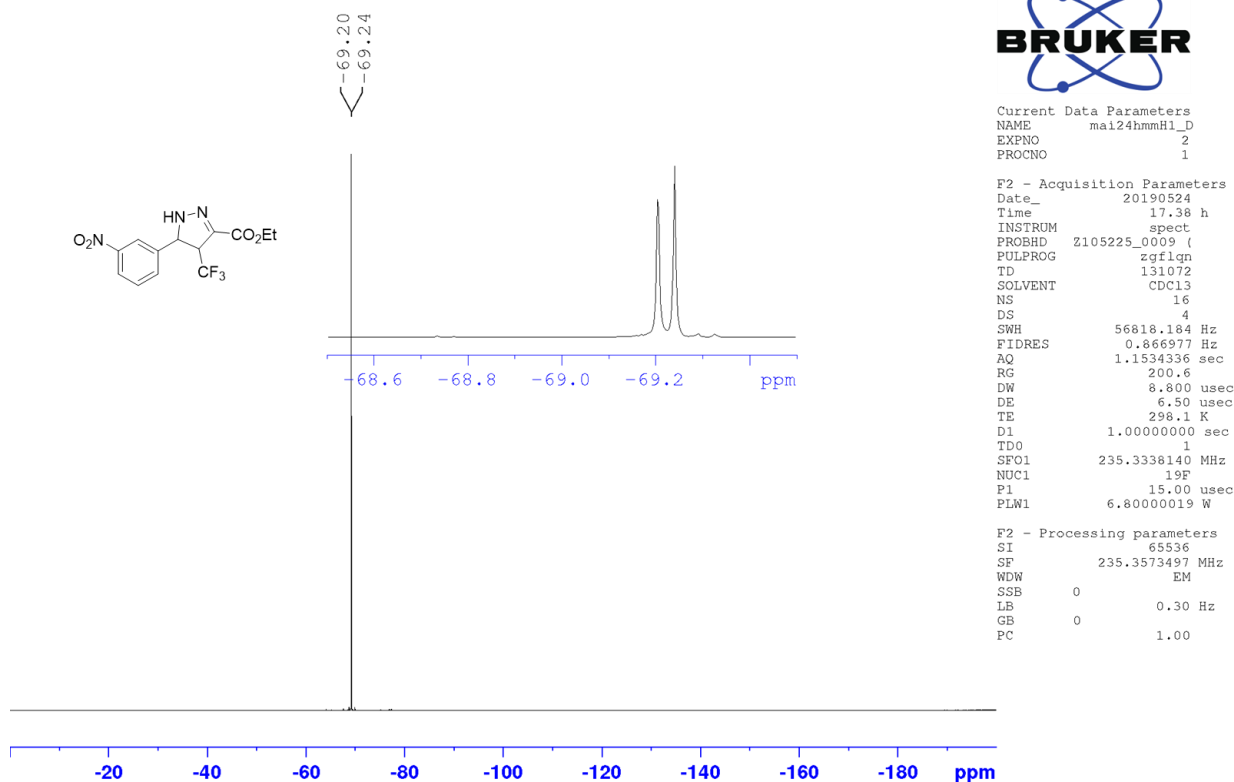


<sup>13</sup>C NMR spectrum in DMSO-d<sub>6</sub>, 125 MHz, of compound **7**

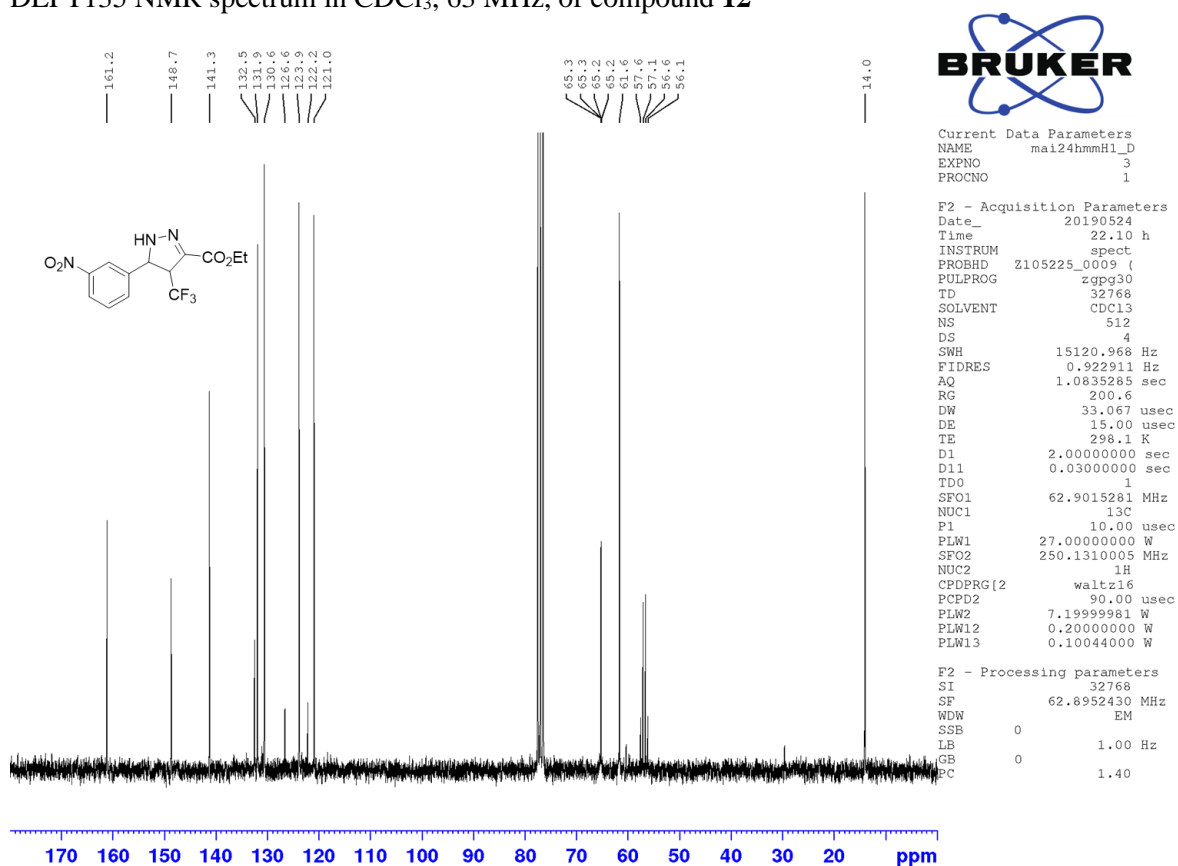
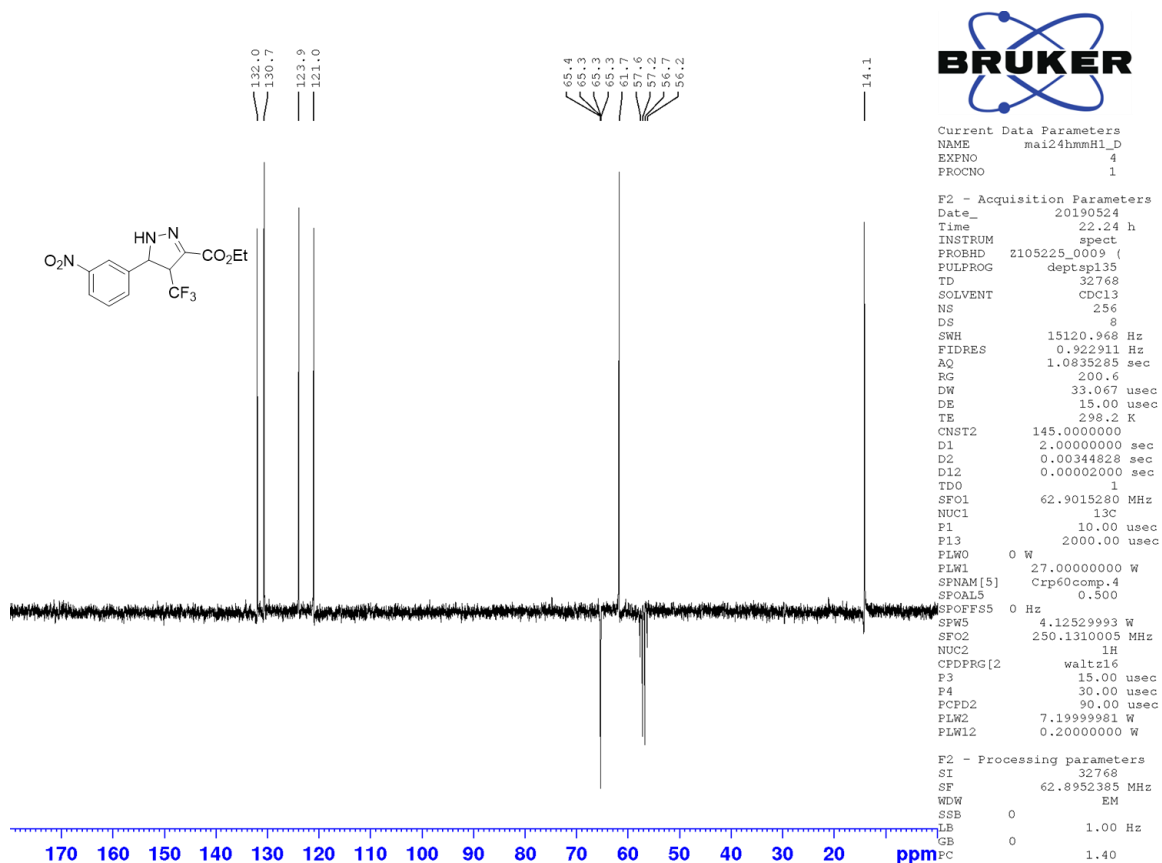
# **Ethyl 5-(3-nitrophenyl)-4-(trifluoromethyl)-4,5-dihydro-1H-pyrazole-3-carboxylate**



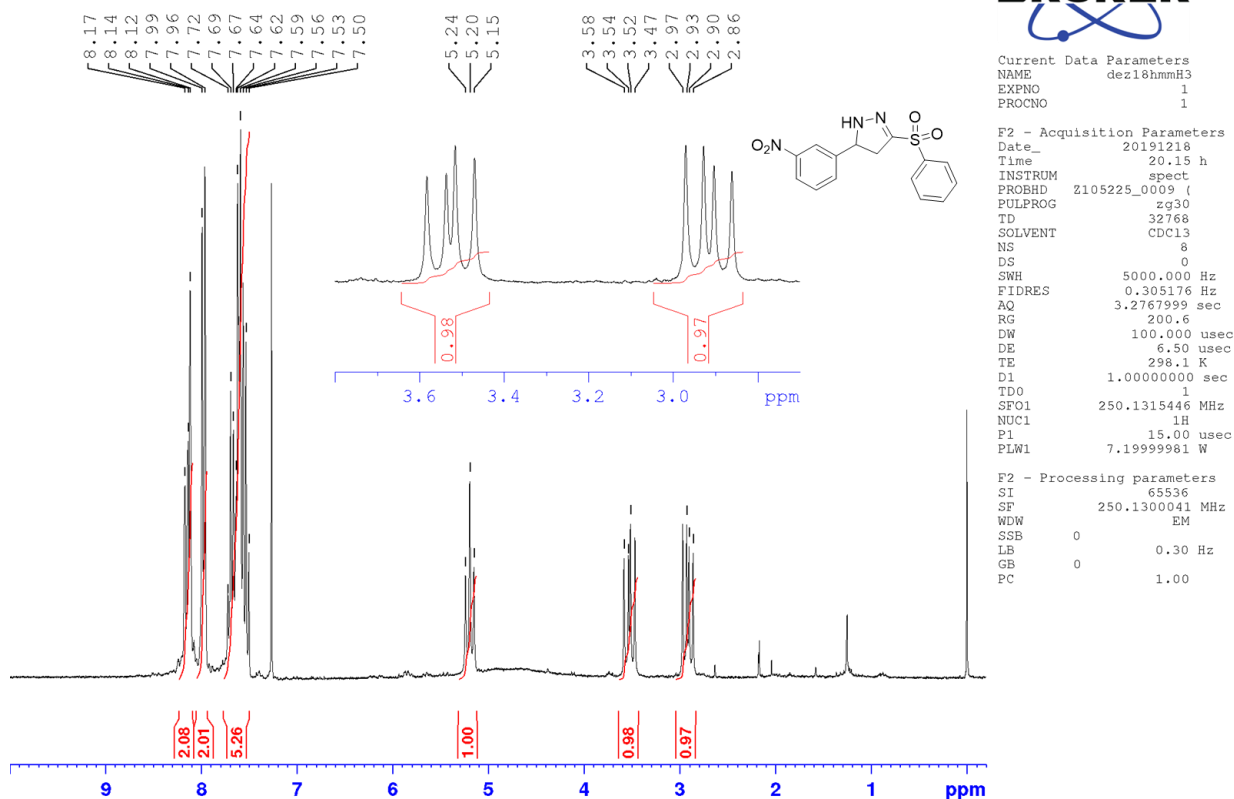
<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound **12**



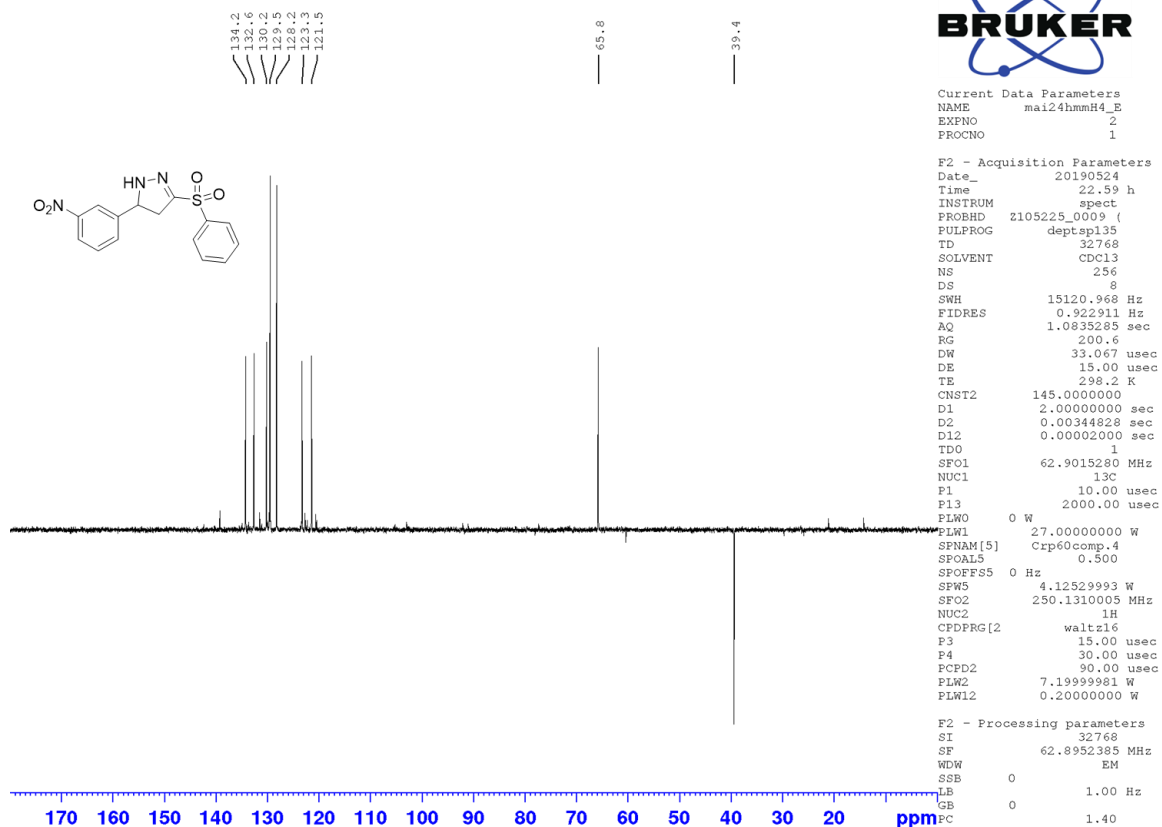
<sup>19</sup>F NMR spectrum in CDCl<sub>3</sub>, 235 MHz, of compound **12**



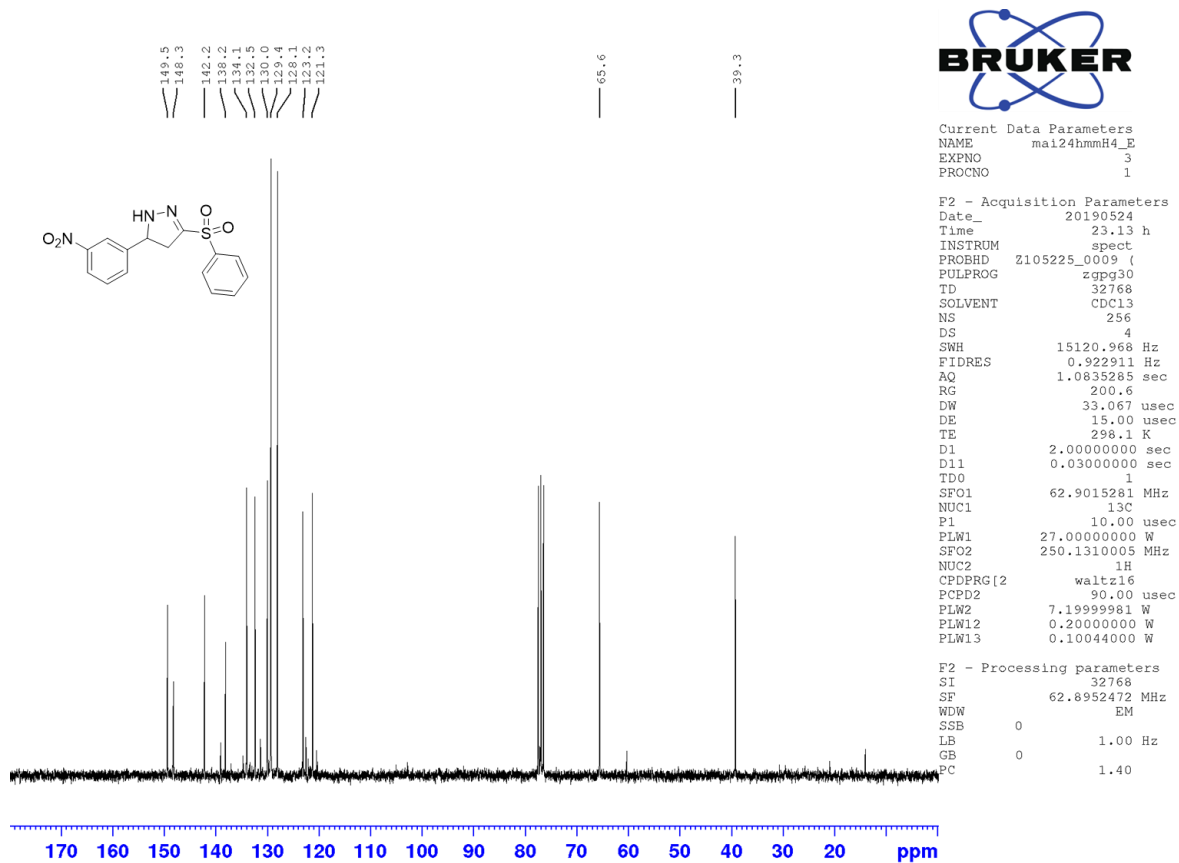
# 5-(3-nitrophenyl)-3-(phenylsulfonyl)-4,5-dihydro-1H-pyrazole



