Supplementary Information

Bent-to-planar Si-rhodamines: a distinct rehybridization lights up NIR-II fluorescence for tracking nitric oxide in the Alzheimer's disease brain

Qingshuang Xu,^{‡a} Yutao Zhang,^{‡a} Mingming Zhu,^b Chenxu Yan,^a Wenle Mao,^a Wei-Hong Zhu^a and Zhiqian Guo^{*a}

^a Key Laboratory for Advanced Materials and Joint International Research Laboratory of Precision Chemistry and Molecular Engineering, Shanghai Key Laboratory of Functional Materials Chemistry, Frontiers Science Center for Materiobiology and Dynamic Chemistry, Frontiers Science Center for Materiobiology and Dynamic Chemistry, Institute of Fine Chemicals, School of Chemistry and Molecular Engineering, East China University of Science & Technology, Shanghai 200237, China.

E-mail: guozq@ecust.edu.cn

^b Division of Gastroenterology and Hepatology, Key Laboratory of Gastroenterology and Hepatology, Ministry of Health, Inflammatory Bowel Disease Research Center, Renji Hospital, School of Medicine, Shanghai Institute of Digestive Disease, Shanghai Jiao Tong University, Shanghai, China.

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1. Experimental Section

Materials and General Methods

Unless special stated, all solvents and chemicals were purchased from commercial suppliers in analytical grade and used without further purification. The ¹H and ¹³C NMR spectra were recorded on a Bruker AM 400 spectrometer, using TMS as an internal standard. High resolution mass spectrometry data were obtained with a Waters LCT Premier XE spectrometer. Absorption spectra were collected on a Varian Cary 500 spectrophotometer, and fluorescence spectra measurements were performed on a Varian Cary Eclipse fluorescence spectrophotometer and a PTI-QM4 steady-stead fluorimeter with a 75 W Xenon arc-lamp and a R928 PMT and an InGaAs photodetector. The time-dependent fluorescence measurements were conducted upon continuous illumination (Hamamatsu, LC8 Lightningcure, 300 W). The measurement of pH values in the context of buffer adjustments was performed on a SevenCompact pH-meter (Mettler Toledo). Femtosecond transient absorption (TA) spectra were collected using commercial femtosecond transient absorption spectrometer (Helios fire, Ultrafast System). In vitro PA images were scanned with a PA equipment (VEVO LAZR-X, Fujifilm VisualSonics, USA). IR images were monitored by a compact thermal imaging camera (FOTRIC 286). Confocal fluorescence images were taken on confocal laser scanning microscope (CLSM, Leica confocal microscope TCS SPS CFSMP and a Leica DMi8 microscope). Fluorescence signals in 96-well plates were recorded using a Series III 900/1700 animal imaging system (NIROPTICS, China). In vivo fluorescence images were measured with MARS-FAST in vivo imaging system (Artemis Intelligent Imaging, Shanghai, China). DEA·NONOate (Enzo Biochem, Inc.) was used as the source of NO, which is commercially available.⁵¹⁻⁵² DEA·NONOate Notes: Dissociates to the free amine and nitric oxide in a pH-dependent manner following first order kinetics. To initiate the release of nitric oxide, add stock alkaline solution of DEA NONOate to excess buffer of pH 7.0 - 7.4. The half life in 0.1 M phosphate buffer, pH 7.4, is $t\frac{1}{2}$ = 16 min. at 22 -25 °C or $t\frac{1}{2}$ = 2 min. at 37 °C. Liberates 1.5 mol of NO per mol of parent compound. Decomposition is nearly instantaneous at pH 5.0.

DCM = dichloromethane

DMSO = dimethylsulfoxide EtOH = ethanol MeCN = acetonitrile MeQH = methanol

THF = tetrahydrofuran

Spectral measurement details of SiRhd probes

B-SiRhd-3 (λ_{ex} = 550 nm) solution (10 µM, including Hg²⁺) in PBS (0.01 M, pH 7.4) was placed into the wells of a quartz cuvette. Absorbance/FL intensities were continuously acquired using Varian Cary 500 spectrophotometer and Varian Cary Eclipse fluorescence spectrophotometer. Absorbance and fluorescence emission spectra (λ_{ex} = 550 nm) of *B*-SiRhd-4 (10 µM) in the presence of phosgene in ethonal; absorbance and fluorescence emission spectra (λ_{ex} = 550 nm) of *P*-SiRhd-1 (10 µM) in the presence of palladium (Pd(PPh₃)₄ in a mixed solution of CH₃CN/PBS (v/v = 1/3). Absorbance

and fluorescence emission spectra (λ_{ex} = 420 nm, λ_{ex} = 808 nm) of *B*-SiRhd-7 (10 µM) in the presence of NO in a mixed solution of CH₃CN/HEPES (v/v =1/1, 0.02 M, pH 7.4).

Photophysical data

The fluorescence quantum yield (ϕ) was measured following the previous method.⁵³ Briefly, ϕ was calculated by using the following equation:

 $\Phi_{s} = \Phi_{r} (A_{r} F_{s} / A_{s} F_{r}) (n_{s}^{2} / n_{r}^{2})$

Where, s and r denote sample and reference, respectively. A is the absorbance. F is the relative integrated fluorescence intensity and n is the refractive index of the solvent. B-SiRhd-1, P-SiRhd-1, B-SiRhd-4, B-SiRhd-3 and P-SiRhd-3 were measured by using rhodamine 6G (Φ = 88% in ethanol) as a standard.⁵⁴ P-SiRhd-7 were measured by using IR-26 (Φ = 0.05% in dichloroethane) as a standard.⁵⁵

Cell Experiment

Cell Lines

The human epithelioid cervical carcinoma cell line HeLa and the mouse macrophage cell line RAW 264.7 were purchased from the Institute of Cell Biology (Shanghai, China). Cells were all propagated in T-75 flasks cultured at 37 ^oC under a humidified 5% CO₂ atmosphere in DMEM medium (GIBCO/Invitrogen, Camarillo, CA, USA), which were supplemented with 10 % fetal bovine serum (FBS, Biological Industry, Kibbutz Beit Haemek, Israel) and 1 % penicillin-streptomycin (10,000 U mL⁻¹ penicillin and 10 mg mL⁻¹ streptomycin, Solarbio life science, Beijing, China).

In vitro Cellular Imaging

The cells at 1×10^5 cells/well were seeded onto glass-bottom petri dishes with complete medium (1.5 mL) for 12 h. The cells pro-incubated with 10 μ M *B*-SiRhd-3 or *B*-SiRhd-7 for 0.5 h. Then the cells incubated with and without inducer. PBS was used to wash cells for three times to clean the background. The images were then photographed by using a confocal laser scanning microscope Leica TCS SP8 (63 × oil lens).

Animals

All animal studies were approved by the Animal Care and Use Committee of East China Normal University in accordance with the guidelines for the care and use of laboratory animals. The 6-week-old female BALB/cA nude mice were produced from Shanghai SLAC Laboratory Animal Co. Ltd., and maintained under standard conditions. The animals were housed in sterile cages within laminar airflow hoods in a specific pathogen-free room with a 12-h light/12-h dark schedule and fed autoclaved chow and water *ad* libitum. Number of qualitative qualification: No.20220004001694. Production Permit No.: SCXK (Shanghai) 2022-0004. The 12-month-old male APP/PS1 transgenic mice and wild-type mice were produced from Jiangsu Huachuang Sino Pharmatech Co., Ltd. (Jiangsu, China), and

maintained under standard conditions. Number of qualitative qualification: No.20220168. Production Permit No.: SCXK (Jiangsu) 2020-0009.

In vivo NIR II fluorescence imaging

Mice were treated with either LPS (1 mg/mL, 4 mg/Kg) or a saline (as control group) intraperitoneally. After 6 h, the B-SiRhd-7@liposome (0.4 mM, 200 μL) were tail intravenous injected into mice for in vivo imaging. NIR-II images of mice anesthetized with isoflurane were recorded at various time points post-injection through MARS-FAST in vivo imaging system (Artemis Intelligent Imaging, Shanghai, China). The excitation wavelength was 808 nm, and the NIR-II fluorescence signal was collected with 1100 nm LP filters (Thorlabs).

Synthesis of Intermediate Compound 1-7 and *B*-SiRhd-1, *P*-SiRhd-1, *P*-SiRhd-2, *B*-SiRhd-3, *B*-SiRhd-4, *B*-SiRhd-5, *B*-SiRhd-6 and *B*-SiRhd-7

The compound **O-SiRhd**^{S6} was synthesized by the established procedures from our group.



