

1 **Supporting Information**

2 **Local microenvironment tuning induces switching between**
3 **electrochemical CO₂ reduction pathways**

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36 **Methods**

37 **Materials.** Potassium hydroxide (ACS reagent, $\geq 85\%$), potassium formate (ReagentPlus, 99%),
38 potassium bicarbonate ($\geq 99.95\%$ trace metals basis) and 8-Aminopyrene-1,3,6-trisulfonic acid
39 trisodium salt (APTS, $\geq 96.0\%$), were purchased from Sigma-Aldrich. Nickel foam and anion
40 exchange membrane (Fumasep FAA-3-PK-130) were purchased from Fuel Cell Store.
41 Hydrophobic PTFE substrates of various pore sizes were obtained from Beijing Zhongxingweiye
42 Instrument Co. Ltd. The custom designed electrochemical flow cell and Ag/AgCl (3M KCl)
43 reference electrodes were procured from Tianjin Aida Hengsheng Technology Development Co.
44 For confocal microscopy experiments, a leakless Ag/AgCl reference electrode from eDAQ and a
45 Pt mesh (99.9%, 0.0726 mm diameter wires) from Alfa Aesar were used. Ltd. Aminex HPX-87H
46 columns were used for high performance liquid chromatography and these were purchased from
47 Bio-Rad Laboratories. The standard calibration gas mixtures for calibrating the gas
48 chromatography system were obtained from CitiSafe Pte. Ltd. All materials were used as
49 received without any further purification or treatment.

50 **Fabrication of Ag/PTFE and Cu/PTFE.** Samples were fabricated by sputter deposition of 325
51 nm Ag or Cu onto PTFE substrates of various pore sizes, using a DC sputtering system (Denton
52 Discovery D18). The Ag and Cu sputtering targets (Latech Scientific Supply Pte. Ltd.) used both
53 have a diameter of 3", thickness of 3 mm and a purity of 99.99%.

54 **Materials characterization.** XRD analysis of Ag/PTFE samples was carried out with a Bruker
55 D8 Discover diffractometer using Cu $K\alpha$ radiation under 40 kV and 40 mA. Catalyst
56 morphology was studied using scanning electron microscopy (SEM) using a JEOL 7600F. For
57 cross-section SEM images, focus ion beam (FIB) scanning electron microscopy (FEI Helios
58 NanoLab 450) was used. Before the FIB cut can be made, each PTFE/Ag sample is first coated

59 with 100 nm of carbon using a Leica model EM ACE200, followed by another ~15 nm of Au on
60 top of it to prevent charging of sample during the FIB cutting process. In the FIB system, a strip
61 of Pt of 500 nm thickness is deposited onto the desired area to be cut before starting the milling
62 process. The sample is then milled down slowly, moving deeper into the depth of the sample
63 until the desired SEM cross-section is achieved. A current of 2.5 nA was used for the milling
64 process.

65 **Electrochemical CO₂ reduction.** The Ag/PTFE samples were tested in a custom designed
66 electrochemical flow cell system (Fig. S13), with an active area of 1 cm² (cathode). This was
67 operated under ambient pressure and temperature. An Autolab PGSTAT302N potentiostat
68 system was used to control the current and measure the voltage supplied to the electrochemical
69 cell. The potentiostat system was equipped with a FRA32M module for performing
70 electrochemical impedance spectroscopy (EIS) measurements. In the flow cell, Ni foam was
71 used as the counter electrode (anode) and an anion exchange membrane was used to separate the
72 cathode and anode chambers. Ag/AgCl (3M KCl) was used as the reference electrode. KHCO₃
73 solution of varying concentrations was employed as the electrolyte used for both the catholyte
74 and anolyte. The electrolyte was stored in external centrifuge tube reservoirs and continuously
75 recirculated through the electrochemical cell using peristaltic pumps.

76 CO₂ was supplied at a flow rate of 20 sccm using a mass flow controller (Alicat Scientific) to the
77 backside of the Ag/PTFE gas diffusion electrode. The effluent CO₂ exiting the cell was directly
78 connected to an online gas chromatography (GC) system (Shimadzu Nexis GC-2030), for
79 analysis of gas products. The GC was equipped with a thermal conductivity detector for analysis
80 of H₂ and a methanizer+flame ionization detector for analysis of CO. Calibration of the GC was
81 performed using standard calibration gas mixtures. At the end of the experiment, the formate in

82 both the catholyte and anolyte were analyzed using high-performance liquid chromatography
83 (Shimadzu LC-2030C NT) equipped with a UV and RID detector. Calibration was carried out by
84 injecting known concentrations of aqueous potassium formate solutions into the system.

85 **Double layer capacitance measurements.** These were performed in the same electrochemical
86 cell as that for CO₂R. 1 M KHCO₃ was employed as the electrolyte and cyclic voltammetry was
87 carried out in a potential window where faradaic processes do not take place. This was carried
88 out under various scan rates of 80, 60, 40 and 20 mV/s. The capacitance current density was
89 plotted vs scan rate and the slope of this graph gives the double layer capacitance. This value is
90 directly proportional to the electrochemically active surface area (ECSA)¹.

91 **Confocal microscopy experiments.** Confocal microscopy experiments were performed with a
92 Zeiss LSM 880 confocal microscope (Fig. S17). A WN Achromplan 63x water immersion
93 objective with a working distance of 1.7 mm was used. A customized electrochemical cell was
94 designed and 3D-printed to be compatible with confocal microscopy and a water immersion
95 objective (Fig 6a). The cell is oriented horizontally and operates without an ion exchange
96 membrane. A rubber gasket between the gas chamber and electrolyte chamber seals the cell. The
97 flow rate of the CO₂ gas stream through the gas chamber was adjusted to 10 sccm. The
98 electrolyte chamber exhibits two perpendicular inlet and outlet tubes. Electrolyte is recirculated
99 through the electrochemical cell at a flow rate of 6 mL/min. The active surface area of the
100 working electrode is 0.2 cm². A leakless Ag/AgCl reference electrode and a Pt mesh counter
101 electrode are both immersed into the electrolyte. 200 μM APTS was dissolved in 1 M KHCO₃
102 electrolyte and filled into the electrolyte chamber. A 458 nm laser scans the sample and excites
103 the fluorescent APTS dye molecules. The laser power is set to 100%, the pinhole to 57.1 μm and
104 the gain to 800. The emission is collected separately in the wavelength intervals 480 – 550 nm

105 and 551 – 754 nm. The ratio between the two signals collected is a measure of the local pH
106 value. The pH value can be calculated with a previously determined sigmoidal calibration curve:

107

$$pH = \frac{1}{2.743} \ln \left(-1 + \frac{5.005}{(Ratio\ of\ Emission)_{APTS} - 0.1041} \right) - 11.95.$$

108 More details about the calibration of APTS can be found elsewhere². Before each experiment, a
109 potentiostatic electrochemical impedance spectroscopy (PEIS) was performed with a Biologic
110 SP-200 potentiostat to determine the solution resistance of the cell. A constant current was then
111 applied with the potentiostat while performing an automatic 85% IR electronic compensation of
112 the electrochemical potential. The system was allowed to equilibrate for 15 seconds before
113 taking fluorescence measurements. The laser beam scanned the electrolyte in the vicinity of the
114 electrode in the plane perpendicular to the electrode surface by scanning the laser line by line and
115 moving the stage in the z-direction in 0.5 μm increments. The first line was placed a few
116 micrometers below the electrode surface. The dimensions of the resulting map are 120 μm in x
117 and 50 μm in z. The measuring speed was adjusted so that capturing one frame takes
118 approximately 30 seconds. In between measurements, the electrolyte was removed from the
119 electrochemical cell and replaced with electrolyte containing fresh APTS. Each measurement
120 was performed at least six times.

121 **Multiphysics simulations.** The model employed was previously reported by Weng et al. and full
122 details can be found in the corresponding publication³. The simulations were implemented using
123 COMSOL Multiphysics software. This model is a stationary, isothermal and 1D axisymmetric
124 model for the cathodic compartment of the cell, which contains the nanoparticle Ag catalyst
125 layer and diffusion medium. CO formation and hydrogen evolution reaction are the two
126 electrochemical reactions focused on. Triangular meshes were applied.

127 The governing equation is mass balance, which is written as

128
$$R_i = \nabla \cdot n_i = \nabla \cdot J_i + u \cdot \nabla c_i$$

129 where R_i is reaction rate, n_i is mass flux, J_i is diffusive mass flux, u is velocity and c_i is
130 concentration.

131 For charged chemical species in liquid phase, the Nernst–Planck equation is the conservation of
132 mass equation. With assumption that the convection is negligible, the equation can be written as:

133
$$n_i = -D_i \nabla c_i - z_i u_{m,i} F c_i \nabla \varphi_l$$

134 where D_i is the diffusivity of the chemical species, z_i is the valence of ionic species, $u_{m,i}$ is the
135 mobility in medium m, F is the Faraday's constant, and φ_l is the potential of liquid phase.

136 The reaction rate is composed of three elements,

137
$$R_i = R_{ec,i} + R_{dis,i} + R_{b,i}$$

138 where $R_{ec,i}$, $R_{dis,i}$ and $R_{b,i}$ represents electrochemical reaction, CO₂ dissolution and
139 homogeneous bulk reactions respectively.