<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound **17**

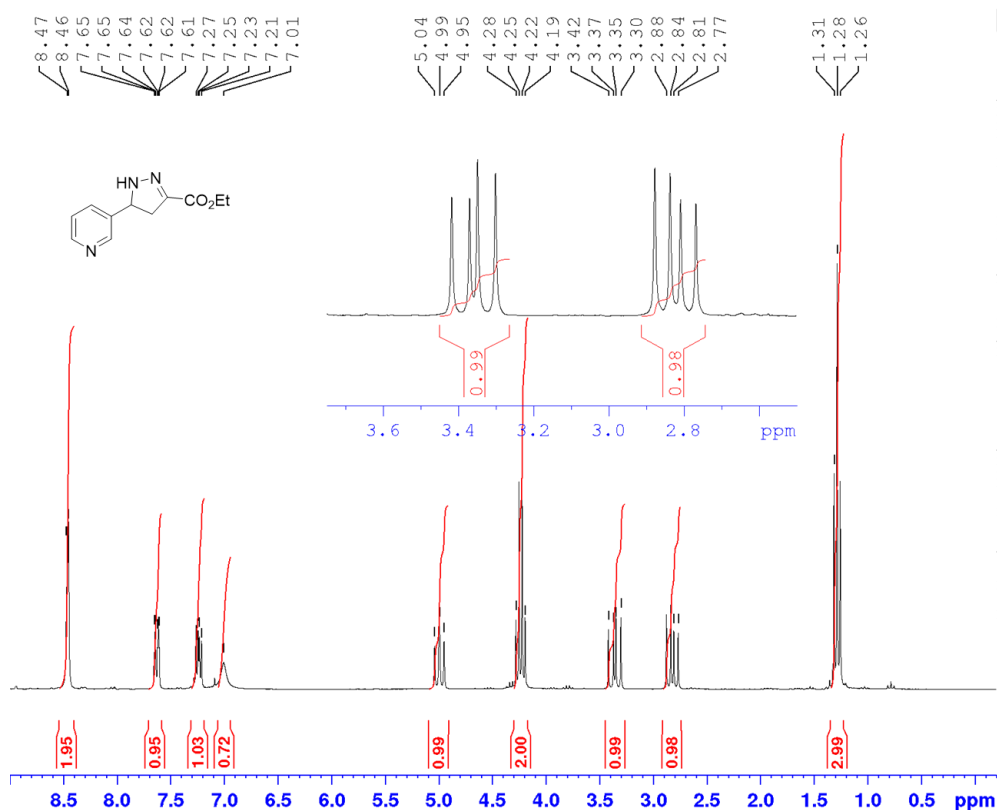


DEPT135 NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **17**



<sup>13</sup>C NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **17**

# Ethyl 5-(pyridin-3-yl)-4,5-dihydro-1H-pyrazole-3-carboxylate

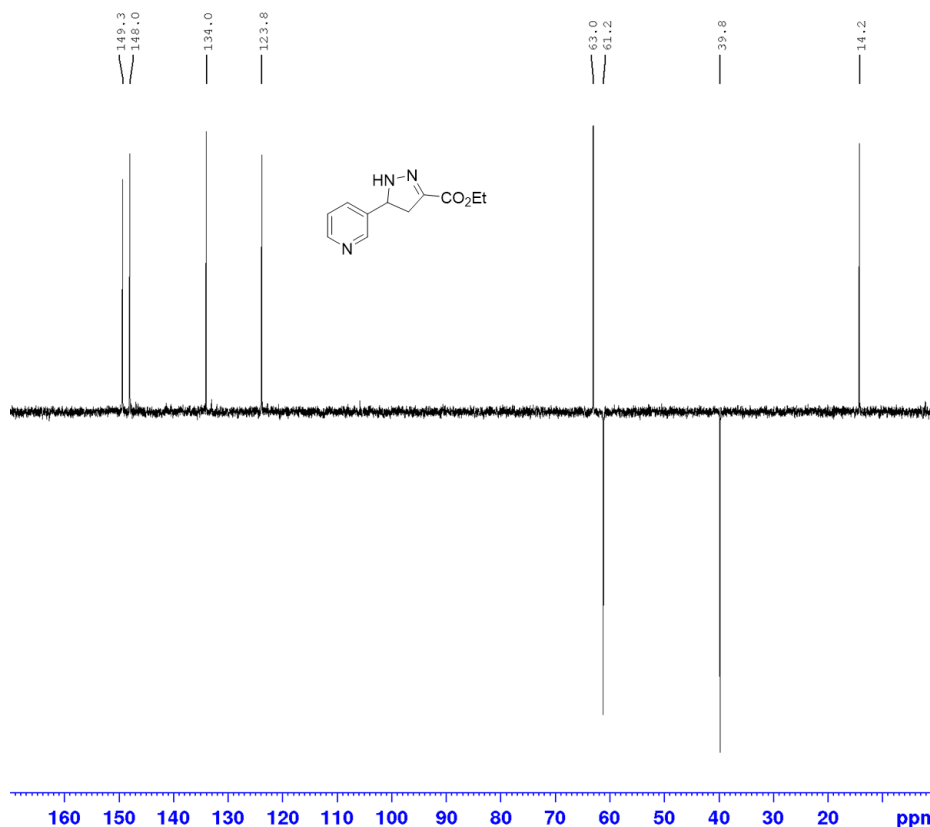


Current Data Parameters  
NAME mai21hmmH1  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190521  
Time 16.14 h  
INSTRUM spect  
PROBHD Z105225\_0009 (   
PULPROG zg30  
TD 32768  
SOLVENT CDCl3  
NS 8  
DS 0  
SWH 5000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 147.17  
DW 100.000 usec  
DE 6.50 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 250.1315446 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 7.19999981 W

F2 - Processing parameters  
SI 55536  
SF 250.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound **3**



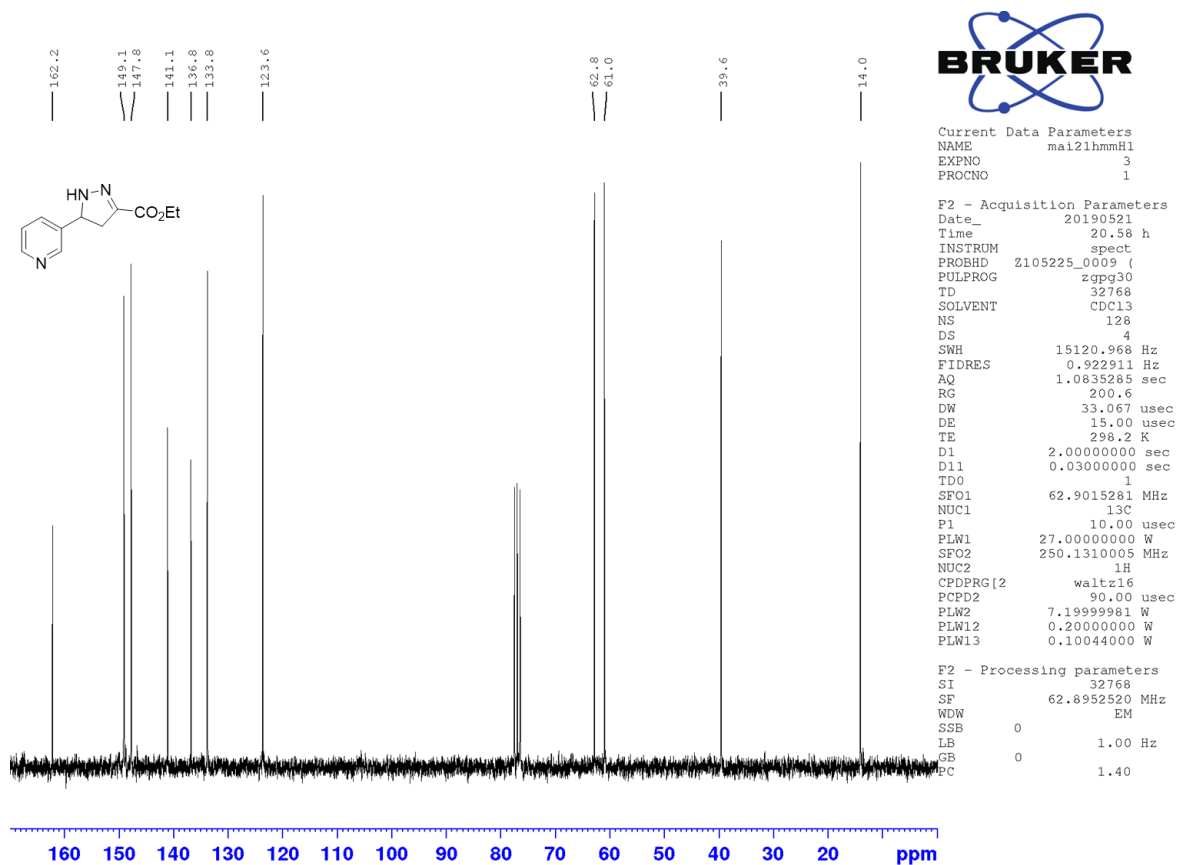
Current Data Parameters  
NAME mai21hmmH1  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190521  
Time 18.47 h  
INSTRUM spect  
PROBHD Z105225\_0009 (   
PULPROG deptsp135  
TD 32768  
SOLVENT CDCl3  
NS 55  
DS 8  
SWH 15120.968 Hz  
FIDRES 0.922911 Hz  
AQ 1.0835285 sec  
RG 200.6  
DW 33.067 usec  
DE 15.00 usec  
TE 298.1 K  
CNST2 145.0000000  
D1 2.00000000 sec  
D2 0.00344828 sec  
D12 0.00002000 sec  
TD0 1  
SFO1 62.9015280 MHz  
NUC1 13C  
P1 10.00 usec  
P13 2000.00 usec  
PLW0 0 W  
PLW1 27.00000000 W  
SPNAM[5] Crp60comp.4  
SFOAL5 0.500  
SPOFFS5 0 Hz  
SPW5 4.12529993 W  
SFO2 250.1310005 MHz  
NUC2 1H  
CFDPRG[2] waltz16  
P3 15.00 usec  
P4 30.00 usec  
PCPD2 90.00 usec  
PLW2 7.19999981 W  
PLW12 0.20000000 W

F2 - Processing parameters  
SI 32768  
SF 62.8952385 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

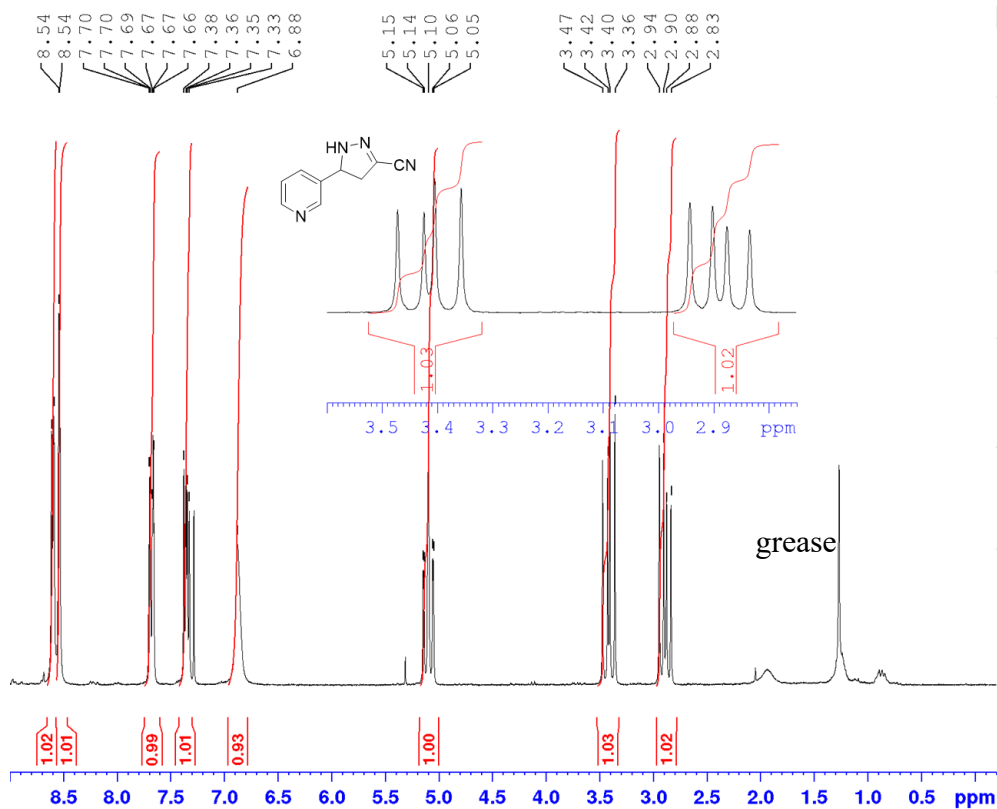
DEPT135 NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **3**





<sup>13</sup>C NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **3**

# 5-(pyridin-3-yl)-4,5-dihydro-1H-pyrazole-3-carbonitrile

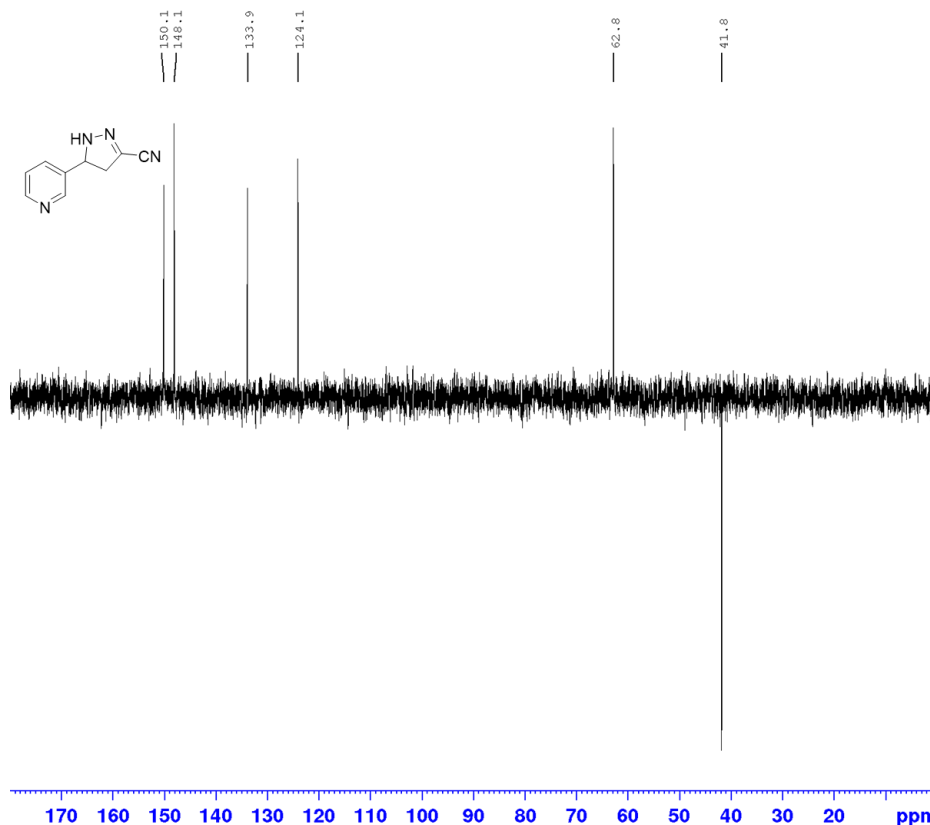


Current Data Parameters  
NAME mai21hmmH2  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190521  
Time 16.18 h  
INSTRUM spect  
PROBHD Z105225\_0009 (Z105225\_0009)  
PULPROG zg30  
TD 32768  
SOLVENT CDCl3  
NS 8  
DS 0  
SWH 5000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 200.6  
DE 100.000 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 250.1315446 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 7.19999981 W

F2 - Processing parameters  
SI 55536  
SF 250.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound **8**

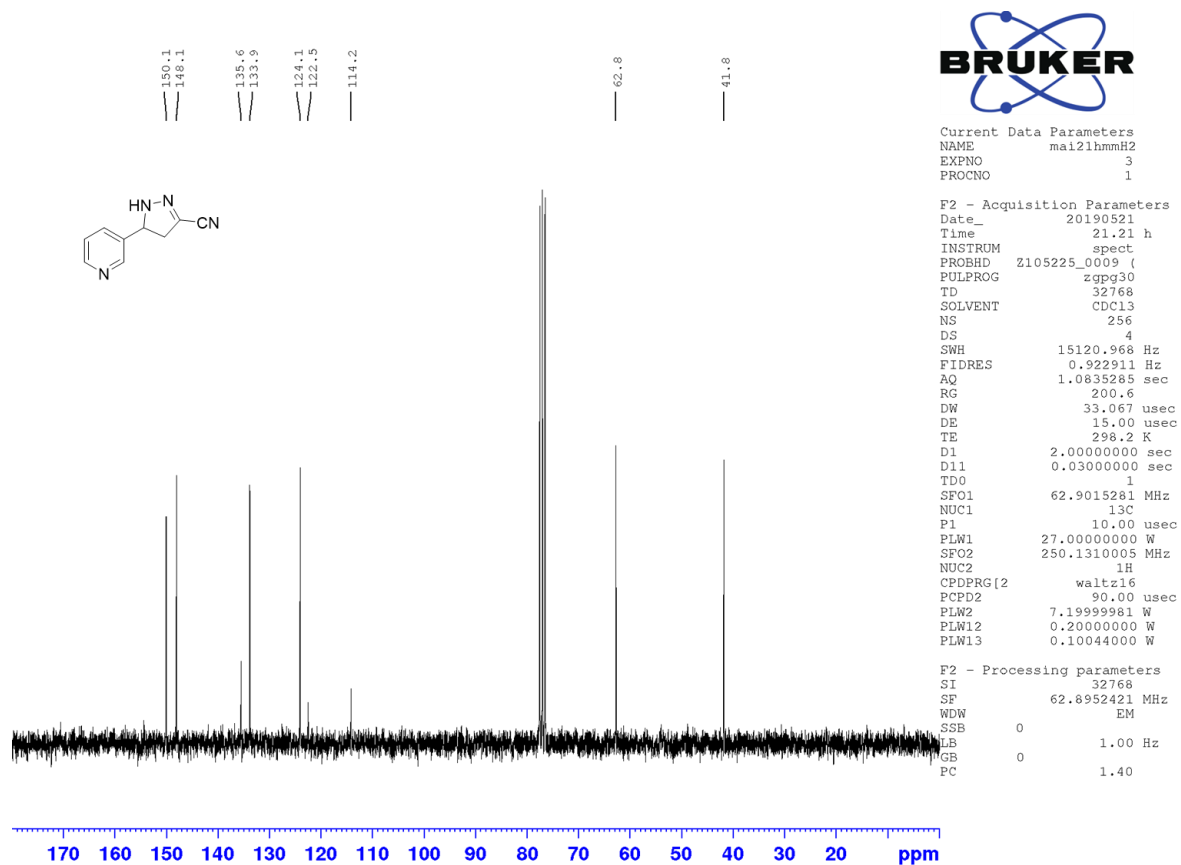


Current Data Parameters  
NAME mai21hmmH2  
EXPNO 2  
PROCNO 1

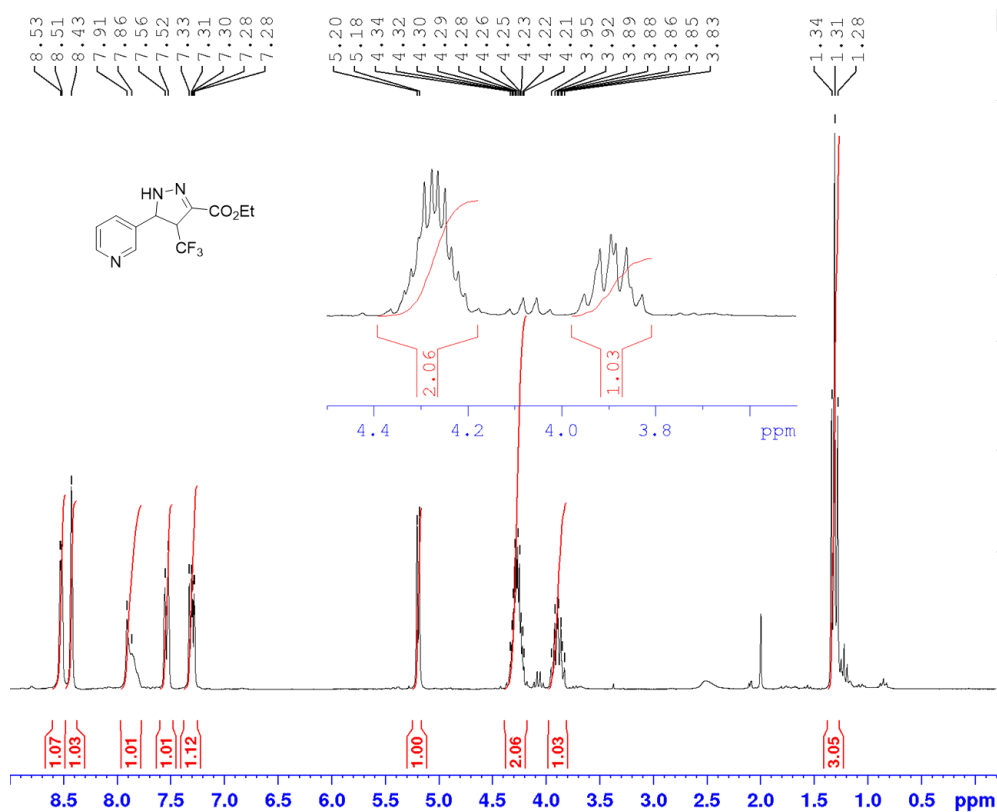
F2 - Acquisition Parameters  
Date\_ 20190521  
Time 18.57 h  
INSTRUM spect  
PROBHD Z105225\_0009 (Z105225\_0009)  
PULPROG deptsp135  
TD 32768  
SOLVENT CDCl3  
NS 64  
DS 8  
SWH 15120.968 Hz  
FIDRES 0.922911 Hz  
AQ 1.0835285 sec  
RG 200.6  
DE 33.067 usec  
TE 298.1 K  
CNST2 145.0000000  
D1 2.00000000 sec  
D2 0.00344828 sec  
D12 0.0002000 sec  
TD0 1  
SFO1 62.9015280 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 2000.00 usec  
PLW0 0 W  
PLW1 27.00000000 W  
SPNAM[5] Crp60comp.4  
SFOAL5 0.500  
SPOFFS5 0 Hz  
SPW5 4.12529993 W  
SFO2 250.1310005 MHz  
NUC2 1H  
CFDPRG[2] waltz16  
P3 15.00 usec  
P4 30.00 usec  
PCPD2 90.00 usec  
PLW2 7.19999981 W  
PLW12 0.20000000 W

