Supplementary Information

for

Designing multi-target-directed flavonoids: A strategic approach to Alzheimer's disease

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Experimental Section

Materials and Methods. All chemical reagents were purchased from commercial suppliers and used as received unless otherwise stated. Chrysin (1) was obtained from TCI Chemicals (Tokyo, Japan). 6,7-Dihydroxyflavone (2) and 5,7-dihydroxyisoflavone (1b) were purchased from Carbosynth (Compton, UK). 5,6-Dihydroxyflavone (3) and baicalein (4) were gained from Alfa Aesar (Ward Hill, MA, USA). 5.7-Dimethoxyflavone (1a) and 5.6.7-Trimethoxyflavone (4a) were purchased from Sigma-Aldrich (St. Louis, MO, USA) and AA Blocks LLC (San Diego, CA, USA), respectively. 8-Chlorobaicalein (5) and 5,6,7-trihydroxyisoflavone (4b) were prepared following previously reported procedures.^{1,2} A_{β40} (DAEFRHDSGYEVHHQKLVFFAEDVGSNKGAIIGLMVG-GVV) and A_{β42} (DAEFRHDSGYEVHHQKLVFFAEDVGSNKGAIIGLMVGGVVIA) were obtained from Peptide Institute, Inc. (Osaka, Japan). HEPES [2-(4-(2-hydroxyethyl)piperazin-1-yl)ethanesulfonic acid] was purchased from Sigma-Aldrich. The buffered solution was prepared in doubly distilled water [ddH₂O; a Milli-Q Direct 16 system (18.2 MΩ·cm; Merck KGaA)]. Trace metal contamination was removed from all solutions used for A β experiments by treating with Chelex (Sigma-Aldrich) overnight. Cellular ROS detection assay and AChE assay kits were purchased from Abcam (Cambridge, UK) and Invitrogen (Carlsbad, CA, USA), respectively. ¹H and ¹³C NMR spectra of 5, 4b, and 3-(3,4-dihydroxyphenyl)-5,6,7-trihydroxy-4H-chromen-4-one (6) were recorded on a Bruker AV400 NMR spectrometer (Bruker BioSpin, Rheinstetten, Germany; Department of Chemistry, KAIST, Daejeon, Republic of Korea). High-resolution mass spectrometric analysis of 5, 4b, and 6 was carried out by micrOTOF-QII [Bruker Daltonics, Bremen, Germany; KAIST Analysis Center for Research Advancement (KARA), Daejeon, Republic of Korea]. Absorbance and fluorescence values for biological assays were measured on a Molecular Devices SpectraMax M5e microplate reader (Sunnyvale, CA, USA). Images gained through gel/Western blot were visualized by a ChemiDoc MP imaging system (Bio-Rad, Hercules, CA, USA). Morphologies of A β aggregates produced from the aggregation experiments were taken on a Tecnai F20 and G2 F30 S-Twin transmission electron microscope (FEI Company, Eindhoven, Netherlands; KARA). The change in the secondary structure of peptides was analyzed by a JASCO-815 150-L CD spectropolarimeter (Jasco Inc., Tokyo, Japan). Optical spectra were recorded on an Agilent 8453 UV-Vis spectrophotometer (Santa Clara, CA, USA). ITC measurements were performed by a VP-ITC instrument (Malvern Panalytical Ltd., Malvern, UK; KBSI, Ochang, Republic of Korea). 2D SOFAST-HMQC NMR spectra of uniformly ¹⁵N-labeled $A\beta_{40}$ (rPeptide, Georgia, GA, USA) and $A\beta_{42}$ with and without flavonoids were collected by a Bruker AVANCE II-800 MHz NMR spectrometer equipped with a cryoprobe (Bruker BioSpin: KBSI). Mass spectrometric analysis of the interactions of **6** with A β_{40} and A β_{42} in the absence and presence of Cu(II) was conducted by an Agilent 6530 Accurate-Mass and Waters Xevo G2-XS quadrupole time-of-flight liquid chromatography/mass spectrometry.

Calculation of Redox Potentials. All calculations were performed based on the density functional theory (DFT)³ with the Jaguar 9.1 suite⁴ at B3LYP⁵⁻⁷ level of theory. The optimization of flavonoids' structures was carried out with the 6-31G^{**} basis set.⁸⁻¹⁰ Following the geometry optimization, electronic energies of the optimized flavonoid structures were recalculated with a high quality triple- ζ basis set cc-pVTZ(-f).¹¹ All atoms in the molecules were treated with the LACV3P^{**} basis set, one of the classes in LACVP basis functions where the exponents were decontracted to match with the triple- ζ functions of the main group elements. Vibrational frequencies for the optimized structures were calculated at the same level of theory as the geometry optimization procedure. Vibrational entropy correction along with the zero-point vibrational energies was considered for proper thermodynamic approximations. Based on the optimized gas phase geometries, solvation correction energies were deduced. Self-consistent reaction field (SCRF)¹²⁻¹⁴ approximations were considered to calculate the linearized Poisson-Boltzmann equations with the dielectric constant ε . The solvation energy used in the system was treated with EtOH (ε = 24.5). The Gibbs free energies in solution phase were computed as the following equations:

G(sol) = G(gas) + G(solv)	(eq 1)
G(gas) = H(gas) – TS(gas)	(eq 2)
H(gas) = E(SCF) + ZPE	(eq 3)

G(Sol) represents the Gibbs free energy with solvation correction G(solv) from the gas phase free energy G(gas); H(gas) is the enthalpy of the molecule in gas phase; T is the temperature (298.15 K); S(gas) is the entropy of the molecule in the gas phase; E(SCF) is the self-consistent field converged electronic energy; ZPE represents the vibrational zero-point energy. To calculate the redox potential, the free energy of the one-electron oxidized form of the flavonoid was deduced by the free energy of its neutral form.

TEAC Assay. The free radical scavenging capacity of flavonoids, relative to that of the vitamin E analog, Trolox, known as an antioxidant,¹⁵ was determined by the TEAC assay based on the decolorization of ABTS [2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid)diammonium salt] cation radicals. The TEAC assay was conducted in ethanol (EtOH) following previously reported

methods.^{16,17} Blue ABTS⁺ cation radicals were generated by dissolving ABTS (7 mM) with potassium persulfate (2.5 mM) in H₂O (5 mL) and incubating the solution for 16 h at room temperature in the dark, and then the solution was diluted with EtOH to make the absorbance value of ca. 0.7 at 734 nm. The resultant ABTS⁺⁺ solution (200 µL) was added into a clear 96-well plate. Various concentrations of flavonoids were added into a 96-well plate and incubated at 25 °C for 1, 3, and 6 min. The final concentrations of flavonoids were as follows: Trolox (0.24, 0.48, 0.97, 1.9, 3.9, 7.7, 16, 31, 62, 125, and 248 μM); **2** (0.10, 0.19, 0.39, 0.77, 1.6, 3.1, 6.2, 12, 25, 50, and 99 μM); **3** and **6** (0.39, 0.77, 1.6, 3.1, 6.2, 12, 25, 50, 99, 149, and 198 μM); **4** and **5** (0.077, 0.15, 0.31, 0.62, 1.2, 2.5, 5.0, 10, 15, 20, and 25 μM); **1a** and **1b** (0.03, 0.06, 0.12, 0.23, 0.47, 0.93, 1.9, 3.7, 7.4, 15, and 30 µM); **4a** (0.19, 0.39, 0.77, 1.6, 3.1, 6.2, 12, 19, 25, 37, and 50 µM); **4b** (0.019, 0.039, 0.077, 0.15, 0.31, 0.62, 1.2, 2.5, 5.0, 9.9, and 20 µM). Percent inhibition was calculated based on the measured absorbance at 734 nm [% inhibition = $100 \times (A_0 - A)/A_0$; A_0 = absorbance of the control well without flavonoids; A = absorbance of the wells treated with flavonoids] and plotted as a function of flavonoids' concentration. The TEAC values of each time point were calculated as the ratio between the slope of flavonoids and the slope of Trolox. All measurements were carried out in triplicate.