Scheme S1. Synthetic route of compound Cl-SiRhd, B-SiRhd-1, P-SiRhd-1, P-SiRhd-2, B-SiRhd-3, B-SiRhd-4, B-SiRhd-5.



Scheme S2. Synthetic route of compound Cl-SiRhd-II, B-SiRhd-6 and B-SiRhd-7.

Synthesis of compound Cl-SiRhd

A solution of compound **O-SiRhd** (100 mg, 0.26 mmol) in CH₃CN (10 mL) was stirred at 0 °C under N₂ for 10 min, and then (COCl)₂ (0.10 mL) was added dropwise over 1 min. The reaction mixture was stirred for 10 min. The completion of reaction was monitored by TLC. After completion of the reaction, the solvents were removed under reduced pressure and the residue was purified by silica gel chromatography using dichloromethane/methanol (v/v, 20:1) as the eluent to afford **Cl-SiRhd** (30 mg): Yield 34%. ¹H-NMR (400 MHz, CD₃CN, ppm): δ 0.63 (s, 6H, -SiCH₃), 1.2 (t, *J* = 7.2 Hz, 12H, - CH₂CH₃), 3.69 (q, *J* = 7.2 Hz, 8H, -CH₂CH₃), 7.87 (dd, *J*₁ = 8.8 Hz, *J*₂ = 2.4 Hz, 2H, Ph-H), 8.14 (d, *J* = 2.4 Hz, 2H, Ph-H), 8.54 (d, *J* = 8.4 Hz, 2H, Ph-H). High resolution mass spectrometry (ESI-MS, m/z): [M]⁺ Calc for C₂₃H₃₂N₂SiCl, 399.2023, Found, 399.2025.

Synthesis of compound *B*-SiRhd-1

A solution of compound **O-SiRhd** (100 mg, 0.26 mmol) in CH₃CN (10 mL) was stirred at 0 °C under N₂ for 10 min, and then (COCl)₂ (0.10 mL) was added dropwise over 1 min. The reaction mixture was stirred for 10 min. The solvents were removed under reduced pressure and the crude compound was used without further purification. To a solution crude compound in CH₃CN (10 mL) at 0 °C was added phenyl n-propylamine (23.30 mg, 0.39 mmol). The mixture was stirred overnight at 25 °C. The completion of reaction was monitored by TLC. After completion of the reaction, the solvents were removed under reduced pressure and the residue was purified by silica gel chromatography using dichloromethane/methanol (v/v, 30:1) as the eluent to afford **B-SiRhd-1** as a red solid (30 mg): Yield 27%. ¹H-NMR (400 MHz, DMSO-*d*₆, ppm): δ 0.49 (s, 6H, -SiCH₃), 0.82 (t, *J* = 7.2 Hz, 3H, -CH₂CH₃), 1.14 (q, *J* = 6.4 Hz, 12H, -CH₂CH₃), 1.79 (q, *J* = 7.2 Hz, 2H, -CH₂CH₂CH₃), 3.50 (t, 8H, -CH₂CH₃), 3.85 (q, *J* = 6.4 Hz, 2H, -CH₂CH₂CH₃), 6.82-6.88 (m, 2H, Ph-H), 6.98 (d, *J* = 2.4 Hz, 1H, Ph-H), 7.06 (d, *J* = 2.4 Hz, 1H, Ph-H), 7.62 (d, *J* = 9.2 Hz, 1H, Ph-H), 7.79 (d, *J* = 9.2 Hz, 1H, Ph-H). High resolution mass spectrometry (ESI-MS, m/z): [M]⁺ Calc for C₂₆H₄₀N₃Si, 422.2992, Found, 422.2997.

Synthesis of compound *B*-SiRhd-2

A solution of compound **O-SiRhd** (100 mg, 0.26 mmol) in CH₃CN (10 mL) was stirred at 0 °C under N₂ for 10 min then (COCl)₂ (0.10 mL) was added dropwise over 1 min. After stirring 10 min of the reaction mixture, the solvents were removed under reduced pressure and the crude compound was used without further purification. To a solution crude compound in CH₃CN (10 mL) at 0 °C was added phenyl methylamine (0.10 mL). The mixture was stirred overnight at 25 °C. The completion of reaction was monitored by TLC. After completion of the reaction, the solvents were removed under reduced pressure and the residue was purified by silica gel chromatography using dichloromethane/methanol (v/v, 30:1) as the eluent to afford **B-SiRhd-2** as an orange solid (30 mg): Yield 30%. ¹H-NMR (400 MHz, CDCl₃, ppm): δ 0.46 (s, 6H, -SiCH₃), 1.18-1.26 (m, 12H, -CH₂CH₃), 3.42-3.51 (m, 8H, -CH₂CH₃), 3.67 (s, 3H, -NCH₃), 6.70-6.73 (m, 1H, Ph-H), 6.76 (d, *J* = 2.8 Hz, 1H, Ph-H), 6.88-6.91 (m, 2H, Ph-H), 7.57 (d, *J* = 8.8 Hz, 1H, Ph-H), 8.45 (d, *J* = 9.2 Hz, 1H, Ph-H). High resolution mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₂₄H₃₆N₃Si, 394.2679, Found, 394.2679.

Synthesis of compound *B*-SiRhd-3

A solution of compound **O-SiRhd** (100 mg, 0.26 mmol) in CH₃CN (10 mL) was stirred at 0 °C under N₂ for 10 min then (COCI)₂ (0.10 mL) was added dropwise over 1 min. The reaction mixture was stirred for 10 min then ethylenediamine (0.35 mL, 5.25 mmol) was added. The mixture was stirred 30 min at room temperature. The solvents were removed under reduced pressure and the crude compound was used without further purification. To a solution crude compound in ethanol (10 mL) at 0 °C was added phenyl isothiocyanate (0.10 mL). The mixture was stirred overnight at 25 °C. The completion of reaction was monitored by TLC. After completion of the reaction, the solvents were removed under reduced pressure and the residue was purified by silica gel chromatography using dichloromethane/methanol (v/v, 20:1) as the eluent to afford **B-SiRhd-3** as a red solid (12 mg): Yield 8.2%. ¹H-NMR (400 MHz, DMSO-*d*₆, ppm): δ 0.46 (s, 6H, -SiCH₃), 1.14 (t, 12H, *J* = 6.8 Hz, -CH₂CH₃), 3.49 (d, 8H, *J* = 6.8 Hz, -CH₂CH₃), 4.05 (d, 4H, -CH₂-), 6.79-6.88 (m, 2H, Ph-H), 6.97 (s, 1H, Ph-H), 7.04 (s, 1H, Ph-H), 7.10 (d, 1H, Ph-H), 7.24 (s, 4H, Ph-H), 7.66 (s, 1H, Ph-H), 7.83 (s, 1H, Ph-H), 8.03 (s, 1H, -NH-), 9.93 (s, 1H, -NH-). ¹³C NMR (100 MHz, DMSO-*d*₆, ppm): δ = -2.57, 12.32, 43.13, 43.86, 123.21, 124.27, 124.60, 128.63, 139.02, 180.57. Mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₃₂H₄₄N₅SSi, 558.3087, Found, 558.3079.

Synthesis of compound B-SiRhd-4

A solution of compound **O-SiRhd** (100 mg, 0.26 mmol) in CH₃CN (10 mL) was stirred at 0 °C under N₂ for 10 min then (COCl)₂ (0.10 mL) was added dropwise over 1 min. After stirring 10 min of the reaction mixture, the solvents were removed under reduced pressure and the crude compound was used without further purification. To a solution crude compound in DMF (30 mL) was added 4-aminobutyric acid (41 mg, 0.39 mmol). The mixture was heated to reflux in an argon atmosphere for 5 h. The completion of reaction was monitored by TLC. After completion of the reaction, the solvents were removed under reduced pressure and the residue was purified by silica gel chromatography using dichloromethane/methanol (v/v, 20:1) as the eluent to afford **B-SiRhd-4** as a red solid (30 mg): Yield 27%. ¹H-NMR (400 MHz, DMSO- d_6 , ppm): δ 0.43 (s, 6H, -SiCH₃), 1.12 (s, 12H, -CH₂CH₃), 1.85 (s, 2H, -CH₂CH₂CC₂CH₂COOH), 3.41 (s, 8H, -CH₂CH₃), 3.77 (s, 2H, -CH₂CH₂CH₂COOH), 6.72 (s, 2H, Ph-H), 6.83 (s, 1H, Ph-H), 6.94 (s, 1H, Ph-H), 7.41 (d, *J* = 8.4 Hz, 1H, Ph-H), 7.66 (d, *J* = 8 Hz, 1H, Ph-H). Mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₂₇H₄₀N₃O₂Si, 466.2890, Found, 466.2885.

