

Supplementary Information for

Computational Mesoscale Framework for Biological Clustering and Fractal Aggregation

S1 Supplementary Equations

The thermal fluctuation is included in the model by

$$m \frac{d\tilde{\mathbf{r}}_i}{dt} = \sum_j \left(A_{ij} d\bar{\mathbf{W}}_{ij} + B_{ij} \frac{1}{D} \text{tr}[d\mathbf{W}_{ij}] \right) \cdot \mathbf{e}_{ij}, \quad (\text{S1})$$

where $\bar{\mathbf{W}}_{ij}$ is a matrix of independent increments of a Wiener process for each pair i, j of particles, and $\bar{\mathbf{W}}_{ij}$ is its traceless symmetric part, given by

$$d\bar{\mathbf{W}}_{ij} = \frac{1}{2} [d\mathbf{W}_{ij} + d\mathbf{W}_{ij}^T] - \frac{\delta^{\alpha\beta}}{D} \text{tr}[d\mathbf{W}_{ij}],$$

where D is the dimensionality of the system. To satisfy the fluctuation-dissipation balance the amplitude of the thermal noises A_{ij} and B_{ij} are related to the friction coefficients a and b through

$$A_{ij} = \left[4k_B T a \frac{F_{ij}}{\rho_i \rho_j} \right]^{1/2}, \quad (\text{S2})$$

$$B_{ij} = \left[4k_B T \left(b - a \frac{D-2}{D} \right) \frac{F_{ij}}{\rho_i \rho_j} \right]^{1/2}, \quad (\text{S3})$$

To describe the variation of the pressure with the density of the system we adopt the Cole equation (a.k.a. Tait's equation of state) given by $p_i = c^2 \rho_0 / 7 [(\rho_i / \rho_0)^7 - 1] + p_b$ (S4) where c is the speed of sound on the fluid, and ρ_0 is the reference density. The term $c^2 \rho_0 / 7$ corresponds to the reference pressure of the system, given by $c^2 = \partial p / \partial \rho|_{p=p_0}$. The parameter p_b is a background pressure, that provides numerical stability by keeping the pressure of the system always positive.

For the interpolant function, we adopt the Lucy kernel¹ typically used in SDPD

$$W(r) = \begin{cases} \frac{w_0}{h^D} (1 + \frac{3r}{h}) (1 - \frac{r}{h})^3, & r/h < 1. \\ 0, & r/h > 1, \end{cases} \quad (\text{S5})$$

where $w_0 = 5/\pi$ or $w_0 = 105/16\pi$ for two or three dimensions, respectively. For a comprehensive description of the SDPD method, the reader is referred to.²

S2 MSD calculation

The time-averaged MSD is calculated from the trajectory of moving (S) particles in the range $t = 0, \dots, T$.

$$\langle \delta^2(\Delta) \rangle = \frac{1}{T-\Delta} \int_0^{T-\Delta} [\mathbf{r}(t+\Delta) - \mathbf{r}(t)]^2 dt \quad (\text{S6})$$

where Δ is the so-called lag time, which defines the size of a window slid along the trajectory $\mathbf{r}(t)$. The trajectory length T is also referred to as measurement time. Besides the individual time traces $\langle \delta^2(\Delta) \rangle$ we also considered the average over N individual trajectories:

$$\langle \langle \delta^2(\Delta) \rangle \rangle = \frac{1}{N} \sum_{k=1}^N \langle \delta^2(\Delta) \rangle \quad (\text{S7})$$

In normal diffusion, $\langle \langle \delta^2(\Delta) \rangle \rangle \sim D\Delta$, where the diffusion coefficient D is constant. In pure anomalous subdiffusion, $\langle \langle \delta^2(\Delta) \rangle \rangle \sim \Delta^\alpha$, $\alpha < 1$ at all times, where α is the anomalous diffusion exponent. The diffusion coefficient is, therefore, time-dependent, $D(t) \sim 1/t^{1-\alpha}$, appropriately modified to give the proper limit at $t = 0$, say $D(t) = D_0/(1+t^{1-\alpha})$. The case of interest here is transient anomalous subdiffusion, in which there is a crossover from anomalous subdiffusion at short times to normal diffusion at long times,

$$\langle \langle \delta^2(\Delta) \rangle \rangle \sim \begin{cases} t^\alpha & \text{for } t \ll t_{CR2} \\ t & \text{for } t \gg t_{CR2} \end{cases} \quad (\text{S8})$$

where t_{CR2} is the crossover time.