F2 - Processing parameters  
SI 32768  
SF 62.8952385 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

DEPT135 NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **8**



# **Ethyl 5-(pyridin-3-yl)-4-(trifluoromethyl)-4,5-dihydro-1H-pyrazole-3-carboxylate**

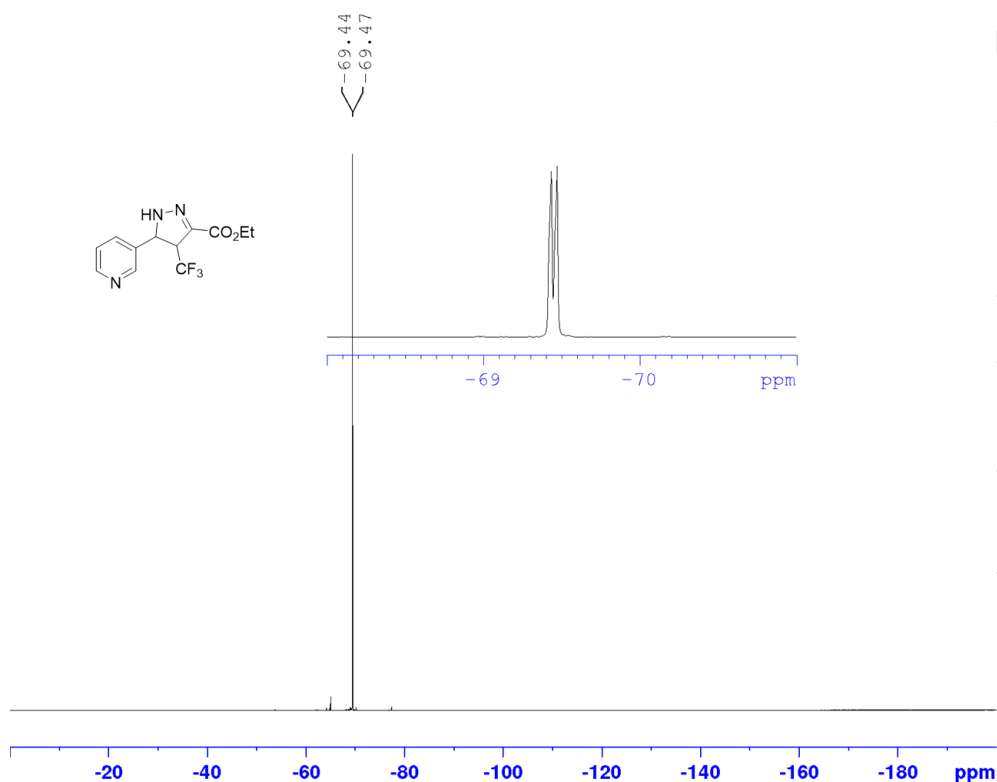


Current Data Parameters  
NAME mais21hmmH3  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190521  
Time 16.23 h  
INSTRUM spect  
PROBHD Z105225\_0009 ( )  
PULPROG zg30  
TD 32768  
SOLVENT CDCl3  
NS 8  
DS 0  
SWH 5000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 200.6  
DW 100.000 usec  
DE 6.50 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 250.1315446 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 7.19999981 W

F2 - Processing parameters  
SI 65536  
SF 250.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound **13**

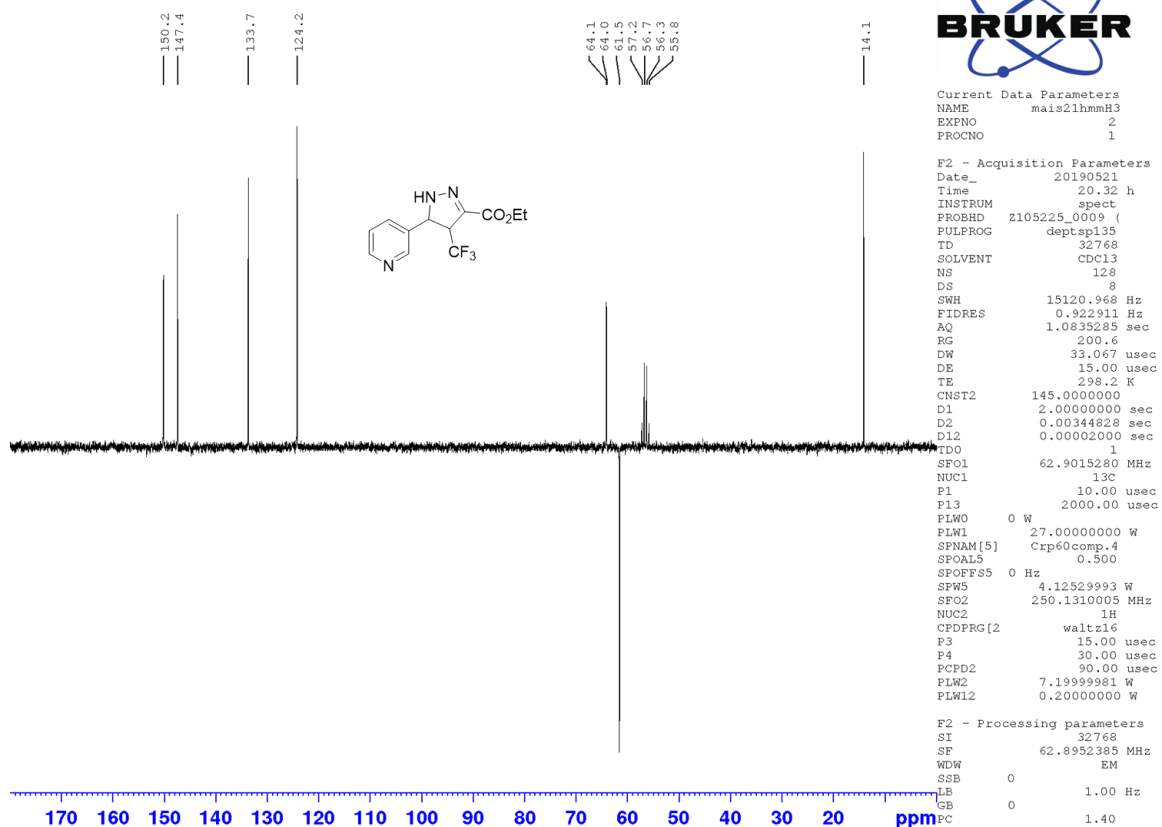


Current Data Parameters  
NAME mais21hmmH3  
EXPNO 4  
PROCNO 1

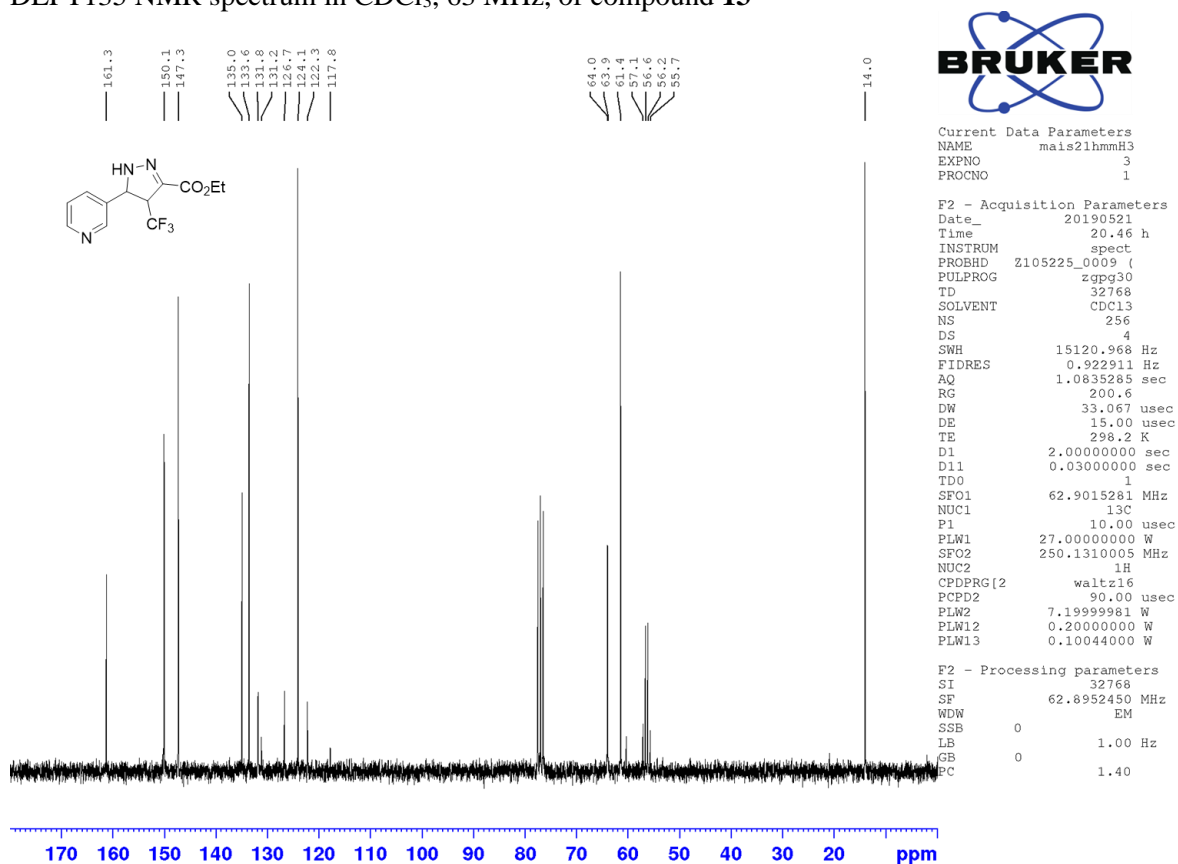
F2 - Acquisition Parameters  
Date\_ 20190521  
Time 21.03 h  
INSTRUM spect  
PROBHD Z105225\_0009 ( )  
PULPROG zgfgln  
TD 131072  
SOLVENT CDCl3  
NS 16  
DS 4  
SWH 56818.184 Hz  
FIDRES 0.866977 Hz  
AQ 1.1534336 sec  
RG 200.6  
DW 8.800 usec  
DE 6.50 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 235.3338140 MHz  
NUC1 19F  
P1 15.00 usec  
PLW1 6.80000019 W

F2 - Processing parameters  
SI 65536  
SF 235.3573497 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