140 For each electrochemical reaction, the reaction rate is:

141
$$R_{ec,i,k} = -\frac{\nu_i a_v i_k}{nF}$$

142 where ν_i is stoichiometric number, a_v is specific surface area and n is electron transferred in
143 reaction. i_k is current density of reaction k, which is calculated by the generic concentration
144 dependent Butler-Volmer type kinetics,

145
$$i_k = -i_{0,k} C_O \exp\left(-\frac{\alpha_{c,k} F}{RT} \eta_k\right)$$

146 where is C_O dimensionless concentration, $i_{0,k}$ is exchange current density and $\alpha_{c,k}$ is cathodic
 147 charge transfer coefficient. The overpotential η_k is:

148
$$\eta_k = \varphi_s - \varphi_l - (U_{0,k} - 0.059pH)$$

149 where $U_{0,k}$ is reference potential and φ_s is the potential of gas phase. Moreover, for
 150 electrochemical reactions, the process is governed by charge conservation and Ohm's law as well,

151
$$\nabla \cdot i_s = -\nabla \cdot i_l = a_v \sum_k i_k$$

152
$$i_s = -\sigma_s \nabla \varphi_s$$

153 where σ_s is the electrical conductivity. Here, Bruggeman relationship is applied to obtain the
 154 effective σ_s as the diffusion medium is porous.

155 The dissolution rate of CO₂ is obtained by Fick's law:

156
$$R_{dis,CO_2} = \frac{D_{CO_2(l)} a_v \Delta p}{\delta_{TF}}$$

157 where Δp is difference in CO₂ partial pressure across the thin electrolyte film with thickness δ_{TF} .

158 The homogenous buffering reactions are mainly carbonate and water dissociation reactions:

159
$$R_{B,i,k} = k_n \prod_{v_i < 0} c_i - k_{-n} \prod_{v_i > 0} c_i$$

160 where k_n and k_{-n} are rate constants for forward and reverse homogeneous reaction.

161 For gas phase, the mass flux is composed of diffusion and convection:

162
$$n_i = j_i + \rho_i u_g$$

163 where ρ_i is gas density.

164 The reactions in the gas phase only includes electrochemical reactions and CO₂ dissolution.

165
$$R_i = R_{ec,i} + R_{dis,i}$$

166
$$R_{dis,CO_2} = -\frac{D_{CO_2(l)} a_v \Delta p}{\delta_{TF}}$$

167
$$R_{ec,i,k} = -\frac{v_i a_v i_k}{nF}$$

168 The diffusive flux j_i is calculated using a mixture averaged diffusion model,

169
$$j_i = -\rho_i D_i^{eff} \nabla \omega_i - \rho_i D_i^{eff} \omega_i \frac{\nabla M_n}{M_n}$$

170 where ω_i is mass fraction, M_n is the average molar mass of gas mixture. The effective diffusion
171 coefficient is corrected by Bruggeman correlation.

172 The model was validated under various potential ranging from -0.8 to -2.0 V vs RHE. The order
173 of magnitude of calculated current densities is similar to reference based on experimental data
174 and shows similar tendency along with the change of applied potential.

175 For the purposes of this work, the porosity values of the diffusion media and catalyst layer were
176 set to be identical to each other and varied across the values of 0.4, 0.5, 0.6, 0.7 and 0.8. A
177 parametric sweep was used to facilitate this simulation process. The CO₂ concentration and pH
178 profile were determined for each porosity value, at cathodic current densities of 100, 200, 300,
179 400 and 500 mA/cm².

180 List of symbols used in the model:

181

182 $\alpha_{c,k}$ Cathodic charge transfer coefficient

183 Δp Difference in CO₂ partial pressure across the thin electrolyte film with thickness, atm

184 δ_{TF} Thin electrolyte film thickness

185 η_k Overpotential

186 ν_i Stoichiometric number

187 ρ_i Density, g/cm³

188 σ_s Electrical conductivity, S/m

189 φ_l Potential of liquid phase, V

190 ω_i Mass fraction

191 a_v Specific surface area, 1/m

192 C_o Dimensionless concentration

193 c_i Concentration, mol/m³

194 D_i Diffusivity, m²/s

195 F Faraday's constant, C/mol

196 i Current density, mA/cm²

197 $i_{0,k}$ Exchange current density, mA/cm²

198 J_i Diffusive mass flux, m²/s

199 k_n, k_{-n} Rate constants for forward and reverse homogeneous reaction

200 M_n Average molar mass of gas mixture, g/mol

201 n Numbers of electron transferred in reaction

202 n_i Mass flux, g/(m²·s)

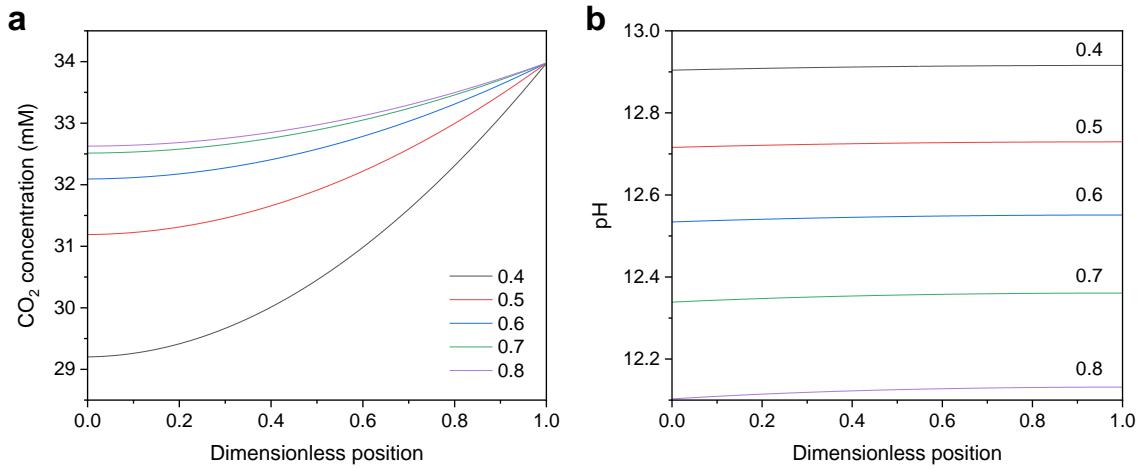
203 R_i Reaction rate, g/(m³·s)

204 u Velocity, m/s

205 $u_{m,i}$ Mobility in medium m, s/(mol·kg)

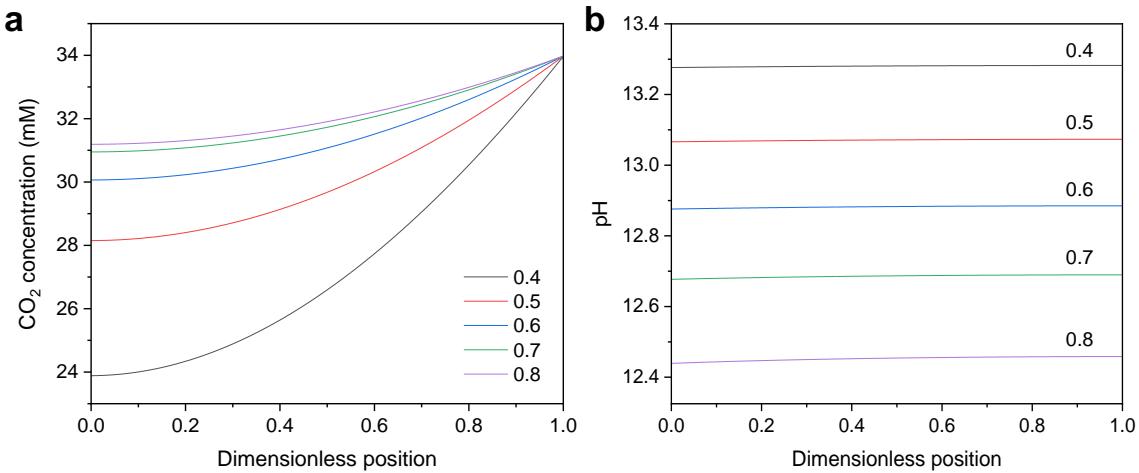
206 $U_{0,k}$ Reference potential, V

207 z_i Valence of ionic species



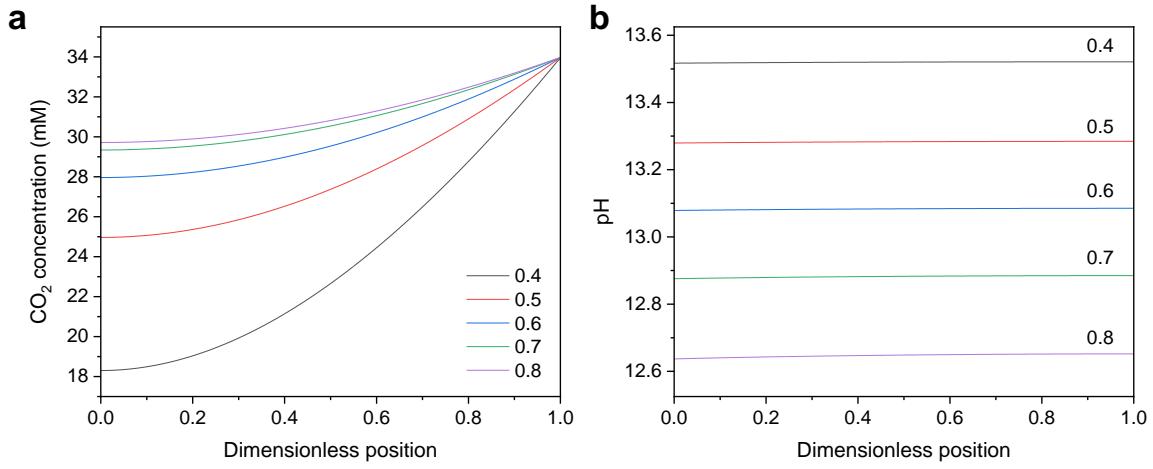
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209 **Fig. S1.** Simulated (a) CO₂ concentration and (b) pH profile in the boundary layer at various
210 GDL porosity values. Applied cathodic current density is 100 mA/cm².