Cellular ROS Detection Assay. The prooxidant activity of flavonoids in living cells was measured by a cellular ROS detection assay kit (Abcam) following the manufacturer's protocol. The human neuroblastoma 5Y cells were maintained in media containing 50% v/v Dulbecco's Modified Eagle Medium and 50% v/v Nutrient Mixture F12 (phenol red-free DMEM/F12; GIBCO, Grand Island, NY, USA) supplemented with 10% v/v fetal bovine serum (GIBCO) and 100 U/mL penicillin-streptomycin (GIBCO). The cells were grown and maintained at 37 °C in a humidified atmosphere with 5% CO₂. Cells were seeded in a 96-well plate (50,000 cells/100 µL). After 24 h incubation, a ROS-staining working solution (100 µL) was added to each well and the plate was incubated for 1 h at 37 °C. Cells were then treated with the samples {[flavonoid] = 0.01, 0.1, 0.5, and 1 mM (1% v/v DMSO)}. The cellular ROS levels upon the treatment of samples were measured based on the fluorescence intensity ($\lambda_{ex}/\lambda_{em} = 520/605$ nm) of the wells that was detected after 2 h incubation and calculated, relative to that of the cells containing an equivalent amount of DMSO. All experiments were performed in triplicate.

AChE Activity Assay. The AChE inhibitory activity of flavonoids was measured by a fluorometric AChE assay kit (Invitrogen) following the manufacturer's protocol with slight modifications.

Electric eel acetylcholinesterase (*ee*AChE) was dissolved and diluted with a working solution. The *ee*AChE solution (50 µL) was added into a 96-well plate. Various concentrations of flavonoids were then introduced into the 96-well plate. The final concentrations of flavonoids were as follow: **1**, **3**, **4**, **5**, **1a**, **1b**, **4a**, **4b**, and **6** (0.010, 0.50, 5.0, 50, and 250 µM); **2** (0.32, 1.6, 8, 40, and 200 µM). After shaking, the mixture of *ee*AChE and flavonoids was pre-incubated for 15 min. A reaction mixture containing acetylcholine, an AChE probe, and AbRedTM was added into the 96-well plate to initiate the reaction. The AChE activity was calculated based on the fluorescence intensity ($\lambda_{ex}/\lambda_{em} = 571/585$ nm) of the wells that was detected after 15 min incubation. Data were normalized to the control (no inhibitor). All experiments were performed in triplicate.

Docking Studies. Flexible ligand docking studies using AutoDock Vina 1.5.6 software¹⁸ for flavonoids were conducted against *ee*AChE (PDB 1C2B¹⁹). The MMFF94 energy minimization in ChemBio3D Ultra 11.0 was used to optimize the structure of flavonoids for docking studies. The structural files of flavonoids and the proteins were prepared in Auto Dock Tools,²⁰ imported into PyRx, and used to run AutoDock Vina.¹⁸ The exhaustiveness for the docking runs was set at 1,024. Docked models of flavonoids with the proteins were visualized using Pymol 2.0.7.

Aβ **Aggregation Experiments.** Aβ was dissolved in ammonium hydroxide [1% w/w NH₄OH (aq)] and the resulting solution was aliquoted, lyophilized overnight, and stored at -80 °C. A stock solution of A_β was then prepared by dissolving the lyophilized peptide with NH₄OH (1% w/w, ag; 10 μ L) and diluting with ddH₂O. All A β samples were prepared following previously reported procedures.^{21,22} The concentration of the peptide solution was determined by measuring its absorbance at 280 nm (ε = 1,450 M⁻¹cm⁻¹ for A β_{40} ; ε = 1,490 M⁻¹cm⁻¹ for A β_{42}). The buffered solution (20 mM HEPES, pH 7.4, 150 mM NaCl) was used for preparing Aβ samples. For the inhibition studies, flavonoids (final concentration, 50 µM; 1% v/v DMSO) were added to the samples of A_β (25 μ M) in the absence and presence of CuCl₂ or ZnCl₂ (25 μ M) followed by incubation for 24 h at 37 °C with constant agitation. For the disaggregation studies, A β (25 μ M) was incubated with and without CuCl₂ or ZnCl₂ (25 µM) for 24 h at 37 °C with constant agitation to generate preformed A β aggregates. The resultant A β aggregates were then treated with flavonoids (50 µM) and incubated for additional 24 h with constant agitation. For the additional disaggregation experiments, preformed A β_{42} aggregates (25 μ M) or A β_{42} seeds (10% v/v) with or without metal ions (25 µM for CuCl₂ and ZnCl₂) were incubated with flavonoids (50 µM) for 24 h with constant agitation. The resultant A β aggregates were then treated with freshly prepared A β_{42}

(25 μ M) and incubated for additional 24 h with constant agitation.

Gel/Western Blot. The resultant A β species from the aggregation experiments were analyzed by gel/Western blot using an anti-A β antibody (6E10, Covance, Princeton, NJ, USA).^{21,22} Each sample (10 µL) was separated on a 10–20% tricine gel (Invitrogen). Following separation, the proteins were transferred onto nitrocellulose membranes and blocked with bovine serum albumin (BSA, 3% w/v; Sigma-Aldrich) in Tris-buffered saline (TBS) containing 0.1% v/v Tween-20 (Sigma-Aldrich) (TBS-T) for 4 h at room temperature or overnight at 4 °C. The membranes were incubated with 6E10 (1:2,000) in the solution of BSA (2% w/v in TBS-T) for 4 h at room temperature. After washing with TBS-T (3x, 10 min), a horseradish peroxidase-conjugated goat anti-mouse secondary antibody (1:5,000 in 2% w/v BSA in TBS-T; Cayman Chemical Company, Ann Arbor, MI, USA) was added for 2 h at room temperature. A homemade ECL kit^{21–23} was used to visualize gel/Western blots on a ChemiDoc MP Imaging System (Bio-Rad).

TEM. Samples for TEM were prepared following previously reported methods.^{22,24} Glowdischarged grids (Formvar/Carbon 300-mesh; Electron Microscopy Sciences, Hatfield, PA, USA) were treated with Aβ samples (25 μM; 7.5 μL) for 2 min at room temperature. Excess sample was removed using filter paper followed by washing with ddH₂O (3x). Each grid incubated with uranyl acetate (1% w/v ddH₂O; 7.5 μL) for 2 min was blotted off and dried overnight at room temperature. Images for each sample were taken on a TEM (200 kV; 29,000x magnification; 300 kV; 29,500x magnification; KARA). The location of samples on the grids were randomly picked for taking more than 20 images per each grid.

Cu(II)-binding Studies. The interaction of flavonoids with Cu(II) was detected by Abs spectroscopy in 20 mM HEPES, pH 7.4, 150 mM NaCl. The solutions of flavonoids were titrated with 1 equiv of CuCl₂ at room temperature. The mixture solution was allowed to equilibrate for 5 min after the addition of CuCl₂ at room temperature before the spectra were recorded.

CD Spectroscopy. Metal-free A β_{42} (100 μ M) was incubated with and without **5**, **4b**, and **6** (200 μ M) for 24 h in 20 mM HEPES, pH 7.4, 150 mM NaF at 37 °C with constant agitation (250 rpm). CD spectra were collected in the range from 195 nm to 255 nm. The digital integration time, the bandwidth, and the scanning speed were 4 s, 2 nm, and 20 nm/min, respectively. Each spectrum was smoothed by Fourier transforms. The change in the secondary structure of metal-free A β_{42}

treated with a flavonoid was analyzed following the previously reported BeStSel method.²⁵ The measurements were conducted in triplicate.

Dot Blot Assay. The samples containing Aβ₄₂ (100 μM) with or without CuCl₂ or ZnCl₂ (100 μM) in the absence and presence of flavonoids (200 μM; 1% v/v DMSO) were prepared in 20 mM HEPES, pH 7.4, 150 mM NaCl and incubated for various time points (0, 1, 2, 3, 4, 6, 12, and 24 h). The resulting solution (2.5 μL) was spotted on a nitrocellulose membrane and blocked with a BSA solution (3% w/v) in 0.01% TBS-T at room temperature for 12 h. The membrane was incubated with a primary antibody [*i.e.*, 6E10²⁶ for Aβ species (1:2,000), A11²⁶ for Aβ oligomers (1:1,000; Invitrogen, Carlsbad, CA, USA), or OC²⁶ for Aβ fibrils (1:2,000; Merch Millipore, Billerica, MA, USA)] in the solution of BSA (2% w/v in TBS-T) for 4 h at room temperature. After washing with 0.01% TBS-T (3x, 7 min), the horseradish peroxidase-conjugated goat anti-mouse (1:5,000; for 6E10) or goat anti-rabbit (1:50,000; for A11 and OC) secondary antibody in the solution of BSA (2% w/v in TBS-T) was added to the membrane and incubated for 1 h at room temperature. A homemade ECL kit^{21–23} was used to visualize the results on a ChemiDoc MP Imaging System. The intensity observed from A11- or OC-detectable Aβ species was quantified, relative to that detected from 6E10, by the ImageJ software.

