Electronic Supplementary Material (ESI) for Journal of Materials Chemistry A. This journal is © The Royal Society of Chemistry 2023

1	Supporting information						
2	Assessing the Potential of Biochar Derived from Sewage Sludge for Photoelectrode						
3	Fabrication: Methods, Mechanisms, and Conclusions						
4	Chuangxin Peng <sup>a1</sup> , Jing Huang <sup>b1</sup> , Min Ruan <sup>c</sup> , Haihao Peng <sup>a</sup> , Meiying Jia <sup>d</sup> , Jing Tong <sup>a</sup> ,						
5	Weiping Xiong <sup>a*</sup> , Zhaohui Yang <sup>a*</sup>						
6							
7	a College of Environmental Science and Engineering, Hunan University, Changsha 410082, P.R.						
8	China;						
9	b State Key Laboratory of Utilization of Woody Oil Resource, Hunan Academy of Forestry,						
10	Changsha 410004, PR China;						
11	c School of Energy and Power Engineering, Changsha University of Science & Technology,						
12	Changsha 410076, PR China;						
13	d College of Environmental Science and Engineering, Central South University of Forestry						
14	Technology, Changsha 410004, P.R. China;						
15							
16	* Corresponding author at: College of Environmental Science and Engineering, Hunan University, Changsha 410082, P.R. China;						
17	E-mail address: xiongweiping@hnu.edu.cn (W.P. Xiong)						
18	College of Environmental Science and Engineering, Hunan University, Changsha 410082, P.R. China;						
19	E-mail address: yzh@hnu.edu.cn (Z.H. Yang)						
20	1 These authors contributed equally to this article.						

Text 1: To explore the effect of different biochar dosing amounts on TC degradation,
four gradients of 200mg, 300mg, 400mg, and 500mg were set for the specific biochar
dosing levels. To explore the effect of different annealing temperatures on the effect of
TC degradation, three temperature gradients of 250°C, 350°C, and 450°C were set to
fabricate the electrodes.

Text 2 : Samples were filtered through 0.22  $\mu$ m cellulose acetate syringe membrane filters, and TC concentration was determined by (HPLC, Agilent 1260 series, USA) and Agilent TC-C18 column (4.6 × 250 mm, 5  $\mu$ m). The mobile phase was a mixture of 80% formic acid and 20% acetonitrile. The flow rate and injection volumes were 0.5 mL/min and 20  $\mu$ L, respectively. The wavelength of the detector was set to 354 nm. The retention time of TC was 3.5 min.

Text 3: Determination of the photodegradation intermediates of TC was carried out on 32 an LC-MS system equipped with a 6460 HPLC (Agilent, USA) and an API 3000 mass 33 analyzer. The HPLC column was a Kromasil C18 column (4.6 × 100 mm) from Agilent. 34 35 0.1% of formic acid aqueous solution and acetonitrile were used as mobile phases A and B, respectively. The eluent flow rate keeps at 0.45 mL/min. An injection volume 36 of 10 µL was used in the analysis, and the column temperature at 302 K (30°C). Linear 37 gradient elution as follows: The gradient solvent was as followed: A=90% (0 min), 10% 38 (10 min), 10% (14 min), 90% (15 min), 90% (16 min). Max pressure limit: 1300 bar. 39 Text 4: The biochar electrode is used as the anode, the Platinum sheet electrode as the 40 41 counter electrode, and Ag/AgCl as the reference electrode. The biochar electrode and 42 the platinum sheet electrode were placed at an angle of 120°. The light-facing side of

43 the biochar electrode is connected to the platinum sheet of the platinum sheet electrode
44 clip. A bias voltage of 1.5V was applied after the open circuit voltage was stabilized.
45 The lamp is turned on after the current is stabilized.

46 Text 5: The cost of the photoelectrode includes chemicals, equipment, and time. In this part, we considered the cost of chemical and electricity used for corresponding 47 equipment. The chemicals used include foam nickel (\$3.79/0.06m<sup>3</sup>), gelatin 48 (\$7.58/500g), sodium dodecyl sulfate (\$3.94/500g), high-purity nitrogen gas 49 50 (\$16.83/40L), high-purity argon gas (\$22.44/40L), ethanol (\$5.61/5L) and acetone 51 (\$4.14/500ml). Each foam nickel can prepare 45 substrates. To prepare 4 photocathodes, the following are needed: 0.4 grams of gelatin, 10mg of sodium dodecyl 52 sulfate, 0.045L nitrogen gas (0.03Mpa), and 1.071L argon gas (0.03Mpa); 40 ml of both 53 54 ethanol and acetone each. In the end, the chemical cost of producing each photoelectrode is \$0.12. The power of the tube furnace is 1.2kw, the power of the freeze 55 dryer is 0.85kw, and the cost of electricity is \$0.125 per Kw\*h. To prepare 1 56 photocathode, the tube furnace consumes 0.02754 Kw\*h of electricity, and the freeze 57 dryer consumes 0.3699 Kw\*h of electricity. Therefore, the electricity cost of producing 58 each photoelectrode is \$0.25. Without taking into account equipment loss and labor 59 costs, the production cost of each photocathode is only \$0.37. For a large synthesis, the 60 61 average cost could be lower.

