Supplementary Information

Enantioselective Total Syntheses of Six Natural and Two Proposed Meroterpenoids from *Psoralea Corylifolia*

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1. General Information

All moisture- or oxygen-sensitive reactions were carried out under an argon atmosphere in oven flasks. The solvents were purified by distillation over the drying agents indicated and were transferred under argon: toluene, THF and Et₂O from Na; MeOH from Mg and I₂; CH₂Cl₂, Et₃N and DMF from CaH₂. All reactions were monitored by thin-layer chromatography (TLC) on silica gel GF₂₅₄ plates using UV light as visualizing agent (if applicable), and a solution of phosphomolybdic acid (50 g/L) in EtOH followed by heating as developing agents. The products were purified by flash column chromatography on silica gel (200-300 meshes) from the Qingdao Marine Chemical Company (China).

¹H NMR and ¹³C NMR spectra were recorded in CDCl₃, (CD₃)₂CO or CD₃OD solution on a Bruker AM 400 MHz instrument or 600 MHz NMR instrument. Chemical shifts were denoted in ppm (δ), and calibrated by using residual undeuterated solvent (CDCl₃ (7.26 ppm), (CD₃)₂CO (2.05 ppm), CD₃OD (3.31 ppm) or tetramethylsilane (0.00 ppm)) as internal reference for ¹H NMR and the deuterated solvent (CDCl₃ (77.00 ppm), (CD₃)₂CO (29.84 ppm), CD₃OD (49.00 ppm) or tetramethylsilane (0.00 ppm)) as internal standard for ¹³C NMR. The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, br = broad, dd = double doublet, td = triple doublet, dt = double triplet, m = multiplet.

The high-resolution mass spectral analysis (**HRMS**) data were measured on Thermo Fisher Orbitrap Elite Mass Spectrometer or a LCT Premier XE (Waters) mass spectrometer (Waters, Milford, MA, U.S.) by means of the ESI technique.

Electron ionization mass spectra (**EI-MS**) were measured on a Shimadzu GCMSQP2010SE spectrometer by direct inlet at 70 eV and the corresponding signals were given in m/z with relative intensity (%) in brackets.

Melting points (m.p.) were measured on a Kolfer melting point apparatus without calibration (Beijing Tech Instrument Co., LTD).

The IR spectra were recorded on Nicolet Nexus 670 FT-IR spectrometer.

The **X-ray** single-crystal determination was performed on an Agilent Super Nova single crystal X-ray diffractometer.

Optical rotations were detected on RUDOLPH A21202-J APTV/GW.

The **enantiomeric excesses** (ee) value of the products was determined by Ultra Performance Convergence Chromatography (UPC²) equipped with Waters 2998 Photodiode Array Detector instruments.

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2. Synthesis of precursors 2 and 2'

Scheme S1 Synthesis of precursors 2 and 2'.

2.1 Synthesis of compound δ-keto ester 5



Compound δ -keto ester **5** was prepared according to the literature by a modified procedure¹. To a solution of **Cat. 1** (6.0 mmol, 20.0 mol %) in toluene (20.0 mL) were added the 2-methylcyclohexanone (**6**) (3.64 mL, 30.0 mmol) and methyl acrylate (**7**) (5.40 mL, 60.0 mmol, 2.0 equiv.) at room temperature, and the resulting mixture was stirred at 90 °C. After 84 h, the residue was purified by chromatography on silica gel, eluting with 10-60% Et₂O/pentane to give the Michael product δ -keto ester **5** as a colorless oil (4.75 g, 80% yield, 92% ee. R_f = 0.4 (10% Et₂O in pentane)).



Compound 5: Colorless oil; $[\alpha]_D^{25} = -32.0$ (c = 3.0 in EtOH).

¹H NMR (400 MHz, CDCl₃) δ: 3.59 (s, 3H), 2.34-2.29 (m, 2H), 2.27-2.21 (m, 1H), 2.13-2.05 (m, 1H), 2.01-1.93 (m, 1H), 1.80-1.65 (m, 6H), 1.56-1.51 (m, 1H), 1.00 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ: 214.8,

173.7, 51.3, 47.6, 38.9, 38.4, 32.2, 28.7, 27.1, 22.1, 20.7. **HRMS** (ESI) m/z found 221.1449, calculated for C₁₁H₁₆O₃Na [M+Na]⁺ 221.1448. **MS** (EI) m/z (%): 198 (9), 154 (36), 96 (52), 83 (54), 69 (62), 112 (98), 55 (100). **IR** (KBr plate) v_{max}: 2937, 2866, 1739, 1705, 1437, 1377, 1306, 1257, 1172, 1123, 1097 cm⁻¹. **Enantiomeric excess** was determined by **UPC²** (CHIRALPAK[®] IC-3 column, CO₂/Methanol = 97/3, flow rate: 2 mL/min, 40 °C, λ = 297.3



nm), 92% ee, $t_R = 2.177$ min (minor), 2.312 min (major).

2.2.1 Synthesis of chiral thiourea catalyst 1 (Cat. 1)



Catalyst **1** (**Cat. 1**) were synthesized according to the literature procedures.² An isothiocyanate (2.12 g, 10.0 mmol) was dropped to a solution of an ethylenediamine (1.33 mL, 10.0 mmol) in CH₂Cl₂ (20.0 mL), and the mixture was stirred at room temperature for 24 h. When the starting material disappears (monitored by TLC), the solvent was removed under reduced pressure, the residue was purified by column chromatography to afford the thiourea catalysts **1** (**Cat. 1**) as white solid (2.89 g, 80% yield. $R_f = 0.4$ (10% MeOH in DCM)).



Catalyst 1 (**Cat. 1**): White solid; $[\alpha]_D^{26} = -71.8$ (c = 1.0 in CHCl₃), [lit², $[\alpha]_D^{24} = -74.3$ (c = 1.0 in CHCl₃)]; **m.p.**: 66-68 °C. ¹**H NMR** (600 MHz, CDCl₃) δ : 7.43 (brs, 1H), 7.29-7.22 (m, 11H), 7.13

¹H NMR (600 MHz, CDCl₃) δ: 7.43 (brs, 1H), 7.29-7.22 (m, 11H), 7.13 (brs, 4H), 6.50 (brs, 1H), 5.11 (brs, 1H), 4.60 (brs, 1H), 4.52-4.94 (m,

1H), 4.29 (brs, 1H), 1.57 (brs, 2H); ¹³**C NMR** (150 MHz, CDCl₃) δ: 181.8, 141.7, 139.6, 137.1, 128.8, 128.7, 128.6, 127.7, 127.6, 127.4, 126.6, 63.9, 60.2, 48.4. **MS** (EI) m/z (%):

361 (0.1), 327 (4), 256 (15), 194 (3), 165 (7), 106 (100), 91 (31), 79 (12). **IR** (KBr plate) v_{max}: 3268, 3061, 3029, 2964, 1538, 1495, 1453, 1347, 1287, 1262, 1076, 1028, 799, 757, 699 cm⁻¹.

2.2 Synthesis of compound 4

Method I:



To a stirred solution of δ -keto ester **5** (39.6 mg, 0.2 mmol) in dichloromethane (2 mL) was added freshly distilled Et₃N (41.7 µL, 0.3 mmol) and TMSOTf (36.3 µL, 0.2 mmol) at 0 °C and was stirred at the same temperature for 1 h. The reaction was quenched by addition of saturated aqueous NaHCO₃ and the aqueous phase was extracted with diethyl ether (3 × 20 mL). The combined organic extracts were washed with brine, dried over anhydrous Na₂SO₄, and concentrated to give **Int 5** as a yellow oil.

To a stirred solution of the crude product **Int 5** in anhydrous CH₃CN (2 mL) was added Na₂CO₃ (63.6 mg, 0.6 mmol) and Pd(OAc)₂ (48.5 mg, 0.2 mmol). The resulting mixture was stirred at the room temperature for 10 hours, treated with Et₃N (0.31 mL, 2.40 mmol) and then concentrated. The residue was purified by column chromatography to afford **5'** as a yellow oil (25.5 mg, 65%). $R_f = 0.4$ (silica gel, petroleum ether: ethyl acetate = 5:1). **Method II:**



To a stirred solution of **5** (6.6 g, 31.0 mmol) in DMSO:toluene (3:1/v:v) was added IBX (26.1 g, 93.3 mmol), and the mixture was stirred at 85 °C.³ After the starting material disappears (monitored by TLC), aqueous saturated NaHCO₃ (300 mL) and diethyl ether (300 mL) were added to the reaction mixture at room temperature, which was filtered through a Celite pad. The organic layer was separated and the aqueous layer was extracted with diethyl ether (3×100 mL). The combined organic phase was washed with brine, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure to give a residue **5'** as a yellow oil. The crude product **5'** was used for next step without purification.

To a solution of the crude product **5'** (5.22 g, 26.6 mmol) in MeOH (11.0 mL) was added aqueous 2.0 mol/L NaOH (26.6 mL, 53.2 mmol, 2.0 equiv.). The reaction mixture was

stirred at room temperature. When the starting material disappears (monitored by TLC), MeOH was evaporated under reduced pressure. The aqueous layer was washed with Et₂O (1×100 mL), the pH value of the aqueous layer was adjusted to pH = 4 using HCl (1.0 mol/L) before being extracted with ethyl acetate (3×100 mL). The combined extracts were washed with brine, dried over Na₂SO₄, filtered, and concentrated *in vacuo*. The crude product was purified by flash column chromatography to afford carboxylic acid **4** as a yellow oil (4.68 g, 83% yield from **5**, two steps.). R_f = 0.2 (25% ethyl acetate-petroleum ether).



Compound 5': Light yellow oil; $[\alpha]_D^{25} = 2.1$ (c = 0.5 in MeOH). ¹H NMR (400 MHz, CDCl₃) δ : 6.89 (dt, *J* = 10.0, 4.0 Hz, 1H), 5.91 (dt, *J* = 10.0, 2.0 Hz, 1H), 3.66 (s, 3H), 2.45-2.34 (m, 2H), 2.31-2.21 (m, 2H), 1.95-1.77 (m, 4H), 1.10 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ :

202.9, 173.7, 148.4, 128.1, 51.3, 43.6, 33.3, 31.0, 28.8, 22.8, 21.4. **HRMS** (ESI) m/z found 219.0990, calculated for C₁₁H₁₆O₃Na [M+Na]⁺ 219.0992. **MS** (EI) m/z (%): 196 (75), 191 (60), 180 (37), 165(40), 137 (92), 105 (70), 106 (54). **IR** (KBr plate) v_{max} : 2953, 2931, 2873, 1738, 1674, 1436, 1382, 1200, 1174 cm⁻¹. **Enantiomeric excess** was determined by **UPC²** (CHIRALPAK[®] IC-3 column, CO₂/Methanol = 95/5, flow rate: 2 mL/min, 40 °C, λ = 220.0 nm), 92% ee, t_R = 2.445 min (major), 3.775 min (minor).





Compound 4: Yellow oil; $[\alpha]_D^{25} = -0.9$ (c = 0.5 in MeOH).

¹**H NMR** (400 MHz, CDCl₃) δ : 11.18 (brs, 1H), 6.87 (dt, J = 10.0, 4.0 Hz, 1H), 5.91 (dt, J = 10.4, 1.6 Hz, 1H), 2.40-2.26 (m, 4H), 1.92-1.76 (m, 4H), 1.09 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ : 203.4, 179.6,

148.8, 128.4, 43.8, 33.6, 31.1, 29.1, 23.0, 21.6. **HRMS** (ESI) m/z found 205.0835, calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 205.0832. **MS** (EI) m/z (%): 182 (53), 164 (23), 148(75), 138 (42), 110 (100), 101 (58). **IR** (KBr plate) v_{max}: 3433, 2961, 2928, 2872, 1710, 1672, 1454, 1389, 1284, 1225, 1077 cm⁻¹. **Enantiomeric excess** was determined by **UPC**² (CHIRALPAK[®] IC-3 column, CO₂/Methanol = 90/10, flow rate: 2 mL/min, 40 °C, λ = 210.0 nm), 94% ee, t_R = 1.637 min (major), 1.871 min (minor).



2.3 Synthesis of compound 3



PdCl₂(PPh₃)₂ (0.2 mol%), Xantphos (0.24 mol%), (*t*-Bu)₄biphenol(1 mol%) Ac₂O (1.2+0.5+0.3+0.2 equiv)

neat, 2 h vacuum distillation



Compound **3** was synthesized following the literature procedure.⁴ A flame-dried 25.0 mL round-bottom flask, PdCl₂(PPh₃)₂ (15.4 mg, 0.022 mmol, 2% equiv.), Xantphos (15.0 mg, 0.026 mmol, 2.4% equiv.), (*t*-Bu)₄biphenol (45.2 mg, 0.11 mmol, 0.01 equiv.) and carboxylic acid 4 (2.0 g, 11.0 mmol, 1.0 equiv.) were added under the argon atmosphere. The flask was equipped with adistillation head and a 10 mL round-bottom receiving flask. The system was evacuated and backfilled with argon three times, and the first portion of acetic anhydride (6.0 mmol, 1.2 equiv.) was added via syringe through the septum that seals the top of the distillation head. The flask was immersed in a pre-heated 60 °C oil bath and rapidly heated to 130 °C. When the oil bath temperature reached 120 °C, switched to vacuum to allow distillation of resultant acetic acid and trace product into a receiving flask. which was cooled to -78 °C. When the oil bath temperature reached 130 °C. The system a vacuum was drawn and the mixture was stirred for approximate 30 min at 130 °C. The system was backfilled with argon atmosphere, and the second portion of acetic anhydride (2.5 mmol, 0.5 equiv.) was added via syringe. The system was then gradually resubjected to a vacuum. Acetic anhydride was added as follows (0.3, 0.2 equiv.) in the same manner every 30 min. The reaction was stopped about 2 h and allowed to cool to the room temperature under the argon atmosphere. The distillate was added to a saturated aqueous solution of NaHCO₃, stirred for 30 min, and the biphasic mixture and residual dark red reaction mixtures were extracted with CH₂Cl₂ (3×20 mL). The combined extracts were washed with brine, dried over Na₂SO₄. The solvents were evaporated and the residue was purified by flash column chromatography on silica gel to afford vinyl ketone 3 (1.05 g, 70% yield) as a colorless oil. ($R_f = 0.5$, 16% Et₂O in pentane).

Compound 3: Colorless oil; $[\alpha]_D^{26} = -71.8$ (c = 1.0 in CHCl₃).



¹**H NMR** (400 MHz, CDCl₃) δ : 6.93-6.88 (m, 1H), 5.97 (dt, *J* = 10.0, 2.0 Hz, 1H), 5.93 (dd, *J* = 17.6, 10.8 Hz, 1H), 5.05 (dd, *J* = 10.8, 0.4 Hz, 1H), 5.00 (dd, *J* = 17.6, 0.4 Hz, 1H), 2.44-2.31 (m, 2H), 2.03-1.97 (m, 1H), 1.93-1.86

(m, 1H), 1.20 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 201.4, 149.1, 140.3, 128.5, 114.0, 47.9, 34.5, 23.3, 22.6. HRMS (ESI) m/z found 137.0962, calculated for C₁₁H₁₇O₃ [M+H]⁺ 137.0961. **MS** (EI) m/z (%): 136 (72), 128 (39), 120 (34), 107 (60). **IR** (KBr plate) v_{max}: 2958, 2925, 2854, 1730, 1515, 1469, 1337, 1289, 1068, 749 cm⁻¹. **Enantiomeric excess**

was determined by **UPC**² (CHIRALPAK[®] IG-3 column, CO₂/Methanol = 99/1, flow rate: 2 mL/min, 40 °C, λ = 238.0 nm), 94% ee, t_R = 1.552 min (major), 1.719 min (minor).





Method I:

According to the literature⁵, to a stirred suspension of CuCN (89.6 mg, 1.0 mmol, 1.0 equiv.) in Et₂O (15 mL) was slowly added via syringe a solution of isopropenyllithium (5.0 mL, 0.4 M in Et₂O, 2.0 equiv.) at -78 °C under an argon atmosphere. The mixture was stirred for approximate 30 min at -78 °C until a homogeneous solution was observed, and enone **3** (136.0 mg, 1.0 mmol) was then added at **-78** °C. After the starting material disappeared, the reaction mixture was allowed to warm to -20 °C, diluted with dry DMF (10.0 mL),

Cu(OAc)₂ (181.6 mg, 1.0 mmol, 1.0 equiv.) and the diaryliodonium triflate (4-MeOC₆H₄)₂IOTf (1.024 g, 2.0 mmol, 2.0 equiv.) was sequentially added. The reaction mixture was stirred for 12 h at -20 °C, and quenched with a saturated aqueous solution of NH₄Cl. The mixture was extracted with ethyl acetate (3×30 mL) and the combined organic layers were washed with water and brine, dried over Na₂SO₄, filtered, and concentrated under vacuum. The resultant crude products (dr~8:1) were purified by flash column chromatography to give the desired product **2** (101.0 mg, 35.6% yield) and its isomer **2**' (12.5 mg, 4.4% yield). (R_f = 0.50 and 0.51, respectively, 10% ethyl acetate-petroleum ether).

Method II:

According to the literature⁵, to a stirred suspension of CuCN (89.6 mg, 1.0 mmol, 1.0 equiv.) in Et₂O (15.0 mL) was slowly added *via* syringe a solution of isopropenyllithium (5.0 mL, 0.4 M in Et₂O, 2.0 equiv.) at -78 °C under an argon atmosphere. The mixture was stirred for approximate 30 min at -78 °C until a homogeneous solution was observed. After the reaction mixture was allowed to warm to room temperature, and enone **3** (136.0 mg, 1.0 mmol) was then added at room temperature. After the starting material disappeared, **CsF** (608.0 mg, 4.0 mmol, 4.0 equiv.) was added and the reaction mixture was stirred for 10 minutes. Subsequently, diaryliodonium triflate (4-MeOC₆H₄)₂IOTf (1.024 g, 2.0 mmol, 2.0 equiv.) in anhydrous DMF (6.5 mL) was added via syringe. The reaction mixture was stirred for 24 h at room temperature, and quenched with a saturated aqueous solution of NH₄Cl. The mixture was extracted with ethyl acetate (3x30 mL) and the combined organic layers were washed with water and brine, dried over Na₂SO₄, filtered, and concentrated under vacuum. The resultant crude products (dr = 1.5:1) were purified by flash column chromatography to give the product **2** (102.0 mg, 35.9% yield) and its isomer **2'** (68.4 mg, 24.1% yield). (R_f = 0.50 and 0.51, respectively, 10% ethyl acetate-petroleum ether).