Abs Spectroscopy. Flavonoid (50 μ M; 1% v/v DMSO) was added into the buffered solution (20 mM HEPES, pH 7.4, 150 mM NaCl) with or without A β_{40} (25 μ M) in the absence and presence of CuCl₂ or ZnCl₂ (25 μ M). Abs spectra of the samples were collected for 24 h at 37 °C.

Synthesis of 6.

6c [3-(3,4-Dihydroxyphenyl)-5,6,7-trimethoxy-4*H*-chromen-4-one]. 3,4,5-Trimethoxyphenol (**6a**; 1.2 g, 6.3 mmol) and 3,4-dihydroxyphenylacetic acid (**6b**; 1.1 g, 6.3 mmol) were dissolved in boron trifluoride diethyl etherate (BF₃·OEt₂, 10 mL) and refluxed at 75 °C. After 4 h, the reaction mixture was extracted in a solution of ethyl acetate (EtOAc) and saturated sodium acetate (NaOAc; 0.12 g/mL). The organic layer was collected, washed with brine, dried with sodium sulfate (NaSO₄), filtered, and concentrated. The crude product was purified by column chromatography (SiO₂; EtOAc:hexanes = 1:1) to yield **6c** [R_f = 0.3; yellow powder; 650 mg, 2.0 mmol (yield = 31%)] that was used for the next step without further purification. ¹H NMR [400 MHz, DMSO- d_6 , δ (ppm)]: 6.58 (1H, d, J = 8.0 Hz), 6.54 (1H, dd, J = 8.0, 2.1 Hz), 6.40 (1H, d, J = 2.1 Hz), 6.29 (1H, s), 4.00 (2H, s), 3.77 (3H, s), 3.65 (3H, s), 3.62 (3H, s). ¹³C NMR [100 MHz, DMSO-

 d_6 , δ (ppm)]: 202.9, 158.3, 158.0, 153.4, 144.9, 143.9, 134.4, 125.8, 120.5, 117.1, 114.5, 110.0, 96.2, 92.8, 61.3, 60.6, 56.0, 55.6, 48.6. HRMS: *m*/*z* Calcd. for C₁₇H₁₈O₇ [M – H]⁻: 333.1053; found: 333.2388.

6d [3-(3,4-Dihydroxyphenyl)-5,6,7-trimethoxy-4H-chromen-4-one]. 6c (650 mg, 2.0 mmol) was dissolved in dry *N*,*N*-dimethylformamide (DMF, 8 mL), and BF₃·OEt₂ (1.4 mL, 20 mmol) was added dropwise and heated to 50 °C for 1 h followed by adding methanesulfonyl chloride (MeSO₂Cl; 231 µL, 2.0 mmol) dropwise. The reaction mixture was refluxed at 100 °C for 2 h, cooled down to room temperature, and extracted with EtOAc containing saturated NaOAc (0.12 g/mL). The organic layer was collected, washed with brine, dried with NaSO₄, filtered, and concentrated. The crude product was purified by column chromatography (SiO₂; EtOAc:hexanes = 3:1). **7d** was obtained by washing the solid with CH₃OH [*R_f* = 0.4; yellow powder; 74 mg, 0.22 mmol (yield = 11%)]. ¹H NMR [400 MHz, CD₃OD, *δ* (ppm)]: 8.05 (1H, s), 7.03 (1H, d, *J* = 8.1 Hz), 6.97 (1H, dd, *J* = 8.1, 2.2 Hz), 6.83 (1H, d, *J* = 2.2 Hz), 3.99 (3H, s), 3.92 (3H, s), 3.87 (3H, s). ¹³C NMR [100 MHz, CD₃OD, *δ* (ppm)]: 176.0, 158.4, 155.0, 152.4, 151.7, 145.3, 144.7, 140.4, 125.5, 123.4, 120.5, 116.3, 114.8, 112.7, 96.3, 61.1, 60.4, 55.6. HRMS: *m/z* Calcd. for C₁₈H₁₆O₇Na [M + Na]⁺: 367.0794; found: 367.0786.

6 [3-(3,4-Dihydroxyphenyl)-5,6,7-trihydroxy-4*H*-chromen-4-one]. **6**d (74 mg, 0.22 mmol) was dissolved in dry dichloromethane (CH₂Cl₂) and stirred in an ice bath. A solution of boron tribromide (BBr₃) in CH₂Cl₂ (1 M, 250 μ L) was added into the solution of **6d** dropwise and then the ice-bath was removed to stir the reaction mixture at 60 °C for 18 h followed by refluxing at 100 °C for 2 h. After cooling down to room temperature, the mixture was extracted with EtOAc and H₂O. The organic layer was collected, washed with brine, dried with NaSO₄, filtered, and concentrated. The crude was purified by column chromatography (SiO₂; CH₃OH:CH₂Cl₂ = 1:30). **6** was obtained by recrystallization using a mixture of acetone, hexanes, and CH₂Cl₂ [*R_f* = 0.2; white powder; 36 mg, 0.12 mmol (yield = 55%)]. ¹H NMR [400 MHz, CD₃OD, δ (ppm)]: 8.04 (1H, s), 7.02 (1H, d, *J* = 2.2 Hz), 6.86 (1H, dd, *J* = 2.2, 8.1 Hz), 6.83 (1H, d, *J* = 8.1 Hz), 6.46 (1H, s). ¹³C NMR [100 MHz, CD₃OD, δ (ppm)]: 181.0, 153.5, 153.2, 151.0, 147.0, 145.3, 144.8, 129.1, 122.7, 122.6, 120.3, 116.1, 114.9, 105.1, 93.2. HRMS: *m/z* Calcd. for C₁₅H₁₀O₇Na [M + Na]⁺: 325.0324; found: 325.0317.

Cell Viability Studies. Human neuroblastoma 5Y cells were maintained in media containing 50% v/v minimum essential medium (MEM; GIBCO) and 50% v/v F12 (GIBCO) and supplemented with 10% v/v fetal bovine serum (Sigma-Aldrich) and 100 U/mL penicillin with 100 mg/mL streptomycin (GIBCO). The cells were grown and maintained at 37 °C in a humidified atmosphere with 5% CO₂.

Cell viability was determined by the MTT assay [MTT = 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide]. Cells were seeded in a 96-well plate (15,000 cells/100 µL) and treated with the samples {[flavonoid] = 5, 10, 25, 50, and 100 µM (1% v/v DMSO)}. After 24 h incubation, 5Y cells were washed with phosphate buffered saline (PBS; GIBCO). MTT [5 mg/mL in PBS (pH 7.4, GIBCO); 25 µL] was added to each well and the plate was incubated for 4 h at 37 °C. The formazan produced by cells was solubilized using an acidic solution of DMF (pH 4.5, 50% v/v, aq) and sodium dodecyl sulfate (SDS; 20% w/v) overnight at room temperature in the dark. Absorbance was measured at 600 nm by a microplate reader (Molecular Devices). Cell viability, relative to that of the cells containing an equivalent amount of DMSO, was calculated. The measurements were conducted in triplicate.

ITC. The solutions of A β and **6** were subjected to degassing under vacuum for 3 min prior to loading into the instrument. The solution of **6** [800 μ M; 20 mM HEPES, pH 7.4 (1% v/v DMSO, 1% w/w NH₄OH)] in the syringe was titrated into the solution containing A β [40 μ M; 20 mM HEPES, pH 7.4 (1% v/v DMSO, 1% w/w NH₄OH)] in the cell at 10 °C with 25 injections at a constant interval of 360 s (2 μ L for the first injection and 11 μ L for the following injections). The cell was continuously stirred at 264 rpm. The initial delay and the reference power were 60 s and 10 μ cal/s, respectively. Dilution heat of **6** only was also measured under the same experimental conditions. The ITC thermogram and binding isotherm were shown after the subtraction of the dilution heat. The binding isotherm after baseline correction was fitted to the one-set-of-sites binding model using the MicroCal Origin software (version 7.0).

Expression and Purification of ¹⁵N-labeled Recombinant A β_{42} . ¹⁵N-labeled recombinant A β_{42} was expressed in *E. coli* and purified as previously reported.²⁷ Briefly, the DNA sequence coding the human A β_{42} peptide was artificially constructed using codons preferred by *E. coli* with an extra Met residue at the *N*-terminal of the peptide. The DNA construct was inserted into pAED4 vector²⁸ and the *E. coli* BL21 (DE3) pLysS strain (Novagen, Inc., Madison, WI, USA) was utilized in the protein expression. Transformed cells were incubated in LB media containing ampicillin and chloramphenicol until the cultures reach 3.5 McFarland turbidity. Cells were then harvested by centrifugation and the cell pellets were resuspended in M9 minimal media containing isotopically labeled 1 g/L ¹⁵N ammonium chloride, and induced by addition of 1 mM IPTG for 4 h. The peptide formed inclusion bodies in the bacterial cells. The purified inclusion bodies were dissolved in 20 mM NaOH and the peptide was further purified by repeated cycles of amyloid growth at low pH

and monomerization in hexafluoroisopropanol (HFIP) combined with centrifugation steps.

