# Chemical and Linguistic Considerations for Encoding Chinese Characters: An Embodiment Using Chain-End Degradable SequenceDefined Oligourethanes Created by Consecutive Solid Phase Click Chemistry 

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## I. General procedure and equipment

All materials used in the synthesis of each compound and related tests, were purchased from SigmaAldrich Chemical Co., Acros Organics, Tokyo Chemical Industry, Chem Impex International, etc. and used without further purification. Solvents (DCM, NMP, chloroform, DMSO, DMF, MeOH, MeCN, Isopropanol) were of reagent grade or HPLC grade quality and purchased from Fischer Scientific. NMR solvents ( $\mathrm{CDCl}_{3}-d$, MeOD- $d_{4}$ ) were purchased from Cambridge Isotope Laboratories.
Column chromatography was performed using silica gel 60 ( $230 \pm 400$ mesh, $0.040 \pm 0.063 \mathrm{~mm}$ ) from Dynamic Adsorbents.

TLC analyses were carried out using Silica TLC Plates Glass Backing 20 by 20 cm sheet UV active at 254 nm .

Reverse phase column chromatography and HPLC purifications were performed on Shimadzu Prominence HPLC system equipped with Zorbax SB-C18 preparatory column ( $21.2 \times 250 \mathrm{~mm}$ ) with 7.0 $\mu \mathrm{m}$ packing material. Analytical HPLC traces were also carried out using a Zorbax SB-C18 analytical column ( $4.6 \times 250 \mathrm{~mm}$ ) with $5.0 \mu \mathrm{~m}$ packing material. $5-95 \%$ gradient elution ( $\mathrm{MeCN} / \mathrm{H}_{2} \mathrm{O}$ with $0.1 \%$ formic acid). Hydrophobic urethanes utilized $30-95 \%$ gradient elution (MeCN/H2O with $0.1 \%$ formic acid).
${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ spectra were recorded on Bruker AVIII with a BBFO Prodigy liquid nitrogen CryoProbe 600 MHz NMR spectrometers. The NMR spectra were referenced to solvent and the spectroscopic solvents were purchased from Cambridge Isotope Laboratories.
Liquid Chromatography/Mass spectra were recorded on an Agilent Technologies 6120 Single Quadrupole or 6125B Single Quadrupole mass spectrometer interfaced with an Agilent 1200 series liquid chromatography system equipped with a diode-array detector. Column: Agilent ZORBAX Eclipse Plus S2 C18 narrow bore column; 2.1 mm internal diameter; 50 mm length; 5 micron particle size; P.N. 959746902. Resulting spectra were analysed using Agilent LC/MSD ChemStation. Separations were achieved
with a gradient elution from 5 to $95 \%$ organic, using MeCN and Water w/ 50 mM ammonium acetate as the eluents. High resolution mass spectrometry was performed by the UT-Austin Mass Spectrometry Facility using an Agilent Technologies 6530 Accurate-Mass Q-TOF (G6530A) with an Agilent Technologies Jet Stream ESI source, interfaced with an Agilent Technologies 1260 Infinity liquid chromatography system (G1312B). An Agilent Technologies 6546 Accurate-Mass Q-TOF (G6546A) with an Agilent Technologies Dual Jet Stream ESI source, interfaced with an Agilent Technologies 1260 Infinity II liquid chromatography system (G7112B), was used.

## II. Synthesis and characterization

## II(a). Synthesis and characterization of the monomer



To a stirred solution of Fmoc-L-azidolysine ( $1.972 \mathrm{~g}, 5 \mathrm{mmol}$ ) in anhydrous THF ( 19 mL ) was added N,Ncarbonyldiimidazole ( $1.08 \mathrm{~g}, 6.7 \mathrm{mmol}$ ) at room temperature. The reaction stirred for at least 10 minutes and was then cooled to $0^{\circ} \mathrm{C}$. Next, a solution of $\mathrm{NaBH}_{4}(311.7 \mathrm{mg}, 8.24 \mathrm{mmol})$ in $\mathrm{H}_{2} \mathrm{O}(8.32 \mathrm{~mL})$ was added. The solution was stirred for at least 30 minutes, up to 1.5 hours. The reaction was quenched by addition of 1 M HCl and extracted with EtOAc ( $3 \times 65 \mathrm{~mL}$ ). The combined organics were washed $1 \times$ with brine, dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$, and concentrated under vacuum. The crude product was purified by silica gel chromatography ( $1: 1$ Hexanes:EtOAc) to furnish a white solid ( $1.69 \mathrm{~g}, 89 \%$ ). ${ }^{1} \mathrm{H}$ NMR ( 600 MHz , MeOD$\left.d_{4}\right) \delta 7.79(\mathrm{~d}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 7.66(\mathrm{~d}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 7.38(\mathrm{t}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 7.30(\mathrm{t}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 4.40-$ $4.43(\mathrm{~m}, 1 \mathrm{H}), 4.30-4.34(\mathrm{~m}, 1 \mathrm{H}), 4.21(\mathrm{t}, \mathrm{J}=6 \mathrm{~Hz}, 1 \mathrm{H}), 3.56-3.59(\mathrm{~m}, 1 \mathrm{H}), 3.43-3.50(\mathrm{~m}, 2 \mathrm{H}), 3.27(\mathrm{t}, J=6$ $\mathrm{Hz}, 2 \mathrm{H})$, , 1.55-1.64 (m, 3H), 1.35-1.49 (m, 3H). ${ }^{13} \mathrm{C}$ NMR ( 125 MHz , MeOD- $\mathrm{d}_{4}$ ) ס 159.13, 145.70, 145.62, 142.93, 129.05, 128.42, 126.52, 126.47, 121.21, 67.82, 65.70, 54.45, 52.65, 49.83, 32.02, 30.03, 24.56. HRMS +ESI: calculated $\left(\mathrm{C}_{21} \mathrm{H}_{24} \mathrm{~N}_{4} \mathrm{O}_{3} \mathrm{Na}^{+}\right)$403.1746, found 403.1741. [ ${ }^{+}+\mathrm{Na}$ ].