(4-MeOC₆H₄)₂IOTf were prepared according to the reported procedures.⁶

Isopropenyllithium was prepared according to the following manner⁷: To a precooled (-78 °C) solution of 2-bromopropene (281.7 μ L, 3.2 mmol) in dry Et₂O (2.8 mL) was added dropwise *tert*-butyllithium (4.92 mL, 1.3 *M* in pentane, 6.4 mmol). The resulting solution was stirred at -78 °C for 30 min, and then used immediately.

2.4.1 The Optimization of the Cu-mediated one-pot Michael addition/arylation reaction

Unless otherwise noted, the reaction was performed according to the Method I and Method II, and the details were listed in Table S1-S4

2.4.1.1 The screening of additives

	о — з	Isopropenyllithium CuCN, Et ₂ O, T $^{\circ}$ C then Additives, (4-MeOC ₆ H ₄) ₂ IOTf DMF, rt 2 (major)	2' (minor)	
Entry	Temp.	Additives (equiv.)	dr (2 : 2')	Yield ^b
1	-78 °C	CuCl (1.0)		25%
2	-78 °C	Cul (1.0)		20%
3	-78 °C	CuOAc (1.0)		32%
4	-78 °C	Cu(OTf) ₂ (1.0)		34%
5	-78 °C	Cu(OAc)₂ (1.0)	8:1	40%
6	-78 °C	Cu(OAc) ₂ (1.0)	8:1	32%
7	rt	Cu(OAc) ₂ (1.0)	1.4:1	35%
8	40 °C	Cu(OAc) ₂ (1.0)	1.7:1	20%
9	-78 °C to rt	LiHMDS (2.0)	4:1	44%
10	-78 °C to rt	LiCA (2.0)	2:1	38%
11	-78 °C to rt	Cs ₂ CO ₃ (2.0)	1.8:1	25%
12	-78 °C to rt	CsOH·H ₂ O (2.0)	1.7:1	20%
13	-78 °C to rt	NaH (2.0)		ND
14	-78 °C to rt	DBU (2.0)		trace
15	-78 °C to rt	Na ₂ CO ₃ (2.0)		trace
16	-78 °C to rt	DMAP (2.0)		NR
17	-78 °C to rt	DIPEA (2.0)		NR
18	-78 °C to rt	HMPA (5.0), Cu(OAc)₂ (1.0)		21%
19	-78 °C to rt	TMEDA (5.0), Cu(OAc) ₂ (1.0)		25%
20	-78 °C to rt	CsF (2.0)	3:1	50%
21	rt	CsF (2.0)	1.51:1	50%
22	rt	CsF (3.0)	1.51:1	53%
23	rt	CsF (4.0)	1.49:1	60.0%
24	rt	CsF (5.0)	1.49:1	59.6%

Table S1. The screening of additives^a

^aReaction was performed using isopropenyllithium (0.2 mmol) and CuCN (0.1 mmol) in 1.5 mL Et₂O at -78 °C and stirred for 30 min; then enone **3** (0.1 mmol) was added to the reaction mixture. After the starting material (**3**) disappeared, additives and (4-MeOC₆H₄)₂IOTf (0.2 mmol) in DMF was added to the reaction mixture at the note temperature. ^bIsolated yield; rt = Room temperature; DMF = *N*,*N*-dimethylformamide; LiCA = Lithium cyclohexylisopropylamide; HMPA = Hexamethylphosphoramide; DMAP = 4dimethylaminopyridine; DBU = 1,8-diazabicyclo[5.4.0]undec-7-ene; DIPEA = *N*,*N*-diisopropylethylamine; TMEDA = *N*,*N*,*N*,*N*-tetramethylethylenediamine.

2.4.1.2 The screening of diaryliodonium salts

	3 Isopropenyllithium CuCN, Et ₂ O, -78 °C then Cu(OAc) ₂ , Diaryliodonium salts DMF, rt	MeO Me	0 0 1 1 2' (minor)	
Entry	Diaryliodonium salts	dr (2 : 2')	Yield ^b	
1	(4-MeOC ₆ H₄)₂IOTf	8:1	40%	
2	(4-MeOC ₆ H ₄) ₂ IBF ₄	4.8:1	27%	
3	(4-MeOC ₆ H ₄) ₂ IOTs	8:1	10%	
4	(4-MeOC ₆ H ₄) ₂ IBF ₆	8:1	30%	

Table S2. The screening of diaryliodonium salts^a

^aReaction was performed using isopropenyllithium (0.2 mmol) and CuCN (0.1 mmol) in 1.5 mL Et₂O at -78 °C and stirred for 30 min, then enone **3** (0.1 mmol) was added to the reaction mixture, after the starting material (**3**) disappeared. Then Cu(OAc)₂ (0.1 mmol) and diaryliodonium salts (0.2 mmol) in DMF was added to the reaction mixture at rt. ^bIsolated yield; rt = Room temperature; DMF = *N*,*N*-Dimethylformamide. **2.4.1.3 The screening of chiral ligands**

Table S3. The screening of chiral ligands^a Isopropenyllithium MeO MeC CuCN, Et₂O, -78 °C Liquids then Cu(OAc)₂ (1.0 eq.) (4-MeOC₆H₄)₂IOTf DMF, rt 2' (minor) 2 (major) Ρĥ -tBu tBu L3 L4 L5 L6 12 Yield^b Entry Liquids Additives (equiv.) dr (2:2') 1 L1 Cu(OAc)₂(1.0) 2.9:1 11.7% 2 L2 Cu(OAc)₂(1.0) 10.5% 3:1 3 Cu(OAc)₂(1.0) L3 trace ___ 4 L4 Cu(OAc)₂(1.0) trace 5 L5 Cu(OAc)₂(1.0) trace __ L6 $Cu(OAc)_2(1.0)$ 6 -trace 7 CsF (4.0) 31.7% L1 4.5:1 8 L2 CsF (4.0) 28.7% 3:1 9 L3 CsF (4.0) trace --CsF (4.0) 10 L4 trace --11 L5 CsF (4.0) trace 12 L6 CsF (4.0) trace

^aReaction was performed using isopropenyllithium (0.2 mmol) and CuCN (0.1 mmol) in 1.5 mL Et₂O at -78 °C and stirred for 30 min; then chiral ligand (L) was added to the reaction mixture and stirred for 1 h; enone **3** (0.1 mmol) was added. After the starting material (**3**) disappeared, Cu(OAc)₂ (0.1 mmol) and (4-MeOC₆H₄)₂IOTf (0.2 mmol) in DMF was added to the reaction mixture; ^bIsolated yield; rt = Room temperature; DMF = *N*,*N*-Dimethylformamide.

2.4.1.4 The screening of organmetallic reagents

1) IsopropenylMgBr:



^aReaction was performed using isopropenylmagnesium bromide (0.12 mmol) and CuBr·SMe₂ in 1.5 mL solvent at the note temperature and stirred for 30 min; then enone **3** (0.1 mmol) was added to the reaction mixture. After the starting material (**3**) disappeared, additives and (4-MeOC₆H₄)₂IOTf (0.2 mmol) in DMF were added to the reaction mixture; ^bIsolated yield; ND = No detected; TMEDA = N,N,N,N-tetramethylethylenediamine; HMPA = Hexamethylphosphoramide; rt = Room temperature; IsopropenylMgBr = Isopropenylmagnesium bromide; HMPA = Hexamethylphosphoramide; LiHMDS = Lithium bis(trimethylsilyl)amide; MTBE = *tert*-Butyl methyl ether; DMF = N,N-Dimethylformamide; THF = Tetrahydrofuran; 2-MeTHF = 2-Methyltetrahydrofuran.

To a stirred suspension of CuBr·SMe₂ (2.05 mg, 0.01 mmol, 0.1 equiv.) and HMPA (174.0 μ L, 0.5 mmol, 5.0 equiv.) in Et₂O (1.0 mL) was slowly added *via* syringe a solution of isopropenylmagnesium bromide (0.4 mL, 0.5 M in THF, 2.0 equiv.) at -78 °C under an argon atmosphere. The mixture was stirred for approximate 30 min at -78 °C until a homogeneous solution was observed. Enone **3** (13.6 mg, 1.0 mmol) was then added at -78 °C. After the starting material disappeared, **CsF** (60.8 mg, 0.4 mmol, 4,0 equiv.) was

added and the reaction was stirred for 10 minutes. Subsequently, diaryliodonium triflate (4- $MeOC_6H_4$)₂IOTf (102.4 mg, 0.2 mmol, 2.0 equiv.) in anhydrous DMF (1.5 mL) was added *via* syringe. The reaction mixture was stirred for 24 h at room temperature, and quenched with a saturated aqueous solution of NH₄Cl. The mixture was extracted with diethyl ether (3×30 mL) and the combined organic layers were washed with water and brine, dried over Na₂SO₄, filtered, and concentrated under vacuum. The resultant crude products (dr = 4:1) were purified by flash column chromatography on silica gel, eluting with 0.5-1% Et₂O/pentane to only afford the Michael addition product **Int 2** (11.9 mg, 67.0% yield) and its isomer **Int 2'** (3.2 mg, 18.0% yield). (R_f = 0.50 and 0.51, respectively, 20% Et₂O/pentane).

2) Diisopropenylzinc:



Cu(OTf)₂ ((4.0 mg, 0.01 mmol, 0.1 equiv.) and chiral ligands (0.01 mmol, 0.12 equiv.) were suspended in dry THF (1.0 mL) and stirred at room temperature for 30 min before cooling to -30 °C, diisopropenylzinc (0.61 mL, 0.33 mol/L in THF, 2.0 equiv.) was added dropwise. The mixture was stirred for approximate 30 min at -30 °C, and enone **3** (13.6 mg, 0.1 mmol) was then added. The mixture was slowly warmed up to room temperature and stirred for 12 h. **CsF** (60.8 mg, 0.4 mmol, 4.0 equiv.) was added and the reaction was stirred for 10 minutes, then diaryliodonium triflate (4-MeOC₆H₄)₂IOTf (102.4 mg, 0.2 mmol, 0.2 equiv.) in anhydrous DMF (1.5 mL) was added *via* syringe. The reaction mixture was stirred for 24 h

at room temperature, and quenched with a saturated aqueous solution of NH₄Cl. The mixture was extracted with diethyl ether ($3 \times 30 \text{ mL}$) and the combined organic layers were washed with water and brine, dried over Na₂SO₄, filtered, and concentrated under vacuum. The resultant crude products (dr~1.4:1) were purified by flash column chromatography on silica gel, eluting with 0.5-1.0% Et₂O/pentane to only afford the Michael addition product **Int 2** (6.4 mg, 36.0% yield) and its isomer **Int 2'** (4.6 mg, 25.7% yield), and without the isolation of expected products **2** and **2'**,

Preparation of diisopropenylzinc reagent⁸

Preparation of isopropenyIMgBr-LiCI

In an oven-dried 10 mL flask, Mg turnings (97.2 mg, 4.0 mmol), I₂ (5 mg, 0.02 mmol) and LiCI (84.8 mg, 2.0 mmol) were weighed. The system was evacuated and backfilled with argon three times. THF (2.0 mL) was added at room temperature and stirred for 5 minutes. The mixture was then cooled to 0 °C and 2-bromopropene (0.18 mL, 2.0 mmol) was added dropwise. The mixture was removed from the ice bath and allowed to stir for 1 h at room temperature.

Preparation of diisopropenylzinc

In an oven-dried flask, $ZnCl_2$ (1.0 mL, 1.0 mol/L in THF, 1.0 equiv., 1.0 mmol) was added under the argon atmosphere. Then, isopropenylMgBr·LiCl (2.0 mL, 2.0 equiv., 2.0 mmol) was added dropwise *via* syringe. A dense-grey solution was formed and used without further titration (c = 0.33 mol/L).



Precursor 2: Yellow oil; $[α]_D^{25} = 82.3$ (c = 0.5 in MeOH). ¹**H NMR** (400 MHz, CDCl₃) δ: 6.93 (d, J = 8.6 Hz, 2H), 6.81 (d, J =

8.6 Hz, 2H), 6.23 (dd, J = 17.6, 10.8 Hz, 1H), 5.10 (d, J = 10.8 Hz, 1H), 5.04 (d, J = 17.6 Hz, 1H), 4.60 (brs, 2H), 3.84 (d, J = 12.4 Hz, 1H), 3.77 (s, 3H), 2.74 (td, J = 12.4, 3.6 Hz, 1H), 2.19-2.08 (m, 1H),

2.03-1.84 (m, 3H), 1.59 (s, 3H), 1.44 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ : 212.1, 158.2, 145.8, 142.9, 130.3, 129.0, 113.4, 112.8, 112.3, 56.0, 55.1, 53.1, 50.5, 36.7, 27.3, 23.0, 18.8. **HRMS** (ESI) m/z found 285.1846, calculated for C₁₈H₂₅O₂ [M+H]⁺ 285.1849. **MS** (EI) m/z (%): 284 (33), 257 (30), 236 (25), 186 (28), 160 (36), 121(100), 107 (30). **IR** (KBr plate) v_{max} : 2935, 2868, 2836, 1707, 1643, 1612, 1514, 1461, 1249, 1179, 1038, 914, 894, 817, 783 cm⁻¹.



Precursor 2': Yellow oil; $[\alpha]_D^{24}$ = -125.6 (c = 0.5 in MeOH).

¹**H NMR** (600 MHz, CDCl₃) δ : 6.93 (d, J = 8.4 Hz, 2H), 6.82 (d, J = 8.6 Hz, 2H), 6.06 (dd, J = 17.4, 10.2 Hz, 1H), 5.26 (d, J = 10.8 Hz, 1H), 5.07 (d, J = 18.0 Hz, 1H), 4.59 (brs, 1H), 4.57 (brs, 1H), 3.90 (d, J = 12.6 Hz, 1H), 3.78 (s, 3H), 2.74 (t, J = 13.2 Hz, 1H),

2.17-2.08 (m, 2H), 1.81-1.75 (m, 2H), 1.55 (s, 3H), 1.17 (s, 3H); ¹³**C NMR** (150 MHz, CDCl₃) δ : 211.2, 158.2, 145.8, 142.8, 130.3, 129.0, 116.0, 113.4, 112.6, 56.8, 55.1, 53.9, 52.2, 39.0, 28.2, 24.8, 18.6. **HRMS** (ESI) m/z found 285.1847, calculated for C₁₈H₂₅O₂ [M+H]⁺ 285.1849. **MS** (EI) m/z (%): 284 (30), 173 (24), 159 (19), 135 (21), 121 (100), 91 (12). **IR** (KBr plate) v_{max}: 2933, 2867, 1709, 1615, 1514, 1458, 1249, 1179, 1067, 966, 892, 817, 750 cm⁻¹.



Int 2: Colorless oil; $[\alpha]_D^{18} = 19.2$ (c = 1.0 in CHCl₃).

¹**H NMR** (600 MHz, CDCl₃) δ : 6.07 (dd, J = 18.0, 10.8 Hz, 1H), 5.12 (d, J = 11.4 Hz, 1H), 5.02 (d, J = 18.0 Hz, 1H), 4.80 (brs, 1H), 4.71 (brs, 1H), 2.56-2.52 (m, 1H), 2.48-2.44 (m, 2H), 1.91-1.86 (m, 1H), 1.82-1.75 (m,

3H), 1.73 (s, 3H), 1.23 (s, 3H); ¹³**C NMR** (150 MHz, CDCl₃) *δ*: 213.4, 147.1, 142.7, 113.4, 110.6, 50.7, 44.9, 43.0, 36.0, 25.5, 22.9, 21.0. **HRMS** (ESI) m/z found 285.1848, calculated for C₁₈H₂₅O₂ [M+H]⁺ 285.1849. **MS** (EI) m/z (%): 178 (59), 121 (78), 111 (84), 107 (63), 105 (100).



Int 2': Colorless oil; $[\alpha]_D^{18} = 96.9$ (c = 1.0 in CHCl₃).

¹**H NMR** (600 MHz, CDCl₃) δ : 5.91 (dd, J = 17.4, 10.2 Hz, 1H), 5.15 (d, J = 10.8 Hz, 1H), 5.00 (d, J = 18.0 Hz, 1H), 4.75 (brs, 1H), 4.71 (brs, 1H), 2.55 (t, J = 13.8 Hz, 1H), 2.36-2.31 (m, 2H), 2.06 (dt, J = 13.8, 3.6

Hz, 1H), 1.78-1.74 (m, 2H), 1.72 (s, 3H), 1.62-1.57 (m, 1H), 1.14 (s, 3H); ¹³**C** NMR (150 MHz, CDCl₃) δ : 212.6, 147.5, 142.3, 115.6, 109.7, 51.6, 46.7, 44.1, 38.8, 27.1, 24.0, 20.4. HRMS (ESI) m/z found 285.1849, calculated for C₁₈H₂₅O₂ [M+H]⁺ 285.1849. MS (EI) m/z (%): 178 (1), 121 (9), 111 (6), 107 (5), 105 (100).



3. Collective total syntheses of $7\alpha\mbox{-aryl-12}\beta\mbox{-alkyl}$ type meroterpenoids

Scheme S2 Collective total syntheses of 7α -aryl-12 β -alkyl type meroterpenoids.

3.1 Synthesis of natural (+)-psoracorylifol F (1a)

3.1.1 Synthesis of compounds 8 and 8'

Method:

To a solution of the precursor **2** (5.0 mg, 0.018 mmol) in solvents (1.0 mL) was added reductants (0.09 mmol, 5.0 equiv.) under an argon atmosphere at the noted temperature, respectively. When the reaction completed (monitored by TLC), the reaction mixture was quenched with a saturated aqueous solution of NH₄Cl, and extracted with CH₂Cl₂ (3×20 mL). The combined organic phase was washed with brine, dried over Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure, and the dr value was measured by ¹H NMR (entries 1-6, Table S5).

MeO		MeO Reductants Solvent, T °C	OH 8	MeO	OH
Entry	reductors	Solvents	Temp.	dr (8 :8')	yield ^b
1	LiAIH ₄	THF	-100 °C	14:1	98%
2	LiAIH ₄	Et ₂ O	-100 °C	4:1	98%
3	BH₃•Me₂NH	DME	rt	1:4.6	96%
4	BH₃•Me₂NH	DME	-60 °C	1:4	95%
5	BH₃∙ <i>i</i> Pr₂NH	DME	-60 °C	1:3	90%
5	BH₃- <i>i</i> Pr₂NH	DME	rt	1:7	95%
6	BH₃∙ <i>i</i> Pr₂NH	DME	90 °C	1:7	56%

Table S5. Optimization of the reduction conditions of the precursor 2

^aThe reactions were performed using the precursor **2** (0.018 mmol), reductants (5.0 equiv.) in 1.0 mL THF; ^bIsolated yield; rt = Room temperature; THF = Tetrahydrofuran; DME = 1,2-dimethoxyethane.