2D ¹H–¹⁵**N SOFAST-HMQC NMR Experiments.** To prepare a stock solution of ¹⁵N-labeled A_{β40} uniformly ¹⁵N-labeled A_{β40} was dissolved in NH₄OH (1% w/w, aq) and the resulting solution was lyophilized for 48 h and stored at –80 °C. To remove possible preformed aggregates, the lyophilized peptide was then redissolved in diluted NaOH (pH 10) and passed through a filter (0.22 µm). On the other hand, the stock solution of ¹⁵N-labeled A_{β42} based on the previous report.²⁷ Briefly, uniformly ¹⁵N-labeled A_{β42} was solubilized in diluted NaOH (pH 10) at a concentration of ~200 µM. The undissolved precipitates of A_{β42} were removed by centrifugation at 40,000 rpm for 1 h. The solution of ¹⁵N-labeled A_β (33 µM) was prepared with and without **6** (100 µM; 1% v/v DMSO) in 20 mM HEPES, pH 7.4 (10% v/v D₂O) for NMR experiments. All spectra were acquired with 180 scans at 10 °C. Dimensions of the spectra were 1,024 (¹H) × 128 (¹⁵N) points. Experimental data were processed with NMRpipe²⁹ and Sparky 3.115.³⁰ The ¹H and ¹⁵N assignments of ¹⁵N-labeled A_β were determined based on the previous results.^{21,31} The chemical shift perturbation was calculated following the equation (eq 4).³² Note that $\Delta \Delta_{\rm H}$ and $\Delta \delta_{\rm N}$ represent the alterations in the chemical shifts of ¹H and ¹⁵N of ¹⁵N-labeled A_β, respectively.

$$\Delta \delta_{\rm NH} = \sqrt{(\Delta \delta_{\rm H})^2 + \left(\frac{\Delta \delta_{\rm N}}{6.5}\right)^2} \qquad ({\rm eq} \ 4)$$

ESI–MS. A β (100 μ M) was incubated with **6** (200 μ M; 1% v/v DMSO) in the absence and presence of CuCl₂ (100 μ M) in 20 mM ammonium acetate, pH 7.4 at 37 °C with constant agitation. The incubated samples were diluted by 10-fold with ddH₂O and then injected into the mass spectrometer. The capillary voltage, nozzle voltage, and gas temperature were set to 5.8 kV, 2 kV, and 300 °C, respectively. ESI–MS² analyses were additionally conducted on the singly oxidized A β produced by treating **6** with metal-free and Cu(II)-bound A β . ESI parameters and experimental conditions were the same as above. Collision-induced dissociation was carried out by applying the collision energy in the trap at 65 V. The isolation width was set as narrow (*ca.* 1.3 *m/z*), and more than 200 spectra were obtained for each sample and averaged for the analysis. For the experiments under anaerobic conditions, all samples were prepared following the same procedure described above for the aerobic samples in a N₂-filled glove box.



Fig. S1 ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of 5 in DMSO- d_6 .



Fig. S2 ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of 4b in DMSO- d_6 .



Fig. S3 Prooxidant activity of **2–6** and **4b** in 5Y cells. The total concentration of ROS in the cells incubated with various concentrations of the flavonoids was measured by a cellular ROS detection assay. Conditions: [flavonoid] = 0.01, 0.1, 0.5, and 1 mM (1 % v/v DMSO); $[H_2O_2]$ = 10 and 100 μ M.



Fig. S4 Possible interactions of flavonoids against AChE. Nine docked models of **1–5**, **1a**, **1b**, **4a**, **4b**, and **6** against AChE with binding energies ranging from –10.8 to –8.5 kcal/mol were obtained. The representative docked models with the highest binding affinity towards AChE are visualized in the figure. Hydrogen bonds between flavonoids and AChE are indicated with dashed lines (2.2–3.7 Å). N, O, H, Cl (from **5**), and C (from Ser203) atoms are depicted in blue, red, white, brown, and green, respectively.



Fig. S5 Effects of **1**, **1a**, **1b**, **4**, **4a**, and **4b** on the formation of metal-free A β_{42} and metal-A β_{42} aggregates. Analyses of the (a) MW distribution and (b) morphology of the resultant A β_{42} species were carried out by gel/Western blot with an anti-A β antibody (6E10) and TEM, respectively. Lanes: (<u>C</u>) A β_{42} ± Cu(II) or Zn(II); (**1**) <u>C</u> + **1**; (**1a**) <u>C</u> + **1a**; (**1b**) <u>C</u> + **1b**; (**4**) <u>C</u> + **4**; (**4a**) <u>C</u> + **4a**; (**4b**) <u>C</u> + **4b**. Conditions: [A β_{42}] = 25 µM; [M(II)] = 25 µM; [flavonoid] = 50 µM (1% v/v DMSO); 20 mM HEPES, pH 7.4, 150 mM NaCI; 37 °C; 24 h; constant agitation. Scale bars = 200 nm.



Fig. S6 Influence of **1**, **1a**, **1b**, **4**, **4a**, and **4b** on the formation of metal-free A_{β40} and metal–A_{β40} aggregates. Analyses of the (a) MW distribution and (b) morphology of the resultant A_{β40} species were performed by gel/Western blot with an anti-A_β antibody (6E10) and TEM, respectively. Lanes: (\underline{C}) A_{β40} ± Cu(II) or Zn(II); (**1**) \underline{C} + **1**; (**1a**) \underline{C} + **1a**; (**1b**) \underline{C} + **1b**; (**4**) \underline{C} + **4**; (**4a**) \underline{C} + **4a**; (**4b**) \underline{C} + **4b**. Conditions: [A_{β40}] = 25 µM; [M(II)] = 25 µM; [flavonoid] = 50 µM (1% v/v DMSO); 20 mM HEPES, pH 7.4, 150 mM NaCl; 37 °C; 24 h; constant agitation. Scale bars = 200 nm.



Fig. S7 Impact of **1**–**6** and **4b** on preformed metal-free A β and metal–A β aggregates. (a) Scheme of the disaggregation experiments. (b and c) Analyses of the MW distribution and morphology of the resultant (b) A β_{42} and (c) A β_{40} species by gel/Western blot with an anti-A β antibody (6E10) and TEM, respectively. Lanes: (<u>C</u>) A β ± Cu(II) or Zn(II); (**1**) <u>C</u> + **1**; (**2**) <u>C</u> + **2**; (**3**) <u>C</u> + **3**; (**4**) <u>C</u> + **4**; (**5**) <u>C</u> + **5**; (**4b**) <u>C</u> + **4b**; (**6**) <u>C</u> + **6**. The original gel images are shown in Fig. S9b. Conditions: [A β] = 25 µM; [M(II)] = 25 µM; [flavonoid] = 50 µM (1% v/v DMSO); 20 mM HEPES, pH 7.4, 150 mM NaCI; 37 °C; 24 h; constant agitation. Scale bars = 200 nm.



Fig. S8 Effects of **1–6** and **4b** on preformed metal-free and metal-added A β_{42} aggregates and seeds in the presence of freshly prepared A β_{42} . (a and c) Scheme of the aggregation experiments. (b and d) Analyses of the MW distribution and morphology of the resultant A β_{42} species by gel/Western blot with an anti-A β antibody (6E10) and TEM, respectively. Lanes: (<u>C</u>) A β ± Cu(II) or Zn(II); (**1**) <u>C</u> + **1**; (**2**) <u>C</u> + **2**; (**3**) <u>C</u> + **3**; (**4**) <u>C</u> + **4**; (**5**) <u>C</u> + **5**; (**4b**) <u>C</u> + **4b**; (**6**) <u>C</u> + **6**. Conditions: [A β_{42}] = 25 µM; A β_{42} seeds (10% v/v); [M(II)] = 25 µM; [flavonoid] = 50 µM (1% v/v DMSO); 20 mM HEPES, pH 7.4, 150 mM NaCI; 37 °C; 24 h; constant agitation. Scale bars = 200 nm.



Fig. S9 Original gel images of Figs. 4 and S7.



