Supporting Information

Photoredox catalyzed stereo- and regioselective vicinal fluorosulfonyl-borylation of unsaturated hydrocarbons

Heyin Li,[†] Mengjun Huang,[†] Zhenlei Zou,[†] Zhen Wang,[†] Yifan Li,[†] Chao Sun,[†] Wangzhe Chen,[†] Yi Pan,[†] Weigang Zhang,^{*,†} and Yi Wang ^{*,†}

[†]State Key Laboratory of Coordination Chemistry, Jiangsu Key Laboratory of Advanced Organic Materials, School of Chemistry & Chemical Engineering, Nanjing University, 163 Xianlin Avenue, Nanjing, 210023, China.

*E-mail: wgzhang@nju.edu.cn; yiwang@nju.edu.cn.

Table of Contents

I. General Methods	S1
II. Synthesis of Starting Materials	S2
III. Synthesis of Sulfonyl fluoride imidazolium salt (2a-2d)	S2
IV. Cyclic Voltammetry Studies for 2a-2d	S4
V. Optimizations of the Reaction Conditions	S7
VI. General Procedure for the Synthesis of 4, 6-9, 11, 13-15, 17-18, 20-21	S16
VII. Mechanistic experiments	S20
VIII. References	S26
IX. Characteristic Data	S27
X. NMR Spectra of 4, 6-9, 11, 13-15, 17-18, 20-21	S46

Supplementary Methods

I. General Methods

All reactions were performed in flame-dried glassware with magnetic stirring bar and sealed with a rubber septum. The solvents were distilled by standard methods. Reagents were obtained from commercial suppliers and used without further purification unless otherwise noted. Silica gel column chromatography was carried out using silica Gel 60 (230-400 mesh). Analytical thin layer chromatography (TLC) was done using silica Gel (silica gel 60 F254). TLC plates were analyzed by an exposure to ultraviolet (UV) light and/or submersion in phosphomolybdic acid solution or submersion in KMnO₄ solution or in I₂. NMR experiments were measured on a Bruker AVANCE III-400 or 500 spectrometer and carried out in chloroformd (CDCl₃) or acetonitrile-d₃ (CD₃CN). ¹H NMR and ¹³C NMR spectra were recorded at 400 MHz or 500 MHz and 100 MHz or 125 MHz spectrometers, respectively.19F NMR spectra were recorded at 376 MHz or 470 MHz spectrometers. Chemical shifts are reported as δ values relative to internal TMS (δ 0.00 for ¹H NMR), chloroform (δ 7.26 for ¹H NMR), acetonitrile (δ 1.94 for ¹H NMR), chloroform (δ 77.00 for ¹³C NMR), and acetonitrile (δ 1.32 or 118.26 for ¹³C NMR) in parts per million (ppm). The following abbreviations are used for the multiplicities: s: singlet, d: doublet, dd: doublet of doublet, t: triplet, q: quadruplet, m: multiplet, br: broad signal for proton spectra; Coupling constants (J) are reported in Hertz (Hz). Melting points were uncorrected. Infrared spectra were obtained on agilent Cary630. HRMS were recorded on a Bruker miccOTOF-Q111. GC-MS spectra were performed on Agilent 5977B.

Medium-sized screw-cap test tubes (8 mL) were used for all 0.10 mmol scale reactions: Fisher 13 x 100 mm tubes (Cat. No.1495935C)



Supplementary Figure 1. Fisher 13 x 100 mm tubes

Cap with Septa: Thermo Scientific ASM PHN CAP w/PTFE/SIL (Cat. No.03378316)



Supplementary Figure 2. Cap with Septa

II. Synthesis of Starting Materials

Substrates **1a-1e**, **1m-1x**, **5a**, **5f-5m** were purchased from commercial sources (Alfa, TCI, Energy, Bide and Macklin) and used as received.

Substrates 1f-1l, 5n were prepared according to the literature.^[1]

Substrates 5b-5e were prepared according to the literature.^[2]

III. Synthesis of Sulfonyl fluoride imidazolium salt (2a-2d)^[3]



General Procedure:

1) Sodium hydride (60% dispersion in mineral oil.) (36 mmol, 1.2 equiv.) was added to corresponding imidazole (30 mmol, 1 equiv.) in N,N-Dimethylformamide (100 mL). The mixture was stirred at room temperature for 1 hour; A balloon volume of sulfuryl fluoride gas was then added to the reaction system. After the reaction was completed by TLC monitoring, the reaction mixture was evaporated in *vacuo*. Then, the reaction mixture was quenched with water and extracted with ethyl acetate (60 mL x 3). The organic layer was dried over Na₂SO₄, and evaporated in vacuo. The product was purified by flash column chromatography on silica gel with n-pentane/ethyl acetate as eluent to give the corresponding intermediate A.

2) To a solution of the corresponding intermediate A in DCM (50 mL) was added dropwise MeOTf (45 mmol) at 0 °C. Then, the mixture was stirred at room temperature for 12 hours, while monitoring by TLC. After that time, the mixture was concentrated under rotary evaporation to give a white solid (or a viscous liquid) crude product, to which *tert*-butyl methyl ether (30 mL) was added. With vigorous stirring, a solid precipitate was formed. The precipitate was washed with *tert*-butyl methyl ether (30 mL × 3) and dried in *vacuo* to yield the title compound (**2a-2d**) as a white solid.

1-(fluorosulfonyl)-3-methyl-2-(4-(trifluoromethyl)phenyl)-1H-benzo[d]imidazol-3-ium trifluoromethanesulfonate (2a)



69%; white solid: m.p. 165-166 °C; ¹H NMR (400 MHz, Acetonitrile-*d*₃) δ 8.24 – 8.20 (m, 1H), 8.17 – 8.11 (m, 1H), 8.09 (d, J = 0.9 Hz, 4H), 8.03 – 7.95 (m, 2H), 3.96 (s, 3H). ¹³C NMR (101 MHz, Acetonitrile-*d*₃) δ 152.5, 136.1 (q, J = 33.1 Hz), 132.8, 132.6, 132.0, 130.7, 130.6, 127.6 (q, J = 3.8 Hz), 125.9, 124.8, 123.1, 122.0 (q, J = 320.8 Hz), 116.0, 115.9, 35.6. ¹⁹F NMR (376 MHz, Acetonitrile- d_3) δ 64.62, -63.88, -79.31.; HRMS(ESI): called for C₁₅H₁₁F₄N₂O₂S⁺ [M]⁺ 359.0472; found 359.0471.

1-(fluorosulfonyl)-3-methyl-2-phenyl-1H-benzo[d]imidazol-3-ium trifluoromethanesulfonate (2b)

 $\begin{array}{l} \tilde{\textbf{OTf}}_{\textbf{N}} \quad \textbf{Me} \\ \tilde{\textbf{N}}_{\textbf{N}} \quad \textbf{N}_{\textbf{N}} \quad \textbf{N} \quad \textbf{N}_{\textbf{N}} \quad \textbf{N} \quad \textbf{N} \\ \textbf{N}_{\textbf{N}} \quad \textbf{N}_{\textbf{N}} \quad \textbf{N} \quad$

35.5. ¹⁹F NMR (376 MHz, Acetonitrile-*d*₃) δ 64.76, -79.23. HRMS(ESI): caled for C₁₄H₁₂FN₂O₂S⁺ [M]⁺ 291.0598; found 291.0596.

2-(4-fluorophenyl)-1-(fluorosulfonyl)-3-methyl-1H-benzo[d]imidazol-3iumtrifluoromethanesulfonate (2c)



60%; white solid: m.p. 148-149 °C; ¹H NMR (400 MHz, Acetonitrile- d_3) δ 8.24 – 8.16 (m, 1H), 8.14 – 8.04 (m, 1H), 8.01 – 7.95 (m, 2H), 7.94 – 7.89 (m, 2H), 7.59 – 7.48 (m, 2H), 3.95 (s, 3H). ¹³C NMR (101 MHz, Acetonitrile- d_3) δ 168.4, 165.8, 153.4, 134.6 (d, J = 9.8 Hz), 132.7, 131.8, 130.6, 122.0 (q, J = 320.7 Hz), 118.1, 116.9 (d, J = 3.4 Hz), 116.0, 115.9, 35.49.

 ^{19}F NMR (376 MHz, CDCl₃) δ 64.65, -79.30, -104.18– -104.25 (m). HRMS(ESI): caled for $C_{14}H_{11}F_2N_2O_2S^+$ [M]+ 309.0504; found 309.0505.

1-(fluorosulfonyl)-2-(4-methoxyphenyl)-3-methyl-1H-benzo[d]imidazol-3iumtrifluoromethanesulfonate (2d)



43%; white solid: m.p. 162-163 °C; ¹H NMR (400 MHz, Acetonitrile- d_3) δ 8.23 – 8.13 (m, 1H), 8.12 – 8.00 (m, 1H), 7.99 – 7.89 (m, 2H), 7.85 – 7.74 (m, 2H), 7.33 – 7.25 (m, 2H), 3.96 (s, 3H), 3.95 (s, 3H). ¹³C NMR (101 MHz, Acetonitrile- d_3) δ 165.4, 154.7, 133.7, 132.7, 131.4, 130.6, 130.3, 122.1 (q, *J* = 320.9 Hz), 116.1, 116.0, 115.7, 111.9,

56.8, 35.4. ¹⁹F NMR (376 MHz, Acetonitrile- d_3) δ 64.72, -79.28. HRMS(ESI): caled for C₁₅H₁₄FN₂O₃S⁺ [M]⁺ 321.0704; found 321.0704.

Synthesis of 3,4-Diethyl-3,4-hexanediol^[4]



According to the method previously reported by Roberto Sanz et al^[4], in an oven dried Schlenck flask (100 mL), the corresponding ketone (20 mmol) and anhydrous THF (50 mL) were added under an inert nitrogen atmosphere. The resulting solution was cooled to -60 °C and TiCl₄ (3.3 mL, 30 mmol) was added slowly via a syringe. The mixture was stirred for 30 min and Zn dust (3.93 g, 60 mmol) was added. Then, the obtained suspension was stirred and refluxed (70 °C) for 3 h. The reaction mixture was cooled to 0 °C, saturated aqueous K₂CO₃ (25 mL) was added slowly and stirred for 30 min. The resulting mixture was filtered through a pad of Celite and washed with EtOAc. The filtrate was extracted with EtOAc. The organic layer was washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography (hexane/EtOAc = 20:1), 3,4-diethyl-3,4-hexanediol which was obtained pure without further purification.

All data matched that reported in the literature.^[4]

IV. Cyclic Voltammetry Studies for 2a-2d

Unless otherwise noted, the cyclic voltammetry measurements were conducted on a MPI-A multi-functional electrochemical and chemiluminescent system (Shanghai CH Instrument Ltd. Co., China) at room temperature, with a polished Pt plate as the working electrode, platinum thread as the counter electrode and Ag-AgNO₃ (0.1 M) in CH₃CN as the reference electrode, tetrabutylammonium tetrafluoroborate (0.1 M) was used as the supporting electrolyte, using Fc+/Fc as the internal standard, the scan rate was 0.2 V/s.



Supplementary Figure 4. Cyclic voltammograms of 2b



Supplementary Figure 6. Cyclic voltammograms of 2d

V. Optimizations of the Reaction Conditions

	- IMSF + B ₂ Cat ₂ - t	4CzIPN (2 mol%) EA(2 mL), 60 W Blue LEDs hen 3,4-diethyl-3,4-hexanediol 1h	B(EPin) SO ₂ F
1a, 0.1 mmol	2a, 0.25 mmol 3 0.25 mmol \overline{OTf} Me N^+ SO_2F	ŌTf Me N ⁺ SO ₂ F	4a OTf Me N ⁺ N ⁺ SO ₂ F
2a	2b	2c	2d
Entry	Change of conditions	Yield of 4a ^[b]	Z/E ratio ^[c]
1	None	23%	> 20:1
2	2b instead of 2a	8%	> 20:1
3	2c instead of 2a	9%	> 20:1
4	2d instead of 2a	5%	5.6:1

Supplementary Table 1: Optimization of IMSF-reagents^[a]

[a] All reactions were carried out with **1a** (12.2 mg, 0.10 mmol), **2** (0.25 mmol, 2.5 equiv.), **3** (0.25 mmol, 2.5 equiv.), 4CzIPN (2 mol%) in EA (2.0 mL) under Ar and 60 W blue LEDs, the reaction was completed as monitored by TLC analysis, after which 3,4-diethyl-3,4-hexanediol was added to the reaction system and stirred for 1 hour. [b] Yields determined by GC using dodecane as an internal standard. [c] The *Z*/*E* ratio was determined by ¹H NMR and ¹⁹F NMR.