<sup>19</sup>F NMR spectrum in CDCl<sub>3</sub>, 235 MHz, of compound **13**

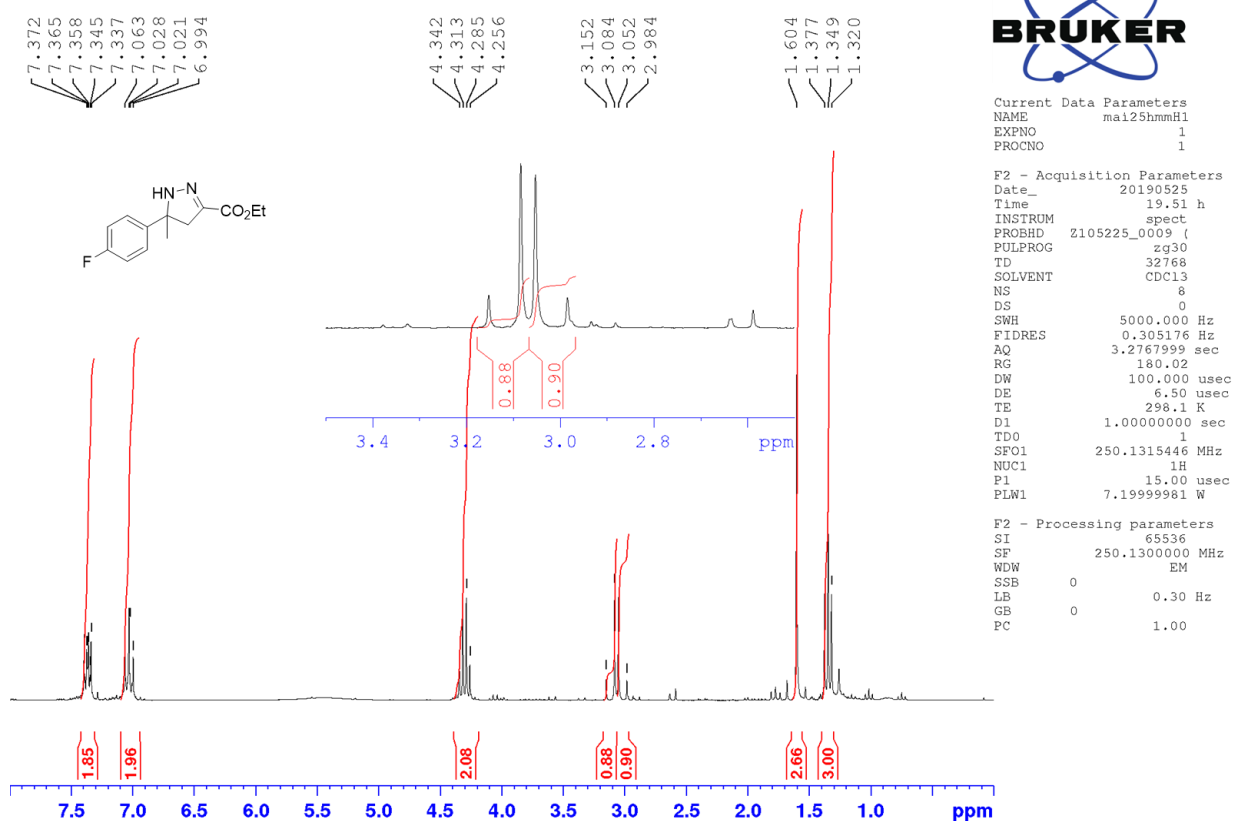


DEPT135 NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound 13

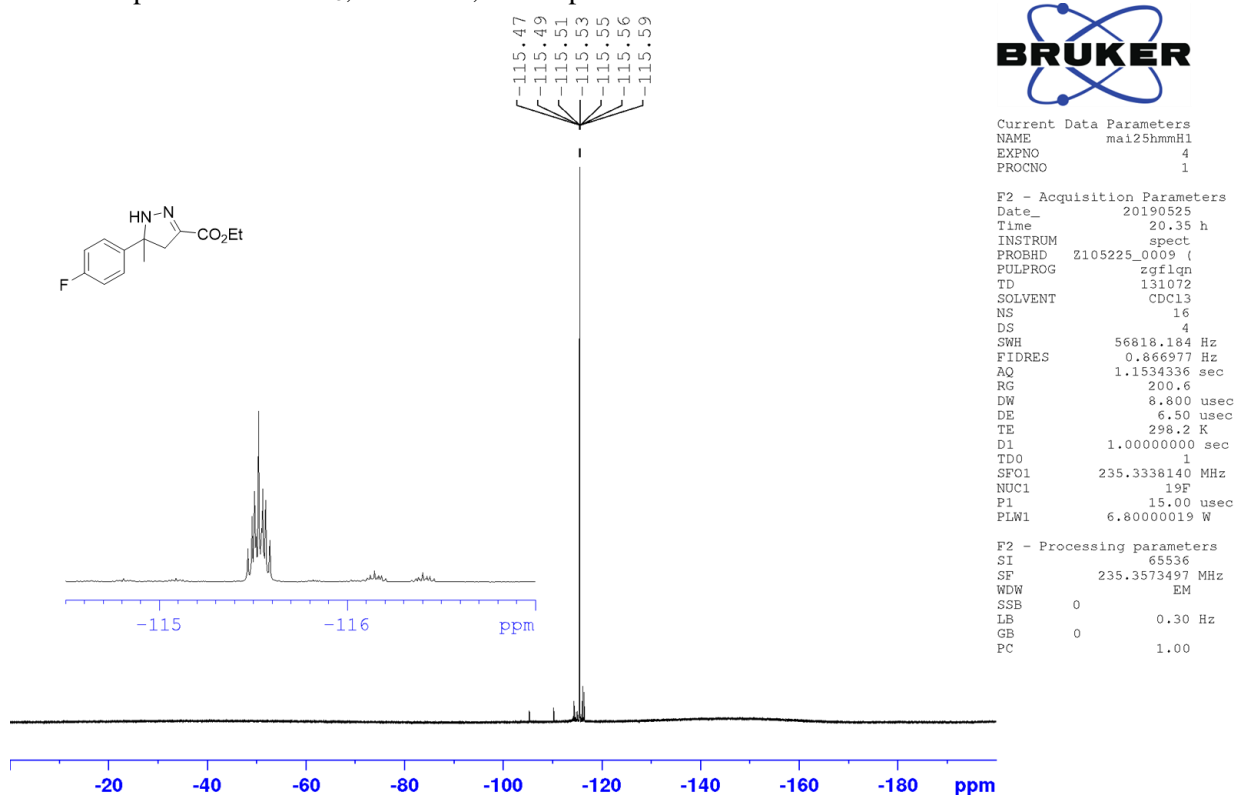


<sup>13</sup>C NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound 13

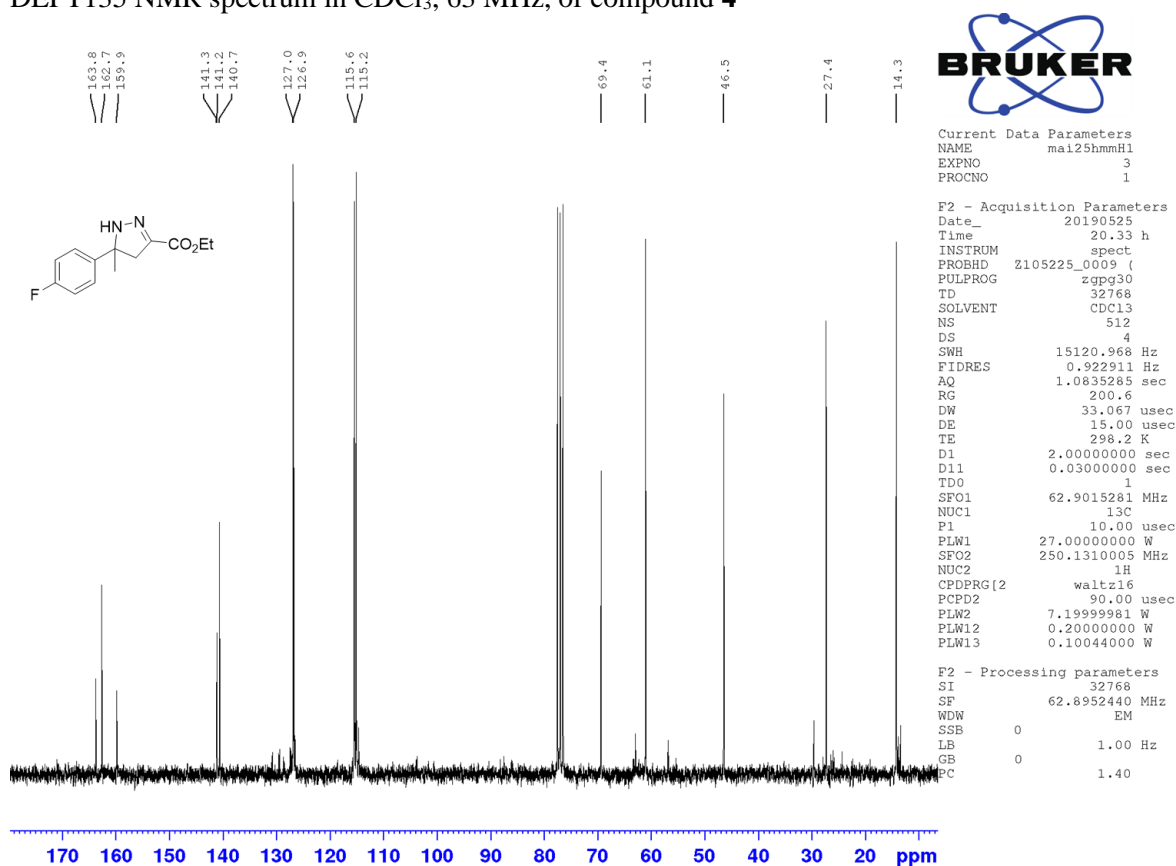
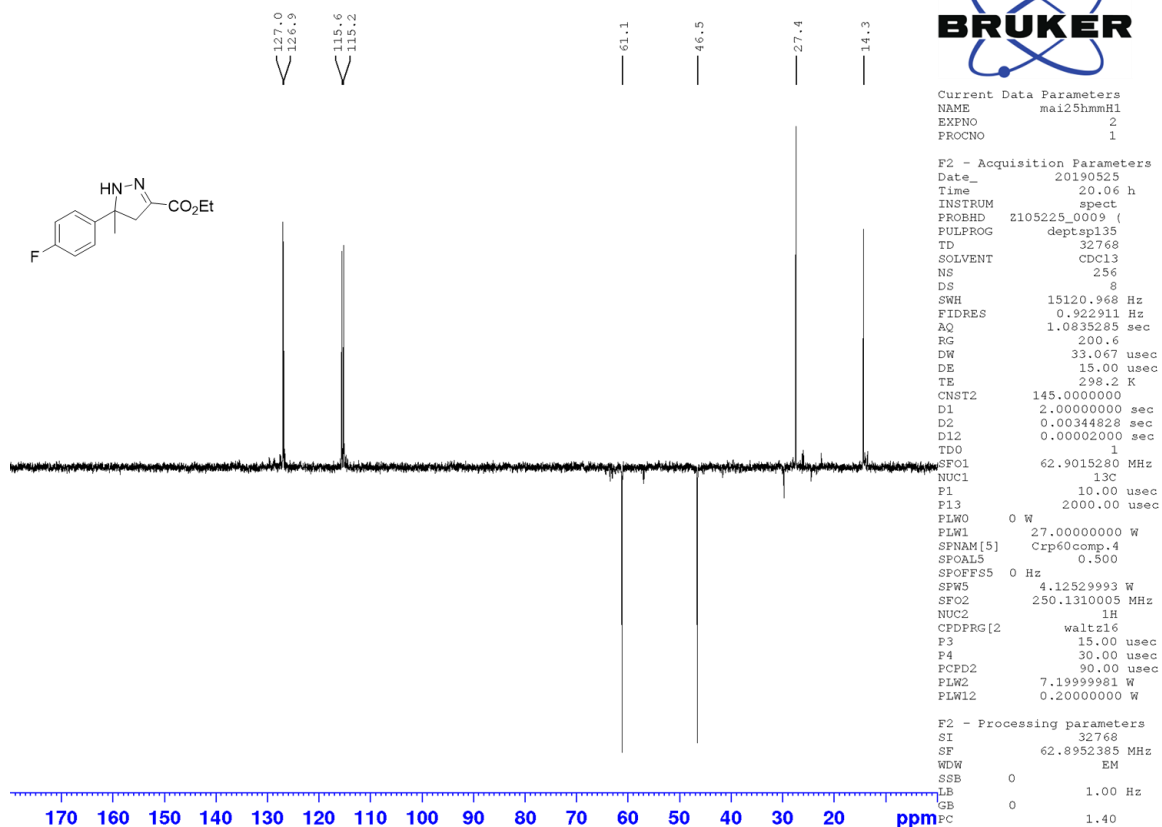
# Ethyl 5-(4-fluorophenyl)-5-methyl-4,5-dihydro-1H-pyrazole-3-carboxylate



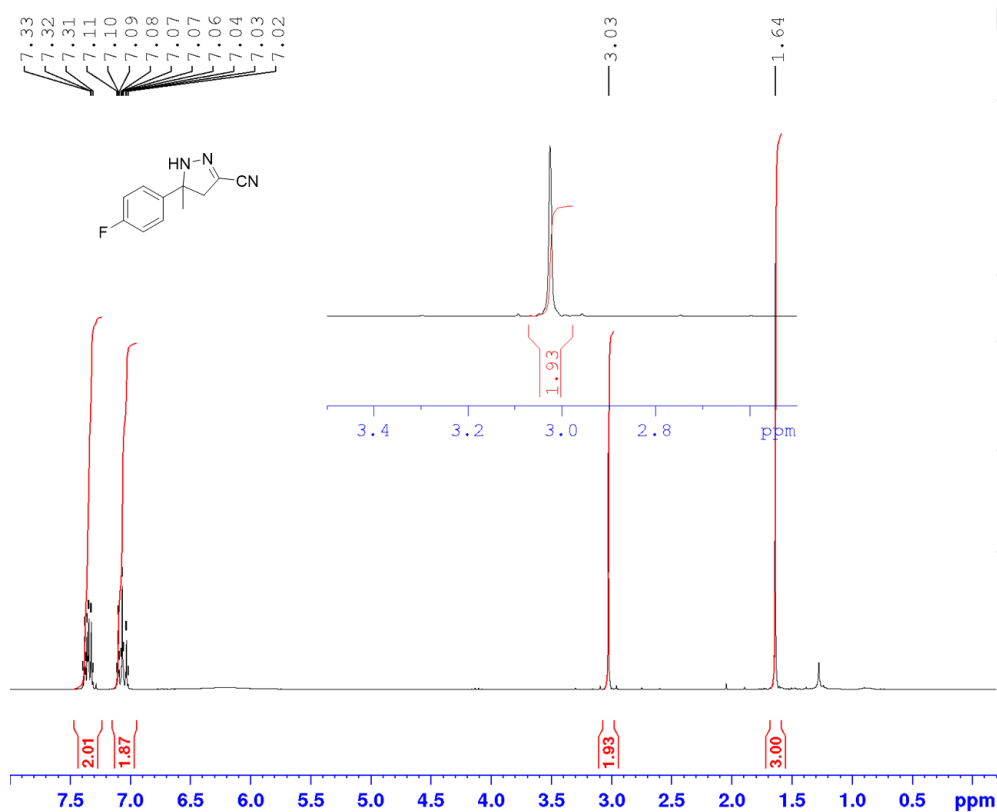
<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound 4



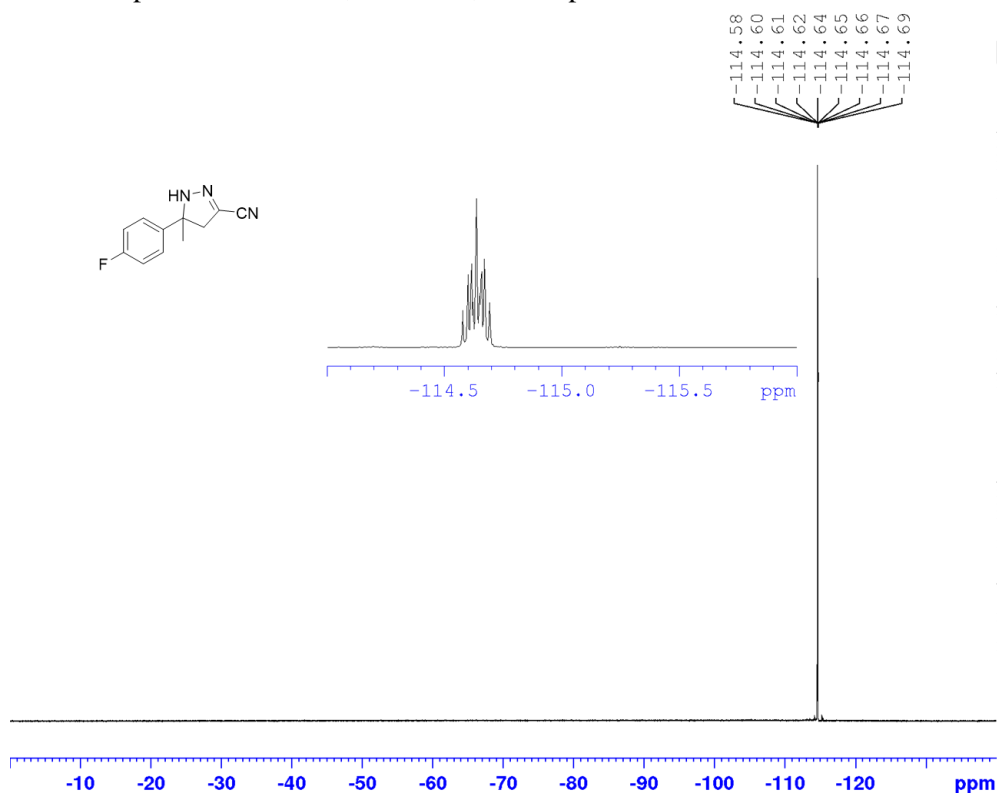
<sup>19</sup>F NMR spectrum in CDCl<sub>3</sub>, 235 MHz, of compound 4



# 5-(4-fluorophenyl)-5-methyl-4,5-dihydro-1H-pyrazole-3-carbonitrile



<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound **9**



<sup>19</sup>F NMR spectrum in CDCl<sub>3</sub>, 235 MHz, of compound **9**



Current Data Parameters  
NAME mai25hmmH2  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190525  
Time 21.15 h  
INSTRUM spect  
PROBHD Z105225\_0009 (   
PULPROG zg30  
TD 32768  
SOLVENT CDCl3  
NS 8  
DS 0  
SWH 5000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 180.02  
DW 100.000 usec  
DE 6.50 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 250.1315446 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 7.19999981 W

F2 - Processing parameters  
SI 65536  
SF 250.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

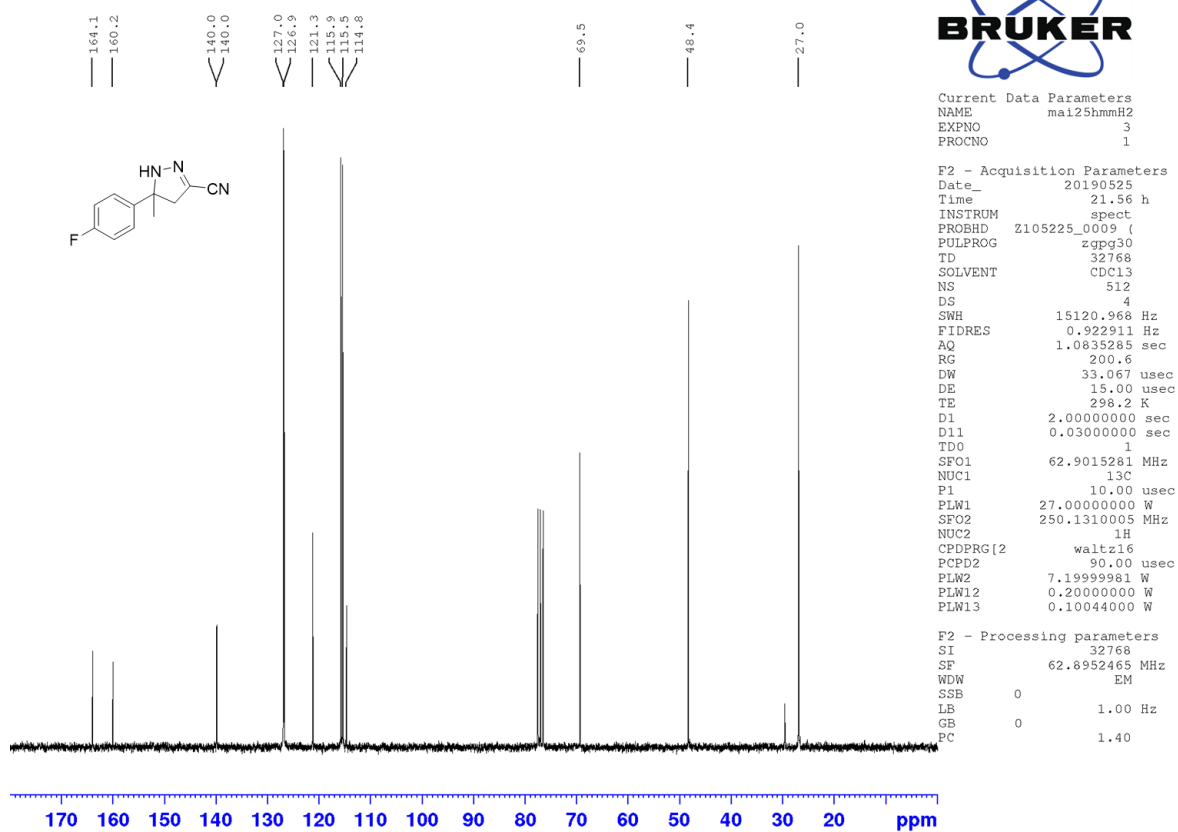
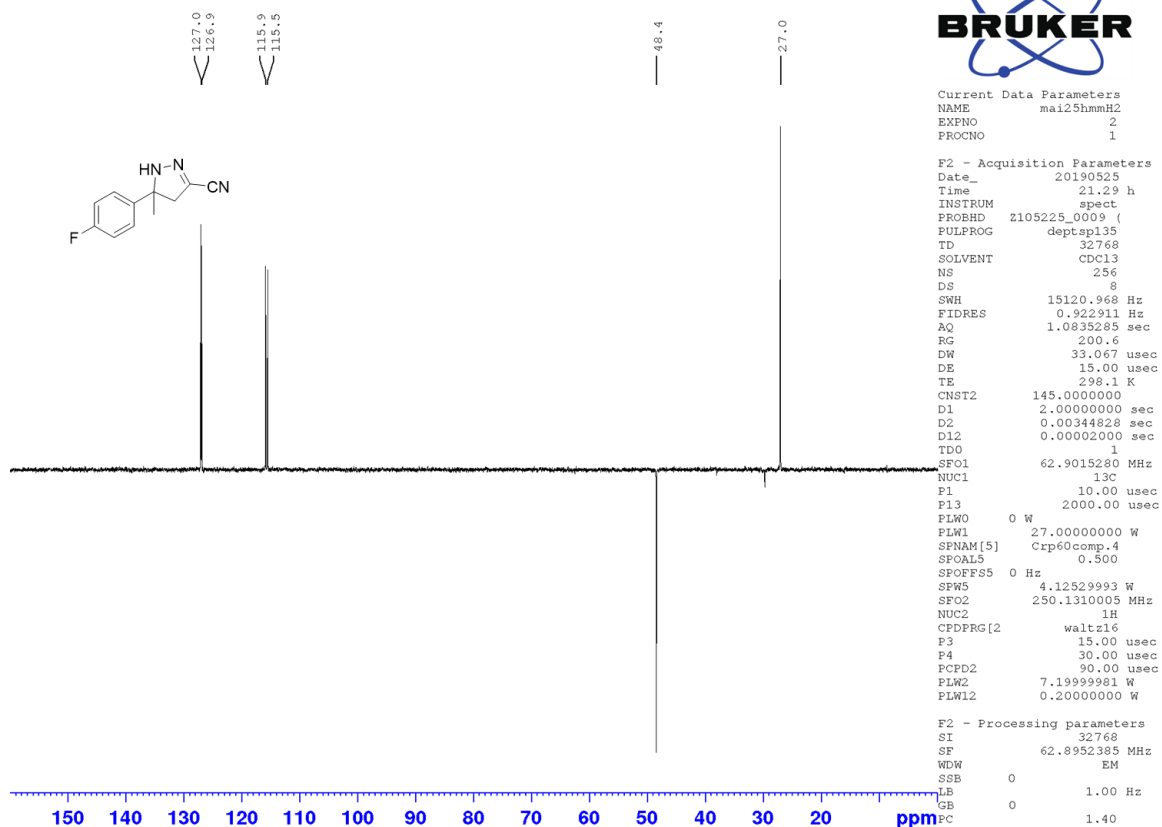