Fig. S10 Interaction of the flavonoids with Cu(II) monitored by Abs spectroscopy. Conditions: [flavonoid] = 50 μ M; [Cu(II)] = 50 μ M; 20 mM HEPES, pH 7.4, 150 mM NaCl; room temperature.

(a)	$- Metal-free A\beta_{42}$	(b)	Metal-free $A\beta_{42}$	+ 5	+ 4b	+ 6
cm -1)		α-Helix (%)	19.2 (± 2.1)	1.2 (± 1.2)	0.4 (± 0.4)	1.5 (±1.5)
ε (M ^{−1} e	-5 -	β -Sheet (%)	73.7 (± 0.7)	66.7 (± 15.3)	35.5 (± 5.7)	46.7 (± 2.6)
Ā.		Turn (%)	5.7 (± 1.0)	12.5 (± 0.7)	10.3 (± 1.8)	38.1 (± 7.6)
	195 210 225 240 255 Wavelength (nm)	Ramdom Coils (%)	1.5 (± 1.5)	19.7 (± 15.5)	53.8 (± 5.9)	13.6 (± 5.6)

Fig. S11 Change in the secondary structure of metal-free A β_{42} upon incubation with **5**, **4b**, or **6**. (a) CD spectra of the samples containing metal-free A β_{42} in the absence and presence of **5**, **4b**, or **6**. (b) Structural composition of the samples from each CD spectrum analyzed by BeStSel.²⁵ Conditions: [A β_{42}] = 100 μ M; [flavonoid] = 200 μ M (1% v/v DMSO); 20 mM HEPES, pH 7.4, 150 mM NaF; 37 °C; 24 h; constant agitation.

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(a)) Metal-free																							
	Anti-oligomer Antibody (A11)					Anti-fibril Antibody (OC)					Anti-Aβ Antibody (6E10)													
Time (h)	0	1	2	3	4	6	12	24	0	1	2	3	4	6	12	24	0	1	2	3	4	6	12	24
<u>C</u>	•	•		•	0			•	•	•	٠	•	•	•	•	٠	٠	٠	•	•	•	•	•	•
+ 1	•	•	•	•					٠	•	•	•	•	•	•	•	٠	٠	•	•	•	•	•	•
+ 2	٠	•	٠	۰	•	•	0	٠	•	•	•	•	•	•	•	•	•	٠	•	٠	•	•	•	•
+ 3		•	•	•	•	•			•	•	•	•	٠	•	•	•	•	•	•	•	•	•	•	•
+ 4	•	•	•						•	•	•				e.	2	•	٠	٠	•	•		•	
+ 5	٠	•							•	•	•	•		1		24	•	٠	٠	٠	•	•	•	6
+ 1a	•		•	•			0		•	•	•	٠	٠	•	•	•	•	•	•	•	٠	•	•	•
+ 1b	•	•	•			0		•		•	٠	•		٠	٠	٠	•	•	•	•	•	•	•	•
+ 4a	•	•	•	•	•	•		•	•	•	•	•	٠	•	•	•	•	•	•	•	•	•	•	•
+ 4b							-		•	•	•					9	•	•	•	•	•	•	•	•
+ 6	0	۰	•	-				-	•	•		4				-	•	•	•	•	•	•	•	
												Cu	(II)											
		Anti-	oligo	mer	Antik	ody	(A11)		Ar	nti-fib	ril Ar	tiboo	dy (O	C)			Ar	nti-Aß	8 Anti	body	(6E	10)	
Time (h)	0	1	2	3	4	6	12	24	0	1	2	3	4	6	12	24	0	1	2	3	4	6	12	24
<u>C</u>	•	•	•	•	•		•		•	•	٠	٠	٠	•	٠	٠	•	•	•	•	•	•	•	•
+ 1	•	•	•	•	•			٠	•	•	•	•	٠	٠	•	•	•	•	•	•	٠	•	•	•
+ 2	•	•	•	•	•	•	•		•	•	•	•	•	٠	•	•	٠	•	•	•	•	•	•	•
+ 3	•	•		•	•			*	•	•	6	•	۰.		3	0	•	•	•	•	•	•	•	•
+ 4	•	•	•		٠				•	•		•	•	•	٠		٠	•	•	•	•	•	•	•
+ 5	•	•						-	•	•			9				•	•	٠	•	•	•	•	•
+ 1a	•	•	•	•				•	•	•	•	*•	•	•		•	•	•	•	٠	•	•	•	•
+ 1b	•	•	•				•	•	•	•	•	•			÷.	٠	•	•	•	•	•	•	•	•
+ 4a		0	•	•	0	•	•	•	•	•	•	•	٠	•	•	•	•	•	•	•	•	•	•	•
+ 4b	•	•	•					•	•	•	•	•		•	•	•	•	•	•	•	•	•	•	•
+ 6									•	•		C	6			0	•	•	•	•	•	•	•	۲
												Zn	(II)											
Anti-oligomer Antibody (A11) Anti-fibril Antibody (OC) Anti-Aβ Antibody (6E10)																								
Time (h)	0	1	2	3	4	6	12	24	0	1	2	3	4	6	12	24	0	1	2	3	4	6	12	24
<u>C</u>	•		•	•	•	•			•	٠	٠	•	,	٠	٠	٠	•	•	•	•	•	٠	•	•
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+ 2			•	•	•	•	•	0	۲	•	•			•			•	•	•	•	•	•	•	•
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Fig. S12 Formation of metal-free A β_{42} and metal–A β_{42} oligomers and fibrils upon treatment of flavonoids monitored by the dot blot assay. (a) Dot blots of metal-free A β_{42} and metal–A β_{42} species produced by incubation with flavonoids detected by anti-oligomer (A11), anti-fibril (OC), and anti-A β (6E10) antibodies. Note that the anti-fibril antibody, OC, may monitor both monomeric and aggregated forms of A β .²⁶ (b) Quantification of A11- and OC-detectable A β species. The value of *I* / *I*_{6E10} indicates the intensity of A11- or OC-detectable A β species quantified, relative to that detected from 6E10. Conditions: [A β_{42}] = 100 µM; [M(II)] = 100 µM; [flavonoid] = 200 µM (1% v/v DMSO); 20 mM HEPES, pH 7.4, 150 mM NaCl; 37 °C; 0, 1, 2, 3, 4, 6. 12, and 24 h; constant agitation.



Fig. S13 Absorbance spectra of **4**–**6** and **4b**. The compounds' Abs was shown to interfere with the excitation and emission wavelengths of ThT ($\lambda_{em}/\lambda_{ex} = 440/490$ nm). Conditions: [flavonoid] = 50 μ M (1% v/v DMSO); 20 mM HEPES, pH 7.4, 150 mM NaCl.



Fig. S14 1 H (400 MHz) and 13 C (100 MHz) NMR spectra of **6c** in DMSO- d_6 .



Fig. S15 1 H (400 MHz) and 13 C (100 MHz) NMR spectra of 6d in CD₃OD.



Fig. S16 1 H (400 MHz) and 13 C (100 MHz) NMR spectra of 6 in CD₃OD.



Fig. S17 Cytotoxicity of **1–6** and **4b** in 5Y cells. Viability (%) of cells incubated with various concentrations of **1–6** and **4b** was determined by the MTT assay. Cell survival was calculated in comparison to that of the cells treated with an equivalent amount of DMSO (1 % v/v). Error bars indicate the standard error from three independent experiments. Conditions: [flavonoid] = 5, 10, 25, 50, and 100 μ M (1 % v/v DMSO); 37 °C; 24 h.



Fig. S18 Chemical transformations of **6**. (a) Optical changes of **6** in the absence and presence of A β_{40} with or without metal ions, monitored by Abs spectroscopy. Conditions: $[A\beta_{40}] = 25 \ \mu$ M; $[M(II)] = 25 \ \mu$ M; [**6**] = 50 μ M; 20 mM HEPES, pH 7.4, 150 mM NaCl; 37 °C; 0–24 h; no agitation. (b) Change in the normalized absorbance of **6** with or without A β_{40} at 268 nm and 276 nm in the absence and presence of Cu(II), respectively, for 24 h.



