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Direct Conversion of Amino Acids to Oxetanol Bioisosteres via Photoredox Catalysis

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A. SUPPLEMENTARY TABLES S1 & S2 AND SUPPLEMENTARY FIGURES S1 & S2

Supplementary Table S1. Complete Data on Optimization of Cr-mediated Reaction



entry	photocatalyst	base	additive	Cr catalyst	solvent	yield (%)ª
1	4CzIPN (1 mol%)	-	-	CrCl₃ (5 mol%)	DMA	0
2	4CzIPN (1 mol%)	CsOAc	-	CrCl₃ (5 mol%)	DMA	7
3	4CzIPN (1 mol%)	CsOAc	TMSCI (0.5 equiv)	CrCl₃ (5mol%)	DMA	22
4	4CzIPN (1 mol%)	CsOAc	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMF	25
5	4CzIPN (2.5 mol%)	CsOAc	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMF	22
6	4CzIPN (5 mol%)	CsOAc	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMF	25
7	[Ir(dF(CF ₃)ppy) ₂ (dtbbpy)] [PF ₆] (1 mol%)	CsOAc	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMF	17
8	[Ir(dF(CF ₃) ₂ ppy) ₂ (bpy)] [PF6] (1 mol%)	CsOAc	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMF	24
9	4CzIPN (1 mol%)	CsOAc	TMSCI (0.5 equiv)	CrCl ₃ (10 mol%)	DMF	25
10	4CzIPN (1 mol%)	CsOAc	TMSCI (0.5 equiv)	CrCl ₃ (25 mol%)	DMF	0
11	4CzIPN (1 mol%)	CsOAc	TMSCI (0.5 equiv)	CrCl₃ (0.5 equiv)	DMF	0
12	4CzIPN (1 mol%)	CsOAc	TMSCI (0.25 equiv)	CrCl₃ (5 mol%)	DMF	18
13	4CzIPN (1 mol%)	CsOAc	TMSCI (1 equiv)	CrCl₃ (5 mol%)	DMF	0
14	4CzIPN (1 mol%)	CsOAc	TMSCI (2 equiv)	CrCl₃ (5 mol%)	DMF	0
15	4CzIPN (1 mol%)	NaOAc	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMF	29
16	4CzIPN (1 mol%)	LiOAc	TMSCI (1 equiv)	CrCl ₃ (10 mol%)	DMF	25

entry	photocatalyst	base	additive	Cr catalyst	solvent	yield (%) ^a
17	4CzIPN (1 mol%)	KOAc	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMF	27
18	4CzIPN (1 mol%)	Cs ₂ CO ₃	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMF	0
19	4CzIPN (1 mol%)	Li ₂ CO ₃	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMF	0
20	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMF	33
21	4CzIPN (1 mol%)	NaHCO ₃	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMF	11
22	4CzIPN (1 mol%)	KHCO ₃	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	DMF	8
23	4CzIPN (1 mol%)	NaCO ₂ C ₂ H ₅	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMF	21
24	4CzIPN (1 mol%)	NaBz	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	DMF	19
25	4CzIPN (1 mol%)	NaCO ₂ C ₃ H ₇	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	DMF	19
26	4CzIPN (1 mol%)	CsCO ₂ Ad	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	DMF	31
27	4CzIPN (1 mol%)	DBU	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	DMF	0
28	4CzIPN (1 mol%)	2,6-lutidine	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	DMF	0
29	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	CH₃CN	40
30	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	THF	38
31	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	DMSO	23
32	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	DMA	29
33	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	DCM	33
34	4CzIPN (1 mol%)	CsOPiv	TIPSCI (0.5 equiv)	CrCl ₃ (5 mol%)	CH₃CN	37
35	4CzIPN (1 mol%)	CsOPiv	TESCI (0.5 equiv)	CrCl ₃ (5 mol%)	CH₃CN	37
36	4CzIPN (1 mol%)	CsOPiv	TESOTf (0.5 equiv)	CrCl₃ (5 mol%)	CH₃CN	38
37	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	-	CH₃CN	32

entry	photocatalyst	base	additive	Cr catalyst	solvent	yield (%)ª
38	4CzIPN (1 mol%)	CsOPiv	-	-	CH₃CN	25
39	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	CH₃CN	60 ^b
40	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	CH₃CN	55 ^c
41	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	CH₃CN	50 ^d
42	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	-	CH₃CN	50 ^b
43	4CzIPN (1 mol%)	CsOPiv	-	CrCl ₃ (5 mol%)	CH₃CN	49 ^b
44	4CzIPN (1 mol%)	-	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	CH₃CN	0 ^b
44	4CzIPN (1 mol%)	CsOPiv	-	-	CH₃CN	47 ^b
45	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	CH₃CN	42 ^{b,e}
46	4CzIPN (1 mol%)	CsOPiv	TMSCI (0.5 equiv)	CrCl ₃ (5 mol%)	CH₃CN	0 <i>b,f</i>
47	-	CsOPiv	TMSCI (0.5 equiv)	CrCl₃ (5 mol%)	CH₃CN	0 <i>b</i>

^aYields based on ¹H-NMR analysis of crude reaction product and are relative to *N*-phenyl glycine. ^b1 equiv *N*-phenyl glycine to 1 equiv 3-oxetanone. ^c1 equiv *N*-phenyl glycine to 2 equiv 3-oxetanone. ^d1 equiv *N*-phenyl glycine to 3 equiv 3-oxetanone. ^eReaction mixture was not degassed with Ar. ^fReaction mixture was not irradiated with blue LEDs.

Supplementary Table S2. Complete Data on Optimization of Cr-free Reaction



entry	3-oxetanone (equiv)	photocatalyst	base (1.2 equiv)	solvent	yield (%) ^a
1	1	4CzIPN (1 mol%)	CsOPiv	CH₃CN	15 ^b
2	1	4CzIPN (1 mol%)	CsOPiv	CH₃CN	6 ^{<i>c</i>}
3	1	4CzIPN (1 mol%)	CsOPiv	CH₃CN	9 <i>d</i>
4	1	4CzIPN (1 mol%)	CsOPiv	CH₃CN	8
5	2	4CzIPN (1 mol%)	CsOPiv	CH₃CN	10
6	3	4CzIPN (1 mol%)	CsOPiv	CH₃CN	11
7	3	4CzIPN (2 mol%)	CsOPiv	CH₃CN	13
8	3	4CzIPN (2 mol%)	CsOPiv	DCE	57
9	3	4CzIPN (2 mol%)	CsOPiv	CH_2CI_2	89
10	3	4CzIPN (2 mol%)	CsOPiv	iPrOH	78
11	3	4CzIPN (2 mol%)	CsOPiv	DMF	0
12	3	4CzIPN (2 mol%)	CsOPiv	toluene	82
13	3	4CzIPN (2 mol%)	Na ₂ CO ₃	CH_2CI_2	24
14	3	4CzIPN (2 mol%)	DIPEA	CH_2CI_2	21
15	3	5CzBN (2 mol%)	CsOPiv	CH_2CI_2	66
16	3	3CzCI-IPN (2 mol%)	CsOPiv	CH_2CI_2	63
17	3	3DPACI-IPN (2 mol%)	CsOPiv	CH_2Cl_2	50
18	3	3DPAF-IPN (2 mol%)	CsOPiv	CH_2Cl_2	74
19	3	3DPA2FBN (2 mol%)	CsOPiv	CH_2CI_2	31
20	3	10-Phen-MesAcr (2 mol%)	CsOPiv	CH_2Cl_2	30
21	3	2,7-dimethyl-10-Phen-MesAcr (2 mol%)	CsOPiv	CH ₂ Cl ₂	55
22	3	10-Me-MesAcr (2 mol%)	CsOPiv	CH_2CI_2	5
23	3	[Ir{dF(CF ₃) ₂ ppy} ₂ (bpy)]PF ₆ (2 mol%)	CsOPiv	CH ₂ Cl ₂	43
24	3	[Ir{dF(CF ₃) ₂ ppy} ₂ (dtbbpy)]PF ₆ (2 mol%)	CsOPiv	CH ₂ Cl ₂	30

25	3	[Ir(dtbbpy)(ppy)2]PF6 (2 mol%)	CsOPiv	CH_2CI_2	63
26	3	[Ir{dF(Me) ₂ ppy}2(dtbbpy)]PF ₆ (2 mol%)	CsOPiv	CH_2CI_2	62
27	3	fac-[lr(dFppy)₃] (2 mol%)	CsOPiv	CH_2CI_2	56
28	3	fac-[lr(dppy)3] (2 mol%)	CsOPiv	CH_2CI_2	50
29	3	4CzMeBN (2 mol%)	CsOPiv	CH ₂ Cl ₂	80

^{*a*}Determined by ¹H-NMR analysis using mesitylene as an internal standard. ^{*b*}1.0 equiv of TMSCI was used as an additive. Reaction time = 20 h. ^{*c*}1.0 equiv of CrCl₃ was used as an additive. ^{*d*}Reaction time = 20 h. DCE = dichloroethane, DMF = N, N-dimethylformamide.