Synthesis of compound B-SiRhd-5

A solution of compound **O-SiRhd** (100 mg, 0.26 mmol) in CH₃CN (10 mL) was stirred at 0 °C under N₂ for 10 min then (COCl)₂ (0.10 mL) was added dropwise over 1 min. After stirring10 min of the reaction mixture, the solvents were removed under reduced pressure and the crude compound was used without further purification. To a solution crude compound in CH₃CN (10 mL) at 0 °C was added *o*-phenylenediamine (0.32 g, 3 mmol). The mixture was stirred overnight at 25 °C. After completion of the reaction, the solvents were removed under reduced pressure and the residue was purified by silica gel chromatography using dichloromethane/methanol (v/v, 30:1) as the eluent to afford *B*-SiRhd -5 as a red solid (30 mg): Yield 30%. ¹H-NMR (400 Hz, CDCl₃, ppm): δ 0.51(s, 6H, -SiCH₃), 1.18 (t, *J* = 7.2 Hz, 12H, -CH₂CH₃), 3.41(q, *J* = 7.2 Hz, 8H, -CH₂CH₃), 6.56-6.58 (m, 2H, Ph-H), 6.81-6.82 (m, 3H, Ph-H), 7.02-7.12 (m, 3H, Ph-H),

7.73 (s, 2H, Ph-H). Mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₂₉H₃₉N₄Si, 471.2944, Found, 471.2946.

Synthesis of compound P-SiRhd-1

DBU (94 mg, 0.62mmol) was added dropwise to the solution of compound *B*-SiRhd-1 (130 mg, 0.31 mmol) in CH₃CN (10 mL). Then allyl chloroformate (75 mg, 0.62 mmol) was added over 1 min at 0 °C. The reaction mixture was stirred for 2h at room temperature. The completion of reaction was monitored by TLC. After completion of the reaction, CH_2CI_2 (100 mL) was added to the mixture, the combined organic layer was washed with brine. The combined organic layer was dried over Na_2SO_4 and evaporated to dryness. The residue was purified by silica gel chromatography using dichloromethane/methanol (v/v, 30:1) as the eluent to afford *P*-SiRhd-1 as a green solid (43 mg): Yield 35.9%. ¹H-NMR (400 MHz, DMSO-*d*₆, ppm): δ 0.55 (d, *J* = 10.8 Hz, 6H, -SiCH₃), 0.80-0.84 (m, 3H, -CH₂CH₂CH₃), 1.21-1.23 (t, 12H, *J* = 7.2 Hz, -CH₂CH₃), 1.54-1.60 (m, 2H, -CH₂CH₂CH₃), 3.52-3.56 (m, 2H, -CH₂CH₂CH₃), 3.72-3.74 (m, 8H, -CH₂CH₃), 4.49 (q, 2H, *J* = 1.6 Hz, -COOCH₂-), 4.68 (d, 1H, *J* = 5.2 Hz, -OCH₂CH=CH₂), 4.90-5.02 (m, 2H, -OCH₂CH=CH₂), 7.06 (dd, *J*₁ = 9.6 Hz, *J*₂ = 2.8 Hz, 2H, Ph-H), 7.38 (d, *J* = 2.4 Hz, 2H, Ph-H), 7.59-7.62 (d, 2H, Ph-H). High resolution mass spectrometry (ESI-MS, m/z): [M]⁺ Calc for C₃₀H₄₄N₃O2Si, 506.3205, Found, 506.3205.

Synthesis of compound *P*-SiRhd-2

Triethylamine (63 mg, 0.62 mmol) was added dropwise to the solution of compound *B*-SiRhd-1 (130 mg, 0.31 mmol) in CH₃CN (10 mL). Then acryloyl chloride (57 mg, 0.62 mmol) was added over 1 min at 0 °C. The reaction mixture was stirred for 2h at room temperature. The completion of reaction was monitored by TLC. After completion of the reaction, CH_2Cl_2 (100 mL) was added to the mixture, the combined organic layer was washed with brine. The combined organic layer was dried over Na₂SO₄ and evaporated to dryness. The residue was purified by silica gel chromatography using dichloromethane/methanol (v/v, 20:1) as the eluent to afford *P*-SiRhd-2 as a green solid (66 mg): Yield 58.5%. ¹H-NMR (400 MHz, DMSO-*d*₆, ppm): δ 0.56 (s, 3H, -SiCH₃), 0.62 (s, 3H, -SiCH₃), 0.82 (t, *J* = 7.2 Hz, 3H, -CH₂CH₂CH₃), 1.22 (t, *J* = 7.2 Hz, 12H, -CH₂CH₃), 1.52-1.58 (m, 2H, -CH₂CH₂CH₃), 3.05-3.08 (m, 2H, -CH₂CH₂CH₃), 3.73 (s, 8H, -CH₂CH₂CH₃), 5.62-5.65 (m, 1H, -CH=CH₂), 5.96-6.03 (m, 1H, -CH=CH₂), 6.26-6.31 (m, 1H, -CH=CH₂), 7.04-7.08 (m, 2H, Ph-H), 7.42 (d, *J* = 2.4 Hz, 2H, Ph-H), 7.49 (d, 2H, Ph-H). High resolution mass spectrometry (ESI-MS, m/z): [M]⁺ Calc for C₂₉H₄₂N₃OSi, 476.3097, Found, 476.3092.

Synthesis of compound 1

3-bromoaniline (2.00 g, 11.70 mmol), Etl (7.30 g, 46.80 mmol), and K₂CO₃ (1.60 g, 11.70 mmol) were dissolved in anhydrous CH₃CN (50 mL). The mixture was heated to reflux in an argon atmosphere for 11 h. The completion of reaction was monitored by TLC. After completion, the reaction mixture was cooling to room temperature, the mixture was filtered. The organic layer was concentrated under vacuum, and then the crude product was purified by silica gel chromatography using dichloromethane/petroleum ether (v/v, 1:10) as the eluent to afford **compound 1** as a colorless liquid (1.6 g): Yield 60.3%. ¹H NMR (400 MHz, CDCl₃, ppm): δ 1.15 (t, 6H, *J* = 7.2 Hz, -CH₂CH₃), 3.32 (q, 4H, *J* = 7.2 Hz, -CH₂CH₃), 6.57 (dd, *J*₁ = 8.4 Hz, *J*₂ = 2.4 Hz, 1H, Ph-H), 6.72-6.77 (m, 2H, Ph-H), 7.03 (t, *J* = 8 Hz, 1H, Ph-H). ¹³C NMR (100

MHz, CDCl₃, ppm): δ = 12.57, 44.44, 110.26, 114.26, 117.95, 123.74, 130.56, 149.04. Mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₁₀H₁₅NBr, 228.0388, Found, 228.0380.

Synthesis of compound 2

0.25 mL POCl₃ was added dropwise in anhydrous DMF (5 mL) in 0 °C and the reaction mixture was stirred at room temperature for 30 min. **Compound 1** (1.00 g, 4.40 mmol) was added in the reaction mixture. The mixture was stirred in an argon atmosphere at room temperature for 6 h. The reaction mixture was poured into 200 mL cool water. The mixture was filtered. The residue was washed by water for three times to afford **compound 2** as a white solid. Yield 41.24%. ¹H-NMR (400 MHz, CDCl₃, ppm): δ 1.22 (t, *J* = 7.2 Hz, 6H, -CH₂CH₃), 3.42 (q, *J* = 7.2 Hz, 4H, -CH₂CH₃), 6.59-6.62 (dd, *J*₁ = 8.8 Hz, *J*₂ = 2.4 Hz, 1H, Ph-H), 6.76 (d, *J* = 2.4 Hz, 1H, Ph-H), 7.78 (d, *J* = 8.8 Hz, 1H, Ph-H), 10.05 (d, *J* = 0.4 Hz, 1H, -CHO). ¹³C NMR (100 MHz, CDCl₃, ppm): δ = 12.43, 44.81, 110.26, 114.32, 121.50, 131.32, 152.50, 190.03. Mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₁₁H₁₅NOBr, 256.0337, Found, 256.0334.