Method I (entry 1, Table S6):



To a solution of the precursor **2** (5.0 mg, 0.018 mmol) in THF (1.0 mL) was added LiAlH₄ (3.4 mg, 0.09 mmol) under an argon atmosphere at -100 °C, and the mixture was stirred for 2 h. When the reaction completed (monitored by TLC), the reaction mixture was quenched with a saturated aqueous solution of NH₄Cl, and extracted with CH₂Cl₂ (3×20 mL). The combined organic phase was washed with brine, dried over Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure. The resulting crude product **8** and **8'** (dr = 14:1) was purified by flash column chromatography (silica gel, petroleum ether/ethyl acetate (50:1)) to give the desired product **8** (4.6 mg, 91% yield) as a white solid and its isomer **8'** (0.35 mg, 7% yield).

Method II (entry 5, Table S6):



To a solution of the precursor **2** (5.0 mg, 0.018 mmol) in DME (1.0 mL) was added $BH_3 \cdot iPrNH$ (10.3 mg, 0.09 mmol) under an argon atmosphere at -60 °C, and the mixture

was stirred for 24h. When the reaction completed (monitored by TLC), the reaction mixture was quenched with a saturated aqueous solution of NH₄Cl, and extracted with CH_2Cl_2 (3×20 mL). The combined organic phase was washed with brine, dried over Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure. The resulting crude product **8** and **8'** (dr = 1:7) was purified by flash column chromatography (silica gel, petroleum ether/ethyl acetate (50:1)) to give the desired product **8'** (4.2 mg, 83% yield) as a white solid and its isomer **8** (0.6 mg, 12% yield).



Compound 8: Colorless needle; $[\alpha]_D^{25} = 3.9$ (c = 0.5 in MeOH); **m.p.**: 105-107 °C.

¹**H NMR** (400 MHz, CDCl₃) δ: 7.12 (d, J = 8.4 Hz, 2H), 6.84 (d, J = 8.4 Hz, 2H), 5.92 (dd, J = 17.6, 10.8 Hz, 1H), 5.11 (dd, J = 17.6,

1.2 Hz, 1H), 5.07 (dd, J = 10.8, 0.8 Hz , 1H), 4.53 (brs, 2H), 4.78 (s, 3H), 3.45 (d, J = 10.4 Hz, 1H), 2.68 (t, J = 11.6 Hz, 1H), 2.37 (td, J = 11.6, 3.6 Hz, 1H), 1.79-1.64 (m, 1H), 1.63-1.56 (m, 3H), 1.54 (s, 3H), 1.18 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ : 158.2, 147.9, 147.0, 133.0, 129.3, 113.8, 112.1, 111.7, 78.9, 55.1, 50.9, 49.0, 41.7, 36.2, 27.3, 19.5, 15.2. **HRMS** (ESI) m/z found 309.1826, calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 309.1825. **MS** (EI) m/z (%): 286 (87), 253 (5), 204 (8), 150 (100), 137 (22), 121(91). **IR** (KBr plate) v_{max}: 3506, 3069, 2975, 2928, 2903, 2852, 1720, 1611, 1512, 1298, 810 cm⁻¹.



Compound 8': White solid; $[\alpha]_D^{25} = 35.6$ (c = 0.5 in MeOH); **m.p.**: 82-83 °C.

¹**H NMR** (400 MHz, CDCl₃) δ: 7.15 (d, *J* = 8.4 Hz, 2H), 6.83 (d, *J* = 8.4 Hz, 2H), 5.88 (dd, *J* = 17.6, 10.8 Hz, 1H), 5.11 (dd, *J* = 4.4, 0.8

Hz, 1H), 5.08 (dd, J = 11.2, 1.2 Hz, 1H), 4.70 (brs, 1H), 4.63 (brs, 1H), 3.77 (s, 3H), 3.39 (brs, 1H), 3.00 (dd, J = 12.4, 2.0 Hz, 1H), 2.94-2.90 (m, 1H), 2.05-1.99 (m, 1H), 1.73-1.71 (m, 2H), 1.56 (s, 3H), 1.40 (dt, J = 13.2, 2.8 Hz, 1H), 1.20 (s, 3H); ¹³**C** NMR (100 MHz, CDCI₃) δ : 158.0, 147.9, 147.0, 134.1, 129.8, 113.4, 112.8, 111.9, 77.5, 55.1, 45.4, 42.0, 41.4, 28.5, 28.0, 22.4, 19.2. HRMS (ESI) m/z found 309.1827, calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 309.1825. **MS** (EI) m/z (%): 286 (23), 150 (71), 137 (16), 121 (100), 68 (13), 55 (10). **IR** (KBr plate) v_{max}: 3461, 3075, 2927, 2858, 1643, 1611, 1583, 1511, 1456, 1248, 1178, 1038, 912, 887, 821 cm⁻¹.

3.1.2 Synthesis of natural (+)-psoracorylifol F (1a)

Method I:



To a stirred solution of alcohol **8** (28.4 mg, 0.1 mmol) in dry CH₂Cl₂ (2.0 mL) at -78 °C was added BBr₃ (0.6 mL, 0.6 mmol, 1 M in CH₂Cl₂, 6.0 equiv.). After stirred for at -40 °C for 12 h, and gradually warmed to room temperature for 16 h, after which reaction was quenched by careful addition of H₂O (1.0 mL) at 0 °C. The pH of the aqueous layer was adjusted to pH 8 using sat. NaHCO₃ before being extracted CH₂Cl₂ (3×20 mL). The combined organic layer was washed with brine, dried with Na₂SO₄, and concentrated in vacuum. The crude product was purified by preparative thin layer chromatography (petroleum ether/ethyl acetate) to afford (+)-psoracorylifol F (**1a**) (20.7 mg, 76% yield) as a white solid. **Method II**:



To a suspension solution of alcohol **8** (19.4 mg, 0.067 mmol) and K₂CO₃ (3.7 mg, 0.027 mmol) in NMP (1.0 mL) was added PhSH (27.3 μ L, 0.27 mmol). The mixture was stirred under reflux for 12 h, and then cooled to room temperature and quenched with H₂O. The resulting mixture was extracted with ethyl acetate (3×20 mL). The combined organic phase was washed with brine, dried over Na₂SO₄, and concentrated. The crude product was purified by chromatography on silica gel (petroleum ether/ethyl acetate) to give (+)-psoracorylifol F (**1a**) (17.7 mg, 97% yield) as a white solid.



(+)-psoracorylifol F (1a): White solid; $[\alpha]_D^{25} = +6.7$ (c = 0.1 in MeOH); m.p.: 157-159 °C.

¹**H NMR** (600 MHz, methanol- d_4) δ : 6.91 (d, J = 8.4 Hz, 2H), 6.57 (d, J = 8.4 Hz, 2H), 5.88 (dd, J = 18.0, 10.8 Hz, 1H), 4.94 (dd, J = 17.4, 1.2 Hz, 1H), 4.87 (dd, J = 10.8, 1.8 Hz, 1H), 4.40 (brs, 1H),

4.37 (brs, 1H), 3.37 (d, J = 10.8 Hz, 1H), 2.54 (t, J = 10.8 Hz, 1H), 2.23 (td, J = 12.0, 3.6 Hz, 1H), 1.69-1.60 (m, 1H), 1.56-1.51 (m, 1H), 1.48-1.45 (m, 1H), 1.44-1.40 (m, 1H), 1.41 (s, 3H), 1.04 (s, 3H); ¹³**C NMR** (150 MHz, methanol- d_4) δ : 156.8, 150.1, 149.1, 134.7, 131.0,

116.1, 112.3, 111.9, 80.6, 53.4, 51.0, 43.3, 37.6, 28.9, 20.1, 16.6. **HRMS** (ESI) m/z found 295.1669, calculated for $C_{18}H_{24}O_2Na$ [M+Na]⁺ 295.1669. **MS** (EI) m/z (%): 272 (89), 245 (56), 220 (60), 187 (43), 121 (63), 113 (69), 100 (55). **IR** (KBr plate) v_{max}: 3344, 2969, 2924, 2866, 1643,1615, 1600, 1514, 1245, 1081, 822 cm⁻¹.

Comparison of ¹H NMR data for synthetic and natural (+)-psoracorylifol F



	(+)-psorac	orylifol F (1a)	
Position	Natural psoracorylifol F ^{a,9}	Synthetic psoracorylifol F ^b	Δδ (ppm)
1	· -	-	-
2,5	6.90, d (<i>J</i> = 8.5 Hz)	6.91, d (<i>J</i> = 8.4 Hz)	-0.01
3,6	6.58, d (<i>J</i> = 8.5 Hz)	6.57, d (<i>J</i> = 8.4 Hz)	0.01
4	-	-	-
7	2.54, t (<i>J</i> = 11.0 Hz)	2.54, t (<i>J</i> = 10.8 Hz)	0.00
8	3.37, d (<i>J</i> = 10.5 Hz)	3.37, d (<i>J</i> = 10.8 Hz)	0.00
9	-	-	-
10	1.44, m	1.44-1.40, m (1.42, m)	0.02
	1.64, m	1.69-1.59, m (1.64, m)	0.00
11	1.48, m	1.48-1.45, m (1.47, m)	0.01
	1.54, o ^c	1.56-1.51, m (1.54, m)	0.00
12	2.24, m	2.23, td (<i>J</i> = 12.0, 3.6 Hz)	0.01
13	-	-	-
14	4.36, s	4.37, brs	-0.01
	4.40, s	4.40, brs	0.00
15	1.41, s	1.41, s	0.00
16	5.87, dd (<i>J</i> = 17.5, 11.0 Hz)	5.88, dd (<i>J</i> = 18.0, 10.8 Hz)	-0.01
17	4.86, dd (<i>J</i> = 11.0, 1.5 Hz)	4.87, dd (<i>J</i> = 10.8, 1.8 Hz)	-0.01
	4.94, dd (<i>J</i> = 18.0, 1.5 Hz)	4.94, dd (<i>J</i> = 17.4, 1.2 Hz)	0.00
18	1.04, s	1.04, s	0.00

^a NMR data measured at 500 MHz in methanol-*d*₄; ^b NMR data measured at 600 MHz in methanol-*d*₄; ^c "o" means overlapped.





Comparison of ¹³C NMR data for synthetic and natural (+)-psoracorylifol F



(+)-psoracorylifol F (1a)

Position	Natural psoracorylifol F a,9	Synthetic psoracorylifol F ^b	Δ δ (ppm)
1	134.9	134.7	0.2
2	131.0	131.0	0.0
3	116.1	116.1	0.0
4	156.8	156.8	0.0
5	116.1	116.1	0.0
6	131.0	131.0	0.0
7	51.0	51.0	0.0
8	80.6	80.6	0.0
9	43.3	43.3	0.0
10	37.6	37.6	0.0
11	28.9	28.9	0.0
12	53.4	53.4	0.0
13	149.1	149.1	0.0
14	112.3	112.3	0.0
15	20.2	20.1	0.1
16	150.0	150.1	-0.1
17	111.9	111.9	0.0
18	16.6	16.6	0.0

^a NMR data measured at 125 MHz in methanol-*d*₄; ^b NMR data measured at 150 MHz in methanol-*d*₄.



Comparison of ¹³C NMR spectra of (+)-psoracorylifol F





To a suspension solution of alcohol **8'** (10.0 mg, 0.035 mmol) and K₂CO₃ (1.9 mg, 0.014 mmol) in NMP (1.0 mL) was added PhSH (17.9 μ L, 0.175 mmol). The mixture was stirred under reflux for 12 h, then cooled to room temperature and quenched with H₂O. The resulting mixture was extracted with ethyl acetate (3×20 mL). The combined extracts were washed with brine, dried over Na₂SO₄. The solvent was removed under vacuum, and the residue was purified by flash column chromatography (petroleum ether/ethyl acetate, 20:1-10:1) on silica gel to give *pseudo*-natural (-)-8α-hydroxy-psoracorylifol F (**9**) (9.0 mg, 95% yield) as a white solid.



Pseudo-natural (-)-8α-hydroxy-psoracorylifol F (9): White solid. $[\alpha]_D^{21} = -40.7$ (c = 1.0 in MeOH). m.p.: 114-116 °C. ¹H NMR (600 MHz, CDCl₃) δ: 7.09 (d, J = 8.4 Hz, 2H), 6.74 (d, J = 8.4 Hz, 2H), 5.88 (dd, J = 18.0, 11.4 Hz, 1H), 5.10 (d, J = 10.8 Hz, 1H), 5.08 (d, J = 16.8 Hz, 1H), 4.68 (s, 1H), 4.61 (brs, 2H), 3.38 (s,

1H), 2.98 (dd, J = 12.6, 1.8 Hz, 1H), 2.90-2.86 (m, 1H), 2.05-1.99 (m, 1H), 1.72-1.68 (m, 2H), 1.54 (s, 3H), 1.40 (dt, J = 13.2, 3.0 Hz, 1H), 1.19 (s, 3H); ¹³**C NMR** (150 MHz, CDCl₃) δ : 153.9, 147.9, 147.0, 134.4, 130.0, 115.0, 112.9, 111.9, 77.5, 45.4, 42.0, 41.4, 28.5, 28.0,

22.4, 19.2. **HRMS** (ESI) m/z found 295.1671, calculated for $C_{18}H_{24}O_2Na$ [M+Na]⁺ 295.1669. **MS** (EI) m/z (%): 272 (17), 136 (87), 123 (21), 107 (100), 68 (17), 55 (11). **IR** (KBr plate) v_{max} : 3394, 3076, 2963, 2922, 2861, 1641, 1613, 1514, 1451, 1374, 1224, 1174, 963, 912, 888, 821 cm⁻¹.

3.3 Synthesis of natural (-)-7α,8β-hydroxy-12β-cyclobakuchiol C (1b)



To an ice-cold suspension solution of NaHCO₃ (21.0 mg, 0.25 mmol) and *m*-CPBA (13.4 mg, 80%, 0.06 mmol) in CH₂Cl₂ (1.0 mL) was added a solution of (+)-psoracorylifol F (**1a**, 14.8 mg, 0.05 mmol) in CH₂Cl₂ (1.0 mL) at 0 °C. The mixture was stirred at 5 °C for 4 h and the reaction was quenched by addition of Me₂S (5.3 µL, 0.07 mmol) and saturated NaHCO₃ with vigorous stirring. The layers were separated and the aqueous layer was extracted with CH₂Cl₂ (3×20 mL). The combined extracts were washed with brine, dried over Na₂SO₄, and concentrated to give a residue, which was directly used for the next step. To an ice-cold solution of above oxirane in THF (2.0 mL) was added LiAlH₄ (9.5 mg, 0.25 mmol) slowly at 0 °C. The resulting mixture was stirred for 3 h under reflux, then cooled to room temperature and quenched with H₂O. The resulting mixture was extracted with ethyl acetate (3×20 mL). The combined extracts were washed with saturated NaHCO₃ and brine, dried over Na₂SO₄, and concentrated. The residual oil was purified by column chromatographed on silica gel (petroleum ether/ethyl acetate) to give (-)-7α,8β-hydroxy-12β-cyclobakuchiol C (**1b**) (11.0 mg, 76% yield for 2 steps from natural (+)-psoracorylifol F (**1a**)).



(-)-7α,8β-hydroxy-12β-cyclobakuchiol C (1b): Colorless needles; $[\alpha]_D^{25} = -22.0$ (c = 0.1 in MeOH), [lit¹⁰, $[\alpha]_D^{25} = -27.2$ (c = 0.1 in MeOH)]. m.p.: 104-106 °C.

¹H NMR (600 MHz, acetone-*d*₆) δ: 8.13 (s, 1H), 7.09 (d, *J* = 7.8 Hz, 2H), 6.73 (d, *J* = 8.4 Hz, 2H), 5.97 (dd, *J* = 17.4, 10.8 Hz, 1H), 4.96

(dd, J = 17.4, 1.8 Hz, 1H), 4.86 (dd, J = 10.8, 1.2 Hz, 1H), 3.40 (dd, J = 9.6, 4.2 Hz, 1H), 2.55 (t, J = 11.4 Hz, 1H), 2.44 (d, J = 4.2 Hz, 1H), 2.21 (s, 1H), 1.90-1.87 (m, 1H), 1.82 (td, J = 11.4, 4.2 Hz, 1H), 1.53-1.51 (m, 2H), 1.44-1.36 (m, 1H), 1.04 (s, 3H), 0.96 (s, 3H), 0.79 (s, 3H); ¹³**C NMR** (100 MHz, acetone- d_6) δ : 156.8, 149.9, 135.1, 131.1, 116.1, 110.9, 80.4,

73.2, 52.9, 49.4, 42.0, 36.0, 27.6, 23.5, 16.2. **HRMS** (ESI) m/z found 313.1774, calculated for $C_{18}H_{26}O_3Na$ [M+Na]⁺ 313.1774. **MS** (EI) m/z (%): 290 (6), 214 (23), 161 (32), 133 (100), 107 (83), 59 (56). **IR** (KBr plate) v_{max}: 3530, 2920, 2851, 1614, 1514, 1461, 1376, 1229, 1172, 960, 897, 827, 784 cm⁻¹.

Comparison of 1H NMR data of synthetic and natural (-)-7 $\alpha,8\beta$ -hydroxy-12 β -cyclobakuchiol C (1b)



D	Natural	Synthetic	Δδ
Position	7α ,8 β -hydroxy-12 β -cyclobakuchiol C ^{a,10}	7α ,8 β -hydroxy-12 β -cyclobakuchiol C b	(ppm)
1	-	-	-
2,6	7.09, d (<i>J</i> = 8.0 Hz)	7.09, d (<i>J</i> = 7.8 Hz)	0.00
3,5	6.74, d (<i>J</i> = 8.5 Hz)	6.73, d (<i>J</i> = 8.4 Hz)	0.01
4	-	-	-
7	2.55, t (<i>J</i> = 11.0 Hz)	2.55, t (<i>J</i> = 11.4 Hz)	0.00
8	3.41, d (<i>J</i> = 10.0 Hz)	3.40, dd (<i>J</i> =9.6, 4.2 Hz)	0.01
9	-	-	-
10 _{ax}	1.53, m	1.53-1.51, m (1.52, m)	0.01
10 _{eq}	1.53, m	1.53-1.51, m (1.52, m)	0.01
11 _{ax}	1.90, m	1.90-1.87, m (1.89, m)	0.01
11 _{eq}	1.40, m	1.44-1.36, m (1.40, m)	0.00
12	1.83, m	1.82, td (<i>J</i> = 11.4, 4.2 Hz)	0.01
13	-	-	-
14	0.99, s	0.96 (s)	0.03
15	0.76, s	0.79 (s)	-0.03
16	5.97, dd (<i>J</i> = 17.5, 10.5 Hz)	5.97, dd (<i>J</i> = 17.4, 10.8 Hz)	0.00
17 _{ax}	4.97, dd (<i>J</i> = 17.5, 1.5 Hz)	4.96, dd (<i>J</i> =17.4, 1.8 Hz)	0.01
17 _{eq}	4.88, dd (<i>J</i> = 11.0, 1.5 Hz)	4.86, dd (<i>J</i> = 10.8, 1.2 Hz)	0.02
18	1.05, s	1.04 (s)	0.01

^a NMR data measured at 500 MHz in acetone-d₆; ^b NMR data measured at 600 MHz in acetone-d₆.



