# Electronic Supplementary Information: Dynamic-bond-induced sticky friction tailors non-Newtonian rheology

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#### S1. Characterization techniques

- Dynamic light scattering (DLS): A dynamic light scattering instrument (Wyatt Technology's Mobius<sup>™</sup>) was used to measure the hydrodynamic radius of the synthesized thiol-functionalized silica particle and silica seed particle.
- Nuclear magnetic resonance spectroscopy: A Bruker Avance nanobay III HD 400 MHz nuclear magnetic resonance spectrometer was used to characterize the functionalized silica particles and the synthesized Michael-acceptors. The residual solvent CHCl<sub>3</sub> peak at  $\delta = 7.26$  ppm was used as a reference peak.

# S2. Characterization of the synthesized materials

• Functionalized silica particle characterization

The functionalized silica particles were characterized with <sup>1</sup>H NMR following the method reported by Crucho *et al.*<sup>1</sup>. 20 mg of silica particles was dissolved in a solution of 0.5 M NaOD in D<sub>2</sub>O. To quantify the functional group density, 35 mg of 1,3,5-trioxane was added as an internal standard. The intensity of the (ONa)<sub>3</sub>SiCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>SH at  $\delta = 2.43$  ppm was compared to that of the internal standard, giving 1.3 SH groups/nm<sup>2</sup> surface area of silica particles.

DLS was used to measure the size of synthesized silica particles suspended in water. The synthesized particle density ( $\rho = 1.92 \text{ g/mL}$ ) was measured with the bromoform ( $\rho = 2.89 \text{ g/ml}$ ) and methanol ( $\rho = 0.79 \text{ g/ml}$ ) mixture at which the particle does not sediment or float.

• Characterization of Michael-acceptors

The synthesized 1M, 1H, and 1N molecules were characterized with <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub> in Fig. S1):

 $\label{eq:R-OCH3} \mbox{(1M): } \delta = 8.23 \mbox{ (s, 1H, C=CH), 7.67 (d, 2H, Ar-H), 6.91 (d, 2H, Ar-H), 3.82 (s, 3H, Ar-OCH_3, 3.42-3.77 (m, -CH_2-CH-), 1.25 (m, -CH_3) ppm.$ 

 $\begin{array}{l} {\rm R}{\rm =-H} \; ({\bf 1H}){\rm :}\; \delta = 8.30\; ({\rm s},\, 1{\rm H},\, {\rm C}{\rm =CH}),\, 7.73\; ({\rm t},\, 3{\rm H},\, {\rm Ar}{\rm -H}),\, 7.40\; ({\rm d},\, 2{\rm H},\, {\rm Ar}{\rm -H}),\, 3.42{\rm -}3.77\; ({\rm m},\, {\rm -CH}_2{\rm -CH}{\rm -}),\\ {\rm 1.25\; ({\rm m},\, {\rm -CH}_3)\; \rm ppm}. \end{array}$ 

R=–NO<sub>2</sub> (**1N**):  $\delta$  = 8.37 (s, 1H, C=CH), 8.24 (d, 2H, Ar-H), 7.88 (d, 2H, Ar-H), 3.42–3.77 (m, -CH<sub>2</sub>-CH-), 1.25 (m, -CH<sub>3</sub>) ppm.

• Solvent viscosity

The viscosity of Michael-acceptor endcapped polymer fluids was measured using MCR301 or MCR702 rheometer (Anton Paar). In the measured shear rate range  $(1 - 100 \text{ s}^{-1})$ , the fluid viscosity exhibited Newtonian behavior.

$$\eta_{1M_{230}} = 2564 \text{ mPa} \cdot \text{s}, \ \eta_{1M_{2000}} = 579 \text{ mPa} \cdot \text{s}, \ \eta_{1M_{4000}} = 1195 \text{ mPa} \cdot \text{s}$$

$$\begin{split} \eta_{\mathbf{1H}_{230}} &= 880 \text{ mPa} \cdot \text{s}, \, \eta_{\mathbf{1H}_{2000}} = 528 \text{ mPa} \cdot \text{s}, \, \eta_{\mathbf{1H}_{4000}} = 1048 \text{ mPa} \cdot \text{s} \\ \eta_{\mathbf{1N}_{2000}} &= 980 \text{ mPa} \cdot \text{s}, \, \eta_{\mathbf{1N}_{4000}} = 1393 \text{ mPa} \cdot \text{s} \end{split}$$