Fig. S19 Interaction of **6** with monomeric A_{β40}. (a) Binding of **6** with A_{β40} recorded by ITC. The ITC thermogram (left) and binding isotherm (right) of **6** with A_{β40} monomer are shown. The solid line indicates the best fit of the ITC data to a one-set-of-sites binding model. Conditions: $[Aβ_{40}] = 40 \ \mu$ M; [**6**] = 800 μ M (1% v/v DMSO); 20 mM HEPES, pH 7.4; 10 °C. (b) 2D ¹H–¹⁵N SOFAST-HMQC NMR (800 MHz) spectra of ¹⁵N-labeled A_{β40} with and without **6**. (c) CSPs and peak intensity ratios of amino acid residues in ¹⁵N-labeled A_{β40} upon addition of **6**. Amino acid residues involved in the β-turn motif and the self-recognition site are highlighted in bold/green and bold/underline, respectively. The unresolved peaks (*i.e.*, Asp1, Ala2, Phe4, His6, His13, His14, and Asn27) are indicated with asterisks. Two horizontal lines indicate the average chemical shift (solid line) plus one standard deviation (dashed line). Conditions: [¹⁵N-labeled A_{β40}] = 33 μM; [**6**] = 100 μM (1% v/v DMSO); 20 mM HEPES, pH 7.4; 10% v/v D₂O; 10 °C.



Fig. S20 Interactions of **6** with metal-free and Cu(II)-added A β_{42} in the absence of O₂. Conditions: [A β_{42}] = 100 μ M; [Cu(II)] = 100 μ M; [**6**] = 200 μ M (1% v/v DMSO); 20 mM ammonium acetate, pH 7.4; 37 °C; 3 h; no agitation. The samples were prepared in a N₂-filled glove box and diluted by 10-fold with ddH₂O before injection into the mass spectrometer.



Fig. S21 Detailed analysis of ESI–MS² spectra shown in Fig. 7.



Fig. S22 Interactions of **6** with metal-free and Cu(II)-added A β_{40} . ESI–MS spectra of +3-charged A β_{40} and ESI–MS² analyses of the singly oxidized A β_{40} (1,449 *m*/*z*) were obtained upon treated of **6** in the (a and b) absence and (c and d) presence of Cu(II). The peaks highlighted as a red circle correspond to the singly oxidized A β_{40} . Detailed analysis of tandem spectra is shown in Fig. S23. Conditions: [A β_{40}] = 100 µM; [Cu(II)] = 100 µM; [**6**] = 200 µM (1% v/v DMSO); 20 mM ammonium acetate, pH 7.4; 37 °C; 3 h; constant agitation. The samples were diluted by 10-fold with ddH₂O before injection into the mass spectrometer.



Fig. S23 Detailed analysis of ESI–MS² spectra presented in Fig. S22.

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Appendix A. Cartesian coordinates of the optimized geometries.

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	С	-2.737648487	-4.718838215	-3.502676487
	Н	-0.722081840	-4.134303570	-0.823851526
	Н	-3.278165579	-4.202028751	-4.287439823
	С	-1.310209513	-6.073299408	-1.518036962
	С	-2.724905252	-6.105212212	-3.468658447
	Н	-0.746842980	-6.613808632	-0.764802456
	С	-2.017660141	-6.799302101	-2.471786261
	0	-2.085692406	3.868039131	-3.069872618
	Н	-2.097781658	4.269024849	-3.953572273
	0	-2.010292768	-8.160394669	-2.448388577
	Н	-2.539152145	-8.469491959	-3.200191259
	0	-3.382389307	-6.907441616	-4.373646736
	Н	-3.796572208	-6.357590199	-5.050311089

0	-2.129260063	2.857430220	-5.576642513
Н	-2.142997026	2.333509684	-6.392791271

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С	-2.227174282	0.591237605	-4.709803581			
С	-2.170015335	1.984547019	-4.525145054			
С	-2.025994062	2.529259443	-3.232281685			
С	-1.939639330	1.686829209	-2.118846416			
С	-1.994070649	0.311421514	-2.324744225			
С	-2.136181593	-0.275873810	-3.591377258			
С	-2.199601889	-1.709892035	-3.766739130			
С	-2.064286470	-2.514111996	-2.527297497			
С	-1.948624849	-1.817075610	-1.342328429			
0	-1.913680673	-0.493892968	-1.224406004			
н	-1.834000707	2.103016615	-1.124538183			
0	-2.344529867	-2.213070869	-4.901741982			
0	-2.366115093	0.142951161	-5.956882477			
н	-2.393357515	-0.855203450	-5.901386738			
н	-1.902377844	-2.285854816	-0.367777348			
С	-2.057842016	-3.970853567	-2.524805069			
С	-1.481456757	-4.690269470	-1.428758740			
С	-2.617203236	-4.712084770	-3.591741800			
н	-0.991438210	-4.152709484	-0.625745773			
н	-3.034947634	-4.191376686	-4.443635464			
С	-1.481178522	-6.065736294	-1.384778976			
С	-2.623768330	-6.091571331	-3.551775694			
н	-1.032920122	-6.612483501	-0.562942743			
С	-2.057865381	-6.793273449	-2.440963507			
0	-1.972460270	3.856505632	-3.055270910			
н	-2.044802904	4.304683685	-3.913393974			
0	-2.063628912	-8.119333267	-2.404462814			
Н	-2.489494324	-8.469717979	-3.206907511			
0	-3.144028664	-6.908358097	-4.495245934			
н	-3.495731831	-6.415612698	-5.250398159			
0	-2.246359587	2.850774765	-5.556621075			
н	-2.344753504	2.362206221	-6.389840603			

Appendix B. Vibrational 1	frequencies of the	optimized structures.
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====== 1 				-	
 24.49	51.72	95.37	106.71	140.05	201.85
233.04	255.42	257.96	270.13	295.82	350.96
367.67	379.61	411.25	427.87	469.73	504.48
517.28	569.29	619.27	621.92	632.84	641.48
641.97	664.70	675.32	703.29	720.29	747.31
755.72	789.06	813.15	821.70	852.58	864.70
895.16	919.52	941.84	981.97	1000.08	1007.90
1016.36	1048.88	1062.23	1115.15	1127.09	1137.83
1179.91	1191.28	1213.84	1216.03	1263.31	1276.42
1302.26	1338.89	1359.78	1370.16	1404.95	1435.48
1475.71	1492.59	1510.46	1538.67	1559.21	1625.79
1638.95	1655.07	1661.77	1677.09	1719.79	3178.85
3185.81	3195.80	3198.84	3206.74	3216.66	3232.09
3244.47	3249.46	3821.29			

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====== 19.00	======= 53.42	 101.80	 106.74	= 138.28	190.09		
228.52	247.82	258.11	258.75	277.59	331.09		
377.33	397.18	405.67	452.47	479.92	500.39		
506.93	564.34	577.99	611.27	618.88	625.52		
630.49	635.85	668.30	684.32	712.24	724.95		
739.85	793.21	822.55	840.70	849.72	867.15		
904.81	956.21	987.09	994.13	1011.20	1026.67		
1039.30	1055.10	1093.67	1113.43	1125.52	1146.17		
1180.19	1199.00	1220.83	1224.25	1259.31	1271.39		
1295.81	1306.54	1354.85	1383.00	1411.03	1435.44		
1464.23	1490.05	1512.55	1522.18	1540.59	1557.50		
1582.47	1607.96	1625.69	1648.57	1712.25	2462.03		
3205.85	3214.95	3221.66	3228.03	3230.26	3238.74		
3244.28	3264.77	3790.19					

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49.29	100.26	104.16	122.46	183.91		
227.68	235.40	260.81	277.62	313.71		
401.39	407.90	412.32	449.27	469.89		
486.01	543.45	572.42	614.91	632.13		
662.65	687.93	689.32	702.56	717.42		
780.71	791.29	849.35	849.90	853.47		
900.98	924.66	941.19	984.01	1008.70		
1045.85	1062.51	1083.98	1117.59	1158.59		
1191.30	1217.00	1229.81	1238.22	1261.61		
	49.29 227.68 401.39 486.01 662.65 780.71 900.98 1045.85 1191.30	49.29100.26227.68235.40401.39407.90486.01543.45662.65687.93780.71791.29900.98924.661045.851062.511191.301217.00	49.29100.26104.16227.68235.40260.81401.39407.90412.32486.01543.45572.42662.65687.93689.32780.71791.29849.35900.98924.66941.191045.851062.511083.981191.301217.001229.81	49.29100.26104.16122.46227.68235.40260.81277.62401.39407.90412.32449.27486.01543.45572.42614.91662.65687.93689.32702.56780.71791.29849.35849.90900.98924.66941.19984.011045.851062.511083.981117.591191.301217.001229.811238.22		

1276.59	1330.60	1340.27	1360.53	1369.29	1402.33
1442.10	1490.78	1511.60	1537.84	1560.54	1621.06
1637.77	1659.80	1665.08	1684.33	1731.62	3175.15
3180.98	3193.82	3197.01	3205.31	3225.22	3235.52
3251.64	3771.52	3841.35			