Comparison of ¹H NMR spectra of (-)-7 α ,8 β -hydroxy-12 β -cyclobakuchiol C (1b)





Position	Natural 7α,8β-hydroxy-12β-cyclobakuchiol C ^{a,10}	Synthetic 7α,8β-hydroxy-12β-cyclobakuchiol C ^b	Δδ (ppm)
1	134.8	135.1	-0.3
2,6	131.1	131.1	0.0
3,5	116.0	116.1	-0.1
4	156.8	156.8	0.0
7	49.3	49.4	-0.1
8	80.4	80.4	0.0
9	42.0	42.0	0.0
10	36.1	36.0	0.1
11	23.3	23.5	-0.2
12	52.8	52.9	-0.1
13	73.4	73.2	0.2
14	27.0	27.6	-0.6
15	29.6	29.2	0.4
16	149.8	149.9	-0.1
17	111.0	110.9	0.1
18	16.2	16.2	0.0

^a NMR data measured at 125 MHz in acetone-d₆; ^b NMR data measured at 100 MHz in acetone-d₆.



Comparison of ¹³C NMR spectra of (-)-7α,8β-hydroxy-12β-cyclobakuchiol C (1b)

3.4 Synthesis of the proposed corypsoriol J (1c)

3.4.1 Synthesis of compound 10

MeO



To a suspension solution of alcohol **8** (70.0 mg, 0.24 mmol) and NaH (38.4 mg, 60%, 0.96 mmol) in THF (5.0 mL) was added MeI (60.0 μ L, 0.96 mmol). The mixture was stirred at room temperature for 20 h, and quenched with H₂O. The resulting mixture was extracted with ethyl acetate (3×20 mL). The combined extracts were washed with brine, dried over Na₂SO₄, and filtered, and concentrated in vacuo. The crude residue was purified by flash column chromatography (petroleum ether/ethyl acetate = 100:1) on silica gel to give methylether **10** (68.4 mg, 95% yield) as a colorless oil.



Compound 10: Colorless oil; $[\alpha]_D^{23} = 22.7$ (c = 0.5 in MeOH). ¹H NMR (600 MHz, CDCl₃) δ : 7.12 (d, J = 8.4 Hz, 2H), 6.79 (d, J = 8.4 Hz, 2H), 5.97 (dd, J = 17.4, 10.8 Hz, 1H), 5.04 (dd, J = 17.4, 1.2 Hz, 1H), 4.98 (dd, J = 10.8, 1.2 Hz, 1H), 4.55 (brs, 1H), 4.53 (brs,

1H), 3.77 (s, 3H), 2.89 (d, J = 10.2 Hz, 1H), 2.70 (t, J = 10.8 Hz, 1H), 2.70 (s, 3H), 2.38 (td, J = 11.4, 3.6 Hz, 1H), 1.69-1.55 (m, 4H), 1.53 (s, 3H), 1.13 (s, 3H); ¹³**C** NMR (150 MHz, CDCl₃) δ : 157.8, 148.3, 147.1, 134.2, 129.5, 113.2, 111.7, 111.0, 90.8, 61.2, 55.0, 50.8, 48.5, 42.2, 35.9, 27.2, 19.3, 16.4. HRMS (ESI) m/z found 323.1981, calculated for C₂₀H₂₈O₂Na [M+Na]⁺ 323.1982. **MS** (EI) m/z (%): 300 (65), 255 (20), 179 (42), 164 (100), 121 (46). **IR** (KBr plate) v_{max}: 3076, 3033, 2970, 2929, 2864, 2830, 1645, 1613, 1512, 1455, 1269, 1247, 1176, 1126, 1038, 911, 888, 818 cm⁻¹.

3.4.2 Synthesis of compound 11

MeO



To a suspension solution of methylether **10** (42.0 mg, 0.14 mmol) and K₂CO₃ (8.1 mg, 0.06 mmol) in NMP (2.0 mL) was added PhSH (58.0 μ L, 0.56 mmol). The mixture was stirred under reflux for 14 h. The reaction was quenched by addition of HCI (1.0 mol/L) and extracted with ethyl acetate (3×10 mL). The combined extracts were washed with brine, dried (Na₂SO₄), and concentrated. The crude product was purified by column chromatography on silica gel (petroleum ether/ethyl acetate) to give phenol **11** (37.2 mg, 93% yield).



Compound 11: White solid; [α]²⁵_D = 20.0 (c = 0.5 in MeOH); **m.p.**: 94.2-95.3 °C.

¹**H NMR** (400 MHz, CDCl₃) δ : 7.07 (d, J = 8.4 Hz, 2H), 6.72 (d, J = 8.8 Hz, 2H), 5.96 (dd, J = 17.6, 10.8 Hz, 1H), 5.04 (dd, J = 17.6,

1.2 Hz, 1H), 4.99 (dd, J = 10.8, 1.2 Hz, 1H), 4.95 (s, 1H), 4.54 (brs, 1H), 4.53 (brs, 1H), 2.91 (d, J = 10.4 Hz, 1H), 2.73 (s, 3H), 2.75-2.67 (m, 1H), 2.37 (td, J = 11.6, 3.2 Hz, 1H), 1.71-1.56 (m, 4H), 1.53 (s, 3H), 1.13 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ : 153.8, 148.2, 147.0, 134.3, 129.7, 114.8, 111.7, 111.2, 90.9, 61.2, 50.9, 48.5, 42.3, 36.0, 27.2, 19.3, 16.4. **HRMS** (ESI) m/z found 309.1823, calculated for C₁₉H₂₆O₂Na [M+Na]⁺ 309.1825. **MS** (EI) m/z (%): 286 (12), 241 (13), 186 (11), 150 (100), 135 (23), 107 (64). **IR** (KBr plate) v_{max}: 3077, 2973, 2931, 2867, 1644, 1614, 1515, 1449, 1413, 1375, 1225, 1103, 913, 890, 827 cm⁻¹.

3.4.3 Synthesis of the proposed corypsoriol J (1c)



To a solution of phenol **11** (17.0 mg, 0.06 mmol) in CHCl₃/MeOH (3:1 v/v, 1.2 mL) was added IBX (20.2 mg, 0.07 mmol) at 0 °C under an argon atmosphere. A yellow-to-orange color was observed and the mixture was stirred for 2 h, then NaBH₄ (20.5 mg, 0.54 mmol) was added at 0 °C under vigorous stirring for 30 min.¹¹ The reaction was quenched by addition of HCl (1.0 mol/L) and extracted with ethyl acetate (3×10 mL). The combined extracts were washed with brine, dried over Na₂SO₄, and filtered. The filtrate was

concentrated under reduced pressure, and the residue was purified by column chromatography on silica gel using ethyl acetate-petroleum ether (1:1) to give the proposed corypsoriol J (**1c**) (17.2 mg, 95% yield).



Proposed corypsoriol J (1c): White solid; $[\alpha]_D^{25} = 13.3$ (c = 0.1 in MeOH); [lit¹², $[\alpha]_D^{25} = 30.0$ (c = 0.1 in MeOH)]. **m.p.**: 160-163 °C. ¹**H NMR** (600 MHz, methanol-*d*₄) δ : 6.65 (brs, 1H), 6.64 (brs, 1H), 6.54 (d, *J* = 7.2 Hz, 1H), 5.98 (dd, *J* = 17.6, 10.8 Hz, 1H), 5.04 (dd, *J* = 17.6, 1.2 Hz, 1H), 4.97 (dd, *J* = 10.8, 1.2 Hz, 1H), 4.55 (d, J = 10.8, 1.2 Hz, 1H), 4.5 (

1.2 Hz, 1H), 4.50 (d, J = 1.2 Hz, 1H), 3.00 (d, J = 10.4 Hz, 1H), 2.77 (s, 3H), 2.58 (t, J = 10.8 Hz, 1H), 2.38 (td, J = 12.0, 3.6 Hz, 1H), 1.71-1.64 (m, 2H), 1.53 (s, 3H), 1.52-1.49 (m, 2H), 1.11 (s, 3H); ¹³**C** NMR (150 MHz, methanol- d_4) δ : 149.7, 148.5, 145.7, 144.4, 135.0, 121.4, 115.8, 115.8, 112.3, 111.7, 92.2, 61.7, 52.4, 50.1, 43.4, 37.3, 28.4, 19.7, 16.7. HRMS (ESI) m/z found 325.1772, calculated for C₁₉H₂₆O₃Na [M+Na]⁺ 325.1774. MS (EI) m/z (%): 302 (22), 271 (50), 257 (11), 202 (13), 198 (63), 166 (100), 123 (39). IR (KBr plate) V_{max}: 3339, 2972, 2929, 2856, 1642, 1565, 1520, 1444, 1375, 1281, 1194, 1105, 912, 886, 751 cm⁻¹.

Comparison of ¹³ H NMR	data for synthetic and	natural corypsoriol J (1c)
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Position	Natural corypsoriol J ^{a,12}	Synthetic corypsoriol J ^b	Δδ (ppm)
1	-	-	-
2	6.61 (d, <i>J</i> = 2.0 Hz)	6.64, brs	-0.03
3	-	-	-
4	-	-	
5	6.61 (d, <i>J</i> = 8.0 Hz)	6.64, brs	-0.03
6	6.50 (dd, <i>J</i> = 8.0, 2.0 Hz)	6.54, d (<i>J</i> = 7.2 Hz)	-0.04
7	2.03 (t, <i>J</i> = 9.4 Hz)	2.58, t (<i>J</i> = 10.8 Hz)	-0.55
8	3.75 (d, <i>J</i> = 9.4 Hz)	3.00, d (<i>J</i> = 10.4 Hz)	0.75
9	-	-	-
10α	1.70, m	1.65, m	0.05
10β	1.52, m	1.52, m	0
11α	1.92, m	1.70, m	0.22
11β	1.37, o ^c	1.49, m	-0.12
12	2.30 (td, <i>J</i> = 9.7, 4.1 Hz)	2.38, td (<i>J</i> = 12.0, 3.6 Hz)	-0.08
14	4.14, brs	4.55, d (<i>J</i> = 1.2 Hz)	-0.41
	4.08, brs	4.50, d (<i>J</i> = 1.2 Hz)	-0.42
15	1.36, s	1.53, s	-0.17
16	6.15 (dd, <i>J</i> = 18.0, 10.5 Hz)	5.98, dd (<i>J</i> = 17.6, 10.9 Hz)	0.17
17a	5.04 (dd, <i>J</i> = 10.5, 1.7 Hz)	4.97, dd (<i>J</i> = 10.8, 1.2 Hz)	0.07
17b	5.07 (dd, <i>J</i> = 18.0, 1.7 Hz)	5.04, dd (<i>J</i> = 17.6, 1.2 Hz)	0.03
18	1.28, s	1.11, s	-0.17
8-OCH₃	3.00, s	2.77, s	0.23

^a NMR data measured at 500 MHz in methanol- $d_{4;}$ ^b NMR data measured at 600 MHz in methanol- $d_{4;}$ ^c

"o" means overlapped.

Comparison of ¹³H NMR data for synthetic and natural corypsoriol J (1c)



Position	Natural corypsoriol J ^{a,12}	Synthetic corypsoriol J ^b	Δδ (ppm)
1	134.0	135.0	-0.1
2	115.7	115.8	-0.1
3	146.1	143.0	3.1
4	146.1	143.0	3.1
5	116.7	115.8	0.9
6	121.5	121.4	-1.4
7	59.4	50.1	9.3
8	88.2	92.2	-4
9	50.1	43.4	6.7
10	41.1	37.3	4.1
11	31.7	28.4	3.3
12	51.3	52.4	-1.1
13	149.2	148.5	0.7
14	110.4	112.3	-1.9
15	19.8	19.7	0.1
16	145.5	149.7	-4.2
17	111.9	111.7	0.2
18	28.4	16.7	11.7
8-OCH ₃	55.8	61.7	-5.9

^a NMR data measured at 125 MHz in methanol-d₄; ^b NMR data measured at 150 MHz in methanol-d₄.

3.5 Synthesis of proposed corypsoriol J's derivatives

3.5.1 Synthesis of compound 1c-1



To a suspension solution of the proposed corypsoriol J (**1c**, 13.3 mg, 0.044 mmol), Et₃N (12.2 μ L, 0.088 mmol) and DMAP (0.5 mg, 0.0044 mmol) in DCM (5.0 mL) was added TsCl

(16.4 mg, 0.099 mmol) at 0 °C under an argon atmosphere. The mixture was stirred at room temperature for 2 h, and quenched with H₂O. The resulting mixture was extracted with ethyl acetate (3×20 mL). The combined extracts were washed with brine, dried over Na₂SO₄, and filtered, and concentrated in vacuo. The crude residue was purified by flash column chromatography (petroleum ether/ethyl acetate = 50:1) on silica gel to give compound **1c-1** (25.2 mg, 94% yield) as a white foamy solid.



Compound 1c-1: White foamy solid; $[\alpha]_D^{20} = -1.3$ (c = 0.5 in CHCl₃). ¹H NMR (600 MHz, CDCl₃) δ : 7.58 (d, J = 8.4 Hz, 2H), 7.53 (d, J =7.8 Hz, 2H), 7.24 (d, J = 8.4 Hz, 2H), 7.19 (d, J = 8.4 Hz, 2H), 7.14 (brs, 1H), 7.13 (d, J = 8.4 Hz, 1H), 7.03 (dd, J = 8.4, 1.8 Hz, 1H),

5.93 (dd, J = 18.0, 10.8 Hz, 1H), 5.05 (dd, J = 17.4, 1.2 Hz, 1H), 5.01 (dd, J = 10.8, 1.2 Hz, 1H), 4.54 (brs, 1H), 4.52 (brs, 1H), 2.89 (d, J = 10.2 Hz, 1H), 2.72 (s, 3H), 2.70 (t, J = 10.2 Hz, 1H), 2.44 (s, 3H), 2.41 (s, 3H), 2.29 (td, J = 11.4, 3.6 Hz, 1H), 1.68-1.61 (m, 1H), 1.60-1.52 (m, 3H), 1.51 (s, 3H), 1.11 (s, 3H); ¹³**C** NMR (150 MHz, CDCl₃) δ : 147.7, 146.0, 145.53, 145.50, 143.3, 140.7, 139.1, 132.0, 131.7, 129.6, 129.5, 128.62, 128.57, 123.4, 112.3, 111.6, 90.3, 61.2, 50.9, 49.1, 42.2, 35.7, 26.7, 21.74, 21.70, 19.3, 16.2. HRMS (ESI) m/z found 633.1963, calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 633.1961. **MS** (EI) m/z (%): 610 (10), 565 (7), 423 (13), 355 (10), 319 (66), 155 (76), 135 (28), 91 (100).

3.5.2 Synthesis of compound 1c-2



To a suspension solution of the proposed corypsoriol J (**1c**, 20.0 mg, 0.066 mmol), Et₃N (18.0 μ L, 0.13 mmol) and DMAP (0.8 mg, 0.007 mmol) in DCM (5.0 mL) was added 4bromobenzoyl chloride (28.6 mg, 0.13 mmol) at 0 °C under an argon atmosphere. The mixture was stirred at room temperature for 2 h, and quenched with H₂O. The resulting mixture was extracted with ethyl acetate (3×20 mL). The combined extracts were washed with brine, dried over Na₂SO₄, and filtered, and concentrated in vacuo. The crude residue was purified by flash column chromatography (petroleum ether/ethyl acetate = 50:1) on silica gel to give compound **1c-2** (41.0 mg, 93% yield) as a white foamy solid.



Compound 1c-2: White foamy solid; $[\alpha]_D^{20} = 14.5$ (c = 0.5 in CHCl₃).

¹**H NMR** (600 MHz, CDCl₃) δ: 7.90 (d, J = 8.4 Hz, 2H), 7.88 (d, J = 8.4 Hz, 2H), 7.54 (d, J = 4.8 Hz, 2H), 7.53 (d, J = 4.8 Hz, 2H), 7.27 (d, J = 7.8 Hz, 1H), 7.25 (d, J = 1.8 Hz, 1H),

7.19 (dd, J = 8.4, 1.8 Hz, 1H), 5.97 (dd, J = 18.0, 10.8 Hz, 1H), 5.05 (dd, J = 17.4, 1.2 Hz, 1H), 5.00 (dd, J = 10.8, 1.2 Hz, 1H), 4.63 (brs, 1H), 4.61 (brs, 1H), 2.96 (d, J = 10.2 Hz, 1H), 2.85 (s, 3H), 2.83 (t, J = 10.8 Hz, 1H), 2.42 (td, J = 12.0, 3.6 Hz, 1H), 1.73-1.67 (m, 1H), 1.64-1.54 (m, 3H), 1.58 (s, 3H), 1.15 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ : 163.5, 147.9, 146.2, 141.7, 140.3, 131.9, 131.47, 131.45, 129.0, 127.8, 122.5, 122.4, 111.4, 90.6, 61.4, 50.6, 49.1, 42.3, 35.8, 27.1, 19.5, 16.4. HRMS (ESI) m/z found 691.0484, calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 691.0488. **MS** (EI) m/z (%): 668 (2), 532 (8), 183 (100), 179 (20), 157 (12), 147 (9), 104 (8), 93 (6), 68 (6).

3.5.3 Synthesis of compound 1c-3



To a suspension solution of the proposed corypsoriol J (**1c**, 12.0 mg, 0.04 mmol) and Cs_2CO_3 (19.6 mg, 0.06 mmol) in DMF (2.0 mL) was added CH_{2l_2} (4.8 µL, 0.06 mmol) at room temperature. The mixture was stirred at 110 °C for 20 h, and quenched with H₂O at room temperature. The resulting mixture was extracted with ethyl acetate (3×20 mL). The combined extracts were washed with brine, dried over Na₂SO₄, and filtered, and concentrated in vacuo. The crude residue was purified by flash column chromatography (petroleum ether/ethyl acetate = 20:1) on silica gel to give compound **1c-3** (8.2 mg, 65% yield) as a colorless oil.



Compound 1c-3: Colorless oil; $[\alpha]_D^{20} = 0.8$ (c = 0.5 in CHCl₃).