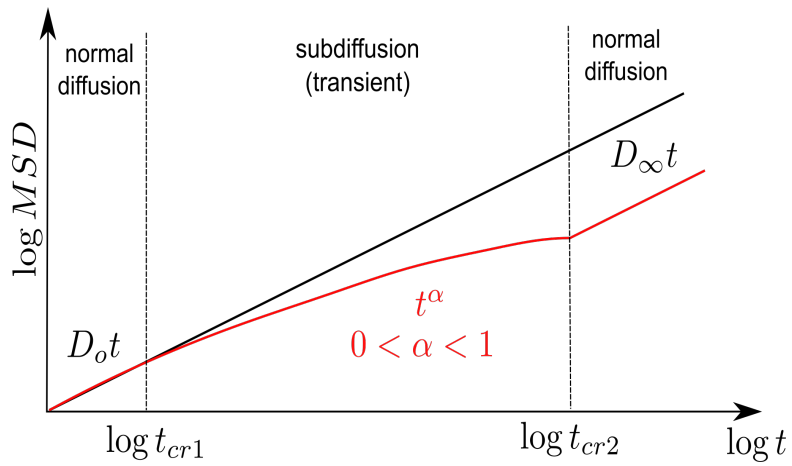


Fig. S1 Behavior of MSD in the complex clusters

S3 Supplementary Results

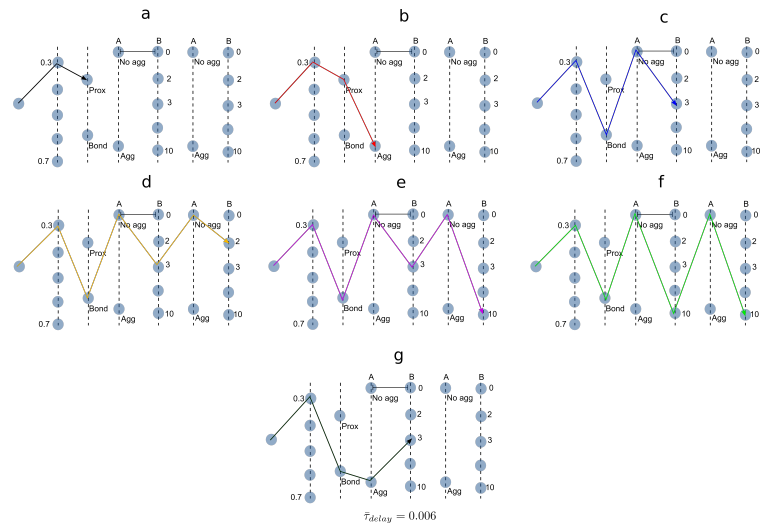


Fig. S2 The complete set of mechanisms (a)–(g) paths

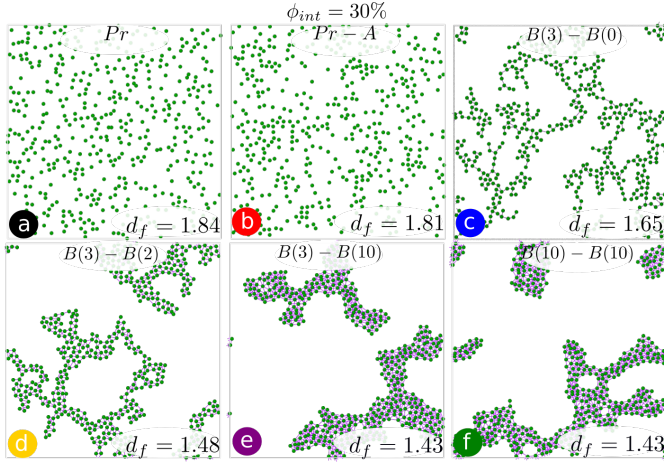


Fig. S3 Final cluster morphology and d_f are shown for six alternative mechanisms (a,b,c,d,e,f) and $\phi_{int} = 30\%$