Supplementary Table 2: Optimization of photocatalysts^[a]

1a , 0.1 mmol	+ IMSF + B ₂ Cat ₂ 2a, 0.25 mmol 3 0.25 mmol	4CzIPN (2 mol%) EA(2 mL), 60 W Blue LEDs then 3,4-diethyl-3,4-hexanediol 1h	B(EPin) 4a SO ₂ F
Entry	Change of conditions	Yield of 4a ^[b]	Z/E ratio ^[c]
1	None	23%	> 20:1
2	<i>fac</i> -Ir(ppy)₃	11%	> 20:1
3	<i>fac</i> -Ir[<i>d</i> -F-(<i>p</i> - <i>t</i> -Bu)ppy]₃	9%	5.6:1
4	Ir{[dF(CF₃)ppy]₂(dtbbpy) }PF	6 8%	6:1

5	lr(mppy)₃	11%	1.3:1
6	4-DPA-iPN	7%	> 20:1
7	Ir[(bpy)2dtbbpy]PF6	7%	> 20:1
8	2-Isopropylthioxanthone	11%	4.9:1

[a] All reactions were carried out with **1a** (12.2 mg, 0.10 mmol), **2a** (0.25 mmol, 2.5 equiv.), **3** (0.25 mmol, 2.5 equiv.), photocatalysts (2 mol%) in EA (2.0 mL) under Ar and 60 W blue LEDs, the reaction was completed as monitored by TLC analysis, after which 3,4-diethyl-3,4-hexanediol was added to the reaction system and stirred for 1 hour. [b] Yields determined by GC using dodecane as an internal standard. [c] The *Z/E* ratio was determined by ¹H NMR and ¹⁹F NMR.

Supplementary Table 3: Optimization of solutions^[a]

	+ IMSF + B ₂ Cat ₂	4CzIPN (2 mol%) EA(2 mL), 60 W Blue LEDs then 3 4-diethyl-3 4-bexanediol 1b	B(EPin)
1a, 0.1 mmol	2a , 0.25 mmol 3 0.25 mmol		4a SO ₂ F
Entry	Change of additives	Yield of 4a ^[b]	Z/E ratio ^[c]
1	None	23%	> 20:1
2	Methyl acetate	7%	> 20:1
3	1,4-dioxane	0	-
4	CH₃CN	6%	> 20:1
5	Mesitylene	trace	-
6	Acetone	0	-
7	CH ₃ CH ₂ OH	0	-

[a] All reactions were carried out with **1a** (12.2 mg, 0.10 mmol), **2a** (0.25 mmol, 2.5 equiv.), **3** (0.25 mmol, 2.5 equiv.), 4CzIPN (2 mol%) in solution (2.0 mL) under Ar and 60 W blue LEDs, the reaction was completed as monitored by TLC analysis, after which 3,4-diethyl-3,4-hexanediol was added to the reaction system and stirred for 1 hour. [b] Yields determined by GC using dodecane as an internal standard. [c] The *Z/E* ratio was determined by ¹H NMR and ¹⁹F NMR.

$\left(\right)$	// + IMSE + B.Cat	4CzIPN (2 mol%)	B(EPin)
\checkmark		EA(2 mL), 60 W Blue LEDs hen 3,4-diethyl-3,4-hexanediol 1h	
1a, 0.1 mm	ol 2a , 0.25 mmol 3 0.25 mmol		4a SO ₂ F
Entry	Change of conditions	Yield of 4a ^[b]	Z/E ratio ^[c]
1	None	23%	> 20:1
2	30W Blue LEDs	32%	> 20:1
3	90W Blue LEDs	46%	> 20:1
4	EA (1 mL)	36%	> 20:1
5	EA (1.5 mL)	33%	> 20:1
6	EA (2.5 mL)	25%	> 20:1
7	Isopropyl acetate (1 mL), 90W Blue	e LEDs 60%	> 20:1
8	Isopropyl acetate (0.6 mL), 90W LEDs	Blue 95% (70%) ^[d]	> 20:1
9	Isopropyl acetate (0.8 mL), 90W LEDs	Blue 49%	> 20:1
10	Isopropyl acetate (1.2 mL), 90W LEDs	Blue 31%	> 20:1
11	In darkness	0	-
12	w/o 4CzIPN	0	-

Supplementary Table 4: Optimization of conditions^[a]

[a] All reactions were carried out with 1a (12.2 mg, 0.10 mmol), 2a (0.25 mmol, 2.5 equiv.), 3 (0.25 mmol, 2.5 equiv.), 4CzIPN (2 mol%) in solution (X mL) under Ar and light irradiation, the reaction was completed as monitored by TLC analysis, after which 3,4-diethyl-3,4-hexanediol was added to the reaction system and stirred for 1 hour. [b] Yields determined by GC using dodecane as an internal standard. [c] The *Z*/*E*ratio was determined by ¹H NMR and ¹⁹F NMR.
[d] Isolated yield.

Supplementary Table 5: Optimization of solvents under the optimal condition^[a]



Entry	Change of additives	Yield of 4a ^[b]	Surplus of 1a ^[b]
1	None	95% (70%) ^[c]	5%
2	EtOH	0%	100%
3	CH ₃ CN	27%	73%
4	EA	84%	16%
5	THF	35%	65%
6	DCM	44%	56%
7	Toluene	17%	83%
8	DMF	0%	100%
9	Methyl acetate (MA)	82%	18%
10	DME	38%	62%

[a] All reactions were carried out with **1a** (12.2 mg, 0.10 mmol), **2a** (0.25 mmol, 2.5 equiv.), **3** (0.25 mmol, 2.5 equiv.), 4CzIPN (2 mol%) in solvents (0.6 mL) under Ar and 90 W blue LEDs, the reaction was completed as monitored by TLC analysis, after which 3,4-diethyl-3,4-hexanediol was added to the reaction system and stirred for 1 hour. [b] Yields determined by GC using dodecane as an internal standard. [c] Isolated yield.

1r , 0.1 mmol	+ IMSF + B ₂ Cat ₂ 2a, 0.25 mmol 3 0.25 mmol	Ir(ppy) ₃ (2 mol%) EA(2 mL), 90 W Blue LEDs then 3,4-diethyl-3,4-hexanediol 1h	B(EPin) SO ₂ F
Entry	Change of conditions	Yield of 4r ^[b]	Z/E ratio ^[c]
1	None	41	> 20:1
2	30W, 24h	41	> 20:1
3	60W, 24h	38	> 20:1
4	2b , 90W, 24h	34	> 20:1

Supplementary Table 6: Optimization of conditions for aryl alkynes^[a]

5	2b , 60W, 24h	32	> 20:1
6	2b , 30W, 24h	30	> 20:1
7	4CzIPN 30W, 12h	31	> 20:1
8	4CzIPN 90W, 12h	7	> 20:1
9	2a , 4CzIPN 30W, 12h	24	> 20:1
10	2a , 4CzIPN 90W, 12h	42	> 20:1
11	2a , 4CzIPN 90W, 24h	36	> 20:1
12	2a , 4CzIPN 30W, 24h	50	> 20:1
13	2a , 4CzIPN 60W, 24h	55	> 20:1
14	2a (3.0 equiv), 4CzIPN 30W, 24h	40	> 20:1
15	2a , 4CzIPN 60W, 12h	54	> 20:1
16	2a , 4CzIPN 60W, 6h	49	> 20:1
17	2a:3 =3:3, 60W, 12h	52	> 20:1
18	2a:3 =3:3, 60W, 24h	44	> 20:1
19	Isopropyl acetate instead EA	26	> 20:1
20	Methyl acetate instead EA	48	> 20:1
21	2a, Isopropyl acetate (0.6 mL), 4CzIPN 90W, 13h	69(50) ^d	> 20:1

[a] All reactions were carried out with **1r** (10.2 mg, 0.10 mmol), **2a** (0.25 mmol, 2.5 equiv.), **3** (0.25 mmol, 2.5 equiv.), 4CzIPN (2 mol%) in solution (X mL) under Ar and light irradiation, the reaction was completed as monitored by TLC analysis, after which 3,4-diethyl-3,4-hexanediol was added to the reaction system and stirred for 1 hour. [b] Yields determined by GC using dodecane as an internal standard. [c] The E/Z ratio was determined by ¹H NMR and ¹⁹F NMR. [d] Isolated yield.

Supplementary Table 7: Optimization of solvents for fluorosulfonyl-borylation of olefins^[a]

+ 5a, 0.1 mmol	$ \begin{array}{c} \overline{OTf} & Me \\ N^{+} & Ph + B_2Cat_2 \\ SO_2F \\ 2b, 0.25 \text{ mmol} & 3 0.30 \text{ mmol} \end{array} $	EA(2 mL), 90 W Blue LEDs then pinacol, NEt ₃ , 1h
Entry	Change of reagents	Yield of 6a ^[b]
1	None	54
2	Methyl acetate	49
3	Isopropyl acetate(IA)	68
4	DCM	nd
5	THF	nd
6	CH₃CN	13
7	DME	7
8	Acetone	11
9	DCE	3

[a] All reactions were carried out with **5a** (13.2 mg, 0.10 mmol), **2b** (0.25 mmol, 2.5 equiv.), **3** (0.3 mmol, 3.0 equiv.), BEt₃ (0.03 mmol, 0.3 equiv.), *fac*-Ir(ppy)₃ (2 mol%), in Solvents (2.0 mL) under Ar and 90 W blue LEDs, the reaction was completed as monitored by TLC analysis, after which pinacol and triethylamine was added to the reaction system and stirred for 1-2 h. [b] Yields determined by GC using dodecane as an internal standard.

Supplementary Table 8: Optimization of photocatalysts for fluorosulfonyl-borylation of olefins^[a]

5a , 0.1 mmol	+ B ₂ Cat ₂ Ir(ppy) ₃ (2 mol%), BEt ₃ (0.3 IA(2 mL), 90 W Blue LE then pinacol, NEt ₃ , 1 2b , 0.25 mmol 3 0.30 mmol	equiv) EDs h 6a
Entry	Change of Photocatalysts	Yield of 6a ^[b]
1	None	68
2	<i>fac</i> -Ir[<i>d</i> -F-(<i>p</i> - <i>t</i> -Bu)ppy]₃	46
3	Ir[{dF(CF ₃)ppy} ₂ (dtbbpy)]PF ₆	38

4	4CzIPN	52
5	lr(mppy) ₃	61
6	4-DPA-iPN	29
7	lr[dF(CF₃)ppy]₃	41
8	lr[dFppy]₃	48

[a] All reactions were carried out with **5a** (13.2 mg, 0.10 mmol), **2b** (0.25 mmol, 2.5 equiv.), **3** (0.3 mmol, 3.0 equiv.), BEt₃ (0.03 mmol, 0.3 equiv.), photocatalysts (2 mol%), in IA (2.0 mL) under Ar and 90 W blue LEDs, the reaction was completed as monitored by TLC analysis, after which pinacol and triethylamine was added to the reaction system and stirred for 1-2 h. [b] Yields determined by GC using dodecane as an internal standard.

Supplementary Table 9: Optimization of material ratio for fluorosulfonyl-borylation of olefins^[a]

$\bigcirc \frown \frown$	+ B_2Cat_2 $\frac{Ir(ppy)_3 (2 \text{ mol}\%)}{IA(2 \text{ mL}), 90 \text{ W}}$ then pinaced	Et ₃ (0.3 equiv) Blue LEDs
5a, 0.1 mmol	2b , 0.25 mmol 3 , 0.30 mmol	6a
Entry	Change of material ratio(5a:2a:3 :BEt ₃)	Yield of 6a ^[b]
1	None	68
2	1:1.5:2:0.2	28
3	1:2:2:0.2	19
4	1:2.5:2:0.2	28
5	1:2.5:1.5:0.2	10
6	1:2.5:2.5:0.2	23
7	1:2.5:3:0.2	64
8	1:2.5:3:0.15	47
9	1:2.5:3:0.25	69
10	1:2.5:3:0.35	66
11	1:2:2:0.15	49
12	1:2:3:0.3	19
13	1:2:2:0.3	49
14	1:2:3:0.2	49
15	1:1.5:3:0.3	17

[a] All reactions were carried out with **5a** (13.2 mg, 0.10 mmol), **2b** (x mmol, x equiv.), **3** (y mmol, y equiv.), BEt₃ (z mmol, z equiv.), *fac*-Ir(ppy)₃ (2 mol%), in IA (2.0 mL) under Ar and 90 W blue LEDs, the reaction was completed as monitored by TLC analysis, after which pinacol and

triethylamine was added to the reaction system and stirred for 1-2 h. [b] Yields determined by GC using dodecane as an internal standard.