Synthesis of compound 3

Compound 2 (300 mg, 1.18 mmol) was dissolved in methanol (6.00 mL), and then sodium borohydride (67.11 mg, 1.76 mmol) was added into the solution. The reaction mixture was stirred for 2 h at room temperature. H₂O was added to the residue, and the aqueous layer was extracted with CH₂Cl₂. The combined organic layer was washed with brine, dried over Na₂SO₄ and evaporated to dryness. The residue was purified by column chromatography using dichloromethane/petroleum ether (v/v, 1:2) as the eluent to afford **compound 3** as a colorless liquid (250 mg): Yield 83.36%. ¹H-NMR (400 MHz, CDCl₃, ppm): δ 1.15 (t, *J* = 7.2 Hz, 6H, -CH₂CH₃), 1.81 (s, 1H, -CH₂OH), 3.33 (q, *J* = 7.2 Hz, 4H, -CH₂CH₃), 4.62 (s, 2H, -CH₂OH), 6.58 (dd, *J*₁ = 8.4 Hz, *J*₂ = 2.4 Hz, 1H, Ph-H), 6.83 (d, *J* = 2.8 Hz, 1H, Ph-H), 7.21 (d, *J* = 8.8 Hz, 1H, Ph-H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ = 12.46, 44.43, 64.98, 110.75, 115.17, 124.75, 125.97, 130.69, 148.48. Mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₁₁H₁₇NOBr, 258.0494, Found, 258.0494.

Synthesis of compound 4

1,2-Cyclohexanedione (500 mg, 4.46 mmol) and 4-Bromobenzene-1,2-diamine (1.00 g, 5.53 mmol) was dissolved in acetonitrile. The reaction mixture was then stirred at 80 °C for 8 h. The completion of reaction was monitored by TLC. After completion, the reaction mixture was cooling to room temperature. The solvent was removed in a rotary evaporator and crude product was purified by column chromatography using ethanldan/petroleum ether (v/v, 1:4) as the eluent to afford **compound 4** as a pale yellow solid (890 mg): Yield 76.23%. ¹H-NMR (400 MHz, CDCl₃, ppm): δ 2.03-2.06 (m, 4H, -CH₂(CH₂)₂CH₂-), 3.15 (dd, J_1 = 4.8 Hz, J_2 = 2.4 Hz, 4H, -CH₂(CH₂)₂CH₂-), 7.73 (dd, J_1 = 8 Hz, J_2 = 2 Hz, 1H, Ph-H), 7.83 (d, J = 8 Hz ,1H, Ph-H), 8.15 (d, J = 2 Hz, 1H, Ph-H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ = 22.65, 33.19, 122.66, 129.68, 130.67, 132.47, 139.87, 141.74, 154.62, 155.21. Mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₁₂H₁₂N₂Br, 263.0184, Found, 263.0181.

Synthesis of compound 5

Compound 4 (0.45 g, 1.70 mmol) was dissolved in dry toluene (80 mL) and cooled to 0 °C. To this cold solution was added sodium borohydride (0.65 g, 17.00 mmol) over a period of 15 min. The pale yellow slurry thus obtained was stirred for 10 min. Glacial acetic acid (3.40 mL, 0.60 mmol) was added to it drop wise over a period of 1 h maintaining the temperature 5-10 °C. The resulting brown slurry was stirred for another 1 h at 10 °C and allowed to attain room temperature. It was then heated to gentle reflux for 5 h. On cooling, a thick red resinous mass was obtained. To this red resinous mass water (250 mL) was added. The toluene layer formed was separated and aqueous layer was extracted with ethyl acetate (3 × 100 mL). Combined extracts and toluene layer were washed repeatedly with dilute sodium carbonate solution and then with water, dried over anhydrous sodium sulphate, filtered and vacuum evaporated. The dark brown oil obtained was purified by column chromatography using dichloromethane/petroleum ether (v/v, 1:10) as the eluent to afford **compound 5** as a pale yellow oil (0.17 g): Yield 31.05%. ¹H-NMR (400 MHz, CDCl₃, ppm): δ 1.09-1.16 (m, 6H, -CH₂CH₃), 1.34-1.38 (m, 2H, -CH₂-), 1.54-1.57 (m, 4H, -CH₂-), 1.79-1.86 (m, 2H, -CH₂-), 3.11-3.44 (m, 8H, -CH₂CH₃, -NCHCH₂-), 6.39 (d, *J* = 8.8 Hz, 1H, Ph-H), 6.60 (d, *J* = 2 Hz, 1H, Ph-H), 6.67 (dd, *J*₁ = 8.4 Hz, *J*₂ = 2 Hz, 1H, Ph-H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ = 11.53, 22.39, 27.00, 27.63, 41.92, 54.65, 55.35, 109.54, 112.06, 113.16, 119.28, 133.99, 136.47. Mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₁₆H₂₄N₂Br, 322.1123, Found, 322.1120.

Synthesis of compound 6

Compound 3 (1.95 g, 7.58 mmol) and **compound 5** (2.29 g, 7.11 mmol) were dissolved in methanol (6.00 mL), and then BF₃·OEt₂ (1.92 mL, 15.16 mmol) was driped into the solution. The reaction mixture was stirred for 24 h at room temperature, diluted with water and extracted with CH₂Cl₂. The organic layer was dried over Na₂SO₄ and evaporated to dryness. The residue was purified by column chromatography using dichloromethane/petroleum ether (v/v, 1:10) as the eluent to afford **compound 6** as a pale yellow oil (1.50 g): Yield 37.6%. ¹H-NMR (400 MHz, CDCl₃, ppm): δ 0.99-1.03 (m, 3H, -CH₂CH₃), 1.11-1.16 (m, 9H, -CH₂CH₃), 1.32-1.37 (m, 2H, -CH₂-), 1.50-1.57 (m, 4H, -CH₂-), 1.79-1.84 (m, 2H, -CH₂-), 3.01-3.42 (m, 10H, -NCH₂CH₃, -NCHCH₂-), 3.94 (s, 2H, -PhCH₂Ph-), 6.28 (s, 1H, Ph-H), 6.52 (dd, *J*₁ = 8.8 Hz, *J*₂ = 2.8 Hz, 1H, Ph-H), 6.67 (s, 1H, Ph-H), 6.86-6.88 (m, 2H, Ph-H). ¹³C NMR (100 MHz, CDCl₃, ppm): δ = 11.42, 12.49, 22.26, 27.47, 40.16, 41.97, 44.42, 55.11, 111.40, 113.25, 113.98, 135.38, 125.66, 126.61, 127.26, 130.83, 134.45, 134.81, 147.16. Mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₂₇H₃₈N₃Br₂, 562.1432, Found, 562.1435.

Synthesis of O-SiRhd-II

To a dried flask flushed with argon, **compound 6** (1.50 g, 2.67 mmol) and anhydrous THF (17 mL) were added. The solution was cooled to -78 °C, 2.4 M n-BuLi (3.20 mL, 8.01 mmol) was added, and the mixture was stirred for 0.5 h. At the same temperature, a solution of $SiMe_2Cl_2$ (3.47 mmol) in anhydrous THF (10 mL) was slowly added, and the mixture was slowly warmed to room temperature, then stirred for 8 h. After completion, the reaction was quenched by addition of saturated NH₄Cl aq and extracted with CH₂Cl₂. The organic layer was washed with brine and dried over

Na₂SO₄. The solvent was removed under reduced pressure and the crude compound was used without further purification. To a solution crude compound in acetone (50 mL) at 0 °C was added KMnO₄ (1.26 g, 8.01 mmol) in small portions over a period of 1 h with stirring. The mixture was stirred for another 1 h at the same temperature, then diluted with CH₂Cl₂ (50 mL), filtered through paper filter and evaporated to dryness. The residue was purified by column chromatography using dichloromethane/petroleum ether (v/v, 2:1) as the eluent to afford **O-SiRhd-II** as a yellow solid (10 mg): Yield 1%. ¹H-NMR (400 MHz, DMSO-*d*₆, ppm): δ 0.38 (d, *J* = 6 Hz, 6 H, -SiCH₃), 1.08-1.16 (m, 12H, - CH₂CH₃), 1.34-1.38 (m, 2H, -CH₂-), 1.47-1.48 (m, 2H, -CH₂-), 1.56-1.61 (m, 2H, -CH₂-), 1.72-1.82 (m, 2H, -CH₂-), 3.20-3.64 (m, 10H, -NCH₂CH₃, -NCHCH₂-), 6.68 (s, 1H, Ph-H), 6.80 (d, *J* = 9.2 Hz, 2H, Ph-H), 7.48 (s, 1H, Ph-H), 8.08 (d, *J* = 8.4 Hz, 1H, Ph-H). ¹³C NMR (100 MHz, DMSO-*d*₆, ppm): δ = 0.00, 12.30, 13.52, 44.68, 111.43, 128.25, 129.03, 131.01, 136.22, 138.76, 141.60, 149.61, 184.54. Mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₂₉H₄₂N₃OSi, 476.3097, Found, 476.3097.