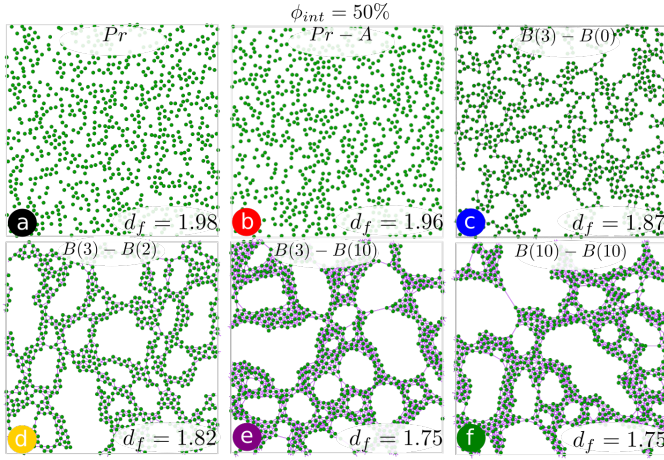


Fig. S4 Final cluster morphology and d_f are shown for six alternative mechanisms (a,b,c,d,e,f) and $\phi_{int} = 50\%$

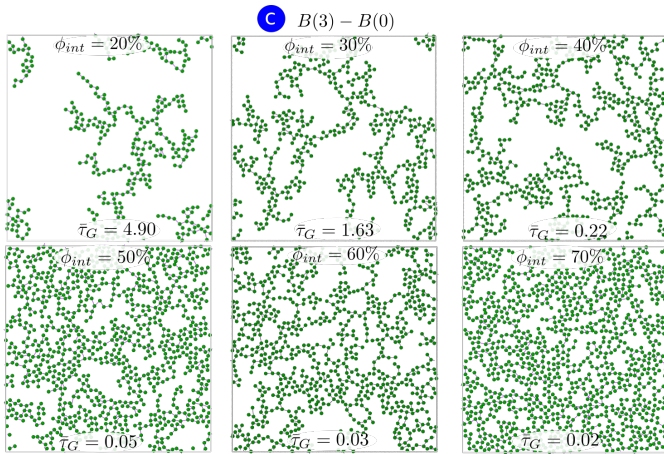


Fig. S5 Final cluster morphology and τ_G are shown for six alternative concentrations $\phi_{int} = 20\%, 30\%, 40\%, 50\%, 60\%$ and mechanism c ($B(3) - B(0)$)

S3.1 Effect of first-stage bonding number (n) on cluster morphology

We explore the morphology of clusters for three distinct mechanisms where the second-stage bonding or consolidation stage is off ($m = 0$), in order to examine the effect of the first-stage bond number. We do this by altering n bonds number created at the initial stage of bonding for $n = 2, 3, 10$ (mechanisms: $B(2)_B(0), B(3)_B(0), B(10)_B(0)$) and three different concentrations of $\phi_{int} = 30\%, 40\%, 50\%$ as shown in Fig. S6. We define the parameter N_B/N_A (the average number of bonds formed between (A) particles) as a biomarker for studying the cluster morphology. One might observe that N_B/N_A does not exceed 1 for all mechanisms and there is no difference between the structure of clusters for $n > 2$. However, because there are only two allowed bonds in the mechanism $B(2)_B(0)$, the cluster morphology resembles a chain, and the activation time for every (P) particle is quite long, especially for lower concentration values. There are still (P) particles in the system at all three concentrations of this mechanism as depicted in Fig. S6. It is possible to deduce that the characterization of a cluster is unaffected by the number of bonds in the first-stage bonding for $n > 2$. However, regardless of how large n is, N_B/N_A is equal to 1.