5a, 0.1 mmol	$\begin{array}{c} \overline{OTf} \overset{Me}{\overset{N+}}{\overset{N+}{\overset{N+}{\overset{N+}{\overset{N+}{\overset{N+}{\overset{N+}{\overset{N+}{\overset{N+}{\overset{N+}}{\overset{N+}}{\overset{N+}}}}}}}}}}$	by) ₃ (2 mol%) _, BEt ₃ (0.25 equiv) IA (2 mL), 90 W Blue LEDs then pinacol, NEt ₃ , 1h 6a
OTf + Me	CF ₃	ŌTf + Me N SO ₂ F
2a	2c	2d
Entry	PC	Yield of 6a ^[b]
1	None	69
2	2a instead of 2b	68
3	2c instead of 2b	59
4	2d instead of 2b	8
5	w/o lr(ppy)₃	nd
6	In the drakness	nd

Supplementary Table 10: Optimization of IMSF reagents and control experiments for fluorosulfonyl-borylation of olefins^[a]

[a] All reactions were carried out with **5a** (13.2 mg, 0.10 mmol), **2** (0.25 mmol, 2.5 equiv.), **3** (0.3 mmol, 3 equiv.), BEt₃ (0.025 mmol, 0.25 equiv.), *fac*-Ir(ppy)₃ (2 mol%), in IA (2.0 mL) under Ar and 90 W blue LEDs, the reaction was completed as monitored by TLC analysis, after which pinacol and triethylamine was added to the reaction system and stirred for 1-2 h. [b] Yields determined by GC using dodecane as an internal standard.

Supplementary Table 11: Optimization of IMSF reagents and other conditons for fluorosulfonyl-borylation of olefins^[a]

5a , 0.1 mmol	+ $\overrightarrow{N}_{SO_2F}^{OTf, Me}$ + $B_2Cat_2 \frac{lr(pp)}{l}$ 2b, 0.25 mmol 3, 0.30 mmol	() ₃ (2 mol%) BEt ₃ (0.25 equiv) A (2 mL), 90 W Blue LEDs then pinacol, NEt ₃ , 1h 6a
	CF ₃	
Entry	Change of conditions	Yield of 6a ^[b]
1	None	69
2	30W, 12h	67

3 60W, 12h 62 4 90W, 24h 25 5 2a , 90W, 24h 80 6 3 equiv of 2b , 12h 54 7 3.5 equiv of 2b , 12h 62 8 3.0 equiv of 2b , 12h, 30W 21 10 3.0 equiv of 2a , 12h , 30W 86(73) ^c 11 3.0 equiv of 2b , 12h, 60W 50			
4 90W, 24h 25 5 2a , 90W, 24h 80 6 3 equiv of 2b , 12h 54 7 3.5 equiv of 2b , 12h 62 8 3.0 equiv of 2b , 12h, 30W 21 10 3.0 equiv of 2a , 12h , 30W 86(73) ^c 11 3.0 equiv of 2b , 12h, 60W 50	3	60W, 12h	62
5 2a, 90W, 24h 80 6 3 equiv of 2b, 12h 54 7 3.5 equiv of 2b, 12h 62 8 3.0 equiv of 2b, 12h, 30W 21 10 3.0 equiv of 2a, 12h, 30W 86(73)° 11 3.0 equiv of 2b, 12h, 60W 50	4	90W, 24h	25
6 3 equiv of 2b, 12h 54 7 3.5 equiv of 2b, 12h 62 8 3.0 equiv of 2b, 12h, 30W 21 10 3.0 equiv of 2a, 12h, 30W 86(73)° 11 3.0 equiv of 2b, 12h, 60W 50	5	2a , 90W, 24h	80
7 3.5 equiv of 2b, 12h 62 8 3.0 equiv of 2b, 12h, 30W 21 10 3.0 equiv of 2a, 12h, 30W 86(73)° 11 3.0 equiv of 2b, 12h, 60W 50	6	3 equiv of 2b , 12h	54
8 3.0 equiv of 2b, 12h, 30W 21 10 3.0 equiv of 2a, 12h, 30W 86(73) ^c 11 3.0 equiv of 2b, 12h, 60W 50	7	3.5 equiv of 2b , 12h	62
103.0 equiv of 2a, 12h, 30W86(73)°113.0 equiv of 2b, 12h, 60W50	8	3.0 equiv of 2b , 12h, 30W	21
11 3.0 equiv of 2b , 12h, 60W 50	10	3.0 equiv of 2a, 12h, 30W	86(73)°
	11	3.0 equiv of 2b , 12h, 60W	50

[a] All reactions were carried out with **5a** (13.2 mg, 0.10 mmol), **2** (x mmol, x equiv.), **3** (0.3 mmol, 3 equiv.), BEt₃ (0.025 mmol, 0.25 equiv.), *fac*-Ir(ppy)₃ (2 mol%), in IA (2.0 mL) under Ar and Light irradiation, the reaction was completed as monitored by TLC analysis, after which pinacol and triethylamine was added to the reaction system and stirred for 1-2 h. [b] Yields determined by GC using dodecane as an internal standard. [c] Isolated Yields.

VI. General Procedure for the Synthesis of the Products 4, 6-9, 11, 13-15, 17-18, 20-21.

General Procedure for the synthesis of products 4a, 4c-4e. Condition A: Under argon, to a solution of 4CzIPN (2 mol%), 2,2'-Bis-1,3,2-benzodioxaborole **3** (0.25 mmol, 2.5 equiv.) and IMSF reagent **2a** (0.2 mmol, 2 equiv.) in dried isopropyl acetate (0.6 mL) was added corresponding alkynes **1a, 1c-1e** (0.1 mmol) at room temperature. After that, the tube was exposed to a 90 W blue LEDs about 40min-12h, after which 3,4-diethyl-3,4-hexanediol was added to the reaction system and stirred for 1 h. The reaction was completed as monitored by TLC analysis and the reaction mixture was evaporated in *vacuo*. The crude products were directly purified by flash chromatography on silica gel to give the desired products.

General Procedure for the synthesis of products 4b, 4f-4l. Condition B: Under argon, to a solution of 4CzIPN (2 mol%), 2,2'-Bis-1,3,2-benzodioxaborole **3** (0.25 mmol, 2.5 equiv.) and IMSF reagent **2a** (0.2 mmol, 2 equiv.) in dried isopropyl acetate : ethyl acetate= 2:1 (0.8 mL) was added corresponding alkynes **1b**, **1f-1l** (0.1 mmol) at room temperature. After that, the tube was exposed to a 90 W blue LEDs about 40min-12h, after which 3,4-diethyl-3,4-hexanediol was added to the reaction system and stirred for 1 h. The reaction was completed as monitored by TLC analysis and the reaction mixture was evaporated in *vacuo*. The crude products were directly purified by flash chromatography on silica gel to give the desired products.

General Procedure for the synthesis of products 4m-4x. Condition C: Under argon, to a solution of 4CzIPN (2 mol%), 2,2'-Bis-1,3,2-benzodioxaborole 3 (0.25 mmol, 2.5 equiv.) and IMSF reagent 2a (0.2 mmol, 2 equiv.) in dried isopropyl acetate (0.6 mL) was added corresponding alkynes 1m-1x (0.1 mmol) at room temperature. After that, the tube was

exposed to a 90 W blue LEDs about 13 h, after which 3,4-diethyl-3,4-hexanediol was added to the reaction system and stirred for 1 h. The reaction was completed as monitored by TLC analysis and the reaction mixture was evaporated in *vacuo*. The crude products were directly purified by flash chromatography on silica gel to give the desired products.

General Procedure for the synthesis of product 6. Condition D: Under argon, to a solution of *fac*-Ir(ppy)₃ (2 mol%), BEt₃ (0.025 mmol, 0.25 equiv.), 2,2'-Bis-1,3,2-benzodioxaborole **3** (0.3 mmol, 3.0 equiv.) and IMSF reagent **2a** (0.3 mmol, 3.0 equiv.) in dried isopropyl acetate (2 mL) was added corresponding olefins **5** (0.1 mmol) at room temperature. After that, the tube was exposed to a 30 W blue LEDs about 12 h, after which pinacol and triethylamine was added to the reaction system and stirred for 1-2 h. The reaction was completed as monitored by TLC analysis and the reaction mixture was evaporated in *vacuo*. The crude products were directly purified by flash chromatography on silica gel to give the desired products.

General Procedure for the synthesis of product 7 and 8^[5]**. Condition E:** Under argon, to a solution of **4m** (0.05 mmol) in dried MeOH (1 mL) was added CuX₂ (1.0 equiv.) at room temperature. After that, the tube was heated to 80 °C about 12 h, then until the reaction was completed as monitored by TLC analysis. The reaction mixture was evaporated in *vacuo*. The crude products were directly purified by flash chromatography on silica gel to give the desired products.

General Procedure for the synthesis of product 9^[6]**. Condition F:** Under argon, to a solution of **4m** (0.05 mmol), Pd(OAc)₂ (5 mol%), P(o-Tol)₃ (10 mol%), in dried toluene (0.5 mL) was added H₂O (2.5 equiv.) at room temperature. After that, the tube was heated to 80 °C about 12

h, then until the reaction was completed as monitored by TLC analysis. The reaction mixture was evaporated in *vacuo*. The crude products were directly purified by flash chromatography on silica gel to give the desired products.

General Procedure for the synthesis of product 11, 13, 17, 20^[7]**. Condition G:** Under argon, to a solution of **4m** (0.05 mmol), Pd(OAc)₂ (15 mol%), SPhos (35 mol%), K₃PO₄ (1.5 equiv.) in dried toluene (0.5 mL) was added corresponding aryl bromides **12, 16, 19** (0.06 mmol) or 3,6-dihydro-2H-pyran-4-yl trifluoromethanesulfonate **10** (0.06 mmol) at room temperature. After that, the tube was heated to 80 °C about 12 h, then until the reaction was completed as monitored by TLC analysis. The reaction mixture was evaporated in *vacuo*. The crude products were directly purified by flash chromatography on silica gel to give the desired products.

General Procedure for the synthesis of product 14^[8]**. Condition H:** To a solution of sesamol (1.0 equiv.), HMDS (1.0 equiv.), and BTMG (20 mol%) in dried MeCN (0.5 mL) was added corresponding alkenes **13** at room temperature. After that, the tube was heated to a 60 °C about 1 h, then until the reaction was completed as monitored by TLC analysis. The reaction mixture was evaporated in *vacuo*. The crude products were directly purified by flash chromatography on silica gel to give the desired products.

General Procedure for the synthesis of product 15, 21^[9]**. Condition I:** To a solution of amines (2.0 equiv.), Ca(NTf)₂ (1.0 equiv.) in t-amyIOH (0.5 mL) was added corresponding alkenes **13** or **20** at room temperature. After that, the tube was heated to a 60 °C about 24 h, then until the reaction was completed as monitored by TLC analysis. The reaction mixture was

evaporated in *vacuo*. The crude products were directly purified by flash chromatography on silica gel to give the desired products.

General Procedure for the synthesis of product 18^[3]**. Condition J:** To a solution of estrone (2.0 equiv.), KOH (2.0 equiv.) in MeCN (0.5 mL) was added corresponding alkene **17** at room temperature. After that, the tube was heated to a 50 °C about 12 h, then until the reaction was completed as monitored by TLC analysis. The reaction mixture was evaporated in vacuo. The crude products were directly purified by flash chromatography on silica gel to give the desired products.

VII. Mechanistic experiments

A. Luminescence quenching experiment

The luminescence quenching experiment was taken using a F-4600 FL Spectrophotometer (Hitachi, Japan). The experiments were carried out in 5 x 10⁻⁴ mol/L of Ir(ppy)₃ in CH₃CN at 25 ^oC with an excitation wavelength of 380 nm and an excitation and emission bandwidth of 5 nm, The scanspeed was set at 1200 nm/min and the PMT voltage was set to 500 V. The concentrations of quencher (**5a**, IMSF-**2a**, B₂Cat₂) in CH₃CN were 0.01 mmol/mL. The concentrations of quencher IMSF-**2a** in CH₃CN was 4 mmol/L, 8 mmol/L, 12 mmol/L, 16 mmol/L. (see supplementary figure **7** and supplementary figure **8-9**)

To determine whether a reductive or oxidative quenching cycle is operative in the reaction, fluorescence quenching studies were conducted. Based on the above data, photoexcited $Ir(ppy)_{3}^{*}$ can be quenched by IMSF-**2a**, which involving a oxidative quenching cycle.