Supplementary Figure S1. Competition experiments with CH₃OD (olive). The alkyl-Cr $(8 + 2 \rightarrow 10)$ and carbanion $(9 + 2 \rightarrow 11)$ pathways are shown separately for clarity.



Supplementary Figure S2. Proposed flux through competing reaction pathways, consistent with the experimental data. See Supplementary Figure S1 above for full-scale reaction scheme. Reaction arrows are in blue; propagation cycles are in teal; relative flux is represented by arrow size. (a) The Crmediated reaction proceeds primarily through the alkyl-Cr addition pathway ($8 + 2 \rightarrow 10$), including the propagation cycle via Cr-carboxylate 12. (b) In the presence of methanol but absence of oxetanone, only the protodecarboxylation products 13a/b can be formed. (c) In the presence of both methanol and oxetanone, flux increases through the radical pathway, thus paradoxically decreasing protodecarboxylation to 13a/b. (d) In the absence of TMSCI, the Cr-dependent propagation cycle is blocked or decreased, thus decreasing the quantum yield ($\Phi = 1.6$ vs 5.1 with TMSCI); propagation may still occur to some extent via TMSCI-independent formation of Cr-carboxylate 12 and/or by the SET propagation cycle in the radical addition pathway. (e) The Cr-free reaction proceeds primarily through the radical addition pathway (6 + 2 \rightarrow 7), including the SET propagation cycle that regenerates radical 6. (f) In the presence of methanol but absence of oxetanone, the radical intermediate 6 has no productive pathway forward other than to shunt to photocatalyzed reduction to carbanion intermediate 9 and then to protodecarboxylation products 13a/b. (g) In the presence of both methanol and oxetanone, flux increases through the radical addition pathway, resulting in increased yield of the 3-oxetanol product 3a.

B. MATERIALS AND METHODS

Reagents

Reagents were obtained from Aldrich Chemical (www.sigma-aldrich.com) or Acros Organics/Tokyo Chemical Industry (www.fishersci.com) and used without further purification. Commercially available α -amino acids were obtained from the following suppliers and used without further purification: *N*-phenyl glycine (**1a**) from Sigma Aldrich (www.sigmaaldrich.com); (2-(phenylamino)propanoic acid (**1b**), 1-phenylpyrrolidine-2-carboxylic acid (**1h**), 2-(2,3-dihydro-1*H*-indol-1-yl)acetic acid hydrochloride) (**1i**), 2-methyl-2-(phenylamino)propanoic acid (**1k**), 2-[(4-methylphenyl)amino]acetic acid (**1u**), *N*-(4-bromophenyl)-glycine (**1v**), and 2-[(carboxymethyl)-(phenyl) amino] acetic acid (**1y**) from Enamine (www.enamine.net); *N*-(4-fluorophenyl)-glycine (**1w**) and from Fisher Scientific (www.fishersci.com); *N*-methyl *N*-phenylalanine (**1j**), morpholin-4-yl acetic acid (**4a**), (4-methyl-piperazin-1-yl acetic acid (**4b**), Boc-(4-carboxymethyl)piperazine (**4c**), and piperidin-1-yl-acetic acid hydrochloride (**4d**), from Matrix Scientific (www.fishersci.com); *N*,*N*-dipropyl-L-alanine (**4e**) from Tokyo Chemical Industry (https://www.tcichemicals.com/US/en/). Optima or HPLC grade solvents were obtained from Fisher Scientific (www.fishersci.com) acid and purified on a solvent drying system as described unless otherwise indicated.¹

Reactions

All reactions were performed in flame-dried glassware under positive Ar pressure with magnetic stirring unless otherwise noted. Liquid reagents and solutions were transferred thru rubber septa via syringes flushed with Ar prior to use.

Photochemical reactions were performed using a PR160 Rig with fan kit and irradiated with four PR160L LEDs (40 W, $\lambda_{max} = 456$ nm) from Kessil (www.kessil.com). This apparatus was enclosed within an aluminum covered box. The reaction vials were placed 5 cm from the LEDs and the temperature was measured to be between 25 °C to 30 °C using this configuration.



Supplementary Figure S2. Photochemical reaction apparatus using Kessil PR160L LEDs (40 W, λ_{max} = 456 nm) Rig and fan kit.



Supplementary Figure S3. Emission spectrum and intensity provided by the manufacturer, Kessil. Model PR160L 456 nm used for the reactions described.

Chromatography

TLC was performed on 0.25 mm E. Merck silica gel 60 F254 plates and visualized under UV light (254 nm) or by staining with potassium permanganate (KMnO₄), or cerium ammonium molybdenate (CAM). Silica flash chromatography was performed manually on E. Merck 230–400 mesh silica gel 60 or on an ISCO CombiFlash Rf+ instrument with RediSep silica gel normal phase columns or RediSep Gold silica gel normal phase columns with UV detection at 254 nm.

Analytical Instrumentation

NMR spectra were recorded on a Bruker UltraShield Plus 500 MHz Avance III NMR or UltraShield Plus 600 MHz Avance III NMR with DCH CryoProbe at 24 °C in CDCl₃ unless otherwise indicated. Chemical shifts are expressed in ppm relative to TMS (¹H, 0 ppm) or solvent signals: CDCl₃ (¹³C, 77.0 ppm), C₆D₆ (¹H, 7.16 ppm; ¹³C, 128.0 ppm) or acetone-d₆ (¹³C, 206.2 ppm); coupling constants are expressed in Hz. NMR spectra were processed using Bruker TopSpin. Mnova (www.mestrelab.com/software/mnova-nmr). or nucleomatica iNMR (www.inmr.net) software. Differential pulse voltammetry experiments were recorded using a 660E potentiostat/galvanostat from CH Instruments (https://www.chinstruments.com/). Mass spectra were obtained at the MSKCC Analytical Core Facility on a Waters Acuity SQD LC-MS. High resolution mass spectra were obtained on a Waters Acuity Premiere XE TOF LC-MS by electrospray ionization (ESI).

C. SYNTHESIS OF N-ARYL-α-AMINO ACIDS (1c-g,l-t,x)

N-Aryl α -amino acids **1c–g**, **1l–t**, and **1x**, which were not commercially available, were synthesized using a literature protocol² with the minor modification of running the reaction at 60 °C instead of rt.



General Procedure for Ullmann Coupling

In a 50-mL roundbottom flask, CuI (23 mg, 0.25 mmol, 10 mol%), Cs₂CO₃ (1.63 g, 5 mmol, 2 equiv), and the appropriate α -amino acid (3 mmol, 1.2 equiv) were combined. The mixture was evacuated and refilled with Ar three times, then dissolved in DMF (3.7 mL). The appropriate aryl iodide (2.5 mmol, 1 equiv) and 2-isobutyrylcyclohexanone (0.82 mL, 0.5 mmol, 20 mol%) were added. The reaction mixture was heated to 60 °C and stirred until complete conversion as determined by TLC and ¹H-NMR analysis. The mixture was allowed to cool to rt, then diluted with 1*N* HCl to pH = 4, and extracted with 25 mL EtOAc (3x). The combined organic extracts were washed with 30 mL of water (2x) and brine, dried (Na₂SO₄), filtered, and concentrated by rotary evaporation. Purification by silica flash chromatography (10–30% EtOAc in hexanes, with 1% AcOH) afforded the *N*-aryl α -amino acid product **1**.



N-Phenyl-L-leucine (1c): Prepared from iodobenzene and L-leucine in 78% yield as a white solid. The NMR spectra of 1c matched those reported in literature.²



*N***-Phenyl-L-phenylalanine (1d)**: Prepared from iodobenzene and L-phenylalanine in 66% yield as a white solid. The NMR spectra of **1d** matched those reported in literature.²



*N***-Phenyl-L-tryptophan (1e)**: Prepared from iodobenzene and L-tryptophan in 51% yield as a brown solid. The NMR spectra of **1e** matched those reported in literature.²



N-Phenyl-L-valine (1f): Prepared from iodobenzene and L-valine in 72% yield as a light yellow solid. The NMR spectra of **1e** matched those reported in literature.²



N-Phenyl-L-isoleucine (1g): Prepared from iodobenzene and L-isoleucine in 70% yield as a light yellow solid. The NMR spectra of **1f** matched those reported in literature.²



*N***-Phenyl-L-methionine (11)**: Prepared from iodobenzene and L-methionine in 78% yield as an off-white solid. The NMR spectra of **11** matched those reported in literature.²



 N^{6} -(*tert*-butoxycarbonyl)- N^{2} -phenyl-L-lysine (1m): Prepared from iodobenzene and N^{e} -(*tert*-butoxycarbonyl)-L-lysine in 92% yield as an off-white solid. The NMR spectra of 1m matched those reported in literature.³



*O***-benzyl-***N***-phenyl-L-serine (1n)**: Prepared from iodobenzene and *O*-benzyl-L-serine in 45% yield as an light yellow solid. **TLC**: R_f 0.30 (1% acetic acid in 2:3 EtOAc/Hexanes). ¹H NMR (600 MHz, CD₂Cl₂) δ 7.39 – 7.27 (m, 5H), 7.23 – 7.15 (m, 2H), 6.84 – 6.75 (m, 1H), 6.74 – 6.60 (m, 2H), 4.57 (s, 2H), 4.20 (s, 1H, N–H), 3.96 (dd, J = 9.5, 4.0 Hz, 1H), 3.83 (dd, J = 9.5, 4.2 Hz, 1H).¹³C NMR (151 MHz, CD₂Cl₂) δ 146.7, 137.8, 129.7, 128.9, 128.2, 119.6, 114.2, 73.8, 70.1, 57.9. **ESI-MS**: Calcd for C₁₆H₁₈NO₃ ([M+H]⁺) 272.1287; found 272.1284.