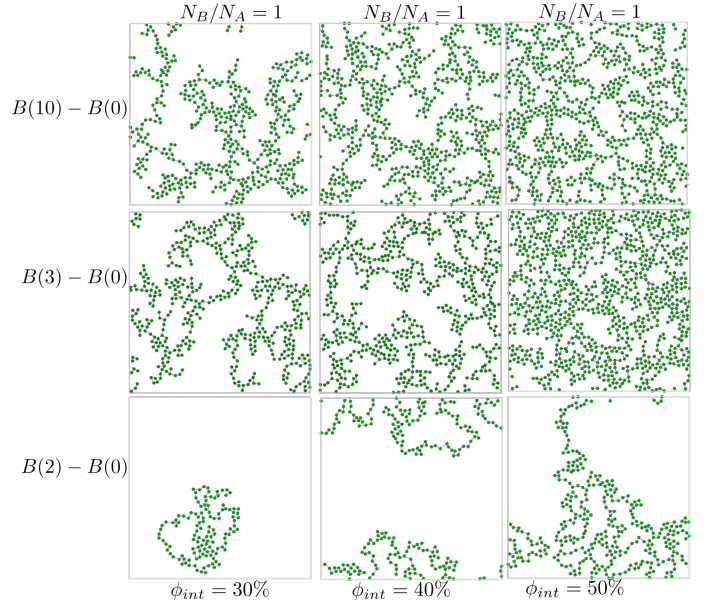


Fig. S6 Effect of the first-stage bond numbers on cluster morphology for various mechanisms: $B(2)_B(0), B(3)_B(0), B(10)_B(0)$ and $\phi_{int} = 30\%, 40\%, 50\%$. The average number of bonds formed between (A) particles (N_B/N_A) are shown.

S3.2 Effect of second-stage bonding number (m) on cluster morphology

To investigate the effect of the second-stage bond number, the first-stage bonding number, n is fixed to 3, and the bond number in the second stage, m is adjusted to $m = 0, 2$, and 10, equal to mechanism types c, d and e ($B(3)_B(0), B(3)_B(2)$ and $B(3)_B(10)$). In Fig. S7 the cluster morphologies of these three mechanisms and different concentrations $\phi_{int} = 30\%, 40\%, 50\%$ are shown. When there is no

bond in the second stage ($m = 0$), the average number of bonds formed between (A) particles, $N_B/N_A = 1$ whereas for $m = 2$ and $m = 10$, the cluster morphology becomes more branched, solid and stable with $N_B/N_A > 1$, resulting the lower value of d_f . It implies that we can only imitate mature clots using second-stage bonding. However, it appears that the second-stage bond number has altered not only the fractality of the cluster but also the cluster kinetic characteristics, as evaluated by MSD measurement (outlined in this study as a new biomarker).

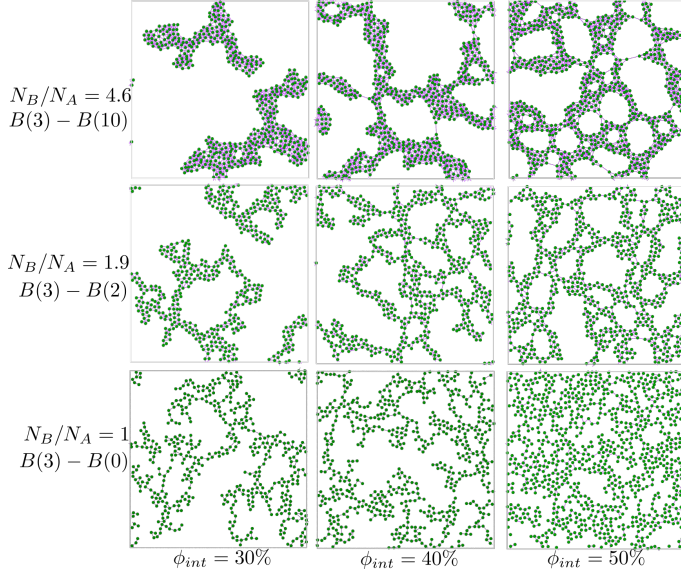


Fig. S7 Effect of the second-stage bond numbers on cluster morphology for various mechanisms of (c,d,e) and $\phi_{int} = 30\%, 40\%, 50\%$. The average number of bonds formed between (A) particles (N_B/N_A) are shown.