Supplementary Figure 7. The data of fluorescence quenching of Ir(ppy)₃, B₂Cat₂, 5a, IMSF-

2a



Supplementary Figure 8. The data of fluorescence quenching of Ir(ppy)₃ by different concentrations of IMSF-2a



Supplementary Figure 9. Stern-Volmer plot of Ir(ppy)3 at different concentrations of IMSF-2a

B. ¹¹B NMR experiments

All ¹¹B NMR data was recorded at 25 °C. See supplementary figure **10-13** for details of reagents and solvents; supplementary figure **10** shows B_2Cat_2 in CD_3CN ; supplementary figure **11** shows B_2Cat_2 : Isopropyl acetate (IA) = 1:1 in CD_3CN ; supplementary figure **12** shows B_2Cat_2 : IMSF-**2a** = 1:1 in CD_3CN ; supplementary Figure **13** shows B_2Cat_2 : 1-methyl-2-(4-(trifluoromethyl)phenyl)-1H-benzo[d]imidazole = 1:1 in CD_3CN .

0 -10 -20 -30 -40 -50 -60 -70 -80 f1 (ppm) 90 80 70 60 50 40 30 20 10 -90

Supplementary Figure 10. B₂Cat₂ in CD₃CN



Supplementary Figure 12. B₂Cat₂: IMSF-2a =1:1 in CD₃CN



Supplementary Figure 13. B₂Cat₂: 1-methyl-2-(4-(trifluoromethyl)phenyl)-1Hbenzo[d]imidazole =1:1 in CD₃CN

¹¹B NMR experiments reveal that there is just a single signal whether only B₂Cat₂ in CD₃CN, mix isopropyl acetate with B₂Cat₂ in CD₃CN or mix IMSF-**2a** with B₂Cat₂ in CD₃CN (supplementary figure 10-12); when mix imidazole residue with B₂Cat₂ in CD₃CN, the latter shows one upfield signal (supplementary figure 13). The upfield shifting supports the ligation of imidazole residue with diboron species.



Under argon, to a solution of 4CzIPN (2 mol%), B₂Cat₂ (2.5 equiv.), TEMPO (3.0 equiv.) and **IMSF-2a** (0.25 mmol, 2.5 equiv.) in dried IA (0.6 mL) was added corresponding alkyne **1a** (0.1 mmol) at room temperature. After that, the tube was exposed to a 90 W blue LEDs about 40min until the reaction was completed as monitored by TLC analysis. Subsequently, the reaction mixture was analyzed by GC. GC showed that no major product **4a** was formed after addition of 0.3 mmol of TEMPO.



Under argon, to a solution of 4CzIPN (2 mol%), B₂Cat₂ (2.5 equiv), 1,1-diphenylethylene (0.2 mmol, 2.0 equiv.) and **IMSF- 2a** (0.25 mmol, 2.5 equiv.) in dried IA (0.6 mL) was added corresponding alkyne **1a** (0.1 mmol) at room temperature. After that, the tube was exposed to a 90 W blue LEDs about 40min until the reaction was completed as monitored by TLC analysis. Subsequently, the reaction mixture was analyzed by GC. GC showed that trace product **4a** was formed after addition of 0.2 mmol of 1,1-diphenylethylene. In addition, product **13** can be obtained with a 66% separation yield.

D. Proposed mechanism

From the above mechanistic experiments, we speculate on the possible mechanism of the reaction: First, under the irradiation, the cationic reagent **2a** can be reduced by excited state photocatalyst (PC*) to generate radical intermediate I and releases SO₂F radical and imidazole residue II. Then the addition of SO₂F radical to the alkynes regioselectively furnishes vinylic radical intermediate III. Subsequent addition of vinyl radical III to B₂cat₂ afforded a *Z*-vinyl diboron radical species IV. The control of stereoselectivity is governed by steric repulsion between the fluorosulfonyl group and the boronates. Then, the activation of diboron reagent by in situ generated imidazole residues II forms a highly reactive B–N heteroleptic intermediate V, which leads to the desired bifunctional products 4 or 6 and imidazole stabilized boryl radical species VI. Finally, photo-oxidation of VI followed by coupling with ⁻OTf affords boryl imidazolium salt VII and regenerates PC. (supplementary figure 14).



Supplementary Figure 14. Proposed mechanism

VIII. References

- Q. Y. Meng, T. E. Schirmer, K. Katou, B. Konig, *Angew. Chem. Int. Ed.*, 2019, 58, 5723-5728.
- 2. Toshima, H.; Maru, K.; Saito, M.; Ichihara, A. Tetrahedron, 1999, 55, 5793.
- 3. W. Zhang, H. Li, X. Li, Z. Zou, M. Huang, J. Liu, X. Wang, S. Ni, Y. Pan, Y. Wang, *Nat. Commun.*, 2022, **13**, 3515.
- R. Rubio-Presa, S. Suárez-Pantiga, M. R. Pedrosa, R. Sanz, *Adv. Synth. Catal.*, 2018, 360, 2216.
- W. Zhang, Z. Zou, W. Zhao, S. Lu, Z. Wu, M. Huang, X. Wang, Y. Wang, Y. Liang, Y. Zhu, Y. Zheng, Y. Pan, *Nat. Commun.*, 2020, **11**, 2572.
- 6. S. Rao, K. R. Prabhu, Chem. Eur. J., 2018, 24, 13954-13962.
- 7. N. Oka, T. Yamada, H. Sajiki, S. Akai, T. Ikawa, Org. Lett., 2022, 24, 3510-3514.
- C. J. Smedley, J. A. Homer, T. L. Gialelis, A. S. Barrow, R. A. Koelln, J. E. Moses, *Angew. Chem. Int. Ed.*, 2022, **61**, e202112375.
- S. Mahapatra, C. P. Woroch, T. W. Butler, S. N. Carneiro, S. C. Kwan, S. R. Khasnavis, J. Gu, J. K. Dutra, B. C. Vetelino, J. Bellenger, C. W. Am Ende, N. D. Ball, *Org. Lett.*, 2020, 22, 4389-4394.
- 10. X. Nie, T. Xu, Y. Hong, H. Zhang, C. Mao, S. Liao, Angew. Chem. Int. Ed., 2021, 60, 22035.

IX. Characteristic Data

(Z)-3-cyclohexyl-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)prop-1-ene-1-sulfonyl fluoride (4a)



70% (27.2 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 6.86 (s, 1H), 2.59 (d, J = 6.8 Hz, 2H), 1.79 – 1.60 (m, 13H), 1.34 – 1.11 (m, 6H), 0.92 (t, J = 7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 129.8 (d, J = 23.2 Hz), 90.3, 38.1, 37.9, 33.2, 32.8, 26.3, 26.2, 8.7. The signal of the α-B-carbon was not observed. ¹⁹F

NMR (376 MHz, Chloroform-*d*) δ 65.06. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 29.09. HRMS(ESI): caled for C₁₉H₃₅BFO₄S⁺ [M + H]⁺ 389.2328; found 389.2326.

(Z)-5-chloro-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)pent-1-ene-1-sulfonyl fluoride

(4b)



57% (20.9 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 6.91 (s, 1H), 3.56 (t, J = 6.7 Hz, 2H), 2.80 (ddt, J = 9.2, 5.9, 1.3 Hz, 2H), 2.07 – 1.93 (m, 2H), 1.71 (qd, J = 7.3, 4.5 Hz, 8H), 0.92 (t, J =7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 130.8 (d, J =23.6 Hz), 90.6, 44.3, 31.6, 28.5, 26.3, 8.7. The signal of the α-B-

carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 64.99. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 28.87. HRMS(ESI): caled for C₁₅H₂₈BCIFO₄S⁺ [M + H]⁺ 369.1469; found 369.1466.

(Z)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)pent-1-ene-1-sulfonyl fluoride (4c)



54% (18.0 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 6.86 (s, 1H), 2.69 – 2.61 (m, 2H), 1.70 (qd, J = 7.3, 4.3 Hz, 8H), 1.59 – 1.49 (m, 2H), 0.97 (t, J = 7.4 Hz, 3H), 0.92 (t, J = 7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 129.7 (d, J = 23.2 Hz), 90.3, 32.6, 26.3, 22.3, 14.1, 8.7. The signal of the α-B-carbon was not observed. ¹⁹F

NMR (376 MHz, Chloroform-*d*) δ 64.92. ¹¹B NMR (160 MHz, Chloroform-*d*) δ 28.94. HRMS(ESI): caled for C₁₅H₂₉BFO₄S⁺ [M + H]⁺ 335.1858; found 335.1856. (Z)-4-phenyl-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)but-1-ene-1-sulfonyl fluoride (4d)



40% (15.8 mg); colorless oil; ¹H NMR (500 MHz, Chloroform-d) δ 7.32 (dd, J = 8.0, 6.9 Hz, 2H), 7.24 (dd, J = 5.8, 2.7 Hz, 3H), 6.92 (s, 1H), 3.03 – 2.96 (m, 2H), 2.85 – 2.79 (m, 2H), 1.72 (qd, J = 7.3, 4.3 Hz, 8H), 0.96 (t, J = 7.4 Hz, 12H). ¹³C NMR (126 MHz, Chloroform-d) δ 140.7, 130.3 (d, J = 23.3 Hz), 128.5, 128.5, 126.3, 125.3, 90.5, 35.1, 32.9, 26.3, 8.7, The signal of the α-B-carbon was not observed. ¹⁹F NMR

(471 MHz, Chloroform-d) δ 64.98. ¹¹B NMR (160 MHz, Chloroform-d) δ 28.27. HRMS(ESI): caled for C₂₀H₃₁BFO₄S⁺ [M + H]⁺ 397.2015; found 397.2016.

(Z)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)dodec-1-ene-1-sulfonyl fluoride (4e)



51% (22.0 mg); colorless oil; ¹H NMR (500 MHz, Chloroform-d) δ 6.83 (s, 1H), 2.68 - 2.60 (m, 2H), 1.76 - 1.63 (m, 8H), 1.48 (ddd, J =11.4, 8.6, 6.2 Hz, 2H), 1.37 - 1.22 (m, 14H), 0.92 (t, J = 7.5 Hz, 12H), 0.88 (t, J = 6.9 Hz,

3H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 129.4 (d, *J* = 22.9 Hz), 90.3, 31.9, 30.8, 29.6, 29.6, 29.4, 29.3, 29.2, 28.8, 26.3, 22.7, 14.1, 8.7. The signal of the α-B-carbon was not observed. ¹⁹F NMR (471 MHz, Chloroform-*d*) δ 64.94. ¹¹B NMR (160 MHz, Chloroform-*d*) δ 29.41. HRMS(ESI): caled for C₂₂H₄₃BFO₄S⁺ [M + H]⁺ 433.2954; found 433.2952.

(Z)-5-(fluorosulfonyl)-4-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)pent-4-en-1-yl 1fluorocyclopropane-1-carboxylate (4f)



50% (21.8 mg); colorless oil; ¹H NMR (500 MHz, Chloroform-d) δ 6.91 (s, 1H), 4.23 (t, J = 6.2 Hz, 2H), 2.77 - 2.71 (m, 2H), 1.89 (ddt, J = 9.3, 7.8, 6.2 Hz, 2H), 1.70 (m, 8H), 1.39 (s, 2H), 1.37 - 1.35 (m, 2H), 0.92 (t, J = 7.4 Hz, 12H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 170.5 (d, J =

24.2 Hz), 130.8 (d, J = 23.6 Hz), 90.6, 73.8, 64.8, 27.8, 27.3, 26.3, 14.6, 14.5, 8.7. The signal

of the α -B-carbon was not observed. ¹⁹F NMR (471 MHz, Chloroform-*d*) δ 64.96, -197.85. ¹¹B NMR (160 MHz, Chloroform-*d*) δ 28.99. HRMS(ESI): caled for C₁₉H₃₂BF₂O₆S⁺ [M + H]⁺ 437.1975; found 437.1977.