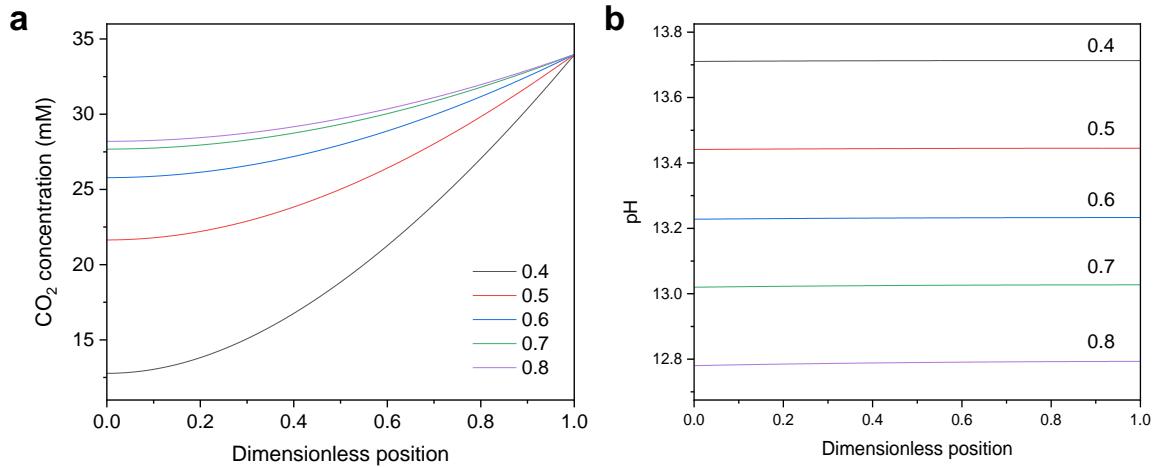
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212 **Fig. S2.** Simulated (a) CO₂ concentration and (b) pH profile in the boundary layer at various
213 GDL porosity values. Applied cathodic current density is 200 mA/cm².



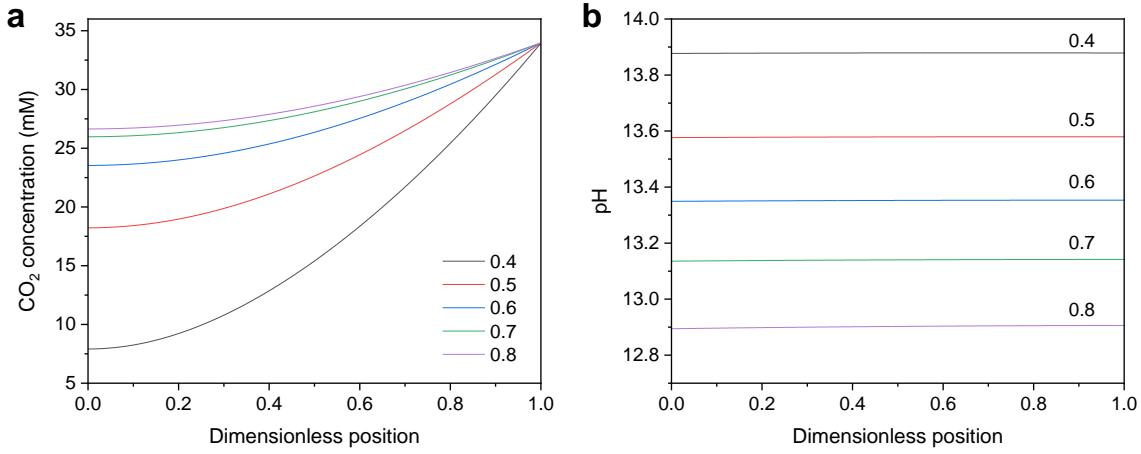
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215 **Fig. S3.** Simulated (a) CO₂ concentration and (b) pH profile in the boundary layer at various
 216 GDL porosity values. Applied cathodic current density is 300 mA/cm².



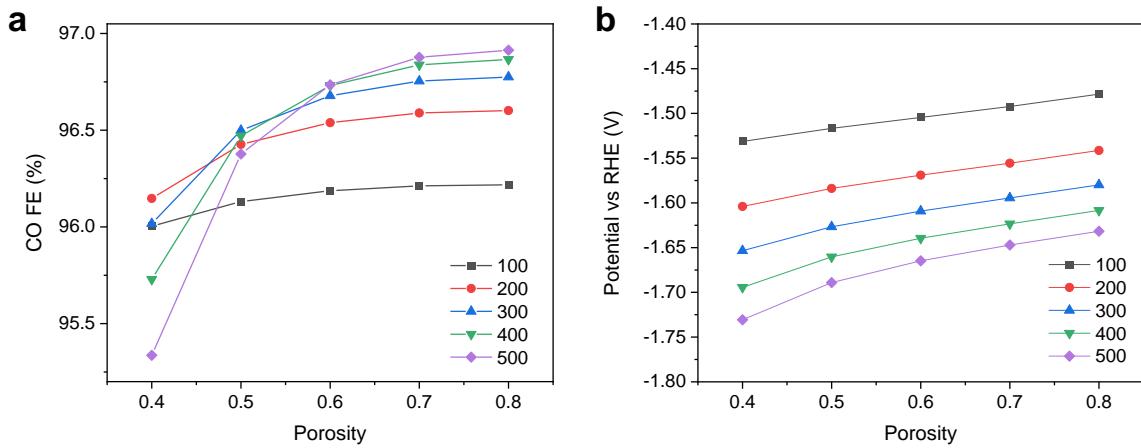
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218 **Fig. S4.** Simulated (a) CO₂ concentration and (b) pH profile in the boundary layer at various
 219 GDL porosity values. Applied cathodic current density is 400 mA/cm².



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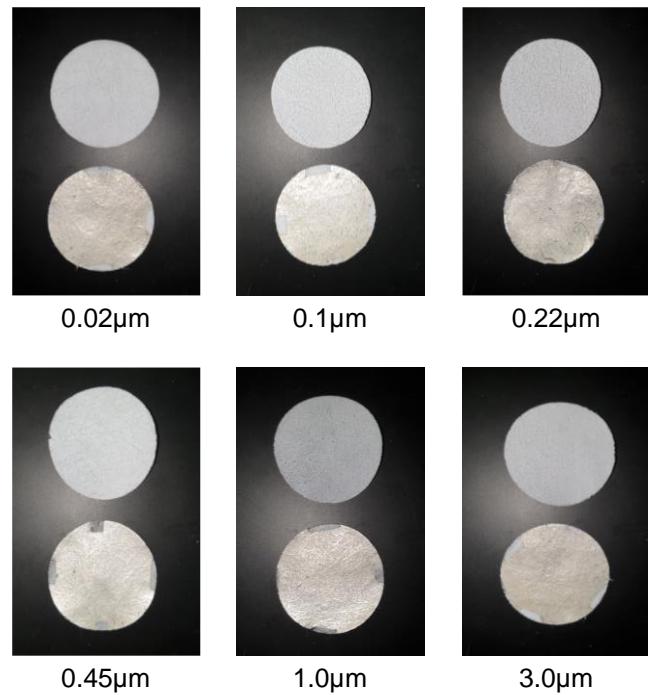
221 **Fig. S5.** Simulated (a) CO_2 concentration and (b) pH profile in the boundary layer at various
222 GDL porosity values. Applied cathodic current density is 500 mA/cm^2 .



223

224 **Fig. S6.** Simulated (a) CO FE and (b) applied potential at various GDL porosity values. The
225 values in the legends of both figures are current densities with units of mA/cm^2 .