S3.3 Effect of delay time between first-stage bonding and aggregation on the cluster morphology and τ_l

This section's primary goal is to examine the impact of the delay time between the aggregation process and the first-stage bonding on the cluster morphology and τ_l . We consider mechanism g ($B(3)A_B(0)$) and $\phi_{int} = 40\%$ to see this effect. The delay time is $\bar{\tau}_{delay} = 0.006$ that is dimensionless with τ_{diff} . In this manner, we apply initiation bonding for every τ_{delay} duration of aggregation. The results in Fig. S8.c show that by delaying the time between bonding and aggregation, $\bar{\tau}_l$ increases in comparison to the case $\tau_{delay} = 0$ (seen in Fig. S8.b) also d_f and final cluster morphology profoundly changes. We also infer that adding aggregation to mechanism c ($B(3)_B(0)$) without delay time (seen in Fig. S8. a, b) has little effect on d_f and τ_l . In Fig. S9, which depicts cluster evolution with time for $\bar{\tau} = 0.006$, can observe how the droplet form of (P) particles (because of aggregation) makes the first-bonding stage slower. Delay time, replicates the activation delay time for platelets during the clot-forming process.

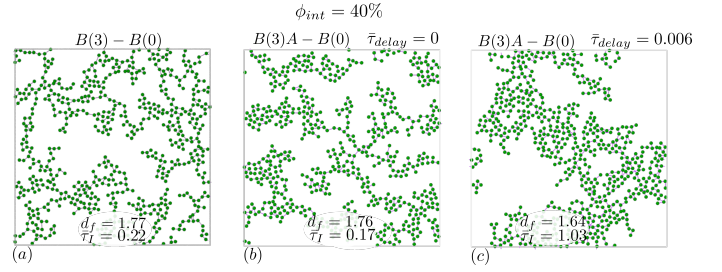


Fig. S8 Effect of delay time between first-stage bonding and aggregation on cluster structure. The final morphology, d_f and $\bar{\tau}_l$ are displayed for $\phi_{int} = 40\%$ and for a) mechanism c, b) mechanism g, and $\bar{\tau}_{delay} = 0$, c) mechanism g, and $\bar{\tau}_{delay} = 0.006$ to see how the delay time between aggregation and activation change cluster morphology.

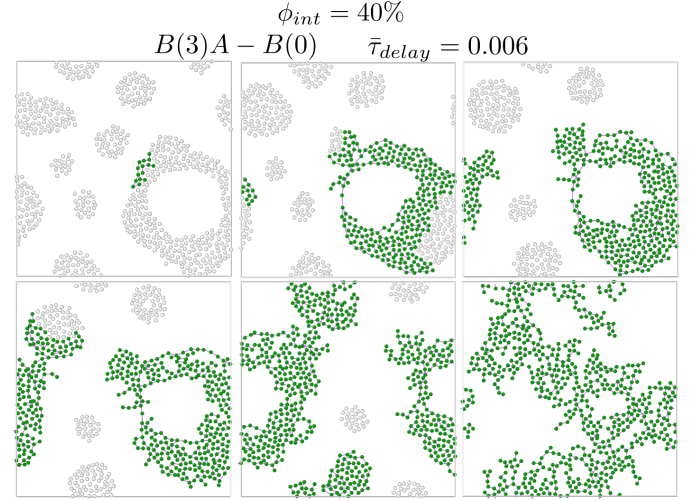


Fig. S9 Cluster evolution with time for mechanism g and $\bar{\tau}_{delay} = 0.006$ of first-stage bonding and aggregation. The activation of passive particles can also be seen in the step-by-step morphology of this mechanism.

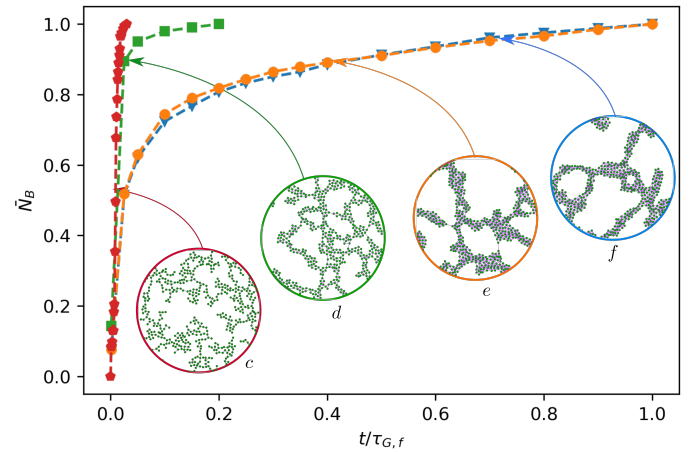


Fig. S10 Dimensionless bonds number \bar{N}_B evolution with time for various mechanisms of c,d,e,f and $\phi_{int} = 40\%$. Each curve is dimensionless with its values in a steady state or gelling state, and time is also dimensionless with τ_G of mechanism f. The final morphology for each mechanism is displayed.