(Z)-5-(fluorosulfonyl)-4-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)pent-4-en-1-yl 3,3difluorocyclobutane-1-carboxylate (4g)



45% (21.1 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 6.91 (s, 1H), 4.16 (t, *J* = 6.1 Hz, 2H), 3.01 – 2.91 (m, 1H), 2.90 – 2.71 (m, 6H), 1.93 – 1.82 (m, 2H), 1.79 – 1.58 (m, 8H), 0.92 (t, *J* = 7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 173.2, 130.8 (d,

J = 23.8 Hz), 90.6, 64.4, 38.7 (t, J = 24.5 Hz), 27.8, 27.3, 26.5 (dd, J = 14.5, 5.0 Hz), 26.3, 8.7. The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 64.93, -82.69-83.26 (m), -96.99-97.67 (m). ¹¹B NMR (128 MHz, Chloroform-*d*) δ 29.01. HRMS(ESI): caled for C₂₀H₃₃BF₃O₆S⁺ [M + H]⁺ 469.2038; found 469.2036.

(Z)-5-(fluorosulfonyl)-4-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)pent-4-en-1-yl cyclobutanecarboxylate (4h)



54% (23.3 mg); colorless oil; ¹H NMR (500 MHz, Chloroform-*d*) δ 6.89 (s, 1H), 4.11 (t, *J* = 6.2 Hz, 2H), 3.13 (pd, *J* = 8.5, 1.1 Hz, 1H), 2.75 (t, *J* = 7.7 Hz, 2H), 2.36 – 2.23 (m, 2H), 2.24 – 2.13 (m, 2H), 2.03 – 1.92 (m, 1H), 1.92 – 1.82 (m, 3H), 1.77 – 1.62 (m, 8H), 0.92 (t, *J* = 7.4

Hz, 12H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 175.4, 130.5 (d, J = 23.3 Hz), 90.5, 63.6, 38.1, 28.0, 27.5, 26.3, 25.3, 18.4, 8.7. The signal of the α-B-carbon was not observed. ¹⁹F NMR (471 MHz, Chloroform-*d*) δ 64.97. ¹¹B NMR (160 MHz, Chloroform-*d*) δ 29.37. HRMS(ESI): caled for $C_{20}H_{35}BFO_6S^+$ [M + H]⁺ 433.2226; found 433.2224.

(Z)-5-(fluorosulfonyl)-4-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)pent-4-en-1-yl cyclopentanecarboxylate (4i)



51% (22.8 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 6.89 (s, 1H), 4.11 (t, *J* = 6.2 Hz, 2H), 2.80 – 2.68 (m, 3H), 1.93 – 1.76 (m, 6H), 1.70 (qd, *J* = 7.3, 3.7 Hz, 9H), 1.58 (tdt, *J* = 6.3, 4.7, 2.2 Hz, 3H), 0.92 (t, *J* = 7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*)

δ 176.7, 130.5 (d, *J* = 23.4 Hz), 90.5, 63.6, 43.9, 30.0, 28.0, 27.6, 26.3, 25.8, 8.7. The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 64.98. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 28.98. HRMS(ESI): caled for C₂₁H₃₇BFO₆S⁺ [M + H]⁺ 447.2383; found 447.2380.

(Z)-5-(fluorosulfonyl)-4-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)pent-4-en-1-yl (1R,2R)-2-phenylcyclopropane-1-carboxylate (4j)



41% (20.3 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.32 – 7.25 (m, 2H), 7.24 – 7.15 (m, 1H), 7.14 – 7.07 (m, 2H), 6.89 (s, 1H). 4.15 (t, *J* = 6.2 Hz, 2H), 2.77 (t, *J* = 7.7 Hz, 2H), 2.53 (ddd, *J* = 9.3, 6.5, 4.1 Hz, 1H), 1.95 – 1.82 (m,

3H), 1.77 – 1.62 (m, 8H), 1.60 (ddd, J = 9.6, 5.3, 4.5 Hz, 1H), 1.32 (ddd, J = 8.4, 6.5, 4.5 Hz, 1H), 0.90 (td, J = 7.5, 2.4 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 173.2, 140.1, 130.5 (d, J = 23.6 Hz), 128.4, 126.5, 126.2, 90.5, 27.9, 27.5, 26.3, 26.2, 24.1, 17.0, 8.7. The signal of the α -B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 65.03. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 29.06. HRMS(ESI): caled for C₂₅H₃₇BFO₆S⁺ [M + H]⁺ 495.2383; found 495.2386.

(Z)-5-(fluorosulfonyl)-4-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)pent-4-en-1-yl 4-

methylcyclohexane-1-carboxylate (4k)



50% (23.7 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 6.89 (s, 1H), 4.10 (t, *J* = 6.1 Hz, 2H), 2.74 (t, *J* = 7.8 Hz, 2H), 2.20 (tt, *J* = 12.3, 3.6 Hz, 1H), 1.95 (dq, *J* = 12.4, 3.5, 3.0 Hz, 2H), 1.89 – 1.79 (m, 2H), 1.80 – 1.63 (m, 11H), 1.50 – 1.28

(m, 4H), 0.92 (t, *J* = 7.5 Hz, 12H), 0.89 (d, *J* = 6.5 Hz, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 176.2, 130.5 (d, *J* = 23.5 Hz), 90.5, 63.4, 43.2, 34.3, 32.0, 29.0, 28.0, 27.6, 26.3, 22.5, 8.7. The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 64.98. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 28.88. HRMS(ESI): caled for $C_{23}H_{41}BFO_6S^+$ [M + H]⁺ 475.2696; found 475.2699.

(Z)-5-(fluorosulfonyl)-4-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)pent-4-en-1-yl 1-(2,2difluorobenzo[d][1,3]dioxol-5-yl)cyclopropane-1-carboxylate (4l)



32% (18.4 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.10 – 7.03 (m, 2H), 6.97 (dd, *J* = 7.8, 0.8 Hz, 1H), 6.88 (s, 1H), 4.08 (t, *J* = 6.0 Hz, 2H), 2.66 – 2.58 (m, 2H), 1.78 – 1.62 (m, 12H), 1.17 (q, *J* =

4.0 Hz, 2H), 0.91 (t, J = 7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 173.9, 135.8, 131.7, 130.8 (d, J = 23.6 Hz), 125.8, 112.0, 108.9, 90.6, 64.5, 29.7, 29.0, 27.9, 27.5, 26.3, 16.9, 8.7. The signal of the α -B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 64.87, -49.90. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 29.30; HRMS(ESI): caled for C₂₆H₃₅BF₃O₈S⁺ [M + H]⁺ 575.2093; found 575.2091.

(Z)-2-phenyl-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)ethene-1-sulfonyl fluoride (4m)



50% (18.4 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.39 (dd, J = 5.1, 2.0 Hz, 3H), 7.37 – 7.30 (m, 2H), 7.08 (s, 1H), 1.71 (hept, J = 7.0 Hz, 8H), 0.91 (t, J = 7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 134.8, 130.4 (d, J = 25.8 Hz), 129.2, 128.1, 127.8 (d, J = 1.3 Hz), 90.8, 26.3, 8.7. The signal of the α-B-carbon was not observed. ¹⁹F NMR

(376 MHz, Chloroform-*d*) δ 64.47. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 28.98. HRMS(ESI): caled for C₁₈H₂₇BFO₄S⁺ [M + H]⁺ 369.1702; found 369.1705.

(Z)-2-(4-(tert-butyl)phenyl)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)ethene-1-





52% (22.0 mg); white solid; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.44 – 7.29 (m, 4H), 7.03 (s, 1H), 1.72 (hept, J = 7.4 Hz, 8H), 1.33 (s, 9H), 0.92 (t, J = 7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform*d*) δ 152.5, 131.6, 129.4 (d, J = 25.9 Hz), 128.1, 125.1, 90.7, 34.8, 31.2, 26.3, 8.7. The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 64.17. ¹¹B NMR (128 MHz,

Chloroform-*d*) δ 29.87. HRMS(ESI): caled for C₂₂H₃₅BFO₄S⁺ [M + H]⁺ 425.2328; found 425.2325.

(Z)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)-2-(p-tolyl)ethene-1-sulfonyl fluoride (40)



62% (23.6 mg); colorless oil; ¹H NMR (500 MHz, Chloroform-*d*) δ 7.35 – 7.29 (m, 2H), 7.24 (d, J = 8.0 Hz, 2H), 7.08 (s, 1H), 2.42 (s, 3H), 1.83 – 1.69 (m, 8H), 0.96 (t, J = 7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 139.4, 131.8, 129.7 (d, J = 25.8 Hz), 128.9, 128.1, 90.7, 26.3, 21.4, 8.7. The signal of the α-B-carbon was not observed.

¹⁹F NMR (376 MHz, Chloroform-*d*) δ 64.38. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 29.18. HRMS(ESI): caled for C₁₉H₂₉BFO₄S⁺ [M + H]⁺ 383.1858; found 383.1859. (Z)-2-([1,1'-biphenyl]-4-yl)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)ethene-1-sulfonyl fluoride (4p)



126.8, 90.9, 26.3, 8.7. The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 64.40. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 29.23. HRMS(ESI): caled for $C_{24}H_{31}BFO_4S^+$ [M + H]⁺ 445.2015; found 445.2018.

(Z)-2-(4-pentylphenyl)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)ethene-1-sulfonyl fluoride (4q)



44% (19.3 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.31 – 7.27 (m, 2H), 7.21 – 7.18 (m, 2H), 7.03 (s, 1H), 2.69 – 2.55 (m, 2H), 1.78 – 1.66 (m, 8H), 1.66 – 1.59 (m, 2H), 1.37 – 1.30 (m, 4H), 0.98 – 0.87 (m, 15H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 144.4, 131.9,

129.48 (d, J = 25.7 Hz), 128.1, 128.1, 90.7, 35.8, 31.6, 30.7, 26.3, 22.5, 14.0, 8.7. The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 64.27. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 28.42. HRMS(ESI): caled for C₂₃H₃₇BFO₄S⁺ [M + H]⁺ 439.2484; found 439.2483.

(Z)-2-(4-fluorophenyl)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)ethene-1-sulfonyl fluoride (4r)



52% (20.0 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.40 – 7.31 (m, 2H), 7.14 – 7.05 (m, 3H), 2.00 – 1.62 (m, 8H), 0.91 (t, J = 7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 164.5, 162.0, 130.6 (d, J = 26.1 Hz), 130.1 (d, J = 9.6 Hz), 115.3 (d, J = 21.8Hz), 90.9, 26.3, 8.7. The signal of the α-B-carbon was not observed.

¹⁹F NMR (376 MHz, Chloroform-*d*) δ 64.37, -111.54 – -111.66 (m). ¹¹B NMR (128 MHz, Chloroform-*d*) δ 29.33. HRMS(ESI): caled for C₁₈H₂₆BF₂O₄S⁺ [M + H]⁺ 387.1608; found 387.1610.

(Z)-2-(4-chlorophenyl)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)ethene-1-sulfonyl fluoride (4s)



50% (20.0 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.40 – 7.33 (m, 2H), 7.30 – 7.26 (m, 2H), 7.09 (s, 1H), 1.79 – 1.63 (m, 8H), 0.91 (t, J = 7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform*d*) δ 135.4, 133.1, 130.9 (d, J = 26.0 Hz), 129.3 (d, J = 1.4 Hz), 128.5, 91.0, 26.3, 8.7. The signal of the α-B-carbon was not

observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 64.50. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 29.19. HRMS(ESI): caled for C₁₈H₂₆BCIFO₄S⁺ [M + H]⁺ 403.1312; found 403.1314.

(Z)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)-2-(o-tolyl)ethene-1-sulfonyl fluoride (4t)



56% (21.4 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.27 – 7.13 (m, 4H), 7.03 – 6.97 (m, 1H), 2.19 (s, 3H), 1.78 – 1.61 (m, 8H), 0.88 (t, J = 7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 135.0, 134.0, 131.8 (d, J = 24.5 Hz), 129.8, 128.3, 126.4, 125.5, 90.6, 26.3, 19.9, 8.6. The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz,

Chloroform-*d*) δ 64.22. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 28.76. HRMS(ESI): caled for C₁₉H₂₉BFO₄S⁺ [M + H]⁺ 383.1858; found 383.1860.