Synthesis of Cl-SiRhd-II

The mixture of **O-SiRhd-II** (100 mg, 0.21 mmol) in CH₃CN (10 mL) was stirred at 0 °C under N₂ for 10 min then (COCI)₂ (0.1 mL) was added dropwise over 1 min. The mixture was stirred 30 min at room temperature. After completion, the solvents were removed under reduced pressure and the residue was purified by HPLC (eluent: CH₃CN/H₂O = 100/0, 14 min) to afford the pure product **CI-SiRhd-II** (60 mg): Yield 58%. ¹H-NMR (400 MHz, CD₃CN, ppm): δ 0.55 (d, *J* = 4 Hz, 6 H, -SiCH₃), 1.16 (t, *J* = 7.2 Hz, 6H, -CH₂CH₃), 1.24-1.26 (m, 3H, -CH₂CH₃), 1.32-1.39 (m, 2H, -CH₂-), 1.44 (t, *J* = 8 Hz, 3H, -CH₂CH₃), 1.49-1.54 (m, 2H, -CH₂-), 1.68-1.76 (m, 2H, -CH₂-), 2.27-2.29 (m, 2H, -CH₂-), 3.39-4.01 (m, 10H, -NCH₂CH₃, -NCHCH₂-), 7.28 (s, 1H, Ph-H), 7.91 (dd, *J*₁ = 8.8 Hz, *J*₂ = 2.4 Hz, 2H, Ph-H), 8.08 (d, *J* = 2.4 Hz, 1H, Ph-H), 8.49 (d, *J* = 8.4 Hz, 1H, Ph-H). Mass spectrometry (ESI-MS, m/z): [M]⁺ Calc for C₂₉H₄₁ClN₃OSi, 494.2758, Found, 494.2754.

Synthesis of *B*-SiRhd-6

The mixture of **O-SiRhd-II** (100 mg, 0.21 mmol) in CH₃CN (10 mL) was stirred at 0 °C under N₂ for 10 min then (COCl)₂ (0.1 mL) was added dropwise over 1 min. The reaction mixture was stirred for 10 min then ethylenediamine (0.35 mL, 5.25 mmol) was added. The mixture was stirred 30 min at room temperature. The completion of reaction was monitored by TLC. After completion, solvents were removed under reduced pressure and the crude compound was used without further purification. To a solution crude compound in ethanol (10 mL) at 0 °C was added phenyl isothiocyanate (0.1 mL). The mixture was stirred overnight at 25 °C. The solvents were removed under reduced pressure and the residue was purified by HPLC (eluent: CH₃CN/H₂O = 90/10 to 80/20, 16 min) to afford the pure product *B*-SiRhd-6 (12 mg): Yield 9%. ¹H-NMR (400 MHz, DMSO-*d*₆, ppm): δ 0.36-0.46 (m, 6H, -SiCH₃), 1.04-1.14 (m, 12H, -CH₂CH₃), 1.41-1.86 (m, 8H, -CH₂-), 3.17-3.63 (m, 10H, -NCH₂CH₃, -NCHCH₂-), 3.97-4.11 (m, 4H, -NCH₂CH₂N-), 6.79 (s, 2H, Ph-H), 7.01-7.11 (m, 2H, Ph-H), 7.25 (s, 4H, Ph-H), 7.64-8.17 (m, 2H, Ph-H). Mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₃₈H₅₃N₆SSi, 653.3822, Found, 653.3827.

Synthesis of B-SiRhd-7

The mixture of **O-SiRhd-II** (100 mg, 0.21 mmol) in CH₃CN (10 mL) was stirred at 0 °C under N₂ for 10 min then (COCl)₂ (0.1 mL) was added dropwise over 1 min. The reaction mixture was stirred for 10 min then o-phenylenediamine (226 mg, 2 mmol) was added. The mixture was stirred overnight at 25 °C. The completion of reaction was monitored by TLC. After completion, the solvents were removed under reduced pressure and the residue was purified by flash chromatography (CH₂Cl₂/MeOH = 50/1) to afford the pure product *B***-SiRhd-7** (10 mg). ¹H-NMR (400 MHz, DMSO-*d*₆, ppm): δ 0.38-0.46 (m, 6 H, -SiCH₃), 1.04-1.14 (m, 12H, -CH₂CH₃), 1.23-1.58 (m, 8H, -CH₂CH₂-), 3.13-3.43 (m, 10H, - NCH₂CH₃, -NCHCH₂-), 4.66 (s, 2H, -NH₂), 6.04 (s, 1H, Ph-H), 6.31-6.38 (m, 2H, Ph-H), 6.62-6.79 (m, 6H, Ph-H). ¹³C NMR (100 MHz, DMSO-*d*₆, ppm): δ -2.61, -0.99, 11.59, 12.32, 12.44, 21.97, 27.10, 41.04, 43.63, 54.10, 54.56, 54.89, 112.14, 113.40, 114.09, 116.68, 118.12, 122.38, 124.82, 128.46, 128.98, 133.67, 134.68, 135.10, 136.71, 139.25, 146.60, 164.01. Mass spectrometry (ESI-MS, m/z): [M+H]⁺ Calc for C₃₅H₄₈N₅Si, 566.3679, Found, 566.3678.

2. Figures of photophysical properties



Figure S1. Absorption (black solid line), normalized emission (red solid lines; λ_{ex} = 460 nm) and normalized excitation (λ_{em} = 615 nm; red dotted line) spectra of *B*-SiRhd-1 in PBS solution (10 μ M, pH 7.4).



Figure S2. (A) Absorption of *B*-SiRhd-2 at various pH values in mixed solution of PBS/CH₃CN (10 μ M, ν/ν = 3/1). (B) Fluorescence intensities of *B*-SiRhd-2 at various pH values in mixed solution of PBS/CH₃CN (10 μ M, ν/ν = 3/1). (C) Normalized fluorescence intensities of *B*-SiRhd-2 at various pH values in mixed solution of PBS/CH₃CN (10 μ M, ν/ν = 3/1).



Figure S3. (A) Absorption of 10 μ M *B*-SiRhd-2 at various pH values in PBS. (B) Fluorescence intensities of 10 μ M *B*-SiRhd-2 at various pH values in PBS. (C) Normalized fluorescence intensities of 10 μ M *B*-SiRhd-2 at various pH values in PBS.



Figure S4. Absorption (black solid line), normalized emission (red solid lines; $\lambda_{ex} = 680$ nm) and normalized excitation ($\lambda_{em} = 702$ nm; red dotted line) spectra of *P*-SiRhd-2 in mixed solution of PBS/CH₃CN (10 μ M, *v*/*v* = 3/1, pH 7.4).



Figure S5. (A) Absorption of *P*-SiRhd-2 at various pH values in mixed solution of PBS/CH₃CN (10 μ M, v/v = 3/1). (B) Fluorescence intensities of *P*-SiRhd-2 at various pH values in mixed solution of PBS/CH₃CN (10 μ M, v/v = 3/1). (C) Normalized fluorescence intensities of *P*-SiRhd-2 at various pH values in mixed solution of PBS/CH₃CN (10 μ M, v/v = 3/1). (C)



Figure S6. (A) Absorption of 10 μ M *P*-SiRhd-2 at various pH values in PBS. (B) Fluorescence intensities of 10 μ M *P*-SiRhd-2 at various pH values in PBS. (C) Normalized fluorescence intensities of 10 μ M *P*-SiRhd-2 at various pH values in PBS.



Figure S7. Absorption (A, C) and fluorescence (B, D) spectra of *B*-SiRhd-2 (A, B) and *P*-SiRhd-2 (C, D) in different solvents. For *P*-SiRhd-2, the solvent effect is negligible on the absorption and emission spectra.



Figure S8. Absorption and fluorescence emission spectra of *B*-SiRhd. (A) Absorption and fluorescence emission spectra ($\lambda_{ex} = 550$ nm) of *B*-SiRhd-1 (10 µM) in a mixed solution of CH₃CN/PBS (v/v = 1/3, pH 7.4, 0.01 M). (B)Absorbance and fluorescence emission spectra ($\lambda_{ex} = 550$ nm) of *B*-SiRhd-3 (10 µM) in PBS solution at pH 7.4. (C)Absorbance and fluorescence emission spectra ($\lambda_{ex} = 550$ nm) of *B*-SiRhd-4 (10 µM) in ethonal. (D)Absorbance and fluorescence emission spectra ($\lambda_{ex} = 550$ nm) of *B*-SiRhd-4 (10 µM) in ethonal. (D)Absorbance and fluorescence emission spectra ($\lambda_{ex} = 385$ nm) of *B*-SiRhd-5 (10 µM) in a mixed solution of CH₃CN/HEPES (v/v = 3/7, pH 7.4, 0.02 M).