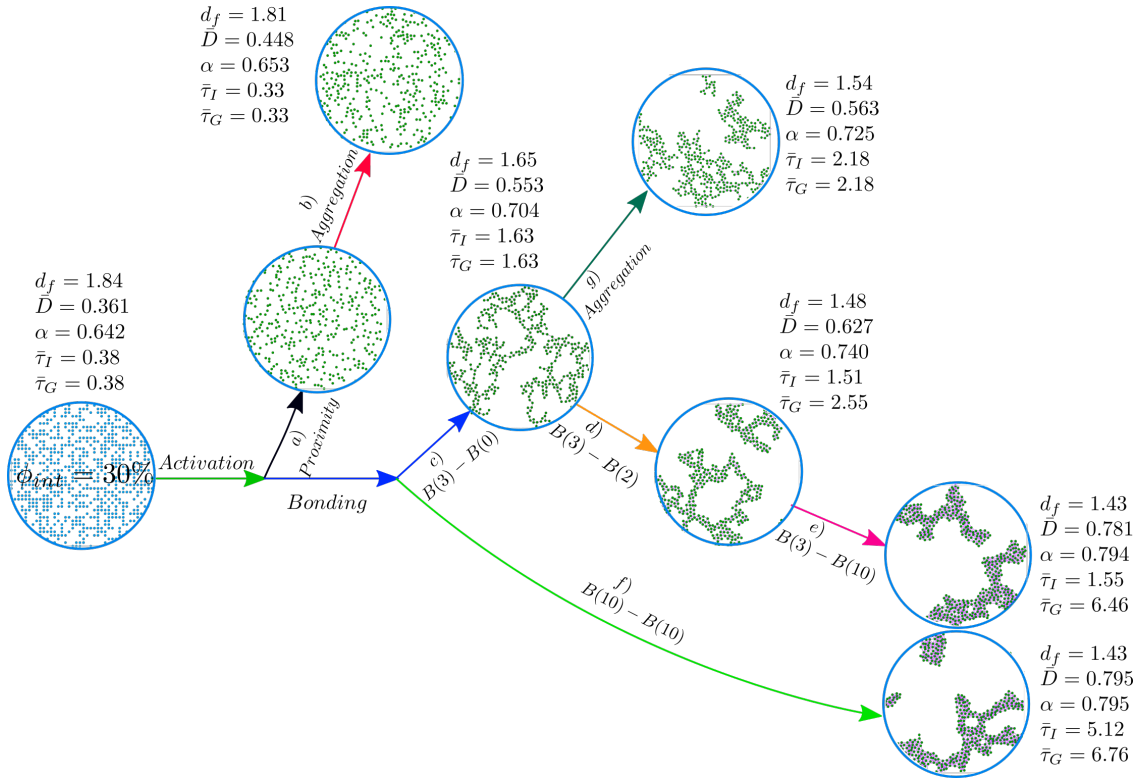


Fig. S11 A biomarker diagram is provided step by step regarding the implementation of different mechanisms in our model for $\phi_{int} = 30\%$. Biomarkers include d_f , \bar{D} , α , $\bar{\tau}_I$ and $\bar{\tau}_G$.