Current Data Parameters  
NAME mai25hmmH2  
EXPNO 4  
PROCNO 1

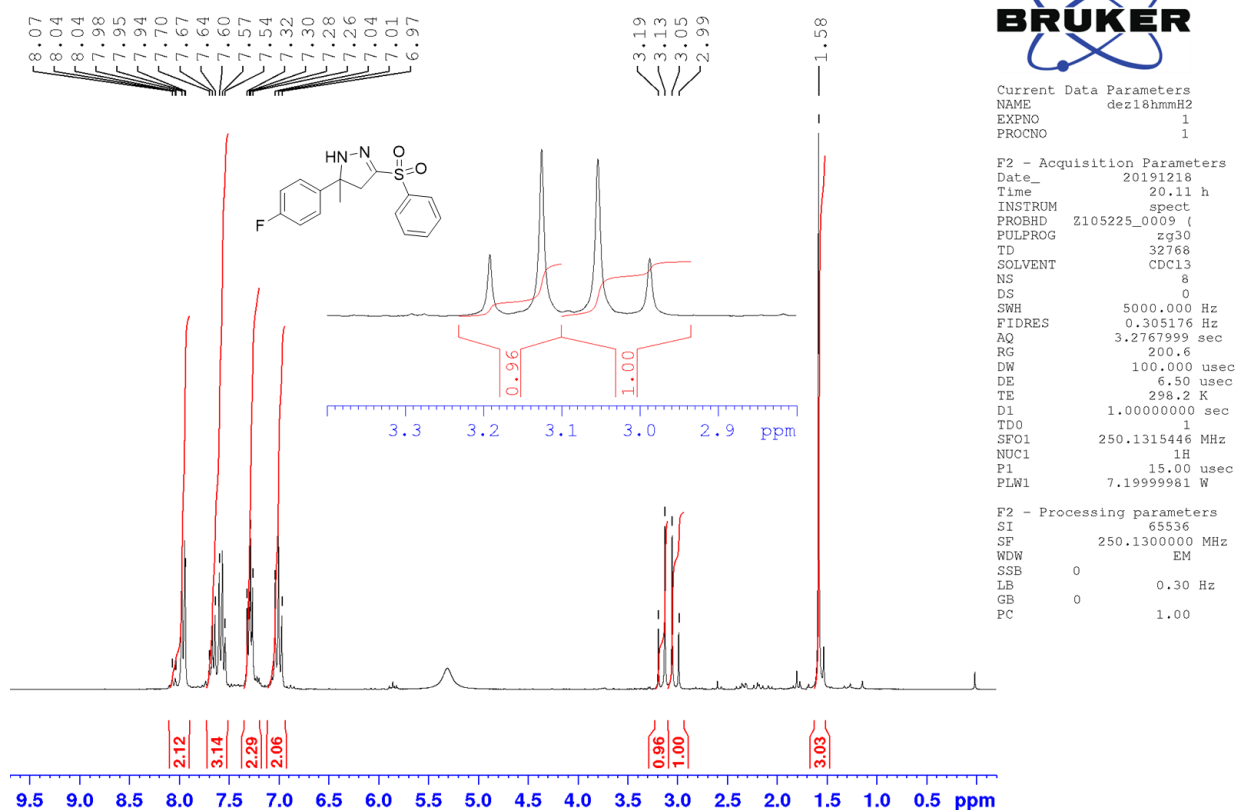
F2 - Acquisition Parameters  
Date\_ 20190525  
Time 21.58 h  
INSTRUM spect  
PROBHD Z105225\_0009 (   
PULPROG zgflgn  
TD 131072  
SOLVENT CDCl3  
NS 16  
DS 4  
SWH 56818.184 Hz  
FIDRES 0.866977 Hz  
AQ 1.1534336 sec  
RG 200.6  
DW 8.800 usec  
DE 6.50 usec  
TE 298.2 K  
D1 1.00000000 sec  
TD0 1  
SFO1 235.3338140 MHz  
NUC1 19F  
P1 15.00 usec  
PLW1 6.80000019 W

F2 - Processing parameters  
SI 65536  
SF 235.3573497 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

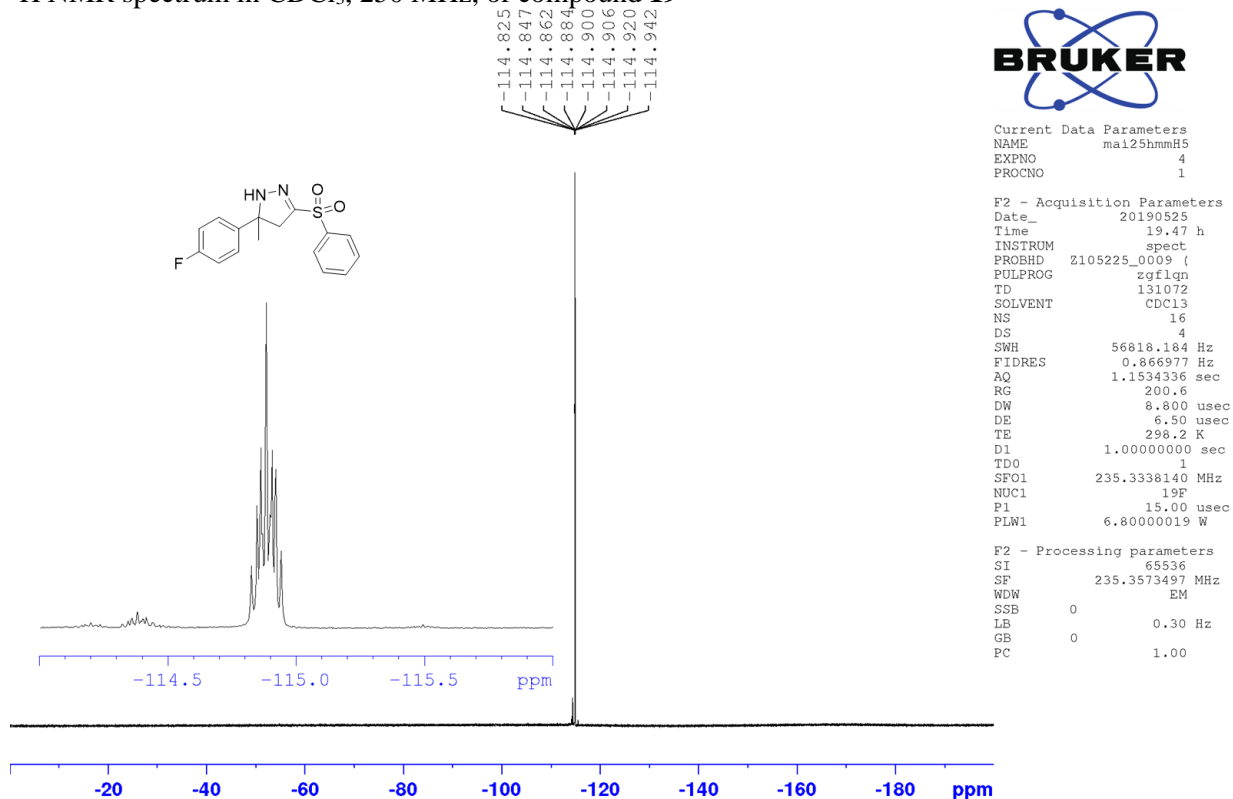




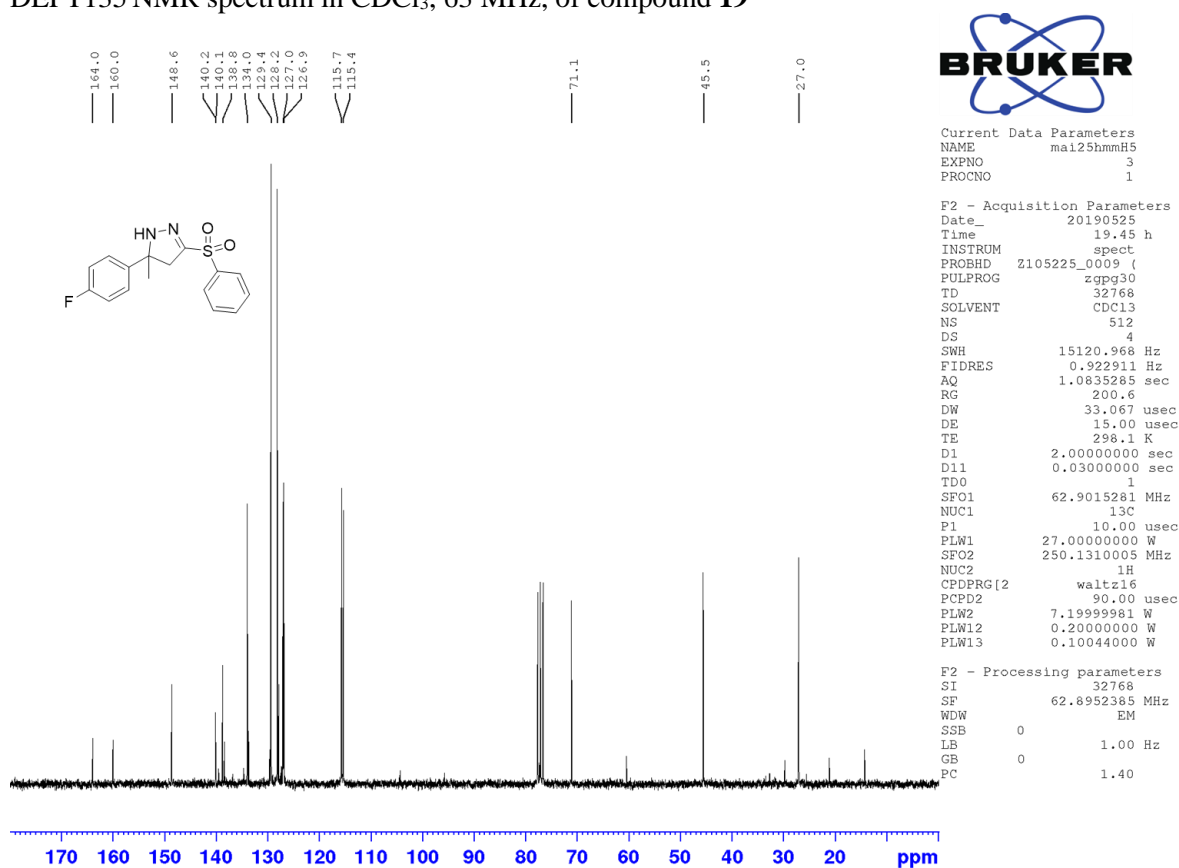
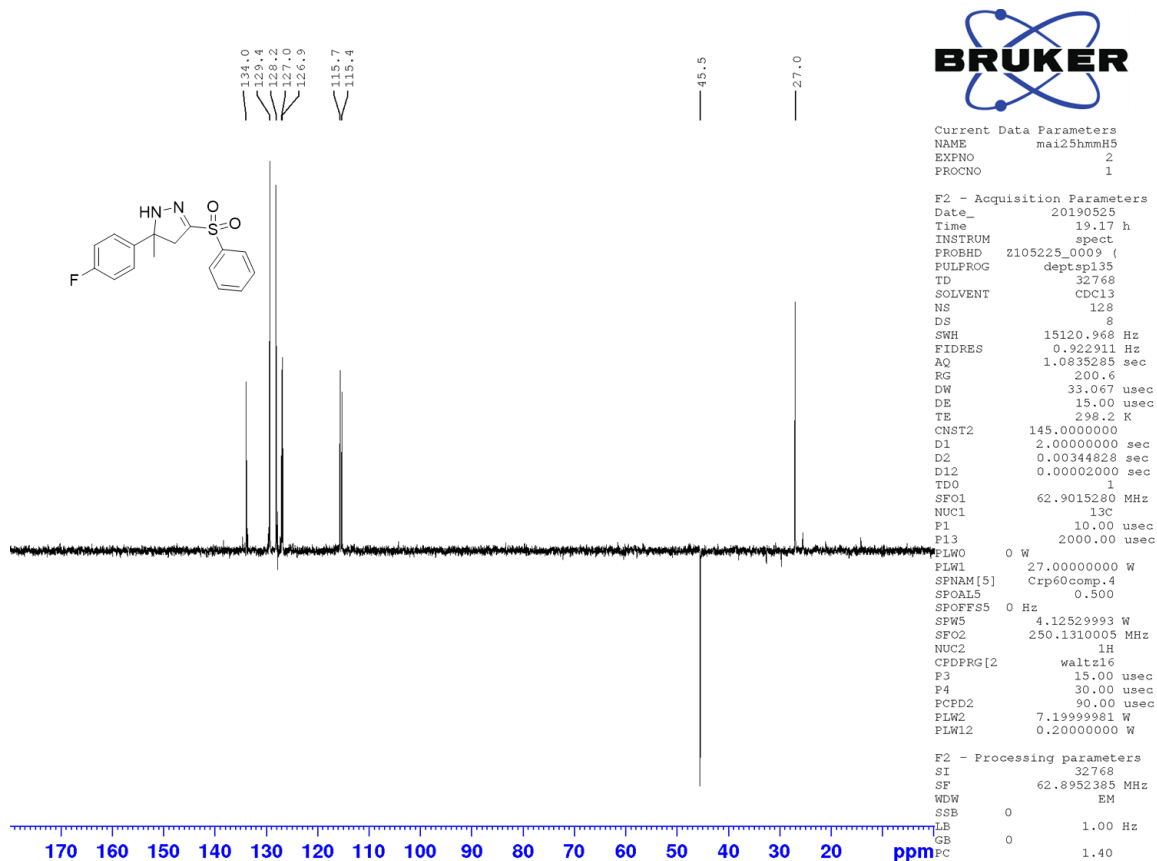
**5-(4-fluorophenyl)-5-methyl-3-(phenylsulfonyl)-4,5-dihydro-1H-pyrazole**



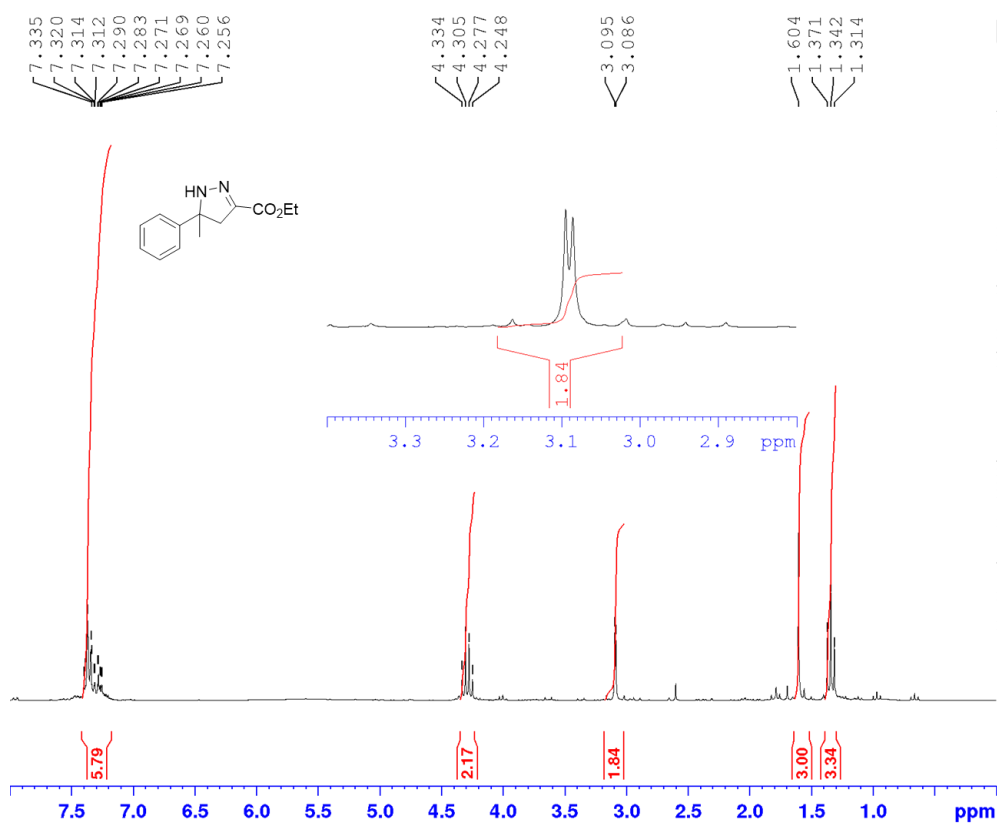
<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound **19**



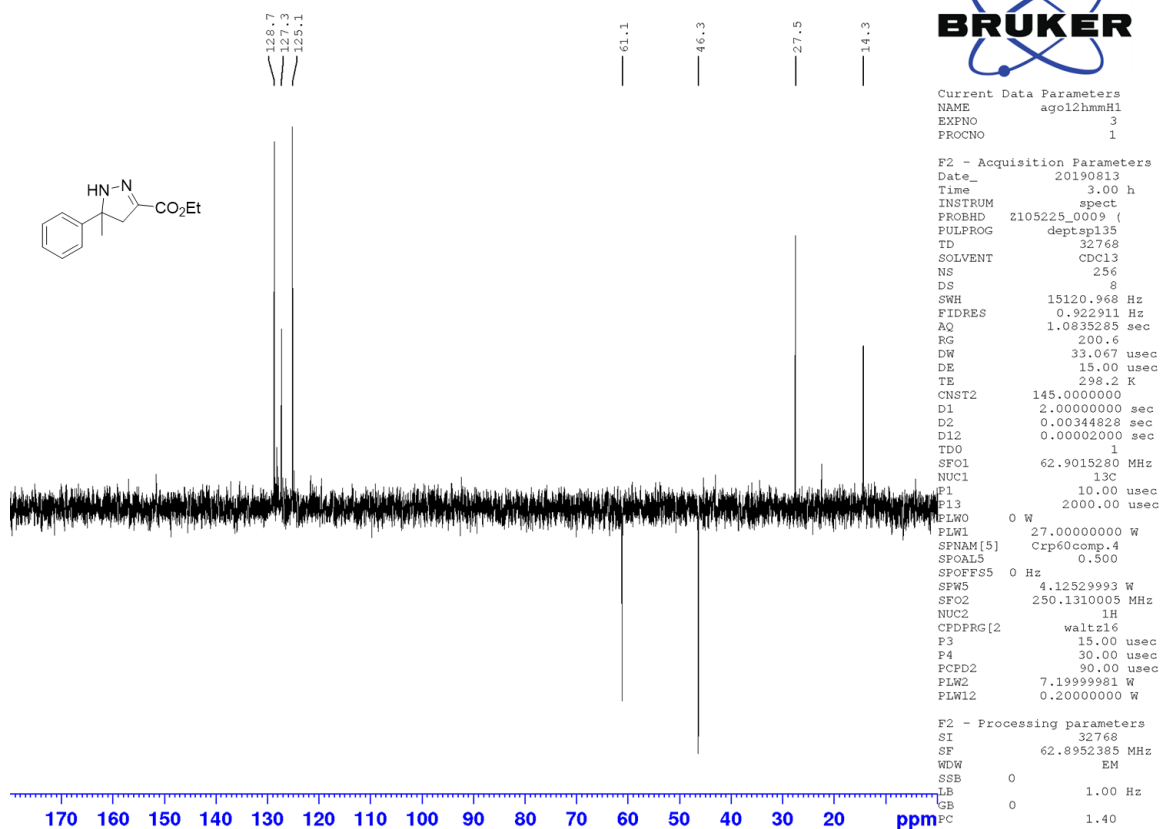
<sup>19</sup>F NMR spectrum in CDCl<sub>3</sub>, 235 MHz, of compound **19**



# Ethyl 5-methyl-5-phenyl-4,5-dihydro-1H-pyrazole-3-carboxylate



<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound **14**



DEPT135 NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **14**



Current Data Parameters  
NAME mai28hmmH1  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190528  
Time 17.24 h  
INSTRUM spect  
PROBHD Z105225\_0009 ( )  
PULPROG zg30  
TD 32768  
SOLVENT CDCl3  
NS 8  
DS 0  
SWH 5000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 114.73  
DW 100.000 usec  
DE 6.50 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 250.1315446 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 7.19999981 W