Figure S9. Absorption and fluorescence emission spectra of *P***-SiRhd.** (a)Absorbance and fluorescence emission spectra ($\lambda_{ex} = 550 \text{ nm}$) of *P*-SiRhd-1 (10 µM) in a mixed solution of CH₃CN/PBS (v/v = 1/3, pH 7.4, 0.01 M). (b) Absorbance and fluorescence emission spectra ($\lambda_{ex} = 680 \text{ nm}$) of *P*-SiRhd-2 (10 µM) in a mixed solution of CH₃CN/PBS (v/v = 1/3, pH 7.4, 0.01 M). (c) Absorbance and fluorescence emission spectra ($\lambda_{ex} = 550 \text{ nm}$) of *P*-SiRhd-3 (10 µM) in PBS solution at pH 7.4. (d) Absorbance and fluorescence emission spectra ($\lambda_{ex} = 550 \text{ nm}$) of *P*-SiRhd-4 (10 µM) in ethonal.



Figure S10. Absorption and fluorescence emission spectra of SiRhd. (A)Absorbance and fluorescence emission spectra (λ_{ex} = 510 nm) of *B*-SiRhd-6 (10 µM) in a mixed solution of DMSO/HEPES (v/v = 1/1, pH 7.4, 0.02 M). (B)Absorbance and fluorescence emission spectra (λ_{ex} = 800 nm) of *P*-SiRhd-6 (10 µM) in a mixed solution of DMSO/HEPES (v/v = 1/1, pH 7.4, 0.02 M). (C)Absorbance and fluorescence emission spectra (λ_{ex} = 420 nm) of *B*-SiRhd-7 (10 µM) in a mixed

solution of CH₃CN/HEPES (v/v = 1/1, pH 7.4, 0.02 M). (D)Absorbance and fluorescence emission spectra (λ_{ex} = 800 nm) of *P*-SiRhd-7 (10 μ M) in a mixed solution of CH₃CN/HEPES (v/v = 1/1, pH 7.4, 0.02 M).



Figure S11. Femtosecond time-resolved transient absorption spectra and kinetics of SiRhd. Femtosecond timeresolved transient absorption spectra (A) and kinetics (B) of *B*-SiRhd-1 (excited at λ_{ex} = 460 nm). Femtosecond timeresolved transient absorption spectra (C) and kinetics (D) of *P*-SiRhd-2 (excited at λ_{ex} = 680 nm).

3. Responsive performance measurement of SiRhd probes



Figure S12. HRMS spectrum of the products from the reaction of *B*-SiRhd-3 with 1 equiv of Hg²⁺.



Figure S13. HRMS spectrum of the products from the reaction of *B*-SiRhd-4 with 1 equiv of phosgene.



Figure S14. HRMS spectrum of the products from the reaction of *P*-SiRhd-1 with 1 equiv of Pd.



Figure S15. Fluorescence intensity ratio of 10 μ M *P*-SiRhd-1 ($I_{605 \text{ nm}}/I_{700 \text{ nm}}$) in the present of different metal ions in a mixed solution of CH₃CN/PBS (v/v =1/3, pH = 7.4, 0.01 M).



Figure S16. Time dependence of fluorescence intensity at 620 nm and 718 nm for *B*-SiRhd-3 (20 μ M) in PBS solution (pH = 7.4) in the presence of Hg²⁺ (20 μ M). λ_{ex} = 546 nm. Data was recorded every 0.5 s.



Figure S17. Fluorescence intensity ratio of 10 μ M *B*-SiRhd-3 ($I_{710 \text{ nm}}/I_{610 \text{ nm}}$) in the present of different metal ions in PBS solution at pH 7.4



Figure S18. Dual-channel linear ratio analysis of *B*-SiRhd-3 for Hg²⁺. A linear correlation between fluorescence ratio I_{710} nm/ I_{610} nm and concentration of Hg²⁺. Note: The detection limit was calculated to be 0.062 μ M·L⁻¹ (3 σ /slope).



Figure S19. Dual-channel linear ratio analysis of *B*-SiRhd-4 for phosgene. A linear correlation between fluorescence ratio $I_{696 \text{ nm}}/I_{595 \text{ nm}}$ and concentration of phosgene.



Figure S20. Dual-channel linear ratio analysis of *P*-SiRhd-1 for palladium. A linear correlation between fluorescence ratio $I_{701 \text{ nm}}/I_{606 \text{ nm}}$ and concentration of palladium.



Figure S21. Initially, HeLa cells (incubated with *B*-SiRhd-3) exhibited bright fluorescence in the 650 nm channel and non-fluorescence in the 710 nm channel (Figure S21a). After incubation with Hg²⁺ (10 μ M), the HeLa cells exhibited a sharply decreased fluorescence intensity in the 650 nm channel and a concomitant increase in the 710 nm channel (Figure S21b), which correspond to emission spectra of *B*-SiRhd-3 in the presence of Hg²⁺ (Figure 4D). Furthermore, similar emission profiles and ratiometric were observed in zebrafish. As shown in Figure S21c, the whole body of zebrafish larva was stained by *B*-SiRhd-3, and exhibited 650 nm fluorescence. Then, zebrafish (pre-stained by *B*-SiRhd-3) was incubated with 1 μ M Hg²⁺ for 12 h, and emission at 710 nm was clearly observed mainly in the intestinal tract and heart (Figure S21d). This dual-channel imaging results suggested the activation of *B*-SiRhd-3 could real-time track the uptaken of Hg²⁺ in zebrafish. All these imaging experiments helped to establish how our bending/planar switchable Si-rhodamines enable cross-talk-free ratiometric analysis of biomolecules in living cells and zebrafish.



Figure S22. (A) Reaction of probe *B*-SiRhd-6 with Hg²⁺ and formation of *P*-SiRhd-6. Absorbance (C) and fluorescence emission (D, E, F) spectra of *B*-SiRhd-6 (10 μ M) in the presence of Hg²⁺ in a mixed solution of DMSO/HEPES (v/v =1/1, PH = 7.4, 0.02 M). Temperature elevation curves (G) and infrared thermal (B) of *B*-SiRhd-6 (20 μ M) with different concentrations of Hg²⁺ (0, 10, 20 and 100 μ M) under 808 nm laser irradiation (2 W/cm², 10 min). (H) The variation of temperatures during five cycles of heating-cooling processes under 808 nm laser irradiation of *B*-SiRhd-6 (20 μ M) with 150 μ M Hg²⁺. (I) PA790 of the *B*-SiRhd-6 in the presence of various representative metal cations. (J) In vitro PA images of the solution of *B*-SiRhd-6 in the presence of various representative metal lons (Na²⁺, K⁺, Mg²⁺, Ca²⁺, Al³⁺, Zn²⁺, Cu²⁺, Ni²⁺, Pb²⁺, Sn²⁺, and Hg²⁺) at 790 nm. (K) Plot of PA₇₉₀ of *B*-SiRhd-6 against the concentration of Hg²⁺.



Figure S23. Absorbance intensity in 785 nm of 10 μ M *B*-SiRhd-6 in the present of different metal ions in a mixed solution of DMSO/HEPES (v/v =1/1, PH = 7.4, 0.02 M).



Figure S24. HRMS spectrum of the products from the reaction of B-SiRhd-6 with 1 equiv of Hg^{2+} .



Figure S25. (A) Absorption spectra of B-SiRhd-6 (black) and P-SiRhd-6 (red) in DMSO. (B) Fluorescence spectra of B-SiRhd-6 (black) and P-SiRhd-6 (red) in DMSO. $\lambda_{ex} = 800$ nm.



Figure S26. (A) Time dependence of absorption at 785 nm for B-SiRhd-6 (10 μ M) in a mixed solution of DMSO/HEPES (v/v =1/1, pH = 7.4, 0.02 M) in the presence of Hg²⁺ (20 μ M). (B) Time dependence of absorption at 798 nm for B-SiRhd-7 (10 μ M) in a mixed solution of CH₃CN/HEPES (v/v = 1/1, pH 7.4, 0.02 M) in the presence of NO (100 μ M).