To a stirring solution of $\mathbf{1}(1.69 \mathrm{~g}, 4.45 \mathrm{mmol})$ in anhydrous DCM $(22 \mathrm{~mL})$ was added pyridine ( $378 \mu \mathrm{~L}$, 4.67 mmol ) dropwise. Next, 4-nitrophenyl chloroformate ( $1.08 \mathrm{~g}, 6.67 \mathrm{mmol}$ ) was added, and the reaction left to stir overnight. Reaction was monitored by TLC ( $2: 1$ Hexanes:EtOAc) and upon consumption of the starting material, was diluted excessively in DCM and transferred to a separatory funnel. The organic layer was washed with $1 \mathrm{M} \mathrm{NaHSO} 4(2 x)$, then $1 \mathrm{M} \mathrm{Na}_{2} \mathrm{CO}_{3}(3 x)$, and finally brine. The organic layer was dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$, filtered, and concentrated in vacuo. The product was purified by silica gel chromatography ( $2: 1$ Hexanes:EtOAc) and isolated as a white solid ( $1.75 \mathrm{~g}, 3.20 \mathrm{mmol}$ ). ${ }^{1} \mathrm{H}$ NMR ( 600 $\left.\mathrm{MHz}, \mathrm{CDCl}_{3}-\mathrm{d}\right) \delta 8.25(\mathrm{~d}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 7.78(\mathrm{~d}, J=6 \mathrm{~Hz}, 2 \mathrm{H}), 7.59(\mathrm{~d}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 7.41(\mathrm{t}, J=12 \mathrm{~Hz}$, $2 \mathrm{H}), 7.36(\mathrm{~d}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 7.31(\mathrm{t}, J=12 \mathrm{~Hz}, 2 \mathrm{H}), 4.82(\mathrm{~d}, J=6 \mathrm{~Hz}, 1 \mathrm{H}), 4.43-4.50(\mathrm{~m}, 2 \mathrm{H}), 4.31-4.34$ $(\mathrm{m}, 1 \mathrm{H}), 4.21-4.26(\mathrm{~m}, 2 \mathrm{H}), 3.99-4.05(\mathrm{~m}, 1 \mathrm{H}), 3.30(\mathrm{t}, \mathrm{J}=6 \mathrm{~Hz}, 2 \mathrm{H}), 1.44-1.67(\mathrm{~m}, 6 \mathrm{H}) .{ }^{13} \mathrm{C}$ NMR ( 125 $\mathrm{MHz}, \mathrm{CDCl}_{3}-$ d) $\delta 156.17,155.50$, 152.62, 145.60, 143.92, 143.88, 141.50, 127.93, 127.22, 125.50, 125.11, 125.08, 121.89, 120.20, 70.78, 66.89, 51.26, 50.02, 47.40, 31.05, 28.64, 23.19. HRMS +ESI: calculated $\left(\mathrm{C}_{28} \mathrm{H}_{2} \mathrm{~N}_{5} \mathrm{O}_{7} \mathrm{Na}^{+}\right) 568.1808$, found 568.1799. [ $\mathrm{M}^{+} \mathrm{Na}$ ].