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63 Fig. S 1 EDS elemental mapping of BCPE-700(a-d); BCPE-800(e-h) and BCPE-



900(i-l).





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Fig. S5 Preparation process diagram of biochar photoelectrode.



70 Fig. S6 Comparison of the morphology of photoelectrodes prepared by different

methods.

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Matarial	BET Surface Area	Total pore volume	
Material	(m <sup>2</sup> /g)	$(cm^3/g)$	
Sludge	16.27	0.05753	
BC-700	67.53	0.1735	
BC-800	83.40	0.1631	
BC-900	141.4	0.2085	

75 Table S2 The composition and ratio of each sample were measured by XPS.

element (%)					2.7/27		o / c
Category	Ν	С	0	Н	N/C	H/C	O/C
Sludge	3.89	26.78	26.56	4.21	0.15	0.16	0.99
BC-700	1.40	15.66	11.11	0.60	0.09	0.04	0.71
BC-800	0.83	15.07	7.74	0.46	0.06	0.03	0.51
BC-900	0.50	14.55	7.13	0.31	0.03	0.02	0.49
Gelatin	16.62	52.65	26.01	5.26	0.32	0.10	0.49
BCPE-700	9.11	33.31	18.71	2.82	0.27	0.08	0.56
BCPE-800	8.55	32.82	17.97	2.70	0.26	0.08	0.55
BCPE-900	8.56	33.27	15.95	2.73	0.26	0.08	0.48

Table S3 Variation of TOC with degradation time.

Time (min)	0	15	30	45	60
TOC (mg/L)	8.962	9.978	2.410	1.611	1.715
Efficiency (%)	0.00	11.34	73.11	82.02	80.86

Parameters	Tap water	River water	Medical wastewater
pH	7.13	7.67	7.72
TOC	0.603 mg•L <sup>-1</sup>	22.61 mg•L <sup>-1</sup>	64.23 mg•L <sup>-1</sup>

79 Table S4 The quality parameters of tap water, river water and medical wastewater.

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81 Table S5 Information and proposed structure of the degradation intermediates from

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TC.

D 1 (	Molecular	1	Proposed Structure	
Products	Formula	m/z		
TC	$C_{22}H_{24}N_2O_8$	445	HO CH <sub>3</sub> <sup>H<sub>3</sub>C<sub>N</sub><sup>CH<sub>3</sub></sup> OH O OH O O OH O OH O O</sup>	
P1	$C_{20}H_{20}N_2O_8$	417	HO CH <sub>3</sub> NH <sub>2</sub> OH O OH O OH O OH	
P2	C <sub>20</sub> H <sub>27</sub> NO <sub>7</sub>	393	CH <sub>3</sub> NH <sub>2</sub> OH OH OH OH OH OH OH	
Р3	$C_{17}H_{16}O_{6}$	317	CH <sub>2</sub> OH OH OH OH O OH O	
Р4	$C_{17}H_{16}O_5$	301	CH <sub>2</sub> OH O OH O	

Р5	$C_{16}H_{18}O_4$	274	
P6	$C_{22}H_{22}N_2O_9$	459	$CH_3 HO H_3C N^{-CH_3} OH $
Ρ7	$C_{20}H_{20}N_2O_8$	415	CH <sub>3</sub> HO NH <sub>2</sub> OH O OH O O
P8	$C_{19}H_{21}N_1O_8$	392.1	CH <sub>3</sub> HO NH <sub>2</sub> OH OH OH OH OHO
Р9	C <sub>19</sub> H <sub>19</sub> NO <sub>5</sub>	340	CH <sub>3</sub> NH <sub>2</sub>
P10	C <sub>16</sub> H <sub>15</sub> NO <sub>5</sub>	301	CH <sub>2</sub> OH O OH O
P11	C <sub>12</sub> H <sub>19</sub> NO <sub>4</sub>	242.1	