O-(*tert*-butyl)-*N*-phenyl-L-serine (10): Prepared from iodobenzene and *O*-(*tert*-butyl)-L-serine in 70% yield as an light yellow solid. TLC: R_f 0.34 (1% acetic acid in 3:2 EtOAc/Hexanes). ¹H NMR (600 MHz, CD₂Cl₂) δ 7.25 – 7.18 (m, 2H), 6.85 – 6.79 (m, 1H), 6.72 – 6.65 (m, 2H), 4.07 (dd, J = 5.9, 4.2 Hz, 1H), 3.88 (dd, J = 9.0, 4.2 Hz, 1H), 3.63 (dd, J = 9.0, 5.9 Hz, 1H), 1.24 (s, 9H). ¹³C NMR (151 MHz, CD₂Cl₂) δ 173.0, 160.9, 146.8, 129.8, 119.8, 114.4, 75.2, 61.9, 58.1, 31.4, 28.5, 27.5. ESI-MS: Calcd for C₁₃H₂₀NO₃ ([M+H]⁺) 238.1443; found 238.1447.



(*S*)-3-(4-(*tert*-butoxy)phenyl)-2-(phenylamino)propanoic acid (1p): Prepared from iodobenzene and *O*-(*tert*-butyl)-L-tyrosine in 75% yield as an light yellow solid. TLC: R_f 0.32 (1% acetic acid in 2:3 EtOAc/Hexanes. ¹H NMR (600 MHz, CDCl₃) δ 7.22 – 7.14 (m, 2H), 7.10 (d, *J* = 8.3 Hz, 2H), 6.98 – 6.89 (m, 2H), 6.79 (dd, *J* = 1497.9, 7.1 Hz, 1H), 6.59 (d, *J* = 7.9 Hz, 2H), 4.27 (t, *J* = 6.6 Hz, 1H), 3.23 (dd, *J* = 14.0, 5.3 Hz, 1H), 3.07 (dd, *J* = 14.1, 7.3 Hz, 1H), 1.33 (s, 9H). ¹³C NMR (151 MHz, CDCl₃) δ 176.5, 154.7, 146.2, 130.7, 129.9, 129.6, 124.6, 119.5, 114.2, 78.7, 58.4, 37.8, 28.9, 23.4, 20.7. ESI-MS: Calcd for C₁₉H₂₂NO₃ ([M–H]⁻)312.1600; found 312.1595.



(*S*)-4-(*tert*-butoxy)-4-oxo-2-(phenylamino)butanoic acid (1q): Prepared from iodobenzene and 4-*O*-(*tert*-butyl)-L-aspartic acid in 70% yield as a light yellow solid. TLC: R_f 0.20 (1% acetic acid in 2:3 EtOAc/Hexanes. ¹H NMR (600 MHz, CDCl₃) δ 7.25 – 7.18 (m, 2H), 6.84 (d, *J* = 1.0 Hz, 1H), 6.73 – 6.66 (m, 2H), 4.40 – 4.29 (m, 1H), 2.95 – 2.86 (m, 1H), 2.82 (dd, *J* = 16.4, 5.4 Hz, 1H), 1.44 (s, 9H). ¹³C NMR (151 MHz, CDCl₃) δ 170.4, 145.86, 129.7, 119.8, 114.5, 114.5, 82.5, 54.3, 37.8, 28.3, 28.2. ESI-MS: Calcd for C₁₄H₁₉NO₄ ([M+H]⁺) 288.1204; found 288.1204.



(*S*)-5-(benzyloxy)-5-oxo-2-(phenylamino)pentanoic acid (1r): Prepared from iodobenzene and (*S*)-2-amino-5-(benzyloxy)-5-oxopentanoic acid in 20% yield as a white solid. TLC: R_f 0.18 (1% acetic acid in 2:3 EtOAc/Hexanes. ¹H NMR (600 MHz, CD₂Cl₂) δ 7.40 – 7.26 (m, 5H), 7.21 – 7.10 (m, 2H), 6.79 – 6.73 (m, 1H), 6.61 (d, *J* = 7.7 Hz, 2H), 5.10 (s, 2H), 4.13 – 4.10 (m, 1H), 2.66 – 2.52 (m, 2H), 2.32 – 2.23 (m, 1H), 2.16 – 2.07 (m, 1H). ¹³C NMR (151 MHz, CD₂Cl₂) δ 177.8, 177.5, 173.2, 146.9, 136.4, 129.8, 128.9, 128.6, 128.6, 119.3, 119.3, 114.0, 114.0, 66.9, 56.6, 56.6, 30.9, 28.0, 28.0. ESI-MS: Calcd for C₁₈H₁₈NO₄ ([M+H]⁺) 312.1236; found 312.1242.



*N*²-**phenyl**-*N*⁴-**trityl**-*L*-**asparagine (1s)**: Prepared from iodobenzene and *N*⁴-trityl-*L*-asparagine in 95% yield as a light yellow solid. **TLC**: *R*_f 0.35 (1% acetic acid in 2:3 EtOAc/Hexanes. ¹**H NMR** (600 MHz, CD₃CN) δ 7.69 (s, 1H), 7.26 – 7.17 (m, 15H), 7.16 – 7.12 (m, 2H), 6.70 (tt, *J* = 7.3, 1.1 Hz, 1H), 6.65 – 6.61 (m, 2H), 5.42 (s, 1H), 4.32 (t, *J* = 6.2 Hz, 1H), 2.84 – 2.75 (m, 2H). ¹³**C NMR** (151 MHz, CD₃CN) δ 174.2, 170.7, 147.8, 145.5, 130.1, 129.6, 128.6, 127.7, 119.0, 114.40, 71.2, 55.3, 54.0, 39.7, 20.6. **ESI-MS**: Calcd for C₂₉H₂₆N₂O₃Na ([M+Na]⁺) 473.1841; found 473.1837.



N-Phenyl-L-aspartic acid (1t): Reaction solvent was changed to methanol. Prepared from iodobenzene and L-aspartic acid in 15% yield as an off-white solid. The NMR spectra of 1t matched those reported in literature.⁴



N-Phenyl(4-trifluoromethane)-glycine (1x): Prepared from 4-trifluoromethane iodobenzene and glycine in 61% yield as a white solid. The NMR spectra of 1f matched those reported in literature.⁵

D. CONVERSION OF *N*-ARYL α-AMINO ACIDS (1) TO 3-OXETANOLS (3)



General Procedure 1 for Cr-mediated decarboxylative addition

In a 4-mL glass vial, 4CzIPN (1.58 mg, 2 μ mol, 1 mol%), CrCl₃ (1.59 mg, 10 μ mol, 5 mol%), cesium pivalate (56 mg, 0.24 mmol, 1.2 equiv), and the appropriate *N*-aryl α -amino acid **1** (0.2 mmol, 1 equiv) were combined. CH₃CN (0.25 mL) was added followed by 3-oxetanone (**2**) (12.8 μ L, 0.2 mmol, 1 equiv). The mixture was degassed with Ar for 1 min, then TMSCl (12.5 μ L, 0.1 mmol, 0.5 equiv) was added. The reaction was exposed to blue LED light (456 nm) at 25 °C for 20 h. The reaction mixture was filtered over celite and concentrated by rotary evaporation. Purification by silica flash chromatography (10–40% EtOAc in hexanes) yielded the 3-oxetanol product **3**.

General Procedure 2 for Cr-free decarboxylative addition

In a 4-mL glass vial, 4CzIPN (1.58 mg, 2 μ mol, 1 mol%), cesium pivalate (56 mg, 0.24 mmol, 1.2 equiv), and the appropriate *N*-aryl α -amino acid **1** (0.2 mmol, 1 equiv) were combined. Methylene chloride (0.4 mL) was added followed by 3-oxetanone (**2**) (38.4 μ L, 0.6 mmol, 3 equiv). The mixture was degassed with Ar for 1 min. The reaction was exposed to blue LED light (456 nm) at 25 °C for 2 h. The reaction mixture was filtered over celite and concentrated by rotary evaporation. Purification by silica flash chromatography (10–40% EtOAc in hexanes) yielded the 3-oxetanol product **3**.