## II(b). General procedure for the solid-supported synthesis of oligourethanes

Coupling: Phenyalaninol loaded ( $0.44 \mathrm{mmol} / \mathrm{gram}, 200-400 \mathrm{mesh}$ ) 2-chlorotrityl polystyrene resin $(40 \mathrm{mg}$, 0.0176 mmol ) was added to a small fritted solid phase synthesis apparatus ( 5 mL ). The apparatus was then evacuated and backfilled with argon. The resin was suspended in 0.5 mL of anhydrous N -methyl-2pyrrolidinone (NMP) and left to swell for 10 minutes. Next, Hunig's base ( $4.4 \mu \mathrm{~L}, 0.025 \mathrm{mmol}$ ) was added, followed by hydroxybenzotriazole ( $13.4 \mathrm{mg}, 0.10 \mathrm{mmol}$ ). The suspension was swirled for 30 seconds until everything was dissolved. Finally, the activated amino alcohol $2(33.2 \mathrm{mg}, 0.05 \mathrm{mmol})$ was added. Reaction was shaken for 8 hours on a shaker at room temperature. Resin was washed with NMP ( $5 \times 3$ $\mathrm{mL})$, then $\mathrm{DCM}(5 \times 3 \mathrm{~mL})$, and finally $\mathrm{Et}_{2} \mathrm{O}(3 \times 2 \mathrm{~mL})$. Resin was dried overnight under vacuum. Test cleavages were effected with $1 \%$ TFA in DCM $(5 \times 0.2 \mathrm{~mL})$ for 20 seconds each. Coupling efficiency was checked by LC/MS. NOTE: depending on the cleavage times and amounts of TFA used, the trifluoroacetic ester (presumably on the terminal alcohol) was observed by LC/MS. This ester was readily hydrolyzed by dissolving the sample in DCM and shaking with saturated $\mathrm{NaHCO}_{3}$.
Solid phase CuAAC click chemistry: To the oligourethanes on the resin (about 40 mg resin, 0.0176 mmol ) in the 5 mL fritted solid phase synthesis apparatus, a terminal alkyne ( 5.0 eq .), sodium ascorbate ( 1.0 eq.), TBTA ( 1.0 eq.) and copper iodide ( 0.5 eq.) were added along with 0.7 mL DMF. The solid phase synthesis apparatus was sealed with a septum and purged with $\mathrm{N}_{2}$ gas for 15 minutes. After degassing, parafilm was used to seal the septum on the solid phase synthesis apparatus to prevent oxygen from going into the apparatus. The reaction was placed on the shaker and agitated for 18 hours at room temperature. The resin was then collected by vacuum filtration and washed with NMP ( $5 \times 3 \mathrm{~mL}$ ), then DCM ( $5 \times 3 \mathrm{~mL}$ ), and finally $\mathrm{Et}_{2} \mathrm{O}(3 \times 2 \mathrm{~mL})$.

Deprotection: Resin loaded with terminal Fmoc-protected oligocarbamates ( 0.0176 mmol ) suspended in $20 \%$ piperidine in DMF ( 3 mL ) and shaken for 2 hours at room temperature. Cleavage of the dibenzofulvene-piperidine adduct was calculated by absorbance at 301 nm using Beer's Law. The resin was washed with DMF ( $5 \times 3 \mathrm{~mL}$ ), DCM $(5 \times 3 \mathrm{~mL})$, $\mathrm{Et}_{2} \mathrm{O}(3 \times 2 \mathrm{~mL})$, and dried overnight under vacuum.
General procedure for labelling of the terminal amine with a NBD-Fluoride (on resin): To a fritted reaction vessel was added the oligomer (dimer, trimer or tetramer) on resin ( 0.0176 mmol ). Vessel was evacuated and backfilled with argon. The resin was suspended and swelled in anhydrous DMF ( 2.4 mL ) for 5 minutes. Next, DIPEA ( $41.76 \mu \mathrm{~L}, 0.24 \mathrm{mmol}$ ) was added, followed by 4 -fluoro-7-nitrobenzofurazan ( $33 \mathrm{mg}, 0.179 \mathrm{mmol}$ ). Reaction was left to shake overnight. Resin was washed with DMF ( $5 \times 16 \mathrm{~mL}$ ), DCM $(5 \times 16 \mathrm{~mL})$, and $\mathrm{Et}_{2} \mathrm{O}(3 \times 8 \mathrm{~mL})$.
Cleavage procedure: Cleavages were effected with $1 \%$ TFA in DCM $(5 \times 1 \mathrm{~mL})$ for 20 seconds each at room temperature. Resin was filtered off, and cleaved product was concentrated. Oligomer was purified by reverse-phase preparatory HPLC ( $5-95 \% \mathrm{MeCN} / \mathrm{H}_{2} \mathrm{O}, 0.1 \%$ formic acid gradient elution).

## II(c). Determination of Conversions of Synthesized Oligourethanes



Oligomer 1: \#6027


Oligomer 2: \#8fd1


Oligomer 3: *BDRW


Oligomer 5: \#9060


Oligomer Z1: *YI


Oligomer 4: \#7fd2


Oligomer 6: *76f8


Oligomer Z2: *YT


Oligomer Z3: *UMC


Oligomer Z5: *FLVV


Oligomer Z4: *PDW


Oligomer Z6_1: \#4e5f

Table 1. The conversions of 12 synthesized oligourethanes

| Oligourethane | Conversion $^{\text {a }}$ | Oligourethane | Conversion $^{\text {a }}$ |
| :---: | :---: | :---: | :---: |
| 1 | $38 \%$ | Z1 | $80 \%$ |
| 2 | $81 \%$ | Z2 | $90 \%$ |
| 3 | $63 \%$ | Z3 | $83 \%$ |
| 4 | $77 \%$ | Z4 | $65 \%$ |
| 5 | $55 \%$ | Z5 | $65 \%$ |
| 6 | $48 \%$ | Z6 | $68 \%$ |

${ }^{\text {a }}$ Conversions were determined by the ratio of the peak area of product to the total peak area in LC trace.