Figure S27. A linear correlation between fluorescence ratio I / I_0 and concentration of NO. Note: The detection limit was calculated to be 0.3 μ M (3 σ /slope). λ_{ex} = 808 nm, λ_{em} = 1040 nm.



Figure S28. (A) Absorption spectra of B-SiRhd-7 (black) and P-SiRhd-7 (red) in DMSO. (B) Fluorescence spectra of B-SiRhd-7 (black) and P-SiRhd-7 (red) in DMSO. λ_{ex} = 800 nm.



Figure S29. HRMS spectrum of the products from the reaction of *B*-SiRhd-7 with 1 equiv of DEA NONOate.



Figure S30. High resolution liquid chromatograph mass (LCMS) traces of probe *B*-SiRhd-7 with excess DEA NONOate. Absorption in 800 nm was verified belonging to *P*-SiRhd-7.



Figure S31. Fluorescence emission (λ_{ex} = 808 nm, 880 nm long-pass filter) of *B*-SiRhd-7 with different concentrations (10, 20, 30, 40, 50, 60 µM) in the presence of DEA NONOate in a mixed solution of CH₃CN/HEPES (v/v =1/1, PH = 7.4, 0.02 M).



Figure S32. Fluorescence emission (λ_{ex} = 808 nm, 880 nm long-pass filter) of *B*-SiRhd-7 (20 μ M) in the presence of different equivalence of DEA NONOate in a mixed solution of CH₃CN/HEPES (v/v =1/1, PH = 7.4, 0.02 M).



Figure S33. Fluorescence emission (λ_{ex} = 808 nm, 880 nm long-pass filter) of *B*-SiRhd-7 (20 µM) over various species (1) Blank; (2) NaClO (0.1 mM); (3) H₂O₂ (0.1 mM); (4) CaCl₂ (0.1 mM); (5) NaNO₂ (1 mM); (6) GSH (1 mM); (7) Glu (1 mM); (8) Cys(0.2 mM); (9) Na₂SO₃(1 mM), and (10) DEA NONOtae in a mixed solution of DMSO/HEPES (v/v =1/1 , PH = 7.4 , 0.02 M).



Figure S34. Photostability of ICG and *P*-SiRhd-7 under continuous-wave laser exposure with a power density of 1 W/cm² in HEPES/CH₃CN solution (1:1, v/v, pH 7.4), λ_{ex} =808 nm. The photostability of NIR-II fluorophores is a key factor for practical application of in vivo bioimaging. These results demonstrated that *P*-SiRhd-7 possess better photostability than ICG.



Figure S35. Photothermal conversion behavior of B-SiRhd-7 (20 μ M) with 500 μ M DEA NONOate after different irradiation at 808 nm laser irradiation (0.8–3 W/cm²). Note: Large excess nitric oxide is added here to only ensure as fast and complete response as possible from the probe, so as to choose the laser with optimal power density, while not priority for the sensitivity of NO with the probe.



Figure S36. Temperature elevation curves of *B*-SiRhd-7 (20 μ M) with different concentrations of DEA NONOate (0, 15, 30 and 100 μ M) under 808 nm laser irradiation (2 W/cm², 10 min).



Figure S37. Infrared thermal of *B*-SiRhd-7 (20 μ M) with different concentrations of DEA NONOate (0, 15, 30 and 100 μ M) under 808 nm laser irradiation (2 W/cm², 10 min).



Figure S38. PA_{800} nm of the *B*-SiRhd-7 in the presence of various representative species (1) Blank; (2) NaClO (0.1 mM); (3) H_2O_2 (0.1 mM); (4) CaCl₂ (0.1 mM); (5) NaNO₂ (1 mM); (6) GSH (1 mM); (7) Glu (1 mM); (8) Cys (0.2 mM); (9) Na₂SO₃(1 mM), and (10) DEA NONOtae in a mixed solution of DMSO/HEPES (v/v =1/1, PH = 7.4, 0.02 M).



Figure S39. Representative size distribution graphs (A) and TEM images (B) of *B*-SiRhd-7@liposome depicting the hydrodynamic diameter after 48 h of synthesis. Insets show the TEM images of the liposomes. (A) Scale bar: 200 nm and (B) Scale bar: 50 nm. (C) Graph shows stability of *B*-SiRhd-7@liposome as a function of change in size distribution and PDI over time, measured by dynamic light scattering during storage condition at 4 °C. Sample size, n=3.



Figure S40. (A) Normalized absorption spectra of B-SiRhd-7@liposome (black) and B-SiRhd-7@liposome+NO (red) in PBS (pH=7.4). (B) Normalized fluorescence spectra of B-SiRhd-7@liposome (black) and B-SiRhd-7@liposome+NO (red) in PBS (pH=7.4). λ_{ex} = 808 nm.



Figure S41. Viability of RAW264.7 cells incubated with B-SiRhd-7@liposome for 24 h.



Figure S42. Comparison of fluorescence intensities at 11h in Figure 6D. Data are presented as the mean value, and the error bars represented the SD from the meanvalue (n = 3). *** P < 0.001.



Figure S43. In vivo circulation of *B*-SiRhd-7@liposome in LPS-pretreated mice in ventral view. (A) The representative time course images after administration of *B*-SiRhd-7@liposome (0.4 mM, 200 μ L) through tail vessel after LPS pretreated 6 h, λ_{ex} = 808 nm, 1100 nm long-pass filter. (B) The normalized fluorescence intensity of *B*-SiRhd-7@liposome in the mice versus time. The maximum fluorescence intensity (11 h in panel A) is defined as 1.0.



Figure S44. In vivo circulation of *B*-SiRhd-7@liposome in LPS-pretreated mice in dorsal view. (A) The representative time course images after administration of *B*-SiRhd-7@liposome (0.4 mM, 200 μ L) through tail vessel after LPS pretreated 6 h, λ_{ex} = 808 nm, 1100 nm long-pass filter. (B) The normalized fluorescence intensity of *B*-SiRhd-7@liposome in the mice versus time. The maximum fluorescence intensity (28 h in panel A) is defined as 1.0.



Figure S45. Bright-field image and NIR-II fluorescence imaging of different organs excised from the LPS-induced mouse at 9 h after tail-vein injection of *B*-SiRhd-7@liposome.



Figure S46. (A) In vivo fluorescence imaging of NO in wild-type and AD-model (APP/PS1) mice brains during 11 h via intravenous injection of 200 μ L 0.4 mM B-SiRhd-7@liposomes, λ_{ex} = 808 nm, 1100 nm long-pass filter. (B) Plots of in vivo fluorescence intensities versus time points for mice in image (A). The maximum fluorescence intensity (11 h) is defined as 1.0. (C) Comparison of fluorescence intensities at 11h for the two groups. Data are presented as the mean value, and the error bars represented the SD from the mean value (n = 3). * P < 0.05.



4. ¹H NMR, ¹³C NMR, and HRMS spectra of the compounds

Figure S47. ¹H NMR spectrum of **O-SiRhd** in CD₃CN.



Figure S48. ¹H NMR spectrum of Cl-SiRhd in CD₃CN.

Monoisotopic Mass, Even Electron Ions 25 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass) Elements Used: C: 0-23 H: 0-36 N: 0-3 Si: 0-2 CI: 0-1 WH-ZHU ZW-XQS-4107 14 (0.300) Cm (14)



Figure S49. HRMS spectrum of Cl-SiRhd.



Figure S50. ¹H NMR spectrum of **B-SiRhd-1** in DMSO-d₆.



Figure S51. HRMS spectrum of **B-SiRhd-1**.



Figure S52. ¹H NMR spectrum of *B*-SiRhd-2 in CDCl₃.



Figure S53. HRMS spectrum of **B-SiRhd-2**.







Figure S55. ¹³C NMR spectrum of **B-SiRhd-3** in DMSO-d₆







Figure S57. ¹H NMR spectrum of **B-SiRhd-4** in DMSO-d₆.











Figure S60. HRMS spectrum of **B-SiRhd-5**.



Figure S61. ¹H NMR spectrum of **P-SiRhd-1** in DMSO-d₆.







Figure S63. ¹H NMR spectrum of *P***-SiRhd-2** in DMSO-*d*₆.

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Figure S65. ¹H NMR spectrum of compound 1 in CDCl₃



Figure S66. ¹³C NMR spectrum of compound 1 in CDCl₃



Figure S67. HRMS spectrum of compound 1.



Figure S68. ¹H NMR spectrum of compound 2 in CDCl₃







Figure S70. HRMS spectrum of compound 2.