3-((Phenylamino)methyl)oxetan-3-ol (3a): Prepared from **1a** by General Procedure 1 in 60% yield or by General Procedure 2 in 60% yield as a light brown solid. **TLC**: R_f 0.16 (2:3 EtOAc/Hexanes). ¹**H NMR** (600 MHz) δ 7.22 (t, J = 7.7 Hz, 2H), 6.80 (t, J = 7.4 Hz, 1H), 6.73 (d, J = 7.5 Hz, 2H), 4.66 (d, J = 7.1 Hz, 2H), 4.58 (d, J = 7.4 Hz, 2H), 3.54 (s, 2H). ¹³**C NMR** (151 MHz, CDCl₃) δ 148.0, 129.6, 118.8, 113.7, 82.5, 73.7, 50.5. **ESI-MS**: Calcd for C₁₀H₁₃NO₂ ([M+H]⁺) 180.1025; found 180.1019.



3b

3-(1-(Phenylamino)ethyl)oxetan-3-ol (3b): Prepared from **1b** by General Procedure 1 in 69% yield or by General Procedure 2 in 96% yield as a light brown solid. **TLC**: $R_f 0.32$ (2:3 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CDCl₃) δ 7.24 – 7.17 (m, 2H), 6.77 (t, 1H), 6.71 (d, 2H), 4.68 – 4.55 (m, 4H), 3.92 (q, J = 6.5 Hz, 1H), 1.24 (d, J = 6.5 Hz, 3H). ¹³**C NMR** (151 MHz, CDCl₃) δ 146.8, 129.6, 118.8, 114.4, 82.6, 81.6, 76.5, 54.0, 13.8. **ESI-MS**: Calcd for C₁₁H₁₅NO₂ ([M+H]⁺) 194.1181; found 194.1184.



3-(2-Phenyl-1-(phenylamino)ethyl)oxetan-3-ol (3c): Prepared from **1c** by General Procedure 1 in 67% yield or by General Procedure 2 in 72% yield as a white solid. **TLC**: R_f 0.42 (2:3 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CD₂Cl₂) δ 7.26 – 7.19 (m, 4H), 7.19 –7.15 (m, 1H), 7.14 – 7.08 (m, 2H), 6.68 – 6.59 (m, 3H), 4.55 (d, J = 7.5 Hz, 1H), 4.41 (d, J = 8.0 Hz, 1H), 4.19 (d, J = 7.7 Hz, 1H), 4.12 (d, J = 7.7 Hz, 1H), 4.08 (t, J = 6.8 Hz, 1H), 2.88 (d, J = 6.7 Hz, 2H). ¹³**C NMR** (151 MHz, CD₂Cl₂) δ 147.8, 138.9 129.7, 129.5, 128.9, 126.9, 118.1, 113.9, 83.6, 82.4, 77.4, 59.9, 36.1. **ESI-MS**: Calcd for C₁₇H₁₉NO₂ ([M+H]⁺) 270.1494; found 270.1493.



3-(3-Methyl-1-(phenylamino)butyl)oxetan-3-ol (3d): Prepared from **1d** by General Procedure 1 in 52% yield or by General Procedure 2 in 99% yield as a white solid. **TLC**: R_f 0.35 (2:3 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CDCl₃) δ 7.21 – 7.12 (m, 2H), 6.75 – 6.64 (m, 3H), 4.65 – 4.59 (m, 2H), 4.56 – 4.51 (m, 2H), 3.86 (dd, J = 10.2, 3.1 Hz, 1H), 1.75 – 1.65 (m, 1H), 1.54 – 1.45 (m, 1H), 1.43 – 1.36 (m, 1H), 0.94 (d, J = 6.7 Hz, 3H), 0.91 (d, J = 6.5 Hz, 3H). ¹³C **NMR** (151 MHz, CDCl₃) δ 148.4, 129.6, 118.0, 113.3, 82.6, 81.7, 56.3, 40.2, 24.9, 23.9, 22.2. **ESI-MS**: Calcd for C₁₄H₂₁NO₂ ([M+H]⁺) 236.1651; found 236.1645.



3-(2-(1*H***-indol-3-yl)-1-(phenylamino)ethyl)oxetan-3-ol (3e):** Prepared from 1e by General Procedure 1 in 72% yield or by General Procedure 2 in 68% yield as a light brown solid. TLC: $R_f 0.34$ (3:7 EtOAc/Hexanes). ¹H NMR (600 MHz, CD₂Cl₂) δ 8.00 (s, 1H–NH), 7.59 (d, J = 7.7 Hz, 1H), 7.35 (d, J = 8.3 Hz, 1H), 7.23 – 7.17 (m, 3H), 7.17 – 7.13 (m, 1H), 7.01 – 6.94 (m, 1H), 6.79 – 6.70 (m, 3H), 4.60 (d, J = 7.4 Hz, 1H), 4.47 (d, J = 7.4 Hz, 1H), 4.36 (q, J = 7.4 Hz, 2H), 4.27 (t, J = 6.3 Hz, 1H), 3.24 – 3.04 (m, 2H). ¹³C NMR (151 MHz, CD₂Cl₂) δ 136.4, 129.7, 127.3, 122.6, 120.0, 118.8, 111.8, 111.5, 83.3, 82.1, 24.9. ESI-MS: Calcd for C₁₉H₂₁NO₂ ([M+H]⁺) 309.1603; found 309.1588.



3-(2-Methyl-1-(phenylamino)propyl)oxetan-3-ol (3f): Prepared from **1f** by General Procedure 1 in 28% yield or by General Procedure 2 in 89% yield as a white solid. **TLC**: R_f 0.28 (1:2 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CDCl₃) δ 7.18 – 7.13 (m, 2H), 6.71 – 6.65 (m, 3H), 4.80 (d, J = 7.4, 0.9 Hz, 1H), 4.68 (d, J = 7.6, 0.9 Hz, 1H), 4.53 (d, J = 7.4, 0.9 Hz, 1H), 4.43 (d, J = 7.5, 0.8 Hz, 1H), 3.62 (d, J = 7.1 Hz, 1H), 2.01 – 1.95 (m, 1H), 0.98 (d, J = 6.8 Hz, 3H), 0.93 (d, J = 6.8 Hz, 3H). ¹³**C NMR** (151 MHz, CDCl₃) δ 149.0, 129.8, 117.7, 113.2, 84.6, 83.3, 78.7, 63.2, 31.5, 21.0, 19.9. **ESI-MS**: Calcd for C₁₃H₁₉NO₂ ([M+H]⁺) 222.1494; found 222.1485.



3-(2-Methyl-1-(phenylamino)butyl)oxetan-3-ol (3g): Prepared from **1g** by General Procedure 1 in 36% yield or by General Procedure 2 in 78% yield as a brown oil. **TLC**: R_f 0.39 (2:3 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CDCl₃) δ 7.18 – 7.13 (m, 4H), 6.71 – 6.63 (m, 6H), 4.81 (d, J = 7.4, 0.9 Hz, 1H), 4.75 (d, J = 7.3, 0.8 Hz, 1H), 4.68 (d, J = 7.5, 1.1 Hz, 2H), 4.56 – 4.52 (m, 2H), 4.44 (t, J = 7.5, 6.5, 0.8 Hz, 2H), 3.77 (d, J = 5.1 Hz, 1H), 3.66 (d, J = 8.0 Hz, 1H), 1.79 – 1.73 (m, 1H), 1.73 – 1.63 (m, 2mH), 1.41 – 1.33 (m, 1H), 1.24 – 1.10 (m, 3H), 0.96 (d, J = 6.8 Hz, 4H), 0.94 – 0.84 (m, 11H). ¹³C **NMR** (151 MHz) δ 148.6, 129.6, 118.0, 113.5, 82.9, 81.9, 59.9, 23.8, 11.6. **ESI-MS**: Calcd for C₁₄H₂₁NO₂ ([M+H]⁺) 236.1651; found 236.1653.