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27.42	47.62	89.45	98.68	125.02	176.42		
207.81	223.97	253.50	264.70	322.38	329.90		
367.77	384.93	405.05	409.76	450.53	456.19		
476.35	519.03	559.25	600.85	605.09	628.87		
629.45	652.87	678.33	691.70	694.29	697.19		
754.58	764.74	787.44	839.54	840.66	849.56		
874.96	889.11	933.85	948.74	988.11	996.55		
1015.12	1023.08	1058.02	1070.93	1117.93	1137.14		
1169.67	1190.26	1201.07	1218.96	1244.71	1259.15		
1273.06	1286.51	1349.73	1374.51	1375.16	1404.19		
1425.46	1490.72	1511.71	1536.30	1540.03	1545.87		
1590.81	1628.26	1650.96	1653.33	1702.57	3201.14		
3208.36	3215.88	3223.60	3223.84	3231.73	3232.38		
3257.60	3703.30	3796.04					

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-------33.97 49.08 106.65 118.93 131.65 185.61 211.82 247.57 265.35 274.32 325.32 365.85 388.85 411.16 433.68 450.30 471.81 485.78 497.49 532.37 566.76 617.74 620.74 632.35 665.53 666.26 681.82 701.12 702.69 749.21 780.91 792.51 820.28 843.85 853.91 867.60 912.72 938.10 943.83 963.98 985.68 1010.90 1013.61 1041.72 1058.43 1095.79 1119.28 1152.56 1191.96 1213.13 1220.23 1259.39 1261.09 1280.14 1326.29 1342.96 1358.53 1369.90 1396.20 1436.48 1451.62 1491.64 1516.09 1529.56 1538.92 1615.97 1639.99 1653.83 1659.09 1672.09 1709.42 3176.97 3182.81 3197.99 3199.74 3206.23 3211.55 3223.80 3227.41 3250.57 3764.93

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38.92	50.32	106.93	113.72	131.22	197.17
210.78	247.82	259.32	261.21	326.97	344.86
366.49	402.04	450.35	461.13	477.39	483.18

557.27	611.89	612.11	626.80	665.42
684.03	692.68	696.44	698.75	746.52
796.42	833.60	845.28	863.53	881.85
958.70	971.00	983.97	997.60	1011.08
1048.09	1060.38	1100.28	1126.72	1162.59
1206.83	1224.33	1254.89	1269.45	1286.23
1365.62	1379.40	1387.94	1409.83	1431.26
1507.15	1527.58	1537.80	1564.52	1579.73
1607.20	1624.94	1649.39	1667.94	3194.96
3209.52	3219.08	3225.40	3227.80	3238.46
3295.42	3616.01			
	557.27 684.03 796.42 958.70 1048.09 1206.83 1365.62 1507.15 1607.20 3209.52 3295.42	557.27611.89684.03692.68796.42833.60958.70971.001048.091060.381206.831224.331365.621379.401507.151527.581607.201624.943209.523219.083295.423616.01	557.27611.89612.11684.03692.68696.44796.42833.60845.28958.70971.00983.971048.091060.381100.281206.831224.331254.891365.621379.401387.941507.151527.581537.801607.201624.941649.393209.523219.083225.403295.423616.011	557.27611.89612.11626.80684.03692.68696.44698.75796.42833.60845.28863.53958.70971.00983.97997.601048.091060.381100.281126.721206.831224.331254.891269.451365.621379.401387.941409.831507.151527.581537.801564.521607.201624.941649.391667.943209.523219.083225.403227.80

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35.13	48.46	100.59	103.16	123.79	168.71	
198.51	247.65	250.01	257.10	284.23	290.72	
330.92	333.63	355.82	392.91	410.59	425.59	
470.09	474.06	484.05	521.05	583.64	600.44	
620.41	623.87	632.58	644.81	659.39	667.36	
701.20	717.03	724.01	737.56	791.50	818.00	
845.32	860.66	867.67	911.77	926.77	943.17	
985.03	1010.56	1013.93	1042.14	1060.29	1103.16	
1117.60	1123.47	1190.80	1191.88	1217.17	1233.22	
1249.83	1269.85	1309.52	1334.12	1365.76	1370.02	
1380.60	1424.71	1441.15	1465.95	1491.60	1521.80	
1537.58	1558.66	1616.97	1639.72	1655.75	1659.71	
1676.34	1720.12	3170.44	3176.21	3182.11	3197.12	
3206.02	3223.34	3241.76	3254.02	3769.13	3782.47	

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25.10	52.36	93.75	101.99	117.30	165.05
211.46	223.70	241.09	255.34	271.41	302.06
327.90	347.10	367.85	402.17	416.00	440.76
451.30	466.18	510.93	544.74	588.33	595.72
626.93	629.71	654.37	668.56	691.75	695.04
699.14	707.21	720.04	722.44	791.94	819.28
842.94	849.77	869.71	908.75	930.91	954.26
991.74	1013.11	1026.88	1039.60	1059.57	1105.10
1124.12	1137.64	1191.64	1199.56	1220.24	1232.24
1247.13	1277.21	1306.52	1344.95	1370.35	1380.58
1392.03	1421.02	1441.17	1484.43	1506.48	1529.37
1538.23	1567.88	1584.79	1604.92	1622.61	1631.44
1652.05	1666.53	3203.55	3211.33	3218.20	3222.54
3226.34	3234.92	3265.31	3354.96	3592.99	3798.94

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40.55	42.72	86.42	100.24	_ 103.03	139.45
167.96	191.25	199.22	239.18	249.86	269.65
294.63	323.43	330.92	344.47	358.56	388.12
396.34	410.45	426.69	472.45	476.22	487.88
530.89	595.32	607.86	630.50	640.72	643.14
662.35	671.63	687.09	701.38	736.05	761.99
792.50	831.20	847.13	860.21	868.81	934.88
944.10	946.26	987.60	1013.16	1013.53	1051.03
1062.88	1117.97	1120.07	1143.05	1192.36	1219.47
1240.30	1244.76	1270.46	1313.04	1335.23	1360.37
1371.63	1375.46	1420.41	1429.46	1461.94	1491.72
1513.28	1537.94	1548.94	1617.76	1637.62	1648.58
1658.96	1669.44	1715.78	3164.69	3176.21	3182.46
3195.78	3206.01	3222.18	3253.50	3762.98	3780.35

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13.60	41.88	85.95	93.36	101.81	134.49
166.52	194.88	211.66	240.25	250.79	267.55
289.89	324.69	325.37	346.41	390.87	399.32
412.36	422.77	459.26	461.96	484.49	519.72
575.99	604.11	625.71	626.48	642.07	651.73
665.26	686.70	691.11	692.80	713.60	770.70
793.44	839.90	856.43	859.15	873.72	933.06
956.00	971.83	993.87	1012.63	1028.41	1053.75
1066.50	1126.03	1132.65	1167.88	1200.73	1222.83
1235.22	1250.97	1280.07	1308.16	1339.95	1352.69
1376.66	1384.72	1419.30	1450.19	1484.07	1504.45
1513.43	1530.69	1539.33	1574.76	1595.46	1606.14
1629.19	1647.73	1662.40	3204.45	3213.11	3219.86
3228.06	3237.29	3274.23	3313.13	3629.92	3761.70

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======	=======	========	=======	=	
30.59	48.94	72.77	84.71	96.36	121.01
168.66	181.06	203.20	221.23	231.45	244.32
275.20	276.04	298.05	340.82	356.11	373.88
412.70	444.67	468.41	492.97	530.61	546.69
561.77	608.51	631.00	632.62	648.21	659.39
665.19	691.79	705.08	715.88	723.92	782.85
791.72	822.76	856.96	872.16	876.70	917.35
940.64	962.08	981.23	988.26	1005.98	1016.29
1054.31	1062.58	1077.21	1114.00	1122.86	1134.46
1177.36	1190.32	1197.88	1213.91	1230.75	1232.11

1246.32	1273.91	1278.54	1285.17	1336.62	1358.57
1368.69	1378.21	1412.74	1465.28	1474.76	1491.56
1503.78	1513.23	1517.03	1517.88	1536.97	1539.67
1550.06	1605.25	1632.92	1661.08	1662.16	1685.71
1728.28	2723.12	2787.92	2836.42	3022.60	3087.05
3155.06	3184.23	3193.74	3204.84	3213.77	3228.23
3235.36	3246.65	3248.61			