(Z)-2-(4-methoxyphenyl)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)ethene-1-sulfonyl fluoride (4u)



52% (18.8 mg); yellow solid; ¹H NMR (400 MHz, Chloroform-d) δ 7.41 – 7.34 (m, 2H), 6.99 (s, 1H), 6.95 – 6.88 (m, 2H), 3.83 (s, 3H), 1.81 - 1.64 (m, J = 7.4 Hz, 8H), 0.92 (t, J = 7.4 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 160.7, 130.2 (d, *J* = 1.7 Hz), 128.6 (d, J = 25.7 Hz), 126.9, 113.6, 90.7, 55.2, 26.3,

8.7. The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 64.19. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 29.66. HRMS(ESI): caled for C₁₉H₂₉BFO₅S⁺ [M + H]⁺ 399.1808; found 399.1805.

(Z)-2-(4-phenoxyphenyl)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)ethene-1-sulfonyl fluoride (4v)



4% (20.2 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-d) δ 7.41 – 7.32 (m, 4H), 7.20 – 7.11 (m, 1H), 7.07 (dt, J = 7.7, 1.1 Hz, 2H), 7.03 (s, 1H), 7.02 - 6.94 (m, 2H), 1.83 - 1.62 (m, J = 7.4 Hz, 8H), 0.92 (t, J = 7.5 Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 158.8, 156.1, 130.1 (d, J = 1.5 Hz), 129.9, 129.4 (d, J = 25.9 Hz), 129.0, 124.0, 119.9, 117.5, 90.8, 26.3, 8.7. The signal of the α -B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-d) δ 64.31. ¹¹B NMR (128 MHz, Chloroformd) δ 29.21. HRMS(ESI): caled for C₂₄H₃₁BFO₅S⁺ [M + H]⁺ 461.1964; found 461.1967.

(Z)-2-(naphthalen-2-yl)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)ethene-1-sulfonyl fluoride (4w)



51% (21.3 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-d) δ 7.88 - 7.80 (m, 4H), 7.53 - 7.38 (m, 3H), 7.15 (s, 1H), 1.82 - 1.62 (m, 8H), 0.92 (t, J = 7.4 Hz, 12H). ¹³C NMR (101 MHz, Chloroformd) δ 133.4, 132.8, 132.4, 130.5 (d, J = 25.9 Hz), 128.6, 127.8 (d, J = 1.7 Hz), 127.7, 126.9, 126.4, 125.4, 125.3, 90.9, 26.3, 8.7. The

signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-d) δ 64.46. ¹¹B
NMR (128 MHz, Chloroform-*d*) δ 29.26. HRMS(ESI): caled for C₂₂H₂₉BFO₄S⁺ [M + H]⁺ 419.1858; found 419.1859.

(Z)-2-(4,4,5,5-tetraethyl-1,3,2-dioxaborolan-2-yl)-2-(thiophen-3-yl)ethene-1-sulfonyl

fluoride (4x)

Et Et O O B SO₂F S 43% (16.0 mg); colorless oil; ¹H NMR (500 MHz, Chloroform-*d*) δ 7.79 (dd, J = 2.9, 1.3 Hz, 1H), 7.38 (dd, J = 5.1, 1.3 Hz, 1H), 7.33 (dd, J = 5.1, 3.0Hz, 1H), 6.99 (s, 1H), 1.80 – 1.67 (m, 8H), 0.94 (t, J = 7.5 Hz, 12H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 134.5, 129.0, 128.5, 128.0 (d, J = 26.1Hz), 125.4, 90.8, 26.3, 8.8. The signal of the α-B-carbon was not observed.

¹⁹F NMR (376 MHz, Chloroform-*d*) δ 62.81. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 29.09. HRMS(ESI): caled for C₁₆H₂₅BFO₄S₂⁺ [M + H]⁺ 375.1266; found 375.1264.

4-phenyl-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)butane-1-sulfonyl fluoride (6a)



73% (25.0 mg); white solid; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.30 (t, *J* = 7.3 Hz, 2H), 7.25 – 7.12 (m, 3H), 3.63 (ddd, *J* = 14.4, 8.1, 6.0 Hz, 1H), 3.48 – 3.34 (m, 1H), 2.78 – 2.59 (m, 2H), 2.03 – 1.80 (m, 2H), 1.76 (ddd, *J* = 14.0, 8.1, 5.9 Hz, 1H), 1.30 (s, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 141.2, 128.5,

128.3, 126.1, 84.3, 52.3 (d, J = 14.6 Hz), 34.4, 31.5, 24.8 (d, J = 13.0 Hz). The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 56.00. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 33.53. HRMS(ESI): caled for C₁₆H₂₄BFO₄SNa⁺ [M + Na]⁺ 365.1364; found 365.1362.

2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-4-(p-tolyl)butane-1-sulfonyl fluoride (6b)



50% (17.8 mg); white solid; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.20 – 6.98 (m, 4H), 3.62 (ddd, *J* = 14.4, 8.2, 6.1 Hz, 1H), 3.42 (ddd, *J* = 14.6, 5.8, 2.9 Hz, 1H), 2.75 – 2.50 (m, 2H), 2.32 (s, 3H), 1.98 – 1.79 (m, 2H), 1.75 (ddd, *J* = 14.0, 8.1, 5.8 Hz, 1H), 1.29 (s, 12H). ¹³C

NMR (101 MHz, Chloroform-*d*) δ 138.1, 135.6, 129.2, 128.2, 84.3, 52.4 (d, J = 14.9 Hz), 33.9, 31.7, 24.8 (d, J = 13.3 Hz), 21.0. The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 55.88 (d, J = 8.4 Hz). ¹¹B NMR (128 MHz, Chloroform-*d*) δ 33.37. HRMS(ESI): caled for C₁₇H₂₇BFO₄S⁺ [M + H]⁺ 357.1702; found 357.1704.

4-(3,5-dimethylphenyl)-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)butane-1-sulfonyl fluoride (6c)



54% (20.0 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 6.99 – 6.71 (m, 3H), 3.62 (ddd, *J* = 14.5, 8.2, 6.1 Hz, 1H), 3.47 – 3.33 (m, 1H), 2.59 (qdd, *J* = 13.5, 10.0, 6.3 Hz, 2H), 2.29 (s, 6H), 2.00 – 1.81 (m, 2H), 1.75 (tt, *J* = 8.1, 5.8 Hz, 1H), 1.28 (s, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 141.1, 138.0, 127.8, 126.2, 84.3,

52.4 (d, J = 14.8 Hz), 34.2, 31.6, 24.8 (d, J = 13.0 Hz), 21.2. The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 55.88 (d, J = 8.9 Hz). ¹¹B NMR (128 MHz, Chloroform-*d*) δ 33.79. HRMS(ESI): caled for C₁₈H₂₉BFO₄S⁺ [M + H]⁺ 371.1858; found 371.1856.

4-([1,1'-biphenyl]-4-yl)-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)butane-1-sulfonyl fluoride (6d)



36% (15.0 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.61 – 7.49 (m, 4H), 7.46 – 7.39 (m, 2H), 7.35 – 7.30 (m, 1H), 7.25 (d, *J* = 8.2 Hz, 2H), 3.64 (ddd, *J* = 14.2, 8.0, 5.9 Hz, 1H), 3.44 (ddd, *J* = 14.7, 6.0, 2.9 Hz, 1H), 2.81 – 2.62 (m, 2H), 1.95 (dddd, *J* = 23.6, 17.3,

13.6, 7.8 Hz, 2H), 1.78 (tt, J = 8.1, 5.9 Hz, 1H), 1.29 (s, 12H). ¹³C NMR (101 MHz, Chloroform*d*) δ 141.0, 140.3, 139.2, 128.8, 128.7, 127.3, 127.1, 127.0, 84.4, 52.4 (d, J = 14.8 Hz), 34.0, 31.5, 24.8 (d, J = 12.7 Hz). The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 56.07 (d, J = 6.3 Hz). ¹¹B NMR (128 MHz, Chloroform-*d*) δ 33.74. HRMS(ESI): caled for C₂₂H₂₉BFO₄S⁺ [M + H]⁺ 419.1858; found 419.1859. 4-(4-(tert-butyl)phenyl)-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)butane-1-sulfonyl fluoride (6e)



40% (15.9 mg); colorless oil; ¹H NMR (500 MHz, Chloroform-*d*) δ 7.33 – 7.30 (m, 2H), 7.12 (d, *J* = 8.3 Hz, 2H), 3.63 (ddd, *J* = 14.5, 8.3, 6.1 Hz, 1H), 3.43 (ddd, *J* = 14.7, 5.8, 2.8 Hz, 1H), 2.74 – 2.56 (m, 2H), 1.98 – 1.82 (m, 2H), 1.76 (tt, *J* = 8.2, 5.8 Hz, 1H), 1.31 (s, 9H), 1.28 (s, 12H). ¹³C NMR (101 MHz,

Chloroform-*d*) δ 149.0, 138.1, 128.0, 125.4, 84.3, 52.4 (d, *J* = 14.9 Hz), 34.4, 33.8, 31.5, 31.4, 24.8 (d, *J* = 12.9 Hz). The signal of the α-B-carbon was not observed. ¹⁹F NMR (471 MHz, Chloroform-*d*) δ 55.84. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 33.68. HRMS(ESI): caled for C₂₀H₃₂BFO₄SNa⁺ [M + Na]⁺ 421.1990; found 421.1989.

2-cyclohexyl-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethane-1-sulfonyl fluoride (6f)



30% (9.6 mg); colorless oil; ¹H NMR (500 MHz, Chloroform-*d*) δ 3.65 (ddd, *J* = 14.6, 10.0, 7.2 Hz, 1H), 3.40 (ddd, *J* = 14.5, 3.9, 2.2 Hz, 1H), 1.82 – 1.61 (m, 5H), 1.29 (d, *J* = 2.1 Hz, 12H), 1.17 – 1.05 (m, 3H), 0.97 – 0.85 (m, 4H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 84.2, 51.2 (d, *J* = 15.0 Hz), 39.3, 32.2, 31.3, 26.5, 26.3 (d, *J* = 13.3 Hz), 24.9 (d,

J = 25.5 Hz). The signal of the α-B-carbon was not observed. ¹⁹F NMR (471 MHz, Chloroform*d*) δ 53.86. ¹¹B NMR (160 MHz, Chloroform-*d*) δ 32.62. HRMS(ESI): caled for C₁₄H₂₇BFO₄S⁺ [M + H]⁺ 321.1702; found 321.1701.

2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)hexane-1-sulfonyl fluoride (6g)



70% (20.6 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*)
δ 3.58 (ddd, J = 14.7, 8.3, 6.4 Hz, 1H), 3.37 (ddd, J = 14.7, 5.8,
2.7 Hz, 1H), 1.75 - 1.62 (m, 1H), 1.61 - 1.52 (m, 2H), 1.38 1.29 (m, 4H), 1.25 (s, 12H). 0.89 (t, J = 6.9 Hz, 3H). ¹³C NMR

(101 MHz, Chloroform-d) δ 84.2, 52.4 (d, J = 14.6 Hz), 30.2, 29.3, 24.7 (d, J = 15.5 Hz), 22.6,

13.9. The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 55.35 (d, J = 6.1 Hz). ¹¹B NMR (128 MHz, Chloroform-*d*) δ 33.33. HRMS(ESI): caled for C₁₂H₂₅BFO₄S⁺ [M + H]⁺ 295.1545; found 295.1543.

6-bromo-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)hexane-1-sulfonyl fluoride (6h)



60% (22.3 mg); colorless oil; ¹H NMR (400 MHz, Chloroformd) δ 3.60 (ddd, J = 14.7, 7.6, 5.8 Hz, 1H), 3.45 – 3.34 (m, 3H), 1.88 (ddt, J = 9.7, 8.0, 6.5 Hz, 2H), 1.75 – 1.67 (m, 1H), 1.66 – 1.50 (m, 4H), 1.26 (s, 12H). ¹³C NMR (101 MHz,

Chloroform-*d*) δ 84.4, 52.3 (d, *J* = 14.8 Hz), 33.4, 32.4, 28.7, 26.5, 24.8 (d, *J* = 14.0 Hz). The signal of the α -B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 55.89. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 33.28. HRMS(ESI): caled for C₁₂H₂₃BBrFO₄SNa⁺ [M + Na]⁺ 395.0470; found 395.0471.

3-phenyl-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)propane-1-sulfonyl fluoride (6i)



63% (20.8 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.30 (tt, J = 7.0, 1.1 Hz, 2H), 7.25 – 7.19 (m, 3H), 3.52 (ddd, J =14.5, 8.4, 6.0 Hz, 1H), 3.36 – 3.28 (m, 1H), 2.97 (dd, J = 14.0, 6.6 Hz, 1H), 2.86 – 2.75 (m, 1H), 2.11 – 1.99 (m, 1H), 1.22 (d, J = 6.8

Hz, 12H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 138.7, 129.0, 128.7, 126.8, 84.4, 51.3 (d, J = 15.2 Hz), 34.8, 24.8 (d, J = 8.0 Hz). The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 55.73. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 33.25. HRMS(ESI): caled for C₁₅H₂₃BFO₄S⁺ [M + H]⁺ 329.1389; found 329.1390.