3-(1-Phenylpyrrolidin-2-yl)oxetan-3-ol (3h): Prepared from **1h** by General Procedure 1 in 56% yield or by General Procedure 2 in 73% yield a yellow solid. **TLC**: R_f 0.40 (2:3 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CD₂Cl₂) δ 7.22 (t, J = 7.8 Hz, 2H), 6.79 – 6.71 (m, 3H), 4.81 (d, J = 6.9 Hz, 1H), 4.65 (d, J = 7.0 Hz, 1H), 4.43 (d, J = 7.0 Hz, 1H), 4.35 (d, J = 6.9 Hz, 1H), 4.12 – 4.05 (m, 1H), 3.71 – 3.64 (m, 1H), 3.28 – 3.19 (m, 1H), 2.07 – 1.99 (m, 2H), 1.99 – 1.92 (m, 1H), 1.91 – 1.81 (m, 1H). ¹³**C NMR** (151 MHz, CD₂Cl₂) δ 149.6, 129.5, 118.3, 114.6, 83.2, 81.3, 78.8, 63.0, 52.7, 27.9, 24.8. **ESI-MS**: Calcd for C₁₃H₁₇NO₂ ([M+H]⁺) 220.1338; found 220.1333.



3i

3-(1-(Methyl(phenyl)amino)ethyl)oxetan-3-ol (3i): Prepared from **1i** by General Procedure 1 in 39% yield or by General Procedure 2 in 37% yield as a clear oil. **TLC**: R_f 0.31 (2:3 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CD₂Cl₂) δ 7.32 – 7.19 (m, 2H), 6.94 (d, J = 8.0 Hz, 2H), 6.89 – 6.80 (m, 1H), 4.79 (d, J = 6.7 Hz, 1H), 4.70 (d, J = 7.0 Hz, 1H), 4.59 (d, J = 6.7 Hz, 1H), 4.52 (d, J = 6.9 Hz, 1H), 3.89 (q, J = 6.8 Hz, 1H), 2.58 (s, 3H), 1.32 (d, J = 6.8 Hz, 3H). ¹³C NMR (151 MHz, CD₂Cl₂) δ 151.4, 129.46, 119.9, 116.8, 81.8, 81.4, 75.7, 61.0, 32.4, 9.2. **ESI-MS**: Calcd for C₁₂H₁₇NO₂ ([M+H]⁺) 208.1338; found 208.1328.



3-(Indolin-2-yl)oxetan-3-ol (3j): Prepared from **1j** by General Procedure 1 in 43% yield or by General Procedure 2 in 42% yield as a light yellow solid. **TLC**: R_f 0.16 (2:3 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CDCl₃) δ 7.10 (d, J = 7.7 Hz, 1H), 7.05 (t, J = 7.7 Hz, 1H), 6.77 (t, J = 7.0 Hz, 1H), 6.67 (d, J = 7.8 Hz, 1H), 4.75 (d, J = 7.0 Hz, 1H), 4.68 (d, J = 6.5 Hz, 1H), 4.54 (d, J = 6.5, 1.3 Hz, 2H), 4.48 (t, J = 9.5 Hz, 1H), 3.27 – 3.18 (m, 1H), 3.06 – 2.98 (m, 1H). ¹³**C NMR** (151 MHz, CDCl₃) δ 150.4, 128.6, 127.7, 125.0, 120.1, 110.3, 83.0, 80.0, 74.4, 63.9, 30.7. **ESI-MS**: Calcd for C₁₁H₁₃NO₂ ([M+H]⁺) 192.1025; found 192.1027.



3k

3-(2-(Phenylamino)propan-2-yl)oxetan-3-ol (3k): Prepared from **1k** by General Procedure 1 in 22% yield or by General Procedure 2 in 29% yield as a clear oil. **TLC**: $R_f 0.37$ (2:3 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CDCl₃) δ 7.25 – 7.20 (m, 2H), 7.05 – 6.98 (m, 1H), 6.84 – 6.79 (m, 2H), 4.68 (d, J = 7.7 Hz, 2H), 4.63 (d, J = 7.7 Hz, 2H), 1.32 (s, 6H). ¹³**C NMR** (151 MHz, CDCl₃) δ 144.6, 129.1, 122.9, 80.7, 78.5, 22.7. **ESI-MS**: Calcd for C₁₂H₁₇NO₂ ([M+H]⁺) 208.1338; found 208.1347.



3-(3-(Methylthio)-1-(phenylamino)propyl)oxetan-3-ol (3l): Prepared from **1l** by General Procedure 1 in 18% yield or by General Procedure 2 in 90% yield as a light yellow oil. **TLC**: $R_f 0.21$ (3:7 EtOAc/Hexanes). ¹H NMR (600 MHz, CDCl₃) δ 7.20 – 7.13 (m, 2H), 6.75 – 6.66 (m, 3H), 4.64 (d, J = 7.1 Hz, 1H), 4.61 (d, J = 7.2 Hz, 1H), 4.56 (d, J = 7.1 Hz, 1H), 4.52 (d, J = 7.3 Hz, 1H), 4.07 (dd, J = 9.2, 3.6 Hz, 1H), 2.67 – 2.58 (m, 1H), 2.57 – 2.49 (m, 1H), 2.07 (s, 3H), 1.98 – 1.89 (m, 1H), 1.89 – 1.77 (m, 1H). ¹³C NMR (151 MHz, CDCl₃) δ 148.2, 129.7, 118.1, 113.4, 82.9, 82.0, 56.7, 31.2, 29.8, 15.7. ESI-MS: Calcd for C₁₃H₂₀NO₂S ([M+H]⁺) 254.1215; found 254.1207.



tert-Butyl (5-(3-hydroxyoxetan-3-yl)-5-(phenylamino)pentyl)carbamate (3m): Prepared from 1m by General Procedure 1 in 77% yield or by General Procedure 2 in 83% yield as a light yellow solid. TLC: $R_f 0.33$ (3:7 EtOAc/Hexanes). ¹H NMR (600 MHz, CD₂Cl₂) δ 7.20 – 7.08 (m, 2H), 6.73 – 6.57 (m, 3H), 4.60 (d, J = 7.0 Hz, 1H), 4.56 (d, J = 7.1 Hz, 1H), 4.50 – 4.44 (m, 2H), 3.79 (dd, J = 8.4, 5.1 Hz, 1H), 3.23 (s, 1H), 3.16 – 3.05 (m, 1H), 3.05 – 2.93 (m, 1H), 1.76 – 1.66 (m, 1H), 1.59 – 1.50 (m, 1H), 1.48 – 1.43 (m, 2H), 1.41 (s, 9H), 1.33 – 1.19 (m, 1H). ¹³C NMR (151 MHz, CD₂Cl₂) δ 156.7, 148.6, 129.8, 118.0, 113.7, 82.7, 82.0, 79.4, 77.4, 58.5, 39.9, 30.8, 30.1, 28.6, 23.6. **ESI-MS**: Calcd for C₁₉H₃₁N₂O₄ ([M+H]⁺) 351.2284; found 351.2271.



3n

3-(2-(Benzyloxy)-1-(phenylamino)ethyl)oxetan-3-ol (3n): Prepared from **1n** by General Procedure 1 in 37% yield or by General Procedure 2 in 40% yield as a brown oil. **TLC**: R_f 0.34 (3:7 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CD₂Cl₂) δ 7.42 – 7.21 (m, 5H), 7.21 – 7.13 (m, 2H), 6.75 – 6.63 (m, 3H), 4.61 – 4.47 (m, 6H), 4.05 – 4.01 (m, 1H), 3.95 – 3.89 (m, 1H, N–H), 3.81 (dd, J = 9.8, 2.8 Hz, 1H), 3.77 (dd, J = 9.8, 4.1 Hz, 1H), 1.21 (d, J = 6.5 Hz, 1H, O–H). ¹³**C NMR** (151 MHz, CD₂Cl₂) δ 147.4, 137.7, 129.8, 129.7, 128.9, 128.4, 128.1, 118.2, 114.0, 82.2, 77.3, 74.1, 68.6, 56.3, 13.7. **ESI-MS**: Calcd for C₁₈H₂₂NO₃ ([M+H]⁺) 300.1600; found 300.1603.



3-(2-(*tert***-Butoxy)-1-(phenylamino)ethyl)oxetan-3-ol (30):** Prepared from 10 by General Procedure 1 in 44% yield or by General Procedure 2 in 30% yield as a brown oil. TLC: R_f 0.40 (2:3 EtOAc/Hexanes). ¹H NMR (600 MHz, CDCl₃) δ 7.19 (dd, J = 259.4, 7.6 Hz, 2H), 6.73 (d, J = 7.1 Hz, 1H), 6.66 (d, J = 8.3 Hz, 2H), 4.68 (d, J = 6.8 Hz, 1H), 4.62 – 4.58 (m, 2H), 4.58 – 4.55 (m, 2H), 4.31 (s, 1H, O–H), 4.01 (s, 1H, N–H), 3.77 – 3.68 (m, 2H), 1.13 (s, 9H).¹³C NMR (151 MHz, CDCl₃) δ 146.8, 129.6, 118.1, 113.9, 82.3, 82.2, 74.9, 60.1, 55.7, 27.4. ESI-MS: Calcd for C₁₅H₂₄NO₃ ([M+H]⁺) 266.1756; found 266.1762.