¹**H NMR** (600 MHz, CDCl₃) δ : 6.72 (brs, 1H), 6.71 (d, J = 8.4 Hz, 1H), 6.66 (d, J = 7.8 Hz, 1H), 5.96 (dd, J = 18.0, 10.8 Hz, 1H), 5.91 (s, 2H), 5.04 (d, J = 17.4 Hz, 1H), 4.97 (d, J = 10.8 Hz, 1H), 4.55 (brs,

2H), 2.89 (d, J = 10.2 Hz, 1H), 2.78 (s, 3H), 2.68 (t, J = 11.4 Hz, 1H), 2.33 (td, J = 12.0, 3.6 Hz, 1H), 1.69-1.59 (m, 2H), 1.57-1.52 (m, 2H), 1.55 (s, 3H), 1.12 (s, 3H); ¹³**C** NMR (150 MHz, CDCl₃) δ : 148.2, 147.2, 146.9, 145.6, 136.2, 122.1, 111.8, 111.2, 108.7, 107.7, 100.7, 90.7, 61.3, 51.0, 49.2, 42.3, 35.9, 27.2, 19.3, 16.4. HRMS (ESI) m/z found 337.1772,

calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 337.1774. **MS** (EI) m/z (%): 314 (34), 178 (90), 149 (69), 135 (49), 105 (41), 84 (100), 57 (77).

4. Total synthesis of 7β-aryl-12β-alkyl type meroterpenoid



Scheme S3 Total synthesis of (-)-7β,8α-hydroxy-12β-psoracorylifol F.

4.1 Synthesis of natural (-)- 7β , 8α -hydroxy-12 β -psoracorylifol F (1d)

4.1.1 Synthesis of compound 12



Under an argon atmosphere, to a solution of the precursor **2** (5.0 mg, 0.018 mmol) in THF (1.0 mL) was added lithium isopropylcyclohexylamide (LiCA, 0.5 mL, 0.09 mmol, 5.0 equiv.) and hexamethylphosphoramide (HMPA, 31.1 μ L, 0.18 mmol, 10.0 equiv.) at -70 °C and the resulting mixture was stirred for 2 h at the same temperature. The reaction mixture was added silica gel (400 mg) and stirred for 30 min. The crude reaction mixture was filtrated through celite and washed with ethyl acetate, then concentrated under vacuum. The crude product was purified by flash column chromatography to give the desired *cis*-product **12** (3.75 mg, 75% yield) as a colorless oil along with the starting material **2** (0.5 mg, 10% yield). The yield based on the recover starting material is 83% yield.

		\rightarrow	T °C, T	HF	
		2			12
Entry	Base (equiv.)	Additive	Temp.	Yield ^c	Remark
1 ^b	KOH (2.0)		rt to 70 °C	NR	Quenched by aq. NH₄Cl
2 ^b	NaOH (2.0)		rt to 70 °C	NR	Quenched by aq. NH₄Cl
3	LDA (2.0)		-78 °C to rt	NR	Quenched by aq. NH₄Cl
4	NaH (2.0)		0°C to 20 °C	29%	Quenched by cold aq. NH₄Cl
5	NaH (5.0)		0°C to 20 °C	30%	Quenched by cold aq. NH ₄ Cl
6	NaH (5.0)		100 °C	33%	Quenched by cold aq. NH ₄ Cl
7	LiHMDS (2.0)		-78 °C to 20 °C	30%	Quenched by cold aq. NH₄Cl
8	LiHMDS (4.0)		-78 °C to 20 °C	50%	Quenched by cold aq. NH ₄ Cl
9	LiCA (5.0)		-70 to 0 °C	NR	Quenched by aq. NH₄Cl
10	LTMP (5.0)		-70 to 0 °C	ND	Quenched by silica gel in 0 °C
11	LiCA (5.0)	HMPA	-70 °C	40%	Quenched by aq. NH₄Cl in -70 °C
12	LiCA (5.0)	HMPA	-70 °C	71%	Quenched by silica gel in -40 °C
13	LiCA (5.0)	HMPA	-70 °C	75%	Quenched by silica gel in -70 °C
14	LiCA (5.0)	HMPA	-70 °C	59%	Quenched by silica gel in -110 °C
15	LTMP (5.0)	HMPA	-70 °C	25%	Quenched by silica gel in -70 °C
16	<i>t</i> -BuLi (5.0)	HMPA	-70 °C	26%	Quenched by silica gel in -70 °C
17	<i>t</i> -BuLi (5.0)	HMPA	-70 °C	45%	Quenched by silica gel in -110 °C

Table S6 Optimization of the enolization/protonation conditions^a

Base, Additve

111

MeC

^aThe reactions were performed using precursor **2** (0.018 mmol), bsae (5.0 equiv.), additive (10.0 equiv.) in 1.0 mL THF;^b MeOH as solvent; ^cIsolated yield; rt = Hoom temperature; THF = Tetrahydrofuran; HMPA = Hexamethylphosphoramide; LDA = Lithium diisopropylamide; LiHMDS = Lithium bis(trimethylsilyl)amide; LiCA = lithium cyclohexylisopropylamide; LTMP = 2,2,6,6-tetramethylpiperidinyl-lithium; NR = No recation; ND = No detected.



Compound 12: Colorless oil; $[\alpha]_D^{23} = -78.8$ (c = 0.5 in MeOH). ¹H NMR (600 MHz, CDCl3) δ : 7.19 (d, *J* = 8.4 Hz, 2H), 6.81 (d, *J* = 9.0 Hz, 2H), 6.03 (dd, *J* = 18.0, 10.8 Hz, 1H), 5.19 (d, *J* = 10.2 Hz, 1H), 5.13 (d, *J* = 18.0 Hz, 1H), 4.96, (brs, 1H), 4.60, (brs, 1H),

4.15 (d, *J* = 5.4 Hz, 1H), 3.78 (s, 3H), 3.08 (dd, *J* = 9.0, 4.8 Hz, 1H), 2.43-2.18 (m, 1H), 2.92-1.93 (m, 2H), 1.91-1.87 (m, 1H), 1.70 (s, 3H), 1.18 (s, 3H); ¹³**C NMR** (150 MHz, CDCl3)
δ: 211.9, 158.0, 143.9, 143.5, 130.7, 129.1, 115.1, 114.3, 113.0, 55.2, 54.8, 52.2, 49.2, 34.9, 26.1, 24.9, 24.0. **HRMS** (ESI) m/z found 307.1668, calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 307.1669. **MS** (EI) m/z (%): 284 (13), 216 (16), 173 (26), 160 (46), 134 (24), 121 (100). **IR** (KBr plate) v_{max}: 3447, 2955, 2926, 2853, 1696, 1609, 1512, 1416, 1375, 1296, 1253, 1184, 1035, 1008, 918, 892, 829 cm⁻¹.

4.1.2 Synthesis of compound 13



To a solution of compound **12** (25.0 mg, 0.09 mmol) in methanol (2.0 mL) was added NaBH₄ (34.2 mg, 0.9 mmol) at -60 °C under an argon atmosphere. The mixture was stirred for 1 h. When the reaction completed (monitored by TLC), 10.0 mL of water were added and then stirred for another 1 h. The methanol of mixture was removed under the reduce pressure, and the resulting aqueous layer was extracted with ethyl acetate (3×20 mL). The combined organic phase was washed with brine, dried over Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure, and the resulting crude product (dr > 20:1) was purified by column chromatography on silica gel using petroleum ether-ethyl acetate (5:1) to afford compound **13** (21.9 mg, 85% yield).



Compound 13: Colorless oil; $[\alpha]_D^{21} = -62.7$ (c = 1.0 in MeOH). ¹H NMR (400 MHz, CDCl₃) δ : 7.67 (d, J = 8.8 Hz, 2H), 6.80 (d, J = 8.8 Hz, 2H), 5.85 (dd, J = 17.6, 10.8 Hz, 1H), 5.04 (dd, J = 10.8, 0.8 Hz, 1H), 4.99 (dd, J = 17.6, 0.8 Hz, 1H), 4.93 (brs, 1H), 4.92 (brs,

1H), 3.79 (s, 3H), 3.66 (s, 1H), 2.88 (brs, 1H), 2.55 (dd, J = 12.4, 3.2 Hz, 1H), 2.26-2.15 (m, 1H), 2.09-2.03 (m, 1H), 1.75 (s, 3H), 1.70-1.66 (m, 2H), 0.65 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃) δ : 158.5, 148.2, 146.1, 132.8, 130.3, 113.8, 112.33, 112.25, 83.4, 77.6, 55.0, 53.8, 42.4, 36.1, 25.4, 25.0, 15.5. **HRMS** (ESI) m/z found 309.1824, calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 309.1825. **MS** (EI) m/z (%): 286 (1), 189 (10), 166 (33), 135 (100), 108 (15), 77 (11), 55 (7). **IR** (KBr plate) v_{max}: 3478, 2959, 2925, 2855, 1611, 1514, 1460, 1414, 1255, 1184, 1086, 1035, 901, 827, 803 cm⁻¹.

4.1.3 Synthesis of natural (-)-7β,8α-hydroxy-12β-psoracorylifol F (1d)



To a suspension solution of compound **13** (7.0 mg, 0.025 mmol) and K₂CO₃ (1.4 mg, 0.01 mmol) in NMP (1.0 mL) was added PhSH (12.6 μ L, 0.12 mmol). The mixture was stirred under reflux for 14 h. The reaction was quenched by addition of HCI (1.0 moL/L) and extracted with ethyl acetate (3×10 mL). The combined extracts were washed with brine, dried (Na₂SO₄), and concentrated. The resulting crude product was purified by flash column chromatography on silica gel (petroleum ether/ethyl acetate, 10:1-2:1) to give (-)-7 β ,8 α -hydroxy-12 β -psoracorylifol F (**1d**) (6.3 mg, 93% yield).



(-)-7 β ,8 α -hydroxy-12 β -psoracorylifol F (1d): White solid; $[\alpha]_D^{25} = -$ 32.0 (c = 0.1 in MeOH), [lit¹⁰, $[\alpha]_D^{25} = -15.1$ (c = 0.1 in MeOH)]. **m.p.**: 164-167 °C.

¹**H NMR** (600 MHz, acetone- d_6) δ : 8.02 (s, 1H), 7.02 (d, J = 8.4 Hz, 2H), 6.70 (d, J = 8.4 Hz, 2H), 6.33 (dd, J = 18.0, 11.4 Hz, 1H), 5.18

(dd, J = 5.4, 1.2 Hz, 1H), 5.15 (dd, J = 12.0, 1.2 Hz, 1H), 4.49 (brs, 1H), 4.42 (brs, 1H), 3.43 (dd, J = 10.2, 4.8 Hz, 1H), 2.94 (s, 1H), 2.78 (s, 1H), 2.73 (d, J = 4.2 Hz, 1H), 2.55 (t, J = 11.4 Hz, 1H), 2.44 (td, J = 12.0, 4.2 Hz, 1H), 1.94-1.92 (m, 1H), 1.78-1.75 (m, 1H), 1.55-1.53 (m, 1H), 1.46 (s, 3H), 1.43-1.41 (m, 1H), 1.13 (s, 3H); ¹³**C** NMR (150 MHz, acetone- d_6) δ : 156.6, 148.7, 142.8, 133.7, 130.6, 115.8, 113.9, 111.8, 81.6, 52.4, 51.7, 43.1, 36.9, 29.7, 29.6, 19.8. HRMS (ESI) m/z found 295.1671, calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 295.1669. MS (EI) m/z (%): 272 (22), 136 (100), 123 (18), 107 (94), 81 (13), 68 (18). IR (KBr plate) v_{max}: 3444, 2923, 2852, 1514, 1461, 1376, 1229, 1015, 914, 819, 720 cm⁻¹.

Comparison of ¹H NMR data of synthetic and natural (-)-7 β ,8 α -hydroxy-12 β -psoracorylifol F (1d)



psoracorylifol F(1d)

Desition	Natural	Synthetic	Δδ
FUSILION	7β,8α-hydroxy-12β-psoracorylifol F ^{a,10}	7β,8α-hydroxy-12β-psoracorylifol F ^b	(ppm)
1	-	-	-
2,6	7.02, d (<i>J</i> = 8.5 Hz)	7.02, d (<i>J</i> = 8.4 Hz)	0.00
3,5	6.70, d (<i>J</i> = 8.5 Hz)	6.70, d (<i>J</i> = 8.4 Hz)	0.00
4	-	-	-
7	2.55, t (<i>J</i> = 11.5 Hz)	2.55, t (<i>J</i> = 11.4 Hz)	0.00
8	3.43, d (<i>J</i> = 10.0 Hz)	3.43, dd (<i>J</i> = 10.2, 4.8 Hz)	0.00
9	-	-	-
10 _{ax}	1.93, m	1.95-1.92, m (1.93, m)	0.00
10 _{eq}	1.54, m	1.55-1.53, m (1.54, m)	0.00
11 _{ax}	1.76, m	1.78-1.75, m (1.77, m)	-0.01
11 _{eq}	1.41, m	1.43-1.41, m (1.42, m)	-0.01
12	2.44, td (<i>J</i> = 12.0, 4.0 Hz)	2.44, td (<i>J</i> = 12.0, 4.2 Hz)	0.00
13	-	-	-
14 _{ax}	4.49, brs	4.49 (brs)	0.00
14 _{eq}	4.41, brs	4.42 (brs)	-0.01
15	1.46, s	1.46 (s)	0.00
16	6.33, dd (<i>J</i> = 16.0, 11.0 Hz)	6.33, dd (<i>J</i> = 18.0, 11.4 Hz)	0.00
17 _{ax}	5.18, dd (<i>J</i> = 4.0, 1.4 Hz)	5.18, dd (<i>J</i> =5.4, 1.2 Hz)	0.00
17 _{eq}	5.15, dd (<i>J</i> = 10.5, 1.5 Hz)	5.15, dd (<i>J</i> = 12.0, 1.2 Hz)	0.00
18	1.13, s	1.13 (s)	0.00

^a NMR data measured at 500 MHz in acetone-d₆; ^b NMR data measured at 600 MHz in acetone-d₆.

Comparison of ¹H NMR spectra of (-)-7β,8α-hydroxy-12β-psoracorylifol F (1d)



Comparison of ^{13}C NMR data of synthetic and natural (-)-7 β ,8 α -hydroxy-12 β -psoracorylifol F (1d)



psoracorylifol F(1d)

Position	Natural	Synthetic	Δδ
FUSILION	7β,8α-hydroxy-12β-psoracorylifol F ^{a,10}	7β,8α-hydroxy-12β-psoracorylifol F ^b	(ppm)
1	133.7	133.7	0.0
2,6	130.6	130.6	0.0
3,5	115.8	115.8	0.0
4	156.5	156.6	-0.1
7	51.7	51.7	0.0
8	81.6	81.6	0.0
9	43.0	43.1	-0.1
10	36.9	36.9	0.0
11	28.8	28.8	0.0
12	52.4	52.4	0.0
13	148.7	148.7	0.0
14	111.8	111.9	-0.1
15	19.8	19.8	0.0
16	142.8	142.8	0.0
17	113.9	113.9	0.0
18	28.9	28.9	0.0

^a NMR data measured at 125 MHz in acetone-*d*₆; ^b NMR data measured at 150 MHz in acetone-*d*₆.

Comparison of ¹³C NMR spectra of (-)-7 β ,8 α -hydroxy-12 β -psoracorylifol F (1d)



5. Collective total syntheses of 7β -aryl-12 α -alkyl type meroterpenoids



Scheme S4 Collective total syntheses of 7β -aryl-12 α -alkyl type meroterpenoids.

5.1 Synthesis of natural (-)-corypsoriol H (1e)

5.1.1 Synthesis of compounds 14 and 14'



Method:

To a solution of the precursor **2'** (5.0 mg, 0.018 mmol) in THF (1.0 mL) was added reductants (0.09 mmol, 5.0 equiv.) under an argon atmosphere at the noted temperature, respectively, and the mixture was stirred. When the reaction completed (monitored by TLC), the reaction mixture was quenched with a saturated aqueous solution of NH₄Cl, and was extracted with CH₂Cl₂ (3×20 mL). The combined organic phase was washed with brine, dried over Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure, and the dr value was measured by ¹H NMR (entries 1-24, Table S7 and entries 1-11, Table S8).

5.1.2 The optimization of the reduction of precursor 2'

5.1.2.1 The screening of reductants

Μ	eO O O O O O O O O O O O O O O O O O O	Reductants THF, T	MeO UH 14	MeO	OH 14'
Entry	Reductants	Solvents	Temp.	dr (14 : 14′)	Yield⁵
1	BH ₃	THF	rt		Decompose
2	BH₃·Me₂S	THF	-70 °C	1:1	38%
3	BH₃·Me₂S	THF	80 °C		ND
4	BH ₃ ·Me ₂ NH	THF	-70 °C	1:4	97%
5	BH₃·Me₂NH	THF	80 °C	1:3.6	92%
6	BH₃·Pyr	THF	rt	1.5:1	38%
7	NaBH ₄	THF	-70 °C	2.7:1	92%
8	NaBH ₄	THF	80 °C	2:1	95%
9	LiBH ₄	THF	rt	3:1	94%
10	ZnBH ₄	THF	rt	1.5:1	95%
11	9-BBN	THF	-70 °C to rt		NR
12	9-BBN	THF	80 °C		ND
13	Red-Al	THF	-70 °C to rt	1:1	50%
14	Red-Al	THF	80 °C	1.4:1	85%
15	DIBAL-H	THF	-70 °C to rt		NR
16	DIBAL-H	THF	80 °C	1:2	90%
17	Al(O ⁱ Pr) ₃	THF	rt		NR
18	LiAIH ₄	THF	-70 °C	12.5:1	96%
19	LiAIH ₄	THF	80 °C	7.9:1	95%
20	L-Selectride	THF	-70 °C to rt		NR
21	L-Selectride	THF	80 °C	1:3	trace
22	K-Selectride	THF	-70 °C to rt		NR
23	K-Selectride	THF	80 °C		ND
24	(<i>R</i>)-CBS, BH₃	THF	-40 °C		ND

Table S7. The screening of reductants^a

^aThe reactions were performed using precursor **2'** (0.018 mmol), reductants (5.0 equiv.) in 1.0 mL THF; ^bIsolated yield. NR = No recation; ND = No detected; 9-BBN = 9-Borabicyclo[3.3.1]nonane; Red-Al = Sodium bis(2-methoxyethoxy)aluminiumhydride; DIBAL-H = Diisobutylaluminium hydride; *L*-Selectride = Lithium triisobutylhydroborate; *K*-Selectride = Potassium *tri*-sec-butylborohydride;

5.1.2.2 The screening of solvents and temperature

Me	2'	Reductants Solvents, T	OH UNIT OH 14	MeO +	OH
Entry	Reductants	Solvents	Temp.	dr (14 : 14′)	Yield ^b
1	BH₃·Me₂NH	Et ₂ O	rt	1:4	93%
2	BH₃·Me₂NH	1,4-dioxane	rt	1:5	90%
3	BH₃·Me₂NH	DME	rt	1:6.6	97%
4	BH₃·Me₂NH	MTBE	rt	1:4	95%
5	BH₃·Me₂NH	Toluene	rt	1:4.5	92%
6	BH₃·Me₂NH	DCM	rt	1:5	90%
7	BH₃· <i>i</i> Pr₂NH	DME	rt	1:6	90%
8	BH₃·Me₂NH	DME	90 °C	1:4	95%
9	BH₃·Me₂NH	DME	-60 °C	1:6.5	98%
10	LiAIH ₄	THF	-100 °C	18:1	98%
11	LiAIH4	Et ₂ O	-100 °C	18:1	9 8%

Table S8. The screening of solvents and temperature^a

^aThe reactions were performed using the precursor 2' (0.018 mmol), Reductants (5.0 equiv.) in 1.0 mL solvent; ^bIsolated yield; rt = Room temperature; THF = Tetrahydrofuran; MTBE = tert-Butyl methyl ether, DCM = Dichloromethane; DME = 1,2-dimethoxyethane.