## III. Sequencing experiments

## III(a). General procedure for self-degradation (sequencing)

The sequencing procedure is adapted from previous work. ${ }^{1}$ The oligomer (measured to be at a final concentration between $0.5-1 \mathrm{mM}$ ) was dissolved in methanol and added to a vial. Next, potassium phosphate tribasic monohydrate was dissolved in water and then added to the reaction solution. The final concentration of base was approximately 30 mM . The final ratio of methanol and water was $1: 2.5$, respectively. Before placing the vial on the heated shaker, the reaction was sampled for LC/MS by taking $50 \mu \mathrm{~L}$ of the reaction mixture and diluted into $50 \mu \mathrm{~L}$ of a 1:1 methanol: water mixture. The reaction was ramped quickly to $70{ }^{\circ} \mathrm{C}$ in the heated shaker and held at the temperature. The reaction was sampled every 60 minutes for 240 minutes.
III(b). Oligomer Sequencing Data:
Oligomer 1: \#6027






After 120 minutes of sequencing, all the following molecules are observed in the solution and analyzed via High-Res MS.








Chemical Formula: $\mathrm{C}_{4} \mathrm{H}_{7} \mathrm{NO}_{2}$
Exact Mass: 101.05
\#



MS Zoomed Spectrum


6

Chemical Formula: $\mathrm{C}_{12} \mathrm{H}_{21} \mathrm{~N}_{5} \mathrm{O}_{2}$
Exact Mass: 267.17

| MS Spectrum Peak List |
| :--- |
| Obs. $\mathbf{m / z} \mathbf{z}$ Calc. $\mathbf{m} / \mathbf{z}$ Charge Abundance Formula Ion Species Tgt Mass Error (ppm) <br> 268.1767 268.1768 1 1189627 C 12 H 21 N 502 $(\mathrm{M}+\mathrm{H})+$ 0.35 <br> 269.1796 269.1794 1 172121 C 12 H 21 N 502 $(\mathrm{M}+\mathrm{H})+$ -0.64 <br> 270.1816 270.1818 1 16955 C 12 H 21 N 502 $(\mathrm{M}+\mathrm{H})+$ 0.49 <br> 271.1826 271.1841 1 1973 C 12 H 21 N 502 $(\mathrm{M}+\mathrm{H})+$ 5.51 |



Chemical Formula: $\mathrm{C}_{14} \mathrm{H}_{22} \mathrm{~N}_{4} \mathrm{O}_{2}$
Exact Mass: 278.17

MS Zoomed Spectrum



Oligomer 2: \#8fd1



After 120 minutes of sequencing, all the following molecules are observed in the solution and analyzed via High-Res MS.

\#8fd1
Chemical Formula: $\mathrm{C}_{65} \mathrm{H}_{93} \mathrm{~N}_{21} \mathrm{O}_{13}$ Exact Mass: 1375.73


8fd1
Chemical Formula: $\mathrm{C}_{61} \mathrm{H}_{86} \mathrm{~N}_{20} \mathrm{O}_{11}$ Exact Mass: 1274.68
 Exact Mass: 965.50

mical Formula: $\mathrm{C}_{32} \mathrm{H}_{4} \mathrm{~N}_{11} \mathrm{O}_{6}$ Exact Mass: 679.36
$\times 106$ Cpd 3: C65 H93 N21 013: +ESI Scan ( 0.22 -0.52 min. 19 Scans) Frag=180.0V MSF22-1488(LZ_2_120min)_hrESlpos2.d Subtrat




Chemical Formula: $\mathrm{C}_{17} \mathrm{H}_{21} \mathrm{~N}_{7} \mathrm{O}_{4}$
Exact Mass: 387.17





Chemical Formula: $\mathrm{C}_{15} \mathrm{H}_{18} \mathrm{~N}_{4} \mathrm{O}_{2}$ Exact Mass: 286.14



Chemical Formula: $\mathrm{C}_{15} \mathrm{H}_{24} \mathrm{~N}_{4} \mathrm{O}_{2}$ Exact Mass: 292.19

Oligomer 3: \#bdrw





After 120 minutes of sequencing, all the following molecules are observed in the solution and analyzed via High-Res MS.





 Exact Mass: 423.17



Exact Mass: 266.17
 Exact Mass: 743.27








Chemical Formula: $\mathrm{C}_{4} \mathrm{H}_{7} \mathrm{NO}_{2}$
Exact Mass: 101.05

## \#



Spectrum Peaks

|  |  |  |  |
| :---: | ---: | :---: | :---: |
| Obs. $\mathbf{m / z}$ | Calc. $\mathbf{m / z}$ | Charge | Abund |
| 102.0549 |  |  |  |
| 103.0584 | 102.0550 | 1 | 30805 |
| 104.0591 | 103.5579 | 1 | 3115 |
|  | 104.0595 | 1 | 236 |
|  |  |  | MassHunter Qual $\mathbf{1 0 . 0}$ |

MassHunter Qual 10.0
(End of Report)


Chemical Formula: $\mathrm{C}_{15} \mathrm{H}_{17} \mathrm{ClN}_{4} \mathrm{O}_{2}$
Exact Mass: 320.10



After 120 minutes of sequencing, all the following molecules are observed in the solution and analyzed via High-Res MS.