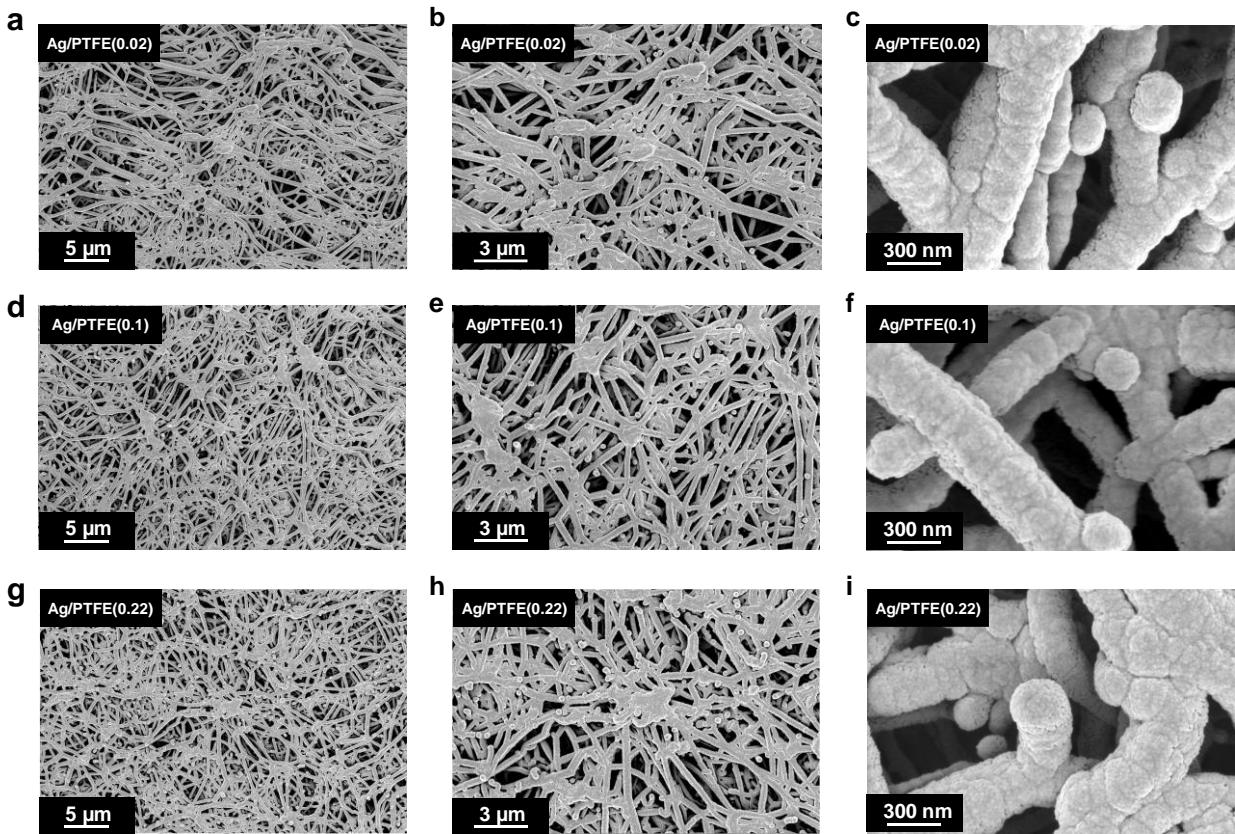
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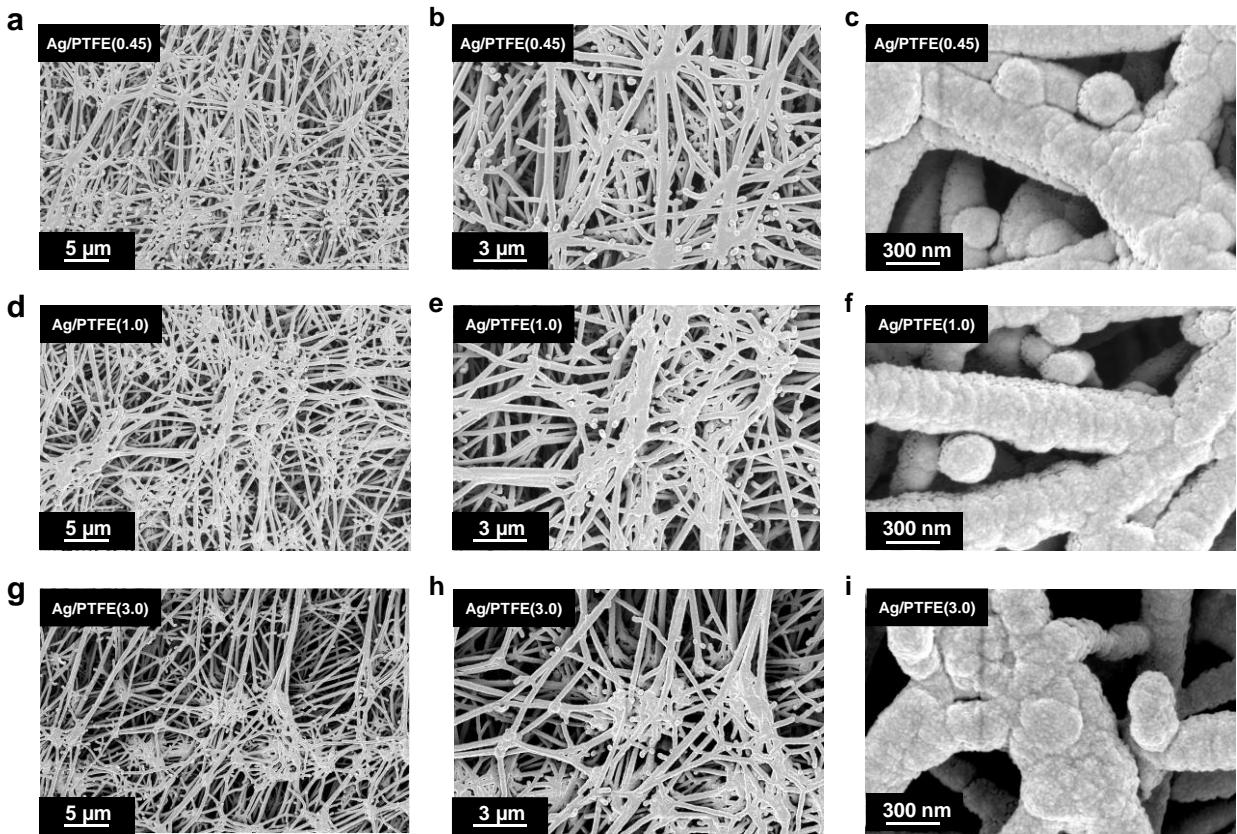


227 **Fig. S7.** Digital photographs of pristine and Ag sputter coated PTFE substrates for each pore size.

228 For each pore size, the pristine case is placed on top and the coated case is placed at the bottom.

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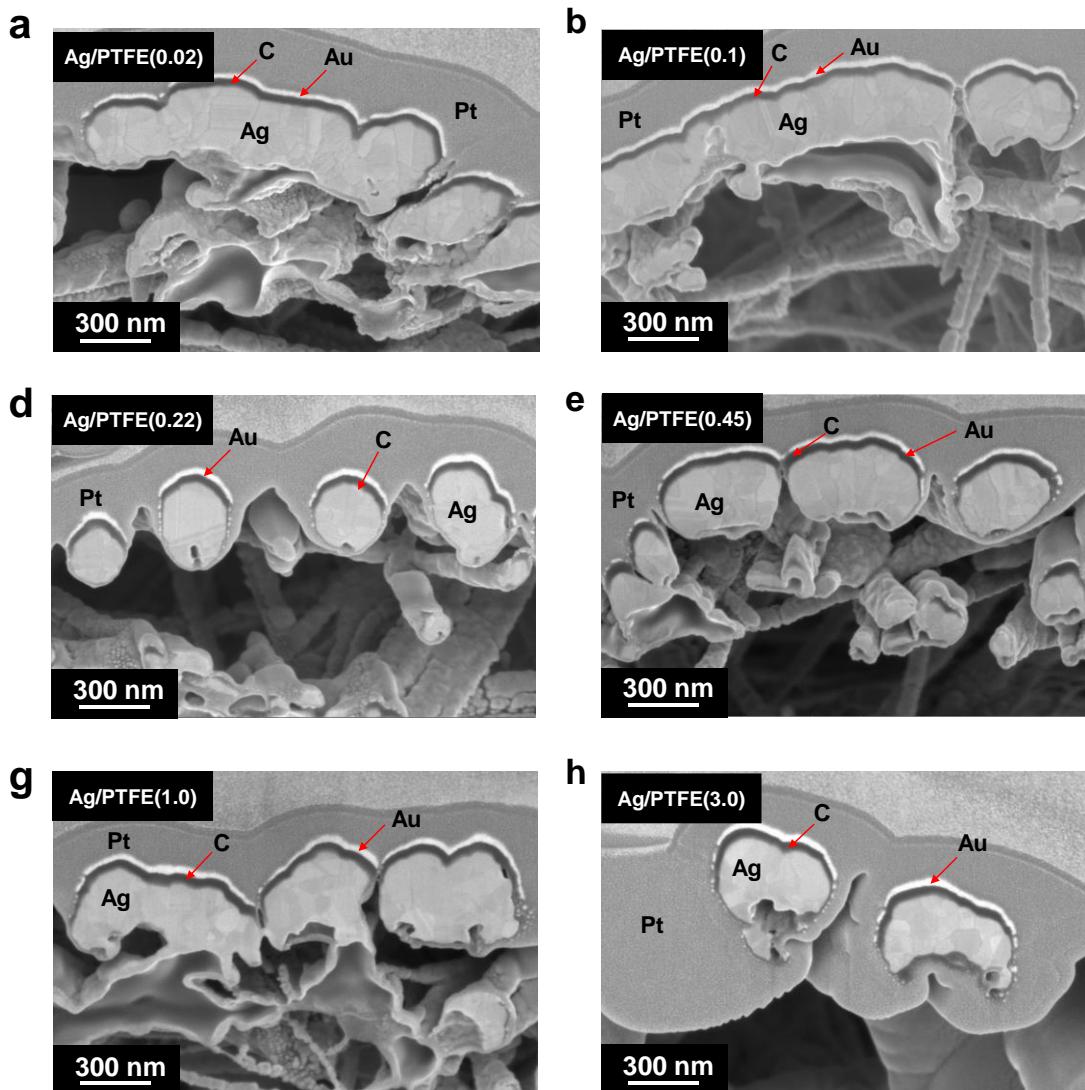




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235 **Fig. S9.** Top-down SEM images of Ag/PTFE with different magnifications. (a), (b) and (c) are
 236 for Ag/PTFE(0.45). (d), (e) and (f) are for Ag/PTFE(1.0). (g), (h) and (i) are for Ag/PTFE(3.0)