Figure S71. ¹H NMR spectrum of compound 3 in CDCl₃



Figure S72. ¹³C NMR spectrum of compound 3 in CDCl₃

Monoisotopic Mass, Even Electron Ions 74 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass) Elements Used: C: 0-25 H: 0-37 N: 0-3 O: 0-2 Br: 0-1



Figure S73. HRMS spectrum of compound 3.



Figure S74. ¹H NMR spectrum of compound 4 in CDCl₃







Figure S76. HRMS spectrum of compound 4.



Figure S77. ¹H NMR spectrum of compound 5 in CDCl₃



Figure S78. ¹³C NMR spectrum of compound 5 in CDCl₃







Figure S80. ¹H NMR spectrum of compound 6 in CDCl₃



Figure S81. ¹³C NMR spectrum of compound 6 in CDCl₃







Figure S83. ¹H NMR spectrum of O-SiRhd-II in DMSO-d₆.



Figure S84. ¹³C NMR spectrum of **O-SiRhd-II** in DMSO-*d*₆.



Figure S85. HRMS spectrum of O-SiRhd-II.



Figure S86. ¹H NMR spectrum of O-SiRhd-II in CD₃CN.







Figure S88. HRMS spectrum of CI-SiRhd-II.







Figure S90. HRMS spectrum of **B-SiRhd-6**.



Figure S91. ¹H NMR spectrum of **B-SiRhd-7** in DMSO-*d*₆.



Figure S92. ¹³C NMR spectrum of **B-SiRhd-7** in DMSO-*d*₆.



Figure S93. HRMS spectrum of **B-SiRhd-7**.

5. Tables of photophysical data and crystal data

Dye	$\lambda_{\scriptscriptstyle abs}({\sf nm})^{[a]}$	$\epsilon_{\lambda abs}$ (M ⁻¹ ·cm ⁻¹) ^[b]	$\lambda_{ ext{em}}$ (nm) $^{[c]}$	Φ _F (%)
B-SiRhd-1 ^[d]	460	17731	610	37 ^[g]
P-SiRhd-1 ^[d]	680	85276	701	31 ^[g]
B-SiRhd-4 ^[e]	460	14042	595	34 ^[g]
B-SiRhd-3 ^[d]	488	9677	610	51 ^[g]
P-SiRhd-3 ^[d]	690	52518	710	55 ^[g]
P-SiRhd-7 ^[f]	780	24035	1040	0.03 ^[f, h]

Table S1. Photophysical data of B-SiRhds and P-SiRhds.

[a] Absorption peak.

[b] Molar absorptivity at the absorption peak.

[c] Fluorescence emission peak.

[d] Measured pH 7.4 PBS.

[e] Measured in methanol.

[f] Measured in a mixed solution of $CH_3CN/HEPES$ (v/v =1/1, pH 7.4).

[g] measured by using rhodamine 6G (ϕ = 88% in ethanol) as a standard.

[h] measured by using IR-26 (ϕ = 0.05% in dichloroethane) as a standard.

Table S2. Crystal data and structure refinement for *B*-SiRhd-5. The X-ray crystallographic coordinates for *B*-SiRhd-5 have been deposited at the Cambridge Crystallographic Data Centre (CCDC), under deposition number: 2119838.

Identification code	<i>B</i> -SiRhd-5		
Empirical formula	C29 H38 N4 Si		
Formula weight	470.72		
Temperature	193(2) K		
Wavelength	avelength 0.71073 Å		
Crystal system	Orthorhombic		
Space group	P 21 21 21	P 21 21 21	
Unit cell dimensions	a = 8.4703(2) Å	α = 90°.	
	b = 13.2994(3) Å	β = 90°.	
	c = 24.4250(7) Å	γ = 90°.	
Volume	2751.47(12) Å ³		
Z	4		
Density (calculated)	1.136 Mg/m ³		
Absorption coefficient	0.108 mm ⁻¹		
F(000) 1016			
Crystal size	0.200 x 0.150 x 0.120 mm ³		
Theta range for data collection	2.545 to 25.999°.		
Index ranges	-10<=h<=9, -16<=k<=16, -28<=l<=30		
Reflections collected	24545		
Independent reflections	5417 [R(int) = 0.0423]		
Completeness to theta = 25.242°	99.5 %		
Absorption correction	Semi-empirical from equivalents		
Max. and min. transmission	0.7456 and 0.6649		
Refinement method	Full-matrix least-squares on	Full-matrix least-squares on F ²	
Data / restraints / parameters	5417 / 2 / 321	5417 / 2 / 321	
Goodness-of-fit on F ²	1.048		
Final R indices [I>2sigma(I)]	R1 = 0.0476, wR2 = 0.1161		
R indices (all data)	R1 = 0.0566, wR2 = 0.1237		
Absolute structure parameter	0.05(5)		
Extinction coefficient	n/a		
Largest diff. peak and hole	0.737 and -0.326 e.Å ⁻³		

 Table S3. Crystal data and structure refinement for P-SiRhd-2. The X-ray crystallographic coordinates for P-SiRhd-2

 have been deposited at the Cambridge Crystallographic Data Centre (CCDC), under deposition number: 2119842.

Identification code	P-SiRhd-2		
Empirical formula	C30 H44 Cl3 N3 O Si		
Formula weight	597.12		
Temperature	293(2) К		
Wavelength	velength 0.71073 Å		
Crystal system	Monoclinic		
Space group	P 21/c		
Unit cell dimensions	a = 10.8497(8) Å	α = 90°.	
	b = 15.4899(12) Å	β = 97.135(2)°.	
	c = 20.3611(14) Å	γ = 90°.	
Volume	3395.4(4) Å ³		
Z	4		
Density (calculated)	1.168 Mg/m ³		
Absorption coefficient	0.331 mm ⁻¹		
F(000)	1272		
Crystal size	0.200 x 0.150 x 0.080 mm ³		
Theta range for data collection	2.407 to 24.999°.		
Index ranges	-12<=h<=12, -16<=k<=18, -24<=	<=24	
Reflections collected	26357		
Independent reflections	5966 [R(int) = 0.0625]		
Completeness to theta = 25.242°	97.2 %		
Absorption correction	Semi-empirical from equivalents	5	
Max. and min. transmission	0.7456 and 0.6574		
Refinement method	Full-matrix least-squares on F^2		
Data / restraints / parameters	5966 / 12 / 369		
Goodness-of-fit on F ²	1.034		
Final R indices [I>2sigma(I)]	R1 = 0.0718, wR2 = 0.1771		
R indices (all data)	R1 = 0.1143, wR2 = 0.2144		
Extinction coefficient	0.0048(13)		
Largest diff. peak and hole 0.739 and -0.462 e.Å ⁻³			

Table S4. Crystal data and structure refinement for CI-SiRhd. The X-ray crystallographic coordinates for CI-SiRhd have been deposited at the Cambridge Crystallographic Data Centre (CCDC), under deposition number: 2132279.

Identification code	Cl-SiRhd			
Empirical formula	C25 H36 Cl6 N2 Si			
Formula weight	605.35			
Temperature	272.0 K			
Wavelength	1.54178 Å	1.54178 Å		
Crystal system	Orthorhombic			
Space group	Pbca			
Unit cell dimensions	a = 22.2241(9) Å	α= 90°.		
	b = 9.0072(3) Å	β = 90° .		
	c = 30.4536(11) Å	γ = 90°.		
Volume	6096.1(4) Å ³			
Z	8			
Density (calculated)	1.319 Mg/m ³			
Absorption coefficient	5.646 mm ⁻¹			
F(000)	2528	2528		
Crystal size	0.1 x 0.05 x 0.02 mm ³			
Theta range for data collection	2.902 to 68.298°.			
Index ranges	ndex ranges -26<=h<=26, -10<=k<=10, -34<=l<=36			
Reflections collected	54262			
Independent reflections	5562 [R(int) = 0.0784]			
Completeness to theta = 67.679°	99.6 %	99.6 %		
Absorption correction	Semi-empirical from eq	Semi-empirical from equivalents		
Max. and min. transmission	nd min. transmission 0.7531 and 0.4474			
Refinement method	ethod Full-matrix least-squares on F ²			
Data / restraints / parameters	aints / parameters 5562 / 41 / 367			
Goodness-of-fit on F ²	1.050			
Final R indices [I>2sigma(I)]	R1 = 0.0568, wR2 = 0.14	R1 = 0.0568, wR2 = 0.1454		
indices (all data) R1 = 0.0912, wR2 = 0.1648				
Extinction coefficient	n/a			
Largest diff. peak and hole	0.340 and -0.453 e.Å ⁻³	0.340 and -0.453 e.Å ⁻³		

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