Chemical Formula: $\mathrm{C}_{15} \mathrm{H}_{18} \mathrm{~N}_{4} \mathrm{O}_{2}$ Exact Mass: 286.14


Spectrum Peaks

| Obs. $\mathbf{m} / \mathbf{z}$ | Calc. $\mathbf{m} / \mathbf{z}$ | Charge |
| :---: | ---: | :---: |
| 287.1506 | 287.1503 | 1 |
| 288.1536 | 288.1531 | 1 |
| 289.1562 | 289.1557 | 1 |
| 290.1586 | 290.1581 | 1 |
| 291.1608 | 291.1606 | 1 |


| Abund | Formula | Ion Species |
| :---: | :---: | :---: |
| 3714401 | C15H18N402 | (M+H)+ |
| 685425 | C15H18N402 | (M+H)+ |
| 75108 | C15H18N402 | (M+H)+ |
| 5989 | C15H18N402 | $(\mathrm{M}+\mathrm{H})+$ |
| 445 | C15H18N402 | $(\mathrm{M}+\mathrm{H})+$ |

(End of Report)


Chemical Formula: $\mathrm{C}_{15} \mathrm{H}_{24} \mathrm{~N}_{4} \mathrm{O}_{2}$
Exact Mass: 292.19




Exact Mass: 1377.74



After 120 minutes of sequencing, all the following molecules are observed in the solution and analyzed via High-Res MS.


 Exact Mass: 278.17







| Obs. m/z | Calc. m/z | Charge | Abundance | Formula | Ion Species | Tgt Mass Error (ppm) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 708.8576 |  |  | 721214 |  |  |  |
| 961.5477 | 961.5479 | 1 | 455760 | C45H68N1608 | (M+H)+ | 0.18 |
| 962.5503 | 962.5506 | 1 | 257267 | C45H68N1608 | (M+H)+ | 0.29 |
| 963.5524 | 963.5532 | 1 | 74346 | C45H68N1608 | (M+H)+ | 0.84 |
| 964.5535 | 964.5558 | 1 | 17695 | C45H68N1608 | (M+H)+ | 2.33 |
| 965.5507 | 965.5582 | 1 | 3226 | C45H68N1608 | (M+H)+ | 7.82 |




Chemical Formula: $\mathrm{C}_{19} \mathrm{H}_{25} \mathrm{~N}_{7} \mathrm{O}_{4}$
Exact Mass: 415.20


Spectrum Peaks

| Obs. $\mathbf{m} / \mathbf{z}$ | Calc. $\mathbf{m} / \mathbf{z}$ | Charge |
| :---: | :---: | :---: |
| 416.2039 | 416.2041 | 1 |
| 417.2070 | 417.2068 | 1 |
| 418.2088 | 418.2092 | 1 |
| 481.2778 |  |  |




Chemical Formula: $\mathrm{C}_{4} \mathrm{H}_{7} \mathrm{NO}_{2}$
Exact Mass: 101.05 \#


| Spectrum Peaks |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Obs. m/z | Calc. m/z | Charge | Abund | Formula | Ion Species | Tgt Mass Error (PPM) |
| 102.0549 | 102.0550 | 1 | 24958 | C4H7NO2 | (M+H)+ | -0.36 |
| 103.0582 | 103.0579 | 1 | 2557 | C4H7NO2 | (M+H)+ | 2.59 |
| 104.0590 | 104.0595 | 1 | 363 | C4H7NO2 | (M+H)+ | -5.09 |
| 922.0036 |  |  | 37361 |  |  |  |
|  | Masshunter Qual(End of Report) |  |  |  |  |  |



Chemical Formula: $\mathrm{C}_{16} \mathrm{H}_{20} \mathrm{~N}_{4} \mathrm{O}_{3}$ Exact Mass: 316.15



## Oligomer 6_1: *76f8







After 120 minutes of sequencing, all the following molecules are observed in the solution and analyzed via High-Res MS.






Exact Mass: 177.08

emical Formula: $\mathrm{C}_{34} \mathrm{H}_{44} \mathrm{~N}_{12} \mathrm{O}$
Exact Mass: 732.35






Chemical Formula: $\mathrm{C}_{12} \mathrm{H}_{21} \mathrm{~N}_{5} \mathrm{O}_{2}$
Exact Mass: 267.17


Oligomer 6_2: *76f8




Oligomer Z1_1: *YI


Exact Mass: 1007.34




After 60 minutes of sequencing, all the following molecules are observed in the solution and analyzed via High-Res MS.





Chemical Formula: $\mathrm{C}_{47} \mathrm{H}_{54} \mathrm{BrN}_{13} \mathrm{O}_{8}$ Exact Mass: 1007.34

MS Zoomed Spectrum


| $\begin{aligned} & \text { MS Spectrum } \\ & \hline \text { Obs. m/z } \end{aligned}$ | Calc. $\mathrm{m} / \mathrm{z}$ | Charge | Abundance | Formula | Ion Species | Tgt Mass Error (ppm) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 330.1927 |  |  | 1631671 |  |  |  |
| 1008.3471 | 1008.3474 | 1 | 139751 | C47H54BrN1308 | (M+H)+ | 0.32 |
| 1009.3488 | 1009.3503 | 1 | 84139 | C47H54BrN1308 | (M+H)+ | 1.51 |
| 1010.3466 | 1010.3466 | 1 | 164479 | C47H54BrN1308 | (M+H)+ | -0.02 |
| 1011.3490 | 1011.3487 | 1 | 80795 | C47H54BrN1308 | (M+H)+ | -0.25 |
| 1012.3585 | 1012.3512 | 1 | 23911 | C47H54BrN1308 | (M+H)+ | -7.14 |
| 1013.3574 | 1013.3538 | 1 | 5160 | C47H54BrN1308 | (M+H)+ | -3.59 |