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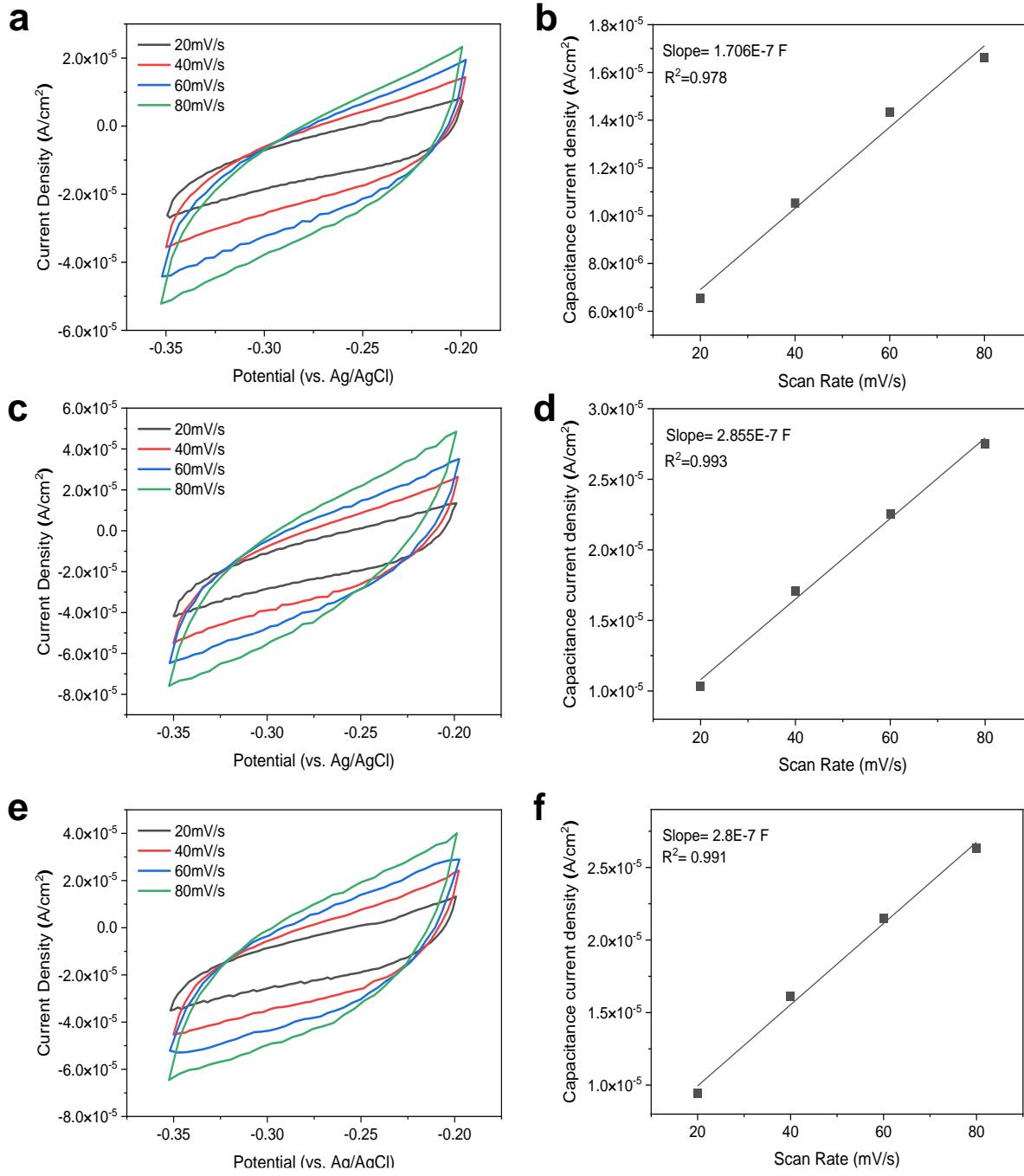


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239 **Fig. S10.** Cross-section SEM images of (a) Ag/PTFE(0.02), (b) Ag/PTFE(0.1), (c)
 240 Ag/PTFE(0.22), (d) Ag/PTFE(0.45), (e) Ag/PTFE(1.0) and (f) Ag/PTFE(3.0). See methods
 241 section for a description of the preparation process.

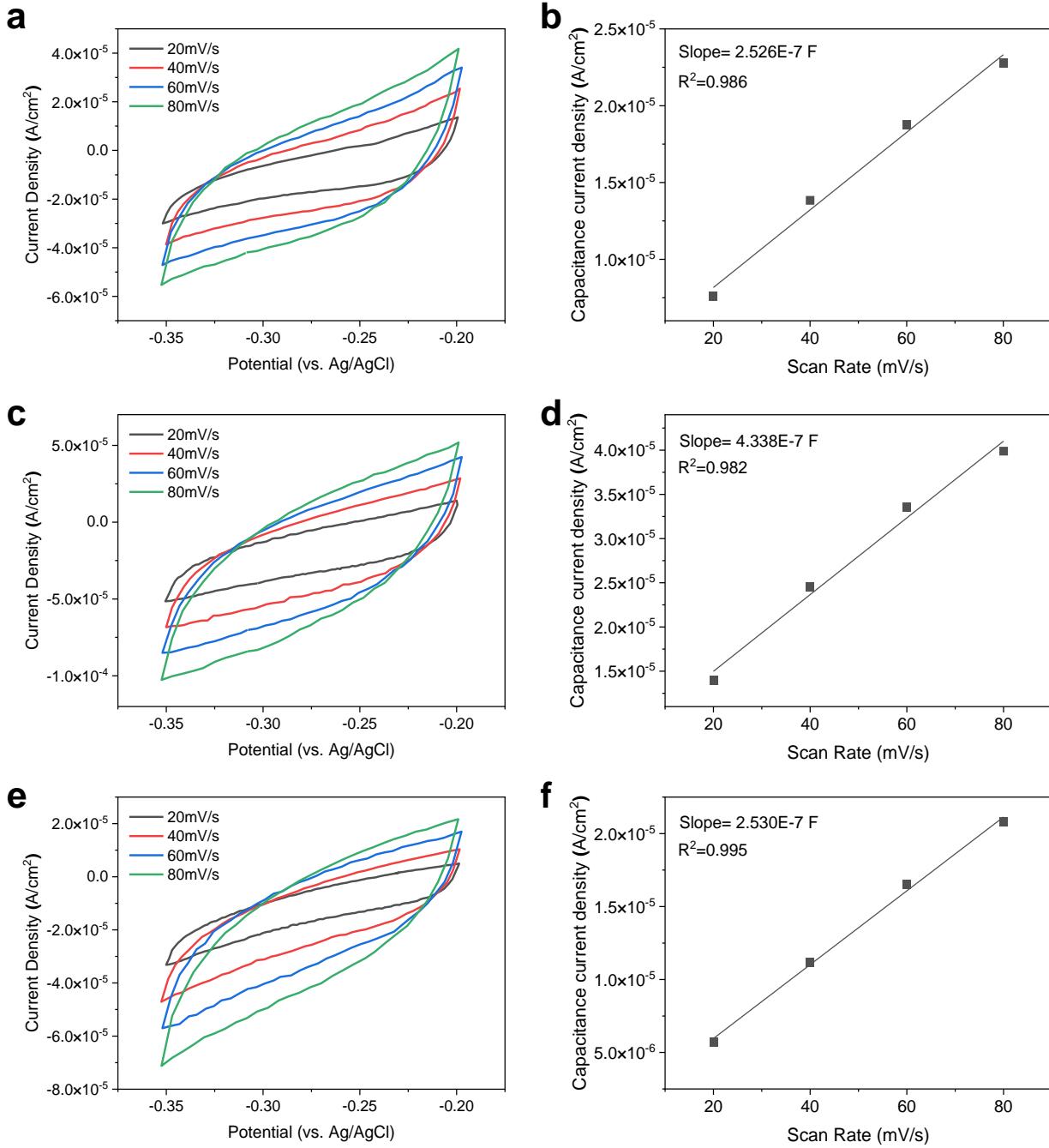
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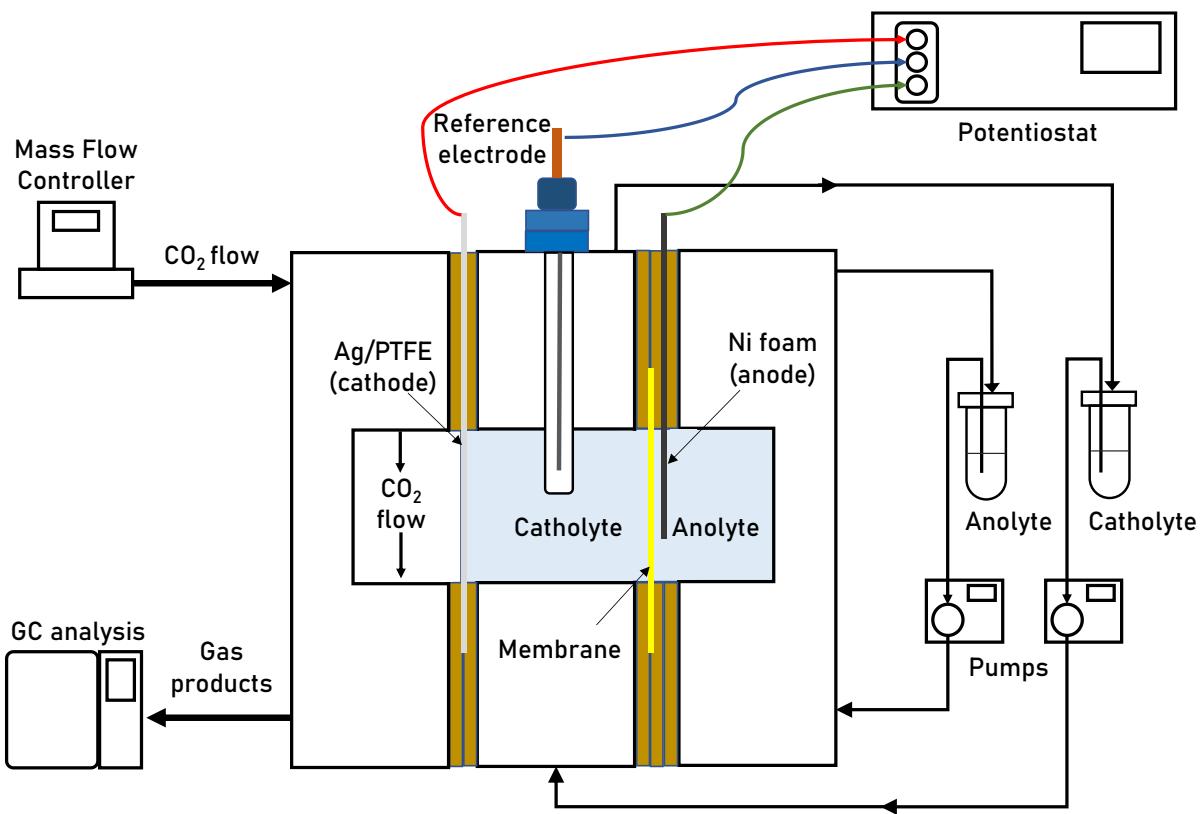
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245 **Fig. S11.** (a), (c) and (e) are cyclic voltammetry results at various scan rates for Ag/PTFE(0.02),
 246 Ag/PTFE(0.1) and Ag/PTFE(0.22) respectively. (b), (d) and (f) are the capacitance current
 247 density plotted against the scan rate for for Ag/PTFE(0.02), Ag/PTFE(0.1) and Ag/PTFE(0.22)
 248 respectively. The slope of the graph gives the capacitance, which is directly proportional to the
 249 electrochemically active surface area (ECSA).