**Fig. S1** <sup>1</sup>H NMR for  $1N_{4000}$ ,  $1H_{4000}$ , and  $1N_{4000}$  (top to bottom).

# S3. Prepartion of piperidinium acetate

Piperidinium acetate was prepared following the reaction in Scheme S1. 40 mmol of piperidine (Sigma-Aldrich, 99%) and 40 mmol of acetic acid (Thermo Fisher Scientific,  $\geq 99.7\%$ ) were added to a 50 mL round-bottom flask with 12 mL of chloroform. The solution was stirred with a magnetic stirrer for 30 min at room temperature. After the reaction, chloroform was removed from the product using a rotary evaporator.

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) spectrum for piperidinium acetate:  $\delta = 8.50$  (s, 2H, -C-NH<sub>2</sub><sup>+</sup>), 3.02 - 2.95 (m, 4H, N-CH<sub>2</sub>), 1.97 (d, 3H, C-C<sub>3</sub>), 1.81 - 1.71 (m, 4H, N-C-CH<sub>2</sub>-C), 1.67 - 1.58 (m, 2H, N-C-C-CH<sub>2</sub>) ppm.



Scheme S1 Synthesis of piperidinium acetate.



Fig. S2 Relative viscosity  $\eta_r = \eta/\eta_s$  for suspensions in  $1M_{2000}$  (a),  $1H_{2000}$  (b), and  $1N_{2000}$  (c) as a function of shear rate  $\dot{\gamma}$ . Closed symbols: increasing stress ramp. Open symbols: decreasing stress on the ramp's return cycle.



Fig. S3 Confirmation of steady-state viscosity. Relative viscosity  $\eta_{\rm r} = \eta/\eta_{\rm s}$  is plotted as a function of shear stress  $\tau$  for  $\phi = 0.52$  thiol functionalized silica particle suspensions in  $1M_{2000}$ . The viscosity is measured at three different equilibrating times (5 s, 30 s, and 90s per each  $\tau$ ).



Fig. S4 Viscosity as a function of time at constant shear rate  $\dot{\gamma}$  for the suspension in  $1H_{2000}$ .



**Fig. S5** The estimated fraction of bonded thiols as a function of  $K_{eq}$  for  $M_{PPG}=230$  (blue), 2000 (orange), and 4000 g/mol (green) at fixed packing fraction ( $\phi = 0.52$ ).

#### S4. The fraction of bonded thiols to form thia-Michael bonds

The equilibrium constant of thia-Michael reaction between thiol (–SH) and Michael-acceptor (–MA) is:

$$K_{\rm eq} = \frac{[\rm tMA]}{[-\rm SH] [-\rm MA]}.$$

The fraction of bounded thiol p at equilibrium suggests  $[tMA] = p[-SH]_0$  and  $[-SH] = (1 - p)[-SH]_0$ , where  $[-SH]_0$  is the initial molar concentration of thiol. Then,  $[-MA] = [-MA]_0 - [tMA] = [-MA]_0 - p[-SH]_0$ . Finally, the equilibrium constant is expressed by:

$$K_{\rm eq} = \frac{p}{(1-p)([-MA]_0 - p[-SH]_0)}$$

Using this equation, p is solved for all Michael-acceptors (Table S1). Based on this expression, we estimated the fraction of bonded thiols as a function of  $K_{eq}$  for all molecular weights in Fig. Fig. S5.

 Table S1
 The fraction of thiol reacted for the thia-Michael adduct formation.

