

Supplementary Information

Plastic recognition and electrogenic uniport translocation of 1st-, 2nd- and 3rd-row transition and post-transition metals by transmembrane P_{1B-2}-type ATPase pumps

Sameera S. Abeyrathna^{1#}, Nisansala S. Abeyrathna^{1#}, Priyanka Basak^{1#}, Gordon W. Irvine^{1#}, Limei Zhang², Gabriele Meloni¹

¹ Department of Chemistry and Biochemistry, The University of Texas at Dallas, Richardson, TX 75080, USA.

² Department of Biochemistry and Redox Biology Center and the Nebraska Center for Integrated Biomolecular Communication, University of Nebraska—Lincoln, Lincoln, NE 68588, USA.

Contributed equally.

Correspondence to: gabriele.meloni@utdallas.edu

Supplementary Figures

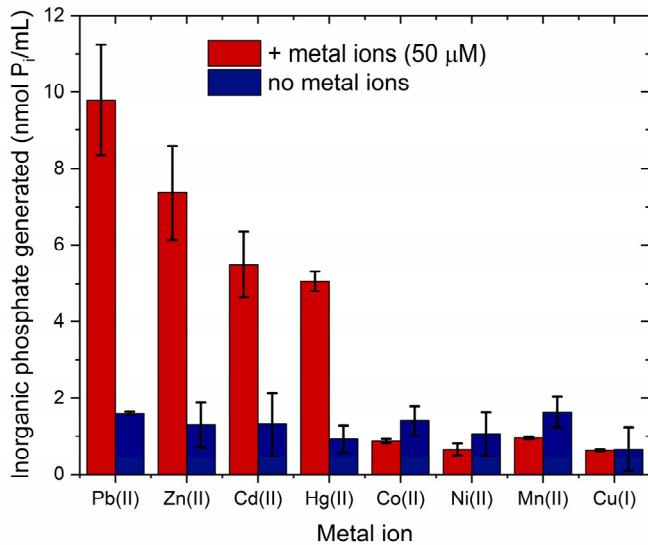


Figure S1: Background Pi and metal-independent ATPase activity.

Metal-stimulated *CmΔZntA* ATPase activity in the presence of different metals in Cymal-7 micelles (metal concentration = 50 μ M; 30 min, 30 °C), compared to the corresponding background Pi generation in the absence of metals. Upon subtracting the Pi originating form buffer and protein independent ATP hydrolysis (approx. 10 nmol Pi/ml) only a minimal metal-independent background ATPase activity by *CmΔZntA* was observed (in average < 10% compared to the maximal activity, either due to uncoupling or background metal contamination). Thus, the metal-dependent stimulation of *CmΔZntA* ATPase activity can be reliably quantified (by subtracting the background in the absence of magnesium).

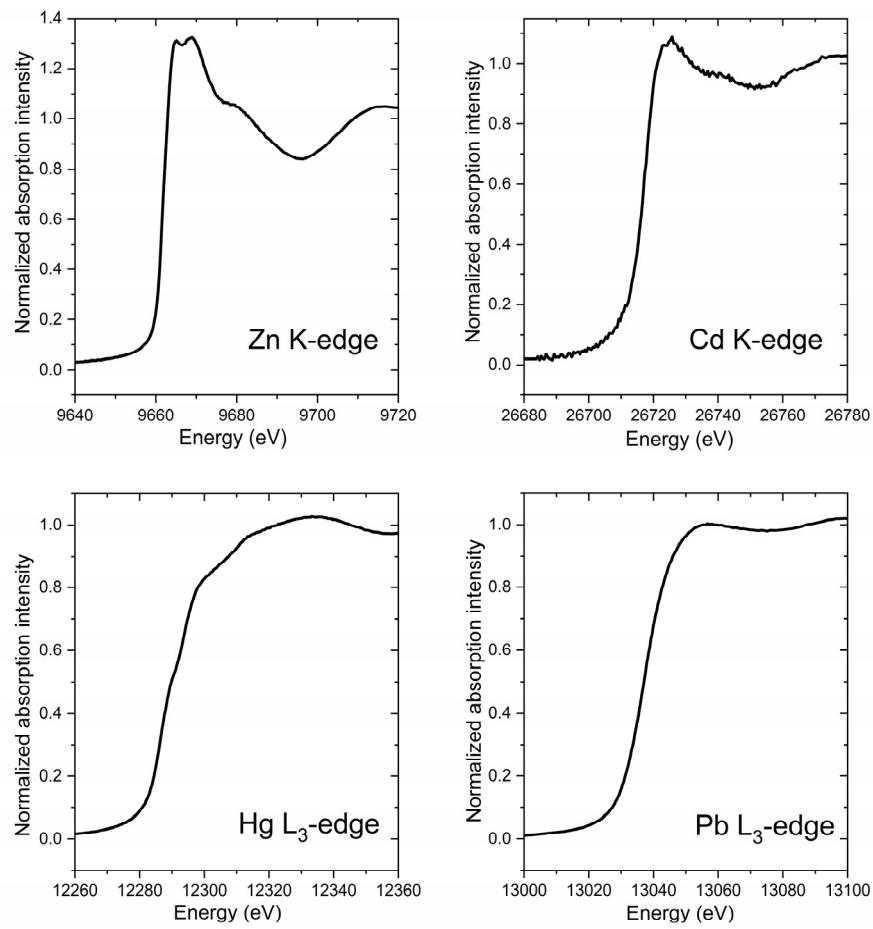


Figure S2: XANES spectra of *CmΔZntA*-M²⁺.