250 **Fig. S12.** (a), (c) and (e) are cyclic voltammetry results at various scan rates for Ag/PTFE(0.45),
251 Ag/PTFE(1.0) and Ag/PTFE(3.0) respectively. (b), (d) and (f) are the capacitance current density
252 plotted against the scan rate for Ag/PTFE(0.45), Ag/PTFE(1.0) and Ag/PTFE(3.0)
253 respectively. The slope of the graph gives the capacitance, which is directly proportional to the
254 electrochemically active surface area (ECSA).
255

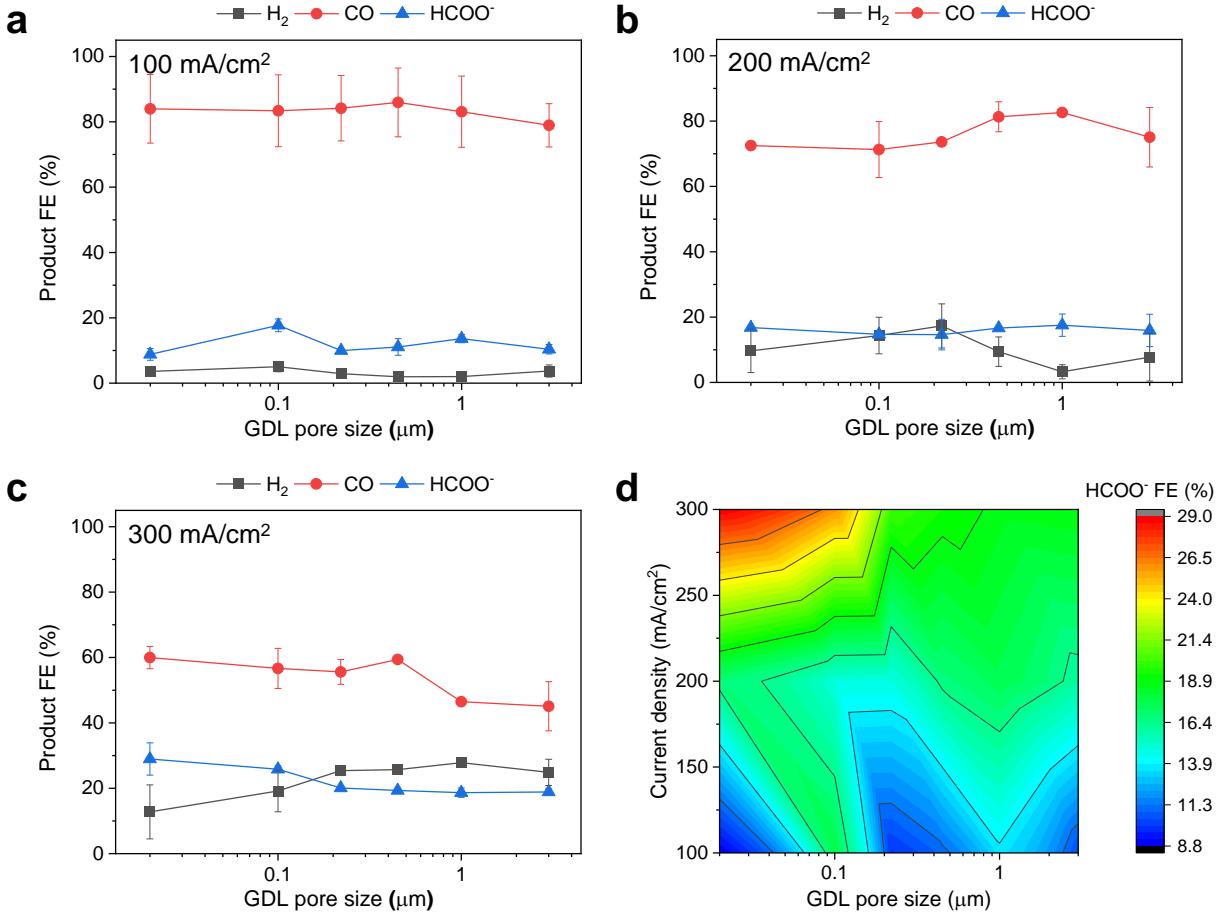
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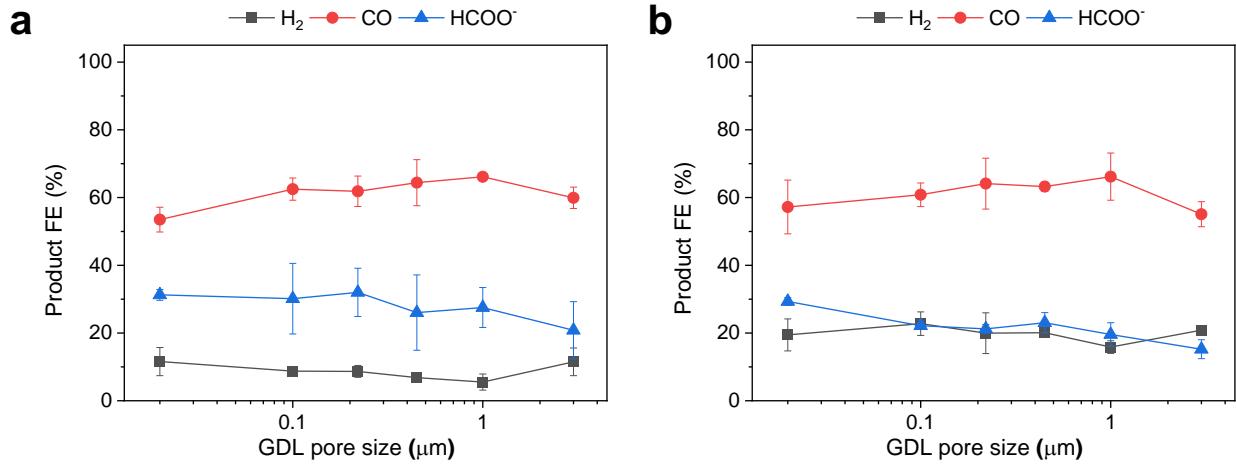
258 **Fig. S13.** Schematic of the experimental setup used to perform electrochemical CO_2 reduction. A
 259 flow type system is employed, with the catholyte and anolyte continuously recirculated through
 260 the cell from an external reservoir using peristaltic pumps. Gas products are analyzed using a gas
 261 chromatograph and formate is analyzed using liquid chromatography. Note: items in the
 262 schematic are not drawn to scale.