2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)dodecane-1-sulfonyl fluoride (6j)



79% (30.0 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 3.58 (ddd, *J* = 14.8, 8.4, 6.4 Hz, 1H), 3.37 (ddd, *J* = 14.7, 5.8, 2.6 Hz, 1H), 1.75 – 1.64 (m, 1H), 1.26 (s,

29H), 0.88 (t, J = 6.8 Hz, 4H). ¹³C NMR (101 MHz, Chloroform-d) δ 84.2, 52.4 (d, J = 14.7 Hz),

31.9, 29.7, 29.6, 29.5, 29.5, 29.4, 29.3, 28.0, 24.8 (d, J = 15.6 Hz), 22.7, 14.1. The signal of the α -B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 55.33 (d, J = 9.6 Hz). ¹¹B NMR (128 MHz, Chloroform-*d*) δ 33.50. HRMS(ESI): caled for C₁₈H₃₇BFO₄S⁺ [M + H]⁺ 379.2484; found 379.2487.

2,4-bis(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)butane-1-sulfonyl fluoride (6k)



38% (15.0 mg); colorless oil; ¹H NMR (500 MHz, Chloroform-*d*) δ 3.57 (ddd, *J* = 14.9, 8.4, 6.6 Hz, 1H), 3.40 (dq, *J* = 14.2, 2.5 Hz, 1H), 1.68 (dt, *J* = 7.4, 3.5 Hz, 3H), 1.25 (s, 12H), 1.24 (s, 12H), 0.88 – 0.79 (m, 2H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 84.2, 83.2, 52.3 (d, *J* = 14.7 Hz), 24.8

(d, *J* = 18.0 Hz), 24.8, 24.2. The signal of the α-B-carbon was not observed. ¹⁹F NMR (471 MHz, Chloroform-*d*) δ 54.97. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 34.02. HRMS(ESI): caled for $C_{16}H_{32}B_2FO_6S^+$ [M + H]⁺ 393.2084; found 393.2085.

(S)-2-((1R,1's,4S,4'R)-4'-butyl-[1,1'-bi(cyclohexan)]-4-yl)-2-(4,4,5,5-tetramethyl-1,3,2dioxaborolan-2-yl)ethane-1-sulfonyl fluoride (6l)



33% (15.1 mg); white soild; ¹H NMR (500 MHz, Chloroform-*d*) δ 3.64 (ddd, *J* = 14.5, 10.2, 7.2 Hz, 1H), 3.39 (ddd, *J* = 14.6, 4.1, 2.1 Hz, 1H), 1.77 (d, *J* = 13.5 Hz, 4H), 1.73

- 1.68 (m, 2H), 1.63 (dt, J = 9.8, 4.6 Hz, 1H), 1.29 (d, J = 2.1 Hz, 12H), 1.22 - 1.11 (m, 3H), 1.03 - 0.94 (m, 6H), 0.95 - 0.83 (m, 14H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 84.2, 51.3 (d, J = 15.0 Hz), 43.3, 43.1, 39.5, 37.9, 37.2, 33.6, 32.4, 31.4, 30.1, 30.0, 29.8, 29.3, 24.9 (d, J =24.8 Hz), 23.0, 14.2. The signal of the α-B-carbon was not observed. ¹⁹F NMR (471 MHz, Chloroform-*d*) δ 53.88. ¹¹B NMR (160 MHz, Chloroform-*d*) δ 32.83. HRMS(ESI): caled for $C_{24}H_{45}BFO_4S^+$ [M + H]⁺ 459.3110; found 459.3113.

(2S)-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-2-((1R,4s)-4'-(p-tolyl)-[1,1'-

bi(cyclohexan)]-4-yl)ethane-1-sulfonyl fluoride (6m)



32% (15.6 mg); white solid; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.09 (d, *J* = 1.6 Hz, 4H), 3.63 (ddd, *J* = 14.5, 10.0, 7.1 Hz, 1H), 3.38 (ddd, *J* = 14.6, 4.2,

2.2 Hz, 1H), 2.39 (tq, J = 7.1, 3.5 Hz, 1H), 2.31 (s, 3H), 1.96 – 1.72 (m, 10H), 1.27 (s, 12H), 1.19 – 0.99 (m, 10H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 144.8, 135.2, 129.0, 126.6, 84.2, 51.2 (d, J = 15.0 Hz), 44.2, 43.0, 42.7, 39.4, 34.6, 32.4, 31.3, 30.3, 30.0, 29.8, 24.9 (d, J = 19.6 Hz), 21.0. The signal of the α -B-carbon was not observed. ¹⁹F NMR (471 MHz, Chloroform-*d*) δ 53.93. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 33.20. HRMS(ESI): caled for C₂₇H₄₃BFO₄S⁺ [M + H]⁺ 493.2954; found 493.2957.

5-(fluorosulfonyl)-4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)pentyl benzoate (6n)



35% (14.0 mg); colorless oil; ¹H NMR (400 MHz, Chloroform-*d*) δ 8.12 – 8.00 (m, 2H), 7.61 – 7.52 (m, 1H), 7.44 (dd, *J* = 8.3, 7.1 Hz, 2H), 4.33 (t, *J* = 4.9 Hz, 2H), 3.73 – 3.57 (m, 1H), 3.47 – 3.37 (m, 1H), 1.94 – 1.71 (m, 5H), 1.25 (s, 12H). ¹³C NMR (126

MHz, Chloroform-*d*) δ 166.5, 132.9, 130.3, 129.6, 128.4, 84.4, 64.4, 52.3 (d, *J* = 15.0 Hz), 27.4, 26.2, 24.8 (d, *J* = 14.8 Hz). The signal of the α-B-carbon was not observed. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 56.11. ¹¹B NMR (128 MHz, Chloroform-*d*) δ 33.37. HRMS(ESI): caled for C₁₈H₂₇BFO₆S⁺ [M + H]⁺ 401.1600; found 401.1603.

(E)-2-bromo-2-phenylethene-1-sulfonyl fluoride (7)



(E)-2-chloro-2-phenylethene-1-sulfonyl fluoride (8)



73% (8.0 mg); colorless oil; ¹H NMR (500 MHz, Chloroform-d) δ 7.53 (dg, J = 6.4, 1.4 Hz, 3H), 7.51 – 7.44 (m, 2H), 6.90 (s, 1H). ¹³C NMR (126) MHz, Chloroform-d) δ 155.4, 133.5, 132.0, 128.6, 128.4, 120.4 (d, J = 31.1 Hz). ¹⁹F NMR (471 MHz, Chloroform-*d*) δ 67.08.

All data matched that reported in the literature.^[10]

(Z)-2-phenylethene-1-sulfonyl fluoride (9)

54% (5.0 mg); white solids; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.60 (dd, н J = 7.6, 2.0 Hz, 2H), 7.50 – 7.42 (m, 3H), 7.38 (dd, J = 11.9, 5.8 Hz, 1H), **SO₂F** 6.51 (dd, J = 11.9, 2.6 Hz, 1H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 146.7, 131.3, 130.1 (d, J = 1.8 Hz), 128.7, 120.3 (d, J = 28.5 Hz). ¹⁹F NMR (376 MHz,

Chloroform-*d*) δ 64.00.

All data matched that reported in the literature.^[3]

(Z)-2-(3,6-dihydro-2H-pyran-4-yl)-2-phenylethene-1-sulfonyl fluoride (11)



60% (8.1 mg); light yellow solids; ¹H NMR (400 MHz, Chloroform-d) δ 7.47 - 7.40 (m, 3H), 7.24 - 7.17 (m, 2H), 6.46 (s, 1H), 5.84 (s, 1H), 4.24 (d, J = 2.7 Hz, 2H), 3.90 (t, J = 5.5 Hz, 2H), 2.39 (dddd, J = 6.7, 4.0, 2.5)1.1 Hz, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 159.2, 139.4, 133.9, 133.2, 129.3, 128.7, 128.1, 116.7 (d, *J* = 27.5 Hz), 66.1, 63.7, 25.5. ¹⁹F

NMR (376 MHz, Chloroform-d) δ 68.89. HRMS(ESI): caled for C₁₃H₁₃FO₃SNa⁺ [M + Na]⁺ 291.0461; found 291.0465.

2,2-diphenylethene-1-sulfonyl fluoride (13)



61% (8.0 mg); white solids; ¹H NMR (400 MHz, Chloroform-d) δ 7.53 -7.44 (m, 4H), 7.40 (dd, *J* = 8.6, 6.9 Hz, 2H), 7.32 (ddd, *J* = 8.7, 6.9, 1.5 Hz, 4H), 6.84 (s, 1H). ¹³C NMR (101 MHz, Chloroform-d) δ 161.3 (d, J = 3.9 Hz), 138.1 (d, J = 2.2 Hz), 135.2, 131.5, 130.2, 129.3, 128.9, 128.8, 128.4, 117.7 (d, J = 28.1 Hz). ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 68.08.

S42

All data matched that reported in the literature.^[3]

benzo[d][1,3]dioxol-5-yl 2,2-diphenylethene-1-sulfonate (14)



89% (17.0 mg); Light yellow solids; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.48 – 7.33 (m, 6H), 7.28 – 7.21 (m, 4H), 6.81 (s, 1H), 6.74 (d, *J* = 8.3 Hz, 1H), 6.71 – 6.61 (m, 2H), 5.98 (s, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 158.5, 148.2, 146.5, 143.4, 139.0, 135.7, 130.8, 129.5, 129.5,

128.8, 128.6, 128.0, 120.4, 115.2, 108.0, 104.6, 102.0. HRMS(ESI): caled for C₂₁H₁₆O₅SNa⁺ [M + Na]⁺ 403.0610; found 403.0608.

4-((2,2-diphenylvinyl)sulfonyl)morpholine (15)



94% (15.6 mg); white solids; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.43 – 7.36 (m, 5H), 7.35 – 7.31 (m, 3H), 7.27 – 7.22 (m, 2H), 6.63 (s, 1H), 3.89 – 3.50 (m, 4H), 3.35 – 2.84 (m, 4H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 155.5, 139.8, 136.4, 130.2, 129.8, 129.1, 128.6, 128.3, 127.9, 121.9, 66.4, 45.5. HRMS(ESI): called for C₁₈H₂₀NO₃S⁺ [M + H]⁺ 330.1159; found 330.1160.

(E)-2-(2-methylquinolin-6-yl)-2-phenylethene-1-sulfonyl fluoride (17)



50% (8.2 mg); white solids; ¹H NMR (400 MHz, Chloroformd) δ 8.02 (t, J = 8.1 Hz, 2H), 7.72 – 7.61 (m, 2H), 7.57 – 7.47 (m, 3H), 7.41 – 7.32 (m, 3H), 6.97 (s, 1H), 2.77 (s, 3H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 161.5, 137.0, 135.1, 135.0,

130.3, 129.6, 129.5, 129.3, 128.5, 128.4, 126.0, 123.2, 120.8, 118.3 (d, J = 28.3 Hz), 115.2, 25.5. ¹⁹F NMR (471 MHz, Chloroform-*d*) δ 68.18. HRMS(ESI): caled for C₁₈H₁₅FNO₂S⁺ [M + H]⁺ 328.0802; found 328.0803.

(8R,9S,13S,14S)-13-methyl-17-oxo-7,8,9,11,12,13,14,15,16,17-decahydro-6H-

cyclopenta[a]phenanthren-3-yl (E)-2-(2-methylquinolin-6-yl)-2-phenylethene-1-sulfonate (18)



51% (7.4 mg); white solids; ¹H NMR (400 MHz, Chloroform-*d*) δ 8.00 (t, J = 8.4 Hz, 2H), 7.63 – 7.57 (m, 2H), 7.50 – 7.27 (m, 7H), 7.00 – 6.92 (m, 3H), 2.89 (dd, J = 9.1, 4.3 Hz, 1H), 2.77 (s, 3H), 2.51 (dd, J = 18.8, 8.7

Hz, 1H), 2.39 (d, J = 11.9 Hz, 1H), 2.28 (t, J = 10.0 Hz, 1H), 2.21 – 2.08 (m, 1H), 2.08 – 1.94 (m, 3H), 1.54 (dddd, J = 34.4, 27.9, 22.3, 11.5 Hz, 7H), 0.91 (s, 3H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 220.5, 161.0, 147.2, 138.8, 138.6, 136.9, 135.5, 129.6, 129.6, 129.3, 129.0, 128.9, 128.7, 128.5, 128.1, 126.7, 126.2, 123.0, 122.4, 121.6, 119.3, 50.4, 47.9, 44.2, 38.0, 35.8, 31.5, 29.7, 29.4, 26.2, 25.8, 25.5, 21.6, 13.8. HRMS(ESI): called for C₃₆H₃₆NO₄S⁺ [M + H]⁺ 578.2360; found 578.2364.