Chemical Formula: $\mathrm{C}_{37} \mathrm{H}_{43} \mathrm{BrN}_{12} \mathrm{O}_{6}$ Exact Mass: 830.26



Chemical Formula: $\mathrm{C}_{20} \mathrm{H}_{20} \mathrm{BrN}_{7} \mathrm{O}_{4}$ Exact Mass: 501.08


Chemical Formula: $\mathrm{C}_{10} \mathrm{H}_{11} \mathrm{NO}_{2}$ Exact Mass: 177.08
*


Chemical Formula: $\mathrm{C}_{17} \mathrm{H}_{23} \mathrm{~N}_{5} \mathrm{O}_{2}$ Exact Mass: 329.19



| MS Spectrum <br> Obs. $\mathbf{m / \mathbf { z }}$ | Calc. m/z | Charge | Abundance | Formula | Ion Species | Tgt Mass Error (ppm) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 330.1927 | 330.1925 | 1 | 1631610 | C17H23N502 | (M+H) + | -0.83 |
| 331.1958 | 331.1953 | 1 | 329494 | C17H23N502 | $(\mathrm{M}+\mathrm{H})+$ | -1.67 |
| 332.1980 | 332.1979 | 1 | 36961 | C17H23N502 | $(\mathrm{M}+\mathrm{H})+$ | -0.44 |
| 333.2098 | 333.2003 | 1 | 5255 | C17H23N5O2 | $(\mathrm{M}+\mathrm{H})+$ | -28.5 |
| 334.2105 | 334.2028 | 1 | 1033 | C17H23N5O2 | (M+H)+ | -23.2 |

Oligomer Z1_2: *YI


Exact Mass: 1007.34


Oligomer Z2: *YT

Exact Mass: 989.45


After 60 minutes of sequencing, all the following molecules are observed in the solution and analyzed via High-Res MS.


Chemical Formula: $\mathrm{C}_{49} \mathrm{H}_{59} \mathrm{~N}_{13} \mathrm{O}_{10}$
Exact Mass: 989.45 Exact Mass: 989.45




Exact Mass: 812.37


Chemical Formula: $\mathrm{C}_{17} \mathrm{H}_{23} \mathrm{~N}_{5} \mathrm{O}_{2}$ Exact Mass: 329.19






## Oligomer Z3: *umc



Exact Mass: 1156.59






After 60 minutes of sequencing, all the following molecules are observed in the solution and analyzed via High-Res MS.



Chemical Formula: $\mathrm{C}_{48} \mathrm{H}_{65} \mathrm{~N}_{15} \mathrm{O}_{8}$
Exact Mass: 979.51


Chemical Formula: $\mathrm{C}_{16} \mathrm{H}_{20} \mathrm{~N}_{4} \mathrm{O}_{2}$ Exact Mass: 300.16


Chemical Formula: $\mathrm{C}_{15} \mathrm{H}_{24} \mathrm{~N}_{4} \mathrm{O}_{2}$
Exact Mass: 292.19





Chemical Formula: $\mathrm{C}_{10} \mathrm{H}_{11} \mathrm{NO}_{2}$ Exact Mass: 177.08



| Obs. m/z | Calc. m/z | Charge | Abundance | Formula | Ion Species | Tgt Mass Error (ppm) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 301.1660 | 301.1659 |  | 452094 | C16H2ON4O2 | (M+H)+ | -0.47 |
| 302.1685 | 302.1688 | 1 | 85743 | C16H2ON4O2 | (M+H)+ | 0.99 |
| 303.1712 | 303.1714 | 1 | 9452 | C16H2ON4O2 | (M+H)+ | 0.69 |
| 304.1762 | 304.1739 | 1 | 828 | C16H2ON4O2 | (M+H)+ | -7.61 |
| 339.1218 |  |  | 1080400 |  |  |  |



## Oligomer Z4: *pdw



Exact Mass: 1182.57
*pdw


After 60 minutes of sequencing, all the following molecules are observed in the solution and analyzed via High-Res MS.




## Oligomer Z5_1: *flvv







After 60 minutes of sequencing, all the following molecules are observed in the solution and analyzed via High-Res MS.







Chemical Formula: $\mathrm{C}_{19} \mathrm{H}_{25} \mathrm{~N}_{7} \mathrm{O}_{4}$ Exact Mass: 415.20



MS Zoomed Spectrum



## Oligomer Z5_2: *flvv







Oligomer Z6_1: *4e5f


Exact Mass: 1515.71


After 60 minutes of sequencing, all the following molecules are observed in the solution and analyzed via High-Res MS.