3-(2-(4-(*tert***-Butoxy)phenyl)-1-(phenylamino)ethyl)oxetan-3-ol (3p):** Prepared from 1p by General Procedure 1 in 70% yield or by General Procedure 2 in 77% yield as a brown solid. TLC: $R_f 0.18$ (2:3 EtOAc/Hexanes). ¹H NMR (600 MHz, CDCl₃) δ 7.20 – 7.13 (m, 2H), 7.13 – 7.09 (m, 2H), 6.93 – 6.85 (m, 2H), 6.74 – 6.66 (m, 1H), 6.66 – 6.60 (m, 2H), 4.62 (d, J = 7.5 Hz, 1H), 4.47 (d, J = 7.5 Hz, 1H), 4.22 (d, J = 7.7 Hz, 1H), 4.13 (d, J = 7.7 Hz, 1H), 4.10 – 4.04 (m, 1H), 2.95 – 2.82 (m, 2H), 1.30 (s, 9H). ¹³C NMR (151 MHz, CDCl₃) δ 154.1, 147.3, 133.3, 129.7, 129.6, 124.7, 118.1, 113.8, 83.9, 82.4, 78.6, 59.8, 35.3, 28.9. ESI-MS: Calcd for C₂₁H₂₈NO₃ ([M+H]⁺) 342.2069; found 342.2068.



tert-Butyl 3-(3-hydroxyoxetan-3-yl)-3-(phenylamino)propanoate (3q): Prepared from 1q by General Procedure 1 in 41% yield or by General Procedure 2 in 83% yield as a yellow solid. TLC: $R_f 0.33$ (3:7 EtOAc/Hexanes). ¹H NMR (600 MHz, CDCl₃) δ 7.23 – 7.14 (m, 2H), 6.79 – 6.71 (m, 1H), 6.71 – 6.65 (m, 2H), 4.63 – 4.56 (m, 4H), 4.27 (d, J = 5.8 Hz, 1H), 2.72 (dd, J = 15.4, 6.1 Hz, 1H), 2.54 (dd, J = 15.4, 5.0 Hz, 1H), 1.36 (s, 9H). ¹³C NMR (151 MHz, CDCl₃) δ 172.4, 146.9, 129.6, 118.5, 113.8, 82.5, 82.2, 82.0, 76.7, 55.0, 36.0, 28.0. ESI-MS: Calcd for C₁₆H₂₄NO₄ ([M+H]⁺) 294.1705; found 294.1705.



Benzyl 4-(3-hydroxyoxetan-3-yl)-4-(phenylamino)butanoate (3r): Prepared from 1r by General Procedure 1 in 35% yield or by General Procedure 2 in 49% yield as a light yellow oil. **TLC**: $R_f 0.19$ (2:3 EtOAc/Hexanes). ¹H NMR (600 MHz, CD₂Cl₂) δ 7.43 – 7.22 (m, 5H), 7.19 – 7.05 (m, 2H), 6.76 – 6.54 (m, 3H), 5.07 (q, J = 12.5 Hz, 2H), 4.60 (d, J = 7.2 Hz, 1H), 4.53 (d, J = 7.3 Hz, 1H), 4.48 (d, J = 7.2 Hz, 1H), 4.45 (d, J = 7.1 Hz, 1H), 3.91 (dd, J = 9.4, 3.9 Hz, 1H), 2.56 – 2.39 (m, 2H), 2.05 – 1.92 (m, 1H), 1.88 – 1.77 (m, 1H). ¹³C NMR (151 MHz, CD₂Cl₂) δ 173.9, 148.4, 136.4, 129.8, 128.9, 128.7, 128.6, 118.2, 113.6, 82.7, 82.0, 77.3, 66.8, 57.7, 31.2, 25.3. ESI-MS: Calcd for C₂₀H₂₄NO₄ ([M+H]⁺) 342.1705; found 342.1712.



3-(3-Hydroxyoxetan-3-yl)-3-(phenylamino)-*N*-tritylpropanamide (3s): Prepared from 1s by General Procedure 1 in 37% yield or by General Procedure 2 in 76% yield as a white solid. TLC: $R_f 0.27$ (3:2 EtOAc/Hexanes). ¹H NMR (600 MHz, CD₃CN) δ 7.34 (s, 1H, N–H), 7.08 – 6.87 (m, 17H), 6.56 (d, J = 8.0 Hz, 2H), 6.48 (t, J = 7.6 Hz, 1H), 4.63 (s, 1H, O–H), 4.33 (d, J = 9.7 Hz, 1H), 4.23 (d, J = 6.7 Hz, 1H), 4.19 (d, J = 6.6 Hz, 1H), 4.17 – 4.13 (m, 2H), 4.10 – 4.04 (m, 1H), 2.47 (dd, J = 15.1, 5.0 Hz, 1H), 2.30 (dd, J = 15.1, 6.5 Hz, 1H). ¹³C NMR (151 MHz, CD₃CN) δ 171.9, 148.9, 145.5, 130.2, 129.6, 128.6, 127.6, 114.0, 82.4, 82.2, 77.4, 71.0, 55.5, 55.5, 37.5. ESI-MS: Calcd for C₃₁H₃₁N₂O₃ ([M+H]⁺) 479.12335; found 479.2352.



3-(3-Hydroxyoxetan-3-yl)-3-(phenylamino)propanoic acid **(3t):** Prepared from **1t** by General Procedure 1 in 34% yield or by General Procedure 2 in 26% yield as an off-white solid. **TLC**: $R_f 0.22$ (3:2 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CDCl₃) δ 7.21 (t, J = 8.2 Hz, 2H), 6.78 (t, J = 7.7 Hz, 1H), 6.71 (d, J = 8.2 Hz, 2H), 4.66 (t, J = 9.1 Hz, 1H), 3.92 (q, J = 12.5 Hz, 2H), 3.76 (d, J = 12.4 Hz, 1H), 3.64 (d, J = 12.5 Hz, 1H), 3.05 (dd, J = 17.9, 9.2 Hz, 1H), 2.70 (dd, J = 17.9, 8.7 Hz, 1H). ¹³**C NMR** (151 MHz, CDCl₃) δ 175.1, 146.7, 129.8, 119.1, 113.7, 90.2, 63.8, 63.3, 52.0, 37.6. **ESI-MS**: Calcd for C₁₂H₁₆NO₄ ([M+H]⁺) 238.1079; found 238.1076.



3-((*p***-Tolylamino)methyl)oxetan-3-ol (3u):** Prepared from **1u** by General Procedure 1 in 80% yield or by General Procedure 2 in 52% yield as a light brown oil. **TLC**: $R_f 0.17$ (2:3 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CDCl₃) δ 7.03 (d, J = 8.4 Hz, 2H), 6.66 (d, J = 8.4 Hz, 2H), 4.66 (d, J = 7.0 Hz, 2H), 4.57 (d, J = 8.0 Hz, 2H), 3.53 (s, 2H), 2.26 (s, 3H). ¹³**C NMR** (151 MHz, CDCl₃) δ 145.5, 130.1, 128.4, 114.1, 82.4, 73.6, 51.2, 20.6. **ESI-MS**: Calcd for C₁₁H₁₅NO₂ ([M+H]⁺) 194.1181; found 194.1181.



3-(((4-Bromophenyl)amino)methyl)oxetan-3-ol (3v): Prepared from **1v** by General Procedure 1 in 47% yield or by General Procedure 2 in 64% yield as a light yellow solid. **TLC**: R_f 0.27 (2:3 EtOAc/Hexanes).¹H NMR (600 MHz, CD₂Cl₂) δ 7.33 – 7.21 (m, 2H), 6.64 – 6.55 (m, 2H), 4.59 (d, J = 7.3 Hz, 2H), 4.54 (d, J = 7.6 Hz, 2H), 3.48 (s, 2H). ¹³C **NMR** (151 MHz, CD₂Cl₂) δ 147.9, 132.3, 115.3, 109.8, 82.5, 74.0, 50.5. **ESI-MS**: Calcd for C₁₀H₁₃NO₂Br ([M+H]⁺) 258.0130; found 258.0132.



3-(((4-Fluorophenyl)amino)methyl)oxetan-3-ol (3w): Prepared from 1k by General Procedure 1 in 53% yield or by General Procedure 2 in 75% yield as a clear oil. TLC: $R_f 0.25$ (2:3

EtOAc/Hexanes). ¹H NMR (600 MHz, CDCl₃) δ 6.91 (t, J = 8.7 Hz, 2H), 6.69 – 6.61 (m, 2H), 4.65 (d, J = 7.0 Hz, 2H), 4.58 (d, J = 6.4 Hz, 2H), 3.49 (s, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 157.4, 155.8, 144.3 (d, J = 2.0 Hz), 116.0 (d, J = 22.4 Hz), 114.8 (d, J = 7.5 Hz), 82.5, 73.7, 51.3. ESI-MS: Calcd for C₁₀H₁₂FNO₂ ([M+H]⁺) 198.1930; found 198.0925.