Method I (entry 10, Table S8):

To a solution of the precursor 2' (5.0 mg, 0.018 mmol) in THF (1.0 mL) was added LiAIH4 (3.4 mg, 0.09 mmol) under an argon atmosphere at -100 °C, and the mixture was stirred for 1 h. When the reaction completed (monitored by TLC), the reaction mixture was quenched with a saturated aqueous solution of NH₄Cl, and was extracted with CH₂Cl₂ (3×20 mL). The combined organic phase was washed with brine, dried over Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure, and the resulting crude product **14** and **14'** (dr = 18:1) was purified by flash column chromatography (silica gel, petroleum ether/ethyl acetate (50:1)) to give the desired product 14 (4.7 mg, 94% yield) as a white solid and its isomer 14' (0.2 mg, 4% yield).



Method II (entry 9, Table S8):

To a solution of the precursor **2'** (5.0 mg, 0.018 mmol) in DME (1.0 mL) was added $BH_3 \cdot Me_2NH$ (5.3 mg, 0.09 mmol) under an argon atmosphere at -60 °C, and the mixture was stirred for 48 h. When the reaction completed (monitored by TLC), the reaction mixture was quenched with a saturated aqueous solution of NH₄Cl, and was extracted with CH₂Cl₂ (3×20 mL). The combined organic phase was washed with brine, dried over Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure, and the resulting crude product **14'** and **14** (dr = 6.5:1) was purified by flash column chromatography (silica gel, petroleum ether/ethyl acetate (50:1)) to give the desired product **14'** (4.25 mg, 85% yield) as a white solid and its isomer **14** (0.65 mg, 13% yield).



Compound 14: Colorless needles; $[\alpha]_D^{25} = -32.0$ (c = 0.5 in MeOH); **m.p.**: 59-62 °C.

¹**H NMR** (600 MHz, CDCl₃) δ : 7.09 (d, J = 7.2 Hz, 2H), 6.83 (d, J = 8.4 Hz, 2H), 6.30 (dd, J = 18.0, 10.8 Hz, 1H), 5.26 (dd, J =11.4, 1.2

Hz, 1H), 5.19 (dd, J = 17.4, 1.2 Hz, 1H), 4.51 (brs, 1H), 4.50 (brs, 1H), 3.78 (s, 3H), 3.40 (d, J = 10.8 Hz, 1H), 2.60 (t, J = 11.4 Hz, 1H), 2.39 (td, J = 12.0, 3.6 Hz, 1H), 1.92-1.88 (m, 1H), 1.81-1.74 (m, 1H), 1.55-1.48 (m, 2H), 1.49 (s, 3H), 1.18 (s, 3H); ¹³**C** NMR (150 MHz, CDCl₃) δ : 158.2, 147.1, 140.6, 132.7, 129.4, 114.5, 113.9, 111.6, 81.2, 55.1, 51.2, 50.5, 41.7, 36.6, 27.7, 27.4, 19.4. HRMS (ESI) m/z found 309.1826, calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 309.1825. **MS** (EI) m/z (%): 286 (26), 150 (65), 121 (100), 91 (12), 81 (10), 68 (15), 57 (14). IR (KBr plate) v_{max}: 3532, 2928, 2892, 2851, 1637, 1611, 1511, 1446, 1243, 1182, 1012, 918, 892, 829, 812, 568 cm⁻¹.



Compound 14': Colorless needles; $[\alpha]_D^{25} = -5.6$ (c = 0.5 in MeOH); **m.p.**: 86-88 °C.

¹**H NMR** (400 MHz, CDCl₃) δ : 7.11 (d, J = 8.8 Hz, 2H), 6.83 (d, J = 8.8 Hz, 2H), 5.95 (dd, J = 17.6, 11.2 Hz, 1H), 5.20 (dd, J =10.8, 0.8

Hz, 1H), 5.15 (dd, *J* =18.0, 1.2 Hz, 1H), 4.67 (brs, 1H), 4.60 (brs, 1H), 3.78 (s, 3H), 3.46 (brs, 1H), 2.97 (dd, *J* =12.0, 1.6 Hz, 1H), 2.86 (td, *J* = 11.6, 3.6 Hz, 1H), 1.86-1.78 (m, 1H), 1.74-1.66 (m, 1H), 1.64-1.56 (m, 2H), 1.49 (s, 3H), 1.07 (s, 3H); ¹³**C NMR** (100 MHz, CDCl₃)

δ: 158.0, 148.1, 144.9, 134.5, 129.5, 113.7, 113.2, 111.8, 78.7, 55.1, 46.5, 41.6, 41.4, 30.5, 28.5, 27.0, 19.1. **HRMS** (ESI) m/z found 309.1824, calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 309.1825. **MS** (EI) m/z (%): 286 (19), 150 (62), 137 (18), 121 (100), 68 (12), 55 (7). **IR** (KBr plate) v_{max}: 3569, 2929, 2861, 2835, 1610, 1511, 1457, 1245, 1177, 1037, 913, 886, 840, 819 cm⁻¹.

5.1.2 Synthesis of natural (-)-corypsoriol H (1e)



To a suspension solution of alcohol **14** (28.6 mg, 0.1 mmol) and K_2CO_3 (5.5 mg, 0.04 mmol) in NMP (1.0 mL) was added PhSH (52.0 μ L, 0.5 mmol). The mixture was stirred under reflux for 14 h, cooled to room temperature, and quenched with H₂O. The resulting mixture was extracted with ethyl acetate (3×20 mL). The combined extracts were washed with brine, dried over Na₂SO₄, and concentrated. The crude residue was purified by flash column chromatography on silica gel (hexane/ethyl acetate) to give (-)-corypsoriol H (**1e**) (24.5 mg, 90% yield) as a colorless solid.



(-)-corypsoriol H (1e): Colorless needles; $[\alpha]_D^{25} = -32.0$ (c = 0.1 in MeOH), [lit¹², $[\alpha]_D^{25} = -120$ (c = 0.1 in MeOH)]. m.p.: 151-153 °C. ¹H NMR (600 MHz, methanol- d_4) δ : 7.01 (d, J = 8.4 Hz, 2H), 6.69 (d, J = 9.0 Hz, 2H), 6.30 (dd, J = 17.4, 10.8 Hz, 1H), 5.24 (d, J = 12.0 Hz, 1H), 5.18 (d, J = 18.0 Hz, 1H), 4.49 (brs, 1H), 4.46 (brs, 1H),

3.43 (d, J = 10.2 Hz, 1H), 2.58 (t, J = 10.8 Hz, 1H), 2.39 (td, J = 12.6, 4.2 Hz, 1H), 1.93 (dt, J = 13.8, 3.0 Hz, 1H), 1.84-1.77 (m, 1H), 1.55 (td, J = 13.8, 4.2 Hz, 1H), 1.48 (s, 3H), 1.46-1.43 (m, 1H), 1.15 (s, 3H); ¹³**C** NMR (150 MHz, methanol- d_4) δ : 156.7, 149.0, 142.5, 134.5, 130.8, 116.0, 114.8, 112.2, 82.4, 53.6, 51.9, 43.6, 37.9, 29.1, 28.7, 20.0. HRMS (ESI) m/z found 295.1670, calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 295.1669. **MS** (EI) m/z (%): 272 (22), 149 (39), 136 (100), 107 (98), 84 (90), 55 (50). **IR** (KBr plate) v_{max}: 3433, 3243, 2960, 2927, 1644, 1614, 1513, 1449, 1370, 1231, 1037, 915, 887, 818, 733 cm⁻¹.

Comparison of ¹H NMR data for synthetic and natural (-)-corypsoriol H (1e)

HO 4 3

	HO 4 1 1 1 1 1 1 1 1 1 1 1 1 1		
Position	Natural corypsoriol H ^{a,12}	Synthetic corypsoriol H ^b	Δ δ (ppm)
1	-	-	-
2,6	7.01, d (<i>J</i> = 8.5 Hz)	7.01, d (<i>J</i> = 8.4 Hz)	0.00
3,5	6.68, d (<i>J</i> = 8.5 Hz)	6.69, d (<i>J</i> = 8.4 Hz)	-0.01
4	-	-	-
7	2.58, t (<i>J</i> = 10.6 Hz)	2.58, t (<i>J</i> = 10.8 Hz)	0.00
8	3.43, d (<i>J</i> = 10.6 Hz)	3.43, d (<i>J</i> = 10.2 Hz)	0.00
9	-	-	-
10	1.55, td (<i>J</i> = 13.5, 3.4 Hz)	1.55, td (13.8, 4.2 Hz)	0.00
	1.94, dt (<i>J</i> = 13.5, 3.4 Hz)	1.93, dt (13.8, 3.0 Hz)	0.01
11	1.80, m	1.84-1.77, m (1.81, m)	-0.01
	1.44, m	1.47-1.43, m (1.45, m)	-0.01
12	2.39, td (<i>J</i> = 12.0, 3.8 Hz)	2.39, td (<i>J</i> = 12.6, 4.2 Hz)	0.00
13	-	-	-
14	4.50, brs	4.49, brs	0.01
	4.46, brs	4.46, brs	0.00
15	1.49, s	1.48, s	0.01
16	6.30, dd (<i>J</i> = 17.9, 11.0 Hz)	6.30, dd (<i>J</i> = 17.4, 10.8 Hz)	0.00
17	5.24, d (<i>J</i> = 11.0 Hz)	5.24, d (<i>J</i> = 12.0 Hz)	0.00
	5.18, d (<i>J</i> = 17.9 Hz)	5.18, d (<i>J</i> = 18.0 Hz)	0.00
18	1.15, s	1.15, s	0.00

^a NMR data measured at 400 MHz in methanol-d₄; ^b NMR data measured at 600 MHz in methanol-d₄.





Comparison of ¹³C NMR data for synthetic and natural (-)-corypsoriol H (1e)



Position	Natural corypsoriol H ^{a,12}	Synthetic corypsoriol H ^b	Δ δ (ppm)
1	134.5	134.5	0.0
2	130.9	130.8	0.1
3	116.1	116.0	0.1
4	156.9	156.7	0.2
5	116.1	116.0	0.1
6	130.9	130.8	0.1
7	53.7	53.6	0.1
8	82.5	82.4	0.1
9	43.7	43.6	0.1
10	37.9	37.9	0.0
11	28.7	28.7	0.0
12	52.0	51.9	0.1
13	149.0	149.0	0.0
14	112.2	112.2	0.0
15	20.0	20.0	0.0
16	142.6	142.5	0.1
17	114.8	114.8	0.0
18	29.2	29.1	0.1

^a NMR data measured at 100 MHz in methanol-*d*₄; ^b NMR data measured at 150 MHz in methanol-*d*₄.

Comparison of ¹³C NMR data for synthetic and natural (-)-corypsoriol H (1e)



5.2. Synthesis of natural (-)-psoracorylifol G (1f)



To a suspension solution of alcohol **14'** (19.4 mg, 0.068 mmol) and K_2CO_3 (3.8 mg, 0.027 mmol) in NMP (1.0 mL) was added PhSH (35.0 µL, 0.34 mmol). The mixture was stirred under reflux for 14 h, cooled to room temperature, and quenched with H₂O. The resulting mixture was extracted with ethyl acetate (3×20 mL). The combined extracts were washed with brine, dried over Na₂SO₄, and concentrated. The crude residue was purified by flash column chromatography on silica gel (hexane/ ethyl acetate) to furnish (-)-psoracorylifol G (**1f**) (16.6 mg, 90% yield).



(-)-psoracorylifol G (1f): Colorless needles; $[\alpha]_D^{25} = -4.7$ (c = 0.1 in MeOH), [lit¹³, $[\alpha]_D^{25} = -20.0$ (c = 0.1 in MeOH)]. **m.p.**: 129-131 °C.

¹H NMR (600 MHz, CDCl₃) δ: 7.05 (d, J = 7.8 Hz, 2H), 6.74 (d, J = 7.2 Hz, 2H), 5.94 (dd, J = 18.0, 11.4 Hz, 1H), 5.20 (d, J = 10.8 Hz, 1H),

5.15 (d, J = 18.0 Hz, 1H), 5.07 (brs, 1H), 4.66 (brs, 1H), 4.59 (brs, 1H), 3.47 (brs, 1H), 2.95 (d, J = 12.6 Hz, 1H), 2.83 (td, J = 10.8 Hz, 1H), 1.80 (t, J = 13.2 Hz, 1H), 1.70-1.56 (m, 4H), 1.48 (s, 3H), 1.07 (s, 3H); ¹³**C** NMR (150 MHz, CDCl₃) δ : 154.0, 148.0, 144.9, 134.5, 129.7, 115.2, 113.3, 111.9, 78.8, 46.4, 41.6, 41.4, 30.5, 28.5, 27.1, 19.0. HRMS (ESI) m/z found 295.1670, calculated for C₁₈H₂₄O₂Na [M+Na]⁺ 295.1669. **MS** (EI) m/z (%): 272 (21), 136 (90), 123 (21), 107 (100), 81 (13), 68 (18), 55 (12). **IR** (KBr plate) v_{max}: 3393, 2963, 2929, 2863, 1642, 1613, 1514, 1452, 1373, 1230, 1173, 1018, 914, 888, 818 cm⁻¹.



Position	Natural psoracorylifol G ^{a,13}	Synthetic psoracorylifol G ^b	Δδ (ppm)
2	7.06, d (<i>J</i> = 8.5 Hz)	7.05, d (<i>J</i> = 7.8 Hz)	0.01
3	6.75, d (<i>J</i> = 8.5 Hz)	6.74, d (<i>J</i> = 7.2 Hz)	0.01
5	6.75, d (<i>J</i> = 8.5 Hz)	6.74, d (<i>J</i> = 7.2 Hz)	0.01
6	7.06, d (<i>J</i> = 8.5 Hz)	7.05, d (<i>J</i> = 7.8 Hz)	0.01
7	2.95, dd (<i>J</i> = 12.0, 1.9 Hz)	2.95, dd (<i>J</i> = 12.6 Hz)	0.00
8	3.47, d (<i>J</i> = 1.9 Hz)	3.47, brs	0.00
10	1.81, m	1.80, t, (<i>J</i> = 13.2 Hz)	0.01
	1.60, m	1.62-1.56, m (1.57, m)	0.03
11	1.69, td (<i>J</i> = 12.0, 4.0 Hz)	1.71-1.64, m (1.68, m)	0.01
	1.59, m	1.62-1.56, m (1.57, m)	0.02
12	2.84, td (<i>J</i> = 12.0, 4.0 Hz)	2.83, td (<i>J</i> = 10.8, 4.2 Hz)	0.01
14	4.66, brs	4.66, brs	-0.01
	4.59, brs	4.59, brs	0.00
15	1.48, s	1.48, s	0.00
16	5.95, dd (<i>J</i> = 17.7, 11.1 Hz)	5.94, dd (<i>J</i> = 18.0, 11.4 Hz)	0.01
17	5.19, dd (<i>J</i> = 11.1, 1.0 Hz)	5.20, d (<i>J</i> = 10.8 Hz)	0.01
	5.15, dd (<i>J</i> = 17.7, 1.0 Hz)	5.15, d (<i>J</i> = 18.0 Hz)	0.00
18	1.07, s	1.07, s	0.00
8-OH	4.78, brs		

^a NMR data measured at 400 MHz in CDCl₃; ^b NMR data measured at 600 MHz in CDCl₃.



Position	Natural psoracorylifol G ^{a,13}	Synthetic psoracorylifol G ^b	Δ δ (ppm)
1	134.7	134.5	0.2
2	129.8	129.7	0.1
3	115.2	115.2	0.0
4	153.9	154.0	-0.1
5	115.2	115.2	0.0
6	129.8	129.7	0.1
7	46.5	46.4	0.1
8	78.7	78.8	-0.1
9	41.4	41.4	0.0
10	30.5	30.5	0.0
11	28.5	28.5	0.0
12	41.7	41.6	0.1
13	148.0	148.0	0.0
14	111.8	111.9	-0.1
15	19.0	19.0	0.0
16	144.9	144.9	0.0
17	113.3	113.3	0.0
18	27.0	27.1	-0.1

(-)-psoracorylifol G (1f)

^a NMR data measured at 100 MHz in CDCl₃; ^b NMR data measured at 150 MHz in CDCl₃.

5.3 Synthesis of natural (-)-8α-hydroxy-cyclobakuchiol C (1g)



To an ice-cold suspension of NaHCO₃ (21.0 mg, 0.25 mmol) and *m*-CPBA (16.1 mg, 80%, 0.75 mmol) in CH₂Cl₂ (1.0 mL) was added a solution of (-)-corypsoriol H (**1e**, 13.6 mg, 0.05 mmol) in CH₂Cl₂ (1.0 mL) at 0 °C. The mixture was stirred at 5 °C for about 4 h. When the

starting material disappeared, the reaction was quenched by addition of Me₂S (5.3μ L, 0.07 mmol) and saturated NaHCO₃ with vigorous stirring. The layers were separated and the aqueous layer was extracted with CH₂Cl₂ (3×20 mL). The combined organic layers were washed with brine, dried over Na₂SO₄, and concentrated to give a residue, which was used for the next step.

To an ice-cold solution of above oxirane in THF (2.0 mL) was added slowly LiAlH₄ (9.5 mg, 0.25 mmol) at 0 °C. The resulting mixture was stirred for 3 h under reflux. After reaction completion, the mixture was cooled to room temperature, quenched by addition of aqueous HCI (1.0 mol/L), and then extracted with ethyl acetate (3×20 mL). The combined organic layers were washed with saturated NaHCO₃ and brine, dried over Na₂SO₄, and concentrated. The residual oil was purified by flash column chromatography on silica gel (hexane/ethyl acetate) to give (-)-8 α -hydroxy-cyclobakuchiol C (**1g**) (11.3 mg, 78% yield from natural (-)-corypsoriol H (**1e**)).