XANES spectra of *CmΔZntA*-M²⁺ (M²⁺= Zn²⁺, Cd²⁺, Hg²⁺ or Pb²⁺) in Cymal-7 detergent micelles.

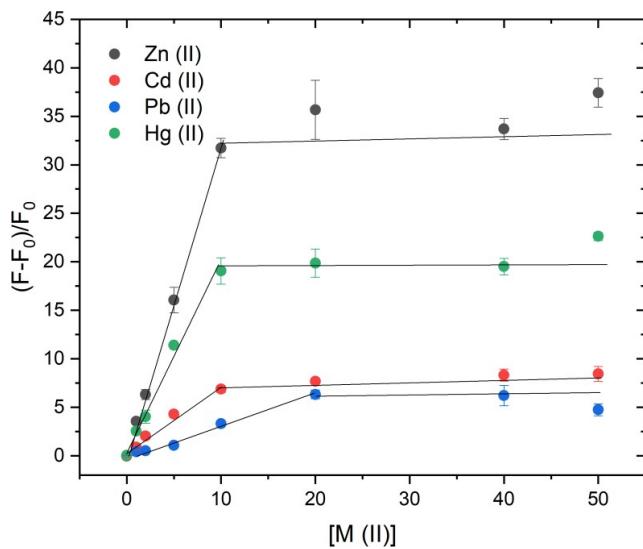


Figure S3: Titration plot of the metal fluorescent turn-on probes with the corresponding metal ions.

Fluozin-3 (10 μM) fluorescence turn-on response upon Zn²⁺ (black) or Cd²⁺ (red) titration ($\lambda_{\text{exc}} = 480 \text{ nm}$; $\lambda_{\text{em}} = 515 \text{ nm}$); Calcium green (10 μM) fluorescence response upon Hg²⁺ (green) titration ($\lambda_{\text{exc}} = 506 \text{ nm}$; $\lambda_{\text{em}} = 531 \text{ nm}$), and Leadmium green (20 μM) fluorescence response upon Pb²⁺ (blue) titration ($\lambda_{\text{exc}} = 490 \text{ nm}$; $\lambda_{\text{em}} = 520 \text{ nm}$), reported as $(F - F_0)/F_0$ (with F_0 the fluorescence in the absence of metals), in the presence of 12.5 mg ml⁻¹ liposomes (200nm). Data are mean ± s.d. (n=3).