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41.13	47.83	70.73	83.11	- 98.24	114.29
139.41	151.27	176.24	186.57	208.76	224.70
239.91	266.74	278.02	294.16	311.51	353.22
373.24	400.68	414.74	455.99	488.53	510.88
551.61	591.80	596.64	612.62	623.38	635.79
653.93	668.05	688.28	702.26	722.01	753.53
793.75	836.45	847.97	868.20	876.85	900.48
940.45	956.47	980.94	996.50	1010.08	1026.89
1033.20	1051.18	1069.93	1090.26	1122.48	1132.31
1151.20	1172.47	1187.56	1198.12	1201.21	1213.99
1223.75	1252.61	1256.62	1285.31	1301.18	1336.99
1355.26	1378.98	1388.86	1414.04	1449.54	1477.79
1482.29	1486.63	1491.06	1505.76	1508.78	1512.85
1524.30	1534.61	1578.46	1606.27	1625.10	1646.76
1681.43	3044.68	3066.87	3121.93	3157.71	3187.82
3193.63	3193.86	3197.98	3208.97	3218.82	3223.98
3241.91	3254.23	3259.74			

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38.33	54.55	84.44	114.06		210.77
232.10	242.50	252.50	285.97	334.98	362.98
384.38	396.54	418.82	421.16	450.25	503.72
520.24	538.66	582.80	601.58	630.41	630.82
638.80	649.88	687.31	695.66	712.25	766.62
797.52	805.70	816.06	824.78	858.65	887.89
893.24	918.47	936.17	976.56	1000.82	1010.17
1016.84	1051.72	1076.32	1084.70	1113.51	1173.79
1188.96	1205.42	1214.66	1232.49	1265.41	1292.89
1333.15	1346.90	1365.37	1366.71	1405.51	1439.53
1471.37	1490.21	1507.97	1539.62	1557.42	1626.27
1636.76	1650.34	1661.63	1676.14	1713.32	3177.14
3185.11	3192.53	3194.93	3199.31	3205.99	3236.77
3238.46	3246.16	3820.79			

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38.42	54.74	95.16	114.89	151.09	200.72
224.51	247.32	258.24	272.77	337.49	352.64
377.69	397.04	419.64	442.66	463.21	498.35
508.18	524.20	570.52	603.63	607.64	622.17
626.53	630.73	669.96	683.32	683.94	765.74
797.06	802.57	832.61	838.37	857.60	862.51
925.06	931.94	961.31	998.51	1001.92	1006.66
1023.00	1040.78	1063.00	1087.05	1127.19	9 1182.17
1193.78	1204.07	1228.45	1238.33	1267.97	1307.58
1328.14	1340.96	1371.56	1387.85	1403.62	2 1433.32
1466.14	1481.59	1504.94	1518.03	1542.54	1565.00
1569.10	1590.29	1632.59	1657.69	1695.68	3 3080.92
3204.56	3210.72	3218.55	3220.56	3227.29	3245.16
3254.04	3272.12	3796.69			

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4a				-	
30.21	======= 31.16	======= 53.14	======= 69.19	= 78.10	83.25
98.80	126.96	131.72	140.08	151.66	179.71
190.38	212.08	230.51	233.00	244.07	277.84
289.36	323.14	339.83	345.82	362.52	412.74
442.01	448.84	480.85	497.84	518.83	563.07
606.29	632.44	634.10	647.56	655.54	669.63
705.27	706.67	734.97	753.46	775.73	788.75
825.31	856.59	865.05	875.40	925.10	940.21
974.22	981.23	1005.39	1013.98	1017.87	1055.15
1062.69	1076.31	1114.14	1129.39	1163.34	1171.47
1173.50	1177.88	1189.78	1201.70	1209.48	1213.66
1222.05	1232.58	1249.51	1273.65	1296.47	1333.25
1337.74	1368.30	1381.80	1396.46	1448.50	1472.47
1485.10	1491.43	1492.30	1498.15	1504.01	1506.44
1515.71	1516.91	1524.72	1527.69	1539.33	1604.93
1632.69	1647.09	1662.60	1680.64	1731.37	3023.09
3023.93	3026.73	3089.49	3096.41	3125.21	3144.52
3155.82	3162.35	3184.09	3193.77	3205.11	3214.70
3226.85	3243.90	3245.56			

4a⁺					
======= 31 81	32 53	======= 55 25	======= 69 28	= 85.54	106 25
118 57	140.39	158 73	160.53	177 62	192.09
206 52	217 58	228 79	235.00	245.23	274 68
292.88	313.04	340.88	364.43	384.64	408.28
421.53	446.14	457.90	484.08	500.34	555.10
608.50	610.87	629.76	642.23	655.32	660.77
	2.5101				

691.79	700.31	708.35	728.46	758.49	788.77
841.57	853.01	876.35	880.24	902.55	944.75
948.10	979.63	987.25	1013.57	1015.33	1020.36
1040.70	1060.76	1120.02	1133.68	1151.54	1157.86
1171.31	1172.48	1196.73	1198.41	1209.13	1214.44
1219.66	1235.71	1260.37	1282.83	1312.07	1339.37
1363.05	1373.41	1374.87	1420.58	1436.87	1448.95
1479.00	1485.55	1491.66	1495.01	1497.85	1499.38
1502.25	1505.53	1523.35	1533.56	1536.82	1548.45
1628.93	1649.21	1655.62	1667.76	1728.22	3050.35
3080.72	3091.92	3130.17	3176.52	3191.52	3192.52
3198.22	3205.52	3205.80	3213.50	3214.11	3221.55
3228.65	3253.41	3258.50			

4b

29.25	32.22	72.77	117.53	127.64	145.39			
185.40	217.55	238.57	264.39	284.40	288.97			
305.40	332.24	338.07	353.21	360.11	395.27			
413.56	446.75	462.33	500.55	520.88	543.78			
586.52	597.81	602.62	629.11	635.91	646.54			
665.37	686.56	710.12	763.45	772.37	783.07			
810.15	814.43	868.19	908.41	933.73	944.88			
987.79	1009.02	1022.83	1047.54	1068.55	1078.89			
1116.86	1128.01	1173.92	1191.91	1206.77	1231.39			
1250.36	1263.52	1301.16	1320.26	1337.89	1352.99			
1379.35	1400.85	1427.92	1438.96	1488.40	1526.25			
1537.95	1543.69	1614.88	1639.46	1656.37	1660.53			
1688.33	1727.23	3127.62	3152.66	3156.33	3195.79			
3211.90	3236.66	3246.68	3751.00	3810.19	3841.19			

4b'⁺ _____ 38.52 56.09 86.31 112.12 127.71 170.67 201.65 231.70 245.32 269.43 311.47 316.32 333.78 336.86 374.28 397.41 406.45 414.18 442.38 460.89 517.54 535.80 560.93 590.46 605.46 611.39 621.66 641.90 652.56 689.56 692.68 702.09 765.39 780.64 800.28 826.85 831.94 850.04 909.16 949.59 951.95 990.99 991.39 1011.53 1019.74 1049.58 1074.41 1114.34 1122.64 1136.80 1168.59 1195.54 1217.25 1229.20 1272.94 1308.62 1332.24 1342.60 1364.87 1377.49 1392.69 1424.55 1427.48 1469.10 1487.20 1528.33 1530.67 1558.63 1571.00 1595.99 1597.88 1635.32 1658.34 1692.64 2870.83 3193.97 3203.98 3213.98

3222.45 3224.96 3254.00 3263.83 3707.84 3818.78

6 _____ 32.83 40.59 64.65 108.26 116.63 156.56 185.03 206.68 216.88 226.28 233.52 276.87 290.14 309.28 310.96 331.95 337.33 365.45 374.43 412.68 414.18 441.84 453.74 465.32 467.19 480.74 488.55 530.69 588.33 599.54 602.37 612.49 632.25 636.20 651.75 661.39 691.03 708.59 767.10 780.96 803.91 821.63 830.07 833.27 855.92 860.74 899.52 918.12 940.90 994.12 1081.30 1084.04 1135.60 1150.68 1182.19 1190.51 1211.69 1234.43 1256.82 1277.81 1306.69 1323.98 1342.32 1353.37 1380.98 1382.50 1411.77 1425.25 1447.94 1466.78 1481.56 1518.45 1556.52 1574.81 1625.51 1652.31 1654.88 1669.4 1672.73 1714.61 3170.99 3193.27 3216.29 3219.62 3239.05 3243.96 3763.48 3773.02 3776.53 3840.53

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149.83								
292.46								
368.56								
464.47								
597.42								
665.45								
826.11								
919.91								
1166.57								
1279.34								
1401.95								
1511.15								
1640.40								
3243.37								
3813.76								