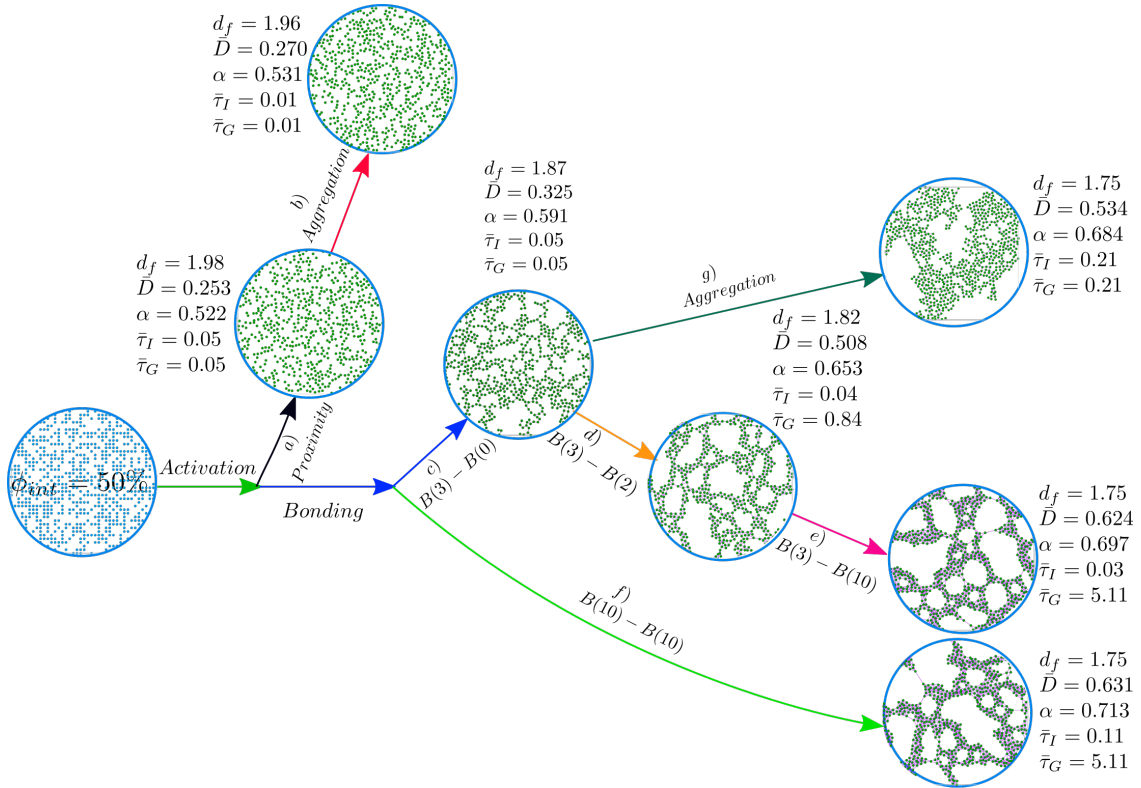


Fig. S12 A biomarker diagram is provided step by step regarding the implementation of different mechanisms in our model for $\phi_{int} = 50\%$. Biomarkers include d_f , \bar{D} , α , $\bar{\tau}_I$ and $\bar{\tau}_G$.

S4 Supplementary Tables

Table S1 Input parameters of the SDPD method

Domain size [$L_x \times L_y$]	$(40dx) \times (40dx)$
Total number of particles (N_t)	1600
Mass(m)	0.04
viscosity(η)	10
$k_B T$	0.1
Density(ρ_0)	1
Pressure(p_0)	50
Speed of sound (c_s)	40
Time step (dt)	10^{-4}
Initial lattice spacing(dx)	0.2
Cutoff radius (h)	$4dx$

Notes and references

1 P. Espanol and M. Revenga, *Physical Review E*, 2003, **67**, 026705.
2 M. Ellero and P. Español, *Applied Mathematics and Mechanics*, 2018, **39**, 103–124.

Table S2 Input parameters of the bond and surface tension potentials

Bond Strength, D_{Morse} (Eq. 6)	Stiffness Parame- ter, α (Eq. 6)	Bond Length, r_0 (Eq. 6)	Surface Tension, σ (Eq. 5)
30	1	0.2	0.5

Table S3 d_f values (with the standard deviation equal to ± 0.01) for dif-
ferent, ϕ_{int} and six different mechanisms

Mechanisms	Concentrations		
	$\phi_{int} = 30\%$	$\phi_{int} = 40\%$	$\phi_{int} = 50\%$
a	$d_f = 1.84$	$d_f = 1.92$	$d_f = 1.98$
b	$d_f = 1.81$	$d_f = 1.90$	$d_f = 1.96$
c	$d_f = 1.65$	$d_f = 1.77$	$d_f = 1.87$
d	$d_f = 1.48$	$d_f = 1.71$	$d_f = 1.82$
e	$d_f = 1.43$	$d_f = 1.56$	$d_f = 1.75$
f	$d_f = 1.43$	$d_f = 1.56$	$d_f = 1.75$