263



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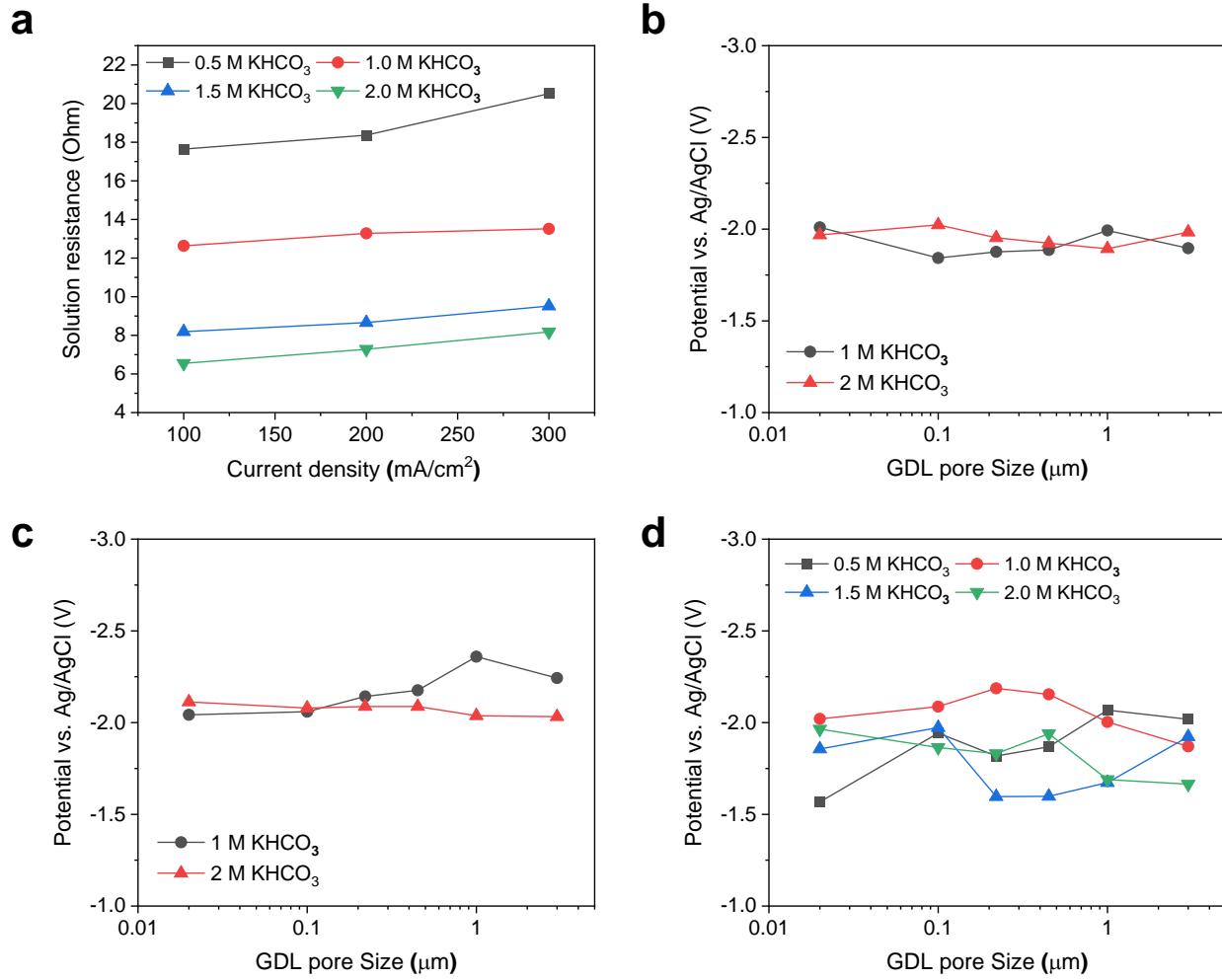
265 **Fig. S14.** Electrochemical CO₂ reduction FE results with 2 M KHCO₃ as the electrolyte. (a), (b)
266 and (c) show the product FE data for Ag/PTFE as a function of GDL pore size under cathodic
267 current densities of 100, 200 and 300 mA/cm² respectively. (d) is the corresponding color
268 contour map of the HCOO⁻ FE data for Ag/PTFE as a function of current density and GDL pore
269 size.



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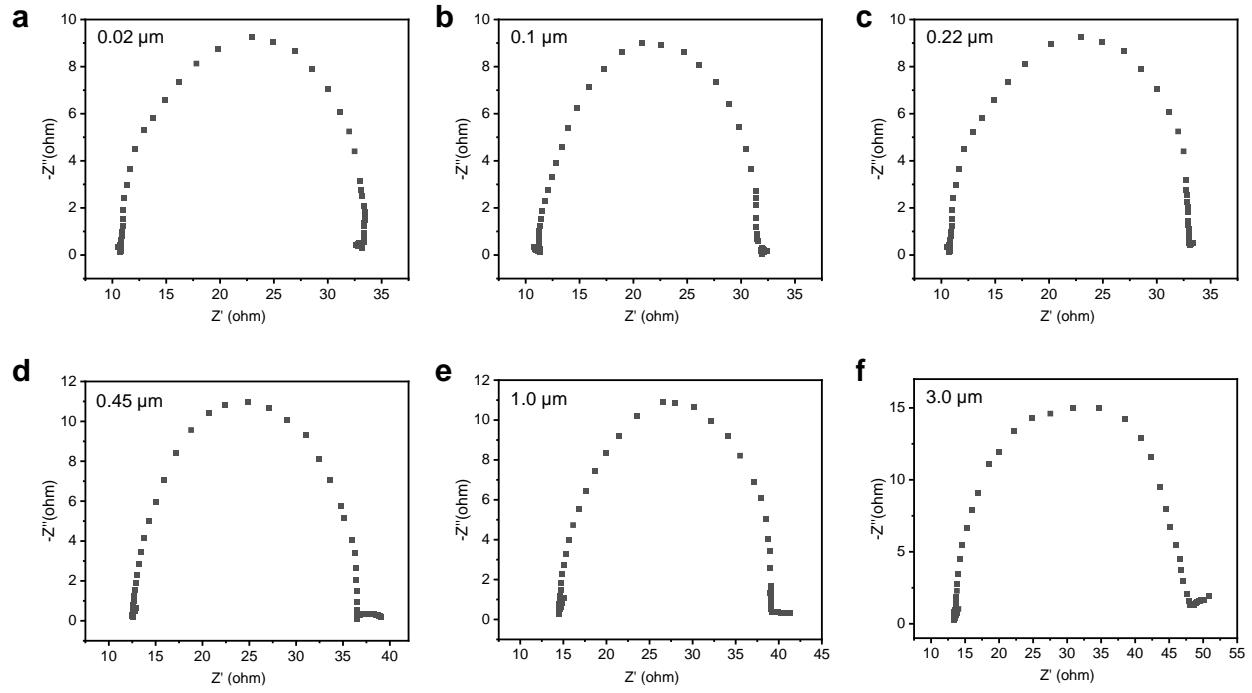
271 **Fig. S15.** Product FE data for Ag/PTFE as a function of GDL pore size at a cathodic current
 272 density of 300 mA/cm² in (a) 0.5 M KHCO₃ and (b) 1.5 M KHCO₃ electrolyte.

273



275 **Fig. S16.** (a) Solution resistance measured using electrochemical impedance spectroscopy (EIS)
 276 at various applied current densities. (b), (c) and (d) are the applied potentials (after IR correction)
 277 as a function of GDL pore size for the different buffer concentrations at cathodic current
 278 densities of 100, 200 and 300 mA/cm² respectively.

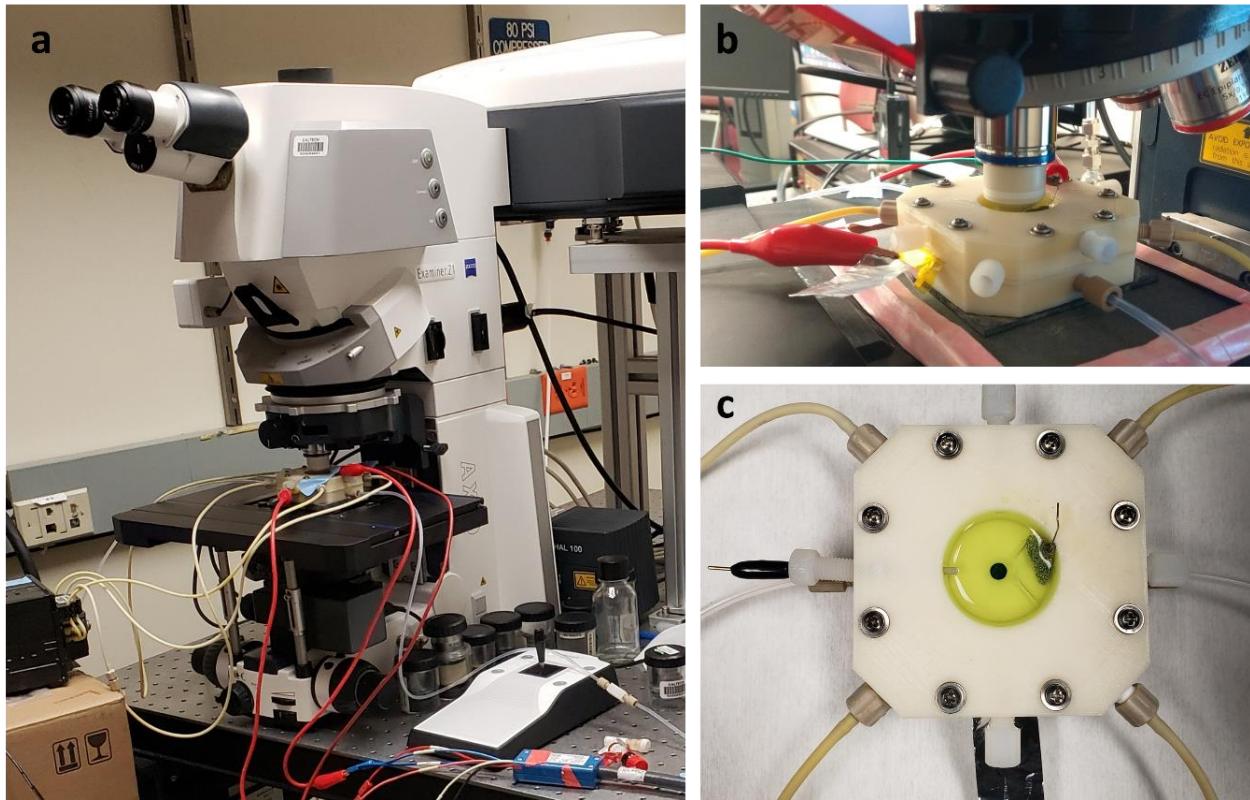
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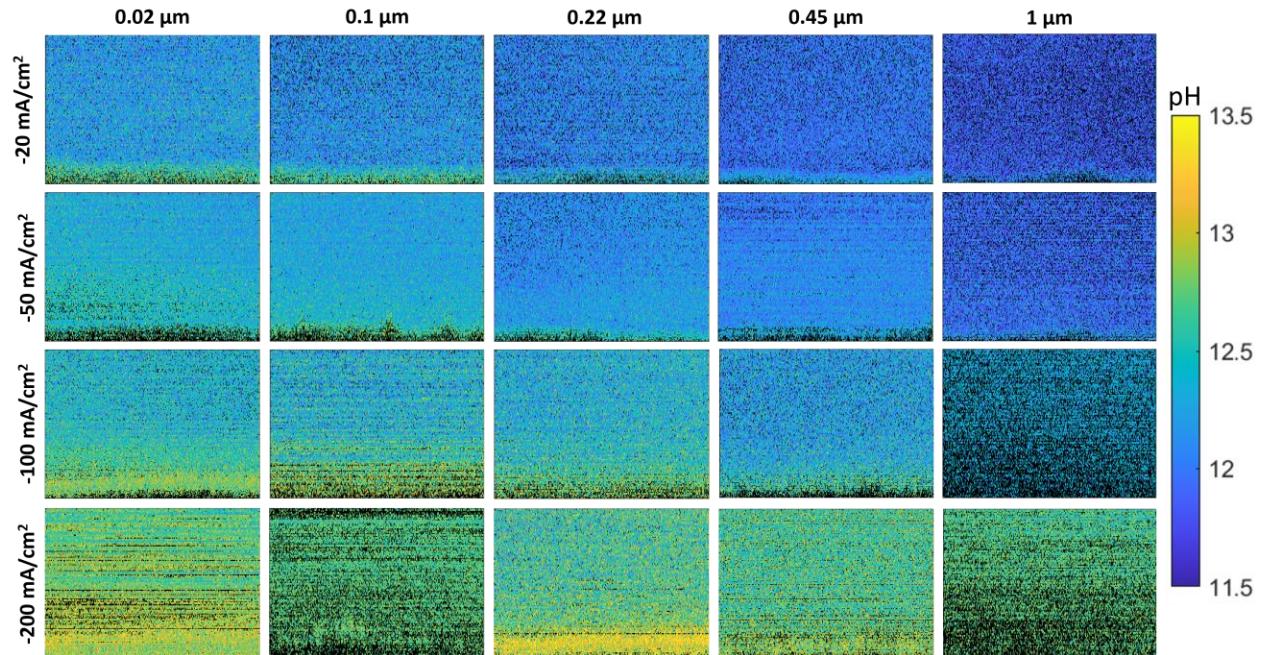
281 **Fig. S17.** Electrochemical impedance spectroscopy (EIS) measurements for the different
 282 Ag/PTFE(X) samples: (a) Ag/PTFE(0.02), (b) Ag/PTFE(0.1), (c) Ag/PTFE(0.22), (d)
 283 Ag/PTFE(0.45), (e) Ag/PTFE(1.0) and (f) Ag/PTFE(3.0).