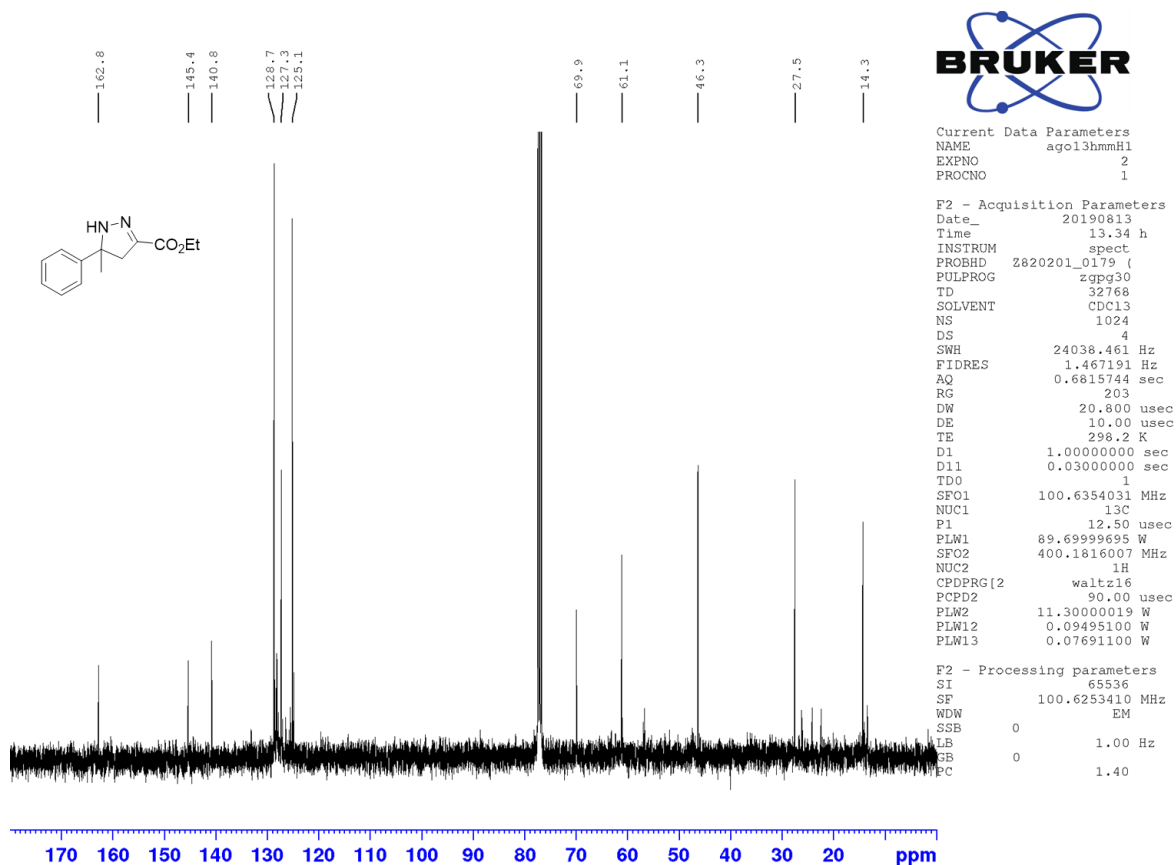
F2 - Processing parameters  
SI 55536  
SF 250.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
NAME agol2hmmH1  
EXPNO 3  
PROCNO 1

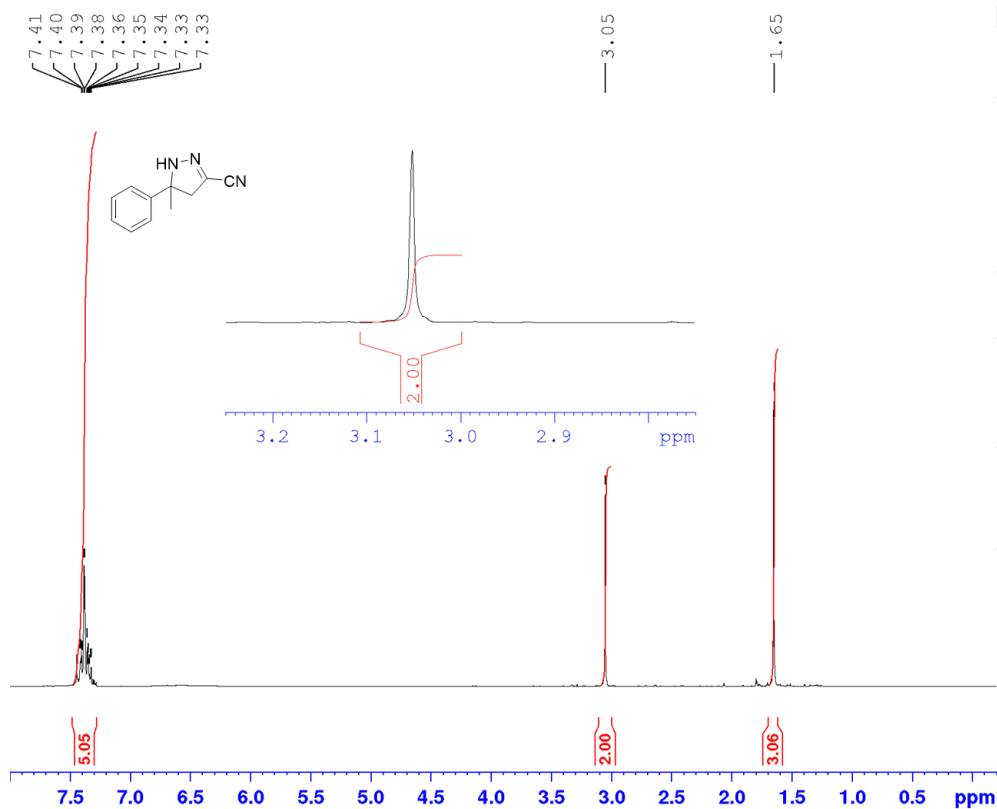
F2 - Acquisition Parameters  
Date\_ 20190813  
Time 3.00 h  
INSTRUM spect  
PROBHD Z105225\_0009 ( )  
PULPROG deptspl35  
TD 32768  
SOLVENT CDCl3  
NS 256  
DS 8  
SWH 15120.968 Hz  
FIDRES 0.922911 Hz  
AQ 1.0835285 sec  
RG 200.6  
DW 33.067 usec  
DE 15.00 usec  
TE 298.2 K  
CNST2 145.0000000  
D1 2.00000000 sec  
D2 0.00344828 sec  
D12 0.00002000 sec  
TD0 1  
SFO1 62.9015280 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 2000.00 usec  
PLW0 0 W  
PLW1 27.00000000 W  
SPNAM[5] Crp60comp.4  
SFOAL5 0.500  
SPOFFS5 0 Hz  
SPW5 4.12529993 W  
SFO2 250.1310005 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
P3 15.00 usec  
P4 30.00 usec  
PCPD2 90.00 usec  
PLW2 7.19999981 W  
PLW12 0.20000000 W

F2 - Processing parameters  
SI 32768  
SF 62.8952385 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



$^{13}\text{C}$  NMR spectrum in  $\text{CDCl}_3$ , 100 MHz, of compound **14**

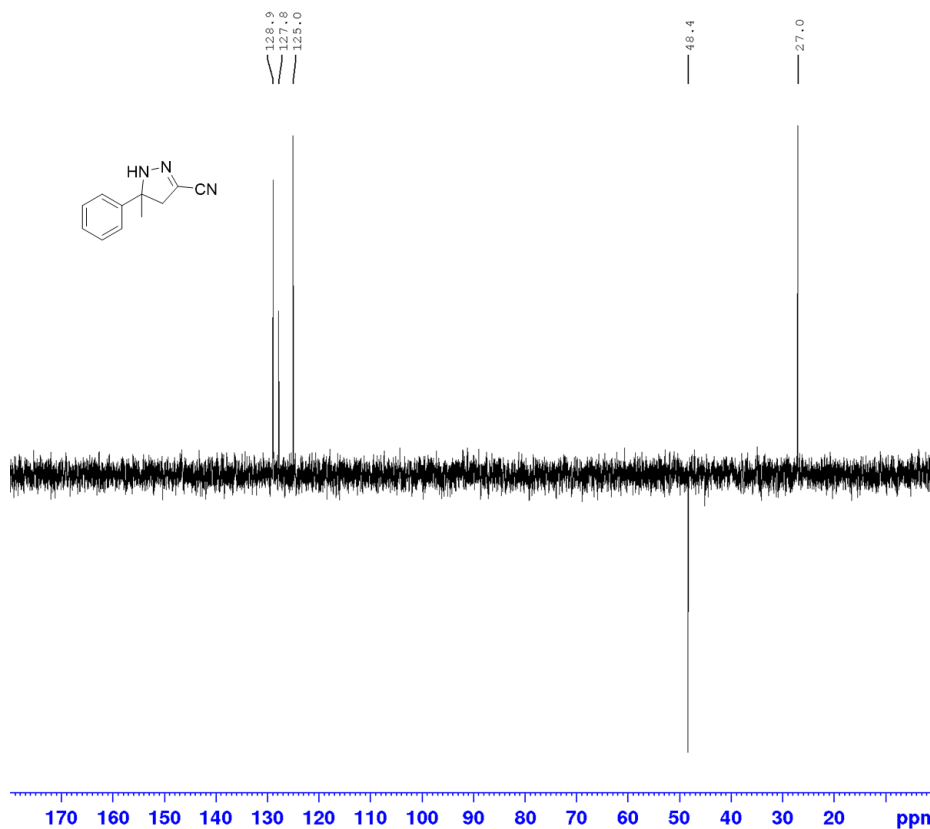
# 5-methyl-5-phenyl-4,5-dihydro-1H-pyrazole-3-carbonitrile



Current Data Parameters  
NAME mai28hmmH2  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190528  
Time 17.29 h  
INSTRUM spect  
PROBHD Z105225\_0009 (Z105225)  
PULPROG zg30  
TD 32768  
SOLVENT CDCl3  
NS 8  
DS 0  
SWH 5000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 162.14  
DW 100.000 usec  
DE 6.50 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 250.1315446 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 7.19999981 W

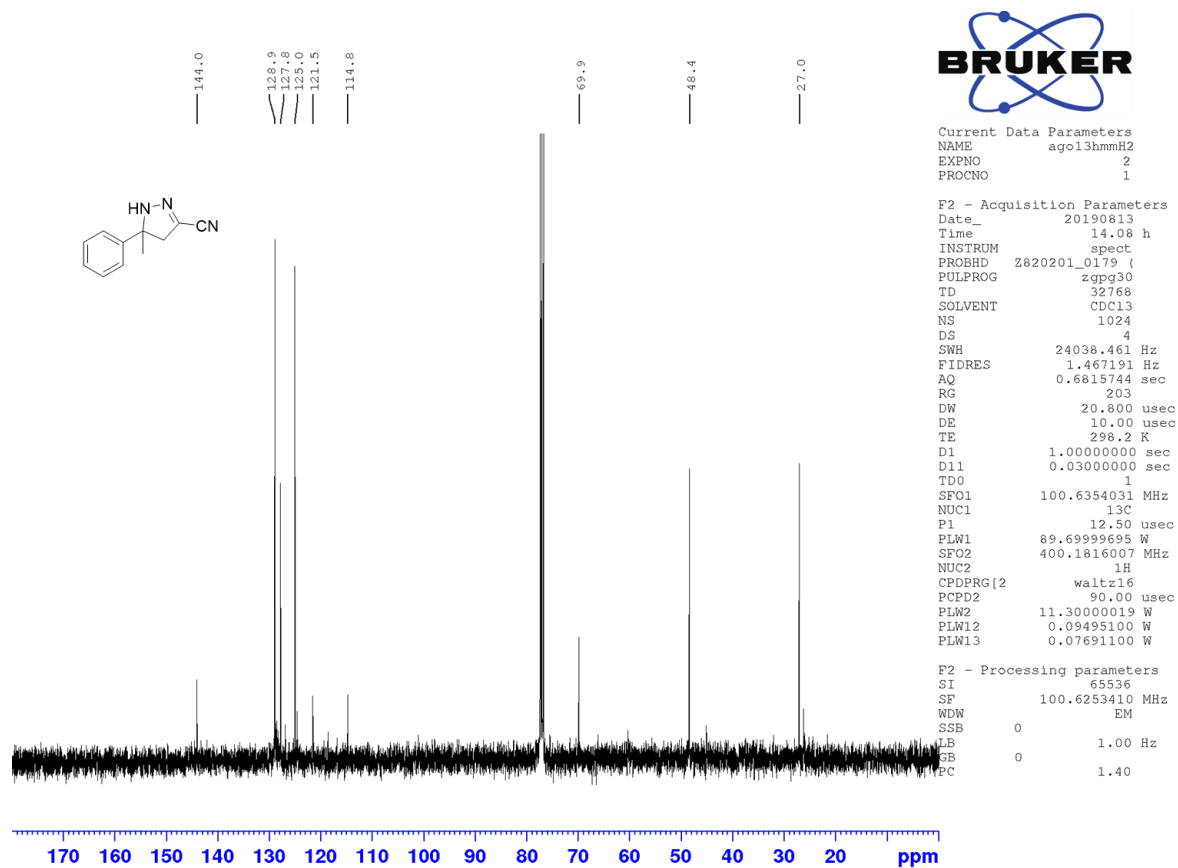
F2 - Processing parameters  
SI 55536  
SF 250.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
NAME agol2hmmH2  
EXPNO 3  
PROCNO 1

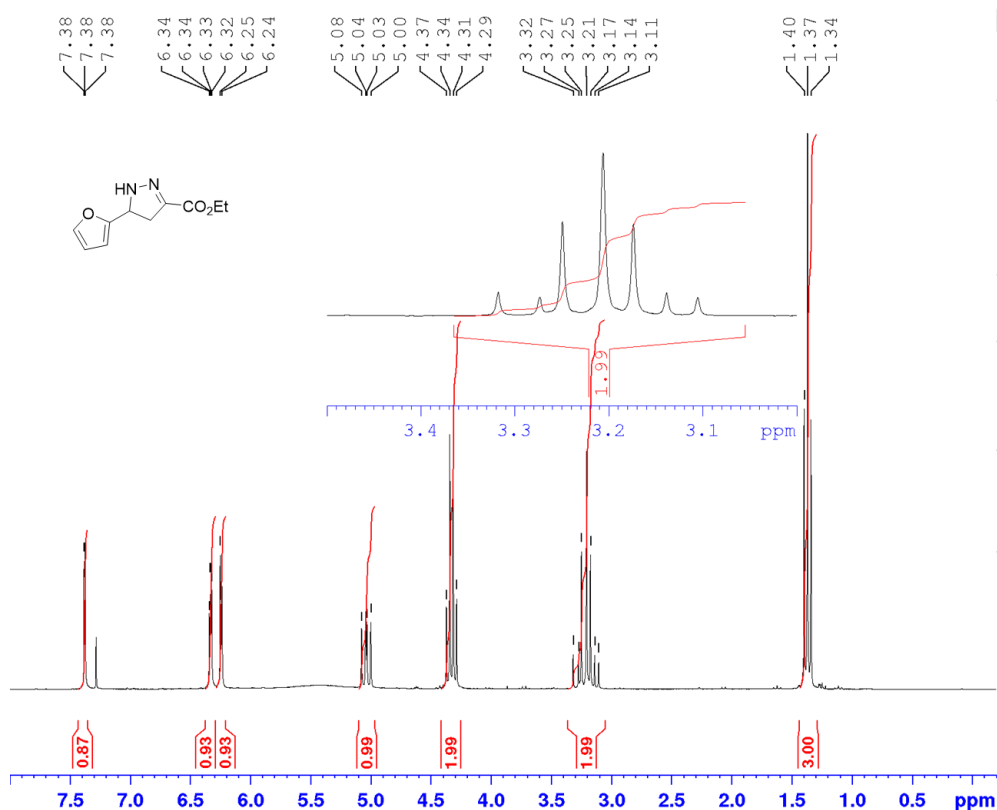
F2 - Acquisition Parameters  
Date\_ 20190813  
Time 3.47 h  
INSTRUM spect  
PROBHD Z105225\_0009 (Z105225)  
PULPROG deptspl35  
TD 32768  
SOLVENT CDCl3  
NS 256  
DS 8  
SWH 15120.968 Hz  
FIDRES 0.922911 Hz  
AQ 1.0835285 sec  
RG 200.6  
DW 33.067 usec  
DE 15.00 usec  
TE 298.2 K  
CNST2 145.0000000  
D1 2.00000000 sec  
D2 0.00344828 sec  
D12 0.00002000 sec  
TD0 1  
SFO1 62.9015280 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 2000.00 usec  
PLW0 0 W  
SPNAM[5] Crp60comp.4  
SFOALS 0.500  
SPOFFS5 0 Hz  
SPW5 4.12529993 W  
SFO2 250.1310005 MHz  
NUC2 1H  
CPDPRG2 waltz16  
P3 15.00 usec  
P4 30.00 usec  
PCPD2 90.00 usec  
PLW2 7.19999981 W  
PLW12 0.20000000 W

F2 - Processing parameters  
SI 32768  
SF 62.8952385 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



<sup>13</sup>C NMR spectrum in CDCl<sub>3</sub>, 100 MHz, of compound **18**

# Ethyl 5-(furan-2-yl)-4,5-dihydro-1H-pyrazole-3-carboxylate

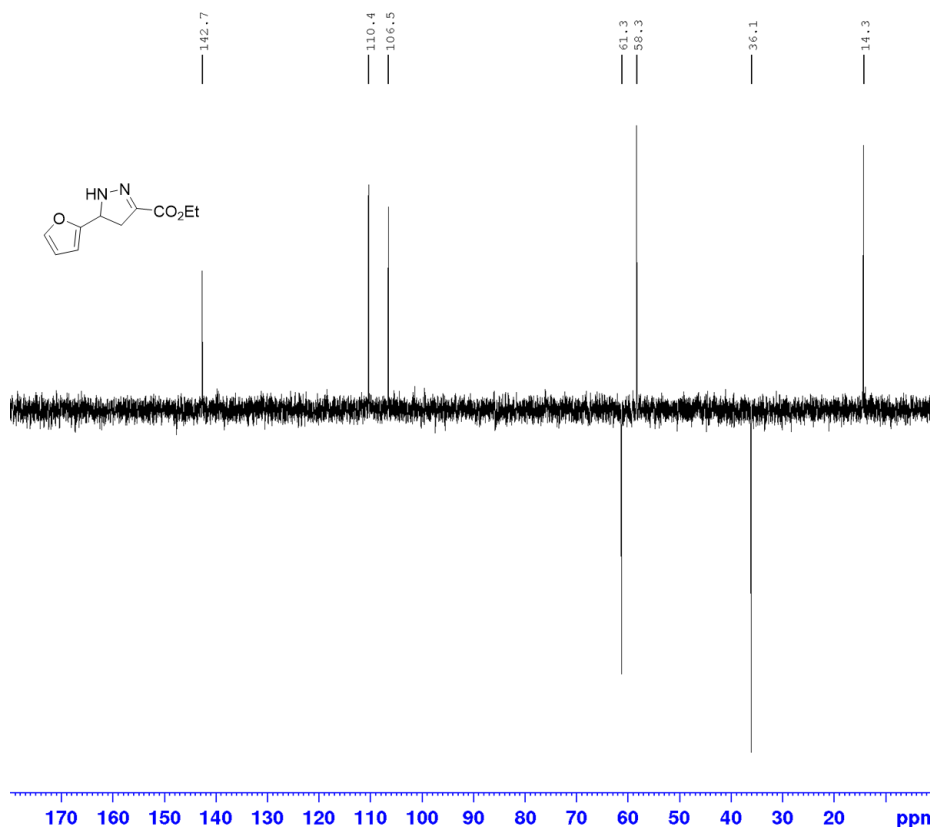


Current Data Parameters  
NAME jull13hmmH1\_A  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190713  
Time 19.05 h  
INSTRUM spect  
PROBHD Z105225\_0009 ( )  
PULPROG zg30  
TD 32768  
SOLVENT CDCl3  
NS 16  
DS 0  
SWH 5000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 200.6  
DE 100.000 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 250.1315446 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 7.19999981 W

F2 - Processing parameters  
SI 55536  
SF 250.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound **5**