(-)-8 α -hydroxy-cyclobakuchiol C (1g): Colorless needles; $[\alpha]_D^{25}$ = -33.4 (c = 0.1 in MeOH), [lit¹⁰, $[\alpha]_D^{25}$ = -18.5 (c = 0.1 in MeOH)]. m.p.: 123-125 °C.

¹**H NMR** (600 MHz, acetone- d_6) δ : 8.16 (s, 1H), 7.08 (brs, 2H), 6.74 (d, J = 7.8 Hz, 2H), 6.23 (dd J = 17.4, 10.8 Hz, 1H), 5.11 (dd, J =

10.8, 1.2 Hz, 1H), 5.09 (dd, J = 10.2, 1.8 Hz, 1H), 3.38 (dd, J = 10.2, 4.2 Hz, 1H), 2.48 (s, 1H), 2.46 (t, J = 10.8 Hz, 1H), 2.12 (s, 1H), 1.88-1.83 (m, 2H), 1.79-1.76 (m, 1H), 1.46-1.38 (m, 2H), 1.07 (s, 3H), 0.93 (s, 3H), 0.75 (s, 3H); ¹³**C** NMR (150 MHz, acetone- d_6) δ : 156.8, 142.5, 134.7, 130.9, 116.1, 113.7, 82.1, 73.2, 53.3, 50.5, 42.4, 36.5, 29.3, 28.6, 27.5, 24.0. HRMS (ESI) m/z found 313.1775, calculated for C₁₈H₂₆O₃Na [M+Na]⁺ 313.1774. **MS** (EI) m/z (%): 290 (5), 272 (6), 214 (38), 161 (32), 133 (82), 107 (100), 59 (55). **IR** (KBr plate) v_{max}: 3546, 2956, 2930, 2868, 1616, 1543, 1513, 1455, 1383, 1256, 1227, 1172, 1010, 928, 907, 827 cm⁻¹.

Comparison of ¹H NMR data of synthetic and natural (-)-8 α -hydroxy-cyclobakuchiol C (1g)



Position	Natural 8α-hydroxy-cyclobakuchiol C ^{a,10}	Synthetic 8α-hydroxy-cyclobakuchiol C ^b	Δδ (ppm)
1	-	-	-
2,6	7.07, d (<i>J</i> = 5.5 Hz)	7.08, d (<i>J</i> = 6.6 Hz)	-0.01
3,5	6.74, d (<i>J</i> = 8.5 Hz)	6.74, d (<i>J</i> = 7.8 Hz)	0.00
4	-	-	-
7	2.45, t (<i>J</i> = 11.0 Hz)	2.46, t (<i>J</i> = 10.8 Hz)	-0.01
8	3.38, d (<i>J</i> = 10.0 Hz)	3.38, dd (<i>J</i> = 10.2, 4.2 Hz)	0.00
9	-	-	-
10	1.89, m	1.88-1.83, m (1.86, m)	0.03
	1.43, m	1.46-1.38, m (1.42, m)	0.01
11	1.79, m	1.79-1.76, m (1.78, m)	0.01
	1.42, m	1.46-1.38, m (1.42, m)	0.00
12	1.89, m	1.85-1.83, m (1.84, m)	0.05
13	-	-	-
14	0.95, s	0.93, s	0.02
15	0.72, s	0.75, s	-0.03
16	6.23, dd (<i>J</i> = 17.5, 11.5 Hz)	6.23, dd (<i>J</i> = 17.4, 10.8 Hz)	0.00
17	5.13, dd (<i>J</i> = 17.5, 1.5 Hz)	5.11, dd (<i>J</i> = 10.8, 1.2 Hz)	0.02
	5.10, dd (<i>J</i> = 11.5, 1.5 Hz)	5.09, dd (<i>J</i> = 10.2, 1.8 Hz)	0.01
18	1.07, s	1.07, s	0.00

^a NMR data measured at 500 MHz in acetone-d₆; ^b NMR data measured at 600 MHz in acetone-d₆.

Comparison of ¹H NMR spectra of (-)-8 α -hydroxy-cyclobakuchiol C (1g)



Comparison of ^{13}C NMR data of synthetic and natural (-)-8 α -hydroxy-cyclobakuchiol C (1g)



Desition	Natural	Synthetic	Δδ
Position	8α -hydroxy-cyclobakuchiol C ^{a,10}	8α-hydroxy-cyclobakuchiol C ^b	(ppm)
1	134.5	134.7	-0.2
2	130.3	130.5	-0.2
3	116.1	116.1	0.0
4	156.8	156.8	0.0
5	116.1	116.1	0.0
6	130.3	130.9	-0.6
7	50.4	50.5	-0.1
8	82.1	82.1	0.0
9	42.4	42.4	0.0
10	36.6	36.5	0.1
11	23.8	24.0	-0.2
12	53.2	53.3	-0.1
13	73.5	73.2	0.3
14	26.9	27.5	-0.6
15	29.4	29.3	0.1
16	142.4	142.5	-0.1
17	114.0	113.7	0.3
18	28.6	28.6	0.0

^a NMR data measured at 125 MHz in acetone-*d*₆; ^b NMR data measured at 150 MHz in acetone-*d*₆.

Comparison of ¹³C NMR spectra of (-)-8α-hydroxy-cyclobakuchiol C (1g)



5.4. Synthesis of the proposed corypsoriol I (1h)

5.4.1 Synthesis of compound 15



To a suspension solution of alcohol **14** (70.0 mg, 0.25 mmol) and NaH (39.2 mg, 60%, 0.98 mmol) in THF (2.0 mL) was added MeI (61.0 μ L, 0.98 mmol). The mixture was stirred at room temperature for 20 h, and quenched with H₂O. The resulting mixture was extracted with ethyl acetate (3×20 mL). The combined organic phase was washed with brine, dried over Na₂SO₄, and filtered, and concentrated in vacuo. The crude residue was purified by flash column chromatography (petroleum ether/ethyl acetate = 10:1) on silica gel to give methylether **15** (71.3 mg, 95% yield) as a colorless oil.



Compound 15: Colorless oil; $[\alpha]_D^{25} = -61.4$ (c = 0.5 in MeOH).

¹**H NMR** (400 MHz, CDCl₃) δ : 7.11 (d, J = 8.4 Hz, 2H), 6.80 (d, J = 8.8 Hz, 2H), 6.23 (dd, J = 18.0, 11.2 Hz, 1H), 5.21 (dd, J = 11.2, 1.6 Hz, 1H), 5.13 (dd, J = 18.0, 1.6 Hz, 1H), 4.54 (brs, 1H), 4.51 (brs,

1H), 3.78 (s, 3H), 2.83 (d, J = 10.4 Hz, 1H), 2.74 (s, 3H), 2.64 (t, J = 11.6 Hz, 1H), 2.45 (td, J = 12.0, 3.6 Hz, 1H), 1.91-1.87(m, 1H), 1.77-1.65 (m, 1H), 1.52-1.43 (m, 2H), 1.49 (s, 3H), 1.13 (s, 3H); ¹³**C** NMR (100 MHz, CDCl₃) δ : 157.7, 147.2, 141.4, 134.1, 129.8, 113.8, 113.1, 111.7, 92.7, 61.4, 55.0, 51.0, 49.4, 43.1, 36.7, 27.8, 27.6, 19.2. HRMS (ESI) m/z found 323.1981, calculated for C₂₀H₂₈O₂Na [M+Na]⁺ 323.1982. **MS** (EI) m/z (%):300 (21), 164 (100), 149 (31), 121 (80), 68 (22), 55 (24). **IR** (KBr plate) v_{max}:3402, 3074, 2970, 2929, 2862, 2830,1644, 1612, 1512, 1456, 1247, 1178, 1106, 1039, 912, 886, 818 cm⁻¹.

5.4.2 Synthesis of phenol 16



To a suspension solution of methylether **15** (42.0 mg, 0.14 mmol) and K_2CO_3 (7.7 mg, 0.056 mmol) in NMP (2.0 mL) was added PhSH (44.0 µL, 0.42 mmol). The mixture was stirred under reflux for 14 h. The reaction was quenched by addition of HCI (1.0 mol/L) and extracted with ethyl acetate (3×20 mL). The combined organic layers were washed with brine, dried (Na₂SO₄), and concentrated. The resulting crude product was purified by flash

column chromatography on silica gel (hexane/ethyl acetate) to furnish phenol **16** (38.0 mg, 95% yield).



Compound 16: White solid; $[\alpha]_D^{21} = -56.7$ (c = 0.1 in MeOH); **m.p.**: 164.2-165.3 °C.

¹**H NMR** (400 MHz, CDCl₃) δ : 7.06 (d, J = 8.0 Hz, 2H), 6.72 (d, J = 8.8 Hz, 2H), 6.22 (dd, J = 18.0, 11.2 Hz, 1H), 5.20 (dd, J = 11.2, 0.8

Hz, 1H), 5.12 (dd, J = 18.0, 1.2 Hz, 1H), 4.82 (s, 1H), 4.54 (brs, 1H), 4.51 (brs, 1H), 2.84 (d, J = 10.4 Hz, 1H), 2.77 (s, 3H), 2.64 (t, J = 11.6 Hz, 1H), 2.43 (td, J = 12.0, 4.0 Hz, 1H), 1.90-1.85 (m, 1H), 1.77-1.65 (m, 1H), 1.51-1.43 (m, 2H),1.49 (s, 3H), 1.13 (s, 3H); ¹³**C NMR** (150 MHz, CDCl₃) δ : 153.7, 147.2, 141.3, 134.3, 130.0, 114.8, 113.9, 111.7, 92.8, 61.4, 51.1, 49.4, 43.1, 36.8, 27.8, 27.6, 19.2. **HRMS** (ESI) m/z found 309.1822, calculated for C₁₉H₂₆O₂Na [M+Na]⁺ 309.1825; **MS** (EI) m/z (%): 286 (9), 242 (6), 203 (6), 150 (100), 107 (56), 84 (40), 57 (37). **IR** (KBr plate) v_{max}: 3421, 3079, 2960, 2925, 2853, 1712, 1616, 1516, 1454, 1374, 1223, 1101, 1016, 910, 886, 800 cm⁻¹.

5.4.3 Synthesis of the proposed corypsoriol I (1h)



To a solution of phenol **16** (18.0 mg, 0.06 mmol) in CHCl₃/MeOH (1.2 mL) (3:1 v/v) was added IBX (20.0 mg, 0.07 mmol) at 0 °C under an argon atmosphere. A yellow-to-orange color developed and the mixture was stirred for 2 h, then NaBH₄ (21.0 mg, 0.54 mmol) was added at 0 °C under vigorous stirring until the color disappeared (usually within 30 min).¹¹ The reaction was quenched by addition of HCl (1.0 mol/L) and extracted with ethyl acetate (3×20 mL). The combined extracts were washed with brine, dried over Na₂SO₄, and filtered. The filtrate was concentrated under reduced pressure, and the residue was subjected to column chromatography on silica gel using ethyl acetate-petroleum ether (1:1) to give the proposed corypsoriol I (**1h**) (16.9 mg, 93% yield).



Proposed corypsoriol I (1h): White solid; $[α]_D^{25} = -50.0$ (c = 0.1 in MeOH), [lit¹², $[α]_D^{25} = -60.0$ (c = 0.1 in MeOH)]. **m.p.**: 175-177 °C. ¹**H NMR** (600 MHz, methanol-*d*₄) δ: 6.65 (d, *J* = 8.4 Hz, 2H), 6.53 (brs, 1H), 6.19, (dd, *J* = 18.0, 11.4 Hz, 1H), 5.17 (dd, *J* = 10.8, 1.2 Hz, 1H), 5.12 (dd, *J* = 18.0, 1.2 Hz, 1H), 4.52 (brs, 1H), 4.48 (brs, 1H), 2.95 (d, *J* = 18.0, 1.2 Hz, 1H), 4.52 (brs, 1H), 4.48 (brs, 1H), 2.95 (d, *J* = 18.0, 1.2 Hz, 1H), 4.52 (brs, 1H), 4.48 (brs, 1H), 2.95 (d, *J* = 18.0, 1.2 Hz, 1H), 4.52 (brs, 1H), 4.48 (brs, 1H), 2.95 (d, *J* = 18.0, 1.2 Hz, 1H), 4.52 (brs, 1H), 4.48 (brs, 1H), 2.95 (d, *J* = 18.0, 1.2 Hz, 1H), 4.52 (brs, 1H), 4.48 (brs, 1H), 4.95 (d, *J* = 18.0, 1.2 Hz, 1H), 4.52 (brs, 1H), 4.48 (brs, 1H), 4.95 (d, *J* = 18.0, 1.2 Hz, 1H), 4.52 (brs, 1H), 4.48 (brs, 1H), 4.95 (d, *J* = 18.0, 1.2 Hz, 1H), 4.52 (brs, 1H), 4.48 (brs, 1H), 4.95 (d, *J* = 18.0, 1.2 Hz, 1H), 4.52 (brs, 1H), 4.48 (brs, 1H), 4.95 (d, *J* = 18.0, 1.2 Hz, 1H), 4.52 (brs, 1H), 4.48 (brs, 1H), 4.95 (d, *J* = 18.0, 1.2 Hz, 1H), 4.52 (brs, 1H), 4.52 (brs, 1H), 4.54 (brs, 1H), 4.55 (brs,

J = 10.2 Hz, 1H), 2.82 (s, 3H), 2.51 (t, J = 11.4 Hz, 1H), 2.44 (td, J = 12.0, 4.2 Hz, 1H),

1.89 (dt, J = 13.8, 3.0 Hz, 1H), 1.75-1.68 (m, 1H), 1.54-1.49 (m, 1H), 1.48 (s, 3H), 1.43-1.39 (m, 1H), 1.11 (s, 3H); ¹³**C NMR** (150 MHz, methanol-*d*₄) δ : 148.6, 145.7, 144.4, 142.9, 135.0, 115.8, 114.2, 112.2, 94.0, 61.8, 52.6, 51.2, 44.3, 37.5, 28.9, 28.6, 19.6. **HRMS** (ESI) m/z found 325.1773, calculated for C₁₉H₂₆O₃Na [M+Na]⁺ 325.1774. **MS** (EI) m/z (%): 302 (19), 257 (9), 202 (10), 166 (100), 123 (37), 68 (18), 55 (17). **IR** (KBr plate) v_{max}: 3368, 2931, 2864, 2829, 1641, 1605, 1518, 1443, 1375, 1278, 1190, 1104, 911, 888, 810 cm⁻¹.

Comparison of ¹H NMR data of synthetic and natural corypsoriol I (1h)

HO $\frac{4}{2}$ OMe ₁₈ HO $\frac{3}{2}$ 7 9 17	
proposed corypsoriol I (1h)	No a la la

Position	Natural corypsoriol I ^{a,12}	Synthetic corypsoriol I ^b	Δδ (ppm)
1	-	-	-
2	6.64 (d, <i>J</i> = 2.0 Hz)	6.64 (d, <i>J</i> = 8.4 Hz)	0.00
3	-	-	-
4	-	-	-
5	6.62 (d, <i>J</i> = 8.0 Hz)	6.64 (d, <i>J</i> = 8.4 Hz)	-0.02
6	6.52 (dd, <i>J</i> = 8.0, 2.0 Hz)	6.53 (d, <i>J</i> = 7.2 Hz)	-0.01
7	2.12 (t, <i>J</i> = 9.2 Hz)	2.51 (t, <i>J</i> = 11.4 Hz)	-0.39
8	3.84 (d, <i>J</i> = 9.6 Hz)	2.95 (d, <i>J</i> = 10.4 Hz)	0.99
9	-	-	-
10α	1.34, m	1.89 (dt, <i>J</i> = 13.8, 3.0 Hz)	-0.55
10β	1.77, m	1.54-1.49, m (1.52, m)	0.25
11α	1.92, m	1.75-1.68, m (1.72, m)	-0.20
11β	1.34, o ^c	1.43-1.39, m (1.41, m)	-0.07
12	2.36 (td, <i>J</i> = 9.7, 4.1)	2.44 (td, <i>J</i> = 12.0, 4.2 Hz)	-0.08
13	-	-	-
14	4.14, brs	4.52, brs	-0.38
	4.09, brs	4.48, brs	-0.39
15	1.37, s	1.48, s	-0.11
16	6.16 (dd, <i>J</i> = 17.8, 10.8 Hz)	6.19 (dd, <i>J</i> = 18.0, 11.4 Hz)	-0.03
17α	4.79 (dd, <i>J</i> = 10.8, 1.2 Hz)	5.12 (dd, <i>J</i> = 10.8, 1.2 Hz)	-0.33
17β	4.90 (dd, <i>J</i> = 17.7, 1.2 Hz)	5.17 (dd, <i>J</i> = 18.0, 1.2 Hz)	-0.27
18	1.14, s	1.11, s	0.03
8-OCH ₃	3.02, s	2.82, s	0.20

^a NMR data measured at 400 MHz in methanol-*d*₄; ^b NMR data measured at 600 MHz in methanol-*d*₄; ^c "o" means overlapped.

Comparison of ¹³C NMR data of synthetic and natural (-)-corypsoriol I (1h)



Position	Natural corypsoriol la,12	Synthetic corypsoriol I ^b	Δδ (ppm)
1	134.0	135.0	-1.0
2	115.8	115.8	0.0
3	146.2	145.7	0.5
4	146.1	144.4	1.7
5	116.6	115.8	0.8
6	121.4	115.8	
7	58.2	51.2	7.0
8	87.8	94.0	-6.2
9	49.6	44.3	5.3
10	41.3	28.9	-12.4
11	31.5	37.5	-6.0
12	50.2	52.6	-2.4
13	149.4	148.6	0.8
14	110.3	112.2	-1.9
15	20.0	19.6	-0.4
16	151.6	142.9	8.7
17	109.1	114.2	-5.1
18	20.4	28.6	-8.2
8-OCH ₃	55.9	61.8	-5.9

^a NMR data measured at 100 MHz in methanol-*d*₄; ^b NMR data measured at 150 MHz in methanol-*d*₄.

6. X-Ray crystallography Data

6.1 X-Ray crystallography of compound 8

The crystal of compound **8** for X-ray diffraction study was obtained through the dissolving of compound in ethyl acetate and petroleum ether, followed by slow evaporation of the solvent at room temperature. X-ray data collections were performed in an Agilent Super Nova.



Figure S1. X-Ray coordinate of compound **8** (CCDC 2226753). Displacement ellipsoids are scaled to the 30% probability level.