(E)-2-(9H-fluoren-3-yl)-2-phenylethene-1-sulfonyl fluoride (20)



54% (9.5 mg); white solids; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.87 - 7.75 (m, 2H), 7.61 - 7.46 (m, 5H), 7.45 - 7.31 (m, 5H), 6.89 (s, 1H), 3.90 (s, 2H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 161.6, 145.2, 144.0, 143.7, 140.3, 136.3, 135.5, 130.1, 129.4, 128.4, 128.1, 128.0, 127.1, 125.5, 125.2, 120.7, 120.0, 116.7 (d, J= 27.8 Hz), 36.9. ¹⁹F NMR (376 MHz, Chloroform-*d*) δ 68.69. HRMS(ESI): caled for C₂₁H₁₅FO₂SNa⁺

[M + Na]⁺ 373.0669; found 373.0666.

(E)-11-(4-((2-(9H-fluoren-3-yl)-2-phenylvinyl)sulfonyl)piperazin-1-yl)-2-

chlorodibenzo[b,f][1,4]oxazepine (21)



63% (10.0 mg); white solids; ¹H NMR (400 MHz, Chloroform-*d*) δ 7.82 – 7.75 (m, 1H), 7.72 (d, J = 8.0 Hz, 1H), 7.58 – 7.51 (m, 1H), 7.48 – 7.29 (m, 9H), 7.28 (d, J = 1.8 Hz, 1H), 7.25 (d,

J = 2.6 Hz, 1H), 7.19 (d, J = 8.7 Hz, 1H), 7.14 – 7.06 (m, 3H), 7.01 (td, J = 7.4, 2.0 Hz, 1H), 6.74 (s, 1H), 3.86 (s, 2H), 3.50 (s, 4H), 3.19 (s, 4H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 159.4, 158.4, 155.7, 151.7, 144.0, 143.9, 143.6, 140.6, 138.0, 136.8, 132.9, 130.5, 129.9, 129.2, 128.8, 128.1, 127.6, 127.5, 127.1, 127.1, 125.9, 125.2, 125.0, 124.7, 122.9, 121.7, 120.5, 120.2, 119.9, 47.2, 45.0, 36.9. HRMS(ESI): caled for C₃₈H₃₁ClN₃O₃S⁺ [M + H]⁺ 644.1769; found 644.1767.



Supplementary Figure 16. ¹³C NMR spectra of product 4a



Supplementary Figure 18. ¹¹B NMR spectra of product 4a



Supplementary Figure 20. ¹H NMR spectra of product 4b



240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 -20 -30 -40 -50 f1 (ppm)

Supplementary Figure 21. ¹³C NMR spectra of product 4b



Supplementary Figure 22. ¹⁹F NMR spectra of product 4b



Supplementary Figure 23. ¹H NMR spectra of product 4c



Supplementary Figure 24. ¹³C NMR spectra of product 4c



Supplementary Figure 25. ¹⁹F NMR spectra of product 4c



Supplementary Figure 26. ¹H NMR spectra of product 4d



Supplementary Figure 27. ¹³C NMR spectra of product 4d



Supplementary Figure 28. ¹⁹F NMR spectra of product 4d



Supplementary Figure 29. ¹H NMR spectra of product 4e



Supplementary Figure 30. ¹³C NMR spectra of product 4e



Supplementary Figure 31. ¹⁹F NMR spectra of product 4e



Supplementary Figure 32. ¹H NMR spectra of product 4f



Supplementary Figure 33. ¹³C NMR spectra of product 4f



Supplementary Figure 34. ¹⁹F NMR spectra of product 4f



Supplementary Figure 35. ¹H NMR spectra of product 4g



Supplementary Figure 36. ¹³C NMR spectra of product 4g



Supplementary Figure 37. ¹⁹F NMR spectra of product 4g



Supplementary Figure 38. ¹H NMR spectra of product 4h



Supplementary Figure 39. ¹³C NMR spectra of product 4h



Supplementary Figure 40. ¹⁹F NMR spectra of product 4h



Supplementary Figure 41. ¹H NMR spectra of product 4i



Supplementary Figure 42. ¹³C NMR spectra of product 4i



Supplementary Figure 43. ¹⁹F NMR spectra of product 4i



Supplementary Figure 44. ¹H NMR spectra of product 4j



Supplementary Figure 45. ¹³C NMR spectra of product 4j



Supplementary Figure 46. ¹⁹F NMR spectra of product 4j



Supplementary Figure 47. ¹H NMR spectra of product 4k



Supplementary Figure 48. ¹³C NMR spectra of product 4k



Supplementary Figure 49. ¹⁹F NMR spectra of product 4k



Supplementary Figure 50. ¹H NMR spectra of product 4I



Supplementary Figure 51. ¹³C NMR spectra of product 4I



Supplementary Figure 52. ¹⁹F NMR spectra of product 4I



Supplementary Figure 53. ¹H NMR spectra of product 4m



Supplementary Figure 54. ¹³C NMR spectra of product 4m



Supplementary Figure 56. ¹¹B NMR spectra of product 4m



Supplementary Figure 58. ¹H NMR spectra of product 4n



Supplementary Figure 59. ¹³C NMR spectra of product 4n



Supplementary Figure 60. ¹⁹F NMR spectra of product 4n



Supplementary Figure 61. ¹H NMR spectra of product 4o



Supplementary Figure 62. ¹³C NMR spectra of product 4o



Supplementary Figure 63. ¹⁹F NMR spectra of product 40



Supplementary Figure 64. ¹H NMR spectra of product 4p



Supplementary Figure 65. ¹³C NMR spectra of product 4p



Supplementary Figure 66. ¹⁹F NMR spectra of product 4p


Supplementary Figure 67. ¹H NMR spectra of product 4q



Supplementary Figure 68. ¹³C NMR spectra of product 4q



Supplementary Figure 69. ¹⁹F NMR spectra of product 4q



Supplementary Figure 70. ¹H NMR spectra of product 4r



Supplementary Figure 71. ¹³C NMR spectra of product 4r



Supplementary Figure 72. ¹⁹F NMR spectra of product 4r



Supplementary Figure 73. ¹H NMR spectra of product 4s



Supplementary Figure 74. ¹³C NMR spectra of product 4s



Supplementary Figure 75. ¹⁹F NMR spectra of product 4s



Supplementary Figure 76. ¹H NMR spectra of product 4t



Supplementary Figure 77. ¹³C NMR spectra of product 4t



Supplementary Figure 78. ¹⁹F NMR spectra of product 4t



Supplementary Figure 79. ¹H NMR spectra of product 4u



Supplementary Figure 80. ¹³C NMR spectra of product 4u



Supplementary Figure 81. ¹⁹F NMR spectra of product 4u



Supplementary Figure 82. ¹H NMR spectra of product 4v







Supplementary Figure 84. ¹⁹F NMR spectra of product 4v



Supplementary Figure 85. ¹H NMR spectra of product 4w



Supplementary Figure 86. ¹³C NMR spectra of product 4w



Supplementary Figure 87. ¹⁹F NMR spectra of product 4w



Supplementary Figure 88. ¹H NMR spectra of product 4x



Supplementary Figure 89. ¹³C NMR spectra of product 4x



Supplementary Figure 90. ¹⁹F NMR spectra of product 4x



Supplementary Figure 91. ¹H NMR spectra of product 6a



Supplementary Figure 92. ¹³C NMR spectra of product 6a



Supplementary Figure 93. ¹⁹F NMR spectra of product 6a



Supplementary Figure 94. ¹H NMR spectra of product 6b



Supplementary Figure 95. ¹³C NMR spectra of product 6b



Supplementary Figure 96. ¹⁹F NMR spectra of product 6b



Supplementary Figure 97. ¹H NMR spectra of product 6c



Supplementary Figure 98. ¹³C NMR spectra of product 6c



Supplementary Figure 99. ¹⁹F NMR spectra of product 6c



Supplementary Figure 100. ¹H NMR spectra of product 6d



Supplementary Figure 101. ¹³C NMR spectra of product 6d



Supplementary Figure 102. ¹⁹F NMR spectra of product 6d



Supplementary Figure 103. ¹H NMR spectra of product 6e



Supplementary Figure 104. ¹³C NMR spectra of product 6e



Supplementary Figure 105. ¹⁹F NMR spectra of product 6e



Supplementary Figure 106. ¹H NMR spectra of product 6f



Supplementary Figure 107. ¹³C NMR spectra of product 6f



Supplementary Figure 108. ¹⁹F NMR spectra of product 6f



Supplementary Figure 109. ¹H NMR spectra of product 6g



Supplementary Figure 110. ¹³C NMR spectra of product 6g



Supplementary Figure 111. ¹⁹F NMR spectra of product 6g



Supplementary Figure 112. ¹H NMR spectra of product 6h



Supplementary Figure 113. ¹³C NMR spectra of product 6h



Supplementary Figure 114. ¹⁹F NMR spectra of product 6h



Supplementary Figure 115. ¹H NMR spectra of product 6i



Supplementary Figure 116. ¹³C NMR spectra of product 6i



Supplementary Figure 117. ¹⁹F NMR spectra of product 6i



Supplementary Figure 118. ¹H NMR spectra of product 6j



Supplementary Figure 119. ¹³C NMR spectra of product 6j



Supplementary Figure 120. ¹⁹F NMR spectra of product 6j



Supplementary Figure 121. ¹H NMR spectra of product 6k



Supplementary Figure 122. ¹³C NMR spectra of product 6k



Supplementary Figure 123. ¹⁹F NMR spectra of product 6k



Supplementary Figure 124. ¹H NMR spectra of product 6I



Supplementary Figure 125. ¹³C NMR spectra of product 6I



Supplementary Figure 126. ¹⁹F NMR spectra of product 6I



Supplementary Figure 127. ¹H NMR spectra of product 6m



Supplementary Figure 128. ¹³C NMR spectra of product 6m



Supplementary Figure 129. ¹⁹F NMR spectra of product 6m



Supplementary Figure 130. ¹H NMR spectra of product 6n



Supplementary Figure 132. ¹⁹F NMR spectra of product 6n



Supplementary Figure 133. ¹H NMR spectra of product 7





Supplementary Figure 135. ¹⁹F NMR spectra of product 7



Supplementary Figure 136. ¹H NMR spectra of product 8







Supplementary Figure 138. ¹⁹F NMR spectra of product 8




Supplementary Figure 140. ¹³C NMR spectra of product 9



Supplementary Figure 141. ¹⁹F NMR spectra of product 9



Supplementary Figure 142. ¹H NMR spectra of product 11



Supplementary Figure 143. ¹³C NMR spectra of product 11



Supplementary Figure 144. ¹⁹F NMR spectra of product 11



Supplementary Figure 145. ¹H NMR spectra of product 13



Supplementary Figure 146. ¹³C NMR spectra of product 13



Supplementary Figure 147. ¹⁹F NMR spectra of product 13



Supplementary Figure 148. ¹H NMR spectra of product 14



Supplementary Figure 149. ¹³C NMR spectra of product 14



Supplementary Figure 150. ¹H NMR spectra of product 15



Supplementary Figure 151. ¹³C NMR spectra of product 15



Supplementary Figure 152. ¹H NMR spectra of product 17



Supplementary Figure 153. ¹³C NMR spectra of product 17



Supplementary Figure 154. ¹⁹F NMR spectra of product 17



Supplementary Figure 155. ¹H NMR spectra of product 18



Supplementary Figure 156. ¹³C NMR spectra of product 18



Supplementary Figure 157. ¹H NMR spectra of product 20



Supplementary Figure 158. ¹³C NMR spectra of product 20



Supplementary Figure 159. ¹⁹F NMR spectra of product 20



Supplementary Figure 160. ¹H NMR spectra of product 21



Supplementary Figure 161. ¹³C NMR spectra of product 21