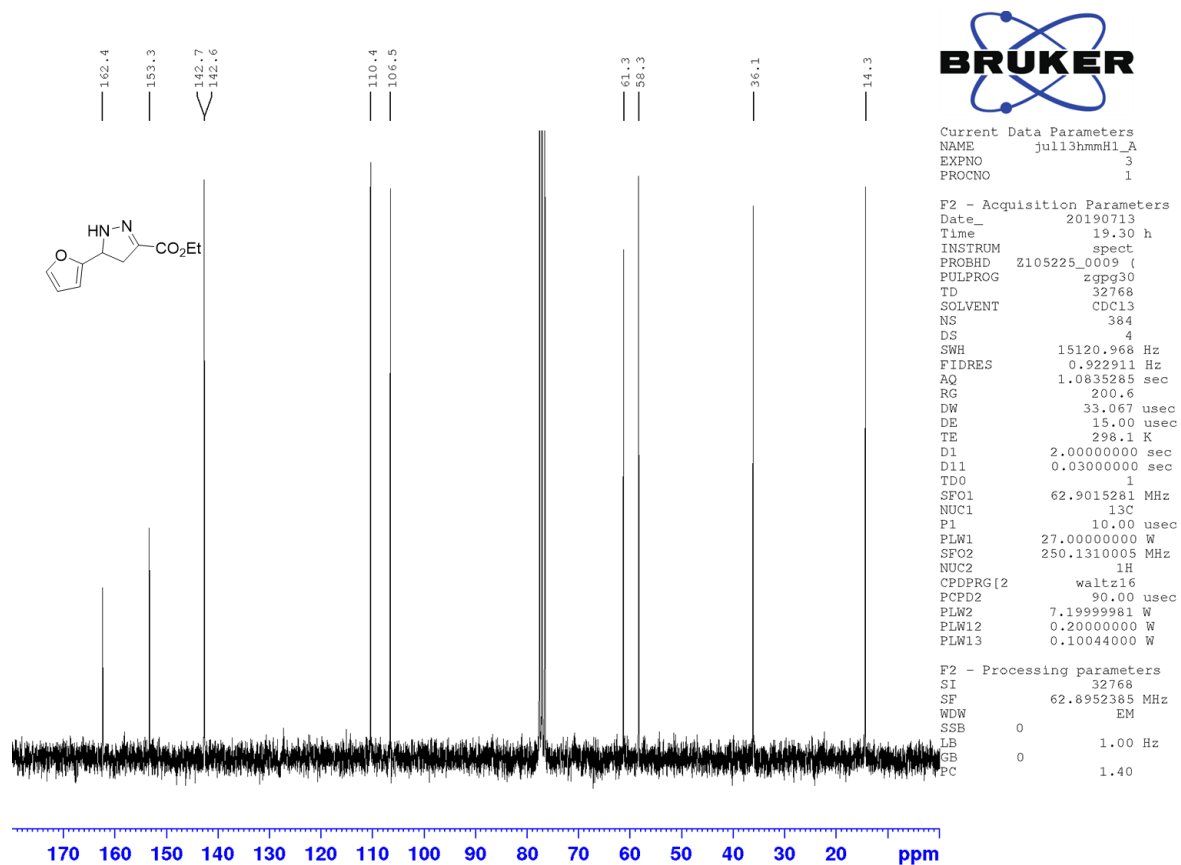
Current Data Parameters  
NAME jull13hmmH1\_A  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190713  
Time 19.09 h  
INSTRUM spect  
PROBHD Z105225\_0009 ( )  
PULPROG deptsp135  
TD 32768  
SOLVENT CDCl3  
NS 64  
DS 8  
SWH 15120.968 Hz  
FIDRES 0.922911 Hz  
AQ 1.0835285 sec  
RG 200.6  
DE 33.067 usec  
TE 298.2 K  
CNST2 145.0000000  
D1 2.00000000 sec  
D2 0.00344828 sec  
D12 0.00002000 sec  
TD0 1  
SFO1 62.9015280 MHz  
NUC1 13C  
P1 10.00 usec  
P13 2000.00 usec  
PLW0 0 W  
PLW1 27.00000000 W  
SPNAM[5] Crp60comp.4  
SFOAL5 0.500  
SPOFFS5 0 Hz  
SPW5 4.12529993 W  
SFO2 250.1310005 MHz  
NUC2 1H  
CFDPRG[2] waltz16  
P3 15.00 usec  
P4 30.00 usec  
PCPD2 90.00 usec  
PLW2 7.19999981 W  
PLW12 0.20000000 W

F2 - Processing parameters  
SI 32768  
SF 62.8952385 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

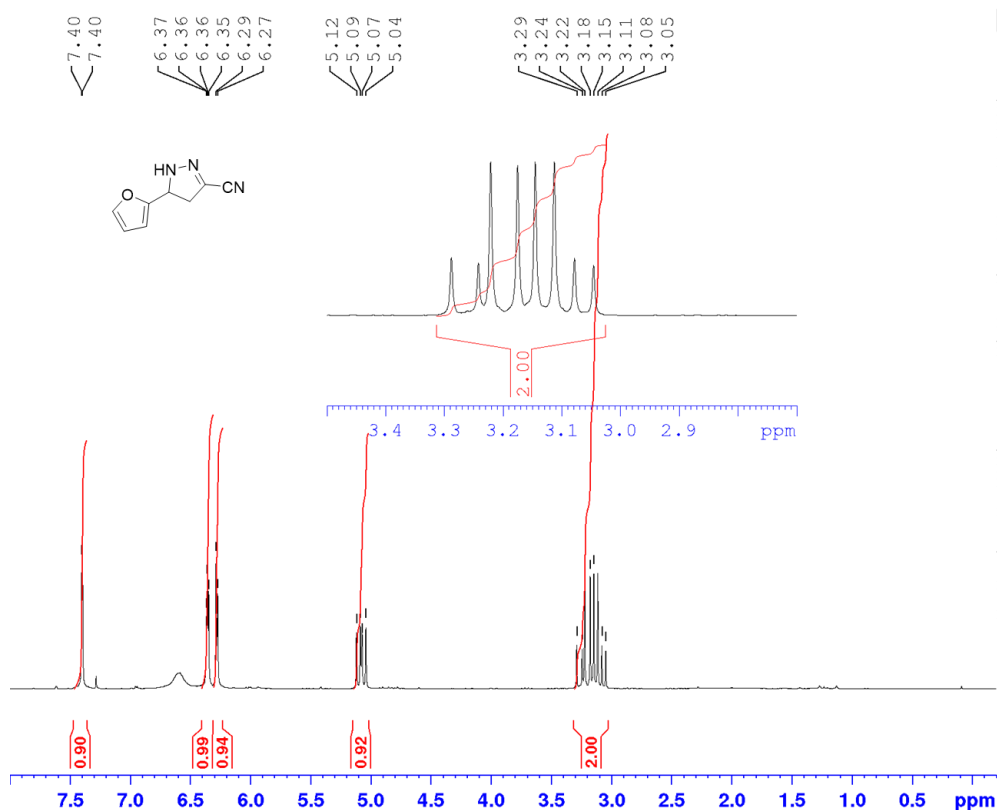
DEPT135 NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **5**



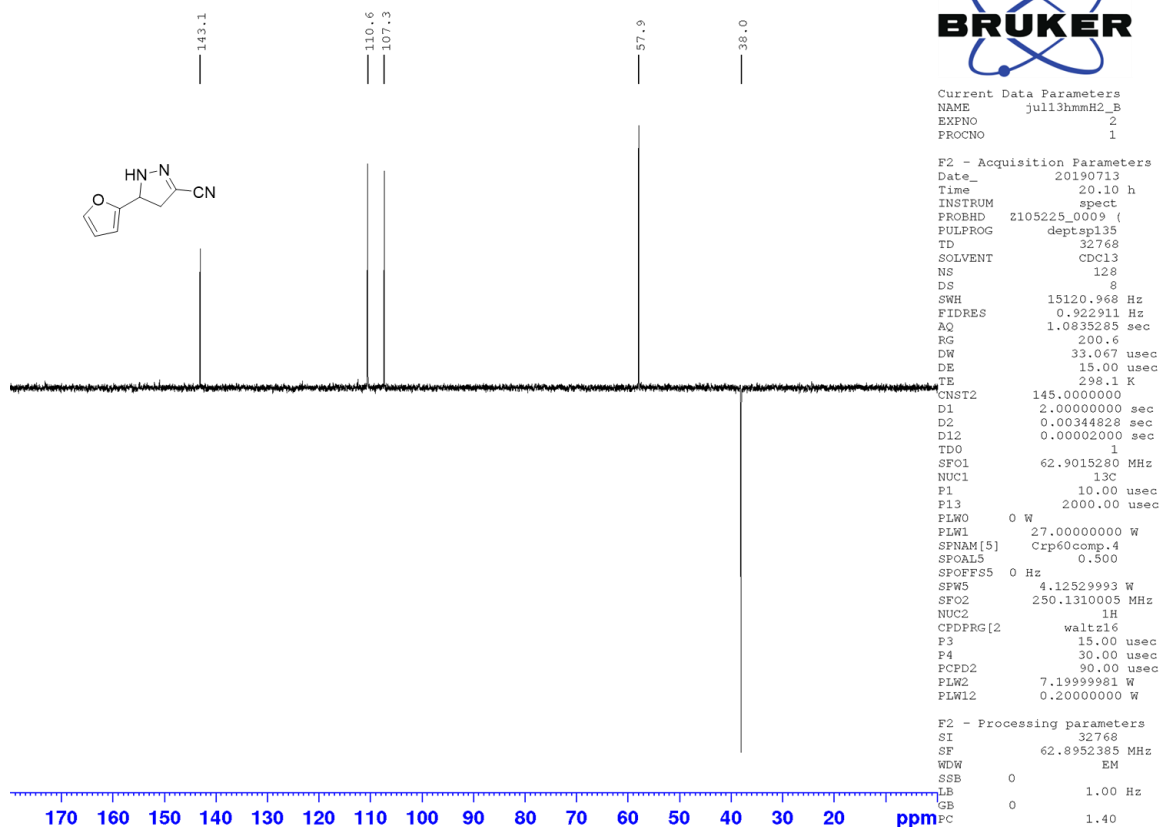


<sup>13</sup>C NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **5**

# 5-(furan-2-yl)-4,5-dihydro-1H-pyrazole-3-carbonitrile



<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound **10**



DEPT135 NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **10**



Current Data Parameters  
NAME jull13hmmH2\_B  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190713  
Time 19.35 h  
INSTRUM spect  
PROBHD Z105225\_0009 (zg30)  
PULPROG zg30  
TD 32768  
SOLVENT CDCl3  
NS 16  
DS 0  
SWH 5000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.2767999 sec  
RG 200.6  
DW 100.000 usec  
DE 6.50 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 250.1315446 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 7.19999981 W

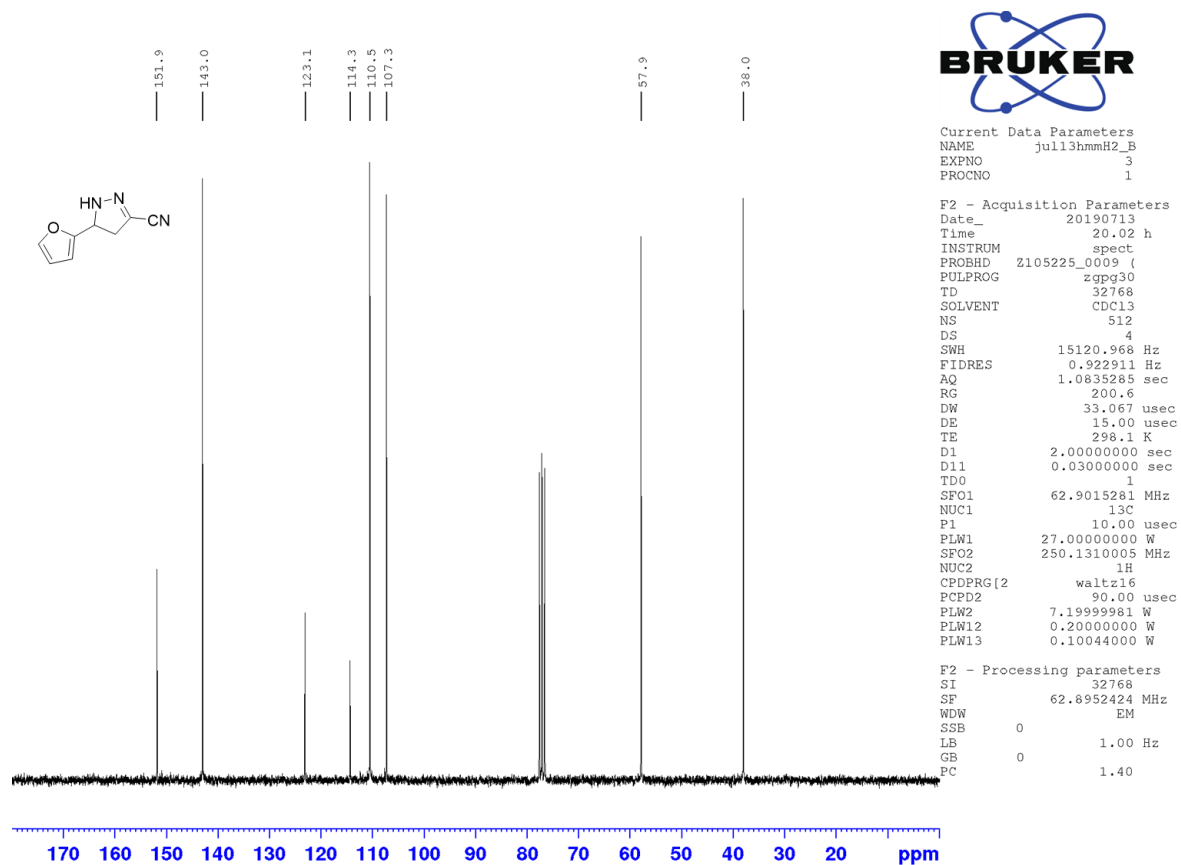
F2 - Processing parameters  
SI 55536  
SF 250.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
NAME jull13hmmH2\_B  
EXPNO 2  
PROCNO 1

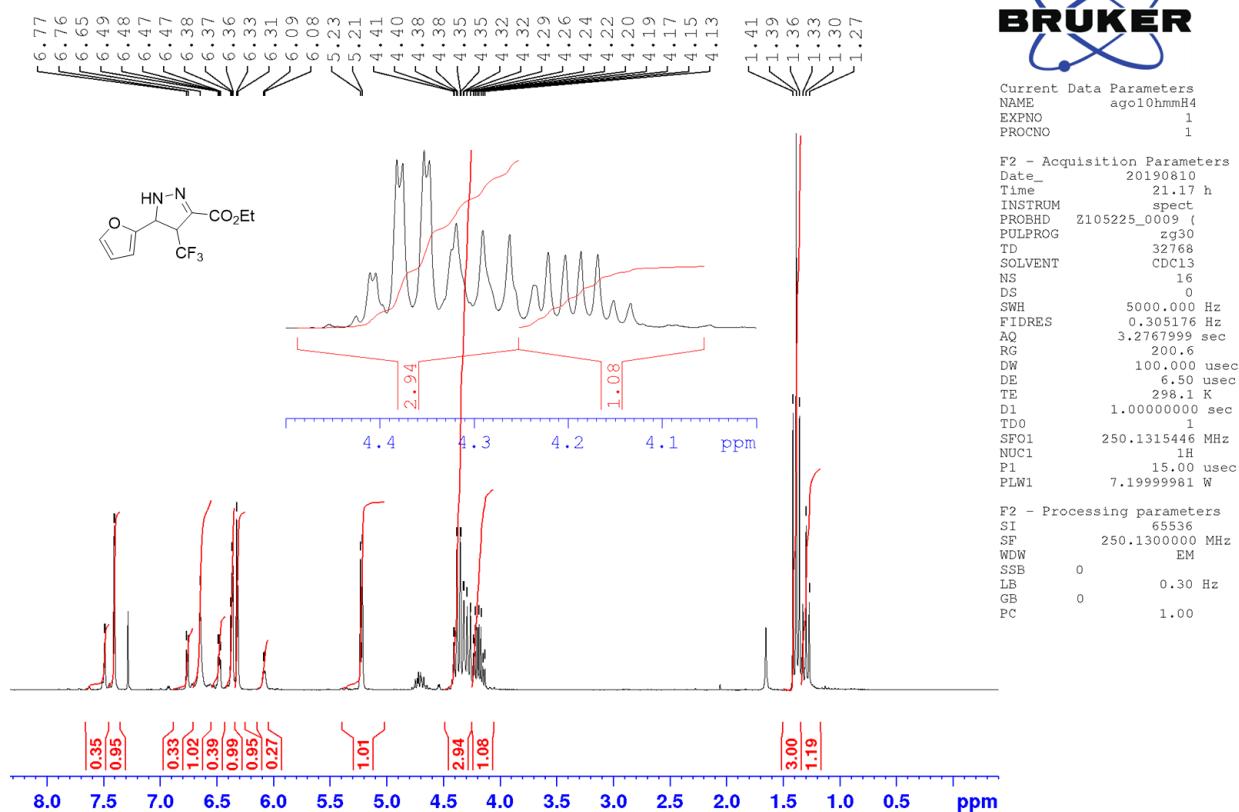
F2 - Acquisition Parameters  
Date\_ 20190713  
Time 20.10 h  
INSTRUM spect  
PROBHD Z105225\_0009 (deptsp135)  
PULPROG deptsp135  
TD 32768  
SOLVENT CDCl3  
NS 128  
DS 8  
SWH 15120.968 Hz  
FIDRES 0.922911 Hz  
AQ 1.0835285 sec  
RG 200.6  
DW 33.067 usec  
DE 15.00 usec  
TE 298.1 K  
CNST2 145.0000000  
D1 2.00000000 sec  
D2 0.00344828 sec  
D12 0.00002000 sec  
TD0 1  
SFO1 62.9015280 MHz  
NUC1 13C  
P1 10.00 usec  
P13 2000.00 usec  
PLW0 0 W  
PLW1 27.00000000 W  
SPNAM[5] Crp60comp.4  
SFOAL5 0.500  
SPOFFS5 0 Hz  
SPW5 4.12529993 W  
SFO2 250.1310005 MHz  
NUC2 1H  
CFDPFRG[2] waltz16  
P3 15.00 usec  
P4 30.00 usec  
PCPD2 90.00 usec  
PLW2 7.19999981 W  
PLW12 0.20000000 W

F2 - Processing parameters  
SI 32768  
SF 62.8952385 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40