Chemical Formula: $\mathrm{C}_{20} \mathrm{H}_{21} \mathrm{~N}_{7} \mathrm{O}_{4}$
Exact Mass: 423.17



Chemical Formula:
$\mathrm{C}_{10} \mathrm{H}_{11} \mathrm{NO}_{2}$
Exact Mass: 177.08
MS Zoomed Spectrum



Chemical Formula: $\mathrm{C}_{14} \mathrm{H}_{25} \mathrm{~N}_{5} \mathrm{O}_{2}$ Exact Mass: 295.20

MS Zoomed Spectrum




Chemical Formula: $\mathrm{C}_{16} \mathrm{H}_{20} \mathrm{~N}_{4} \mathrm{O}_{2}$ Exact Mass: 300.16



Chemical Formula: $\mathrm{C}_{15} \mathrm{H}_{17} \mathrm{ClN}_{4} \mathrm{O}_{2}$
Exact Mass: 320.10


| MS Spectrum <br> Obs. $\mathrm{m} / \mathrm{z}$ | $\frac{\text { Peak List }}{\text { Calc. } \mathrm{m} / \mathrm{z}}$ | Charge | Abundance | Formula | Ion Species | Tgt Mass Error (ppm) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 296.2074 |  |  | 2117257 |  |  |  |
| 343.0932 | 343.0932 | 1 | 19791 | C15H17CIN402 | (M+Na)+ | 0.01 |
| 344.0967 | 344.0961 | 1 | 3716 | C15H17CIN4O2 | (M+Na)+ | -1.85 |
| 345.0903 | 345.0908 | 1 | 6641 | C15H17CIN402 | (M+Na)+ | 1.29 |
| 346.0925 | 346.0934 | 1 | 1370 | C15H17CIN402 | (M+Na)+ | 2.61 |

Oligomer Z6_2: *4e5f






## References

1. Dahlhauser, S. D.; Escamilla, P. R.; Vandewalle, A. N.; York, J. T.; Rapagnani, R. M.; Shei, J. S.; Glass, S. A.; Coronado, J. N.; Moor, S. R.; Saunders, D. P.; Anslyn, E. V. J. Am. Chem. Soc. 2020, 142 (6), 2744-2749.

III(c). Sequencing the oligomers with overlapping truncated oligomers of similar polarities in LC



The sequencing of Oligomer 4 (\#7fd2) is a scenario where multiple peaks overlap. In this case, it can be difficult to identify what peaks are overlapping, and what is the order in which these peaks show up in LCMS. The MS data can help in deconvolute that. The process is shown below:

At 0 min , there is only one peak in the LC trace and the corresponding MS for that peak is shown below:


At 60 min , there are two peaks in the LC trace, and the corresponding MS for these peaks are shown below:


At 120 min , there are three peaks in the LC trace, and the corresponding MS for these peaks are shown below:


As we keep observing the LC trace and the corresponding MS peaks for the 180 and 240 minutes, we can confidently say what peaks are overlapping and what is the order of their appearance and disappearance. From this observation, the accurate sequence of the given oligomer can be determined.

III（d）．The oligomers and corresponding Chinese characters．


6027
性


8fd1
近

$7 f d 2$
習


9060
遠


7648
相


4e5f
也



也
（1）Unicode encoding

UMC
性


YT

(2) Zhengma encoding

III(e). The zoomed in mass spectra of Oligomer Z5 (*flvv) and Z6 (* 4 e 5 f ).


## IV. User manual for Python scripts

To use this program, download the zip file, which can be located by clicking on "Code" on the GitHub website.


## - To convert from Chinese characters to Zhengma codes

1. Go to the "code" folder.

| $\square$ code | Added all Zheng Ma code and data files current as of 2023/06/21 | 3 weeks ago |
| :---: | :---: | :---: |
| - data | Added all Zheng Ma code and data files current as of 2023/06/21 | 3 weeks ago |
| $\square$ drafts | Added all Zheng Ma code and data files current as of 2023/06/21 | 3 weeks ago |
| $\square \mathrm{img}$ | Added all Zheng Ma code and data files current as of 2023/06/21 | 3 weeks ago |
| $\square$ raw | Added all Zheng Ma code and data files current as of 2023/06/21 | 3 weeks ago |
| [) .gitignore | Added MacOS file .DS_Store to .gitignore | 3 weeks ago |
| (1) README.md | Update README.md | 3 weeks ago |

2. Open the "4_converter.ipynb" file.

| Name | Last commit message | Last commit date |
| :---: | :---: | :---: |
| －．． |  |  |
| ［．1＿background．ipynb | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| ［0．2＿data．ipynb | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| ［］3＿tests．ipynb | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| ［］4＿converter．ipynb | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| ［ zm＿helpers．py | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |