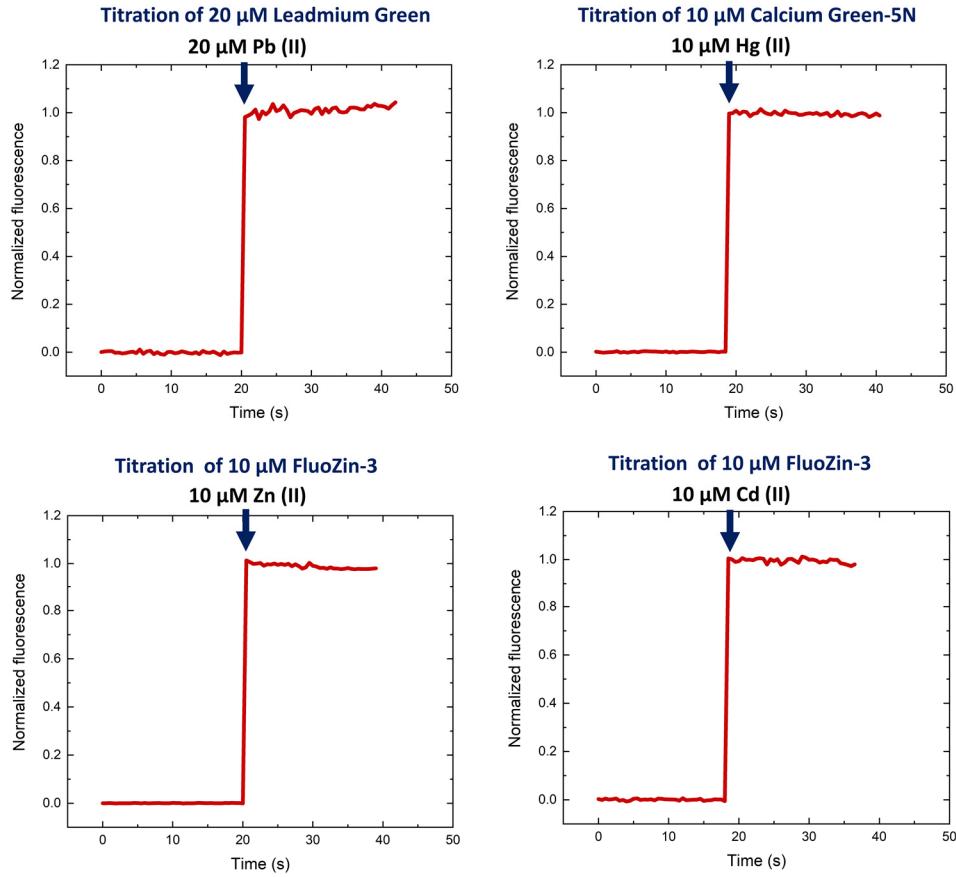


Figure S4: Kinetics of metal binding to the probes.

Fluorescence kinetic traces obtained upon titration of the fluorescent turn-on probes with Zn(II), Cd(II), Hg(II), and Pb(II). Complete stoichiometric binding occurs within the manual mixing-time upon metal addition (see arrow; approx. 5 seconds).

Table S1: Progression to best EXAFS fits for each metal-site

Shell	r (Å)	σ^2 (x10 ⁻³ Å ⁻²)*	R-factor(%)	Red. χ^2
Zn-site				
4N/O	2.11(2)	7(2)	27.5	184.7
4S	2.25(2)	10(1)	14.5	97.2
3N/O	2.03(1)	4(1)		
1S	2.31(1)	1(0)	4.12	35.1
2N/O	1.99(0)	2(0)		
2S	2.29(0)	4(0)	2.91	24.8
Cd-site				
4N/O	2.46(0)	15(59)	98.80	327.41
2S	2.46(0)	1(0)	2.73	9.03
3S	2.46(0)	3(0)	1.48	4.93
4S	2.46(0)	5(0)	4.85	16.09
1N/O	2.24(2)	<i>negative</i> 2(0)		
2S	2.53(2)		1.21	5.01
2N/O	2.32(2)	5(0)		
2S	2.48(2)	2(2)	1.73	6.4
3N/O	2.30(1)	0(0) ^a		
1S	2.57(1)	2(2)	1.21	5.0
4N/O	2.36(1)	1(0)		
1S	2.53(1)	3(1)	1.54	6.4

3N/O	2.30(2)	25(11)		
2S	2.54(1)	1(1)	2.08	8.6
5N/O	2.37(0)	3(1)		
1S	2.51(1)	3(1)	2.39	9.8
4N/O	2.22(3)	29(11)		
2S	2.46(2)	1(0)	2.07	8.6
<u>Hg-site</u>				
4N/O	2.43(0)	neg	4.64	18.7
5N/O	2.43(0)	0(0)	3.43	13.9
6N/O	2.43(0)	1(0)	3.93	15.9
2S	2.33(1)	3(0)	2.21	8.5
3S	2.33(1)	5(0)	8.7	33.3
1N/O	2.44(1)	0(2) ^a		
2S	2.33(1)	4(0)	1.53	7.5
1N/O	2.48(3)	5(0)^b		
2S	2.33(0)	3(0)	1.66	7.1
2N/O	2.49(1)	0(2) ^a		
2S	2.31(1)	5(1)	3.10	15.1
2N/O	2.49(1)	5(0) ^b		
2S	2.31(1)	3(0)	3.31	15.1
<u>Pb-site</u>				
2N/O	2.54(1)	1(1)	18.76	37.1

2S	2.67(2)	9(1)	8.06	15.9
3S	2.65(2)	9(0)	10.94	17.63
2N/O	2.48(6)	10(4)		
1S	2.67(1)	3(1)	8.50	20.5
1N/O	2.52(2)	1(2)		
2S	2.67(2)	9(1)	7.34	17.7
2N/O	2.54(2)	4(3)		
2S	2.65(2)	10(4)	7.92	19.1

r - inter atomic distance (in parentheses are mean-square deviations)

σ^2 - Debye-Waller factor (in parentheses are mean-square deviations in DW factor).

* - values reported as 0 represent small positive numbers after 3 significant figures (e.g. 0.003 with 0.002 error = 3(2); 0.0004 with 0.0001 error = 0(0))

^aUnacceptable negative or null value

^b σ^2 was fixed during fitting.

Table S2: Relative *CmΔZntA* ATPase activity and translocation “efficiency” observed for metal-stimulated ATPase hydrolysis ($V_{max}/K_M, M^{2+}$) and metal translocation ($[\delta F/\delta t]/K_M$) in proteoliposomes, indicating the preferential order for metal substrates ($Pb(II) > Zn(II) > Hg(II) \sim Cd(II)$).

	$V_{max}/K_M, M^{2+}$	$[\delta F/\delta t]/K_M$
Pb(II)	0.34	1.35
Zn(II)	0.15	0.29
Cd(II)	0.13	0.17
Hg(II)	0.12	0.14