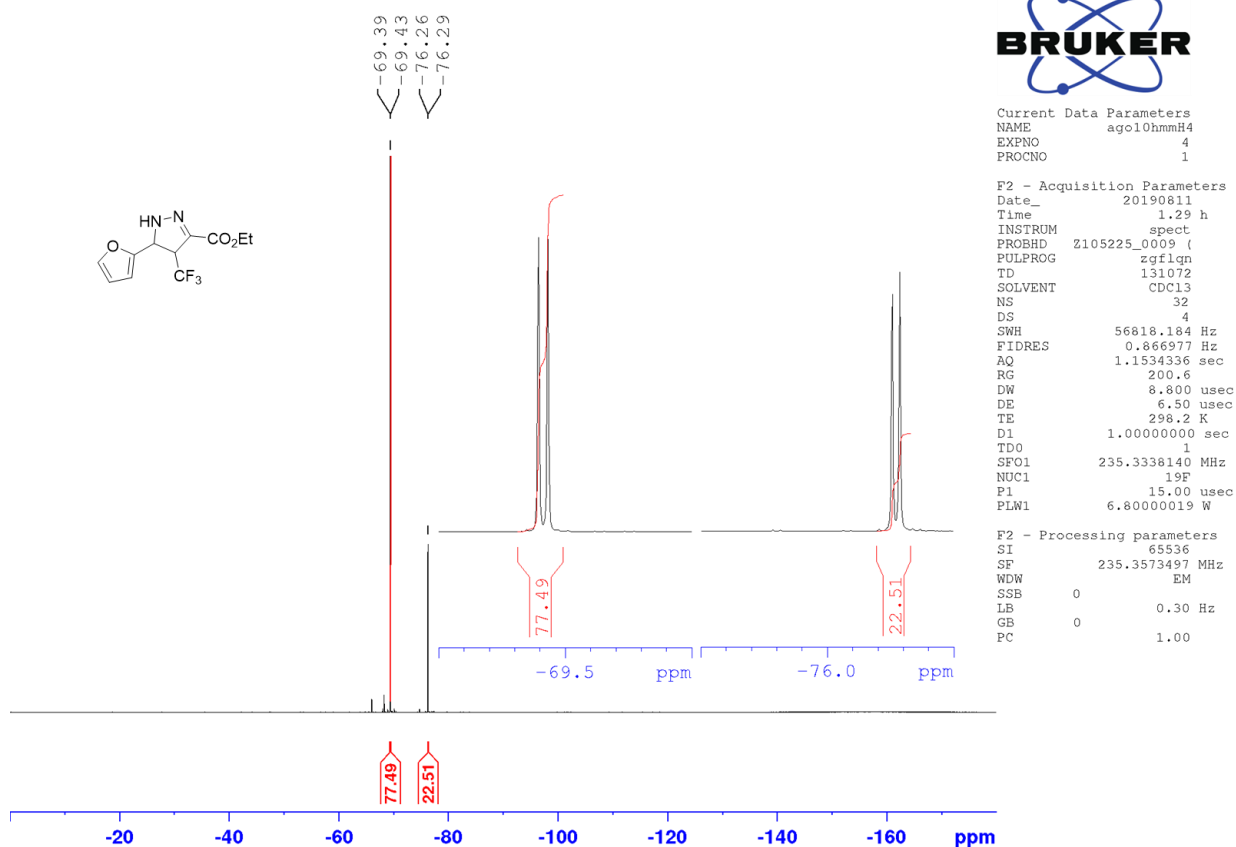


<sup>13</sup>C NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **10**

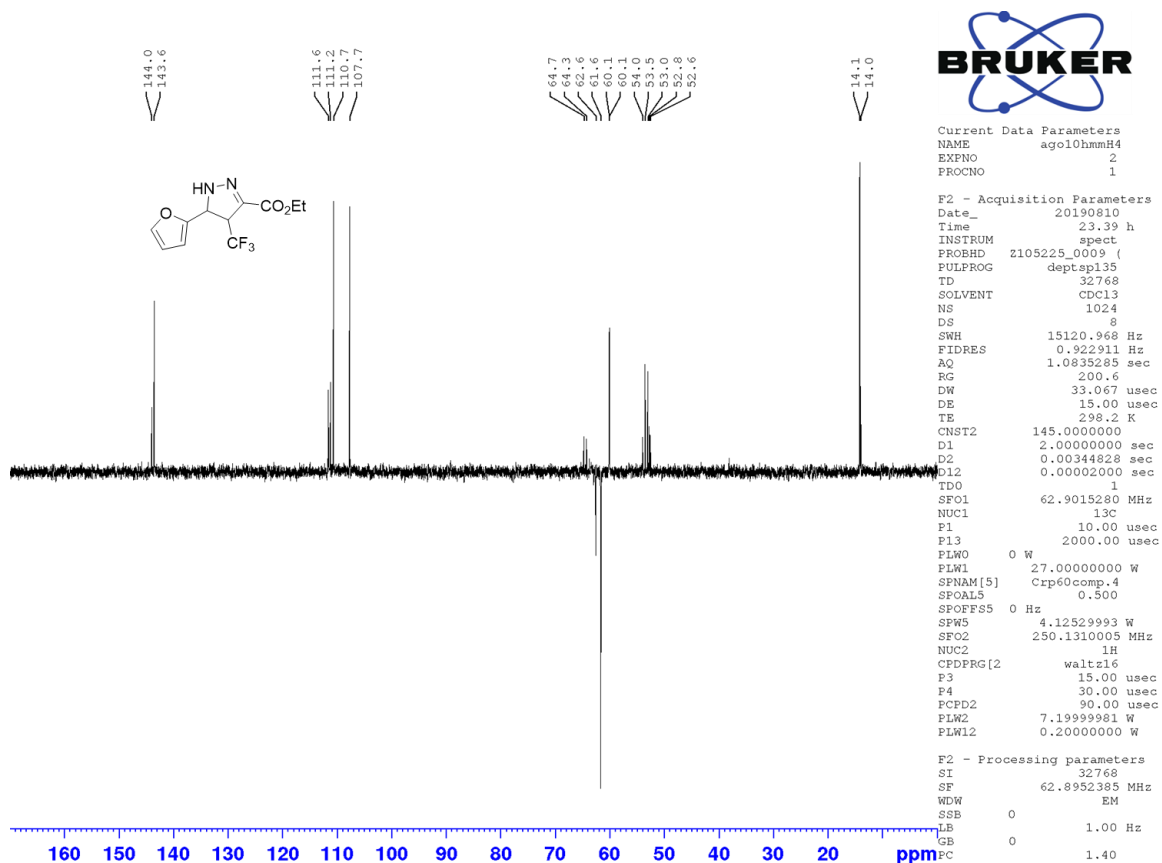
# **Ethyl 5-(furan-2-yl)-4-(trifluoromethyl)-4,5-dihydro-1H-pyrazole-3-carboxylate**



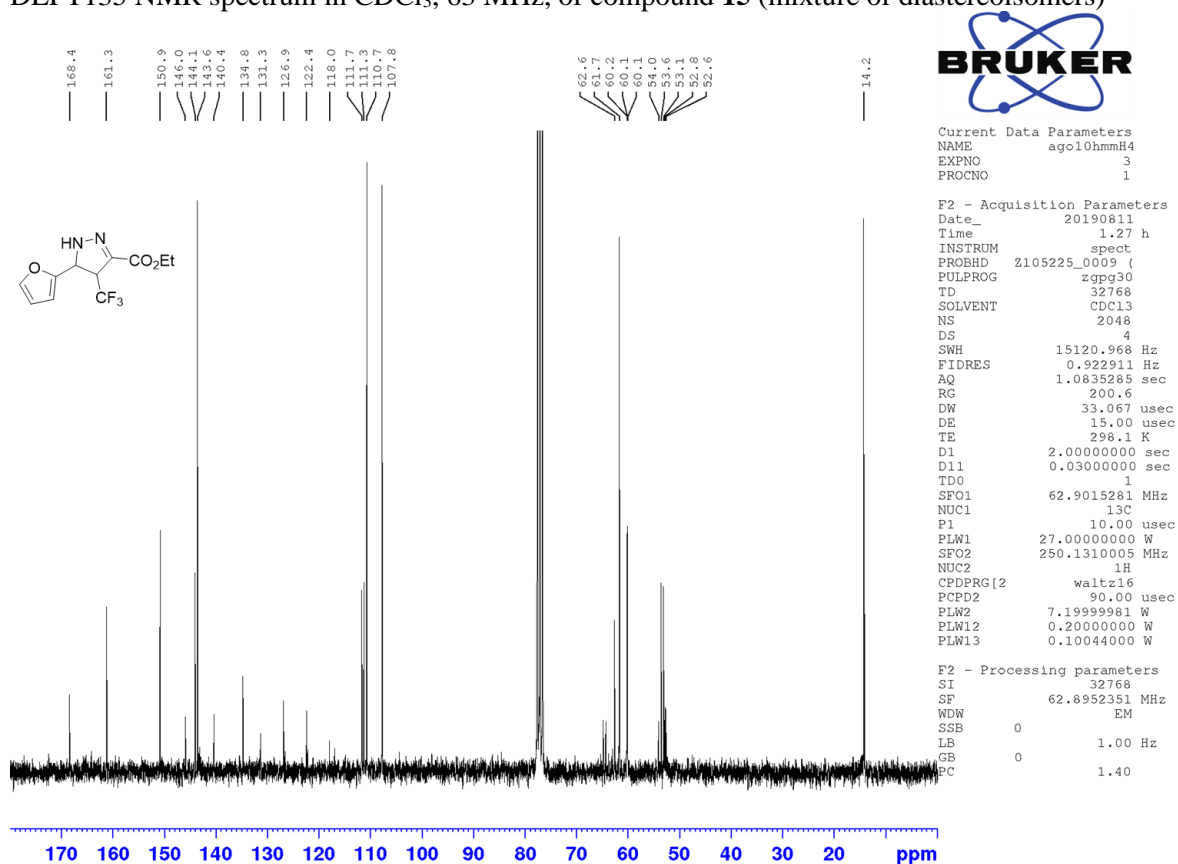
<sup>1</sup>H NMR spectrum in CDCl<sub>3</sub>, 250 MHz, of compound **15** (mixture of diastereoisomers)



<sup>19</sup>F NMR spectrum in CDCl<sub>3</sub>, 235 MHz, of compound **15** (mixture of diastereoisomers)

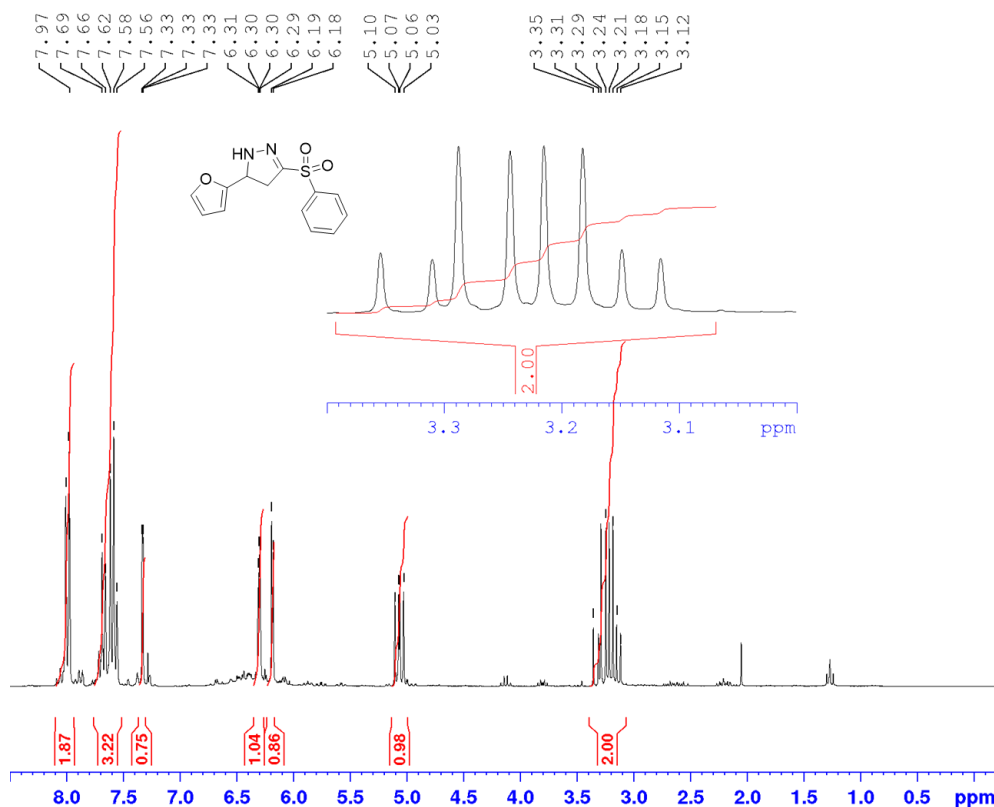


DEPT135 NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **15** (mixture of diastereoisomers)



<sup>13</sup>C NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **15** (mixture of diastereoisomers)

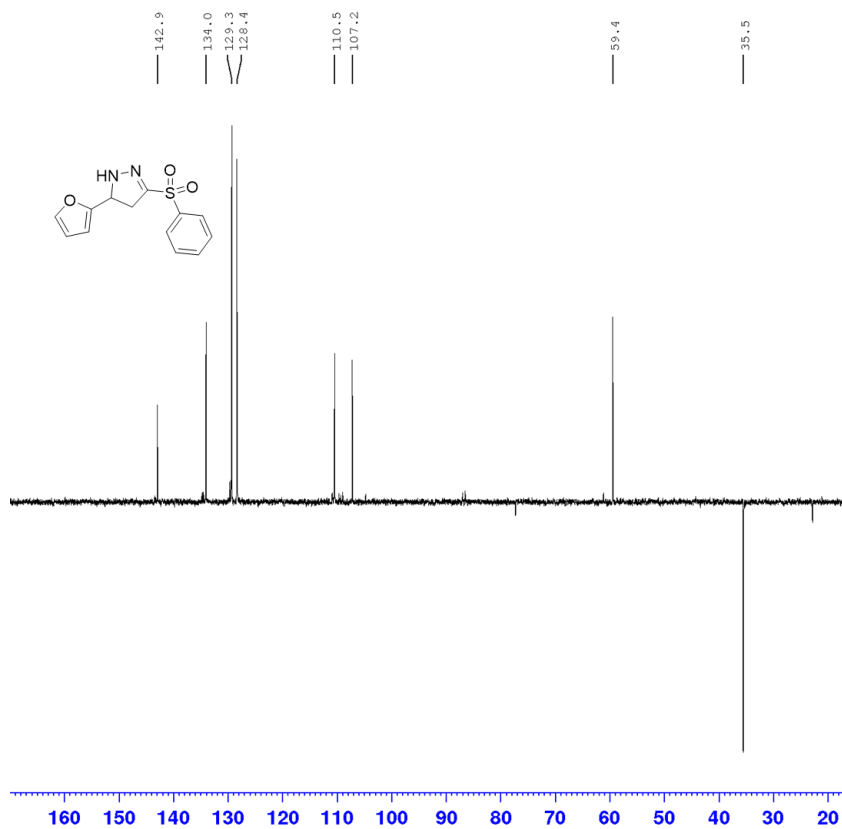
# 5-(furan-2-yl)-3-(phenylsulfonyl)-4,5-dihydro-1H-pyrazole



Current Data Parameters  
NAME agol0hmmH5  
EXPNO 1  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190811  
Time 1.35 h  
INSTRUM spect  
PROBHD Z105225\_0009 ( )  
PULPROG zg30  
TD 32768  
SOLVENT CDC13  
NS 32  
DS 0  
SWH 5000.000 Hz  
FIDRES 0.305176 Hz  
AQ 3.276799 sec  
RG 200.6  
DW 100.000 usec  
DE 6.50 usec  
TE 298.1 K  
D1 1.00000000 sec  
TD0 1  
SFO1 250.1315446 MHz  
NUC1 1H  
P1 15.00 usec  
PLW1 7.19999981 W

F2 - Processing parameters  
SI 55536  
SF 250.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

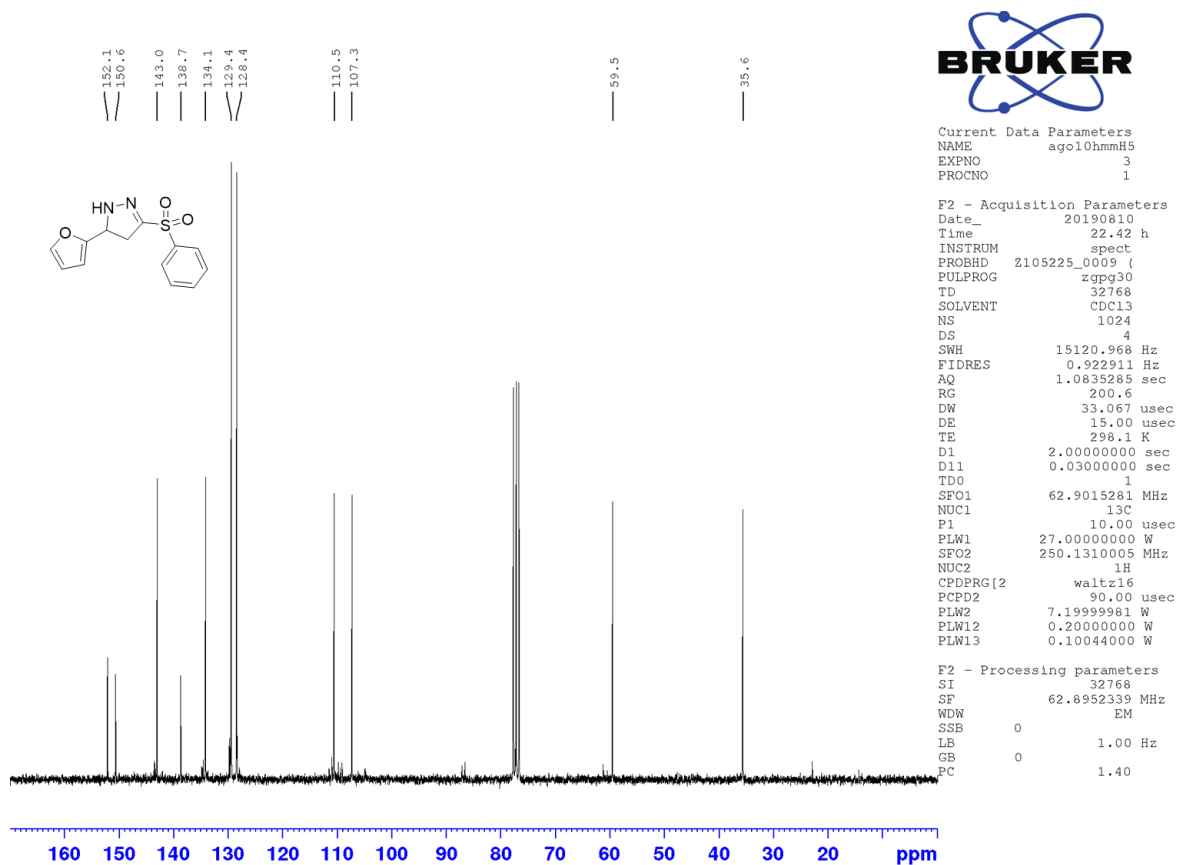


Current Data Parameters  
NAME agol0hmmH5  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190810  
Time 21.48 h  
INSTRUM spect  
PROBHD Z105225\_0009 ( )  
PULPROG deptsp135  
TD 32768  
SOLVENT CDC13  
NS 512  
DS 8  
SWH 15120.968 Hz  
FIDRES 0.922911 Hz  
AQ 1.0835285 sec  
RG 200.6  
DW 33.067 usec  
DE 15.00 usec  
TE 298.2 K  
CNST2 145.0000000  
D1 2.00000000 sec  
D2 0.00344828 sec  
D12 0.00002000 sec  
TD0 1  
SFO1 62.9015280 MHz  
NUC1 13C  
P1 10.00 usec  
PLW1 2000.00 usec

F13 0 W  
PLW1 27.00000000 W  
SPNAM[5] Crp60comp.4  
SFOAL5 0.500  
SPOFFS5 0 Hz  
SPW5 4.12529993 W  
SFO2 250.1310005 MHz  
NUC2 1H  
CFDPFRG2 waltz16  
P3 15.00 usec  
P4 30.00 usec  
PCPD2 90.00 usec  
PLW2 7.19999981 W  
PLW12 0.20000000 W

F2 - Processing parameters  
SI 32768  
SF 62.8952385 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 1.40



<sup>13</sup>C NMR spectrum in CDCl<sub>3</sub>, 63 MHz, of compound **20**

## References

- [1] N. M. Roda, D. N. Tran, C. Battilocchio, R. Labes, R. J. Ingham, J. M. Hawkins, S. V. Ley, *Org. Biomol. Chem.* **2015**, *13*, 2550–2554.