3-(((4-(Trifluoromethyl)phenyl)amino)methyl)oxetan-3-ol (3l): Prepared from **1l** by General Procedure 1 in 30% yield or by General Procedure 2 in 40% yield as a light brown oil. **TLC:** R_f 0.24 (1:1 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CDCl₃) δ 7.47 (d, J = 8.3 Hz, 2H), 6.84 (d, J = 8.4 Hz, 2H), 4.66 (d, J = 7.4 Hz, 2H), 4.58 (d, J = 7.6 Hz, 2H), 3.61 (s, 2H). ¹³**C NMR** (151 MHz, CDCl₃) δ 126.8, 113.6, 82.2, 50.4. **ESI-MS**: Calcd for C₁₁H₁₁F₃NO₂ ([M–H]⁻) 246.0742; found 246.0734.



Зy

3,3'-((Phenylazanediyl)bis(methylene))bis(oxetan-3-ol) (3y): Prepared from **1y** by General Procedure 1 in 10% yield as a clear oil. **TLC**: R_f 0.12 (1:1 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CD₂Cl₂) δ 7.34 – 7.26 (m, 2H), 7.20 – 7.11 (m, 2H), 7.00 (tt, J = 7.3, 1.1 Hz, 1H), 4.43 (d, J = 7.6 Hz, 4H), 4.36 (d, J = 7.7 Hz, 4H), 3.68 (s, 4H). ¹³C **NMR** (151 MHz, CD₂Cl₂) δ 151.2, 129.9, 122.9, 120.6, 82.9, 74.7, 61.7. **ESI-MS**: Calcd for C₁₄H₂₀NO₄ ([M+H]⁺) 266.1392; found 266.1398.



3-((Methyl(phenyl)amino)methyl)oxetan-3-ol (3z): Prepared from **1y** by General Procedure 1 in 44% yield or by General Procedure 2 in 40% yield as a clear oil. **TLC**: R_f 0.28 (2:3 EtOAc/Hexanes). ¹**H NMR** (600 MHz, CD₂Cl₂) δ 7.29 – 7.21 (m, 2H), 6.91 (d, J = 8.1 Hz, 2H), 6.82 (t, J = 7.3 Hz, 1H), 4.66 – 4.57 (m, 4H), 3.62 (s, 2H), 2.88 (s, 3H). ¹³C **NMR** (151 MHz, CD₂Cl₂) δ 151.6, 129.5, 119.2, 115.1, 83.2, 74.2, 60.5, 39.9. **ESI-MS**: Calcd for C₁₁H₁₅NO₂ ([M+H]⁺) 194.1181; found 194.1183.

E. CONVERSION OF N,N-DIALKYL-α-AMINO ACIDS (4) TO 3-OXETANOLS (5)



General Procedure 3 for decarboxylative coupling of N-aliphatic α -amino acids

In a 4-mL glass vial, 4CzIPN (3.155 mg, 4 μ mol, 2 mol%), cesium pivalate (56 mg, 0.24 mmol, 1.2 equiv), and the appropriate α -amino acid **5** (0.2 mmol, 1 equiv) were combined. 2-Propanol (0.5 mL) was added followed by 3-oxetanone (**2**) (12.8 μ L, 0.2 mmol, 1 equiv). The mixture was degassed with Ar for 1 min. The reaction was exposed to blue LED light (456 nm) at 25 °C for 20 h. The reaction mixture was filtered over celite and concentrated by rotary evaporation. Purification by silica flash chromatography yielded the 3-oxetanol product **5**.



5a

3-(morpholinomethyl)oxetan-3-ol (5a): Prepared from **4a** by General Procedure 3 (modification of 3 equiv 3-oxetanone used) in 30% yield as a light brown oil. **TLC**: R_f 0.16 (1:4 Methanol/DCM). ¹**H NMR** (600 MHz, CDCl₃) δ 4.74 (d, J = 6.8 Hz, 2H), 4.49 (d, J = 7.3 Hz, 2H), 3.81 – 3.71 (m, 4H), 2.90 (s, 2H), 2.58 (s, 4H). ¹³C NMR (151 MHz, CDCl₃) δ 83.61, 71.00, 66.46, 64.31, 53.53. **ESI-MS**: Calcd for C₈H₁₆NO₃ ([M+H]⁺) 174.1130; found 174.1130.



5b

3-((4-methylpiperazin-1-yl)methyl)oxetan-3-ol (5b): Prepared from **4b** by General Procedure 3 in 32% yield as a light brown oil. **TLC**: R_f 0.17 (1:4 Methanol/DCM). ¹**H NMR** (500 MHz, CDCl₃) δ 4.73 (d, J = 6.7 Hz, 2H), 4.49 (d, J = 6.7 Hz, 2H), 2.81 (s, 2H), 2.61 – 2.37 (m, 8H), 2.31 (s, 3H). ¹³C **NMR** (151 MHz, CDCl₃) δ 83.92, 71.00, 63.89, 54.92, 52.85, 45.96. **ESI-MS**: Calcd for C₉H₁₉N₂O₂ ([M+H]⁺) 187.1447; found 187.1445.



5c

tert-butyl 4-((3-hydroxyoxetan-3-yl)methyl)piperazine-1-carboxylate (5c): Prepared from 4c by General Procedure 3 in 21% yield as a light brown oil. TLC: $R_f 0.17$ (3:2 EtOAc/Hexanes with 1% triethyl amine). ¹H NMR (600 MHz, CDCl₃) δ 4.67 (d, J = 7.3 Hz, 2H), 4.43 (d, J = 7.4 Hz,

2H), 3.37 (t, J = 4.8 Hz, 4H), 2.74 (s, 2H), 2.34 (d, J = 5.0 Hz, 4H), 1.39 (s, 9H). ¹³C NMR (151 MHz, CDCl₃) δ 154.70, 83.87, 80.12, 71.04, 64.09, 52.91, 28.53. **ESI-MS**: Calcd for C₁₃H₂₅N₂O₄ ([M+H]⁺) 273.1826; found 273.1814.



5d

3-(piperidin-1-ylmethyl)oxetan-3-ol (5d): Prepared from **4d** by General Procedure 3 in 29% yield as a light brown oil. **TLC**: $R_f 0.14$ (1:4 Methanol/DCM). ¹**H NMR** (600 MHz, CD₂Cl₂) δ 4.62 (d, J = 6.0 Hz, 2H), 4.45 (d, J = 1.0 Hz, 2H), 2.89 (s, 2H), 2.68 – 2.34 (m, 4H), 1.65 (p, J = 5.7 Hz, 4H), 1.55 – 1.40 (m, 2H). ¹³C NMR (151 MHz, CD₂Cl₂) δ 83.83, 71.30, 64.39, 54.83, 25.28, 23.70. **ESI-MS**: Calcd for C₉H₁₈NO₂ ([M+H]⁺) 172.1338; found 172.1333.



3-(1-(dipropylamino)ethyl)oxetan-3-ol (5e): Prepared from **4e** by General Procedure 3 in 40% yield as a clear oil. **TLC**: R_f 0.14 (1:4 Methanol/DCM). ¹**H NMR** (500 MHz, CD₂Cl₂) δ 4.72 (d, J = 6.6 Hz, 1H), 4.67 (d, J = 7.0 Hz, 1H), 4.50 (d, J = 6.6 Hz, 1H), 4.42 (d, J = 6.9 Hz, 1H), 2.81 (q, J = 6.9 Hz, 1H), 2.40 – 2.30 (m, 2H), 2.27 – 2.15 (m, 2H), 1.47 (dtt, J = 15.5, 8.0, 2.3 Hz, 2H), 1.43 – 1.35 (m, 2H), 1.28 (d, J = 6.9 Hz, 3H), 0.86 (t, J = 7.4 Hz, 6H). ¹³C NMR (126 MHz, CD₂Cl₂) δ 82.26, 81.01, 74.24, 59.67, 52.39, 21.65, 11.90, 6.69. **ESI-MS**: Calcd for C₁₁H₂₄NO₂ ([M+H]⁺) 202.1807; found 202.1808.

F. MECHANISTIC STUDIES

1. DEUTERIUM QUENCHING AND COMPETITION EXPERIMENTS

General Procedure 1 (SECTION D) was followed for entries 1–3 with the minor modifications of using deuterium exchanged *N*-phenyl glycine with CH₃OD (entries 2,3), excluding 3-oxetanone and adding 327 μ L of CH₃OD (entry 2), or adding 327 μ L CH₃OD (entry 3). General Procedure 2 (SECTION D) was followed for entries 4–6 with the minor modifications of using deuterium exchanged *N*-phenyl glycine with CH₃OD (entries 2,3), excluding 3-oxetanone and adding 327 μ L of CH₃OD (entry 2), or adding 327 μ L CH₃OD (entry 3). Mesitylene and adding 327 μ L of CH₃OD (entry 2), or adding 327 μ L CH₃OD (entry 3). Mesitylene (27.8 μ L, 0.2 mmol, 0.5 equiv) was added as an internal standard for ¹H-NMR analysis.