	$M_{ m M_{PPG}} = 230 \ { m g/mol}$	$M_{\mathrm{M}_{\mathrm{PPG}}} = 2000 \mathrm{~g/mol}$	$M_{\mathrm{M}_{\mathrm{PPG}}} = 4000 \mathrm{~g/mol}$
$\mathbf{1M}_{M_{\mathrm{PPG}}}$	0.992	0.970	0.944
$1\mathbf{H}_{M_{\mathrm{PPG}}}$	0.999	0.996	0.992
$1N_{M_{\rm PPG}}$		0.999	0.999



**Fig. S6** Relative viscosity  $\eta_r$  versus shear stress  $\tau$  of  $\phi = 0.52$  thiol functionalized silica particle suspensions in different molecular weight BCAm-endcapped poly(propylene glycol)  $\mathbf{1N}_{2000}$  (a) and  $\mathbf{1N}_{4000}$  (b).



Fig. S7 Relative viscosity  $\eta/\eta_s$  as a function of shear rate  $\dot{\gamma}$  of the suspension in hydroxyterminated polypropylene glycol (2000 g/mol, PPG<sub>OH</sub>2k) and in a mixture of PPG<sub>OH</sub>2k and 2,2'-(ethylenedioxy)diethanethiol (EDDT) with a 1-to-10 molar ratio. The packing fraction of suspensions is fixed at  $\phi = 0.52$ .



Fig. S8 A Raman spectrum of thiol-functionalized silica particles suspended in a mixture hydroxyterminated polypropylene glycol (2000 g/mol, PPG<sub>OH</sub>2k) and and 2,2'-(ethylenedioxy)diethanethiol (EDDT) (1-to-10 molar ratio). The packing fraction of suspensions is  $\phi = 0.52$ . A free thiol peak clearly appears at 2570 cm<sup>-1</sup>, whereas there is limited-to-no disulfide formation from a peak at 510 cm<sup>-1</sup>.

#### S5. Constraint-based Wyart-Cates model fitting

We fitted the experimental rheology curves using the constraint-based model by Guy *et al.*<sup>2,3</sup>, which extends an earlier model by Wyart and Cates (WC) that does not account for attractive interactions<sup>4</sup>. This modified WC model includes frictional and adhesive force responsible for the sliding and rolling constraints, respectively. Based on the WC model, the relative viscosity is expressed by

$$\eta_{\rm r} = \left[1 - \frac{\phi}{\phi_{\rm J}(a,f)}\right]^{-2} \tag{1}$$

with the jamming volume fraction  $\phi_{\rm J}(a, f)$  that is a function of the fraction of adhesion (a) and friction (f). These parameters can be expressed by  $f(\tau) = \exp\left[-(\tau^*/\tau)^{\beta}\right]$  with the critical stress  $\tau^*$  for dominating friction over particle repulsive interaction and  $a(\tau) = 1 - \exp\left[-(\tau_{\rm a}/\tau)^{\kappa}\right]$  with  $\tau_{\rm a}$ , the stress required to break the adhesion between contacting particles. With a and f,  $\phi_{\rm J}(a, f)$  can be interpolated with all four possible constraints,

$$\phi_{\rm J}(a,f) = af\phi_{\rm alp} + a(1-f)\phi_{\rm acp} + (1-a)f\phi_{\mu} + (1-a)(1-f)\phi_{\rm rcp}$$
(2)

where  $\phi_{\mu} \equiv \phi_{\rm J}(a = 0, f = 1)$ ,  $\phi_{\rm alp} \equiv \phi_{\rm J}(a = 1, f = 1)$ ,  $\phi_{\rm acp} \equiv \phi_{\rm J}(a = 1, f = 0)$ , and  $\phi_{\rm rcp} \equiv \phi_{\rm J}(a = 0, f = 0)$  are the onset packing fraction for shear jamming of frictional particles in the absence of adhesion, the adhesive loose packing fraction, adhesive close packing fraction  $\phi_{\rm acp} \equiv \phi_{\rm J}(a = 1, f = 0)$ , and random close packing for hard spheres, respectively.  $\phi_{\rm rcp} = 0.64$  was fixed, and the other parameters were used to fit both the shear-thinning and -thickening regimes of the measured rheology curve. Using the fitting parameter, yield stress of each suspension was estimated by setting  $\phi_{\rm J} = \phi$  where  $\eta_{\rm r} \to \infty$ . Errors were estimated from fits with varied fitting ranges.

# Supplementary References

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