## 3．Load and run the codes $1-4$ shown below．

```
In [1]: # If running in Google colab
#from google.colab import drive
#drive.mount('/content/gdrive')
##
#path_prefix = "/content/gdrive/My Drive/CoLab Notebooks/zhengma/raw/"
#data_prefix = '/content/gdrive/My Drive/Colab Notebooks/zhengma/data/'
In [2]: # If running on local system
path_prefix = "../raw/"
data_prefix = '../data/'
In [3]: import pickle
# Load pickle
with open(data_prefix + 'df_zm_merged.pkl', 'rb') as pickle_file:
    df_zm_merged = pickle.load(pickle_file)
In [4]: def characters_to_codes_simplistic(cjk_string, zm_dataframe, db_column='RIME Characters', zm_column='ZM Codes'):
    # Input:
    # string of CJK characters
    # database of Zheng Ma codes as a pandas DataFrame
    # name of column to check for characters
    # name of column containing Zheng Ma codes
    # Output:
    # list (dictionary?) of Zheng Ma codes
    # - In case of multiple code correspondences, choose the longest
    characters = cjk_string.strip().replace(' ', '')
    codes = []
    for character in characters:
        # Find any rows in the desired column that have the desired character
        # Take the ZM codes in those rows as a list
        possible_codes = zm_dataframe[zm_dataframe[db_column] == character][zm_column].tolist()
            # Choose the **longest code** in that list of ZM codes
            max_code = max(possible_codes, key=len) if possible_codes else None
            # There could be several, so order alphabetically and pick the first
            desired_codes = [c for c in possible_codes if len(c) == len(max_code)] if max_code else None
            desired_code = sorted(desired_codes)[0] if desired_codes else 'N/A: no match''
            codes.append([character, desired_code])
    return codes
```

4．In code line 5，enter the Chinese characters to be encoded，encased in quotation marks as shown in the red box．The example below reads＂Zhengma Method＂in Chinese．

5．Run code 6 to obtain the Zhengma codes that correlate to the Chinese characters．

In［6］：new＿test＿codes＿output1＝characters＿to＿codes＿simplistic（new＿test＿string1，df＿zm＿merged） print（new＿test＿codes＿output1）
［［＇郑＇，＇uagy＇］，［＇码＇，＇gxvv＇］，［＇输＇，＇heqk＇］，［＇入＇，＇oda＇］，［＇法＇，＇vbzs＇］］

## －To convert from Zhengma codes to Chinese characters

## 1．Go to the＂code＂folder．

| －code | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| :---: | :---: | :---: |
| －data | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| －drafts | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| $\square \mathrm{img}$ | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| $\square$ raw | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| ［）．gitignore | Added MacOS file ．DS＿Store to ．gitignore | 3 weeks ago |
| （1）README．md | Update README．md | 3 weeks ago |

2．Open the＂4＿converter．ipynb＂file．

| Name | Last commit message | Last commit date |
| :---: | :---: | :---: |
| ■ .. |  |  |
| ［0 1＿background．ipynb | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| ［．2＿data．ipynb | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| ［］3＿tests．ipynb | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| ［ 4＿converter．ipynb | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |
| ［ zm＿helpers．py | Added all Zheng Ma code and data files current as of 2023／06／21 | 3 weeks ago |

3．Load and run the codes 1－3 and 8 shown below．

```
In [1]: # If running in Google Colab
#from google.colab import drive
#drive.mount('/content/gdrive')
#
#path_prefix = "/content/gdrive/My Drive/Colab Notebooks/zhengma/raw/"
#data_prefix = '/content/gdrive/My Drive/Colab Notebooks/zhengma/data/'
In [2]: # If running on local system
path_prefix = "../raw/"
data_prefix = '../data/'
import pickle
# Load pickle
with open(data_prefix + 'df_zm_merged.pkl', 'rb') as pickle_file:
    df_zm_merged = pickle.load(pickle_file)
In [8]: def codes_to_characters_simplistic(code_list, zm_dataframe, db_column='RIME Characters', zm_column='ZM Codes'):
    # Inpüt:
    # list of ZM codes
    # database of Zheng Ma codes as a pandas DataFrame
    # name of column to check for characters
    # name of column containing zheng Ma codes
    # Output:
    # string of СJK characters
    # - In case of multiple character correspondences for a code, choose...
    cjk_string = ''
    for code in code_list:
        # Make sure the code is a valid ZM code:
        # - fewer than 5 letters
        # - no spaces
        if ' ' not in code:
            if len(code) < 5:
                    # Get the characters for that code
                    possible_characters = zm_dataframe[zm_dataframe[zm_column] == code][db_column].tolist()
                    # Remove any empty strings
                    viable_characters = [x for x in possible_characters if (len(x) > 0)]
                    # Add the smallest string (hopefully 1 character)
                    # ... watch out: there might be more than one minimum...
                    # ... what does min() do? return the first it finds in the list?
                    cjk_string += min(viable_characters, key=len)
            else:
                    print('Code too long: {}'.format(code))
        else:
            print('Code should not contain spaces: {}'.format(code))
    return cjk_string
```

4．In code line 10，enter the Zhengma codes to be decoded back to Chinese characters as shown in the red box below．Each Zhengma code should be encased in quotation marks as well as separated from the other Zhengma codes by commas．Then，all Zhengma codes should be encased in a pair of brackets．Running this code will return the Chinese characters correlating to the Zhengma codes．

[^0]
[^0]:    In［10］：new＿test＿string＿output1＝codes＿to＿characters＿simplistic［＇uagy＇，＇gxvv＇，＇heqk＇，＇oda＇，＇vbzs＇］，df＿zm＿merged） print（new＿test＿string＿output1）

    郑码输入法