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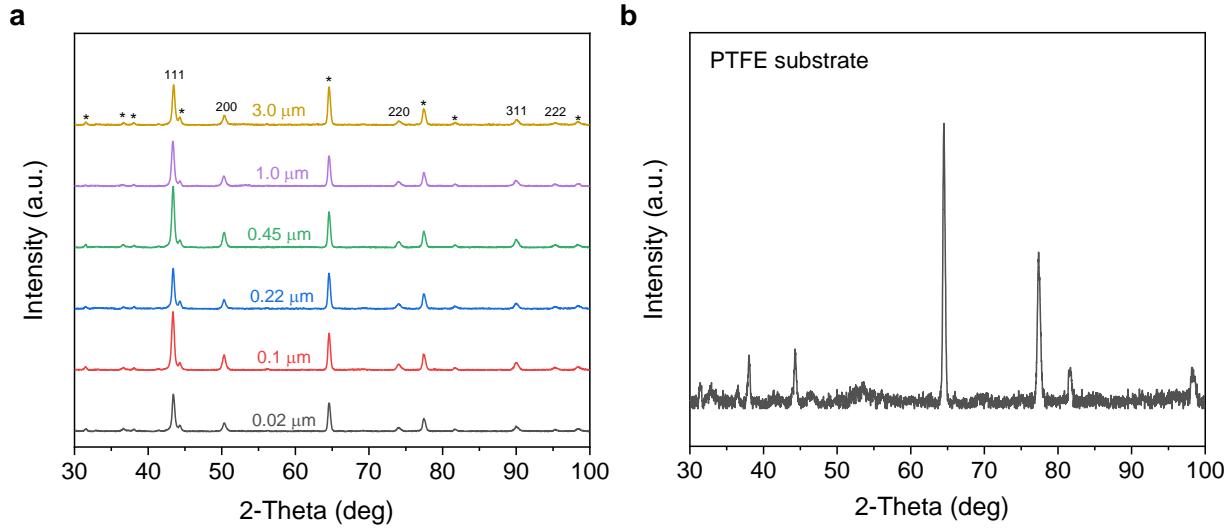


285 **Fig. S18.** Photographs of the confocal microscopy setup used to map the local pH value. (a)
286 Zeiss LSM 880 confocal microscope with a 63x water immersion objective and an
287 electrochemical flow cell. (b) Close-up image of the electrochemical cell. (c) Top view of the
288 electrochemical cell, filled with APTS-spiked electrolyte.

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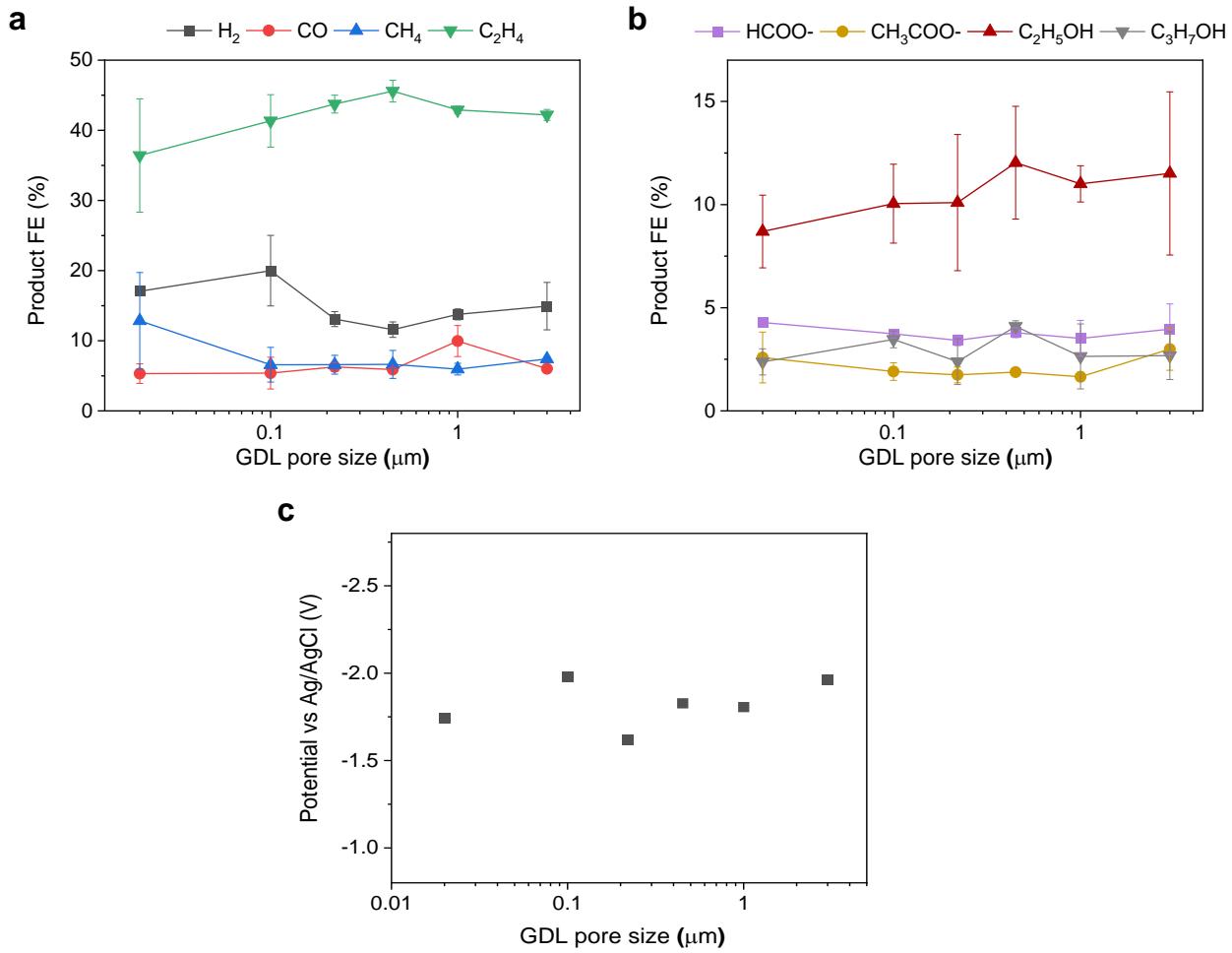


292 **Fig. S19.** Representative pH maps as a cross section through the plane perpendicular to the
 293 electrode surface for different Ag/PTFE samples at four different cathodic current densities
 294 between 20 mA/cm^2 and 200 mA/cm^2 . The dimension of each of the maps is $120 \mu\text{m}$ in x and 50
 295 μm in z.



297 **Fig. S20.** (a) XRD spectrums of Cu sputtered onto PTFE substrates with various pore sizes. The
 298 peaks labelled with a (*) correspond to that of the underlying PTFE substrate. (b) XRD spectrum
 299 of the bare pristine PTFE substrate.

300



301

302 **Fig. S21.** Electrochemical CO_2 reduction results with Cu sputtered on PTFE substrates of various
 303 pore sizes. A constant cathodic current density of $200 \text{ mA}/\text{cm}^2$ was applied and 1 M KHCO_3 was
 304 used as the electrolyte. Multicarbon (C_{2+}) products are ethylene (C_2H_4), acetate (CH_3COO^-),
 305 ethanol ($\text{C}_2\text{H}_5\text{OH}$) and 1-propanol ($\text{C}_3\text{H}_7\text{OH}$). (a) Gas product FE as a function of pore size. (b)
 306 Liquid product FE as a function of pore size. (c) Applied potential as a function of pore size.

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