The following control experiment was also carried out to exclude formation of 4b by H/D-exchange between 4a and CH₃OD in the presence of CsOPiv.



2. DIFFERENTIAL PULSE VOLTAMMETRY (DPV) MEASUREMENTS OF 3-OXETANONE

DPV measurements were performed using a 660E potentiostat/galvanostat model from CH Instruments. A standard three-electrode configuration was used for these experiments, which were a glassy carbon working electrode, a platinum wire counter electrode, and a Ag/AgCl reference electrode. The supporting electrolyte used was tetrabutylammonium tetrafluoroborate (TBAPF₆) dissolved in acetonitrile (0.1 M) and analyte concentration was 20 mM. Standard reduction potentials were obtained against Ag/AgCl and converted against standard calomel electrode (SCE). The 3-oxetanone wave was confirmed using Fc/Fc^+ as an internal standard.



Supplementary Figure S3. Differential pulse voltammetry of 3-oxetanone and 3-oxetanone with 1 equiv. pivalic acid.

3. STERN–VOLMER FLUORESCENCE QUENCHING ANALYSIS

Photocatalyst fluorescence quenching measurements were obtained on a fluorimeter (Fluoromax-4, Horiba Jobin-Yvon). For emission quenching experiments, a 45 μ M solution of 4CzIPN in acetonitrile was added to 3.5 mL quartz cuvettes (10 mm) with varying concentrations of quencher added. The cuvettes were equipped PTFE/Septa screw caps and degassed by argon sparging. The samples were irradiated at 435 nm and fluorescence emission was measured at 535 nm. A Stern–Volmer plot was created by plotting the ratio of emission intensity (I_0 /I) in the absence and presence of individual quenchers at different concentrations.

 I_0 = emission intensity of photocatalyst in the absence of quencher at a given wavelength I = observed emission intensity of photocatalyst with quencher



Supplementary Figure S4. Stern–Volmer plot of 4CzIPN (45 μ M) in acetonitrile with varying concentrations of different quenchers.

This data indicates that N-phenyl glycine (cesium salt), cesium pivalate, 3-oxetanone, and Cr(III)Cl₃, and TMSCl quench the excited state photocatalyst, however, the Stern–Volmer constant of N-phenyl glycine (cesium salt) is much larger than that of the other reagents.



Supplementary Figure S5. Stern–Volmer plot of 4CzIPN (45 μ M) in methylene chloride with varying concentrations of different quenchers.

This data indicates that *N*-phenyl glycine (cesium salt), cesium pivalate, and 3-oxetanone quench the excited state photocatalyst, however, the Stern–Volmer constant of *N*-phenyl glycine (cesium salt) is much larger than that of the other reagents.

4. QUANTUM YIELD DETERMINATION

1.

2.

3.

The quantum yield of the reaction was determined using a published protocol.⁶

Determination of photon flux

The photon flux of the LED (Kessil PR160L, 40 W, $\lambda_{max} = 456$ nm) was measured via standard ferrioxalate actinometry.^{7,8} A solution of ferrioxalate (0.15 M) was prepared by dissolving 2.21 g of potassium ferrioxalate hydrate in 30 mL of 0.05 M H₂SO₄. A buffered solution of 1,10-phenanthroline was prepared by dissolving 25 mg of phenanthroline and 5.63 g of sodium acetate in 25 mL of 0.5 M H₂SO₄. Solutions were stored in the dark. 3 mL of the ferrioxalate solution was added to 4 mL vials and irradiated for 60 seconds. After irradiation, 0.525 mL of the phenanthroline solution was added and the sample was allowed to rest for 1 hour for coordination. Next, the mixture was transferred to a quartz cuvette and the absorbance was measured at 510 nm. Non-irradiated samples as controls were also prepared. Photoconversion of Fe³⁺ to Fe²⁺ was calculated using eq 1.

$$mol \ Fe^{2+} = \frac{V * \Delta A}{l * \varepsilon}$$

V is the total volume (0.003525 L), ΔA is the difference in absorbance at 510 nm between the irradiated and non-irradiated samples, *l* is the path length (1 cm), and ε is the molar absorptivity at 510 nm (11,100 L mol⁻¹ cm⁻¹). After the *mol Fe*²⁺ was calculated from the equation 1, the photon flux was determined using eq 2.

$$photon flux = \frac{mol Fe^{2+}}{\Phi * t * f}$$

 Φ is the quantum yield for the ferrioxalate actinometer (0.846 for 0.15 M at $\lambda = 457.9$ nm),⁹ t is the time (60 seconds), and f is the fraction of light absorbed by the ferrioxalate actinometer at $\lambda = 456$ nm (0.964). The fraction of light absorbed is calculated using eq 3.

$$f = 1 - 10^{-A(at \, 456 \, nm)}$$

The average photon flux from 3 experiments was determined to be 1.79×10^{-8} einsteins per second.

Determination of quantum yield

Conditions 1 (Cr-mediated)



In a 4-mL glass vial, 4CzIPN (1.58 mg, 2 µmol, 1 mol%), CrCl₃ (1.59 mg, 10 µmol, 5 mol%), cesium pivalate (56 mg, 0.24 mmol, 1.2 equiv), and the appropriate *N*-aryl α -amino acid 1 (0.2 mmol, 1 equiv) were combined. CH₃CN (0.25 mL) was added followed by 3-oxetanone (2) (12.8 µL, 0.2 mmol, 1 equiv). The mixture was degassed with Ar for 1 min, then TMSCl (12.5 µL, 0.1 mmol, 0.5 equiv) was added. The reaction was exposed to blue LED light (456 nm) at 25 °C for 3 minutes. After irradiation, the yield was determined using ¹H-NMR analysis of crude reaction product in the presence of mesitylene as an internal standard, relative to limiting reagent 3-oxetanone. The yield obtained was 8% (1.6 x 10⁻⁵ mol of product). The quantum yield of the reaction was calculated using eq 4, where photon flux is 1.79 x 10⁻⁸ einsteins per second, *t* is time (180 seconds), and *f* is the fraction of light absorbed by the reaction mixture in the conditions described (0.95).

4.

quantum yield =
$$\frac{mol \text{ product}}{photon flux * t * f}$$

The quantum yield (Φ) was determined to be 5.2.

Conditions 1 (Cr-mediated) in the absence of TMSCl



In a 4-mL glass vial, 4CzIPN (1.58 mg, 2 µmol, 1 mol%), CrCl₃ (1.59 mg, 10 µmol, 5 mol%), cesium pivalate (56 mg, 0.24 mmol, 1.2 equiv), and the appropriate *N*-aryl α -amino acid **1** (0.2 mmol, 1 equiv) were combined. CH₃CN (0.25 mL) was added followed by 3-oxetanone (**2**) (12.8 µL, 0.2 mmol, 1 equiv). The mixture was degassed with Ar for 1 min. The reaction was exposed to blue LED light (456 nm) at 25 °C for 5 minutes. After irradiation, the yield was determined using ¹H-NMR analysis of crude reaction product in the presence of mesitylene as an internal standard, relative to limiting reagent 3-oxetanone. The yield obtained was 4% (0.8 x 10⁻⁵ mol of product). The quantum yield of the reaction was calculated using eq 4, where photon flux is 1.79 x 10⁻⁸ einsteins per second, *t* is time (300 seconds), and *f* is the fraction of light absorbed by the reaction mixture in the conditions described (0.95).

The quantum yield (Φ) was determined to be 1.6.

Conditions 2 (Cr-free)



In a 4-mL glass vial, 4CzIPN (1.58 mg, 2 µmol, 1 mol%), cesium pivalate (56 mg, 0.24 mmol, 1.2 equiv), and the appropriate *N*-aryl α -amino acid 1 (0.2 mmol, 1 equiv) were combined. Methylene chloride (0.4 mL) was added followed by 3-oxetanone (2) (38.4 µL, 0.6 mmol, 3 equiv). The mixture was degassed with Ar for 1 min. The reaction was exposed to blue LED light (456 nm) at 25 °C for 3 minutes. After irradiation, the yield was determined using ¹H-NMR analysis of crude reaction product in the presence of mesitylene as an internal standard, relative to limiting reagent 3-oxetanone. The yield obtained was 16% (3.2 x 10⁻⁵ mol of product). The quantum yield of the reaction was calculated using eq 4, where photon flux is 1.79 x 10⁻⁸ einsteins per second, *t* is time (180 seconds), and *f* is the fraction of light absorbed by the reaction mixture in the conditions described (0.96).

The quantum yield (Φ) was determined to be 10.3.

G. SUPPORTING INFORMATION REFERENCES

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H.¹H-NMR AND ¹³C-NMR SPECTRA

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ò f1 (ppm)



































100 90 f1 (ppm) θ