Identification code	Compound 8
Empirical formula	C19H26O2
Formula weight	286.40
Temperature/K	149.95(10)
Crystal system	orthorhombic
Space group	P212121
a/Å	6.55997(12)
b/Å	14.3262(3)
c/Å	17.4645(4)
α/°	90
β/°	90
γ/°	90
Volume/Å ³	1641.30(6)
Z	4
ρ _{calc} g/cm ³	1.159
µ/mm ⁻¹	0.569
F(000)	624.0
Crystal size/mm ³	0.13 × 0.13 × 0.12
Radiation	Cu Kα (λ = 1.54184)
2O range for data collection/°	7.982 to 154.694
Index ranges	-7 ≤ h ≤ 3, -17 ≤ k ≤ 18, -21 ≤ l ≤ 21
Reflections collected	5454
Independent reflections	2925 [$R_{int} = 0.0741$, $R_{sigma} = 0.0814$]
Data/restraints/parameters	2925/0/194
Goodness-of-fit on F ²	1.043
Final R indexes [I>=2σ (I)]	R ₁ = 0.0541, wR ₂ = 0.1472
Final R indexes [all data]	R ₁ = 0.0580, wR ₂ = 0.1513
Largest diff. peak/hole / e Å ⁻³	0.33/-0.32
Flack parameter	0.04(18)

6.2 X-Ray crystallography of natural (-)-7 α ,8 β -hydroxy-12 β - cyclobakuchiol C (1b)

The crystal of natural (-)- 7α ,8 β -hydroxy-12 β -cyclobakuchiol C (**1b**) for X-ray diffraction study was obtained through the dissolving of compound in ethyl acetate and petroleum ether, followed by slow evaporation of the solvent at room temperature. X-ray data collections were performed in an Agilent Super Nova.



Figure S1. X-Ray coordinate of natural (-)-7 α ,8 β -hydroxy-12 β -cyclobakuchiol C (1b)

(CCDC 2226760). Displacement ellipsoids ar	re scaled to the 30% probability level.
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Identification code	(-)-7α,8β-hydroxy-12β-cyclobakuchiol C		
	(1b)		
Empirical formula	C37H56O7		
Formula weight	612.81		
Temperature/K	150.00(10)		
Crystal system	monoclinic		
Space group	P21		
a/Å	15.3362(3)		
b/Å	6.95130(12)		
c/Å	16.3195(3)		
α/°	90		
β/°	101.0819(18)		
γ/°	90		
Volume/Å ³	1707.32(5)		
Z	2		
ρcalcg/cm ³	1.192		
µ/mm⁻¹	0.643		
F(000)	668.0		
Crystal size/mm ³	0.14 × 0.13 × 0.12		
Radiation	Cu Kα (λ = 1.54184)		
2O range for data collection/°	5.518 to 154.338		
Index ranges	-19 ≤ h ≤ 19, -7 ≤ k ≤ 8, -20 ≤ l ≤ 20		
Reflections collected	16825		
Independent reflections	6125 [R _{int} = 0.0500, R _{sigma} = 0.0468]		
Data/restraints/parameters	6125/1/445		
Goodness-of-fit on F ²	1.060		
Final R indexes [I>=2σ (I)]	R ₁ = 0.0401, wR ₂ = 0.1058		
Final R indexes [all data]	R ₁ = 0.0421, wR ₂ = 0.1075		
Largest diff. peak/hole / e Å ⁻³	0.22/-0.23		
Flack parameter	0.22(13)		

6.3 X-Ray crystallography of compound 11

The crystal of compound **11** for X-ray diffraction study was obtained through the dissolving of compound in petroleum ether and ethyl acetate, followed by slow evaporation of the solvent at room temperature. X-ray data collections were performed in an Agilent Super Nova.



Figure S1. X-Ray coordinate of compound **11** (CCDC 2226754). Displacement ellipsoids are scaled to the 30% probability level.

Identification code	Compound 11
Empirical formula	C ₁₉ H ₂₆ O ₂
Formula weight	286.40
Temperature/K	149.99(10)
Crystal system	orthorhombic
Space group	P212121
a/Å	6.1934(2)
b/Å	13.5942(4)
c/Å	20.6761(6)
α/°	90
β/°	90
γ/°	90
Volume/Å ³	1740.81(9)
Z	4
$ ho_{calc}g/cm^3$	1.093
µ/mm ⁻¹	0.536
F(000)	624.0
Crystal size/mm ³	0.11 × 0.03 × 0.02
Radiation	Cu Kα (λ = 1.54184)
2Θ range for data collection/°	7.784 to 155.622
Index ranges	-5 ≤ h ≤ 7, -16 ≤ k ≤ 16, -24 ≤ l ≤ 26
Reflections collected	8246
Independent reflections	$3406 [R_{int} = 0.0619, R_{sigma} = 0.0610]$
Data/restraints/parameters	3406/0/194
Goodness-of-fit on F ²	1.103
Final R indexes [I>=2σ (I)]	$R_1 = 0.0519$, $wR_2 = 0.1406$
Final R indexes [all data]	$R_1 = 0.0615$, $wR_2 = 0.1477$
Largest diff. peak/hole / e Å ⁻³	0.26/-0.23
Flack parameter	-0.1(2)

6.4 X-Ray crystallography of compound 14

The crystal of compound **14** for X-ray diffraction study was obtained through the dissolving of compound in petroleum ether and ethyl acetate, followed by slow evaporation of the solvent at room temperature. X-ray data collections were performed in an Agilent Super Nova.



Figure S1. X-Ray coordinate of compound **14** (CCDC 2226756). Displacement ellipsoids are scaled to the 30% probability level.

Identification code	Compound 14
Empirical formula	C ₁₉ H ₂₆ O ₂
Formula weight	286.40
Temperature/K	149.99(10)
Crystal system	orthorhombic
Space group	P212121
a/Å	6.02770(10)
b/Å	10.7851(2)
c/Å	25.2404(4)
α/°	90
β/°	90
γ/°	90
Volume/Å ³	1640.86(5)
Z	4
$ ho_{calc}g/cm^3$	1.159
µ/mm ⁻¹	0.569
F(000)	624.0
Crystal size/mm ³	0.16 × 0.15 × 0.12
Radiation	Cu Kα (λ = 1.54184)
2O range for data collection/°	7.004 to 154.842
Index ranges	$-4 \le h \le 7, -13 \le k \le 13, -31 \le l \le 26$
Reflections collected	8138
Independent reflections	$3260 [R_{int} = 0.0874, R_{sigma} = 0.0730]$
Data/restraints/parameters	3260/0/194
Goodness-of-fit on F ²	1.068
Final R indexes [I>=2σ (I)]	$R_1 = 0.0792, wR_2 = 0.1914$
Final R indexes [all data]	$R_1 = 0.0814$, $wR_2 = 0.1933$
Largest diff. peak/hole / e Å ⁻³	0.45/-0.52
Flack parameter	-0.1(2)

6.5 X-Ray crystallography of compound 14'

The crystal of compound **14'** for X-ray diffraction study was obtained through the dissolving of compound in petroleum ether and ethyl acetate, followed by slow evaporation of the solvent at room temperature. X-ray data collections were performed in an Agilent Super Nova.



Figure S1. X-Ray coordinate of compound **14'** (CCDC 2226759). Displacement ellipsoids are scaled to the 30% probability level.

Identification code	compound 14'
Empirical formula	C ₃₈ H ₅₂ O ₄
Formula weight	572.79
Temperature/K	150.00(10)
Crystal system	monoclinic
Space group	C2
a/Å	25.3714(5)
b/Å	6.05801(10)
c/Å	23.3168(5)
α/°	90
β/°	112.787(2)
γ/°	90
Volume/Å ³	3304.08(12)
Z	4
ρcalcg/cm ³	1.151
µ/mm⁻¹	0.565
F(000)	1248.0
Crystal size/mm ³	0.25 × 0.16 × 0.15
Radiation	Cu Kα (λ = 1.54184)
2⊖ range for data collection/°	4.11 to 154.916
Index ranges	-31 ≤ h ≤ 31, -6 ≤ k ≤ 7, -26 ≤ l ≤ 29
Reflections collected	19488
Independent reflections	6149 [R _{int} = 0.0678, R _{sigma} = 0.0565]
Data/restraints/parameters	6149/2/393
Goodness-of-fit on F ²	1.069
Final R indexes [I>=2σ (I)]	$R_1 = 0.0446$, $wR_2 = 0.1146$
Final R indexes [all data]	R ₁ = 0.0474, wR ₂ = 0.1169
Largest diff. peak/hole / e Å ⁻³	0.18/-0.25
Flack parameter	0.03(18)

6.6 X-Ray crystallography of natural (-)-corypsoriol H (1e)

The crystal of (-)-corypsoriol H (**1e**) for X-ray diffraction study was obtained through the dissolving of compound in MeOH, CH_2Cl_2 and petroleum ether, followed by slow evaporation of the solvent at room temperature. X-ray data collections were performed in an Agilent Super Nova.



Figure S1. X-Ray coordinate of (-)-corypsoriol H (**1e**) (CCDC 2226749). Displacement ellipsoids are scaled to the 30% probability level.

Identification code	(-)-corypsoriol H (1e)
Empirical formula	C ₁₈ H ₂₅ O ₂
Formula weight	281.38
Temperature/K	150.00(10)
Crystal system	monoclinic
Space group	C2
a/Å	17.9138(6)
b/Å	6.1604(2)
c/Å	15.2956(6)
α/°	90
β/°	106.418(4)
γ/°	90
Volume/Å ³	1619.14(10)
Z	4
ρ _{calc} g/cm ³	1.154
µ/mm ⁻¹	0.592
F(000)	612.0
Crystal size/mm ³	0.3 × 0.26 × 0.24
Radiation	Cu Kα (λ = 1.54184)
2O range for data collection/°	6.024 to 154.302
Index ranges	$-22 \le h \le 22, -7 \le k \le 7, -18 \le l \le 19$
Reflections collected	7068
Independent reflections	2909 [$R_{int} = 0.0811$, $R_{sigma} = 0.0806$]
Data/restraints/parameters	2909/1/193
Goodness-of-fit on F ²	1.041
Final R indexes [I>=2σ (I)]	$R_1 = 0.0562, wR_2 = 0.1412$
Final R indexes [all data]	$R_1 = 0.0626$, $wR_2 = 0.1477$
Largest diff. peak/hole / e Å ⁻³	0.31/-0.27
Flack parameter	0.0(2)

6.7 X-Ray crystallography of proposed corypsoriol I (1h)

The crystal of proposed (-)-corypsoriol I (**1h**) for X-ray diffraction study was obtained through the dissolving of compound in CHCl₃, ethyl acetate and petroleum ether, followed by slow evaporation of the solvent at room temperature. X-ray data collections were performed in an Agilent Super Nova.



Figure S1. X-Ray coordinate of proposed corypsoriol I (**1h**) (CCDC 2226751). Displacement ellipsoids are scaled to the 30% probability level.

Identification code	Proposed corypsoriol I (1h)
Empirical formula	C76H104O12
Formula weight	1209.59
Temperature/K	149.99(10)
Crystal system	triclinic
Space group	P1
a/Å	9.51230(10)
b/Å	11.7533(2)
c/Å	16.8232(2)
α/°	109.4870(10)
β/°	90.4410(10)
γ/°	100.6710(10)
Volume/Å ³	1737.54(4)
Z	1
ρ _{calc} g/cm ³	1.156
µ/mm ⁻¹	0.607
F(000)	656.0
Crystal size/mm ³	0.16 × 0.13 × 0.12
Radiation	Cu Kα (λ = 1.54184)
2⊖ range for data collection/°	5.588 to 153.732
Index ranges	-11 ≤ h ≤ 10, -14 ≤ k ≤ 14, -20 ≤ l ≤ 20
Reflections collected	59003
Independent reflections	12367 [Rint = 0.0890, Rsigma = 0.0529]
Data/restraints/parameters	12367/3/859
Goodness-of-fit on F ²	1.025
Final R indexes [I>=2σ (I)]	R ₁ = 0.0558, wR ₂ = 0.1411
Final R indexes [all data]	R ₁ = 0.0592, wR ₂ = 0.1439
Largest diff. peak/hole / e Å ⁻³	0.48/-0.20
Flack parameter	0.15(12)

7. References

- a) J. Y. Kang, R. G. Carter, *Org. Lett.* 2012, **14**, 3178-3181; b) J. Y. Kang, R. C. Johnston, K. M. Snyder, P. H.-Y. Cheong, R. G. Carter, *J. Org. Chem.* 2016, **81**, 3629-3637.
- T. Isobe, K. Fukuda, T. Tokunaga, H. Seki, K. Yamaguchi, T. Ishikawa, *J. Org. Chem.*2000, 65, 7774-7778.
- 3 K. C. Nicolaou, T. Montagnon, P. S. Baran, Y. L. Zhong, J. Am. Chem. Soc. 2002, 124, 2245-2258.
- Y. Liu, S. C. Virgil, R. H. Grubbs, B. M. Stoltz, *Angew. Chem. Int. Ed.* 2015, 54, 11800 11803. *Angew. Chem.* 2015, 127, 11966-11969.
- J.-L. Pan, T. Chen, Z.-Q. Zhang, Y.-F. Li, X.-M. Zhang, F.-M. Zhang, *Chem. Commun.* 2016, **52**, 2382-2385.
- 6 M. Zhu, N. Jalalian, B. Olofsson, *Synlett* 2008, 592-596.
- 7 D. Müller, A. Alexakis, *Chem. Eur. J.* 2013, **19**, 15226-15239.
- X.-G. Liu, C. Zhou, E. Lin, X. Han, S. Zhang, Q. Li, H. Wang, *Angew. Chem. Int. Ed.*2018, **57**,13096-13100.
- 9 G. Xiao, X. Li, T. Wu, Z. Cheng, Q. Tang, T. Zhang, *Fitoterapia* 2012, **83**, 1153-1557.
- 10 M. X. Xiu, Y. M. Zhao, Y. Zhang, D. X. Xiong, D. Wang, H. S. Lee, L. Cui, *Fitoterapia* 2021, **151**, 104881-104885.
- a) A. Pezzella, L. Lista, A. Napolitano, M. d'Ischia, *Tetrahedron Lett.* 2005, 46, 3541-3544;
 b) M. L. Nóvoa, F. J. Salazara, C. Gámez, A. Y. Angarita, E. Tropper, N. Canudas, J. E. Villamizar, *Nat. Prod. Commun.* 2014, 9, 355-358.
- 12 Q.-X. Xu, W. Xu, X.-W. Yang, *Tetrahedron* 2020, **76**, 131343-131348.
- 13 X.-W. Yang, Q. Lü, Q.-X. Xu, W. Xu, Y.-T. Zhang, *Chin. Trad. Herb. Drugs* 2022, **53**, 3269-3279.

8. Copies of ¹H and ¹³C NMR spectra UPC² data of products







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peak information:

	RetTime (min)	Area (µV*s)	Area (%)	Height (µV)
1	2.089	147851	49.60	26859
2	2.339	150206	50.40	21497





peak information:

	RetTime (min)	Area (µV*s)	Area (%)	Height (µV)
1	2.177	6657	3.90	2974
2	2.312	164016	96.10	24747










Sample Information

Sample Name:

Compound-**5**'-C5-9505-xx

Wave Length:

220.0nm

Column:





	RetTime (min)	Area (µV*s)	Area (%)	Height (µV)
1	2.370	5600641	49.91	1413197
2	3.659	5619823	50.09	950410

Empower™3

Sample Information

Sample Name:

Compound -5'-C5-9505

Wave Length: 220.0nm

Column:

AU

1.50

PDA Spectrum PDA 220.0 nm (PDA)





	RetTime (min)	Area (µV*s)	Area (%)	Height (µV)
1	2.445	5752452	96.39	1478514
2	3.775	215153	3.61	39229







Sample Name: Column:

Compound-4-C5-9010xx

Wave Length: 210.0nm





	RetTime (min)	Area (µV*s)	Area (%)	Height (µV)
1	1.633	5653674	50.09	1767231
2	1.848	5633489	49.91	1528376



Compound-4-C5-9010 Sample Name:

Wave Length:

210.0nm

Column:

PDA Spectrum PDA 210.0 nm (PDA)



	RetTime (min)	Area (µV*s)	Area (%)	Height (µV)
1	1.637	3569507	97.05	1025589
2	1.871	108487	2.95	29459







Sample Name:

Column:

Compound -**3**-C7-9901-xx PDA Spectrum PDA 238.0 nm (PDA) Wave Length: 238.0nm



peak information:

	RetTime (min)	Area (µV*s)	Area (%)	Height (µV)
1	1.519	1750624	49.90	534160
2	1.667	1757818	50.10	450094

Reported by User: System Report Method: Sample Information Report Method ID: 2139 Page: 1 of 1

Project Name: ZLH Date Printed: 05/01/2022 20:29:24 PRC

Empower[™]3

Sample Information

Compound-3-C7-9901 Sample Name:

Wave Length:

Column:

238.0nm





peak information:

	RetTime (min)	Area (µV*s)	Area (%)	Height (µV)
1	1.552	521197	96.76	196364
2	1.719	17443	3.24	6956

Reported by User: System Report Method: Sample Information Report Method ID: 2214 Page: 1 of 1

Project Name: ZLH Date Printed: 05/01/2022 20:34:40 PRC

























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s_{u-1} hydroxy. Peoracorylifol F (9) ¹³ C NMR (150 MHz, CDCl ₃)
$s_{\alpha} - hydroxy-Psoracorylifol F (9)$ ¹³ C NMR (150 MHz, CDCl ₃)







$$\begin{array}{c} 157.78 \\ 147.08 \\ 147.08 \\ 147.08 \\ 1129.54 \\ 129.54 \\ 111.04 \\ 111.04 \\ 111.04 \\ 111.04 \\ 111.04 \\ 111.04 \\ 111.04 \\ 111.04 \\ 111.04 \\ 111.04 \\ 111.04 \\ 111.04 \\ 129.53 \\ 120.20 \\ 120$$














fl (ppm)









¹³C NMR (150 MHz, CDCl₃)















$$\begin{array}{c} & 158.46 \\ & 148.22 \\ & 146.05 \\ & 146.05 \\ & 146.05 \\ & 130.32 \\ & 130.32 \\ & 130.32 \\ & 130.32 \\ & 130.32 \\ & 130.32 \\ & 112.25 \\ & 112.25 \\ & 112.25 \\ & 112.25 \\ & 112.25 \\ & 112.25 \\ & 112.25 \\ & 112.25 \\ & 112.25 \\ & 112.25 \\ & 112.25 \\ & 112.25 \\ & 112.25 \\ & 122.49 \\ & 15.49 \\ & 1$$











¹H NMR (600 MHz, acetone- d_6)







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HO ОH

Psoracorylifol G (1f)

¹H NMR (600 MHz, CDCl₃)











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¹³C NMR (100 MHz, CDCl₃)

















